

The underlying mechanisms for the ‘anti-HBc alone’ serological profile

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Abstract The serological pattern, “anti-HBc alone”, characterized by the presence of antibodies against the core antigen of hepatitis B virus (anti-HBc) as the only marker of hepatitis B, is not rare in a diagnostic setting. Depending on the prevalence of HBV infection and the patient group investigated, 1–31% of positive anti-HBc results are isolated positive findings. Anti-HBc alone is frequently observed in intravenous drug addicts, HIV-infected individuals, patients who are coinfecting with HBV and hepatitis C virus, and pregnant women. However, it is not clear how this profile should be interpreted. Several studies have shown that anti-HBc alone is not only compatible with acute and resolved HBV infection but also with chronic infection. The reasons for the lack of HBsAg and anti-HBs in anti-HBc-alone individuals are not clear, but several

mechanisms and possibilities have been suggested that could explain this phenomenon, some of which are delineated in this article.

Introduction

HBcAg is the most immunogenic hepatitis B virus (HBV) component during infection [1]. It is the nucleocapsid of HBV, a 27-nm particle composed of multiple copies of a single polypeptide (P21) [2]. Despite the fact that HBcAg is an internal component of the virion, high titers of HBc antibody (anti-HBc) are produced in virtually all patients who have been exposed to HBV and usually persist, irrespective of ongoing liver disease or clearance of the virus [2, 3]. The high frequency of anti-HBc production is due, at least in part, to the fact that HBcAg can function as a cell-independent antigen [1]. Thus, during the acute phase of infection, anti-HBc of the IgM class predominates. As the infection evolves, anti-HBc IgM levels gradually decline and anti-HBc IgG can persist with slowly decreasing titers for many years [2, 3]. For this reason, antibodies to the core of hepatitis B virus are considered to be the most reliable serological marker of HBV infection [4] and are very cost-effective, especially if compared to molecular markers [5]. Anti-HBc antibodies are called ‘epidemiological markers’ and are widely employed in HBV screening [5]. Once detected, they will guide to additional tests in order to establish the patient’s clinical status [6]. Some anti-HBc-positive patients also carry the hepatitis B surface antigen (HBsAg), indicating ongoing infection, whereas others have the corresponding antibody (anti-HBs), indicating recovery and immunity [7, 8]. However, in a third group of anti-HBc carriers, neither HBsAg nor anti-HBs is detectable in the serum. Subjects that demonstrate this

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serological profile are said to be ‘anti-HBc alone’ [9]. Such a serological profile can be encountered in different population groups and may be explained by different mechanisms, as described in this article.

False positivity to anti-HBc

Depending on the anti-HBc test used and the HBV prevalence where the assay is employed, a certain proportion of individuals with ‘anti-HBc alone’ will be false positive [9]. This false reactivity has been attributed to nonspecific reactions associated with competitive anti-HBc enzyme immunoassays (EIAs), cross-reactivity with interfering serum substances or with IgA or IgM-related molecules produced from nonspecific HBV-activated B-lymphocytes [10]. To minimize nonspecific results, modifications have been made by using different HBcAg antibodies (human, mouse-monoclonal), different detection systems (enzymes, fluorescence, chemiluminescence), and changes in the reaction kinetics (incubation steps, temperature, and time). It has been demonstrated that samples that exhibit reactivity around the cutoff values are most likely to be false positives. Therefore, modifications in cutoff calculations or the use of a broader gray-zone have been proposed [11], but not accepted universally. Cross-reactivity can be diminished by treating samples with reducing agents (dithiothreitol [DTT] or potassium bisulfite [MBS]) [12] and by the employment of neutralization assays, which have been shown to significantly improve the specificity of anti-HBc determination [13].

In vivo studies have allowed the evaluation of the specificity of anti-HBc assays. Ural and Findik [14] observed that almost half of ‘anti-HBc alone’ patients developed primary immune response when challenged with the HBV vaccine. Another study in China also demonstrated that among anti-HBc-alone patients, 72.9% developed a primary anti-HBs response [15], suggesting false reactivity to anti-HBc.

Therefore, because false positive results are very frequently observed, it has been recommended that complementary tests should be employed to confirm anti-HBc reactivity. Such strategies involve the use of another EIA test format, the search for another serological marker (HBsAg, anti-HBs or anti-HBe) or the detection of HBV DNA by an amplification method [9, 11, 16].

Chronic carrier with low levels of HBV

It is generally accepted that the diagnosis of HBV infection is based on the detection of serum hepatitis B surface antigen (HBsAg) and that disappearance of this antigen

indicates the clearance of HBV and resolution of infection. On the other hand, the persistence of HBsAg suggests a chronic-carrier status [17]. However, several studies have shown that HBV DNA has been frequently detected in patients with chronic liver disease who are negative for HBsAg and positive for antibodies to core antigen [17–19], suggesting that a large number of anti-HBc-positive healthy individuals are ‘latent’ carriers of HBV with ongoing viral replication in the liver tissue. Although little is known about the molecular mechanisms of viral persistence, this finding might reflect the difficulty in eliminating HBV covalently closed circular DNA (cccDNA) from the hepatocyte nucleus, and this defines the HBV chronic-carrier state [19, 20].

The persistence of HBV/DNA in the liver after HBsAg clearance has been demonstrated by molecular techniques (PCR and in situ hybridization), suggesting that viral DNA may exist in a complete form in the hepatocyte. In addition, the presence of extrachromosomal HBV DNA has been detected in some patients, suggesting that latent HBV infection may persist in hepatocytes without production of viral particles [21]. Analysis of liver histopathology by immunohistochemistry in anti-HBc-alone patients has revealed that the expression of HBsAg or HBcAg in liver tissue is comparable to that in chronic HBV carriers, except that the signals are lower. This finding could be due to lower expression levels of the viral proteins and/or to a weaker recognition by the immune system of the antigens produced [22]. Thus, some patients may have already had serological markers, but subsequently lost them, while expressing low levels of HBV replication associated with ongoing liver disease. In summary, some anti-HBc-alone individuals are characterized as HBV carriers when the ‘s’ antigen is not present in the bloodstream years after infection or its low levels limit detection by routinely available commercial assays. In this case, the chronic infection diagnosis must be established through HBV DNA detection by PCR or an alternative nucleic acid amplification procedure with detection limits of 10–100 genomes per ml [9]. The vaccination of these individuals against HBV could be worthwhile for their characterization, since they are unable to produce anti-HBs antibodies [15, 23].

Infection by HBV mutants

Several studies have suggested that point mutations, deletions, or rearrangements in some genes of the HBV genome that interfere with gene expression or lead to the production of antigenically modified ‘s’ proteins may be responsible for the lack of HBsAg detection in the bloodstream.

Infection by mutants of the a-determinant region of hepatitis B surface antigen

The "a" determinant of hepatitis B virus consists of a serologically defined region, located between amino acid residues 124 and 147 of the hepatitis B surface antigen (HBsAg), which induces a protective immune response common to all HBV subtypes [24]. Variations in its primary structure have been demonstrated to markedly alter the antigenic conformation and antigenicity of HBsAg [25]. Variant virus with an amino acid substitution at residue 145 from glycine to arginine (G145R) [26] or lysine to glutamic acid at residue 141 (L141E) [27], amongst others [28, 29], within the "a" determinant, have been identified and may escape protection from vaccine-induced anti-HBs-specific neutralizing antibodies. However, it has been demonstrated that variability in the common "a" determinant not only can favor viral escape from neutralizing antibodies but also reduces *in vitro* HBsAg detection in routine immunoassays, promoting mutant HBsAg diagnosis escape when HBsAg is established as the only infection marker for HBV infection [25, 30, 31]. These mutations may promote variations in the tertiary structure of HBsAg, hindering the correct formation of disulphide bonds among cysteine residues of the 'a' determinant, which alters protein immunological properties, leading to false negative results. Overall, it is also important to emphasize that failure in HBsAg variant detection is due to the antigenic exchanges described above and not because of low HBsAg levels in the bloodstream, although exchanges may also affect the overlapping polymerase gene and, consequently, viral replication capacity [30].

Infection by mutants of the polymerase gene of hepatitis B virus

Mutations in the polymerase gene may produce changes in the overlapping 's' gene. Similarly, mutations within the 's' gene may produce changes in the overlapping polymerase gene [30]. Therapy with lamivudine (LMV) often results in exchanges in the viral polymerase catalytic center, some of which induce the selection of HBV mutants that are resistant to treatment with this nucleoside analogue, maintaining low levels of replication. These mutations produce exchanges in the overlapping s gene, which results in reduction in HBsAg antigenicity and expression. The most common amongst them affects the YMDD (tyrosine-methionine-aspartate-aspartate) motif, where the methionine (M) residue at amino acid 204 is replaced by isoleucine (rtM204I) or valine (rtM204V), accompanied by a compensatory leucine-to-methionine exchange at codon 180 (rtL180M/M204V) [32]. This mutation produces I195M and W196S exchanges in the HBsAg, which alter

the structural integrity of the protein and affect the binding to anti-HBs antibodies present in EIA assays, and consequently its *in vitro* detection [33]. LMV-resistant mutants often show reduced replication capacity when compared to wild-type virus, and its emergence is associated with the reappearance of DNA/HBV in the serum. Recently, it was demonstrated that two overlapping s gene mutations (sP120T and sG145R) produce exchanges of rtT128N and rtW153Q in the polymerase protein, which were found to partially restore the replication *in vitro* of the LMV-resistant HBV phenotype, maintaining resistance to the antiviral agent. These exchanges produce a stop codon in the overlapping s gene, resulting in deficient secretion of HBsAg and virus [34]. Other mutations in the polymerase gene have been reported in individuals undergoing antiviral therapy. Such mutations can introduce a stop codon in the surface antigen and include rtM204I (sW194stop) [35, 36] and rt V207I (Sw199stop) [37].

New drugs, such as adefovir, entecavir and tenofovir, have shown efficacy in suppressing viral replication and therefore seem to be promising candidates for treatment of chronic hepatitis B [38]. However, their long-term efficacy in association with their potential to induce mutations has not been widely evaluated [38, 39]. The use of entecavir for LMV-resistant HBV in particular has been questionable, since both drugs may promote HBV mutations by similar mechanisms [40]. Hepatitis B virus resistance to adefovir and tenofovir has also been described. Mutations rtN236T [41] and rtA194T [42] are associated with adefovir and tenofovir resistance, respectively, but do not affect HBsAg expression. However, the mutation rtV191I, which was selected after 24 weeks of adefovir therapy, was shown to simultaneously create a stop codon in the surface antigen (sW182stop), reducing HBsAg levels in the serum [43]. Several HBV mutations associated with drug resistance have been detected and are well-elucidated [44]. There are many other drugs currently under clinical investigation, such as Emtricitabine, Clevudine and Telbivudine [45]. However, new mechanisms of drug resistance will certainly emerge, including those involving changes in the polymerase gene, which may impair secretion, production and/or detection of HBsAg [46, 47].

Infection by mutants of the s-promoter region of hepatitis B virus

Mutations in the HBV regulatory regions can also affect HBsAg secretion and synthesis and lead to the anti-HBc-alone serological profile. The HBV envelope gene is controlled by two different promoters: the pre-S and the S promoter, which regulate transcription of a 2.4- and a 2.1-kb mRNA, respectively. The pre-S promoter is located 5' of the first in frame ATG of the large (pre-S1) envelope

protein. The S-promoter is found in the pre-S gene coding region and is a TATA-less promoter. The two mRNAs transcribed from the S gene code for 3 S-proteins (small, middle, and large) sharing the 226 amino acids at the C-terminus [48]. Deletions in pre-s region can promote S-promoter removal, which blocks HBsAg transcription. This can result in reduction of HBsAg protein synthesis and secretion [49], or even intracellular viral retention, leading to a low concentration of extracellular S gene products in the bloodstream [50] despite active viral replication.

In addition, for appropriate HBV assembly and secretion, a specific ratio between the small and large surface protein is necessary, with the amount of small S-protein predominating [48]. Thus, the reduced synthesis of small surface antigen results in the accumulation of large surface protein in the endoplasmic reticulum, which can only be secreted if small S-protein is present in adequate proportions [51]. On the other hand, overexpression of the large S-protein may result in retention of small S-protein within cells, quantitative decrease of HBsAg in the serum and lack of viral secretion [52]. A single gene-defective HBV population may coexist with a wild-type population, helping in viral rescue. These alterations could be part of a dynamic process in which the existing mutants gradually may die out in order to be replaced by mutants originating from the wild type [48]. This phenomenon may be related with the control of viral replication and the evasion of immune surveillance, which account for occasional life-long persistence of HBV infection and the discrepancy between the presence of HBV DNA and HBsAg in serum [51]. These mutants have been isolated frequently from individuals with persistent viral infection, including those with cirrhosis and hepatocellular carcinoma.

Coinfection with other hepatoviruses

The suppression of HBV replication and its gene expression is known to occur during the late phases of chronic and acute infection and may eventually lead to disappearance of the HBsAg protein in the serum [21, 53]. In the case of HCV/HDV coinfection/superinfection, HBV replication may also be suppressed, leading to the ‘anti-HBc alone’ serostatus.

HBV and HCV are parenterally transmitted and share common routes of infection, and therefore, combined infection may occur, particularly in areas where both agents are endemic and among people or groups with high risk for parenteral infection [54, 55]. Several studies have shown that occult HBV infection can be found in patients with chronic HCV infection [54, 56, 57]. Occult HBV infection is characterized by persistent HBV viremia in HBsAg-negative patients with or without markers of

previous infection (anti-HBc and/or anti-HBs) [58]. In spite of the limited knowledge about the mechanisms involved in the absence of circulating HBsAg protein, it has been demonstrated that there is a causal relationship between HCV infection and the absence of non-anti-HBc markers. This relationship is supported by the direct influence of HCV core protein on HBV, which reduces its replication capacity and HBsAg production dramatically and compromises its detection in the serum of co-infected individuals [59]. Consistent with this, Weber et al. [60], in a study involving 104 anti-HBc-alone-positive individuals from a population of high HCV seroprevalence (patients on maintenance dialysis, organ transplant patients, intravenous drug addicts, HIV-infected individuals and HCV-chronic individuals), demonstrated that the isolated anti-HBc reactivity could be due to HCV interference with HBsAg synthesis (65.4%), with consequent establishment of occult infection associated with a viral interference mechanism [61]. In addition, data from other studies have shown that in HBV/HCV coinfecting patients, there is low DNA polymerase activity, low levels of HBV DNA in the serum and low levels of HBsAg in the bloodstream, which suggest suppression of HBV replication [62].

Formation of HBsAg-anti-HBs immune complexes

The detection of anti-HBs and anti-HBe antibodies in patients with chronic infection (or acute—in the ‘window phase’) has been difficult due to the presence of excess surface antigen (HBsAg), composed of preS/S and S proteins, and of the hepatitis B e antigen (HBeAg—a secreted form of the nucleoprotein) [63]. The anti-HBs and anti-HBe antibodies produced would be consumed by their corresponding antigens, forming immune complexes that could explain the isolated positivity to anti-HBc [23, 64]. These immune complexes (ICs) cannot be detected by the standardized commercial assays, which usually detect anti-HBs and anti-HBe antibodies only after the corresponding antigens have been cleared from the serum, since dissociation of these complexes is necessary for their detection. Joller-Jemelka et al. [64], in a study aiming to identify whether the dissociation of immune complexes in patients’ serum with ‘anti-HBc alone’ could lead to detection of additional serological markers, showed that 31% of the sera examined had HBsAg in the form of immune complexes and detectable HBV-DNA in plasma. Thus, in the early phase of acute hepatitis B, HBV exists in both forms, free and immunoglobulin-bound, but subsequently there is a shift to the immunoglobulin-bound form due to seroconversion from HBsAg to anti-HBs [65]. In another study, a series of 200 HBeAg-positive serum samples from chronically infected hepatitis B patients were analyzed using

novel immunoassays designed to detect antibodies in the presence of excess circulating viral antigens. All patients were seronegative for antibodies directed to envelope antigens and to the secreted nucleoprotein antigen (HBeAg) when the standardized commercial assays were used. In contrast, virtually all chronically infected patients were found to have anti-HBe and anti-envelope antibodies in the form of immune complexes when sera were tested employing more sensitive immunoassays [63].

Low levels of anti-HBs despite immunity to HBV

This serological profile can be found in subjects who have lost their anti-HBs-producing capacity after a long period of infection—years, but most often decades—after resolution of infection, or who have never been able to produce it at a sufficient level for detection [66] despite evidence that T cells can produce a soluble factor that selectively suppresses the production of anti-HBs by B lymphocytes [67].

The occurrence of antibodies directed to the HBs antigen indicates viral clearance and the development of lifelong immunity in patients who have recovered from HBV infection. For HBs antigen vaccine recipients, the presence of HBs antibodies signifies protective immunity. However, little is known about the kinetics and regulation of this HBs-specific antibody response in the clinical course of HBV infection and in vaccine recipients [68]. Likewise, the reasons for decreased levels of antibodies after a long period of infection or following vaccination are not very clear. Several immunological mechanisms that could explain a deficient anti-HBs production have been discussed in detail by Milich and Leroux-Roels [69]. One of them is associated with low HBsAg immunogenicity at the T-cell level. Being a strictly T-cell-dependent immunogen, HBsAg could not be able to induce a strong and long-lasting immune response [69, 70]. However, some studies have demonstrated that T cell memory to HBsAg can be detected by lymphocyte proliferation many years after HBV vaccination, even in the majority of individuals with serum anti-HBs < 10UI/L. This means that protective antibodies could reappear rapidly or that effector cytotoxic T cells could rapidly eliminate virus-infected hepatocytes after exposure to HBV [71]. Bocher et al. [72] observed that patients with acute self-limiting HBV infection had high levels of HBs-specific B cells, despite undetectable anti-HBs serum antibodies. In addition, Bauer and Jilg [73] have shown significant numbers of HBsAg-specific memory T and B cells in vaccine recipients, suggesting that these memory T cells are able to trigger anti-HBs production by B cells once they are activated by revaccination [73]. These findings suggest that despite undetectable anti-

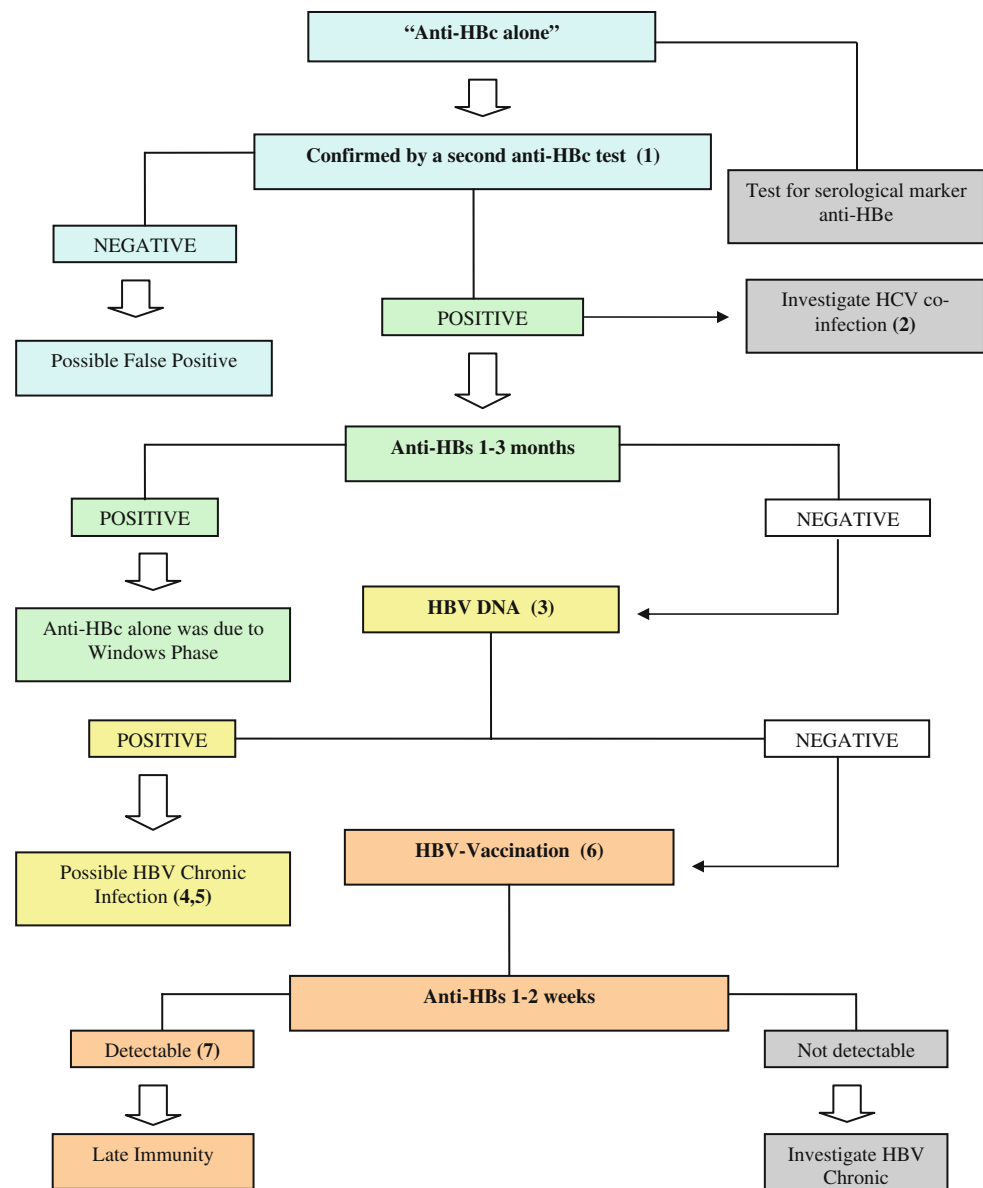
HBs serum antibodies, it is possible that the HBsAg-specific T cell immune response plays an important role in protecting against hepatitis B virus infection and that immunological memory persists beyond the time at which anti-HBs levels are no longer detectable and is able to protect against clinically relevant disease. In the case of HBV exposure, the immune memory rapidly leads to a vigorous anamnestic response, which often prevents acute infection and, most often, acute disease and prolonged viremia.

An additional explanation for the “anti-HBc alone” profile in the scenario of low titers of anti-HBs antibodies is that they can escape detection by the commercially available assays. The EIAs are not adjusted to detect such low levels, since any increase in sensitivity carries the risk of false positivity. Thus, results also depend on assay sensitivity, which is arbitrarily determined by different manufacturers. Greub et al. [74] demonstrated that samples from vaccinated individuals were anti-HBs negative when analyzed in a first assay, and subsequently showed positive results in, respectively, 25, 35 and 38% of the cases when a second, third and fourth assay were used. In this study, it was demonstrated that samples from individuals characterized as ‘non-responders’ and vaccinated individuals that had lost anti-HBs showed significantly higher absorbances than those observed in the group that had not been vaccinated (control). Such findings suggest that subliminal levels of anti-HBs may be present in ‘non-responders’ and vaccinated individuals who have lost anti-HBs, and despite the non-detection by commercially available assays, such subjects may be protected [74, 75]. Regarding susceptibility to HBV re-infection, it is probable that individuals with “anti-HBc alone” as a sign of late immunity do not differ from those who also carry anti-HBs antibodies. Despite the lack of such antibodies, individuals seem to be protected against reinfection by cellular immune mechanisms and immunological memory.

Significance of “anti-HBc alone” in different geographical contexts

The significance of ‘anti-HBc alone’ varies markedly according to the prevalence of HBV infection in a given region. Hepatitis B virus is highly endemic in certain developing regions, such as South East Asia, China, sub-Saharan Africa and the Amazon Basin, where at least 8% of the population are HBV chronic carriers. In these areas, 70–95% of the population shows past or present serological evidence of HBV infection [76]. Most infections occur during infancy or childhood. Hepatitis B is moderately endemic in part of Eastern and Southern Europe, Japan and part of South America. Between 10 and 60% of the

Fig. 1 (1) A different ELISA assay format is recommended. (2) It is also recommended that HCV coinfection should be investigated, especially in high-risk individuals such as haemodialysis patients, organ transplant recipients, intravenous drugs addicts, HIV-infected individuals, in whom HCV seroprevalence is also high. Concurrent HCV infection interferes with HBV replication, favoring the HBV chronic-carrier state with undetectable HBsAg. (3) An HBV DNA test with sensitivity of 10–100 genomes/ml should be employed. (4) Individuals should be tested for alanine aminotransferase activity (ALT). In the case of a clearly elevated ALT level, a biopsy seems appropriate to guide therapy. (5) Individuals with positive HBV DNA and normal ALT values should be assessed in yearly intervals. (6) This approach, named “Diagnostic HBV-Vaccine Booster”, attempts to stimulate immunological memory. (7) The presence of anti-HBs at high levels suggests a secondary response



population show evidence of infection, and 2–7% are chronic carriers [76, 77]. The endemicity of HBV is low in most developed areas, such as North America, Northern and Western Europe and Australia. In these regions, HBV infects 5–7% of the population, and only 0.5–2.0% of the population are chronic carriers [76, 77]. In areas with low endemicity, such as most parts of Europe and United States, “anti-HBc alone” is found in 10–20% of all individuals with HBV markers. In about 10% of these individuals, HBV DNA is detected by PCR [9]. Lower prevalence, 0.08 and 1.5%, has been reported in the UK and Germany, respectively, with the percentage of samples containing HBV DNA ranging between 0 and 7.7% [78]. In areas with a higher prevalence of HBV infection, such as Greece (15.8%), China (70%) and Ghana (83.6%), the

frequency of DNA-positive results in anti-HBc-alone patients tends to increase to 1.9, 2.7 and 12.7%, respectively [78, 79]. As in highly endemic countries, the majority of infections are contracted perinatally or in early childhood, a higher proportion of the infected adults have late chronic HBV with undetectable HBsAg [80], suggesting that the prevalence of ‘anti-HBc alone’ and the proportion of individuals with HBV DNA might be higher in areas with intermediate and high hepatitis B endemicity [9]. However, it should be noted that there has been a great disparity in reported rates among various studies carried out in these areas [81–86], and it has been difficult to elucidate such results for several reasons, such as differences in the epidemiology of the tested populations, sample selection criteria for PCR testing, and differences in

sensitivity and specificity of NAT methods, which promote a great disparity in HBV DNA results [87]. Additionally, the analysis of ‘anti-HBc alone’ prevalence among reports is often difficult because of the low specificity associated with anti-HBc screening and of the algorithms applied to identify true reactivity [78].

Suggested algorithm for investigation of the “anti-HBc alone” pattern

Once the anti-HBc-alone serologic profile is detected, false-positive reactivity should be ruled out by a second anti-HBc test. It is also suggested to test for anti-HBe antibodies. In one-third of the cases, anti-HBe antibodies are also present, representing a good confirming test for hepatitis B infection. Once the anti-HBc-alone profile is confirmed, anti-HBs should be tested after another 1–3 months. If positive, this suggests that the initial testing had been performed during the “window period”. If the anti-HBs test is negative, the presence of viral DNA should be investigated by highly sensitive amplification methods, such as PCR, and if viral DNA is detected, this points to a possible chronic HBV infection. If HBV DNA is negative, HBV vaccination is suggested to test for “late immunity”, which is characterized by the presence of serum anti-HBs antibodies within 1–2 weeks [88]. If these antibodies are not detected after vaccination, chronic hepatitis B infection should be suspected (Fig. 1).

The vaccination approach, named “Diagnostic HBV—Vaccine Booster” attempts to stimulate immunological memory. HBV-immune individuals with undetectable levels of anti-HBs antibodies will develop a strong secondary immune response after the first dose of hepatitis B vaccine. On the other hand, chronic low-level HBV carriers, when challenged by vaccination, are not able to produce anti-HBs antibodies after vaccine doses at 0, 1 and 6 months [15, 23, 89]. In subjects who develop a primary response to hepatitis B vaccine, the isolated anti-HBc positivity is presumably a false positive result [89].

Conclusion

Isolated reactivity to anti-HBc, is a frequent and common laboratory finding that is observed during serological blood donor screening and routine serological assays in the clinical laboratory. In the context of this serological profile, the detection of this marker can represent a false-positive result, but it can also be indicative of acute infection in the window phase. On the other hand, ‘anti-HBc alone’ can represent a resolved infection or even a chronic infection. Other possibilities are not easily distinguished by routine

serological testing, but they can have clinical implications and important consequences, particularly in chronic infection with HBV mutants when HBsAg cannot be detected. HBV vaccination has been an efficient strategy for the diagnosis and management of patients with isolated reactivity to anti-HBc, separating low-level anti-HBs individuals with resolved infection from those chronically infected with undetectable HBV DNA.

The adoption of highly sensitive genome-amplification techniques for detection of viral nucleic acid and sequencing has allowed the identification of HBV chronic carriers and HBV mutants that can escape diagnosis by currently employed immunoassay methods. It is expected that with the advent of the next generation of immunoassays and molecular techniques of higher sensitivity and specificity, new serological and molecular patterns will be identified. These profiles may be able to characterize different infection phases and even revise the current clinical concepts of HBV infection.

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