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# Trimethylsilyldiazomethane – A Mild and Efficient Reagent for the Methylation of Carboxylic Acids and Alcohols in Natural Products

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**Summary.** Esterification of various naturally occurring carboxylic acids with trimethylsilyldiazomethane proceeds nearly quantitatively under very mild conditions. Furthermore, highly sterically hindered alcohols can be converted directly to the corresponding methyl ethers. Improved methods for the preparation and quantification of this reagent are described.

Keywords. Trimethylsilyldiazomethane; Methylation; Carboxylic acids; Alcohols; Natural products.

# Introduction

Diazomethane (CH<sub>2</sub>N<sub>2</sub>) is a well-known reagent for methylations [1], but it is highly toxic, thermally labile, and explosive. These disadvantages can be overcome by replacement of one hydrogen of CH<sub>2</sub>N<sub>2</sub> by a trimethylsilyl group. The resulting safe and stable trimethylsilyldiazomethane (*TMS*CHN<sub>2</sub>, **1**) was initially employed mainly for analytical purposes [2]. In the course of the development of methods for the large-scale preparation of *TMS*CHN<sub>2</sub>, this substitute is increasingly used in synthetic applications [3].

In this paper we focus on the improvement of O-methylation of naturally occurring carboxylic acids and alcohols and demonstrate that  $TMSCHN_2$  is more than a non-toxic alternative to diazomethane. In addition, we developed a simplified method for the preparation and a novel and facile quantification procedure to enable a broad applicability in partial-synthetic work.

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#### **Results and Discussion**

# Preparation of Trimethylsilyldiazomethane

Since *Lappert* and *Lorberth* [4] reported the first preparation of  $TMSCHN_2$  in 1967, several synthetic approaches for the preparation of the title compound **1** have been published. Among these methods, the diazo-transfer reaction of trimethylsilyl-methylmagnesium chloride with diphenyl phosphorylazidate (*DPPA*) [5] (Scheme 1) is the method of choice, because it is most practical and allows a high-yield and large-scale preparation. *DPPA* is commercially available. However, we prepared this precursor in a modified way of the synthesis described in Ref. [6]. The large-scale synthesis of *TMSCHN*<sub>2</sub> is characterised by a very extensive purification followed by a change of the solvent system from  $Et_2O$  to *n*-hexane [5]. We found that the transfer to *n*-hexane is not necessary, because the original  $Et_2O$  solution is also reactive and can be stored without decomposition for several months.

# Determination of the Concentration

The quantification procedure of **1** by NMR (Ref. [5]) is rather complex and impractical. We developed a simple UV-spectroscopic method to determine the concentration of the final etheric solution. The UV-spectrum of **1** shows two maxima at 237.0 and 402.0 nm. To minimize interference with aromatic impurities we chose the less intensive UV-maximum at 402.0 nm ( $\varepsilon = 25.9 \text{ dm}^3 \text{ mol}^{-1} \text{ cm}^{-1}$ ) for this

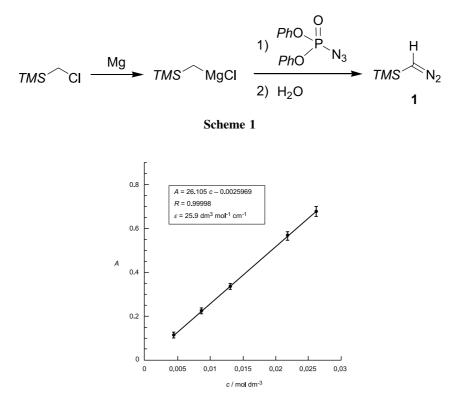


Fig. 1. Calibration graph of  $TMSCHN_2$  in etheric solution

# Trimethylsilyldiazomethane

purpose. The calibration was done with five authentic samples purchased from Sigma-Aldrich. Each one was measured at five dilutions in triplicate or duplicate. The resulting graph shows a high linearity (*R*) in the range between 5–25 mmol dm<sup>-3</sup> with small deviation (Fig. 1), thus the concentration of **1** can be determined via  $\varepsilon$  or the linear equation.

# Preparation of Methyl Esters

Naturally occurring carboxylic acids are cheap, easily available, and enantiomerically pure starting materials for the partial synthesis of biologically active compounds. In many synthesis strategies the conversion to the methyl ester is thereby required [7–11]. The preparation of methyl esters from carboxylic acids can be

| $R - C'_{OH} \xrightarrow{TMS} N_2 \\ R - C'_{OH} \xrightarrow{TMS} R - C'_{OCH_3} \\ 2a - 2g \\ 3a - 3g \\ 3a - 3g$ |  |  |         |  |
|--|--|--|---------|--|
| Entry  | Carboxylic acid  | Product  | Yield/% |  |
| 1  | CH <sub>3</sub> -(CH <sub>2</sub> ) <sub>7</sub> _(CH <sub>2</sub> ) <sub>7</sub> -COOH 2a | CH <sub>3</sub> -(CH <sub>2</sub> )7(CH <sub>2</sub> )7-COOCH <sub>3</sub> 3a        | 96      |  |
| 2  | 2b<br><sup>7</sup> COOH  | 3b<br>COOCH3   | 94      |  |
| 3  | 2c<br>COOH   | 3c<br>COOCH3   | 99      |  |
| 4  | ОН<br>СН <sub>3</sub> -(СН <sub>2)7</sub> -СООН <b>2d</b>                                  | ОН<br>СН <sub>3</sub> -(СН <sub>2</sub> ) <sub>7</sub> -СООСН <sub>3</sub> <b>3d</b> | 96      |  |
| 5  | но 2е  |  | 94      |  |
| 6  | HOOC 2f  | HO<br>HO<br>HO   | 91      |  |
| 7  | COOH<br>OH 2g  | COOCH <sub>3</sub><br>OH 3g  | 99*     |  |

Table 1. Methylation of some natural occurring carboxylic acids

<sup>\*</sup> GC-yield; after CC 83% isolated product

achieved with  $TMSCHN_2$  in methanolic benzene. In the reported procedure a home-made solution of  $TMSCHN_2$  in benzene was used for GC-analysis of fatty acids [2].

We tested the reactivity of the etheric solution of this reagent with various natural compounds (di- and triterpenic acids, fatty acids) and found it to be active even in synthetic scales. Because of its toxicity, benzene was replaced by toluene in our method.

All esterifications proceeded instantaneously and nearly quantitatively without any influence of other functional groups. The reaction was easily monitored by the disappearance of the yellow colour of  $TMSCHN_2$ . The results are summarized in Table 1.

# Preparation of Methyl Ethers

Alcoholic hydroxyl groups can be methylated by diazomethane under very mild conditions [12]. For this reason this reagent was frequently used for the methylation of alcohols containing other sensitive groups. *Aoyama* and *Shioiri* reported the methylation of unhindered and moderately hindered primary, secondary, and tertiary alcohols with  $TMSCHN_2$  in the same way [13]. Yields were comparable with those obtained by diazomethane, but in certain cases the results were significantly better [14].

We report here the methylation of the highly sterically hindered alcohols **4a** and **4b**, which were found to be extremely inert to diazomethane and other electrophilic reagents [12, 15, 16]. We compared the etherification with diazomethane (method described by *Neeman*) and with *TMS*CHN<sub>2</sub> (*Aoyama*'s method). The reaction of **4a** and **4b** with *TMS*CHN<sub>2</sub> yielded the respective methyl ethers in medium yields, whereas the methylation with diazomethane failed. The successful methylation was carried out in CH<sub>2</sub>Cl<sub>2</sub> at 0°C in the presence of *FBA*. The results are summarized in Table 2.

| $R^2 \xrightarrow[R^3]{R^3} OH \xrightarrow{\text{reagent}} R^2 \xrightarrow[R^3]{R^1} OCH_3$ |                              |   |         |  |
|---|------------------------------|---|---------|--|
|   | 4a,4b                        | 5a,5b   |         |  |
| Entry   | Alcohol                      | Reagent   | Yield/% |  |
| 1   | 4a<br>H <sub>3CO</sub><br>4a | <i>TMS</i> CHN <sub>2</sub><br>CH <sub>2</sub> N <sub>2</sub> | 42<br>0 |  |
| 2   | OH 4b                        | <i>TMS</i> CHN <sub>2</sub><br>CH <sub>2</sub> N <sub>2</sub> | 51<br>0 |  |

Table 2. Methylation of some highly sterically hindered alcohols

# Experimental

Melting points were obtained on a digital melting point apparatus Büchi 535. Optical rotation: polarimeter 241 MC (Perkin Elmer). IR spectra: infrared spectrometer system 2000 FT (Perkin Elmer). UV-VIS: UV-160A UV-visible recording spectrophotometer (Shimadzu). NMR spectra: Varian Unity Inova 400 (300 K) 5 mm tubes, *TMS* resonance as internal standard. <sup>1</sup>H- and <sup>13</sup>C-resonances were assigned using <sup>1</sup>H, <sup>1</sup>H- and <sup>1</sup>H, <sup>13</sup>C-correlation spectra. HMBC spectra were optimized for 8 Hz. <sup>1</sup>H- and <sup>13</sup>C-resonances are numbered as given in the formulae. Assignments marked with an asterisk are interchangeable. Mass spectra: GC/MS-Detektor HP 6890, 70 eV electron impact; ESI-MS: Finnigan LCQ Deca XP Plus ion trap mass spectrometer configurated for positive ionisation; HR-MS: PE-SCIEX QStar QTOF mass spectrometer using the Ionspray source in the positive ESI mode, exact mass calibration with quinine giving m/z = 325.1916 (100%), [MH]<sup>+</sup>. Materials: Column-chromatography (CC): silica gel 60 (Merck, 70–230 mesh), pore-diameter 60 Å; solvents: cyclohexane:ethyl acetate (=*CH:EtOAc*), dichloromethane:ethyl acetate; thin-layer chromatography (*TLC*): *TLC* plates (Merck, silica gel 60 F<sub>254</sub>, 0.2 mm, 200×200 mm); the compounds were detected in UV light at 254 nm as well as by spraying with molybdatophosphoric acid and by subsequent heating with a heat gun.

#### Preparation of Trimethylsilyldiazomethane (1, C<sub>4</sub>H<sub>10</sub>N<sub>2</sub>Si)

#### A: Diphenylphosphoryl Azide (DPPA)

To a suspension of 7.22 g (111 mmol) of NaN<sub>3</sub> in 80 cm<sup>3</sup> of anhydrous acetone 14.9 g (55.6 mmol) of diphenylphosphoryl chloride were added and stirred at room temp. for 21 h. The suspension was filtered and the solvent removed under reduced pressure. The residual oil was extracted with anhydrous  $Et_2O$  to give the phosphoryl azide as a nearly colourless oil (12.4 g, 81%).

#### B: Trimethylsilylmethylmagnesium Chloride

Mg turnings (1.46 g, 60.0 mmol) were suspended in  $20 \text{ cm}^3$  of anhydrous  $Et_2O$  and treated with  $20 \text{ mm}^3$  of 1,2-dibromoethane to activate the Mg. After stirring at room temp. for 15 min, 1/10 of a solution of 6.13 g (50.0 mmol) of chloromethyltrimethylsilane in  $20 \text{ cm}^3$  of anhydrous  $Et_2O$  were added at once. When the reaction had started, the remaining solution was added dropwise at such a rate that a gentle reflux was maintained throughout the addition. After complete addition, stirring was continued for 1 h at 40°C to give the *Grignard* reagent.

#### C: Trimethylsilyldiazomethane

A solution of 12.4 g (45.0 mmol) of *DPPA* in 50 cm<sup>3</sup> of anhydrous  $Et_2$ O was cooled to  $-10^{\circ}$ C with a NaCl/ice cooling bath. The *Grignard* reagent prepared above was added dropwise at such a rate that the internal temperature maintained below 0°C. The mixture was stirred at 0°C for 2 h, then allowed to stand in an ice bath overnight. After cooling to  $-10^{\circ}$ C, the mixture was slowly treated with about 5 cm<sup>3</sup> of cold H<sub>2</sub>O. The resulting white precipitate was filtered with suction and washed with cold  $Et_2$ O several times. It is recommended to cool the suction flask during filtration. The combined organic layers were washed with ice-water and dried over Na<sub>2</sub>SO<sub>4</sub> yielding 127 cm<sup>3</sup> of a 0.28 *M* solution of *TMS*CHN<sub>2</sub> (79%). This solution was enriched by distillation over a packed column to a final concentration of approximately 1 *M*. The reagent was used without further purification. It can be stored at 4°C without decomposition for at least 6 months. <sup>1</sup>H NMR (400 MHz,  $Et_2$ O-d<sub>10</sub>):  $\delta = 0.09$  (s, (CH<sub>3</sub>)<sub>3</sub>Si), 2.62 (s, HC=N) ppm; <sup>13</sup>C NMR (100 MHz,  $Et_2$ O-d<sub>10</sub>):  $\delta = -1.0$  ((CH<sub>3</sub>)<sub>3</sub>Si), 19.6 (HC=N) ppm; UV-Vis ( $Et_2$ O):  $\lambda_{max}$  (lg $\varepsilon$ ) = 237.0 (3.98), 402.0 (1.41) nm; EI-MS (70 eV): m/z (%) = 114 (52) [M]<sup>+</sup>, 99 (52), 58 (100).

#### General Procedure for the Preparation of Methyl Esters

To a stirred solution of the carboxylic acid (1 mmol) in approximately  $10 \text{ cm}^3$  of toluene: *Me*OH (3:2) an etheric solution of *TMS*CHN<sub>2</sub> was added dropwise until the yellow colour persisted (1.1–1.5 mmol). The mixture was stirred for 30 min at room temperature and concentrated to give the corresponding methyl ester.

#### Methyl oleate (3a)

The product was purified by CC using CH:EtOAc = 5:1 as eluent. Yield 96%; pale yellow oil;  $R_f = 0.53$  (silica, CH:EtOAc = 5:1); NMR-data are in accordance with those reported in Ref. [17].

#### Methyl abietate (3b)

The product was purified by CC using CH:EtOAc = 9:1 as eluent. Yield 94%; pale yellow oil;  $R_f = 0.40$  (silica, CH:EtOAc = 9:1); NMR-data are in accordance with those reported in Refs. [18, 19].

#### Methyl dehydroabietate (3c)

The product was purified by CC using CH:EtOAc = 9:1 as eluent. Yield 99%; white crystals, mp 61–62°C;  $R_f = 0.48$  (silica, CH:EtOAc = 9:1); NMR-data are in accordance with those reported in Ref. [20].

#### Methyl 13-hydroxylinoleate (3d)

The product was purified by CC using CH:EtOAc = 2:1 as eluent. Yield 96%; pale yellow oil;  $R_f = 0.40$  (silica, CH:EtOAc = 2:1); NMR-data are in accordance with those reported in Ref. [21].

#### Methyl oleanolate (3e)

The product was purified by recrystallization from anhydrous *Me*OH. Yield 94%; white solid, mp 200–202°C;  $R_f = 0.59$  (silica, CH<sub>2</sub>Cl<sub>2</sub>:*Et*OA*c* = 3:1); NMR-data are in accordance with those reported in Refs. [22, 23].

#### Methyl glycyrrhetate (3f)

The product was purified by recrystallization from *Et*OH:H<sub>2</sub>O (1:1). Yield 91%; white crystals, mp 248–250°C;  $R_f = 0.41$  (silica, CH<sub>2</sub>Cl<sub>2</sub>:*Et*OA*c* = 4:1); NMR-data are in accordance with those reported in Ref. [24].

#### Methyl salicylate (3g)

The product was purified by CC using CH:EtOAc = 2:1 as eluent. Yield 83%; colourless oil;  $R_f = 0.50$  (silica, CH:EtOAc = 2:1); NMR-data are in accordance with those reported in Ref. [25].

#### General Procedure for the Preparation of Methyl Ethers

*TMS*CHN<sub>2</sub> (1.0 mmol) was added dropwise to a vigorously stirred solution of the respective alcohol (1 mmol) and *FBA* (1 mmol, 8 *M* in H<sub>2</sub>O) in CH<sub>2</sub>Cl<sub>2</sub> (15–20 cm<sup>3</sup>) at 0°C during 20 min. The yellow colour of *TMS*CHN<sub>2</sub> immediately disappeared with evolution of N<sub>2</sub>. Stirring at 0°C was continued,

and three further amounts of  $TMSCHN_2$  (0.5 mmol/0.25 mmol/0.25 mmol) were added dropwise with intervals of 20 min. The ice-cooled mixture was stirred for further 30 min, neutralized with  $Et_3N$  and concentrated. The residue was redissolved in  $Et_2O$  and washed with cold H<sub>2</sub>O. After drying (Na<sub>2</sub>SO<sub>4</sub>) and concentration to dryness the respective methyl ether was purified as described below.

# *Ethyl* { $1R-(1\alpha,4a\beta,4b\alpha,8a\beta,9\alpha,10a\alpha)$ }-1,2,3,4,4a,4b,5,6,8a,9,10,10a-dodeca-hydro-9-methoxy-1,4a-dimethyl-7-(1-methylethyl)-1-methoxycarbonyl-phenanthrene-9 $\beta$ -yl-acetate (**5a**, C<sub>26</sub>H<sub>42</sub>O<sub>5</sub>)

The product was purified by CC using toluene: EtOAc = 3:1 as eluent. Yield 42%; colourless oil;  $R_f = 0.57$  (silica, toluene: EtOAc = 3:1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta = 0.81$  (s, CH<sub>3</sub>-18), 0.95 (d, J = 6.8 Hz, CH<sub>3</sub>-16), 0.96 (d, J = 6.8 Hz, CH<sub>3</sub>-17), 1.03 (t,  $J = \sim 13$  Hz, H-1<sub>ax</sub>), 1.03–1.14 (m, H-11<sub>ax</sub>), 1.16 (s, CH<sub>3</sub>-19), 1.23 (t, J = 7.2 Hz, CH<sub>3</sub>-25), 1.37 (d,  $J = \sim 13$  Hz, H-6<sub>eq</sub>), 1.40 (t,  $J = \sim 11$  Hz, H-9), 1.50–1.60 (m, H-2<sub>ax</sub>, H-2<sub>eq</sub>), 1.52 (d,  $J = \sim 12$  Hz, H-3<sub>eq</sub>), 1.72 (d,  $J = \sim 13$  Hz, H-1<sub>eq</sub>), 1.73 (d,  $J = \sim 13$  Hz, H-11<sub>eq</sub>), 1.73–1.83 (m, H-3<sub>ax</sub>), 1.85 (t,  $J = \sim 13$  Hz, H-6<sub>ax</sub>), 1.91–1.96 (m, H-12<sub>ax</sub>, H-12<sub>eq</sub>), 2.05 (dd, J = 13.0, 2.3 Hz, H-5), 2.16 (sept, J = 6.8 Hz, H-15), 2.22 (d br, J = 10.6 Hz, H-8), 2.53 (d, J = 13.9 Hz, H-22), 2.69 (d, J = 13.9 Hz, H-22'), 3.05 (s, CH<sub>3</sub>-26), 3.60 (s, CH<sub>3</sub>-21), 4.11 (qd, J = 7.2, 2.8 Hz, H-24, H-24'), 5.39 (s, H-14) ppm; <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta = 13.5$  (C-18), 14.2 (C-25), 16.7 (C-19), 18.0 (C-2), 21.3 (C-16<sup>\*</sup>), 21.7 (C-17<sup>\*</sup>), 22.1 (C-11), 26.3 (C-12), 29.7 (C-6), 35.0 (C-15), 36.0 (C-10), 36.9 (C-3), 37.3 (C-1), 40.5 (C-22), 42.2 (C-8), 43.0 (C-5), 47.1 (C-4, C-9), 48.7 (C-26), 51.7 (C-21), 60.3 (C-24), 76.8 (C-7), 118.6 (C-14), 143.6 (C-13), 171.5 (C-23), 179.2 (C-20) ppm;  $[\alpha]_D^{26} = -3.3^{\circ}$  cm<sup>2</sup>g<sup>-1</sup>; (c = 0.34, CH<sub>2</sub>Cl<sub>2</sub>);  $[\alpha]_{546}^{26} = -6.5^{\circ}$  cm<sup>2</sup>g<sup>-1</sup>; (c = 0.34, CH<sub>2</sub>Cl<sub>2</sub>); IR (KBr):  $\bar{\nu} = 3436$  (s), 2950 (s), 2924 (w), 1728 (s), 1463 (m), 1177 (s) cm<sup>-1</sup>; UV (CH<sub>2</sub>Cl<sub>2</sub>):  $\lambda_{max}$  (lg $\varepsilon$ ) = 234.0 (2.12) nm; ESI-MS: m/z (%) = 457 (56) [M + Na]<sup>+</sup>, 403 (38) [M - CH<sub>3</sub>O]<sup>+</sup>, 343 (100); HR-MS: m/z (%) = 452.3345 (100) [M + NH<sub>4</sub>]<sup>+</sup>, calculated for C<sub>26</sub>H<sub>46</sub>NO<sub>5</sub>: 452.3376.

#### Isoborneol methyl ether (5b)

The product was purified by CC using CH:EtOAc = 5:1 as eluent. Yield 51%; colourless oil;  $R_f = 0.64$  (silica, CH:EtOAc = 5:1); NMR-data are in accordance with those reported in Ref. [26].

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### References

- [1] Black TH (1983) Aldrichimica Acta 16: 3
- [2] Hashimoto N, Aoyama T, Shioiri T (1981) Chem Pharm Bull 29: 1475
- [3] For a review see: Shioiri T, Aoyama T (1993) Adv Use Synthons Org Chem 1: 51
- [4] Lappert MF, Lorberth J (1967) Chem Commun 16: 836
- [5] Shioiri T, Aoyama T, Mori S (1993) Org Synth Coll Vol 8: 612
- [6] Shioiri T, Yamada S (1984) Org Synth 62: 187
- [7] Corey EJ, Cantrall EW (1959) J Am Chem Soc 81: 1745
- [8] Wirthlin T, Wehrli H, Jeger O (1974) Helv Chim Acta 57: 351
- [9] Shimagaki M, Tahara A (1975) Tetrahedron Lett 21: 1715
- [10] Geiwiz J, Haslinger E (1995) Helv Chim Acta 78: 818

- [11] Ullah N, Seebacher W, Haslinger E, Jurenitsch J, Rauchensteiner K, Weis R (2002) Monatsh Chem 133: 139
- [12] Neeman M, Caserio MC, Roberts JD, Johnson WS (1959) Tetrahedron 6: 36
- [13] Aoyama T, Shioiri T (1990) Tetrahedron Lett 31: 5507
- [14] Podlech J (1998) J Prakt Chem 340: 679
- [15] Presser A (2000) PhD Thesis University of Graz, Austria
- [16] Presser A, Pötschger I, Haslinger E, Hüfner A (2002) Monatsh Chem 133: 231
- [17] Stamatov SD, Stawinski J (2000) Tetrahedron 56: 9697
- [18] San Feliciano A, Miguel del Corral JM, Gordaliza M, Salinero MA (1993) Magn Reson Chem 31: 841
- [19] Toki M, Ooi T, Kusumi T (1999) J Nat Prod 62: 1504
- [20] Gigante B, Santos L, Marcelo-Curto MJ, Ascenso J (1995) Magn Reson Chem 33: 318
- [21] Kuklev DV, Christie WW, Durand T, Rossi JC, Vidal JP, Kasyanov SP, Akulin VN, Bezuglov VV (1997) Chem Phys Lipids 85: 125
- [22] Umehara K, Takagi R, Kuroyanagi M, Ueno A, Taki T, Chen YJ (1992) Chem Pharm Bull 40: 401
- [23] Patra A, Mitra AK, Ghosh S, Ghosh A, Barua AK (1981) Org Magn Resonance 15: 399
- [24] Yamada Y, Nakamura A, Yamamoto K, Kikuzaki H (1994) Biosci Biotechnol Biochem 58: 436
- [25] Giumanini AG, Verardo G, Geatti P, Strazzolini P (1996) Tetrahedron 52: 7137
- [26] Fomenko VV, Korchagina DV, Yarovaya OI, Gatilov YV, Salakhutdinov NF, Ione KG, Barkhash VA (1999) Russ J Org Chem (Translation of Zh Org Khim) 35: 1006