BRIEF REPORT

Tomato dwarf leaf virus, a New World begomovirus infecting tomato in Argentina

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Abstract Begomovirus infection is becoming a threat in fresh-market tomato in Argentina, where mixed infections with begomoviruses are common. The complete sequence of a begomovirus isolate infecting tomato sampled in Salta was molecularly characterized. Phylogenetic analysis showed that this virus isolate is closely related to previously reported Brazilian, Bolivian and Argentinean begomoviruses. The associated symptoms in *Nicotiana benthamiana* and *Solanum lycopersicum* were determined by biolistic delivery of infectious DNA-A and DNA-B clones. This begomovirus isolate induced leaf mottling, rugosity and dwarfing, and growth retardation in tomato. Based on these symptoms, we propose the name of tomato dwarf leaf virus (ToDLV) for this new begomovirus.

The *Geminiviridae* are a family of circular, single-stranded DNA plant viruses. Members of this family have twinned icosahedral particles. This family comprises four genera: *Mastrevirus, Curtovirus, Topocuvirus* and *Begomovirus* [1]. Members of the genus *Begomovirus* can have either a mono- or bipartite genome, can infect dicotyledonous plants, and are transmitted by the whitefly *Bemisia tabaci* (Gennadious). Begomoviruses are widespread all over the world and cause economically important diseases to horticultural and fiber crops [2–4]. Most of the New World begomoviruses have bipartite genomes (DNA-A and

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DNA-B) of about 2.6 kb each. These components do not share nucleotide identity, except for the common region (CR), which is about 200 bases long [5]. The CR contains sequences that are essential for replication and transcription of the bipartite genome [6]. DNA-A codes for proteins involved in particle encapsidation (CP), viral replication (Rep and REn), regulation of gene expression (TrAP) and AC4 protein [7–9]. The latter protein performs different functions, such as pathogenicity or RNA silencing suppression, depending on the begomovius species [10, 11]. Some begomoviruses encode an additional AC5 protein [12]. On the other hand, DNA-B encodes two proteins (MP and NSP) involved in virus movement [13–16].

The annual tomato production in Argentina is about 900,000 tons, with a total cultivated area of 14,000 ha [17]. At present, Salta and Jujuy are important fresh-markettomato-producing areas, and begomovirus infection is becoming a major threat. Tomato begomoviruses became economically important due to the migration of the B biotype of *Bemisia tabaci* to Brazil in the mid-1990s [18]. The B biotype has also been detected in Argentina [19]. At least 80 begomovirus species whose members infect tomato have been recognized and proposed, 16 of which are from Brazil [20-22]. Tomato yellow vein streak virus (ToYVSV) and soybean blistering mosaic virus (SbBMV) are the only tomato begomoviruses reported in Argentina [23, 24], where mixed infections with begomoviruses are common [25]. This scenario allows recombination and the emergence of novel viruses [26].

Here, we report the molecular characterization, phylogenetic analysis, and measurement of infectivity of a new begomovirus infecting tomato. We sampled tomato collected from Pichanal, province of Salta, Argentina, that showed mottling, dwarfing and leaf deformation. Total DNA was purified with a Nucleospin[®] Plant II total DNA

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purification kit (Macherey-Nagel). Infected plants were identified by polymerase chain reaction (PCR) with universal primers for DNA-A [27]. The entire genome of positive samples was amplified by rolling-circle amplification (RCA) using *phi*29 polymerase (TempliphiTM, GE, Healthcare) [28]. The DNA-A and DNA-B monomers were obtained by digestion with the restriction enzyme *Apa*I, and then ligated to a digested and dephosphorylated pBluescript SK+ vector (Stratagene). Transformed *E. coli* JM109 competent cell clones were evaluated by PCR. The clones carrying the full genome were sequenced by primer walking (MACROGEN, Korea). Contigs of DNA-A and DNA-B components were assembled using Geneious 5.4.4 (Biomatters Development), and pairwise comparison was done using the MegAlign DNAStar Lasergene 8 package.

We determined the complete bipartite genome sequence of the isolate AR:Pichanal:397:08. Both components have the typical genome organization of bipartite New World begomoviruses. DNA-A (JN564749) is 2540 nt long, and five ORFs (Rep, REn, TrAP, CP, AC4) were identified. By contrast, DNA-B (JN564750) contains two ORFs (MP and NSP) and is 2494 nt long.

The CR of DNA-A and DNA-B of AR:Pichanal:397:08 is about 160 nt long, and pairwise comparison showed 96.1 % identity. A more detailed analysis of the CR sequence showed the elements involved in begomovirus replication: the TATA-box, the iterons (Rep-binding site), and the stem-loop containing the nonanucleotide TAA-TATTAC, which is conserved in the geminivirus family [29]. We indentified two iteron motifs TGGTGACA, and one inverted TGTCACCA located upstream of the TATA box (Figure 1).

The comparison of the complete nucleotide sequence of DNA-A of the isolate studied with those of the previously

reported New World begomoviruses showed highest sequence identity to SoBMV (83 %), ToYVSV (81 %) and solanum mosaic Bolivia virus (SoMBoV) (80 %) (Table 1). For the DNA-B nucleotide sequence, the highest identity percentage was 71 % with ToYVSV and with tomato mild leaf curl virus (ToMiLCV) (Table 1). However, only the DNA-B sequence has been reported for ToMiLCV [30, 31], and DNA-A sequences for SoBMV [24]. It was concluded that this sample contained a member of a new begomovirus species.

Full-length DNA-A and DNA-B sequences were aligned with the selected New World begomoviruses using the

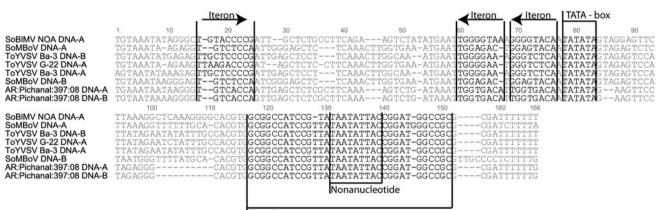
 Table 1
 Pairwise comparison of the complete sequences of DNA-A

 and DNA-B of tomato dwarf leaf virus (ToDLV) with those of the
 most closely related New World begomoviruses

Pairwise identity percentage		
Virus name*	DNA-A	DNA-B
ToYVSV	81	71
SoMBoV	80	68
SiMBoV1	76	62
ToYSV	77	61
ToGMV-YV	79	62
ToMiLCV	-	71
SoBMV	83	-

Nucleotide (nt) percentages were determined

*ToYVSV (EF417915-EF417916): tomato yellow vein streak virus isolate Ba-3; SoMoBoV (HM585435-HM585436): solanum mosaic Bolivia virus; SiMBoV1 (HM585441-HM585442): sida mosaic Bolivia virus 1; ToYSV (DQ336350-DQ336351): tomato yellow spot virus; ToGMV-YV (K02029-K02030): tomato golden mosaic virusyellow vein; ToMiLCV (DQ336352): tomato mild leaf curl virus; SoBMV (EF016486): soybean blistering mosaic virus



Stem loop structure

Fig. 1 Comparison of the common-region sequences of DNA-A and DNA-B between tomato dwarf leaf virus (ToDLV), tomato yellow vein streak virus (ToYVSV), solanum mosaic Bolivia virus (SoM-BoV) and soybean blistering mosaic virus (SoBMV). Vertical lines

demarcate significant replication-related regions: the TATA-box, the iterons, the stem-loop structure, and the conserved nonanucleotide TAATATTAC

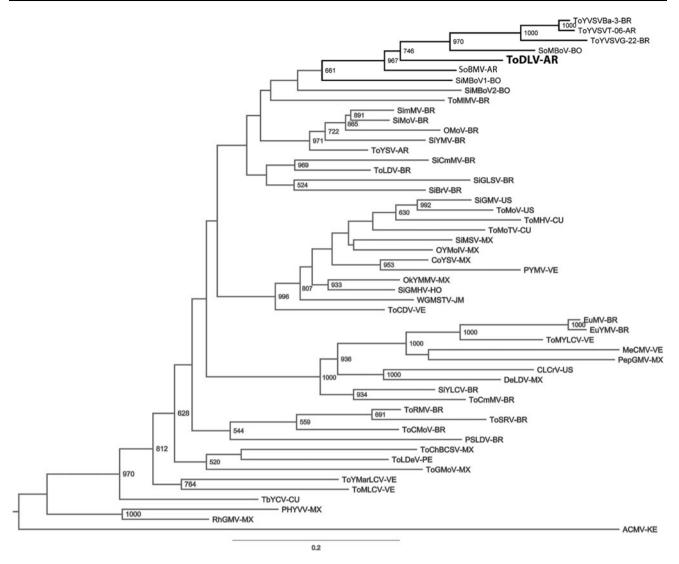


Fig. 2 Maximum-likelihood (ML) phylogenetic trees based on multiple alignments of complete DNA-A sequences of tomato dwarf leaf virus (ToDLV) plus other representative New World begomoviruses (summarized in Supplementary Table S1). Non-parametric

bootstrap values for nodes supported in more than 500 replicates are shown. African cassava mosaic virus was selected as an outgroup

MAFFT algorithm with the iterative E-INS-i method [32, 33]. Phylogenetic analysis of multiple sequence alignments of the complete genomes of New World begomoviruses was done by likelihood mapping analysis with Tree-Puzzle [34, 35] (data not shown).

The phylogenetic relationships between the complete genome sequences of the AR:Pichanal:397:08 isolate obtained in Salta and previously reported New World be-gomoviruses were inferred by the maximum-likelihood criterion using PhyML 3.0 [36] under the GTR+I+G substitution model. Phylogenetic trees with 1000 bootstrap replicates were obtained. Trees constructed from DNA-A sequences showed that AR:Pichanal:397:08 sequences are

grouped into a monophyletic cluster with SoBMV, SoM-BoV and ToYVSV (Figure 2). This close relationship is consistent with the pairwise identity analysis (Table 1) and with previously reported data [37]. This cluster includes all ToYVSV isolates from Brazil and Argentina, as well as SoBMV from Argentina and SoMBoV from Bolivia, suggesting a geographical relationship. Interestingly, the sister group of this cluster contains sida-infecting begomoviruses, showing evidence of a common ancestor that adapts to different hosts.

The DNA-B-based phylogenetic tree confirmed the phylogenetic relationship obtained with DNA-A. The DNA-B sequences of AR:Pichanal:397:08 are most closely

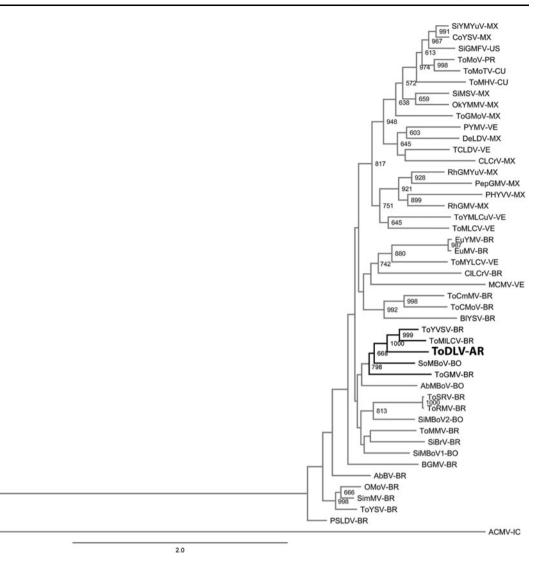


Fig. 3 Maximum-likelihood (ML) phylogenetic trees based on multiple alignments of DNA-B complete sequences of tomato dwarf leaf virus (ToDLV) plus other representative New World begomoviruses

(summarized in Supplementary Table S1). Non-parametric bootstrap values for nodes supported in more than 500 replicates are shown. African cassava mosaic virus was selected as an outgroup

related to those of ToYVSV and SoMBoV. The DNA-B tree displays a rapid and recent burst of begomovirus diversification from the most recent common ancestor, in contrast to the gradual accumulation of diversity of DNA-A (Figure 3).

To evaluate the infectivity of the viral DNAs that were cloned, we performed biolistic inoculation of *Nicotiana benthamiana* and *Solanum lycopersicum* plants using a PDS-1000/He particle delivery system (Bio-Rad). Plasmids containing DNA-A and DNA-B of the AR:Pichanal:397:08 isolate were digested with *ApaI* to purify the monomeric genomes and then re-ligated. The RCA amplification product (5 μ l) of each component was mixed with tungsten particles and then inoculated into six *N. bentamiana* and 11

S. lycopersicum plants with 4-5 leaves, and into the respective control plant [38]. All *N. bentamiana* plants showed chlorosis, wrinkling and leaf deformation. Eight days after inoculation, symptoms developing in eight *S. lycopersicum* plants included leaf mottling, rugosity and dwarfing, and growth retardation (Figure 4). The infection was confirmed by PCR analysis.

We have molecularly characterized a novel member of the genus *Begomovirus* from Pichanal, Salta, Argentina, and established its phylogenetic relationship to other begomoviruses. We have also infected *Nicotiana benthamiana* and *Solanum lycopersicum* plants and determined the symptoms that this novel virus produced on tomato, the original host. Based on these results, we suggest that the

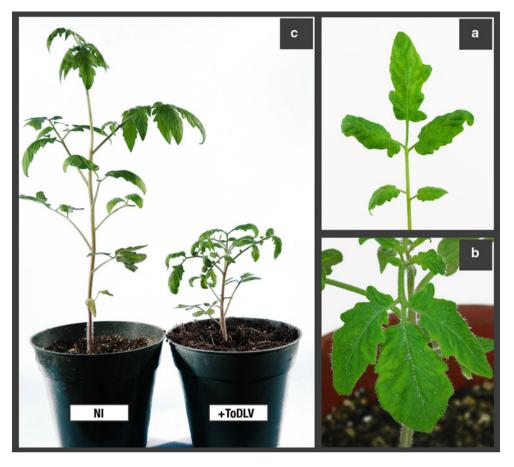


Fig. 4 Detail of leaf symptoms of *Solanum lycopersicum* plants inoculated with DNA-A and DNA-B infectious clones of tomato dwarf leaf virus (ToDLV) by particle bombardment. Dwarfing,

AR:Pichanal:397:08 isolate belongs to a new species and propose the name "Tomato dwarf leaf virus" (ToDLV) for its identification.

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References

- Stanley J, Bisaro DM, Briddon RW, Brown JK, Fauquet CM, Harrison BD, Rybicki EP, Stenger DC (2005) Geminiviridae. In: Fauquet CM MM, Maniloff J, Desselberger U, Ball LA (eds) Virus taxonomy, VIIIth report of the ICTV, Elsevier/Academic Press, London, pp 301–326
- Varma A, Malathi VG (2003) Emerging geminivirus problems: a serious threat to crop production. Ann Appl Biol 142:145–164
- Morales FJ, Anderson PK (2001) The emergence and dissemination of whitefly-transmitted geminiviruses in Latin America. Arch Virol 146:415–441
- Mansoor S, Briddon RW, Bull SE, Bedford ID, Bashir A, Hussain M, Saeed M, Zafar Y, Malik KA, Fauquet C, Markham PG

rugosity (**a**) and mottling (**b**) are shown. Comparison between a non-inoculated (NI) and a ToDLV-inoculated tomato plant. Leaf dwarfing and growth retardation are shown (**c**)

(2003) Cotton leaf curl disease is associated with multiple monopartite begomoviruses supported by single DNA beta. Arch Virol 148:1969–1986

- Hamilton WD, Stein VE, Coutts RH, Buck KW (1984) Complete nucleotide sequence of the infectious cloned DNA components of tomato golden mosaic virus: potential coding regions and regulatory sequences. EMBO J 3:2197–2205
- Fontes EPB, Gladfelter HJ, Schaffer RL, Petty ITD, Hanley-Bowdoin L (1994) Geminivurus replication origins have a modular organization. The Plant Cell 3:405–416
- Hanley-Bowdoin L, Settlage SB, Robertson D (2004) Reprogramming plant gene expression: a prerequisite to geminivirus DNA replication. Mol Plant Pathol 5:149–156
- Settlage SB, See RG, Hanley-Bowdoin L (2005) Geminivirus C3 protein: replication enhancement and protein interactions. J Virol 79:9885–9895
- Sunter G, Bisaro DM (1992) Transactivation of geminivirus AR1 and BR1 gene expression by the viral AL2 gene product occurs at the level of transcription. Plant Cell 4:1321–1331
- Pooma W, Petty IT (1996) Tomato golden mosaic virus open reading frame AL4 is genetically distinct from its C4 analogue in monopartite geminiviruses. J Gen Virol 77(Pt 8):1947–1951
- Dogra SC, Eini O, Rezaian MA, Randles JW (2009) A novel shaggy-like kinase interacts with the Tomato leaf curl virus pathogenicity determinant C4 protein. Plant Mol Biol 71:25–38
- Fontenelle MR, Luz DF, Gomes AP, Florentino LH, Zerbini FM, Fontes EP (2007) Functional analysis of the naturally

recombinant DNA-A of the bipartite begomovirus Tomato chlorotic mottle virus. Virus Res 126:262–267

- Brough CL, Sunter G, Gardiner WE, Bisaro DM (1992) Kinetics of tomato golden mosaic virus DNA replication and coat protein promoter activity in Nicotiana tabacum protoplasts. Virology 187:1–9
- Jeffrey JL, Pooma W, Petty IT (1996) Genetic requirements for local and systemic movement of tomato golden mosaic virus in infected plants. Virology 223:208–218
- Etessami P, Callis R, Ellwood S, Stanley J (1988) Delimitation of essential genes of cassava latent virus DNA 2. Nucleic Acids Res 16:4811–4829
- Sudarshana MR, Berger PH (1998) Nucleotide sequence of both genomic RNAs of a North American tobacco rattle virus isolate. Arch Virol 143:1535–1544
- Programa para el Aumento de la Producción Producción Nacional de Tomate (PACIT). Informe de Progresos 2010-2011. Estadísticas de Producción Nacional, pp 1–13. http://www.inta.gov. ar/laconsulta/. ISSN 1853-6972
- Ribeiro SG, de Ávila AC, Bezerra IC, Fernandes JJ, Faria JC, Lima MF, Gilbertson RL, Maciel-Zambolim E, Zerbini FM (1998) Widespread Occurrence of Tomato Geminiviruses in Brazil, Associated with the New Biotype of the Whitefly Vector. Plant Dis 82:830
- Viscarret MM, Torres-Jerez I, Agostini de Manero E, López SN, Botto EE, Brown JK (2003) Mitochondrial DNA evidence for a distinct New World group of Bemisia tabaci (Gennadius) (Hemiptera:Aleyrodiadae) indigenous to Argentina and Bolivia, and presence of the Old World B biotype in Argentina A. Ann Entomol Soc Am 96(1):65–72
- 20. Fernandes FR, de Albuquerque LC, de Britto Giordano L, Boiteux LS, de Avila AC, Inoue-Nagata AK (2008) Diversity and prevalence of Brazilian bipartite begomovirus species associated to tomatoes. Virus Genes 36:251–258
- Ribeiro SG, Ambrozevicius LP, Avila AC, Bezerra IC, Calegario RF, Fernandes JJ, Lima MF, de Mello RN, Rocha H, Zerbini FM (2003) Distribution and genetic diversity of tomato-infecting begomoviruses in Brazil. Arch Virol 148:281–295
- Fauquet CM, Briddon RW, Brown JK, Moriones E, Stanley J, Zerbini M, Zhou X (2008) Geminivirus strain demarcation and nomenclature. Arch Virol 153:783–821
- 23. Vaghi Medina CG, López Lambertini PM (2010) Genetic diversity of begomovirus infecting tomato in Argentina. In: 6th international geminivirus symposium. 4th international ssDNA comparative virology workshop. Guanajuato, Mexico
- Rodríguez-Pardina PE, Hanada K, Laguna IG, Zerbini FM, Ducasse DA (2011) Molecular characterisation and relative incidence of bean- and soybean-infecting begomoviruses in northwestern Argentina. Ann Appl Biol 158:69–78
- 25. Giachero MS, Bernis ME, López Lambertini PM (2007) Mixed infections with begomovirus is widespread in tomato crops in

Argentina. In: Abstract in 5th International geminivirus symposium. 3rd international ssDNA comparative virology workshop, Ouro Preto, Brazil, p 65

- 26. Davino S, Napoli C, Dellacroce C, Miozzi L, Noris E, Davino M, Accotto GP (2009) Two new natural begomovirus recombinants associated with the tomato yellow leaf curl disease co-exist with parental viruses in tomato epidemics in Italy. Virus Res 143:15–23
- Rojas MR, Gilbertson RL, Maxwell DP (1993) Use of degenerate primers in the polymerase chain reaction to detect whiteflytransmitted geminiviruses. Plant Dis 77:340–347
- Inoue-Nagata AK, Albuquerque LC, Rocha WB, Nagata T (2004) A simple method for cloning the complete begomovirus genome using the bacteriophage phi29 DNA polymerase. J Virol Methods 116:209–211
- 29. Arguello-Astorga GR, Ruiz-Medrano R (2001) An iteron-related domain is associated to Motif 1 in the replication proteins of geminiviruses: identification of potential interacting amino acid-base pairs by a comparative approach. Arch Virol 146:1465–1485
- Albuquerque LC, Martin DP, Avila AC, Inoue-Nagata AK (2010) Characterization of tomato yellow vein streak virus, a begomovirus from Brazil. Virus Genes 40:140–147
- Fernandes JJ, Carvalho MG, Andrade EC, Brommonschenkel SH, Fontes EP, Zerbini FM (2006) Biological and molecular properties of *Tomato rugose mosaic virus* (ToRMV), a new tomatoinfecting begomovirus from Brazil. Plant Pathol 55:513–522
- 32. Katoh K, Misawa K, Kuma K, Miyata T (2002) MAFFT: a novel method for rapid multiple sequence alignment based on fast Fourier transform. Nucleic Acids Res 30:3059–3066
- 33. Katoh K, Kuma K, Toh H, Miyata T (2005) MAFFT version 5: improvement in accuracy of multiple sequence alignment. Nucleic Acids Res 33:511–518
- 34. Schmidt HA, Strimmer K, Vingron M, von Haeseler A (2002) TREE-PUZZLE: maximum likelihood phylogenetic analysis using quartets and parallel computing. Bioinformatics 18:502–504
- 35. Strimmer K, von Haeseler A (1997) Likelihood-mapping: a simple method to visualize phylogenetic content of a sequence alignment. Proc Natl Acad Sci USA 94:6815–6819
- 36. Guindon S, Dufayard JF, Lefort V, Anisimova M, Hordijk W, Gascuel O (2010) New algorithms and methods to estimate maximum-likelihood phylogenies: assessing the performance of PhyML 3.0. Syst Biol 59:307–321
- 37. Wyant PS, Gotthardt D, Schafer B, Krenz B, Jeske H (2011) The genomes of four novel begomoviruses and a new Sida micrantha mosaic virus strain from Bolivian weeds. Arch Virol 156:347–352
- 38. Guenoune-Gelbart D, Sufrin-Ringwald T, Capobianco H, Gaba V, Polston JE, Lapidot M (2010) Inoculation of plants with begomoviruses by particle bombardment without cloning: Using rolling circle amplification of total DNA from infected plants and whiteflies. J Virol Methods 168:87–93