

Detection of cardiac biomarker proteins using a disposable based on a molecularly imprinted polymer grafted onto graphite

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Abstract A low-cost disposable was developed for rapid detection of the protein biomarker myoglobin (Myo) as a model analyte. A screen printed electrode was modified with a molecularly imprinted material grafted on a graphite support and incorporated in a matrix composed of poly(vinyl chloride) and the plasticizer *o*-nitrophenyloctyl ether. The protein-imprinted material (PIM) was produced by growing a reticulated polymer around a protein template. This is followed by radical polymerization of 4-styrenesulfonic acid, 2-aminoethyl methacrylate hydrochloride, and ethylene glycol dimethacrylate. The polymeric layer was then covalently bound to the graphitic support, and Myo was added during the imprinting stage to act as a template. Non-imprinted control materials (CM) were also prepared by omitting the Myo template. Morphological and structural analysis of PIM and CM by FTIR, Raman, and SEM/EDC microscopies confirmed the modification of the graphite support. The analytical

performance of the SPE was assessed by square wave voltammetry. The average limit of detection is 0.79 μg of Myo per mL, and the slope is $-0.193 \pm 0.006 \mu\text{A}$ per decade. The SPE-CM cannot detect such low levels of Myo but gives a linear response at above $7.2 \mu\text{g} \cdot \text{mL}^{-1}$, with a slope of $-0.719 \pm 0.02 \mu\text{A}$ per decade. Interference studies with hemoglobin, bovine serum albumin, creatinine, and sodium chloride demonstrated good selectivity for Myo. The method was successfully applied to the determination of Myo urine and is conceived to be a promising tool for screening Myo in point-of-care patients with ischemia.

Keywords Protein imprinting · Screen-printed electrodes · Cardiac biomarker · Myoglobin · Biosensor · Potentiometry

Introduction

Acute coronary syndrome (ACS) is a spectrum of acute myocardial ischemias ranging from unstable angina to acute myocardial infarction (AMI) [1, 2]. Patients suffering from AMI require a quick diagnosis from emergency medical assistance. The symptoms of chest pain, pressure, shortness of breath, and/or nausea are common to acute myocardial ischemia conditions [3]. The myocytes lose their membrane integrity in response to ischemia thereby releasing molecules into the peripheral circulation known as cardiac biomarkers. Screening cardiac biomarkers may be of great significance for a more accurate and quick diagnosis [4]. Myoglobin (Myo), cardiac troponin T (TnT), troponin I (TnI), and creatinine kinase MB (CK-MB) are the most relevant cardiac biomarkers for this purpose [5].

Serum Myo is a nonspecific biomarker that appears in the peripheral circulation as early as 1–2 h after cardiac damage. It precedes the elevation of TnT and CK-MB. Although Myo

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levels should not be used as the sole biomarker for the diagnosis of ACS, a normal Myo concentration within 2 h after presentation may exclude AMI [4, 6]. Myo cut-off levels range from 100 to 200 ng·mL⁻¹ [7, 8], reaching levels of 420–2000 ng·mL⁻¹ in serum [9] and 450 ng·mL⁻¹ in the urine [10]. Screening Myo in an emergency context is therefore important, and a suitable point-of-care (POC) assay would result in a reduced time to diagnosis and treatment [11].

Several methods have been described for Myo detection in physiological fluids. Some authors have developed immunoassay methods with fluorescence detection of high sensitivity [12–14]. Other immunoassay formats include fluorescence polarization, [15] resonance energy transfer, [16–18] and long-lived lanthanide emission [7, 18, 19], Chemiluminescence [20] and high-performance size-exclusion chromatography [21] have been also reported in the literature. Electrochemical assays based on immune-analysis [22], direct voltammetry in polyacrylamide hydrogels [23] and electrocatalytic reduction of H₂O₂ by myoglobin on carbon nanotube electrodes have also been reported [24]. Overall, the direct detection methods uses a labelled primary antibody to detect protein, displaying some disadvantages concerning stability, batch-to-batch (or clone-to-clone) variability, the requirement for refrigerated transport and storage and the high cost of producing antibodies

The design of synthetic biomimetic materials would therefore be an advantageous approach, and these are most commonly obtained by molecularly imprinted polymer (MIPs) technology. PIM assays have been reported to exhibit good properties in terms of selectivity, sensitivity and short response times for fast screening protein in biological samples. Moreira et al. [25] have reported various approaches to target detection of biomarkers for early PoC detection and the cardiac biomarkers such as troponin and myoglobin [25].

Even the design of Myo binding synthetic materials, also known as plastic antibodies, poses challenges [26]. Surface imprinting is a promising approach for this purpose, especially when coupled to electrochemical transduction. Several different strategies can be used to prepare such materials including: self-assembled monolayer deposition [26] ii) Thin-film polymer-formation using micro-contact imprinting [27] and iii) Designing the imprinted polymer films by grafting on a silicon surface [28]. In this work we describe the imprinting process of Myo on a graphite surface and its inclusion in PVC membranes. These membranes are permselective and allow charged proteins to diffuse through the membrane. The biorecognition element (PIM) was entrapped in the polymeric matrix and deposited on the transducer part of the sensor. In order to obtain the biosensor response, the analyte must penetrate through the membrane to PIM materials. The mass transport through the membrane is mainly driven by the

concentration gradients of species then, diffusive process is dominant [29].

Graphitic materials exhibit good structural, electronic, mechanical, optical, thermal, and chemical properties. The combination of high precision and speed, low cost of analysis, enhanced selectivity and sensitivity over a wide range of concentrations of the ion selective PVC membranes holds great promise for potential applications in biosensors [30]. Besides, measurement of membrane potential does not require the template to be extracted from the membrane, and ionic species do not have to diffuse through the membrane thus overcoming any size restrictions on the template compound [31].

For this purpose, the artificial antibody was produced by attaching Myo to oxidized surface and growing a reticulated polymer around it, by radical polymerization of 4-styrenesulfonic acid (SSA) and 2-aminoethyl methacrylate hydrochloride (AEHM), used as monomers, and ethylene glycol dimethacrylate (EDGMA) employed as crosslinker. The obtained particles, PIM, were used as electroactive materials on the PVC-Myo selective membranes. These membranes were prepared by dispersing the particles in *o*-nitrophenyloctyl ether (*o*NPOE), acting as plasticizer, and PVC and drop coated on carbon screen printed electrodes (SPE). The resulting biomimetic sensor was evaluated by electrochemical and surface characterization techniques and applied to the analysis of myoglobin in a biofluid such as urine.

Experimental

Apparatus

The electrochemical measurements were performed using an Autolab, PGSTAT302N potentiostat (Metrohm). The equipment had an integrated Frequency Response Analyzer (FRA) module and was controlled by NOVA 7.0 software. DRP-C220ATAu-SPEs were procured from DROPSSENS. Working and counter electrodes were made of carbon and the pseudo-reference electrode and electrical contacts made of silver. The diameter of the working electrode was 4 mm. For electrochemical assays the SPEs were placed in a switch box (DROPSSENS) to connect the electrical contacts of the Au-SPE with the electrical connections of the potentiostat.

Reagents

De-ionized (conductivity <0.1 μS·cm⁻¹) or ultrapure water was employed throughout this work and all chemicals were of analytical grade. Potassium hexacyanoferrate III, potassium hexacyanoferrate II trihydrate and sodium hydrogen phosphate dihydrate were obtained from Riedel-deHaen;

potassium dihydrogen phosphate from Panreac (<http://www.panreac.es/>); (4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid) (HEPES), from Sigma (<http://www.sigmaaldrich.com/sigma-aldrich/home.html>); 4-styrenesulfonic acid sodium salt (SSA) from Alfa Aesar (<http://www.alfa.com/pt/gh100w.pgm>), 2-Aminoethyl methacrylate hydrochloride from Aldrich (<http://www.sigmaaldrich.com/sigma-aldrich/home.html>); ethylene glycol dimethacrylate (EDGMA) and sodium dodecyl sulfate (SDS), sodium chloride, ethanol 99.5 %, Myo, *N*-hydroxysuccinimide (NHS), *N*-ethyl-*N*'-(3-dimethylaminopropyl) carbodiimide hydrochloride (EDAC), hemoglobin (Hb), bovine serum albumin (BSA), creatinine (Crea), sodium dodecylsulfate (SDS), *o*-nitrophenyloctyl ether (*o*NPOE) and poly(vinyl chloride) high molecular weight (PVC) from Fluka (<http://www.sigmaaldrich.com/sigma-aldrich/home.html>); styrene and glacial acetic acid from Carlo Erba (<http://www.carloerbareagents.com/default.aspx>); and benzoyl peroxide (BPO) from Himedia (<http://www.himedialabs.com/>).

Design of the plastic antibody on a graphite surface

Oxidation of graphite

Briefly, graphite powder was put placed in a solution of concentrated nitric acid and sulfuric acid (1:2 in volume) at a temperature of 80 °C. When the color of the liquid was transparent, the mixture was cooled down to room temperature and diluted with deionized water and left overnight. The pretreated graphite was then immersed in an ice bath, and potassium permanganate was added carefully. When the temperature stabilized, the vessel was placed into a water bath at a temperature of 38 °C and the mixture was stirred and left for 2 h. The vessel was then placed into an ice bath and deionized water was added drop wise to dilute the 10 times solution. Next, a solution of 30 % of H₂O₂ was added to this, leading to a color changed into brilliant yellow along with bubbling. The mixture was left overnight, to absorb the upper liquid and then filtered with HCl aqueous solution to eliminate residual metal ions, and finally washed with water.

Synthesis of biomimetic polymers

The carboxylic acid groups in the modified graphite were activated by mixing the material in EDAC (50 mmol·L⁻¹) and NHS (25 mmol·L⁻¹). The particles were then washed with ultrapure water and incubated in 1 × 10⁻⁴ mol·L⁻¹ Myo prepared in phosphate buffered saline (PBS) solution, pH 7.0, for 2.5 h, at 4 °C. Then, the material was thoroughly washed with PBS buffer and subsequently ultrapure water to remove any weakly adsorbed Myo (Fig. 1b).

Polymerization in the presence of Myo was carried out by adding 1 mL of 5.0 × 10⁻⁴ mol·L⁻¹ SSA and 5.0 × 10⁻⁴ mol·L⁻¹ of AEHM, 5.0 × 10⁻⁴ mol·L⁻¹ BPO and EDGMA 1.0 × 10⁻² mol·L⁻¹ solution (Fig. 1c and d). The polymerization was carried out at 38 °C, for 3 h (Fig. 1d). The material so obtained was thoroughly washed with deionized water and incubated in a solution of diluted acetic acid 10 % and SDS 10 % for 2 h (Fig. 1e). The imprinted sensor was washed and conditioned in PBS buffer, pH 7.4 during 1 h. Then the sensor was washed several times with ultrapure water.

Preparation of the selective membranes

The Myo-selective membranes were prepared by mixing 71 mg of PVC, 120 mg of plasticizer *o*-nitrophenyloctyl ether (*o*NPOE) and 2.5 mg of polymer, (either PIM or CM). The mixture was stirred until the PVC was well moistened, and then dispersed in 1.0 mL THF. These membranes were cast over the carbon layer on the working electrode area of the SPE. Membranes were left dry for 24 h and conditioned in a 1 × 10⁻⁵ mol·L⁻¹ Myo solution.

Electrochemical assays

Square wave voltammetry (SWV) measurements were conducted in a mixture of 5.0 mmol·L⁻¹ of [Fe(CN)₆]^{3-/4-} prepared in MES buffer, pH 5.0. In these, the potential was scanned from -0.7 to +0.7 V, at 30 mV·s⁻¹. In SWV studies potentials were changed from -0.7 to +0.7 V, at a 0.125 V·s⁻¹, corresponding to a frequency of 20 Hz and step height of 150 mV.

Surface analysis

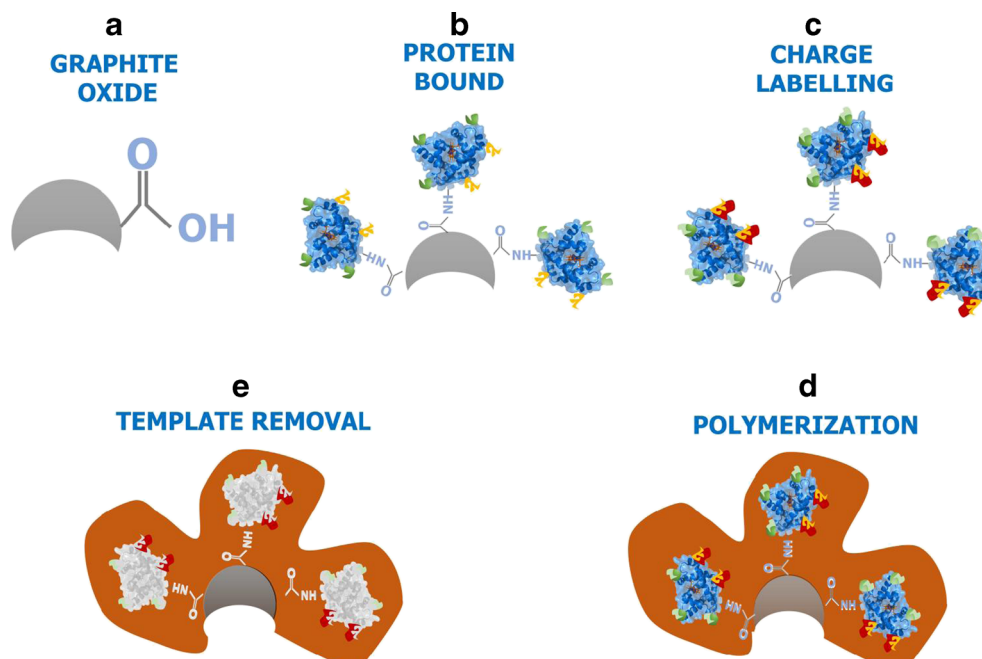
Fourier transform infrared spectroscopy analysis

The Fourier transform infrared spectroscopy (FTIR) spectra of graphite, graphite oxide and PIM particles were measured in a Nicolet 6700 FTIR spectrometer, coupled to a diamond ATR (attenuated total reflectance) prism. The spectra were collected under temperature and humidity control immediately after background correction. The number of scans was 32 for both sample and background.

Raman spectral analysis

Raman microscopy was carried out using a Labram 300 Jobin Yvon spectrometer, equipped with laser operating at 532 nm and with a power of 50 mW. Spectra were recorded as an extended scan. The laser beam was focused either with a 50× or a 100× Olympus objective lens. The laser power at the surface of the samples was varied using a set of neutral density filters (optical densities 0.3, 0.6, 1 and 2).

Fig. 1 The different stages of the molecular-imprinting polymer synthesis



Scanning electron microscope and energy dispersive spectrometer analysis

Scanning electron microscope (SEM) analysis was carried in a high resolution field emission gun scanning electron microscope (LEO Gemini 1525 FEGSEM, Oxford Instruments, UK). SEM images were collected of the dried PIM, PIM with template present, CM and graphitic materials, at room temperature and in low vacuum mode. The samples were metalized with chromium using a SEM sputtering system. Energy Dispersive Spectrometer (EDS) analysis was carried out in several points of each sample.

Selectivity

The interfering species tested were Hb, BSA, Crea and NaCl, prepared in MES buffer pH 4.5 in the concentration range 1.08–14.40 $\mu\text{g}\cdot\text{mL}^{-1}$. The measurements were performed by SWV.

Urine samples analysis

The performance of the plastic antibody was studied using urine samples spiked with Myo in the range 1.08–21.60 $\mu\text{g}\cdot\text{mL}^{-1}$. The urine was 1:10 diluted in MES buffer pH 4.5. The concentration of Myo studied was the same as the calibration curves (1.08–28.8 $\mu\text{g}\cdot\text{mL}^{-1}$).

Design of the biosensor

Several authors have been promoting the elegance of natural antibodies and trying to mimic it. One greatest advantage of

the artificial antibodies is the capability to create and innovate the respective molecular design. Characteristics such as stability, flexibility and permeability can be modulated according to the need. For the plastic antibodies' design several factors need to be considered such as easy preparation and low cost, stability and activity under wide operation conditions, selective and strong binding and guest binding in water.

In this work a novel molecular imprinted polymer on a graphite surface was developed for Myo detection in point of care. The first step of the tailoring process was the graphite oxidation with $\text{H}_2\text{SO}_4/\text{HNO}_3$ mixture and subsequent activation of the carboxylic acids with EDAC and NHS (Fig. 1a). These reagents catalyze the reaction between carboxylic and amine groups to form amide crosslinks. EDAC first activates a carboxylic group, which then reacts with NHS to produce an activate ester that undergoes nucleophilic substitution by amine groups on the protein. This means that the protein was covalently attached under physiological conditions (PBS buffer pH 7.4) supporting the integrity in terms of conformation and structure (Fig. 1b).

The imprinting approach involved self-assembly of the monomer units around the Myo (Fig. 1c) followed by a radical polymerization with the addition of an initiator and finally (Fig. 1d) template removal in acidic medium (Fig. 1e).

The monomeric arrangement was based on the effect of electrostatic charge density on the recognition of the Myo host to tailor the polymer, using AEMH as a positively charged monomer and SSA as a negatively charged monomer (Fig. 1c). The SSA monomer was effectively introduced as a strong negatively charged ion ($-\text{SO}_3^-$) in the polymerization step, enhancing the proton conductivity from the sulfonic acid group involved in the SSA on the plastic antibody surface

[32]. The simultaneous presence of two oppositely charged monomers in the pre-polymerization mixture resulted in imprinted with cavities that contain complementary and specific functional group orientation.

The time and monomer/cross-linker concentrations used in this stage were of particular importance because if the polymer layer is too thick it may entrap the protein irreversibly. The polymerization time was fewer than 3 h avoiding changes in the conformation of the protein. The temperature at which this polymerization reaction took place was also important because the imprinted conformation should match that of Myo in the samples, in this work the temperature was 38 °C near to the physiological conditions (Fig. 1d). Finally, the proteolysis of the attached protein was carried out by reaction with Acetic acid /SDS to empty the imprinted sites (Fig. 1e). Subsequently, several washing steps were performed to remove the released peptide fractions and enzyme from the sensory surface. Non-imprinted polymers were submitted to the same treatment to avoid any differences in comparing to the imprinted polymers.

Results and discussion

Surface analysis of the host-tailored polymers

Raman spectroscopic analysis

Raman spectroscopy is a widely used tool for the characterization of graphite and carbon nanotubes, especially considering the fact that conjugated and carbon-carbon double bonds lead to high Raman intensities.

Raman spectroscopy is used to identify carbon materials and for their characterization after chemical modification [33]. In Fig. 2 we present Raman spectra of Graphite (G), Graphite Oxide (GO), PIM/Myo and CM. In all samples, two prominent peaks are clearly visible, corresponding to the so-called G

peak, which corresponds to the Brillouin-zone-centered LO-phonon around 1590 cm^{-1} , and D peak, which corresponds to the double-resonance excitation of phonons close to the K point in the Brillouin zone around 1330 cm^{-1} [33]. The Raman spectra shows a notable fact, the G band peak is located at a higher frequency in GO than that in graphite (1603.3 vs 1578.5 cm^{-1}). This is a consequence of the extensive chemical modification on graphite surface in the oxidation process.

The intensity ratio (ID/IG) is characteristic of the extent of disorder present within in the material [33]. The ID/IG ratios of the G, GO, PIM/Myo, CM are respectively 0.65, 0.99, 1.10, 1.05 and reflecting the increase in disorder and the occurrence of the polymerization on the GO surface. Furthermore, there is only a little increase in intensity ratio for graphite imprinting and CM. By comparing the G-bands of GO and PIM/Myo, it is clear that G-band of GO–PIM occurs at 1595.4 cm^{-1} , which is downshifted by 8 cm^{-1} compared to that of GO. The Raman shifts of the G band for PIM/Myo, provide evidence for the charge transfer between the GO and PIM, which suggests a strong interaction between the PIM and the GO sensor. Another peak is present at 2665.7 and 2910 cm^{-1} in all sensors except in graphite. The differences in the intensity ratio demonstrated the occurrence of chemical reaction between the GO and the monomers suggesting the occurrence of the polymerization.

Fourier Transform Infrared Spectroscopy analysis

Structural changes in graphite were confirmed by FTIR spectra. The Fig. 3 presents FTIR spectra of Graphite (G), Graphite Oxide (GO) and PIM/Myo.

The bands at 2917 and 2850 cm^{-1} in the PIM material are assigned to the presence of the C–H stretch in the sp^3 hybridized carbons. It is probable that the peak at 2950 cm^{-1} assigned the presence of unsaturated carbon material (sp^2 hybridized carbon) within the PIM polymeric structure.

Fig. 2 Raman analysis of PIM, CM graphite oxide and graphite

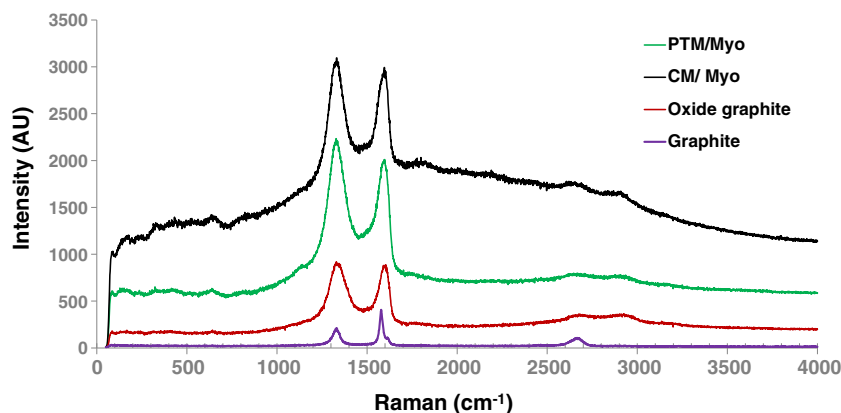
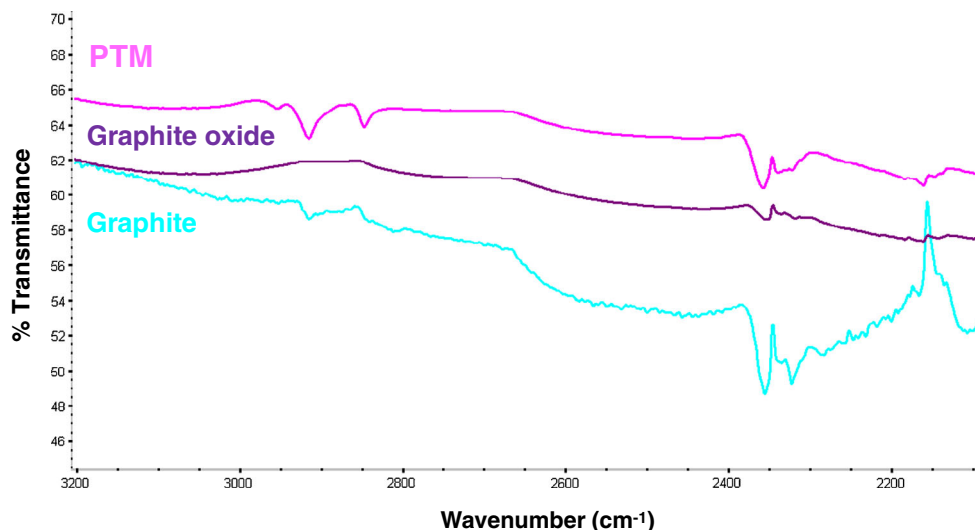


Fig. 3 FTIR spectra of graphite, oxidized graphite and PIM



Scanning electron microscope and energy dispersive spectrometer analysis

The occurrence of imprinting could not be proven by means of SEM, because the PIM and CM materials were similar. However, the chemical modification and imprinting effect of the surface was easily detected by EDC analysis. The chemical modification of the graphite introduced C and O in the overall structure (EDS spectra in Fig. 4). This was present in both PIM/Myo, PIM/Myo without template removal and CM biomaterials. PIM without template removal structures displayed, however, higher levels of these atoms, which probably resulted from prevailing protein structures sequestered inside the imprinted layer.

Analytical performance of the sensors

Square wave voltammetry measurements

Electrochemical assay provides analytical techniques characterized by instrumental simplicity, low cost and portability. This technique is sensitive, selective, rapid and readily applicable to analysis in most areas of analytical chemistry.

Square wave voltammetry (SWV) displayed several advantages such as high effective scan rate thus reducing the scan time, increased analytical sensitivity and relative insensitivity to the presence of dissolved oxygen [34]. Another advantage of SWV is the difference current is larger than either forward or reverse currents, meaning that the height of the peak is usually relatively easy to read, consequently increasing the accuracy.

To evaluate the interaction between PIM and CM sensor and the (Myo), the sensor surface was in contact with different concentrations of Myo. SWV measurements were performed in

MES buffer (pH 4.5) in $5.0 \text{ mmol} \cdot \text{L}^{-1}$ $[\text{Fe}(\text{CN})_6]^{3-}$ and $5.0 \text{ mmol} \cdot \text{L}^{-1}$ $[\text{Fe}(\text{CN})_6]^{4-}$. The SWV readings were carried out by the amplitude 0.15 V.

In SWV assays, Myo binding was shown by a decrease in the typical anodic peak current of redox probe solutions (Fig. 4). Higher Myo concentrations yielded smaller peaks. A linear pattern against $\text{Log}[\text{Myo}]$ was observed for concentrations down to $1.08 \text{ } \mu\text{g} \cdot \text{mL}^{-1}$ (Fig. 4) and with slope $-0.1735 \pm 0.784 \text{ } \mu\text{A decade}^{-1}$ (see Figs. 4 and 5) and maximum overall peak at 0.437 V.

CM polymer displayed linear behavior after $7.2 \text{ } \mu\text{g} \cdot \text{mL}^{-1}$ and the maximum overall peak were situated in a different potential value 0.532 V. These results indicated that the PIM/Myo showed better general performance compared with the CM sensor.

Selectivity study

The selectivity study is crucial for sensor performance evaluation. The interfering species tested were selected among those that may be found in biological fluids, such as Hb, BSA, Crea and NaCl in concentration range within $1.08\text{--}14.40 \text{ } \mu\text{g} \cdot \text{mL}^{-1}$. Each assay was conducted in a different PIM/Myo sensor, in order to avoid a cross contamination from previously adsorbed Myo or interfering compounds. It is important to highlight that the peaks currents observed belonged to the iron redox probe (present in the solution) and not to any other electrochemical active species at this region. So, the interfering study measured the effect of the interfering species upon this redox peak (shift and peak height).

As may be seen in Fig. 6, all species caused a right shift of the peak potential of the Myo reading. Significant interference could be observed at the maximum peak current of Myo assay, but if analytical readings are made at 0.24 V, these interfering

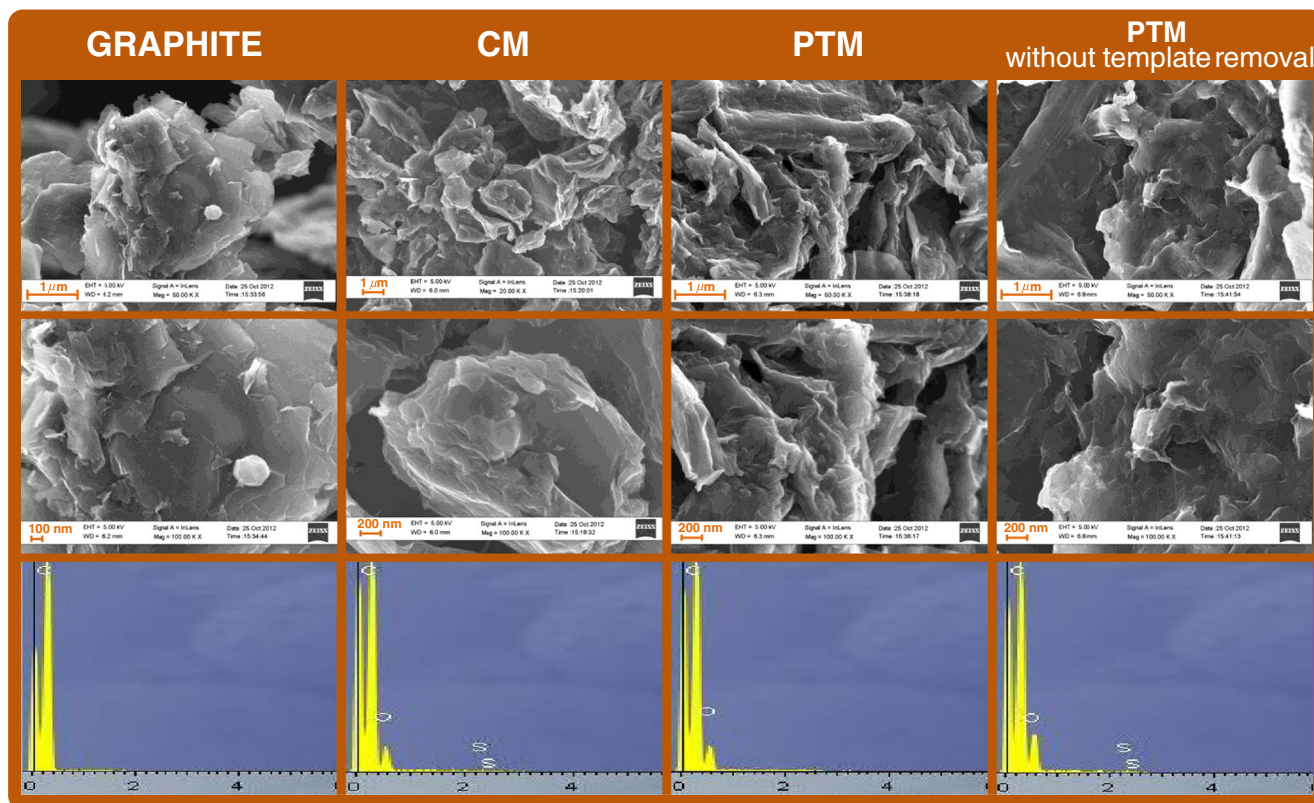


Fig. 4 SEM and EDC images of graphite, CM, PIM with protein and PIM

effects become negligible. The overall peak for Myo, Hb, Crea and BSA were respectively 0.437, 0.523, 0.296 and 0.559 V. We can conclude that the PIM/Myo displayed good features in terms of selectivity, due its different peak potential.

Myoglobin assay

The standard addition method was applied to determine Myo in spiked urine samples, ranging from 1.08-21.60 $\mu\text{g}\cdot\text{mL}^{-1}$

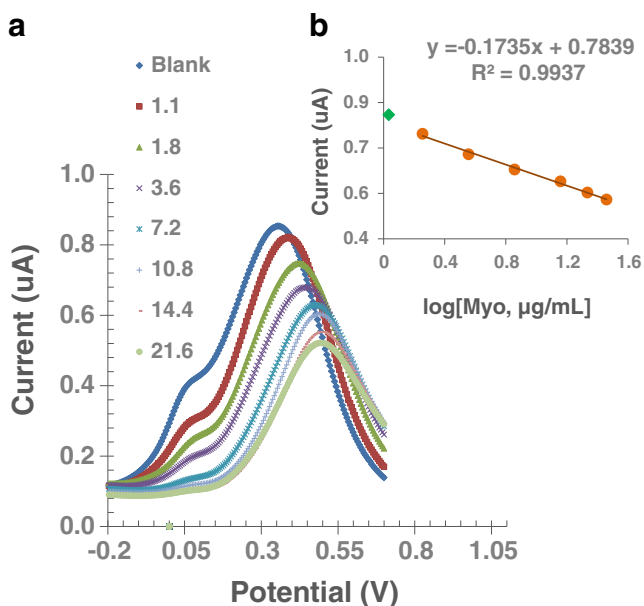


Fig. 5 SWV a of PIM material and the corresponding calibration curve b in 5.0 mM $[\text{Fe}(\text{CN})_6]^{3-}$ and 5.0 mM $[\text{Fe}(\text{CN})_6]^{4-}$, in MES buffer pH 4.5, with different concentrations of Myo

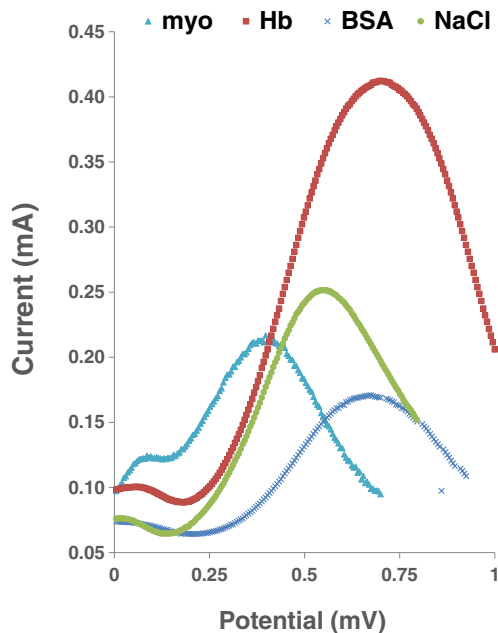


Fig. 6 SWV spectra of the different interfering species Myo, Hb, BSA and d NaCl with 3.6 $\mu\text{g}\cdot\text{mL}^{-1}$, respectively

Table 1 Figures of merit of comparable methods for determination of cardiac biomarker proteins

Transduction platform	Detection range (ng·mL ⁻¹)	Biorecognition element	Comments	Year	Ref.
Fluorescence	31.0	Antibody	pH 7.2 High price (antibody) Needs storage at 4 °C	2014	[1]
Fluorescence	10.0	Antibody	pH 7.2 High price (antibody) High price (indium-tin-oxide-coated glass) Needs storage at 4 °C	2014	[3]
Fluorescence	2.0	Antibody	pH 7.2 High price (antibody) Low price (paper) Need storage at 4 °C	2014	[4]
Faradaic, nanoparticles modified electrodes	17.8	Antibody	pH 7.2 High price (antibody) High price (gold nanoparticles) Need storage at 4 °C	2010	[5]
Faradaic,	5.0	Antibody	pH 7.2 High price (antibody) Low price (nanoparticles modified Fe grafite eletrodes) Need storage at 4 °C	2011	[6]
Polyaniline nanowires/ conductance	1.4	Aptamer	pH 7.4 High price (antibody) Need storage at 4 °C	2010	[7]
Electrochemical SPAM	580.0	Molecular imprinting	pH 5 Low price (plastic antibody) Low price (plastic antibody) Low price (gold/SPE) Storage at room temperature	2013	[8]
Electrochemical Potentiometry	217.0	Molecular imprinting	pH 4 Low price (plastic antibody) Low price (graphite electrodes) Storage at room temperature	2011	[9]
Electrochemical	225.0	Molecular imprinting	pH 5 Low price (plastic antibody) Low price (gold/SPE) Storage at room temperature	2013	[10]
Electrochemical	79.0	Molecular imprinting	pH 5 Low price (plastic antibody) Low price (gold/SPE) Storage at room temperature	–	Present work

Myo. The linear behavior was obtained in the range studied with slope $-0.464 \pm 0.05 \mu\text{A decade}^{-1}$ and LLLR $1.08 \mu\text{g} \cdot \text{mL}^{-1}$, ($Y = -0.45X + 0.989$), according to the calibration curve. These results pointed out the accuracy and the precision of the analytical data, suggesting that it may turn out a successful approach for screening AMI episodes in POC.

A good agreement was found between added and found amounts of Myo. Results of the electrochemical analysis conducted in steady state showed recoveries ranging 102.0–110.2 % with an average relative standard deviation of 1.8 %.

Conclusions

Surface imprinting of Myo tailored on graphite surface was successfully established by merging self-assembled monolayer and molecular imprinting technologies. There are several advantages out coming from this approach, including that there are no requirements of the covalent linkage of the analyte (at the imprinting stage) or/and biorecognition element (in the substrate surface), and the permselectivity characteristics of the plasticized membranes may enhance the selectivity performance.

An extensive comparison of the described method may be found in Table 1. In general, devices using imprinted materials are less expensive and much more stable (thermally and chemically) than immunoassays. Compared to previous devices, the present work offers a greater simplicity at the device construction, because new electrodes may be prepared in ~1 h,

after casting a membrane that has already been prepared (and stable at 4 °C for a minimum of 1 month).

SWV transduction offer suitable selectivity features for practical application and quick responses. Overall, these methods relying on imprinted PVC/PIM/Myo membranes applied on SPE device is simple, of low cost, precise, accurate and inexpensive, and may turn out an alternative method for screening Myo in POC.

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