



# Hollow sphere nickel sulfide nanostructures–based enzyme mimic electrochemical sensor platform for lactic acid in human urine

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## Abstract

An enzyme-free electrochemical sensor platform is reported based on hollow sphere structured nickel sulfide (HS-NiS) nanomaterials for the sensitive lactic acid (LA) detection in human urine. Hollow sphere nickel sulfide nanostructures directly grow on the nickel foam (NiF) substrate by using facile and one-step electrochemical deposition strategy towards the electrocatalytic lactic acid oxidation and sensing for the first time. The as-developed nickel sulfide nanostructured electrode (NiF/HS-NiS) has been successfully employed as the enzyme mimic electrode towards the enhanced electrocatalytic oxidation and detection of lactic acid. The NiF/HS-NiS electrode exhibits an excellent electrocatalytic activity and sensing ability with low positive potential ( $\sim 0.52$  V vs Ag/AgCl), catalytic current density ( $\sim 1.34$  mA), limit of detection (LOD) ( $0.023$   $\mu\text{M}$ ), linear range from  $0.5$  to  $88.5$   $\mu\text{M}$  with a correlation coefficient of  $R^2 = 0.98$ , sensitivity ( $0.655$   $\mu\text{A } \mu\text{M}^{-1} \text{cm}^{-2}$ ), and selectivity towards the lactic acid owing to the ascription of high inherent electrical conductivity, large electrochemical active surface area (ECASA), high electrochemical active sites, and strong adsorption ability. The sensors developed in this work demonstrate the selectivity against potential interferences, including uric acid (UA), ascorbic acid (AA), paracetamol (PA),  $\text{Mg}^{2+}$ ,  $\text{Na}^+$ , and  $\text{Ca}^{2+}$ . Furthermore, the developed sensors show practicability by sensing lactic acid in human urine samples, suggesting that the HS-NiS nanostructures device has promising clinical diagnostic potential.

**Keywords** Nickel sulfide · Hollow architecture · Nanostructures · Modified electrode · Electrocatalysis · Lactic acid detection

## Introduction

The detection of chemically and biologically active species is vital in the area of healthcare and medical diagnosis [1–3]. Numerous enzymatic biosensors were developed for the detection and determination of biomarkers, including glucose, NADH,  $\text{H}_2\text{O}_2$ , cholesterol, lactic acid (LA), etc. through either in vitro or in vivo investigations [4–9]. In particular, lactic acid biosensors are in highly in-demand in various applications such as clinical analysis, biomedical, biological, food industries, and sports for the rapid detection and determination of

lactic acid levels in our bodily fluids [10–14]. The elevated concentrations of lactic acid may not only specify the existence but also indicate the harshness of clinically imperative disorders [12]. Consequently, it is required to design a lactic acid sensor gifted with simplicity to prepare, quickness in response time, low cost, high selectivity, and high sensitivity. Due to the several demerits of the enzyme-based biosensor [15–20], a new design of electrochemical sensors with artificial or enzyme mimic catalysts received much attention. Alongside enzyme-based electrochemical biosensors, the enzyme-free sensors are getting excessive consideration due to the advantages of cost-effective fabrication, enhanced electrochemical redox properties, durability and good reproducibility, etc. [21, 22].

Recently, numerous enzyme-free electrochemical sensors based on various nanostructured materials such as noble metal nanoparticles of gold (Au), platinum (Pt), and silver (Ag) and its alloy–metals [23–25] and transition metal oxides such as nickel oxide (NiO), copper oxide (CuO), manganese oxide ( $\text{MnO}_2$ ), and  $\text{Co}_3\text{O}_4$  were developed [26]. Recently, transition metal chalcogenides (TMC) were considered as promising

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candidates due to their low cost, ease of biocompatible, and layer- and morphological-dependent physico-chemical characteristics [27, 28]. Transition metal chalcogenides have been employed in many potential applications, including electrochemical sensors, electrocatalysis, high-performance supercapacitors, and lithium-ion batteries [29, 30]. In particular, binary nickel sulfides (NiS) possessed enhanced electrocatalytic properties through high electronic conductivity, rich redox chemistry, different atomic configurations, and crystalline structures [31, 32]. For instance, Liu Co-workers developed the various crystal phases and morphologies of NiS<sub>x</sub> through the solvothermal reaction, exhibiting various structure dependent-electrocatalytic properties [33]. Zhang Co-workers reported the microwave-assisted synthesis of NiS<sub>2</sub> with controlled morphologies such as nanocubes, nanospheres, and nanoparticles [34]. Therefore, the development of high-performance and enzyme mimic electrochemical lactic acid sensor based on NiS nanostructures is a key.

In the present study, a novel hollow sphere-architected NiS nanomaterial was directly grown on NiF substrate for the sensitive lactic acid detection in human urine for the first time. To our knowledge, the present enzyme-free electrochemical sensor possessed several advantages, including (i) one-step and simple electrochemical fabrication of sensing electrode strategy; (ii) growing hollow sphere-architected NiS nanomaterials with high dense NiS clusters (~2.0 nm) with free of any templates/capping agents; (iii) time-effective and a green strategy without using of any organic solutions; and (iv) enzyme mimic electrocatalytic activity towards the sensitive detection of lactic acid.

## Experimental section

### Reagents and solutions

Nickel(II)nitrate hexahydrate (Ni(NO<sub>3</sub>)<sub>2</sub>·6H<sub>2</sub>O) was purchased from the Sisco laboratory. Potassium hydroxide pellets (KOH) were purchased from Emplura. Thiourea was purchased from Alfa Aeser. Nitric acid (HNO<sub>3</sub>) was purchased from Rankem analytic. Lactic acid, uric acid, ascorbic acid, paracetamol, magnesium sulfate hexahydrate, sodium chloride, and calcium sulfate pentahydrate were obtained from Sigma Aldrich. All reagents purchased and utilized in this study were of analytical grade and utilized without further purification. Millipore Milli-Q nanopure water (resistivity ≥ 18 MΩ cm) was employed for the preparation of all solutions.

### Characterization of hollow sphere NiS nanostructures

The crystal structures of the as-prepared hollow sphere NiS nanomaterials were primarily characterized using X-ray diffraction (XRD) technique with a Pananalytical Xpert Pro

Diffractionmeter. The surface morphology of the hollow sphere NiS nanomaterials was characterized using a high-resolution scanning electron microscope (FE-SEM) with an FEI Quanta FEG 200, and a high-resolution transmission electron microscope (HR-TEM) with a JEOL 2010F TEM. The elemental composition and distribution of the hollow sphere NiS nanomaterials were studied by using electron dispersive X-ray (EDX) techniques with Hitachi SU-70. X-ray photoelectron spectroscopic measurements were performed to understand the chemical compositions and the valence states of hollow sphere NiS nanomaterials using a XPS-PHI Versaprobe III.

### Electrochemical measurements

All the electrochemical measurements were conducted by employing an Electrochemical Bio Logic SP-300 workstation out at an ambient temperature of 25 ± 3 °C. The NiF modified with HS-NiS nanomaterials was used as the working electrode, a platinum wire employed as the counter electrode and Ag/AgCl (3.0 M KCl) electrode used as the reference electrode. Cyclic voltammetric (CV) technique was employed to study the electrochemical properties and electrocatalytic properties of the NiF/HS-NiS electrode. Chronoamperometric (CA) technique was utilized for the sensing performance of the NiF/HS-NiS electrode.

### Electrochemical sensor fabrication

Electrochemical deposition strategy was adopted to fabricate HS-NiS nanostructures. The pre-cleaned NiF substrate (surface area of ~0.18 cm<sup>2</sup>) was employed as the working electrode to deposit the HS-NiS. Initially, the NiF was cleaned by 3 min sonication in acetone and pure water before to electrochemical deposition process. Typically, there are five continuous electrochemical cyclings of the potential window, starting from 0.6 to -1.2 V (vs Ag/AgCl) in the mixture of 5.0 mM Ni salt precursor + 0.75 M thiourea + 0.1 M nitric acid at a scan rate of 5.0 mV s<sup>-1</sup> (Fig. S1). The electrochemical deposited hollow-structured NiS nanomaterials on NiF, designated as NiF/HS-NiS. The mass loading of HS-NiS was estimated to be 0.024 mg. The developed NiF/HS-NiS electrode will be employed as the sensing electrode for the detection of lactic acid under alkaline electrolytes.

### Preparation of actual human urinary sample

An original human urine sample was collected from a healthy volunteer (~24 years old) and stored in the refrigerator immediately. A small volume (~5.0 mL) serum sample was then centrifuged for 30 min. The supernatant was filtered and then diluted ~10 times using 1.0 M KOH. The resulting solution was transferred into the electrochemical cell for real sample analysis without any further pretreatment. A stock solution of lactic acid (1.0 mM) was stored in a refrigerator, which was further diluted

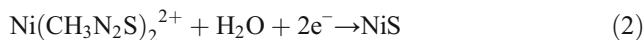
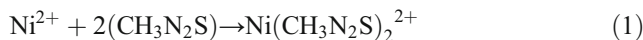
to obtain 0.5, 1.0, and 2.0  $\mu\text{M}$  of lactic acid. For the lactic acid determination in urine samples, the standard addition method (SAM) was employed for the detection of lactic acid concentrations ranged from 0.5 to 2.0  $\mu\text{M}$ . Chronoamperometric technique was used to perform the recovery test for the detection and determination of lactic acid in the human urine samples by adding a defined concentration of lactic acid. Based on our controlled amperometric studies, the concentration of lactic acid is negligible or zero concentration in the actual urine samples.

## Results and discussion

### Morphological characterization of hollow sphere NiS nanostructures

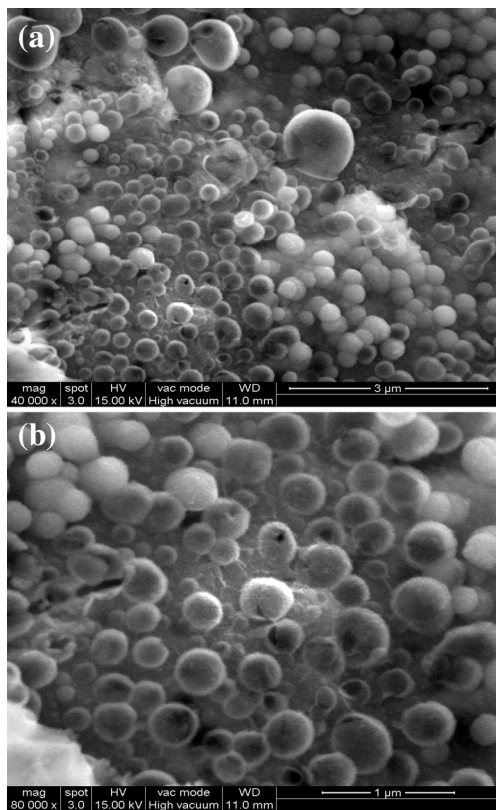
Figure 1 presents the typical FE-SEM images of the developed hollow sphere-structured NiS nanostructures with low (A) and high (B) magnifications. The average size of the formed hollow sphere NiS nanostructures was calculated to be  $\sim 184.0$  nm with hollow size of  $\sim 93.0$  nm as shown in Fig. 1. As can be easily seen in Fig. 1(A), the hollow sphere NiS nanostructures were homogeneously distributed using a single step electrochemical deposition method. As depicted in Fig. S1, the CVs of the NiF electrode recorded in the electrolyte solution of 5.0 mM Ni precursor + 0.75 M thiourea +

0.1 M nitric acid at a scan rate of  $5.0 \text{ mV s}^{-1}$  for five continuous cycles. During the electrochemical deposition, the formed  $\text{Ni}(\text{CH}_3\text{N}_2\text{S})_2^{2+}$  was then reduced to form HS-NiS on the NiF substrate as given in Eqs. (1) and (2) [33]. It may be anticipated the so-formed  $\text{Ni}(\text{CH}_3\text{N}_2\text{S})_2^{2+}$  primarily on edge of the pores of NiF, and grown as hollow sphere NiS nanostructures on the NiF substrate.

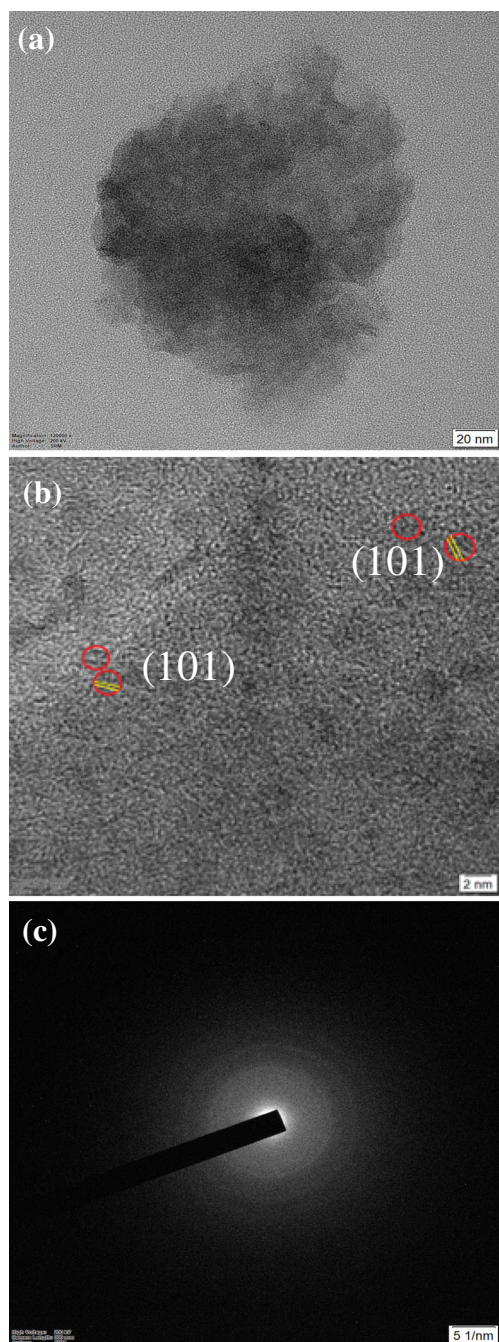


The atomic ratio of Ni to S was measured to be 68.2:31.8 for the hollow sphere NiS nanostructures using the EDX spectrum of the hollow sphere NiS nanostructures (Fig. S2). The elements of Ni and S were homogeneously distributed in hollow sphere NiS nanostructures as depicted in Fig. S3. Figure 2 displays the TEM, HR-TEM, and selected area electron diffraction (SAED) pattern of the developed hollow sphere NiS nanostructures. As depicted in Figs. 2(A) and S4, the sphere-like NiS has the dimension of  $\sim 130.0$  nm, and the size of hollow was found to be  $\sim 35.0$  nm, which agrees with the FE-SEM results of Fig. 1. The hollow structure of the NiS was found based on the bright-field TEM image of Fig. 2(A). It is interesting to note that a high dispersion of cluster-like NiS was found in Fig. S4, and the average size of the cluster was estimated to be  $\sim 2.3$  nm. Figure 2(B) shows the HR-TEM image of the hollow sphere NiS nanostructures. The value of lattice fringes was estimated to be 0.27 nm, correspond to the ascription of the crystalline plane of (101) NiS. The SAED pattern was recorded for the obtained hollow sphere NiS nanostructures and is shown in Fig. 2(C). The resulting SAED pattern of the hollow sphere NiS nanostructures had not shown any clear lattice fringes to investigate further the crystalline facets and  $d$  values, suggesting that the developed NiS in this study was amorphous crystalline structures. Figure S5 shows the EDX spectrum of the hollow sphere NiS nanostructures, illustrated the existence of Ni and S in the region. The atomic ratio was further estimated based on the EDX spectrum of the NiS nanostructures to be 64.5:35.5 which is well agreed with Fig. S2.

The chemical compositions and the valence states of the prepared hollow sphere NiS nanostructures were further analyzed with XPS technique. Figure 3 presents the high-resolution XPS spectra for Ni 2P and S 2P for the hollow sphere NiS nanostructures. The core-level XPS Ni spectrum (Fig. 3(A)) exhibited a couple of major peaks at 855.9 and 861.7 eV, originally generating from  $\text{Ni}2\text{P}_{1/2}$  and  $\text{Ni}2\text{P}_{3/2}$  for  $\text{Ni}^{2+}$  in the NiS nanostructures [33, 35, 36]. The other two peaks at 861.3 and 879.7 eV were ascribed to satellite peaks for NiS nanostructures. Figure 3(B) presents the S 2P spectrum of the NiS nanostructures. The major XPS peaks were observed at

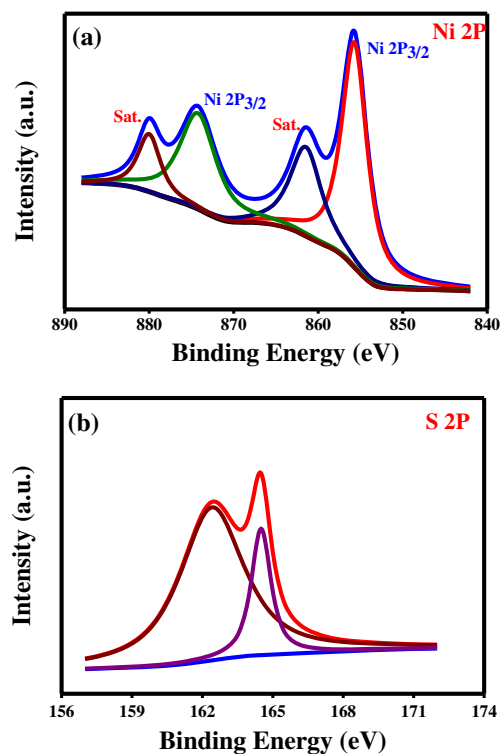


**Fig. 1** FE-SEM images of the NiF/HS-NiS electrode with low (a) and high (b) magnifications



**Fig. 2** TEM (a) and HR-TEM (b) images of the HS-NiS nanomaterials; (c) SAED pattern of the HS-NiS nanomaterials

163.6 and 168.3 eV, corresponding to the binding energies of  $S^{2-}$  [37]. The atomic composition ratio of Ni and S was measured to be 6.5: 3.5 for NiS nanomaterials. The XPS results of Fig. 3 further confirmed the existence of NiS. The crystalline nature of the developed hollow sphere NiS nanomaterials was further studied with XRD analysis. Figure S6 presents the XRD pattern of the developed hollow sphere NiS nanomaterials modified NiF (red curve) and bare NiF (black curve) electrodes. The small peaks observed at 2 values of  $\sim 18.3^\circ$  and  $\sim 27.2^\circ$  were ascribed to (110) and (220) hexagonal crystalline



**Fig. 3** XPS spectra of the Ni 2p (a), and S 2p (b) regions for the HS-NiS nanomaterials

plane of NiS [33, 38]. The other major peaks marked with star (\*) were derived from the NiF substrate.

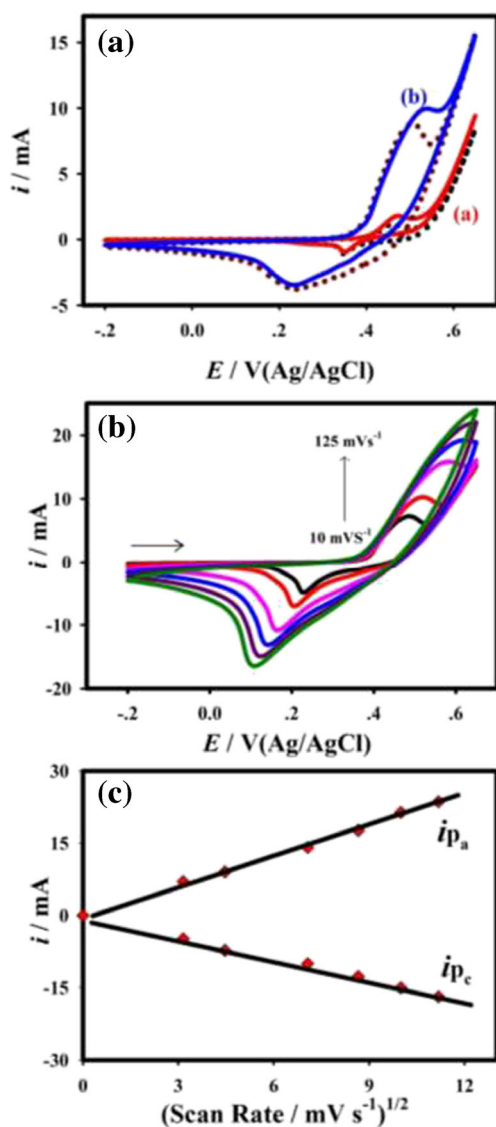
### Electrochemical characterization of hollow sphere NiS nanostructures

The electrochemical characteristics of the hollow sphere NiS nanomaterials were further studied with a cyclic voltammetric technique under alkaline medium. Figure S7 presents the CV curves of the NiF (a) and NiF modified with HS-NiS (NiF/HS-NiS) (c) electrodes recorded at different scan rates, starting from 10 to 125  $\text{mV s}^{-1}$  under 1.0 M KOH in the electrochemical potential window of  $-0.2$  to  $0.65$  V (vs Ag/AgCl). As expected, a pair of electrochemical oxidation and reduction peaks were observed in the potential range of  $0.1$ – $0.5$  V for the NiF and NiF/HS-NiS electrodes, ascribed to the  $\text{Ni}^{2+}/\text{Ni}^{3+}$  redox couple [33]. It is noted that the CV curves area of the NiF/HS-NiS electrode is much higher by  $\sim 4$  times in comparison to bare NiF, suggesting that NiF/HS-NiS electrode possessed a larger double-layer capacitance. As shown in Fig. S7, the anodic and cathodic currents were increased as the square root of scan rates increases, showing that both bare NiF and NiF/HS-NiS electrodes exhibited the diffusion-controlled process under alkaline electrolyte. It is further exposed that there is no substantial change in the amount of the electrochemically active species involved in the electrochemical redox reactions. The electrochemical active surface area (ECASA) of the NiF/HS-

NiS electrode (Fig. S8) was estimated to be  $\sim 1.43 \text{ cm}^2$  based on the equation of  $R_f S$  [39], where  $R_f$  is the roughness factor, and  $S$  stands for the geometric surface area of the electrode.

### Electrocatalytic oxidation of lactic acid at the hollow sphere NiS nanostructures

The electrocatalytic activity of the developed NiF/HS-NiS electrode was tested towards the oxidation of lactic acid. Figure 4(a) displays the CV curves of the bare NiF and NiF/HS-NiS electrodes recorded in the presence (solid curve) and absence (dotted curve) of 10.0 mM lactic acid at a scan rate of  $20 \text{ mV s}^{-1}$  in the potential window of  $-0.2$  to  $0.6 \text{ V}$  (vs Ag/AgCl). As shown in Fig. 4(a), the anodic peak current ( $i_a$ ) was increased about  $\sim 1.34 \text{ mA}$  at the potential of  $\sim 0.52 \text{ V}$  after



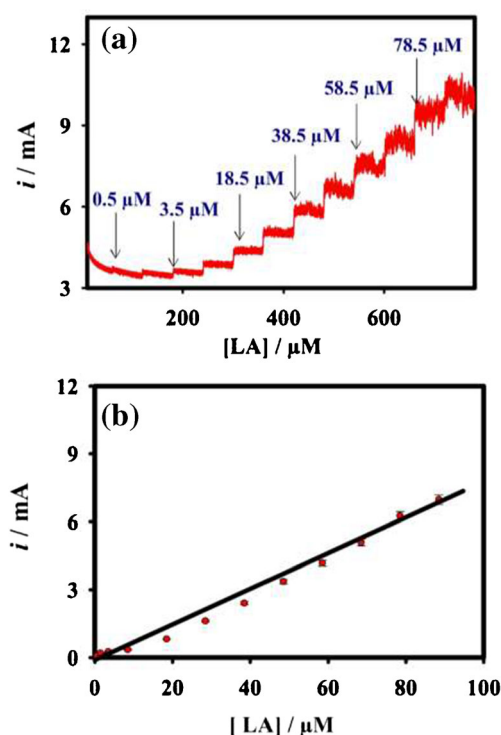
**Fig. 4** CV curves of the bare NiF (curve a, red color) and NiF/HS-NiS (curve b, blue curve) electrodes recorded in the absence (dotted lines) and presence of 10.0 mM lactic acid in 1.0 M KOH at a scan rate of  $20 \text{ mV s}^{-1}$

the addition of 10.0 mM lactic acid due to the ascription of the electrochemical oxidation of lactic acid. However, there was no significant enhancement of the anodic current observed at the bare NiF electrode as presented in Fig. 4(a). Figure 4(b) shows the CV curves of the NiF/HS-NiS electrode recorded with various scan rates in the presence of 10.0 mM lactic acid. As can be seen in Fig. 4(c), a linear plot was observed on the plotting of anodic currents against the square root of the scan rates, suggesting that the electrochemical oxidation of lactic acid was mediated by the diffusion of electroactive species at the NiF/HS-NiS electrode. As expected, the electrochemical oxidation of lactic acid at the NiF/HS-NiS electrode is an irreversible process. The electrochemical rate of the reaction towards the lactic acid oxidation at the NiF/HS-NiS, and bare NiF electrodes was estimated to be  $\sim 1.63 \times 10^{-3}$ , and  $\sim 0.36 \times 10^{-3} \text{ mol s}^{-1} \text{ cm}^2$ , respectively. During the electrochemical oxidation of lactic acid, lactate is formed electrochemically, followed by the chemical reaction of lactate to pyruvic acid (Eqs. 3 and 4). The active  $\text{Ni}^{2+}$  species is initially converted to  $\text{Ni}^{3+}$  species during the anodic scan through the electrochemical oxidation reaction. The as-formed  $\text{Ni}^{3+}$  species rapidly oxidizes the lactic acid molecules to lactate via two-electron transfer process [40].



### Electroanalytical performance of the hollow sphere NiS nanostructures

The electrochemical detection of lactic acid was performed with an amperometric  $i-t$  technique based on the electrocatalytic activity of the NiF/HS-NiS electrode. Figure 5(a) depicts the current–time curves of the NiF/HS-NiS electrode recorded under 1.0 M KOH during the consecutive adding of lactic acid at regular intervals of 60 s at an applied electrode potential ( $E_{\text{app}}$ ) of 0.50 V (vs Ag/AgCl). The applied potential ( $E_{\text{app}}$ ) was chosen based on the electrocatalytic lactic acid oxidation potential (Fig. 4(a)). The anodic current was steadily increased after the addition of various concentrations of lactic acid, starting from 0.5 to 88.5  $\mu\text{M}$  due to the association of lactic acid oxidation at the NiF/HS-NiS electrode. The limit of detection (LOD) was estimated as 0.023  $\mu\text{M}$  based on the equation of  $3\sigma/b$  where  $\sigma$  means the standard deviation of the blank and  $b$  represents the slope of the calibration curve. The steady-state current response was obtained within 3 s following each addition of lactic acid. Figure 5(b) displays the corresponding calibration plot of Fig. 5(a). The linear relationship was attained for the anodic currents against the lactic acid concentration, which was observed in the concentration range from 0.5 to 88.5  $\mu\text{M}$  with a correlation coefficient of  $R^2 = 0.98$  and a sensitivity of



**Fig. 5** (a) Amperometric  $i$ - $t$  response of the NiF/HS-NiS electrode recorded in 1.0 M KOH under various lactic acid concentrations, starting from 0.5 to 88.5  $\mu\text{M}$ ,  $E_{\text{app}}$ : 0.50 V. (b) Corresponding calibration plot of measured anodic current against lactic acid concentrations

$0.655 \mu\text{A} \mu\text{M}^{-1} \text{cm}^{-2}$ . The obtained low calibration regression was due to the high double-layer capacitance of the developed NiF/HS-NiS electrode. To our knowledge, the present sensor based on hollow sphere NiS nanomaterials exhibited such a lowest detection limit and high sensitivity against the detection of lactic acid. To show the merit of this non-enzymatic amperometric sensor, a list of recent electrochemical lactic sensor was

summarized in Table 1. It is highly evident that the hollow sphere NiS nanomaterials demonstrated the lowest detection limit and the highest sensitivity in comparison with reported sensors [41–50]. The attained high performance of the electrochemical sensing attribution was due to the association of the good electrocatalytic aptitude of hollow sphere NiS nanomaterials.

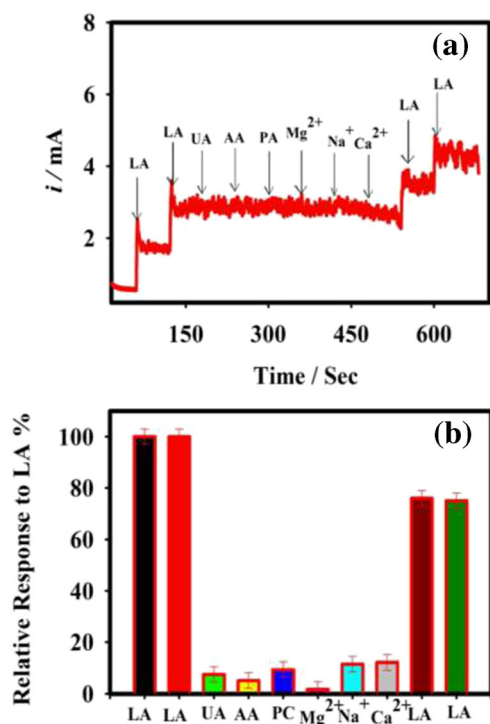
The selectivity of the developed sensor was tested with various interferences such as uric acid (UA), ascorbic acid (AA), paracetamol (PA),  $\text{Mg}^{2+}$ ,  $\text{Na}^+$ , and  $\text{Ca}^{2+}$  with 100-fold higher concentration when compared to lactic acid concentration. The potential interferences were chosen because of their excessive presence in real human urine samples. Figure 6 displays the amperometric  $i$ - $t$  response of the NiF/HS-NiS electrode towards the 10.0  $\mu\text{M}$  lactic acid in the presence of 1.0 mM uric acid (UA), ascorbic acid (AA), paracetamol (PA),  $\text{Mg}^{2+}$ ,  $\text{Na}^+$ , and  $\text{Ca}^{2+}$ . The relative amperometric response to lactic acid at the NiF/HS-NiS electrode in the absence and in the presence of potential interferences are shown in Fig. 6(b). It is suggested that the developed NiF/HS-NiS electrode in the present study exhibited good selectivity to lactic acid without using any enzymes on the electrode surface, mimicking the enzymatic activity. The stability of the lactic acid sensor based on NiF/HS-NiS electrode was performed by conducting the amperometric technique in the presence of 10.0 mM LA at the constant applied potential of 0.50 V over 5000 s under alkaline electrolyte. As presented in Fig. S9, a plot of the measured catalytic current versus time showed that the anodic current was slightly decreased by less than  $\sim 7\%$ , which was due to the ascription of adsorption of oxidized products of lactic acid. The developed electrochemical lactic acid sensor based on NiF/HS-NiS electrode possessed good durability.

The precision of the lactic acid sensor was performed by repeating the electrocatalytic oxidation of 10.0 mM lactic acid

**Table 1** Comparison of the performance of various non-enzymatic lactic acid sensors with the sensor fabricated in the present work sensing electrode

Materials	Technique	Sensitivity	Linear range	LOD	Ref.
AuR-DTSP-LO <sub>x</sub>	Amperometry	$1.49 \mu\text{A} \text{mM}^{-1}$	0.1 mM - 1.3 mM	3.93 $\mu\text{M}$	[44]
MWCNT/MB	Amperometry	$3.46 \mu\text{A} \text{cm}^{-2} \text{mM}^{-1}$	0.10–10.0 mM	7.5 $\mu\text{M}$	[49]
$\text{Fe}_3\text{O}_4/\text{MWCNT}$	Amperometry	$7.67 \mu\text{A} \text{mM}^{-1}$	50.0–500.0 $\mu\text{M}$	5.0 $\mu\text{M}$	[50]
LDH/ $\text{CeO}_2$ /GCE	Amperometry	$571.19 \mu\text{A} \text{mM}^{-1}$	0.2–2.0 mM	50.0 $\mu\text{M}$	[48]
LO <sub>x</sub> /SWCNT	Amperometry	$9.4 \mu\text{A} \text{mM}^{-1} \text{cm}^{-2}$	0.01–0.9 mM	3.0 $\mu\text{M}$	[51]
CoPC-SPCE	Amperometry	–	74.0 $\mu\text{M}$ –1.5 mM	18.3 $\mu\text{M}$	[52]
NiO NPS	Amperometry	–	0.01–5.0 mM	5.7 $\mu\text{M}$	[53]
GCE/MIP	Amperometry	$1.9 \times 10^5 \mu\text{A} \text{M}^{-1}$	0.1–15.0 nM	0.0089 nM	[23]
GCE/ $\text{Co}_3\text{O}_4$	Amperometry	–	0.05–3.0 mM	6.0 $\mu\text{M}$	[11]
HS-NiS	Amperometry	$0.65 \mu\text{A} \mu\text{M}^{-1} \text{cm}^{-2}$	0.5–88.5 $\mu\text{M}$	0.023 $\mu\text{M}$	This Work

Au NPS, gold nanoparticles; GCE, glassy carbon electrode; LDH, lactate dehydrogenase; LO<sub>x</sub>, lactate oxidase; MWCNT, multi-walled carbon nanotubes; PPY, polypyrrole; Pt, platinum; Pttca, poly-5,2'-5'-2"-terthiophene-3'-carboxylic acid; LDA, lactate dehydrogenase



**Fig. 6** Amperometric *i-t* responses of the NiF/HS-NiS electrode recorded in 1.0 M KOH with the addition of 10.0 μM lactic acid, 1.0 mM uric acid, 1.0 mM ascorbic acid, 1.0 mM paracetamol, 1.0 mM Mg<sup>2+</sup>, 1.0 mM Na<sup>+</sup>, 1.0 mM Ca<sup>2+</sup>, and 10.0 μM lactic acid at  $E_{app}$ : 0.50 V. **(b)** Measured relative response of the NiF/HS-NiS electrode in the presence of various interferences

under alkaline electrolyte at the NiF/HS-NiS electrode. The resulting relative standard deviation (RSD) was estimated to be 2.3% for three individual measurements. In addition, the present electrochemical sensor exhibited the repeatability of ~90% for three successive measurements. Thus, the developed NiF/HS-NiS-based lactic acid sensor exhibited good precision.

### Application to human urinary samples

Furthermore, the developed NiF/HS-NiS based sensing electrode was successfully verified to validate its ability towards the detection and determination of lactic acid in real human urine samples by using the standard addition method (SAM). In this SAM, prior to the addition of lactic acid in urine samples was considered as the baseline or zero concentration of analyte for the recovery values of lactic acid concentrations ranged from 0.5 to 2.0 μM. The resulting recovery values of the lactic acid in a human urine sample at the NiF/HS-NiS electrodes are listed in Table 2. The recovery values of the lactic acid in human urinary were reached in the range of 97.3–99.2% with relative standard deviation (RSD %) values between 1.2 and 1.6, suggesting that the established sensing electrode based on hollow sphere NiS nanostructures

**Table 2** Recovery tests of lactic acid in actual human urine samples ( $n = 3$ ) at the NiF/HS-NiS

Material	Added/ μM	Found <sup>a</sup> / μM	Recovery/ %	RSD/ %
HS-NiS	0.50	0.49	99.2	1.49
	1.0	0.97	97.3	1.16
	2.0	1.96	98.2	1.59

<sup>a</sup> Average of three measurements

possessed a strong prospective to empower applications in clinical and biomedical analyses.

### Conclusion

In the present study, we have reported a facile, one-step and green electrochemical approach to fabricate hollow sphere NiS nanostructures based sensing electrode towards the high-performance lactic acid detection for the first time. The developed hollow sphere NiS nanostructures exhibited an outstanding electrocatalytic activity towards the lactic acid oxidation through the unique hollow sphere surface morphology, highly exposed electrochemical active sites in the absence of any binder, enzyme mimic electrocatalytic activity and huge electrochemical active surface area. Thus, the hollow sphere NiS nanostructured sensing electrode delivered a low detection limit, fast response time, high sensitivity, and wide linear range. The present sensor exhibited selectivity without the utilization of any enzymes in the presence of a variety of potential interferences such as uric acid (UA), ascorbic acid (AA), paracetamol (PA), Mg<sup>2+</sup>, Na<sup>+</sup>, and Ca<sup>2+</sup>. The sensor based on NiF/HS-NiS electrode was furthermore successfully performed in real human urine samples to distinguish selectively, revealing its potential practicability. The present study is not only present a novel approach for the electrochemical fabrication of the hollow sphere NiS nanostructured sensing electrode but also offers a promising enzyme-free electrochemical sensor towards the detection of lactic acid in human urine. Moreover, further sensing experiment tests using cell lines, organs, and other biological systems are requisite towards the development of a next-generation electrochemical lactic acid sensor device for various clinical and biomedical applications.

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## Compliance with ethical standards

**Conflict of interest** The author(s) declare that they have no competing interests.

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