

Conditioning training and retrieval increase phospholipase A₂ activity in the cerebral cortex of rats

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Abstract In rats, phospholipase A₂ (PLA₂) activity was found to be increased in the hippocampus immediately after training and retrieval of a contextual fear conditioning paradigm (step-down inhibitory avoidance [IA] task). In the present study we investigated whether PLA₂ is also activated in the cerebral cortex of rats in association with contextual fear learning and retrieval. We observed that IA training induces a rapid (immediately after training) and long-lasting (3 h after training) activation of PLA₂ in both frontal and parietal cortices. However, immediately after retrieval (measured 24 h after training), PLA₂ activity was increased just in the parietal cortex. These findings suggest that PLA₂ activity is differentially required in the frontal and parietal cortices for the mechanisms of contextual learning and retrieval. Because reduced brain PLA₂ activity has been reported in Alzheimer disease, our results suggest that stimulation of PLA₂ activity may offer new treatment strategies for this disease.

Keywords Phospholipase A₂ · Rat · Frontal cortex · Parietal cortex · Learning · Retrieval

Introduction

Phospholipase A₂ (PLA₂) is a family of hydrolytic enzymes that catalyze the cleavage of fatty acids from the

sn-2 position of membrane glycerophospholipids to generate lysophospholipids and free fatty acids (Dennis 1994, 1997). PLA₂-catalyzed hydrolysis of membrane phosphatidylcholine forms lysophosphatidylcholine and free arachidonic acid (AA), which are important mediators in signal transduction (Farooqui et al. 1997). The PLA₂ family is classified into three main groups: secretory (extracellular) Ca²⁺-dependent PLA₂ (sPLA₂), cytosolic Ca²⁺-dependent PLA₂ (cPLA₂), and intracellular Ca²⁺-independent PLA₂ (iPLA₂) (Dennis 1994). The mRNA and/or activity of the three groups have been detected both in human (Chen et al. 1994; Larsson Forsell et al. 1999; Pickard et al. 1999; Gelb et al. 2000; Mancuso et al. 2000; Suzuki et al. 2000) and rat brains (Owada et al. 1994; Molloy et al. 1998; Kishimoto et al. 1999).

Previous studies from our laboratory showed reduced PLA₂ activity in *postmortem* parietal and frontal cortices of Alzheimer disease (AD) patients (Gattaz et al. 1995, 1996). These findings were supported by Ross et al. (1998), who reported reduced cPLA₂ and iPLA₂ activities in *postmortem* parietal and temporal cortices of AD patients, as well as decreased cPLA₂ activity in the hippocampus. Moreover, decreased iPLA₂ activity was found in *postmortem* prefrontal cortex of frontal-variant AD patients (Talbot et al. 2000).

Several studies in laboratory animals have shown that PLA₂ blockade impairs learning and memory, simulating deficits that are found since the earliest phases of AD and represent the most predominant cognitive changes in this disease. For instance, intracerebral infusion of non-selective PLA₂ inhibitors in chicks impaired learning of a passive avoidance task (Holscher and Rose 1994). Additionally, intraperitoneal injection of a non-selective PLA₂ inhibitor in rats impaired spatial learning tested in the Morris water maze (Holscher et al. 1995). Furthermore,

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intracerebroventricular infusion in mice of a non-selective PLA₂ inhibitor or a dual cPLA₂ and iPLA₂ inhibitor impaired memory formation of a step-through inhibitory avoidance task (in which a context, tone, and foot shock are presented together in an associative fashion) (Sato et al. 2007), and a selective iPLA₂ inhibitor impaired spatial learning tested in the Y-maze (Fujita et al. 2000). Recent studies from our group showed that infusion of dual cPLA₂ and iPLA₂ inhibitors or a selective iPLA₂ inhibitor into rat hippocampal CA1 field impaired acquisition of short- and long-term memory (Schaeffer and Gattaz 2005), and retrieval of long-term memory (Schaeffer and Gattaz 2007) of a contextual fear task (step-down inhibitory avoidance [IA], in which fear conditioning is induced by a single exposure to a context followed by an electric foot shock).

Memory training has been clinically performed and reported to be effective in improving memory function in elderly subjects with mild cognitive impairment (Rapp et al. 2002; Belleville et al. 2006; Wenisch et al. 2007) and early-stage AD (Clare et al. 2002; Abrisqueta-Gomez et al. 2004; Avila et al. 2004). Animal research has elucidated some possible brain biochemical mechanisms related to experience-dependent stimulation, and PLA₂ activation seems to be highly implicated here. For example, passive avoidance training was followed by enhanced concentration of AA (Clements and Rose 1996) and prostaglandins (cyclooxygenase products of AA metabolism) (Holscher 1995) in chick brains. Recent studies from our group showed that training of rats in the IA task increased the activity of endogenous PLA₂ in the hippocampal CA1 field (Schaeffer and Gattaz 2005). Additionally, our studies showed that re-exposure of rats to context after training (contextual memory retrieval) also stimulated PLA₂ activity in the CA1 field (Schaeffer and Gattaz 2007). In the present study we extended our previous findings in the hippocampus, by investigating the effects of contextual learning and retrieval on PLA₂ activity in the cerebral cortex of rats.

Materials and methods

One hundred and six male Wistar rats of 270–330 g (Central Animal Laboratory House, Federal University of São Paulo, Brazil) were used in the present study. All the procedures described were approved by the institutional animal ethics committee.

Inhibitory avoidance task

Step-down IA task was carried out as previously described (Vianna et al. 2000). The animals were placed on an 8.0 cm wide, 5.0 cm high platform at the left of a 50 cm

wide, 25 cm deep, 25 cm high IA box (Albarsch, Brazil), whose floor was an electrified grid made of a series of parallel 1.0 mm caliber stainless steel bars spaced 1.0 cm apart. In training sessions, immediately after stepping down from the platform, placing the four paws on the grid, the rats received a 0.4 mA, 4.0 s scrambled foot shock. Latencies to step down were measured. Rats were tested for retrieval 24 h after training. In test sessions, the rats were allowed to stay on the platform up to a ceiling of 180 s, and no foot shock was given. Latencies to step down were measured. Test session step-down latency was taken as a measure of retention.

Three different experiments were carried out.

1. In the first experiment, 31 rats were divided into (a) trained animals: trained in the IA as described above and killed by decapitation immediately after training session; (b) naïve controls: killed by decapitation immediately after withdrawal from their home cages; and (c) shocked controls: placed directly over the electrified grid, given the foot shock, and immediately killed by decapitation. All trained animals stepped down the platform in the training session, showing a mean step-down latency of 9 ± 6 s.
2. In the second experiment, 38 rats were divided into (a) trained animals: trained in the IA and killed by decapitation 3 h after training session; (b) naïve controls: killed by decapitation immediately after withdrawal from their home cages; and (c) shocked controls: placed directly over the electrified grid, given the foot shock, and killed by decapitation 3 h later. All trained animals stepped down the platform in the training session, showing a mean step-down latency of 6 ± 4 s.
3. In the third experiment, 37 rats were divided into (a) trained/tested animals: trained in the IA, tested for retrieval 24 h later, and killed by decapitation immediately after retrieval test session; (b) naïve controls: killed by decapitation immediately after withdrawal from their home cages; and (c) trained controls: trained in the IA and killed by decapitation 24 h later. All trained animals stepped down the platform in the training session. Animals in the trained group showed a mean step-down latency of 8 ± 5 s, and animals in the trained/tested group showed a mean step-down latency of 7 ± 5 s. Animals in the trained/tested group were also tested for retrieval 24 h after training, showing a mean step-down latency of 112 ± 56 s.

Determination of PLA₂ activity

For PLA₂ activity determination, the rat brains were rapidly withdrawn and the frontal association cortex and

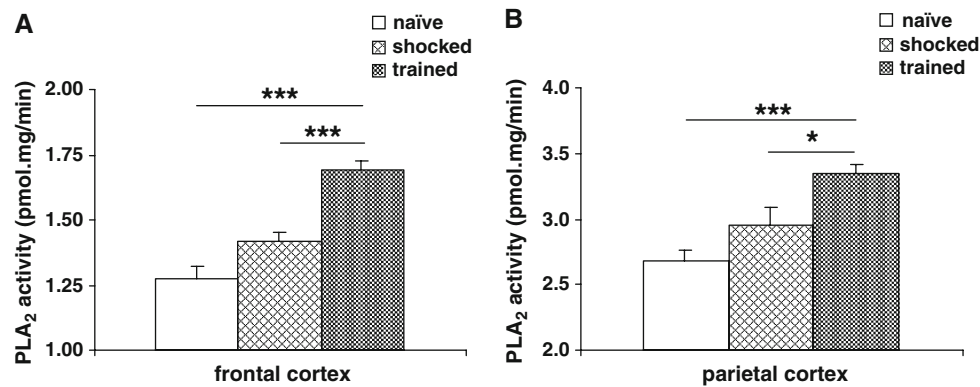


Fig. 1 PLA₂ activity measured immediately after training. PLA₂ activity (pmol mg protein min⁻¹) is given as mean (±SEM). **a** In the frontal cortex, PLA₂ activity was increased in trained animals ($n = 9$) by 32% as compared to naïve controls ($n = 8$), and by 20% as compared to shocked controls ($n = 10$). **b** In the parietal cortex, PLA₂ activity was increased in trained animals ($n = 10$) by 25% as

compared to naïve controls ($n = 11$), and by 13% as compared to shocked controls ($n = 9$). Shocked and naïve controls had similar values of PLA₂ activity in both studies. * $P < 0.05$, *** $P < 0.001$. P values were calculated using Bonferroni's test after ANOVA: Frontal cortex: $F_{(2,24)} = 26.57$, $P < 0.001$; Parietal cortex: $F_{(2,27)} = 14.03$, $P < 0.001$

parietal association cortex were bilaterally dissected according to visual anatomical landmarks and the Atlas of Paxinos and Watson (1998), and immediately stored at -70°C until use. The brain tissue was homogenized in 20 volume of 5 mM Tris-HCl buffer (pH 7.4, 4°C) and stored at -70°C . Prior to PLA₂ assay, total protein levels were determined for each aliquot by the Bio-Rad DC Protein Assay (Bio-Rad, Hercules, CA, USA) modified from the Lowry assay (Lowry et al. 1951). PLA₂ activity was determined by a radioenzymatic assay, as previously described (Schaeffer and Gattaz 2005). Briefly, as enzyme substrate we used L- α -1-palmitoyl-2-arachidonyl-phosphatidylcholine labelled with [$1-^{14}\text{C}$] in the arachidonyl tail at the *sn*-2 position (arachidonyl- $1-^{14}\text{C}$ -PC) (PerkinElmer, Boston, MA, USA). We used optimal assay conditions for measuring cPLA₂ plus iPLA₂ activity in rat brain homogenates, as previously determined by our group. Hence, the assay samples (500 μl) contained 50 mM Tris-HCl (pH 8.5), 1 μM CaCl₂, 300 μg of protein from homogenates, and 0.06 μCi arachidonyl- $1-^{14}\text{C}$ -PC. After an incubation time of 30 min at 37°C , the radioactivity of the liberated [$1-^{14}\text{C}$]arachidonic acid was measured in a liquid scintillation counter (Tri-Carb 2100TR; Packard, Meriden, CT, USA) and used for calculating the PLA₂ activity, which is expressed in pmol mg protein min⁻¹. All determinations of PLA₂ activity were performed in triplicate.

Statistical analysis

One-way analysis of variance (ANOVA) was used to compare the values among groups in each time interval. Post hoc test consisted of the Bonferroni's multiple comparison test. Pearson correlation coefficient was calculated to determine the degree of association between PLA₂

activity and scores on the memory retrieval test of individual animals within the trained/tested group in the third experiment. Two-tailed probabilities < 0.05 were considered significant.

Results

PLA₂ activity measured immediately after training

Frontal association cortex PLA₂ activity was significantly increased in trained animals ($n = 9$) by 32% as compared to naïve controls ($n = 8$), and by 20% as compared to shocked controls ($n = 10$; $P < 0.001$). Shocked controls had similar values of PLA₂ activity as naïve controls ($P > 0.05$). P values were calculated using Bonferroni's test after ANOVA, $F_{(2,24)} = 26.57$, $P < 0.001$ (Fig. 1a).

Parietal association cortex PLA₂ activity was significantly increased in trained animals ($n = 10$) by 25% as compared to naïve controls ($n = 11$; $P < 0.001$), and by 13% as compared to shocked controls ($n = 9$; $P < 0.05$). Shocked controls had similar values of PLA₂ activity as naïve controls ($P > 0.05$). P values were calculated using Bonferroni's test after ANOVA, $F_{(2,27)} = 14.03$, $P < 0.001$ (Fig. 1b).

PLA₂ activity measured 3 h after training

Frontal association cortex PLA₂ activity was significantly increased in trained animals ($n = 11$) by 18% as compared to naïve controls ($n = 10$; $P < 0.01$), and by 13% as compared to shocked controls ($n = 11$; $P < 0.05$).

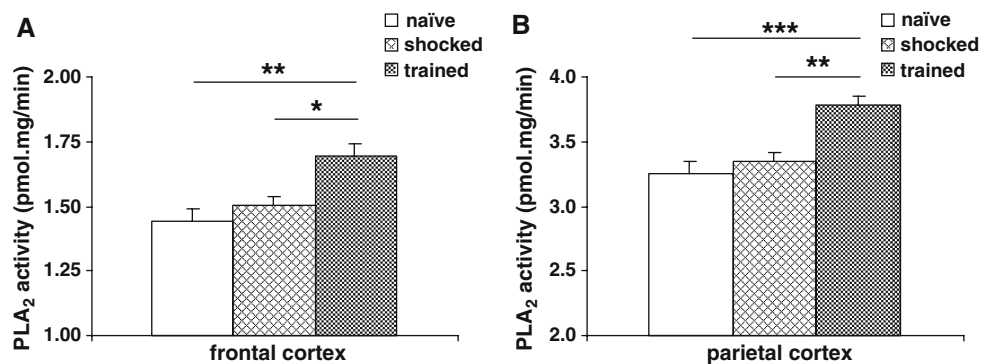


Fig. 2 PLA₂ activity measured 3 h after training. PLA₂ activity (pmol mg protein min⁻¹) is given as mean (\pm SEM). **a** In the frontal cortex, PLA₂ activity was increased in trained animals ($n = 11$) by 18% as compared to naïve controls ($n = 10$), and by 13% as compared to shocked controls ($n = 11$). **b** In the parietal cortex, PLA₂ activity was increased in trained animals ($n = 13$) by 16% as

compared to naïve controls ($n = 13$), and by 13% as compared to shocked controls ($n = 12$). Shocked and naïve controls had similar values of PLA₂ activity in both studies. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$. P values were calculated using Bonferroni's test after ANOVA: Frontal cortex: $F_{(2,29)} = 7.87$, $P < 0.01$; Parietal cortex: $F_{(2,35)} = 12.82$, $P < 0.001$

Shocked controls had similar values of PLA₂ activity as naïve controls ($P > 0.5$). P values were calculated using Bonferroni's test after ANOVA, $F_{(2,29)} = 7.87$, $P < 0.01$ (Fig. 2a).

Parietal association cortex PLA₂ activity was significantly increased in trained animals ($n = 13$) by 16% as compared to naïve controls ($n = 13$; $P < 0.001$), and by 13% as compared to shocked controls ($n = 12$; $P < 0.01$). Shocked controls had similar values of PLA₂ activity as naïve controls ($P > 0.5$). P values were calculated using Bonferroni's test after ANOVA, $F_{(2,35)} = 12.82$, $P < 0.001$ (Fig. 2b).

PLA₂ activity measured immediately after retrieval

Frontal association cortex Trained/tested animals ($n = 12$) had similar values of PLA₂ activity as naïve

($n = 11$; $P > 0.05$) and trained controls ($n = 14$; $P > 0.05$), and trained controls had similar values of PLA₂ activity as naïve controls ($P > 0.5$). P values were calculated using Bonferroni's test after ANOVA, $F_{(2,34)} = 3.29$, $P = 0.05$ (Fig. 3a).

Parietal association cortex PLA₂ activity was significantly increased in trained/tested animals ($n = 12$) by 27% as compared to naïve controls ($n = 11$; $P = 0.01$), and by 18% as compared to trained controls ($n = 14$; $P < 0.05$). Trained controls had similar values of PLA₂ activity as naïve controls ($P > 0.5$). P values were calculated using Bonferroni's test after ANOVA, $F_{(2,34)} = 6.24$, $P < 0.01$ (Fig. 3b).

Animals in the trained/tested group showed a mean step-down latency of 7 ± 5 s in the training session. These animals were also tested for retrieval 24 h after training,

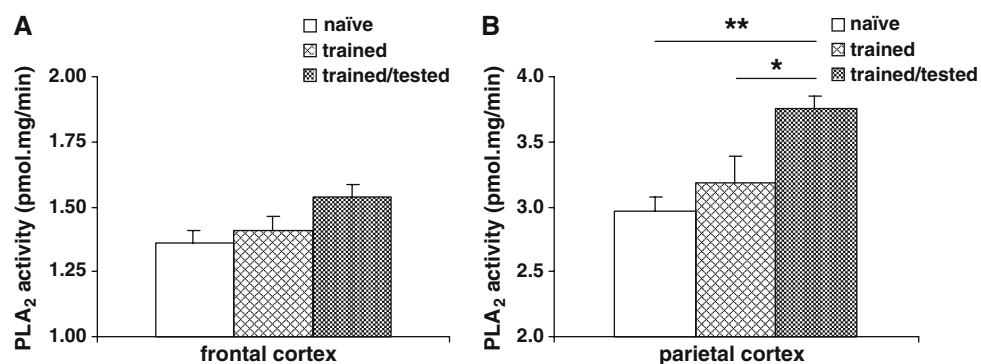


Fig. 3 PLA₂ activity measured immediately after retrieval. PLA₂ activity (pmol mg protein min⁻¹) is given as mean (\pm SEM). **a** In the frontal cortex, trained/tested animals ($n = 12$) had similar values of PLA₂ activity as naïve ($n = 11$) and trained controls ($n = 14$). **b** In the parietal cortex, PLA₂ activity was increased in trained/tested animals ($n = 12$) by 27% as compared to naïve controls ($n = 11$),

and by 18% as compared to trained controls ($n = 14$). Trained and naïve controls had similar values of PLA₂ activity in both studies. * $P < 0.05$, ** $P < 0.01$. P values were calculated using Bonferroni's test after ANOVA: Frontal cortex: $F_{(2,34)} = 3.29$, $P = 0.05$; Parietal cortex: $F_{(2,34)} = 6.24$, $P < 0.01$

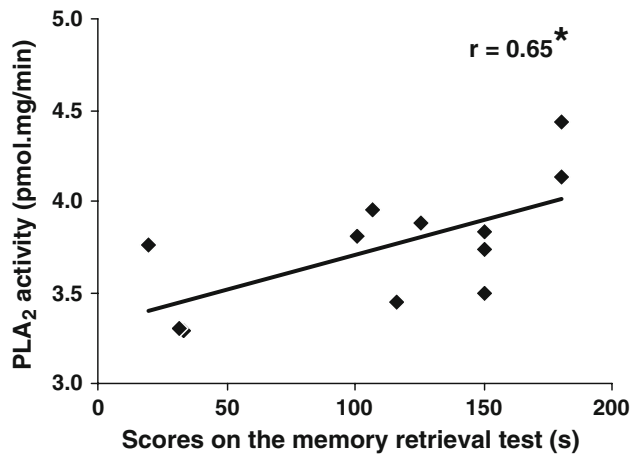


Fig. 4 Correlation between PLA₂ activity and scores on the memory retrieval test. PLA₂ activity (pmol mg protein min⁻¹) and scores on the memory retrieval test (i.e., test session step-down latencies, in seconds) are given as mean. Pearson correlation test showed a positive correlation between PLA₂ activity in the parietal cortex and scores on the memory retrieval test of rats within the trained/tested group ($n = 12$; $r = 0.65$, $*P < 0.05$)

showing a mean step-down latency of 112 ± 56 s, that is 16-fold higher than the mean training session step-down latency. Pearson correlation test showed a positive correlation between PLA₂ activity in the parietal cortex and scores on the memory retrieval test (i.e., test session step-down latencies) of rats within the trained/tested group ($r = 0.65$, $P < 0.05$) (Fig. 4).

Discussion

In our previous studies (Schaeffer and Gattaz 2005, 2007) we found that PLA₂ activity was increased in the CA1 field of rat hippocampus immediately after training and retrieval of the step-down IA task. In the present study we extended

our investigation to the cerebral cortex of rats, and found three major results. PLA₂ activity was increased in both frontal and parietal cortices of rats around the time of training and 3 h after training in the IA. However, PLA₂ activity was increased just in the parietal cortex of rats immediately after retrieval of the IA (Table 1). It should be noticed that, in both time intervals after training, PLA₂ activity was significantly increased in animals trained in the IA when compared to control animals that only received the electric foot shock (shocked controls) associated with the learning paradigm, indicating that increments in PLA₂ were specifically caused by the IA training. In the retrieval studies, we observed that the training effect on PLA₂ (in trained controls) disappeared after 24 h. However, the retrieval of the trained behavior in the IA task (in trained/tested animals) increased again the enzyme activity, indicating that increments in PLA₂ were specifically caused by the IA testing. Moreover, behavioral analysis revealed that animals in the trained/tested group showed a test session step-down latency in the IA 16-fold higher than the training session step-down latency, thus indicating good retention levels and that learning has occurred in this group. Most important, we found that increments in PLA₂ activity in the parietal cortex immediately after retrieval were highly correlated with scores on the memory retrieval test (i.e., test session step-down latencies) of rats within the trained/tested group. These findings support the suggestion that increments in PLA₂ activity immediately and 3 h after training were caused by learning. Altogether, the findings suggest that PLA₂ activity is differentially required in the frontal and parietal cortices of rats for the mechanisms of learning and retrieval of new contextual experience.

Experience-dependent changes have been extensively studied in rodent hippocampus and cerebral cortex in connection with contextual fear memory, and several biochemical mechanisms which are closely connected to

Table 1 Biochemical mechanisms involved in memory formation and retrieval of step-down inhibitory avoidance task in rats

	Memory formation							Memory retrieval
	Around training	30 min post-training	1 h post-training	1.5 h post-training	2 h post-training	3 h post-training	6 h post-training	
Frontal cortex	NMDA AMPA PKC PLA ₂	PKC		AMPA	PKC	NMDA AMPA		
Parietal cortex	AMPA PKC MAPK PLA ₂	PKC	NMDA	NMDA	PKC	NMDA PLA ₂	PKC PKC PKC	NMDA AMPA mGluR MAPK PLA ₂

PLA₂ have been implicated here. In the rat hippocampus, learning of the step-down IA was associated with elevations in the expression of NMDA NR1 subunit (Cammarota et al. 2000), increased activation of protein kinase C (PKC), Ca²⁺/calmodulin-dependent protein kinase II (CaMK II), p38, p42 and p44 mitogen-activated protein kinase (MAPK), and increased [³H]AMPA binding to the AMPA glutamate receptor (Cammarota et al. 1995, 1996, 1997, 1998; Bernabeu et al. 1995, 1997; Alonso et al. 2002, 2003). Learning of the step-through IA was also associated with increased activation of hippocampal PKC in rats (Young et al. 2002). Moreover, re-exposure of mice to context after training (contextual memory retrieval) stimulated the activity of hippocampal p42 and p44MAPK (Chen et al. 2005). Regarding the cerebral cortex, exposure of rats to the step-down IA resulted in increased activation of PKC in the frontal and parietal cortices at varying times after training (immediately, 30 min, and 2 h) (Bernabeu et al. 1995; Cammarota et al. 1997). Several pharmacological studies in rats have been conducted using the step-down IA, adding to the findings above. In the parietal cortex, blockade of NMDA receptors at varying times after training in the IA (1, 1.5 and 3 h) impaired memory consolidation (Zanatta et al. 1996; Izquierdo et al. 1997). In addition, blockade of AMPA receptors before or immediately after training in the IA disrupted memory acquisition and consolidation (Izquierdo et al. 1998). Furthermore, inhibition of MAPK activity immediately after training (Walz et al. 2000), and of PKC activity between 3 and 6 h after training in the IA impaired memory consolidation (Bonini et al. 2005). Finally, blockade of NMDA, AMPA and metabotropic glutamate receptors (mGluR), and inhibition of MAPK impaired memory retrieval of the IA (Quillfeldt et al. 1996; Izquierdo et al. 1997; Barros et al. 2000). In the prefrontal cortex, blockade of NMDA receptors at different times after training in the IA (immediately and 3 h) impaired memory consolidation (Mello et al. 2000). Additionally, blockade of AMPA receptors before training or at varying times after training in the IA (immediately, 1.5 and 3 h) disrupted memory acquisition and consolidation, respectively (Izquierdo et al. 1998, 2007). Blockade of AMPA receptors before or immediately after training also disrupted memory of the step-through IA (Liang et al. 1996). Data on the cerebral cortex described in this paragraph are summarized in Table 1.

As already mentioned, PLA₂ activity is closely connected to all biochemical mechanisms described above. PLA₂-dependent release of AA is a receptor-mediated process. In this way, activation of postsynaptic NMDA receptors raises postsynaptic [Ca²⁺]_i and stimulates cPLA₂, which generates AA, as found in mouse cortical neurons and rat hippocampal neurons and slices (Sanfeliu

et al. 1990; Pellerin and Wolfe 1991; Lazarewicz et al. 1992; Stella et al. 1995). Many studies have demonstrated that PLA₂ can be regulated by a variety of protein kinases. For example, activation of cPLA₂ is regulated by PKC (Wijkander and Sundler 1991; Nemenoff et al. 1993), p38MAPK (Zhou et al. 2003), p42MAPK (Lin et al. 1993; Nemenoff et al. 1993; Gordon et al. 1996) and CaMKII phosphorylation (Muthalif et al. 2001), and iPLA₂ activation is regulated by PKC phosphorylation (Underwood et al. 1998; Akiba et al. 1999). In turn, stimulation of PLA₂ activity in the presence of Ca²⁺ in rodent cortical and hippocampal slices as well as membrane preparations increased [³H]AMPA binding to the AMPA receptor and [³H]glutamate binding to AMPA and mGluR (Massicotte and Baudry 1990; Baudry et al. 1991; Massicotte et al. 1991; Tocco et al. 1992; Catania et al. 1993; Bernard et al. 1995; Chabot et al. 1998; Gaudreault et al. 2004), whereas PLA₂ inhibition and the Ca²⁺ chelator EGTA reduced agonist binding to AMPA and mGluR (Bernard et al. 1993, 1995; Catania et al. 1993). Additionally, PLA₂ inhibition in rat hippocampal slices prevented Ca²⁺-dependent formation of long-term potentiation (LTP; a synaptic model of learning and memory) in the CA1 field, as well as the increase of [³H]AMPA binding to the AMPA receptor that characterizes LTP (Bernard et al. 1994). These findings support the involvement of cPLA₂-mediated AA release in learning and memory. AA has been suggested to be also released by activation of mGluRs. Selective blockade of the mGluR5 subunit inhibited LTP in the CA1 field of rat hippocampal slices, and AA administration restored LTP, suggesting that during LTP group I mGluRs cause AA release that may be mediated by stimulation of iPLA₂ (Izumi et al. 2000). Further studies support a role for iPLA₂ in LTP. Selective inhibition of the iPLA₂-VIB isoenzyme prevented LTP induction in the CA1 field of rat hippocampal slices, as well as the associated increase of [³H]AMPA binding to the AMPA receptor and the up-regulation of AMPA GluR1 subunit levels in crude synaptic fractions (Martel et al. 2006). These findings support the involvement of iPLA₂-mediated AA release in learning and memory. Finally, AA potentiated the current through NMDA receptor channels in cerebellar granule cells, thus amplifying increases in [Ca²⁺]_i caused by glutamate (Miller et al. 1992), and induced a long-lasting potentiation of AMPA receptor currents by increasing Ca²⁺ influx in *Xenopus* oocytes expressing AMPA receptors containing GluR1,3 subunits (Nishizaki et al. 1999).

It is noteworthy that sPLA₂ enzymes (~13–18 kDa) require millimolar [Ca²⁺] for catalytic activity (Farooqui et al. 1999), the activation of 85 kDa cPLA₂ is regulated by nanomolar or micromolar [Ca²⁺]_i (Yoshihara and Watanabe 1990; Underwood et al. 1998), and iPLA₂ enzymes (~88 kDa) do not require Ca²⁺ for catalytic

activity (Larsson et al. 1998; Mancuso et al. 2000; Tanaka et al. 2000). We have previously optimized conditions for measuring cPLA₂ plus iPLA₂ activity or just iPLA₂ activity in rat brain homogenates (Schaeffer and Gattaz 2005). It was not possible to measure the activity of cPLA₂ alone, because iPLA₂ enzymes, which do not require Ca²⁺ for catalytic activity, can also respond to the optimal conditions for cPLA₂, i.e., nanomolar or micromolar [Ca²⁺]_i (Larsson et al. 1998; Mancuso et al. 2000; Tanaka et al. 2000). In fact, in our previous study, using optimized conditions for cPLA₂ (micromolar [Ca²⁺]_i and pH 8.5), we found a dominant activity of iPLA₂, while cPLA₂ activity was about 11-fold lower than the iPLA₂ activity. Accordingly, Yang et al. (1999) reported a dominant iPLA₂ activity over cPLA₂ activity in the rat hippocampus as well as whole brain. However, despite the low activity of cPLA₂ in the rat brain, there is evidence for the involvement of cPLA₂ in the formation of LTP (Bernard et al. 1994; Weichel et al. 1999). Therefore, because both cPLA₂ and iPLA₂ have been implicated in mechanisms of synaptic plasticity and/or learning and memory, we applied in the present study the conditions for measuring cPLA₂ plus iPLA₂ activity previously determined (Schaeffer and Gattaz 2005). Considering the methodological limitations, the findings of the present study, taken together with previous studies described above, allow four major conclusions. In the parietal cortex, (1) activation of cPLA₂ and/or iPLA₂ around the time of training might modulate memory formation through up-regulation of AMPA receptors via a PKC (in the case of cPLA₂ and iPLA₂) and a MAPK-dependent pathway (in the case of cPLA₂); (2) activation of cPLA₂ and/or iPLA₂ 3 h after training might modulate memory formation through up-regulation of NMDA receptors via a PKC-dependent pathway; and (3) activation of PLA₂ (likely cPLA₂) around the time of testing might modulate memory retrieval through up-regulation of NMDA, AMPA, and mGluRs via a MAPK-dependent pathway. In the frontal cortex, (4) activation of cPLA₂ and/or iPLA₂ around the time of training and 3 h after training might modulate memory formation through up-regulation of NMDA and AMPA receptors via a PKC-dependent pathway. We are not aware of any study till date, showing an involvement of sPLA₂ in learning and/or memory. Thus, we did not look at sPLA₂ in the present study.

In the context of AD, where reduced PLA₂ activity has been reported in the frontal and parietal cortices (Gattaz et al. 1995, 1996; Ross et al. 1998; Talbot et al. 2000), the present findings could suggest that reduced PLA₂ activity in the parietal cortex of AD patients might contribute to impairment of context learning and memory retrieval. Regarding the reduced PLA₂ activity in the frontal cortex

of AD patients, it might have a role in the impairment of context learning but not memory retrieval. Interestingly, a very recent study conducted by our group showed that cognitive training, consisting of a four-session memory training intervention for 1 month, increased PLA₂ activity in platelets of healthy elderly individuals, suggesting that memory training may have a modulating effect in PLA₂-mediated biological systems associated with cognitive functions (Talib et al. 2008). Because reduced PLA₂ activity has been reported in the frontal and parietal cortices of AD patients, and lower platelet PLA₂ activity was correlated with the severity of cognitive decline in samples of individuals with AD and mild cognitive impairment (Gattaz et al. 2004), the findings of the present study together with those of Talib et al. (2008) permit to speculate that stimulation of PLA₂ activity might offer new treatment strategies for the memory impairment seen in AD. Collectively, the data support the use of cognitive training as a promising non-pharmacological approach to stimulate PLA₂ at least in healthy elderly subjects for the prevention of cognitive deficits.

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References

- Abrisqueta-Gomez J, Canali F, Vieira VL, Aguiar AC, Ponce CS, Brucki SM, Bueno OF (2004) A longitudinal study of a neuropsychological rehabilitation program in Alzheimer's disease. *Arq Neuropsiquiatr* 62:778–783
- Akiba S, Mizunaga S, Kume K, Hayama M, Sato T (1999) Involvement of group VI Ca²⁺-independent phospholipase A₂ in protein kinase C-dependent arachidonic acid liberation in zymosan-stimulated macrophage-like P388D1 cells. *J Biol Chem* 274:19906–19912
- Alonso M, Viola H, Izquierdo I, Medina JH (2002) Aversive experiences are associated with a rapid and transient activation of ERKs in the rat hippocampus. *Neurobiol Learn Mem* 77:119–124
- Alonso M, Bevilacqua LR, Izquierdo I, Medina JH, Cammarota M (2003) Memory formation requires p38MAPK activity in the rat hippocampus. *Neuroreport* 14:1989–1992
- Avila R, Bottino CM, Carvalho IA, Santos CB, Seral C, Miotto EC (2004) Neuropsychological rehabilitation of memory deficits and activities of daily living in patients with Alzheimer's disease: a pilot study. *Braz J Med Biol Res* 37(11):1721–1729
- Barros DM, Izquierdo LA, Mello e Souza T, Ardenghi PG, Pereira P, Medina JH, Izquierdo I (2000) Molecular signalling pathways in the cerebral cortex are required for retrieval of one-trial avoidance learning in rats. *Behav Brain Res* 114:183–192
- Baudry M, Massicotte G, Hauge S (1991) Opposite effects of phospholipase A₂ on [3H]AMPA binding in adult and neonatal membranes. *Brain Res Dev Brain Res* 61:265–267
- Belleville S, Gilbert B, Fontaine F, Gagnon L, Ménard E, Gauthier S (2006) Improvement of episodic memory in persons with mild

- cognitive impairment and healthy older adults: evidence from a cognitive intervention program. *Dement Geriatr Cogn Disord* 22:486–499
- Bernabeu R, Cammarota M, Izquierdo I, Medina JH (1997) Involvement of hippocampal AMPA glutamate receptor changes and the cAMP/protein kinase A/CREB-P signalling pathway in memory consolidation of an avoidance task in rats. *Braz J Med Biol Res* 30:961–965
- Bernabeu R, Izquierdo I, Cammarota M, Jerusalinsky D, Medina JH (1995) Learning-specific, time-dependent increase in [3H]phorbol dibutyrate binding to protein kinase C in selected regions of the rat brain. *Brain Res* 685:163–168
- Bernard J, Lahsaini A, Baudry M, Massicotte G (1993) The phospholipase A₂ inhibitor bromophenacyl bromide prevents the depolarization-induced increase in [3H]AMPA binding in rat brain synaptoneurosomes. *Brain Res* 628:340–344
- Bernard J, Lahsaini A, Massicotte G (1994) Potassium-induced long-term potentiation in area CA1 of the hippocampus involves phospholipase activation. *Hippocampus* 4:447–453
- Bernard J, Chabot C, Gagne J, Baudry M, Massicotte G (1995) Melittin increases AMPA receptor affinity in rat brain synaptoneurosomes. *Brain Res* 671:195–200
- Bonini JS, Cammarota M, Kerr DS, Bevilaqua LR, Izquierdo I (2005) Inhibition of PKC in basolateral amygdala and posterior parietal cortex impairs consolidation of inhibitory avoidance memory. *Pharmacol Biochem Behav* 80:63–67
- Cammarota M, Izquierdo I, Wolfman C, Levi de Stein M, Bernabeu R, Jerusalinsky D, Medina JH (1995) Inhibitory avoidance training induces rapid and selective changes in 3[H]AMPA receptor binding in the rat hippocampal formation. *Neurobiol Learn Mem* 64:257–264
- Cammarota M, Bernabeu R, Izquierdo I, Medina JH (1996) Reversible changes in hippocampal 3H-AMPA binding following inhibitory avoidance training in the rat. *Neurobiol Learn Mem* 66:85–88
- Cammarota M, Paratcha G, Levi de Stein M, Bernabeu R, Izquierdo I, Medina JH (1997) B-50/GAP-43 phosphorylation and PKC activity are increased in rat hippocampal synaptosomal membranes after an inhibitory avoidance training. *Neurochem Res* 22:499–505
- Cammarota M, Bernabeu R, Levi De Stein M, Izquierdo I, Medina JH (1998) Learning-specific, time-dependent increases in hippocampal Ca²⁺/calmodulin-dependent protein kinase II activity and AMPA GluR1 subunit immunoreactivity. *Eur J NeuroSci* 10:2669–2676
- Cammarota M, de Stein ML, Paratcha G, Bevilaqua LR, Izquierdo I, Medina JH (2000) Rapid and transient learning-associated increase in NMDA NR1 subunit in the rat hippocampus. *Neurochem Res* 25:567–572
- Catania MV, Hollingsworth Z, Penney JB, Young AB (1993) Phospholipase A₂ modulates different subtypes of excitatory amino acid receptors: autoradiographic evidence. *J Neurochem* 60:236–245
- Chabot C, Gagne J, Giguere C, Bernard J, Baudry M, Massicotte G (1998) Bidirectional modulation of AMPA receptor properties by exogenous phospholipase A₂ in the hippocampus. *Hippocampus* 8:299–309
- Chen J, Engle SJ, Seilhamer JJ, Tischfield JA (1994) Cloning and recombinant expression of a novel human low molecular weight Ca²⁺-dependent phospholipase A₂. *J Biol Chem* 269:2365–2368
- Chen X, Garelick MG, Wang H, Lil V, Athos J, Storm DR (2005) PI3 kinase signaling is required for retrieval and extinction of contextual memory. *Nat Neurosci* 8:925–931
- Clare L, Wilson BA, Carter G, Roth I, Hodges JR (2002) Relearning face-name associations in early Alzheimer's disease. *Neuropsychology* 16:538–547
- Clements MP, Rose SP (1996) Time-dependent increase in release of arachidonic acid following passive avoidance training in the day-old chick. *J Neurochem* 67:1317–1323
- Dennis EA (1994) Diversity of group types, regulation, and function of phospholipase A₂. *J Biol Chem* 269:13057–13060
- Dennis EA (1997) The growing phospholipase A₂ superfamily of signal transduction enzymes. *Trends Biochem Sci* 22:1–2
- Farooqui AA, Yang HC, Rosenberger TA, Horrocks LA (1997) Phospholipase A₂ and its role in brain tissue. *J Neurochem* 69:889–901
- Farooqui AA, Litsky ML, Farooqui T, Horrocks LA (1999) Inhibitors of intracellular phospholipase A₂ activity: their neurochemical effects and therapeutical importance for neurological disorders. *Brain Res Bull* 49:139–153
- Fujita S, Ikegaya Y, Nishiyama N, Matsuki N (2000) Ca²⁺-independent phospholipase A₂ inhibitor impairs spatial memory of mice. *Jpn J Pharmacol* 83:277–278
- Gattaz WF, Maras A, Cairns NJ, Levy R, Forstl H (1995) Decreased phospholipase A₂ activity in Alzheimer brains. *Biol Psychiatry* 37:13–17
- Gattaz WF, Cairns NJ, Levy R, Forstl H, Braus DF, Maras A (1996) Decreased phospholipase A₂ activity in the brain and in platelets of patients with Alzheimer's disease. *Eur Arch Psychiatry Clin Neurosci* 246:129–131
- Gattaz WF, Forlenza OV, Talib LL, Barbosa NR, Bottino CM (2004) Platelet phospholipase A₂ activity in Alzheimer's disease and mild cognitive impairment. *J Neural Transm* 111:591–601
- Gaudreault SB, Chabot C, Gratton JP, Poirier J (2004) The caveolin scaffolding domain modifies 2-amino-3-hydroxy-5-methyl-4-isoxazole propionate receptor binding properties by inhibiting phospholipase A₂ activity. *J Biol Chem* 279:356–362
- Gelb MH, Valentin E, Ghomashchi F, Lazdunski M, Lambeau G (2000) Cloning and recombinant expression of a structurally novel human secreted phospholipase A₂. *J Biol Chem* 275:39823–39826
- Gordon RD, Leighton IA, Campbell DG, Cohen P, Creaney A, Wilton DC, Masters DJ, Ritchie GA, Mott R, Taylor IW, Bundell KR, Douglas L, Morten J, Needham M (1996) Cloning and expression of cystolic phospholipase A₂ (cPLA₂) and a naturally occurring variant. Phosphorylation of Ser505 of recombinant cPLA₂ by p42 mitogen-activated protein kinase results in an increase in specific activity. *Eur J Biochem* 238:690–697
- Holscher C (1995) Prostaglandins play a role in memory consolidation in the chick. *Eur J Pharmacol* 294:253–259
- Holscher C, Rose SP (1994) Inhibitors of phospholipase A₂ produce amnesia for a passive avoidance task in the chick. *Behav Neural Biol* 61:225–232
- Holscher C, Canevari L, Richter-Levin G (1995) Inhibitors of PLA₂ and NO synthase cooperate in producing amnesia of a spatial task. *Neuroreport* 6:730–732
- Izquierdo I, Quillfeldt JA, Zanatta MS, Quevedo J, Schaeffer E, Schmitz PK, Medina JH (1997) Sequential role of hippocampus and amygdala, entorhinal cortex and parietal cortex in formation and retrieval of memory for IA in rats. *Eur J NeuroSci* 9:786–793
- Izquierdo I, Izquierdo LA, Barros DM, Mello e Souza T, de Souza MM, Quevedo J, Rodrigues C, Sant'Anna MK, Madruga M, Medina JH (1998) Differential involvement of cortical receptor mechanisms in working, short-term and long-term memory. *Behav Pharmacol* 9:421–427
- Izquierdo LA, Barros DM, da Costa JC, Furini C, Zinn C, Cammarota M, Bevilaqua LR, Izquierdo I (2007) A link between role of two prefrontal areas in immediate memory and in long-term memory consolidation. *Neurobiol Learn Mem* 88:160–166
- Izumi Y, Zarrin AR, Zorumski CF (2000) Arachidonic acid rescues hippocampal long-term potentiation blocked by group I

- metabotropic glutamate receptor antagonists. *Neuroscience* 100:485–491
- Kishimoto K, Matsumura K, Kataoka Y, Morii H, Watanabe Y (1999) Localization of cytosolic phospholipase A₂ messenger RNA mainly in neurons in the rat brain. *Neuroscience* 92:1061–1077
- Larsson Forsell PK, Kennedy BP, Claesson HE (1999) The human calcium-independent phospholipase A₂ gene multiple enzymes with distinct properties from a single gene. *Eur J Biochem* 262:575–585
- Larsson PK, Claesson HE, Kennedy BP (1998) Multiple splice variants of the human calcium-independent phospholipase A₂ and their effect on enzyme activity. *J Biol Chem* 273:207–214
- Lazarewicz JW, Salinska E, Wroblewski JT (1992) NMDA receptor-mediated arachidonic acid release in neurons: role in signal transduction and pathological aspects. *Adv Exp Med Biol* 318:73–89
- Liang KC, Hu SJ, Chang SC (1996) Formation and retrieval of inhibitory avoidance memory: differential roles of glutamate receptors in the amygdala and medial prefrontal cortex. *Chin J Physiol* 39:155–166
- Lin LL, Wartmann M, Lin AY, Knopf JL, Seth A, Davis RJ (1993) cPLA₂ is phosphorylated and activated by MAP kinase. *Cell* 72:269–278
- Lowry OH, Rowebrugh NJ, Farr LA, Randall RJ (1951) Protein measurement with the Folin phenol reagent. *J Biol Chem* 193:265–275
- Mancuso DJ, Jenkins CM, Gross RW (2000) The genomic organization, complete mRNA sequence, cloning, and expression of a novel human intracellular membrane-associated calcium-independent phospholipase A₂. *J Biol Chem* 275:9937–9945
- Martel MA, Patenaude C, Menard C, Alaux S, Cummings BS, Massicotte G (2006) A novel role for calcium-independent phospholipase A in α -amino-3-hydroxy-5-methylisoxazole-propionate receptor regulation during long-term potentiation. *Eur J NeuroSci* 23:505–513
- Massicotte G, Baudry M (1990) Modulation of DL- α -amino-3-hydroxy-5-methylisoxazole-4-propionate (AMPA)/quisqualate receptors by phospholipase A₂ treatment. *Neurosci Lett* 118:245–248
- Massicotte G, Vanderklish P, Lynch G, Baudry M (1991) Modulation of DL- α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid/quisqualate receptors by phospholipase A₂: a necessary step in long-term potentiation? *Proc Natl Acad Sci USA* 88:1893–1897
- Mello E, Souza T, Vianna MR, Rodrigues C, Quevedo J, Moleta BA, Izquierdo I (2000) Involvement of the medial prefrontal cortex in memory consolidation for inhibitory avoidance learning in rats. *Pharmacol Biochem Behav* 66:615–622
- Miller B, Sarantis M, Traynelis SF, Attwell D (1992) Potentiation of NMDA receptor currents by arachidonic acid. *Nature* 355:722–725
- Molloy GY, Rattray M, Williams RJ (1998) Genes encoding multiple forms of phospholipase A₂ are expressed in rat brain. *Neurosci Lett* 258:139–142
- Muthalif MM, Hefner Y, Cnaan S, Harper J, Zhou H, Parmentier JH, Aebersold R, Gelb MH, Malik KU (2001) Functional interaction of calcium/calmodulin-dependent protein kinase II and cytosolic phospholipase A(2). *J Biol Chem* 276:39653–39660
- Nemenoff RA, Winitz S, Qian NX, Van Putten V, Johnson GL, Heasley LE (1993) Phosphorylation and activation of a high molecular weight form of phospholipase A₂ by p42 microtubule-associated protein 2 kinase and protein kinase C. *J Biol Chem* 268:1960–1964
- Nishizaki T, Matsuoka T, Nomura T, Enikolopov G, Sumikawa K (1999) Arachidonic acid potentiates currents through Ca²⁺-permeable AMPA receptors by interacting with a CaMKII pathway. *Molec Brain Res* 67:184–189
- Owada Y, Tominaga T, Yoshimoto T, Kondo H (1994) Molecular cloning of rat cDNA for cytosolic phospholipase A₂ and the increased gene expression in the dentate gyrus following transient forebrain ischemia. *Brain Res Mol Brain Res* 25:364–368
- Paxinos G, Watson C (1998) *The rat brain in stereotaxic coordinates*. Academic Press, San Diego
- Pellerin L, Wolfe LS (1991) Release of arachidonic acid by NMDA-receptor activation in the rat hippocampus. *Neurochem Res* 16:983–989
- Pickard RT, Striffler BA, Kramer RM, Sharp JD (1999) Molecular cloning of two new human paralogs of 85-kDa cytosolic phospholipase A₂. *J Biol Chem* 274:8823–8831
- Quillfeldt JA, Zanatta MS, Schmitz PK, Quevedo J, Schaeffer E, Lima JB, Medina JH, Izquierdo I (1996) Different brain areas are involved in memory expression at different times from training. *Neurobiol Learn Mem* 66:97–101
- Rapp S, Brenes G, Marsh AP (2002) Memory enhancement training for older adults with mild cognitive impairment: a preliminary study. *Aging Ment Health* 6:5–11
- Ross BM, Moszczynska A, Erlich J, Kish SJ (1998) Phospholipid-metabolizing enzymes in Alzheimer's disease: increased lysophospholipid acyltransferase activity and decreased phospholipase A₂ activity. *J Neurochem* 70:786–793
- Sanfeliu C, Hunt A, Patel AJ (1990) Exposure to *N*-methyl-D-aspartate increases release of arachidonic acid in primary cultures of rat hippocampal neurons and not in astrocytes. *Brain Res* 526:241–248
- Sato T, Ishida T, Irifune M, Tanaka K, Hirate K, Nakamura N, Nishikawa T (2007) Effect of NC-1900, an active fragment analog of arginine vasopressin, and inhibitors of arachidonic acid metabolism on performance of a passive avoidance task in mice. *Eur J Pharmacol* 560:36–41
- Schaeffer EL, Gattaz WF (2005) Inhibition of calcium-independent phospholipase A₂ activity in rat hippocampus impairs acquisition of short- and long-term memory. *Psychopharmacology (Berl)* 181:392–400
- Schaeffer EL, Gattaz WF (2007) Requirement of hippocampal phospholipase A₂ activity for long-term memory retrieval in rats. *J Neural Transm* 114:379–385
- Stella N, Pellerin L, Magistretti PJ (1995) Modulation of the glutamate-evoked release of arachidonic acid from mouse cortical neurons: involvement of a pH-sensitive membrane phospholipase A₂. *J Neurosci* 15:3307–3317
- Suzuki N, Ishizaki J, Yokota Y, Higashino K, Ono T, Ikeda M, Fujii N, Kawamoto K, Hanasaki K (2000) Structures, enzymatic properties, and expression of novel human and mouse secretory phospholipase A₂s. *J Biol Chem* 275:5785–5793
- Talbot K, Young RA, Jolly-Tornetta C, Lee VM, Trojanowski JQ, Wolf BA (2000) A frontal variant of Alzheimer's disease exhibits decreased calcium-independent phospholipase A₂ activity in the prefrontal cortex. *Neurochem Int* 37:17–31
- Talib LL, Yassuda MS, Diniz BSO, Forlenza OV, Gattaz WF (2008) Cognitive training increases platelet PLA₂ activity in healthy elderly subjects. *Prostaglandins Leukot Essent Fatty Acids* 78:265–269
- Tanaka H, Takeya R, Sumimoto H (2000) A novel intracellular membrane-bound calcium-independent phospholipase A₂. *Biochem Biophys Res Commun* 272:320–326
- Tocco G, Massicotte G, Standley S, Thompson RF, Baudry M (1992) Phospholipase A₂-induced changes in AMPA receptor: an autoradiographic study. *Neuroreport* 3:515–518
- Underwood KW, Song C, Kriz RW, Chang XJ, Knopf JL, Lin LL (1998) A novel calcium-independent phospholipase A₂, cPLA₂- γ , that is prenylated and contains homology to cPLA₂. *J Biol Chem* 273:21926–21932

- Vianna MR, Barros DM, Silva T, Choi H, Madche C, Rodrigues C, Medina JH, Izquierdo I (2000) Pharmacological demonstration of the differential involvement of protein kinase C isoforms in short- and long-term memory formation and retrieval of one-trial avoidance in rats. *Psychopharmacology (Berl)* 150:77–84
- Walz R, Roesler R, Quevedo J, Sant'Anna MK, Madruga M, Rodrigues C, Gottfried C, Medina JH, Izquierdo I (2000) Time-dependent impairment of inhibitory avoidance retention in rats by posttraining infusion of a mitogen-activated protein kinase inhibitor into cortical and limbic structures. *Neurobiol Learn Mem* 73:11–20
- Weichel O, Hilgert M, Chatterjee SS, Lehr M, Klein J (1999) Bilobalide, a constituent of Ginkgo biloba, inhibits NMDA-induced phospholipase A₂ activation and phospholipid breakdown in rat hippocampus. *Naunyn Schmiedebergs Arch Pharmacol* 360:609–615
- Wenisch E, Cantegreil-Kallen I, De Rotrou J, Garrigue P, Moulin F, Batouche F, Richard A, De Sant'Anna M, Rigaud AS (2007) Cognitive stimulation intervention for elders with mild cognitive impairment compared with normal aged subjects: preliminary results. *Aging Clin Exp Res* 19:316–322
- Wijkander J, Sundler R (1991) An 100-kDa arachidonate-mobilizing phospholipase A₂ in mouse spleen and the macrophage cell line J774. Purification, substrate interaction and phosphorylation by protein kinase C. *Eur J Biochem* 202:873–880
- Yang HC, Mosior M, Ni B, Dennis EA (1999) Regional distribution, ontogeny, purification, and characterization of the Ca²⁺-independent phospholipase A₂ from rat brain. *J Neurochem* 73:1278–1287
- Yoshihara Y, Watanabe Y (1990) Translocation of phospholipase A₂ from cytosol to membranes in rat brain induced by calcium ions. *Biochem Biophys Res Commun* 170:484–490
- Young E, Cesena T, Meiri KF, Perrone-Bizzozero NI (2002) Changes in protein kinase C (PKC) activity, isozyme translocation, and GAP-43 phosphorylation in the rat hippocampal formation after a single-trial contextual fear conditioning paradigm. *Hippocampus* 12:457–464
- Zanatta MS, Schaeffer E, Schmitz PK, Medina JH, Quevedo J, Quillfeldt JA, Izquierdo I (1996) Sequential involvement of NMDA receptor-dependent processes in hippocampus, amygdala, entorhinal cortex and parietal cortex in memory processing. *Behav Pharmacol* 7:341–345
- Zhou H, Das S, Murthy KS (2003) Erk1/2- and p38 MAP kinase-dependent phosphorylation and activation of cPLA₂ by m3 and m2 receptors. *Am J Physiol Gastrointest Liver Physiol* 284:G472–G480