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Effect of alpha lipoic acid on the tardive dyskinesia and oxidative stress induced by haloperidol in rats

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Abstract Haloperidol (HAL) is a widely used neuroleptic drug for the treatment of acute and chronic psychosis. Tardive dyskinesia (TD) is a complex hyperkinetic syndrome consisting of choreiform and athetoid movements, which persists for months or years after withdrawal. Increased levels of thiobarbituric acid reactive products are found in the cerebrospinal fluid and plasma of patients treated with neuroleptics, especially those with movement disorders. Alpha lipoic acid (ALA), a natural metabolic antioxidant, is effective in both prevention and treatment of numerous types of neurological disorders. It is proposed to study the effect of ALA on TD induced by HAL and to correlate it with oxidative stress by studying total antioxidant status and lipid peroxidation (LP). HAL (1 mg/kg/i.p.) was used to induce vacuous chewing movements in rats. ALA was suspended in 0.2% carboxy methyl cellulose at a dose of 25, 50 and 100 mg/kg and was administered orally by oral gavage 1 h before HAL on 21st day of treatment. ALA supplementation significantly decreased HAL-induced TD at a dose of 100 mg/kg and catalepsy dose dependently. ALA improved TD and catalepsy by decreasing HAL-induced LP. ALA and its metabolite dihydro lipoic acid protect against HAL-induced TD and catalepsy by scavenging reactive oxygen species and reactive nitrogen species.

Keywords Tardive dyskinesia · VCMs · TAS · MDA · Antioxidants · Alpha lipoic acid · Lipid peroxidation

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Introduction

Haloperidol (HAL) is a widely used neuroleptic drug for the treatment of acute and chronic psychosis. The use of HAL is limited by its tendency to produce a range of typical antipsychotic extrapyramidal movement disorders such as tardive dyskinesia (TD), akathisia, dystonia and parkinsonism. TD is a complex hyperkinetic syndrome consisting of choreiform, athetoid or rhythmic abnormal involuntary movements. The face, mouth and tongue are most frequently involved (orofacial type), but a variety of less frequent motor abnormalities of the upper, lower limbs and trunk also occur (Kane and Smith [1982](#page-6-0)). The syndrome has usually a delayed onset and the intensity of the disturbance fluctuates over time. The most serious aspect of TD is that it may persist for months or years after drug withdrawal and in some patients it is irreversible (Jeste et al. [1979](#page-6-0); Casey [1985a](#page-5-0), [b;](#page-5-0) Glazer et al. [1990\)](#page-6-0) and the syndrome is a major clinical and ethical issue (APA 1992).

Oxidative stress (OST) resulting from increased production of reactive oxygen species (ROS) and a decrease in antioxidant defense damages biological macromolecules, disrupts normal 5HT-1A metabolism and physiology (Sies [1985](#page-7-0)) and is proposed as a pathogenic mechanism in TD (Cadet et al. [1986;](#page-5-0) Cadet and Kahler [1994\)](#page-5-0).

HAL is metabolized by an oxidase, generates large quantities of oxyradicals and a potent toxic pyridinium-like metabolite (Subramaniam et al. [1990](#page-7-0)) and induces OST (Behl et al. [1995;](#page-5-0) Sagara [1998;](#page-7-0) Post et al. [1998\)](#page-7-0). Blockade of dopamine D_2 receptors by neuroleptics is associated with increased dopamine turnover particularly in catecholamine-rich regions such as basal ganglia (Creese et al. [1976](#page-5-0)). This supports the hypothesis that typical neuroleptics increase the oxyradicals as a result of increased catecholamine and drug metabolism, and contribute to

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oxyradical injury in schizophrenic patients (Cadet and Lohr [1989;](#page-5-0) See [1991\)](#page-7-0).

Increased levels of thiobarbituric acid (TBA) reactive products are found in the cerebrospinal fluid and plasma of patients treated with neuroleptics (Pall et al. [1987](#page-6-0)), especially those with movement disorders (Lohr et al. [1990](#page-6-0); Peet et al. [1993\)](#page-7-0). Amelioration of TD after treatment with Pro-Leu-glycinamide (Sharma et al. [2003\)](#page-7-0) and vitamin E, a lipid soluble antioxidant, confirms (Lohr et al. [1987](#page-6-0); Elkashef et al. [1990;](#page-6-0) Scapicchio et al. [1991](#page-7-0); Egan et al. [1992;](#page-5-0) Adler et al. [1993a](#page-5-0), [b,](#page-5-0) [1998\)](#page-5-0) the involvement of OST in HAL-induced TD.

Alpha lipoic acid (ALA) also known as thioctic acid is a naturally occurring compound synthesized by plants and animals. In humans, it is readily synthesized in the body and is well absorbed from the diet through the stomach and intestine. ALA is a medium length (eight carbon atoms) fatty acid containing two sulfur atoms that are oxidized or reduced. This feature allows ALA to function as a potent antioxidant and cofactor for several important enzymes (Cadenas and Packer [1996\)](#page-5-0). ALA is effective in both prevention and treatment of numerous types of neurological disorders, due to its ability to cross the BBB and accumulates in brain tissue and is an ideal therapeutic antioxidant for treatment of brain and neurodegenerative disorders involving free radical processes (Packer et al. [1995](#page-6-0)). The ability of ALA or its reduced form dihydro lipoic acid (DHLA) to scavenge ROS and chelate transition metals in Fenton reaction restricts the molecular damage by hydroxyl radicals and reduces cellular need for glutathione peroxidase (GPx) and catalase (CAT). ALA or DHLA prevents oxygen-induced DNA damage, exhibits chelating activity, reduces lipid peroxidation (LP) and prevents glycation of proteins (Packer et al. [2001;](#page-6-0) Biewenga et al. [1997;](#page-5-0) Maritim et al. [2003;](#page-6-0) Obrosova et al. [2003a](#page-6-0), [b;](#page-6-0) Maddux et al. [2001](#page-6-0)). The antioxidant responsiveness mediated by ALA has biological significance in eliminating reactive free radicals and restoring antioxidant enzymes that otherwise affect the normal functioning, especially brain.

The most widely used animal model for TD is the rat model in which vacuous chewing movements (VCMs) are induced by chronic neuroleptic treatment (Andreassen and Jorgensen [1995;](#page-5-0) Clow et al. [1979](#page-5-0); Iversen et al. [1980](#page-6-0); Gunne et al. [1982,](#page-6-0) [1986;](#page-6-0) Waddington [1990;](#page-7-0) Egan et al. [1995;](#page-5-0) Hyde et al. [1995](#page-6-0); Kaneda et al. [1992](#page-6-0); Shirakawa and Tamminga [1994;](#page-7-0) Andreassen and Jorgensen [1995](#page-5-0); Andreassen et al. [1996](#page-5-0)).

The present investigation is designed to study the effect of ALA on the selected extrapyramidal movement disorders (parkinsonism, TD) produced by HAL in rats. A dose of 25, 50, 100 mg/kg of ALA is selected, as this dose was effective in increasing antioxidant status of brain and peripheral organs in various studies (Muthuswamy et al. [2006](#page-6-0); Sethumadhavan and Chinnakannu [2006;](#page-7-0) Sahin et al. [2006](#page-7-0); Arivazhagan et al. [2006](#page-5-0)).

Materials and methods

Animals

Male albino Wistar rats weighing 150–200 g bred in Central Animal House facility of our university were used. Male rats were chosen to avoid fluctuations due to estrous cycle-induced antioxidant effects (Sugioka et al. [1987](#page-7-0)). The rats were housed in polypropylene cages (five per cage) under standard laboratory conditions with 12-h light/ dark cycle and were fed with standard laboratory chow (Hindustan Lever Ltd, Mumbai) and water ad libitum. Animals were acclimatized to the laboratory conditions prior to experimentation and all the experiments were carried out between 0900 and 1200 hours. The study protocol was approved by the Institutional Animal Ethical Committee and conducted according to the Indian National Science Academy guidelines for the use and care of experimental animals.

Induction of VCMs and catalepsy

Haloperidol (1 mg/kg/i.p.) was administered chronically to rats for a period of 21 days to induce VCMs and catalepsy (Sasaki et al. [1995](#page-7-0); Naidu and Kulkarni [2001a,](#page-6-0) [b](#page-6-0); Naidu et al. [2003a,](#page-6-0) [b;](#page-6-0) Naidu and Kulkarni [2004](#page-6-0); Thaakur and Jyothi [2007](#page-7-0)). All the behavioral assessments were carried out 24 h after the last dose of HAL.

Assessment of behavioral parameters

Behavioral assessment of VCMs

On the test day, rats were placed individually in a small $(30 \times 20 \times 30 \text{ cm})$ Plexiglass cage for the assessment of VCMs. The floor and back wall of cage consisted of mirrors so as to permit the observation of VCMs (frequencies) when the animal was facing back and down. VCMs were counted with the help of a hand-operated counter by trained observers. VCMs are referred to as single mouth openings in the vertical plane not directed toward a physical material and were counted for a period of 5 min and in all experiments scorer was unaware of the treatment given to the animal and counting was stopped whenever the rat began grooming and restarted when grooming stopped (Sasaki et al. [1995;](#page-7-0) Naidu and Kulkarni [2001a,](#page-6-0) [b](#page-6-0), [2004](#page-6-0); Naidu et al. [2003a](#page-6-0), [b](#page-6-0); Thaakur and Jyothi [2007](#page-7-0)).

Catalepsy

The catalepsy was measured by standard procedure using 3 and 9 cm wooden blocks (Goyal et al. [2006](#page-6-0)). The following scores were assigned to the rats: rats move normally when placed on table, score 0; rats move normally when touched/pushed, score 0.5; front paws of rat were placed on 3 cm block and if it fails to correct the posture in 10 s, a score of 0.5 was assigned to each paw (total 1). If the rat fails to correct the posture within 10 s, when placed on 9 cm block, score for each paw was 1 (total 2); thus for a single rat maximum score assigned was 3.5.

Locomotor activity

Locomotor activity or spontaneous motor activity (SMA) was assessed using Photoactometer (Inco, Ambala, India). The total count over 30-min period was recorded for each animal. Increase in count was regarded as central nervous system stimulant activity, whereas decrease in count was regarded as central nervous system depressant activity.

Biochemical measurements

On 21st day, blood samples were collected from retroorbital plexus under light ether anesthesia for the estimation of total antioxidant status (TAS) and LP.

Estimation of TAS

0.1 ml of serum was deprotonated by the addition of 1 ml of methanol, vortexed for 30 s and centrifuged at 3,000 rpm for 30 min to separate the proteins. To the clear supernatant, 1.5 ml of methanol and 0.5 ml of diphenyl picryl hydrazine solution were added, mixed thoroughly and absorbance was read at 517 nm against blank, which was prepared in an identical way but without the addition of serum (Blios [1958](#page-5-0)).

Estimation of LP

0.1 ml of plasma was treated with 2 ml (0.37%) of TBA, 0.25 N hydrochloric acid and 15% trichloro acetic acid (1:1:1 ratio) and placed in water bath for 15 min, cooled and centrifuged and then clear supernatant was measured at 535 nm against reference blank (Nichans and Samuelson [1968\)](#page-6-0).

Drugs and study protocol

Haloperidol was dissolved in 0.9% saline and injected intraperitoneally in a constant volume of 0.5 ml/kg body weight. Animals were divided into six groups:

- Group I: orally received 0.2% carboxy methyl cellulose in a constant volume of 0.5 ml daily by oral gavage.
- Group II: received HAL 1 mg/kg/i.p. in a constant volume of 0.5 ml/kg body weight for 21 days (0.9% saline).
- Group III: received ALA 100 mg/kg/p.o. suspended in 0.2% CMC and was administered orally by oral gavage once (1 h before 0.9% saline) on the last day of treatment.
- Group IV: received ALA 25 mg/kg/p.o. suspended in 0.2% CMC and was administered orally by oral gavage once (1 h before HAL 1 mg/kg/i.p. in 0.9% saline) on the last day of treatment.
- Group V: received ALA 50 mg/kg/p.o. suspended in 0.2% CMC and was administered orally by oral gavage once (1 h before HAL 1 mg/kg/i.p. in 0.9% saline) on the last day of treatment.
- Group VI: received ALA 100 mg/kg/p.o. suspended in 0.2% CMC and was administered orally by oral gavage once (1 h before HAL 1 mg/kg/i.p. in 0.9% saline) on the last day of treatment.

Statistical analysis

All values were expressed as mean \pm SEM. The data were analyzed using Instat by one-way analysis of variance followed by Dunnett's t test. $P < 0.05$ was considered significant.

Results

Figure 1 shows the effect of ALA on the HAL-induced VCMs. Significant increase in the number of VCMs was observed on the 21st day of treatment in HAL, ALA 25, 50 and 100 mg/kg treated groups when compared with 1st day of treatment. ALA at a dose of 100 mg/kg significantly

Fig. 1 Effect of alpha lipoic acid on the haloperidol-induced vacuous chewing movements. Values are expressed as mean ± SEM of five animals. *** $P\leq0.001$ vs. 1st day of treatment. $^{+++}P\leq0.001$ vs. haloperidol group

decreased the VCMs when compared to HAL treatment group.

Figure 2 shows the effect of ALA on HAL-induced catalepsy. ALA significantly decreased the catalepsy at a dose of 25 mg/kg as compared to control group. When compared to HAL group, ALA significantly decreased the catalepsy at a dose of 25, 50 and 100 mg/kg, at a dose of 100 mg/kg it reversed the catalepsy induced by HAL.

Figure 3 shows the effect of ALA on HAL-induced alterations in locomotor activity. HAL significantly decreased the locomotor activity ($P \lt 0.001$) when compared to control group. ALA significantly reversed the HAL-induced decrease in locomotor activity as compared to HAL group at a dose of 25, 50 and 100 mg/kg $(P<0.001)$.

Figure 4 shows the effect of ALA on HAL-induced changes in TAS. Significant decrease in the total antioxidant levels was observed in HAL, ALA 25, 50, 100 mg/kg treated groups as compared to control group ($P < 0.001$).

Figure 5 shows the effect of ALA on HAL-induced LP. Significant increase in the lipid peroxide levels was observed in HAL, ALA 25, 50 mg/kg treated groups when

Fig. 2 Effect of alpha lipoic acid on of haloperidol-induced catalepsy. Values are expressed as mean \pm SEM of five animals. **P < 0.01 vs. control group. ***P < 0.001 vs. control group. $***P<0.001$ vs. haloperidol group

Fig. 3 Effect of alpha lipoic acid on haloperidol-induced alterations in locomotor activity. Values are expressed as mean \pm SEM of five animals. *** $P < 0.001$ vs. control group. $^{+++}P < 0.001$ vs. haloperidol group

compared to control group $(P\lt 0.01)$, whereas ALA 100 mg/kg reversed the HAL-induced LP.

Discussion

In present study, HAL-treated animals showed increased frequencies of VCMs and catalepsy. Antipsychotic drugs produced two main kinds of motor disturbances in humans, TD and catalepsy, collectively termed as extrapyramidal side effects, which result directly or indirectly from D_2 receptor blockade. These effects constitute one of the main disadvantages for the therapeutic use of typical neuroleptics such as HAL.

Numerous reports indicate that an excessive production of free radicals (OST) is associated with chronic neuroleptic use and contributes to the onset of TD and other movement disorders (Cadet et al. [1986](#page-5-0); Pall et al. [1987](#page-6-0);

Fig. 4 Effect of alpha lipoic acid on haloperidol-induced changes in total antioxidant status. Values are expressed as mean \pm SEM of five animals. *** $P < 0.001$ vs. control group

 CON HAL ALA 100 HAL + ALA 25

Fig. 5 Effect of alpha lipoic acid on the haloperidol-induced alterations in lipid peroxidation. Values are expressed as mean \pm -SEM of five animals. $*P < 0.01$ vs. control group

Lohr et al. [1987](#page-6-0), [1990](#page-6-0); Peet et al. [1993](#page-7-0); Elkashef et al. [1990;](#page-6-0) Scapicchio et al. [1991](#page-7-0); Egan et al. [1992](#page-5-0); Adler et al. [1993a](#page-5-0), [b](#page-5-0), [1998\)](#page-5-0). OST is the shift in balance in cellular oxidation–reduction reactions in favor of oxidation, leading to cellular damage and is indicated by oxidized products of lipids, proteins and nucleic acids (Halliwell et al. [1994\)](#page-6-0).

Neuroleptics act by blocking dopamine receptors (Creese et al. [1976\)](#page-5-0), which in turn increase number of D_2 receptor sites and increase catecholamine levels, leading to excess production of free radicals and as a consequence, hydrogen peroxide resulting in OST (Cohen and Spina [1989;](#page-5-0) Elkashef and Wyatt [1999](#page-6-0)), especially in catecholamine-rich areas such as the basal ganglia. HAL produces ROS in rat primary cortical neurons and the mouse hippocampal cell line HT-2 (Sagara [1998](#page-7-0)), by decreasing the genetic expression of superoxide dismutase (SOD), GPx and CAT and thus decreases the enzymatic activities and protein content of SOD, GPx, CAT (Parikh et al. [2003](#page-7-0); Naidu et al. [2003a](#page-6-0), [b;](#page-6-0) Elkashef and Wyatt [1999](#page-6-0); Shivakumar and Ravindranath [1993;](#page-7-0) Thaakur and Jyothi [2007;](#page-7-0) Von Voigtlander et al. [1990\)](#page-7-0). Among antioxidant enzymes, SOD dismutases superoxide radicals to form hydrogen peroxide, which in turn is decomposed to water and oxygen by GPx, CAT, there by preventing the formation of hydroxyl radicals (Yao et al. [1998](#page-7-0)), these enzymes act co-operatively at different sites in the metabolic pathway of free radicals. Chronic HAL treatment increases LP (Parikh et al. [2003;](#page-7-0) Thaakur and Jyothi [2007\)](#page-7-0) and also protein and nucleic acid peroxidation (Esterbauer et al. [1991\)](#page-6-0), which are mutagenic and alters signal transduction and gene expression (Burcham [1998](#page-5-0); Keller and Mattson [1998\)](#page-6-0); and cytotoxic (Ravindranath and Reed [1990\)](#page-7-0) neuronal loss in the striatum is reported in animals treated chronically with neuroleptics (Nielson and Lyon [1978;](#page-6-0) Skoblenick et al. [2006](#page-7-0); Ukai et al. [2004\)](#page-7-0).

Surprisingly, TAS was not improved at the given doses of ALA, in recent years there is increasing interest in assessing the total antioxidant capacity because of the difficulty in measuring each antioxidant component separately. Acute dose of ALA used in this study might be sufficient to scavenge-free radicals in specific brain areas such as striatum and protect them from OST and TD but might be insufficient to show systemic change in TAS of blood as observed in this study. However, importance of blood in this research area is increasing since it is rich in antioxidants as well as it reflects changes in the antioxidant activity in other less accessible tissues.

In the present study, lipid peroxides were significantly increased in HAL, ALA 25, 50 mg/kg treated groups, but ALA at a dose of 100 mg/kg significantly reversed the HAL-induced LP. LP is one of the well-established indices of cellular peroxidative membrane injury associated with increased OST and reduced membrane fatty acids (Mahadik et al. [1999\)](#page-6-0). Living organisms developed complex antioxidant systems to reduce oxidative damage that include enzymes such as SOD, CAT, GPx, macromolecules such as albumin, ceruloplasmin, ferritin and an array of small molecules including ascorbic acid, alpha-tocopherol, carotene, uric acid, ubiquinol-10, methionine, reduced glutathione (Esterbaur et al. [1990](#page-6-0); Pascand [1993](#page-7-0)). ALA is water insoluble and its reduced metabolite, DHLA, forms a redox couple. Free ALA is rapidly taken up by cells and reduces to dithiol (Freeman and Capro [1982\)](#page-6-0); DHLA contributes to the antioxidant activity of ALA in vivo and its β oxidation products, bis nor and tetra nor lipoic acid, also contribute to the antioxidant activity (Biewenga et al. [1997\)](#page-5-0). DHLA prevents oxidative damage by interacting with ROS and reactive nitrogen species (RNS) (Packer et al. [2001\)](#page-6-0). ALA scavenges hydroxyl radicals, singlet oxygen and hypochlorous acid, and regenerates other antioxidants such as glutathione, vitamin C, ubiquinol (coenzyme Q_{10}) and indirectly, vitamin E (Cadenas and Packer [1996](#page-5-0); Roy and Packer [1998\)](#page-7-0). In addition, DHLA is capable of reducing the oxidized forms of vitamin C, glutathione and coenzyme Q10, which regenerates oxidized alpha-tocopherol (vitamin E), forming an antioxidant network (Kramer and Packer [2001](#page-6-0); Scholich et al. [1989\)](#page-7-0). Certain free metal ions such as iron and copper induce oxidative damage by catalyzing reactions that generate highly reactive free radicals. Both ALA and DHLA chelate metal or bind ions in a way that it prevents them from generating free radicals (Zhang and Frei [2001](#page-7-0)).

HAL-induced TD and OST are effectively prevented by dietary supplementation with antioxidants and essential fatty acids (Mahadik and Gowda [1996;](#page-6-0) Mahadik and Scheffer [1996;](#page-6-0) Reddy and Yao [1996;](#page-7-0) Mahadik et al. [2001](#page-6-0)). Vitamin E (Adler et al. [1993a](#page-5-0), [b](#page-5-0), [1998;](#page-5-0) Gupta et al. [1999](#page-6-0); Soares and McGrath [2000](#page-7-0); Elkashef and Wyatt [1999](#page-6-0); Barak et al. [1998](#page-5-0)), melatonin (Shamir et al. [2000,](#page-7-0) [2001](#page-7-0); Naidu et al. [2003a](#page-6-0), [b;](#page-6-0) Faria et al. [2005\)](#page-6-0) and vitamin C are reported to be effective against HAL-induced oral dyskinesia. HAL-induced orofacial dyskinesia is attenuated by inhibiting nitric oxide synthase (Naidu et al. [2003a,](#page-6-0) [b](#page-6-0); Chen et al. [2001;](#page-5-0) Raso et al. [2001\)](#page-7-0) and monoamino oxidase B activity (Lee et al. [2001\)](#page-6-0); nitric oxide- and mono amine-induced free radicals are involved in the pathophysiology of orofacial dyskinesia (Naidu and Kulkarni [2001a\)](#page-6-0). Our findings are in agreement with previous investigations which reported decrease in frequencies of HAL-induced TD by various antioxidants such as vitamin E, melatonin, carvedilol, selegiline, quercetin and faria ebselen.

Neuroleptic-induced catalepsy is used as an animal model for the study of extrapyramidal side effects such as parkinsonism, associated with antipsychotic use in humans

(Naidu et al. [2003a](#page-6-0), [b;](#page-6-0) Pires et al. [2005](#page-7-0)). Catalepsy is linked to the blockade of post-synaptic striatal dopamine D_1 and D_2 receptors (Sanberg et al. [1988](#page-7-0)). Surprisingly, our results suggest that free radicals are also involved in the incidence of catalepsy; chronic blockade of $D₂$ inhibitory DA receptors localized on glutaminergic terminals in the striatum leads to the persistent enhanced release of glutamate that kills the striatal output neurons (Naidu and Kulkarni [2001a](#page-6-0), [b\)](#page-6-0).

Casey (1985a, b) proposed that chronic blockade of inhibitory dopamine receptors by HAL enhances catecholamine and/or glutamate release in the striatum, leading to chronic excitotoxic neurodegeneration. The competitive NMDA receptor antagonist SDZ 250-581, LY235959 (Chartoff et al. 1999) and MK-801, a non-competitive NMDA receptor antagonist reduced the cataleptogenic effect of HAL (Verma and Kulkarni [1992;](#page-7-0) Chartoff et al. 1999). Raised Ca^{2+} levels increase glutamate release, thereby activation of nitric oxide synthase, while low concentrations of NO are neuroprotective, high concentrations in the presence of ROS generate peroxynitrite and hydroxyl free radicals, which damages many important biomolecules including membrane lipids, proteins and DNA causing neurodegeneration.

Since antipsychotics are the drugs of choice for the treatment of psychiatric disorders, understanding their effects on OST and oxidative cell injury may be very important. In conclusion, ALA improves TD and catalepsy by scavenging hydroxyl radicals, singlet oxygen and hypochlorous acid, and regenerating other antioxidants such as glutathione, vitamin C, ubiquinol (coenzyme Q 10) and indirectly, vitamin E, its metabolite DHLA reduces oxidized forms of vitamin C, glutathione and coenzyme Q10, which regenerates oxidized alpha-tocopherol (vitamin E), forming an antioxidant network scavenges ROS and RNS.

Further research on the effect of chronic administration of ALA in the improvement of TD, catalepsy and its correlation with the indicators of OST in various brain regions may be needed to draw better conclusions.

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