ORIGINAL PAPER

Nonenzymatic amperometric sensing of glucose using a glassy carbon electrode modified with a nanocomposite consisting of reduced graphene oxide decorated with $Cu₂O$ nanoclusters

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Abstract We describe a simple solvothermal method for preparation of reduced graphene oxide nanosheets decorated with uniform $Cu₂O$ nanoclusters by using poly(vinyl pyrrolidone)-poly(methacrylamide)-poly(vinyl imidazole) triblock co-polymer as a shape-directing agent and L-glutamic acid as a reducing agent. The resulting nanocomposite was deposited on a glassy carbon electrode where it displays improved electrocatalytic activity toward glucose oxidation in 0.5 M NaOH. This observation was exploited to construct a nonenzymatic amperometric sensor for glucose. It has a detection limit as low as 1.0 μ M, high sensitivity (23.058 μ A mM⁻¹), and a dynamic (analytical) range that extends from 5.0 to 9595 μM at a working potential of 600 mV (vs. SCE).

Keywords Reduced graphene oxide .Nanoclusters .Triblock copolymer . Glucose . Electrocatalysis

Introduction

A variety of noble metal nanostructures with desired physical and/or chemical properties has been prepared [[1\]](#page-6-0), but their

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high price and rare storage seriously confine their commercial applications. Therefore, it is essential to search for lowcost and plentiful metals (e.g., Ni, Cu, Co, and Fe) as alternatives.

 $Cu₂O$ is one of the most investigated semiconductors and has wide applications in CO oxidation, biosensors, solar energy conversion, and photocatalysis [[2\]](#page-6-0). Its catalytic properties are tightly correlated with the respective structural features, size, and morphology [[3\]](#page-6-0). As a result, it is highly desired to prepare novel $Cu₂O$ nanocatalysts with fine-controlled structure, dimension, and shape [[4\]](#page-6-0). Accordingly, many shape-controlled $Cu₂O$ nanostructures have been prepared, including cages [[5](#page-6-0)], cubes [\[6](#page-6-0)], and spheres [[7\]](#page-6-0).

Many supports have been introduced to endow the above catalysts with well distribution, especially graphene oxide (GO) nanosheets [\[7](#page-6-0)] or reduced graphene oxide (r-GO) [[7\]](#page-6-0), owing to their enlarged surface area, high electrical conductivity, better chemical stability, and strong adhesion to the catalysts [[8\]](#page-6-0). To date, many nanocomposites, especially metal oxide-graphenes, have been synthesized, including $Cu₂O-$ graphene [\[9\]](#page-6-0), NiO-graphene [\[10](#page-6-0)], TiO₂-graphene [\[11\]](#page-6-0), and MnO₂-graphene [\[12\]](#page-6-0).

Enzyme-based electrochemical biosensors display high sensitivity and selectivity [[13](#page-6-0)]. Nevertheless, they suffer from poor stability and reproducibility. Alternatively, non-enzymatic sensors possess better durability and stability against external environment toward glucose oxidation [[14](#page-6-0)]. For example, Zhou et al. fabricated monodisperse porous $Cu₂O/r-GO$, which showed excellent catalytic activity and good selectivity for glucose detection [[15](#page-6-0)]. In another example, Gao's group prepared mesocrystalline Cu₂O hollow nanocubes for construction of glucose sensors [\[14\]](#page-6-0). However, their relatively higher detection limit and narrow linear range are unsatisfied during accurately monitoring glucose.

Polymers have strong interactions and sometimes form coordination complexes with nanocrystals, and thereby influence the growth kinetics of the nanocrystals as capping agents and shape-directing agents [[16](#page-6-0)]. A variety of polymers (e.g., cationic, anionic and nonionic polymers) has been used for shape-controlled synthesis of nanocrystals [[17](#page-7-0)]. Polyvinylpyrrolidone (PVP) is the most frequently used nonionic polymer in this field [[18](#page-7-0)]. Copolymers, containing two or more different monomers, have more functional properties [[19](#page-7-0)]. For example, poly(ethylene glycol)-block-poly(propylene glycol)-block-poly(ethylene glycol) Pluronic® P123 was used as a shape-directing agent for shape-controlled synthesis of mesoporous silica [[20](#page-7-0)].

Herein, a facile solvothermal method is described for synthesis of uniform $Cu₂O$ nanoclusters on r-GO (denoted as $Cu₂O$ NCs/r-GO), using poly(vinyl pyrrolidone)poly(methacrylamide)-poly(vinyl imidazole) triblock copolymer (Fig. S1, Electronic Supplementary Material, ESM) and L-glutamic acid as a shape-directing agent and a reducing agent. The catalytic performance of $Cu₂O$ NCs/r-GO was investigated through the fabrication of a nonenzymatic glucose sensor.

Experimental

Reagents and materials

Copper(II) nitrate trihydrate, sodium hydroxide, and glucose were purchased from Shanghai Sigma-Aldrich (Shanghai, China, www.sigmaaldrich.com). Poly(vinyl pyrrolidone) poly(methacrylamide)-poly(vinyl imidazole) triblock copolymer (Luviset^R Clear), graphite powder (99.95 %, 8000 mesh), L-glutamic acid, ascorbic acid (AA), dopamine (DA), uric acid (UA), lactic acid, glutaric acid, acetaminophen, and NaBH4 were bought from Shanghai Aladdin Chemical Reagent Company (Shanghai, China, [www.chemicalbook.com\)](http://www.chemicalbook.com/). The other chemicals were of analytical grade and used without further purification. Double distilled-water was used to prepare all of the aqueous solutions throughout the whole experiments.

Preparation of $Cu₂O$ NCs/r-GO and of $Cu₂O$ nanoparticles

Typically, GO was prepared from natural graphite powder by a modified Hummers' method, and experimental details are given in the previous work [[21](#page-7-0)].

For typical preparation of $Cu₂O$ NCs/r-GO, 0.5 g of poly(vinyl pyrrolidone)-poly(methacrylamide)-poly(vinyl imidazole) triblock copolymer was dissolved into 15 mL of ethanol under stirring. Then, 5 mL of GO $(1 \text{ mg } \text{mL}^{-1})$, 0.121 g

of $Cu(NO₃)₂·3H₂O$, and 0.3 g of *L*-glutamic acid were put into the mixed solution, respectively. After homogeneously stirring, the mixed solution was transferred into a Teflon lined stainless steel autoclave (25 mL), heated at 160 °C for 6 h, and cooled to 25 °C in air. The final black precipitates were collected by centrifugation, thoroughly washed with water and ethanol for several times, and dried in vacuum at 60 °C for further characterization.

Similarly, $Cu₂O$ nanoparticles were fabricated without GO for comparison, and pure r-GO were obtained by reducing GO with NaBH₄, while other conditions were kept unchanged.

Characterization

The morphology of the samples was characterized by transmission electron microscopy (TEM) and high-resolution transmission electron microscopy (HRTEM), which were performed on a JEM-2100 F transmission electron microscope [\(www.jeol.co.jp/en.com\)](http://www.jeol.co.jp/en.com) equipped with the selective area electron diffraction (SAED). The crystal structures were determined by X-ray diffraction (XRD, Rigaku Dmax-2000 diffractometer) using Cu-Kα radiation (Bruker Co., Germany, www.rigaku.com). X-ray photoelectron spectra (XPS) were recorded on a Thermo Scientific Escalab 250 XPS spectrometer ([www.pharmaceuticalonline.com\)](http://www.pharmaceuticalonline.com/) with Al Kα X-ray radiation (1486.6 eV). Fourier transform infrared (FT-IR) analysis was performed on a Nicolet NEXUS670 spectrometer [\(http://web.ysu.edu/gen/stem](http://web.ysu.edu/gen/stem)). Raman spectra were acquired on a micro-Raman system (Renishaw RM1000 spectrometer) with an excitation wavelength of 633 nm [\(www.antpedia.](http://www.antpedia.com/) [com\)](http://www.antpedia.com/). Thermogravimetric analysis (TGA) was conducted with a simultaneous thermo-gravimetric analyzer (Netzsch, STA 449C, www.brain-power.com). The samples were heated in air from 25 to 800 °C at a heating rate of 10 °C min⁻¹.

Electrochemical measurements

The electrochemical measurements were conducted on a CHI660D electrochemical workstation (Chenhua Instruments Co., Shanghai, China, www.chinstruments.com), and performed with a conventional three-electrode system, including a platinum wire as the counter electrode, a saturated calomel electrode (SCE, saturated KCl) as the reference electrode, and a bare or modified glassy carbon electrode (GCE, 3.0 mm in diameter) as the working electrode.

For typical construction of $Cu₂O$ NCs/r-GO modified electrode (Cu₂O NCs/r-GO/GCE), 2.0 mg of Cu₂O NCs/r-GO was dispersed into 1.0 mL water under ultrasonication to form a homogeneous suspension. Next, 8 μL of the suspension was casted on the clean GCE and dried naturally, followed by casting another layer of Nafion (0.05 wt %) with the volume of 5 μL to fix the deposit on the electrode surface. For comparison, individual r -GO and Cu₂O modified electrodes

were prepared in a similar way, denoted as r-GO/GCE and Cu₂O NCs/GCE, respectively.

The electrocatalytic activity of the above catalysts modified electrodes was determined by linear sweep voltammetry toward glucose oxidation in 0.5 M NaOH at a scan rate of 50 mV s^{-1} . The stability of Cu₂O NCs/r-GO/GCE was examined by amperometry at an applied potential of 600 mV in 0.5 M NaOH. The electrochemical experiments were carried out at 25 °C, if not stated otherwise.

Results and discussion

Characterization

As seen from TEM images (Fig. 1a and b), the product mainly consists of numerous well-defined nanoclusters across the whole section, without any agglomerating, which are uniformly dispersed on r-GO. The nanoclusters have a very narrow size distribution from 1.2 to 3.9 nm (Fig. 1d), with the average size of 2.6 nm. Their polycrystalline nature is demonstrated by the corresponding SAED pattern (inset in Fig. 1b). HRTEM image (Fig. 1c) shows the well-defined lattice fringes with the inter-planar spacing of ca. 0.19 nm, which is in good accordance with the (200) crystal planes of $Cu₂O$.

It is known that surfactants have great effects on the morphology and size of nanocrystals [\[16](#page-6-0), [17](#page-7-0)]. As expected, only some irregular, agglomerated, and large $Cu₂O$ particles (Fig. [2a, b](#page-3-0)) are obtained without or using smaller amount of the triblock copolymer (0.1%) . When the amounts of the

triblock copolymer are 0.5% (Fig. [2c\)](#page-3-0) and 1.0% (Fig. [2d\)](#page-3-0), numerous $Cu₂O$ nanoparticles with decreased sizes are observed on r-GO. These results indicate that the amounts of the triblock copolymer played the essential role in controlling the morphology and size of $Cu₂O$ NCs.

Fig. S2 (ESM) provides the XRD pattern of $Cu₂O$ NCs/r-GO, using r-GO as a reference. Clearly, there are four representative diffraction peaks emerged at 36.5, 42.6, 61.5, and 73.6°, which are well assigned to the (111), (200), (220), and (311) planes of the face-centered cubic (fcc) structure of $Cu₂O$ [[22](#page-7-0), [23](#page-7-0)]. These values are matched well with the Joint Committee Powder Diffraction Standard of pure $Cu₂O$ (JCPDS No. 05–0667). The corresponding (111) diffraction peak is stronger and sharper than those of the other planes, showing the preferential growth of $Cu₂O$ NCs along the (111) directions [\[15\]](#page-6-0). Furthermore, there are no any other peaks of impurities (e.g., CuO and Cu) emerged, indicating high purity of Cu2O NCs. In addition, a new broad peak is observed at 2θ =19.4° for Cu₂O NCs/r-GO (Fig. S2, ESM, curve a), which is consistent with that of r-GO (Fig. S2, ESM, curve b, $2\theta = 20.9^{\circ}$), suggesting the efficient removal of oxygencontaining functional groups and well dispersion of $Cu₂O$ NCs on r-GO [[24\]](#page-7-0).

The composition and surface states of $Cu₂O$ NCs/r-GO were examined by XPS measurements. The XPS survey spectrum (Fig. [3a](#page-3-0)) demonstrates the coexistence of Cu, O, N, and C elements in $Cu₂O$ NCs/r-GO. As for high-resolution Cu XPS spectrum (Fig. [3b](#page-3-0)), there are two peaks detected at 933.7 and 953.8 eV, which are assigned to the binding energies of Cu $2p_{3/2}$ and Cu $2p_{1/2}$ of Cu(I) [\[25,](#page-7-0) [26](#page-7-0)], respectively. A

using 2.5 % triblock copolymer. d The corresponding sizedistribution histograms. Inset in B shows the corresponding SAED pattern

Fig. 2 TEM images of $Cu₂O$ nanoparticles obtained with different concentrations of the triblock copolymer a 0 %, b 0.1 %, c 0.5 %, and **d** 1.0 %

weak peak shows up at 943.7 eV, suggesting that there is a very minimal amount of Cu(II) present in this system, which may be derived from the un-reacted precursor of $Cu(NO₃)₂$ [\[27\]](#page-7-0). By measuring their relative intensities, Cu(I) is the predominant species, revealing the formation of Cu₂O NCs.

High-resolution C 1 s XPS spectrum of $Cu₂O$ NCs/r-GO (Fig. 3c) can be well fitted into four peaks at 284.70, 285.70, 287.58 eV, and 288.67 eV, which are attributed to the C-C $(sp²)$, C-O, C = O, and O-C = O groups, respectively [[28\]](#page-7-0). Particularly, the peaks related to the oxygen functionalities are much weaker in comparison with those of GO (Fig. 3d),

Fig. 3 a Survey, high-resolution b Cu 2p, and C 1 s XPS spectra of c Cu₂O NCs/r-GO and d GO, respectively

showing the formation of r-GO under hydrothermal conditions. This assumption is strongly supported by FT-IR spectra of $Cu₂O$ NCs/r-GO (Fig. 4A). Notably, the peak intensities at 1046 cm⁻¹ (O–H) and 1730 cm⁻¹ (C = O) are greatly decreased for $Cu₂O$ NCs/r-GO (curve a) as compared to those of GO (curve b).

Similarly, Raman spectra (Fig. 4B) display two characteristic peaks at 1362 and 1608 cm⁻¹ for the D and G bands in $Cu₂O NCs/r-GO$ (curve a) and GO (curve b), respectively. The integral area ratio of the two bands (denoted as I_D/I_G) is 1.5 for $Cu₂O NCs/r-GO$, which is larger than that of $GO(0.82)$, indicating the efficient formation of r-GO [[29](#page-7-0)].

TGA analysis was performed to test the thermal stability and estimate the metal loading of $Cu₂O$ NCs/r-GO (Fig. S3, ESM). After heating from 25 to 800 °C, there is no weight left for GO because of the evaporation of water molecules, the removal of some oxygen-containing functional groups, and the decomposition of the carbon skeleton groups [\[30](#page-7-0)]. However, $Cu₂O$ NCs/r-GO exhibit much lower mass loss and faster achievement of the thermal plateau under the identical conditions, manifesting their better thermal stability and efficient reduction of GO to r-GO [[31](#page-7-0)]. Additionally, the mass loading of Cu₂O is 30.7 wt. $\%$.

Electrochemical properties of $Cu₂O$ NCs/r-GO/GCE

Firstly, Cu₂O NCs/r-GO modified electrode was constructed to examine their electrocatalytic properties, using GCE and pure r-GO/GCE as referenced electrodes. Electrochemical impedance spectroscopy was employed to investigate the electrical conductivity of the nanocomposites to study the synergistic effects between r -GO and Cu₂O. Figure 5 shows Nyquist plots obtained on GCE (curve a), r-GO/GCE (curve b), and $Cu₂O$ NCs/r-GO/GCE (curve c) in 0.1 M KCl containing 5.0 mM $[Fe(CN)_6]^{3-/4-}(1:1)$ solutions. The diameter of a semicircle of Cu₂O NCs/r-GO modified electrode in highfrequency zone is larger than those of r-GO/GCE or bare GCE under the identical conditions. It means the larger

Fig. 5 EIS of GCE (curve a), r-GO/GCE (curve b), $Cu₂O$ NCs/r-GO/GCE (curve c), and Cu₂O/GCE (curve d) in 0.1 M KCl containing 5.0 mmol L⁻¹ $[Fe(CN)_6]^{3-4}$ solutions. Frequency range: 0.01 to 1×10^5 Hz

resistance for electron transfer on Cu₂O NCs/r-GO modified electrode as compared to those of the other two cases. This is ascribed to the fact that the decorated $Cu₂O$ NCs as a semiconductor increase the resistance in $Cu₂O$ NCs/r-GO as contrast to that of r-GO, reflecting the decreased conductivity of $Cu₂O$ NCs/r-GO in the present work.

Herein, a non-enzymatic glucose sensor was constructed to investigate the electrocatalytic activity of $Cu₂O$ NCs/r-GO (Fig. [6A](#page-5-0), curve a) in 0.5 M NaOH containing 5.0 mM glucose, using individual r -GO (curve b) and Cu₂O NCs (curve c) as references. There is almost no anodic current observed at bare GCE. Clearly, Cu₂O NCs/r-GO modified electrode shows more negative onset potential and higher anodic currents in contrast to r -GO and Cu₂O NCs. It means the enhanced electrocatalytic performance of $Cu₂O$ NCs/r-GO. This is attributed to the fact that the present redox pair of $Cu(II)$ Cu(III) leads to the easy oxidation of glucose in strongly alkaline media [[32](#page-7-0)] and the fast electron transfer provided by r-GO [[33\]](#page-7-0).

Figure [6B](#page-5-0) illustrates a series of linear sweep voltammograms (LSVs) at $Cu₂O$ NCs/r-GO modified electrode, displaying the enhanced anodic peak currents with glucose in the range of $10.0 \sim 35.0$ mM. This observation confirms

Fig. 4 A FT-IR and **B** Raman
spectra of Cu₂O NCs/r-GO (*curve* a) and GO (curve b)

Fig. 6 A LSVs of Cu₂O NCs/r-GO (curve a), r-GO (curve b), and Cu₂O (curve c) modified electrodes in 0.5 M NaOH + 5.0 mM glucose at a scan rate of 50 mV s⁻¹. **B** LSVs of Cu₂O NCs/r-GO modified electrode without (curve a) and with 10.0, 15.0, 20.0, 25.0, 30.0, and 35.0 mM glucose (curve b–g)

the improved catalytic activity of $Cu₂O$ NCs/r-GO for glucose oxidation. The maximum catalytic current is observed at the applied potential of 600 mV upon the addition of glucose, which is applied for amperometric determination of glucose.

Determination of glucose

Typical amperometric curves of $Cu₂O$ NCs/r-GO modified electrode were recorded at 600 mV with successive addition of glucose into 0.5 M NaOH under constant stirring. As depicted in Fig. 7a, the amperometric response is less than 3 s, indicating rapid responses toward glucose, owing to the improved catalytic activity and fast electron transfer of $Cu₂O$ NCs/r-GO with the electrode [\[4](#page-6-0)]. The plot shown in Fig. 7b does not fit a conventional linear equation in the whole concentration range. The linear regression equations are I (μA) =1.6301+37.5518C (\mathbb{R}^2 =0.9950) and I (μA) $=$ 37.7961+23.0580 C (R²=0.9968) for determining glucose over the two linear concentration ranges of $0.005 \sim$ 2.095 mM and 2.595~9.595 mM, respectively. The detection limit is 1.0 μ M at the signal-to-noise ratio (3S/N), which is much lower than most previously reported values [[14](#page-6-0), [15,](#page-6-0) $34-37$ $34-37$ $34-37$]. It is notable that Cu₂O NCs/r-GO modified electrode

Fig. 7 a Amperometric i–t *curves* of $Cu₂O$ NCs/r-GO modified electrode after successive addition of glucose to 0.5 M NaOH at 600 mV. b The corresponding calibration plot

displays the enhanced catalytic performance for glucose sensing in terms of sensitivity and the limits of detection (LOD), compared to those of $Cu₂O$ -based materials previously reported (Table 1) [\[14,](#page-6-0) [15,](#page-6-0) [34](#page-7-0)–[37](#page-7-0)].

Repeatability and stability

The repeatability of the sensor used in the measurement was obtained by recording the responses to 5.0 mM glucose in 0.5 M NaOH. The relative standard deviations (RSDs) were

Table 1 Comparison of $Cu₂O$ -based sensors for glucose determination

Electrodes	Sensitivity $(\mu A \text{ mM}^{-1})$	Linear range (mM)	LOD (mM)	Ref.
Porous Cu ₂ O nanosphere-r-GO	185	$0.01 - 6.0$	0.05	[14]
Hollow Cu ₂ O nanocubes	52.5	$0.001 - 1.7$	0.87	[15]
$Cu-Cu2O NWs$	859.14	$0.1 - 12$	0.05	$\lceil 33 \rceil$
Cu/Cu ₂ O	33.63	$0.22 - 10.89$	0.05	$\left[34\right]$
Au/LDH-CNTs-G	141.22	$0.01 - 6.1$	0.001	$\left[35\right]$
$MCo-CFs (M=Cu,$ Fe, Ni, and Mn)	35.82	$0.02 - 11$	0.001	[36]
$Cu2O$ NCs/r-GO	37.55; 23.06	$0.005 - 2.095$; $2.595 - 9.595$	0.001	This work

calculated to be 2.0 % $(n=5)$ for Cu₂O NCs/r-GO/GCE. Additionally, to evaluate the electrode-to-electrode reproducibility, they were independently prepared using the same way and revealed an acceptable reproducibility with low RSD values of 1.4 % $(n=5)$. The stability of the sensor was examined by intermittently measuring the current responses to 5.0 mM glucose over a 35-day storage period (Fig. S4, ESM). The sensors were stored at 25 °C when not in use. The amperometric responses remained 95.2 % of their initial values after a storage period of 35 days, revealing long-term stability of the resulting sensor. The obtained sensor can be continuously detected without regenerating between each measurement.

Interference study and applications to human serum samples

To evaluate the selectivity of the sensor, potentially interfering biomolecules (i.e., UA, AA, and DA) were detected at $Cu₂O$ NCs/r-GO modified electrodes, which usually coexist with glucose in real samples such as human blood. The interference study was investigated in the presence of 5.0 mM glucose. The criterion for interference was a relative error of less than \pm 5 %. It was found that 500 folds of CaCl₂, MgCl₂, KCl, and NaCl, 50 folds of lactic acid and glutaric acid, 15 folds of UA and AA, 10 folds of DA and acetaminophen, and 5-fold concentrations of fructose and maltose had no interference in the determination of glucose. These results demonstrate good selectivity of the resulting sensor for glucose assay.

Glucose in human serum samples were determined by $Cu₂O$ NCs/r-GO modified electrode at 600 mV. As shown in Table S1 (ESM), the as-obtained results are consistent with those determined by a hospital-used blood sugar instrument (Accu-Chek Performa), and the RSD values range from 1.23 to 2.37 %. These results display that the nonenzymatic sensor can be employed practically for routine analysis of glucose in real biological samples.

Conclusions

In summary, a facile solvothermal method was developed for preparation of $Cu₂O$ NCs/r-GO, with the assistance of the triblock copolymer (Luviset^R Clear) and *L*-glutamic acid as a shape-directing agent and a reducing agent, respectively. The nanocomposites exhibit better catalytic activity than those of Cu2O nanoparticles and r-GO toward glucose oxidation. The sensor has wide linear range (0.005 to 9.595 mΜ), high sensitivity (23.058 μA mM⁻¹), and selectivity. The improved performances of $Cu₂O$ NCs/r-GO make it a promising candidate for the assay of glucose.

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