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A magnetic metal-organic framework as a new sorbent for solid-phase extraction of copper(II), and its determination by electrothermal AAS

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Abstract We report on the synthesis of Fe₃O₄-functionalized metal-organic framework (m-MOF) composite from Zn(II) and 2-aminoterephthalic acid by a hydrothermal reaction. The magnetic composite is iso-reticular and was characterized by FTIR, X-ray diffraction, SEM, magnetization, and TGA. The m-MOF was then applied as a sorbent for the solid-phase extraction of trace levels of copper ions with subsequent quantification by electrothermal AAS. The amount of sorbent applied, the pH of the sample solution, extraction time, eluent concentration and volume, and desorption time were optimized. Under the optimum conditions, the enrichment factor is 50, and the sorption capacity of the material is 2.4 mg g^{-1} . The calibration plot is linear over the 0.1 to 10 μ g L⁻¹ Cu(II) concentration range, the relative standard deviation is 0.4 % at a level of 0.1 μ g L⁻¹ (for *n*=10), and the detection limit is as low as 73 ng L^{-1} . We consider this magnetic MOF composite to be a promising and highly efficient material for the preconcentration of metal ions.

Keywords $Fe_3O_4 \cdot IRMOF-3 \cdot Electrothermal atomic absorption spectrometry <math>\cdot$ Copper

Introduction

Solid-phase extraction (SPE) is becoming increasingly used as a sample pretreatment technology in the laboratory because of its merits of simplicity, low cost, low consumption of organic

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Y. Wang (⊠) · J. Xie · Y. Wu · X. Hu (⊠) College of Chemistry and Chemical Engineering, Yangzhou University, Yangzhou 225002, China e-mail: wangyzu@126.com e-mail: xyhu@yzu.edu.cn solvents, and high enrichment factor [1]. Recently, a new procedure for SPE, magnetic solid-phase extraction (MSPE), has been developed [2]. It is based on the use of magnetic or magnetically modified sorbents. In the MSPE method, the magnetic sorbents with adsorbed analytes can be easily separated from the sample matrix by the use of an external permanent magnet. Consequently, MSPE makes separation faster and easier, and avoids the time-consuming column passing or filtration operations encountered in SPE.

To date, many types of magnetic materials have been used for MSPE, such as magnetic carbon nanotubes [3], magnetic graphene nanocomposite [4], magnetic C18 microspheres [5], alumina-coated magnetite nanomaterials [6], and silica-coated magnetite microspheres [7]. Nevertheless, an important core task for enabling the widespread use of the MSPE method is to develop novel magnetic sorbents. For example, Zhou et al. reported the exploration of coordination polymers as a sorbent for SPE for the first time [8]. Cui et al. demonstrated the first example of the utilization of MOFs for SPME [9]. Yu et al. used a covalent bonding approach to fabricate a robust metalorganic framework ZIF-90 coating for SPME [10].

Metal-organic frameworks (MOFs), built up from organic linkers and inorganic connectors, have emerged as a new class of structured microporous material. The application of MOFs in hydrogen storage, gas sorption and separation, catalysis, and drug delivery has been reported. Recently, MOFs have been employed as stationary phases for gas chromatography [11–14], high-performance liquid chromatography [15–17] and capillary electrokinetic chromatography [18] to separate and determine organic pollutants in environmental matrices, such as polycyclic aromatic hydrocarbons (PAHs) [19], benzene homologues [20, 21], alkane isomers [22, 23] and organic phosphonates [24]. Furthermore, MOFs exhibit high activity and selectivity in adsorption, and there is growing interest particularly in the use of MOFs for SPME to adsorb and separate metal ions. Bagheri et al. utilized (Fe₃O₄-Pyridine)/ $Cu_3(BTC)_2$ for preconcentration of Pd(II) for detection by flame atomic absorption spectrometry [25]. Sohrabi et al. utilized magnetic MOF Fe₃O₄-pyridine nanocomposite as an adsorbent for separation and preconcentration of Cd(II) and Pb(II) ions [26]. Taghizadeh et al. prepared a novel magnetic MOF DHz nanocomposite for the preconcentration of heavy metal ions [27].

Yaghi's group have developed iso-reticular MOFs (IRMOFs) to demonstrate that the three-dimensional porous system can be functionalized with the groups -Br, $-NH_2$, $-OC_3H_7$, $-OC_5H_{11}$, $-C_2H_4$, and $-C_4H_4$ and its pore size can be expanded with the long molecular struts biphenyl, tetrahydropyrene, pyrene, and terphenyl [28]. Among IRMOFs, IRMOF-3 (Zn₄O(BDC-NH₂)₃) is of particular interest due to the presence of the free amine functionality in the linkers [29]. The amino group on the 1,4-benzenedicarboxylate allows chemical reactions to occur in the pores of MOF materials. However, the application of IRMOF-3 to the analysis of heavy metals has not been reported before.

In this work, an MSPE method with Fe₃O₄/IRMOF-3 particles as sorbent for the separation and preconcentration of copper was established before sensitive determination by electrothermal atomic absorption spectrometry (ETAAS). It has been recognized that copper is an indispensable micronutrient in human health, which has important effects on the blood, central nervous system and immune system. High intake of copper, however, can be fatal: a concentration of 1.3 ppm can cause immediate effects on the human body in the form of vomiting, diarrhea, and nausea [30]. Consequently, copper was chosen as a model analysis material. To the best of our knowledge, this may be the first report on the determination of copper by using MSPE with Fe₃O₄/IRMOF-3 as sorbent. All the important parameters of the method were studied and optimized to enhance its effectiveness. The method was applied in the analysis of environmental water samples and certified reference materials.

Experimental

Apparatus

A Zeenit700 atomic absorption spectrometer (Germany, http://www.analytik-jena.com) was employed for the determination of copper. A copper hollow cathode lamp was used as the radiation source at 324.8 nm. Measurements were carried out in the integrated absorbance (peak area) mode at 6 mA, using a spectral bandwidth of 0.2 nm. A Tensor 27 spectrometer (Germany, http://www.bruker-axs.com) was used to obtain Fourier transform infrared (FTIR) spectra. X-ray diffraction (XRD) patterns were recorded on a D8 Advance X-ray diffractometer (Germany, http://www.bruker-axs.

com) at room temperature. Field emission scanning electron micrographs (SEM) were obtained with a Hitachi S-4800 microscope (Japan, http://www.hitachi.com) at an acceleration voltage of 15 kV. The magnetic property was analyzed by using a vibrating sample magnetometer (America, http://www.ade.com). Thermogravimetric analysis (TGA) was performed by means of a Pyris 1 TGA (America, http://www.perkinelmer.com), under N₂ at a scan rate of 10 °C min⁻¹. A permanent magnet was used to enable the isolation of the analytes from the complicated matrix.

Reagents and materials

The stock standard solution containing 1,000 mg L^{-1} of Cu^{2+} was prepared by dissolving 1.9024 g Cu(NO₃)₂·3H₂O in 500 mL 0.5 mol L^{-1} nitric acid and working standard solutions were obtained by appropriate stepwise dilution of the stock standard solution. 2-Aminoterephthalic acid (R₃-BDC, America, http://www.sigma-aldrich.com); zinc nitrate, ferrous chloride, ferric chloride, ethanol, and dimethylformamide (DMF) were at least of analytical reagent grade and obtained from Sinopharm Chemical Reagent Co., Ltd (Shanghai, China, http://www.reagent168.cn) unless otherwise stated. Double de-ionized water (18 M Ω cm) was used throughout the experiments. The reference materials, which were stream sediment (GBW07303a) and soil (GBW07405), were purchased from BHH Biotechnology Co., Ltd (Beijing, China, http://www.gbw114.org). All containers were treated with 10 % HNO₃ for at least 24 h, rinsed with de-ionized water, and dried at room temperature before usage.

Preparation of IRMOF-3

The fabrication process was established using an established procedure with minor modifications [31]: Briefly, 0.5950 g (2 mmol) $Zn(NO_3)_2 \cdot 6H_2O$ in 5 mL DMF (solution A) and 0.0906 g (0.5 mmol) 2-aminoterephthalic acid in 5 mL DMF (solution B) were prepared. Solution A was then poured into solution B and mixed in a vial under ultrasound for 5 min. The mixed solution was poured into a 20 mL Teflon liner, placed in an autoclave, and heated to 383 K for 24 h. After reaction, the crystal was collected by filtration, washed with DMF (10 mL×3) and water (10 mL×3), and finally dried at 80 °C for 4 h.

Preparation of iron oxide

The magnetic particles were synthesized by the chemical coprecipitation method. FeCl₃·6H₂O (5.4 g) and FeCl₂· 4H₂O (2.0 g) were dissolved in a 6.0 mol L⁻¹ hydrochloric acid solution (25 mL), which was then sufficiently degassed with a nitrogen stream. After that, a 25 % (ν/ν) ammonium hydroxide solution (30 mL) was added to the solution with

vigorous stirring at 60 °C for 30 min under a nitrogen atmosphere. After cooling to ambient temperature, the resultant magnetic particles were attracted to the bottom by a magnet. The black products were washed several times sequentially with sufficient volumes of water and ethanol and then dried at 60 °C for 6 h.

Preparation of Fe₃O₄/IRMOF-3

For the synthesis of Fe₃O₄/IRMOF-3, 0.5 g R₃-BDC was dissolved in 40 mL DMF, and the mixture was poured into a 250 mL round-bottomed flask. Under magnetic stirring, Fe₃O₄ (70 mg) dissolved in 20 mL DMF was added and refluxed for a while at 90 °C. Next, 1.0 g Zn(NO₃)₂·6H₂O dissolved in 40 mL DMF were added to the above solution. After refluxing for 4 h, the product was collected by vacuum suction filtration, washed with DMF (10 mL×3) and water (10 mL×3), and finally dried at 80 °C for 4 h.

Sample extraction and preconcentration procedure

15 mg of Fe₃O₄/IRMOF-3 were added to 10 mL of sample solution containing the analyte ions and the pH of the solution was adjusted to 5 with 0.1 mol L^{-1} HNO₃ or 0.1 mol L^{-1} NaOH solutions. The mixed solution was then shaken at room temperature for 15 min to facilitate adsorption of Cu²⁺ onto the sorbent. Subsequently, the magnetic sorbent with adsorbed copper was separated from the mixture using a permanent magnet. The magnet was removed and 1 mL of 0.1 mol L^{-1} NaCl solution (pH=2)was added as eluent and stirred for 15 min. Finally, the magnet was used again to settle the magnetic sorbent, and the eluate was transferred into a test tube for subsequent ETAAS analysis.

Samples and sample pretreatment

The water samples were collected locally. The samples were filtered immediately after sampling, and stored at 4 °C. Before the experiment, 10 mL of the water samples were diluted to 100 mL with de-ionized water. Certified reference materials stream sediment (GBW07303a): 0.5000 g of stream sediment were mixed with 10 mL HCl and heated at 150 °C for 1 h. After preliminary decomposition, 5 mL HNO₃ and 3 mL HClO₄ were added and the mixtures were heated at 250 °C until the solution was nearly dried. After cooling, the residue was diluted to 50 mL with de-ionized water. Certified reference materials soil (GBW07405): 0.5000 g of soil is mixed with 10 mL HNO3 and 5 mL HClO4. After soaking for one night at room temperature, 1 mL H₂O₂ and 1 mL HCl were added and heated at 180 °C to dissolve the soil until the residues become milky. After cooling, 10 mL HNO₃ was added to make the solution clear and transparent. Finally, the content was diluted to 50 mL with de-ionized water.

Results and discussion

Characterization of the sorbent

FTIR spectrums of IRMOF-3 and Fe₃O₄/IRMOF-3 in the range of 4,000–400 cm⁻¹ were recorded using the KBr pellet method (Fig. S1, Electronic Supplementary Material, ESM). As for IRMOF-3, two peaks appeared at 3,126 and 3,317 cm⁻¹, corresponding to the presence of the amino groups. The two sharp bands at 1,560 and 1,388 cm⁻¹ correspond to the asymmetric and symmetric vibrations of carboxyl groups, respectively. The peak appearing at 1,232 cm⁻¹ can be assigned to C–N vibrations. The FTIR spectrum of Fe₃O₄/IRMOF-3 is similar to that of IRMOF-3, except for a stronger peak at 575 cm⁻¹, which can be assigned to the stretching of Fe–O bond, indicating that Fe₃O₄ was successfully introduced into IRMOF-3.

Figure 1 shows X-ray diffraction patterns of IRMOF-3 and $Fe_3O_4/IRMOF-3$. The XRD pattern (Fig. 1a) of IRMOF-3 is very similar to that reported in the literature [32]. When combined with Fe_3O_4 , the similar peak positions show that the structure of IRMOF-3 was preserved, although the peak intensities decreased. The decrease in the peak intensities after modification can be attributed to the decrease in the crystal-linity. Meanwhile, the change in the peak intensities indicated the successful synthesis of magnetic IRMOF-3.

The hysteresis curves measured at T=293 K for Fe₃O₄ and Fe₃O₄/IRMOF-3 are compared in Fig. 2. Neither coercivity nor remanence was observed, suggesting that the two particles are superparamagnetic. The saturation magnetization value was measured to be 80.92 emu g⁻¹ for Fe₃O₄ and 20.25 emu g⁻¹ for Fe₃O₄/IRMOF-3. Compared with Fe₃O₄, the magnetic strength of Fe₃O₄/IRMOF-3 decreased due to the presence of IRMOF-3. However, it was sufficient for magnetic separation with common magnets.

To investigate the surface morphology of IRMOF-3 and Fe₃O₄/IRMOF-3, the samples were characterized by SEM (Fig. S2, ESM). IRMOF-3 (Fig. S2-a) is obtained as well-formed cubic crystals with a rather smooth surface. In comparison with IRMOF-3, the SEM image of Fe₃O₄/IRMOF-3 (Fig. S2-b) keeps the crystal morphology, and the surface of Fe₃O₄/IRMOF-3 tends to be rougher after immobilization by Fe₃O₄. When combined with Fe₃O₄, the size of crystals became obviously smaller. It may be that the addition of Fe₃O₄ restricted the growth of the crystals, or it may be due to the different synthesis methods used in the cases of IRMOF-3 and Fe₃O₄/IRMOF-3.

Thermogravimetric analysis (TGA) of IRMOF-3 and Fe₃O₄/IRMOF-3 in nitrogen was also investigated (Fig. S3, ESM). IRMOF-3 shows an initial weight loss of about 6 wt% below 350 °C, due to the release of the remaining DMF, and a second weight loss of ca. 48 wt% due to the decomposition of organic linkers of the framework between 350 and 550 °C. As



Fig. 1 XRD patterns of IRMOF-3(a) and Fe₃O₄/IRMOF-3 (b)

shown in Fig. S3-b, $Fe_3O_4/IRMOF$ -3 has the same thermal behavior as IRMOF-3, indicating that the $Fe_3O_4/IRMOF$ -3 is stable up to 350 °C. Furthermore, the remaining amount of $Fe_3O_4/IRMOF$ -3 is larger than that of IRMOF-3, which due to the presence of Fe_3O_4 particles.

Optimization of instrument measurement conditions

Preliminary experiments indicated that the temperature program suggested by manufacturers could not be efficient for this method. As key parameters in the atomization process, the pyrolysis and the atomization temperatures were investigated in this work.

The optimal pyrolysis temperature needs to be high enough to cause the interfering components to volatilize completely. In addition, in order to avoid losing the analytes during pyrolysis, the pyrolysis temperature should be as low as possible. The pyrolysis temperature of the sample solutions containing 50 μ g L⁻¹ of copper ions was adjusted in the range of 500–900 °C (Fig. 3a). The absorbance increased as the temperature increased, and reached a maximum value at 800 °C. With temperatures higher than this, loss of analyte occurred. Therefore, a pyrolysis temperature of 800 °C was selected.



Fig. 2 VSM magnetization curves of Fe_3O_4 (a) and Fe_3O_4 /IRMOF-3 (b)

The atomization temperature of the sample solutions containing 50 μ g L⁻¹ of copper ions was studied in the range of 1,700–2,200 °C (Fig. 3b). With the temperature rise, the absorbance increased, and remained unchanged at 2,000– 2,200 °C. The lower atomization temperature is beneficial in terms of prolonging the service life of the graphite tube. Therefore, the best atomization temperature is 2,100 °C.

Optimization of the adsorption conditions

The pH of the solution is one of the most important parameters influencing the adsorption process of metal ions on the sorbent because of the protonation or deprotonation reaction for the sorbent and the hydrolysis reaction for the metal ions in the alkaline condition. Therefore, pH is the first parameter to be optimized. In this series of experiments, the solution pH was varied over the range of 2 to 7 with 0.1 mol L^{-1} HNO₃ or 0.1 mol L^{-1} NaOH solutions. As shown in Fig. 4, the adsorption efficiency increased with the increasing pH of the solution. Adsorption efficiency was calculated using the following equation:

Adsorption efficiency% =
$$\left(\left(C_a - C_b \right) \middle/ C_a \right) \times 100$$

where C_a and C_b are initial and final concentrations of copper ions in the solution, respectively.

The change in adsorption characteristics with solution pH may be more clearly understood from the relevant equations (see supplementary material) [33], which depict the major characteristic reactions that can take place at the solid-solution interface of Fe₃O₄/IRMOF-3. With increasing solution pH, it can be reasonably speculated that the adsorption of Cu²⁺ can occur in three ways: (1) coordination between $-NH_2$ from Fe₃O₄/IRMOF-3 and Cu²⁺; (2) electrostatic attraction between negative $-NH_2OH^-$ and positive Cu²⁺ (or CuOH⁺); (3) an ion exchange process between -mFeOH and Cu²⁺. When pH continues to increase, the precipitate of Cu(OH)₂ appears. Hence, the subsequent studies were carried out using a sample pH of 5.

Adsorption time is one of the main factors affecting the adsorption process. If the adsorption time is not sufficient, the sorbent could adsorb the metal ion incompletely. The effect of adsorption time on the adsorption efficiency of the solution of $1.0 \ \mu g \ L^{-1} \ Cu^{2+}$ in the range of 5–25 min was studied. As the adsorption time increased from 5 to 15 min, the adsorption efficiency for Cu²⁺ increased, and further prolonging of the adsorption time did not significantly increase the adsorption efficiency (Fig. S4, ESM). Thus, 15 min is the best adsorption time.

For solid phase extraction processes, a smaller amount of sorbent may result in incomplete adsorption, whereas a larger



0.25 b 0.20 Absorbance 0.15 0.10 1700 1800 1900 2000 2100 2200 Atomization temperature (°C)

Fig. 3 Pyrolysis (a) and atomization temperature (b) curves for copper. Conditions: 50 μ g L⁻¹ Cu²⁺ solution, pH=5; instrument conditions: wavelength: 324.8 nm, lamp current: 6 mA, spectral bandpass: 0.2 nm,

drying temperature: 90 °C, cleaning temperature: 2,200 °C, argon purge gas flow rate: 1,000 mL min⁻¹, determination mode: peak area

amount of sorbent increases the interference of the coexisting metal ions. Thus, the amount of sorbent is one of the most important parameters influencing the adsorption process of metal ions on the sorbent. In order to determine the optimal amount of sorbent, different amounts (5-20 mg) of the sorbent were added to the solution of 10 mL 1.0 μ g L⁻¹ Cu²⁺ to examine the effect on adsorption of Cu^{2+} ion on Fe₃O₄/ IRMOF-3 (Fig. S5, ESM). The experimental results show that the adsorption efficiency increased with an increasing amount of absorbent due to the increase of the amount of adsorption surface area and the number of functional groups involved in adsorption. When the sorbent was above 15 mg, the adsorption efficiency increased slowly and reached equilibrium. Therefore, 15 mg of Fe₃O₄/IRMOF-3 was selected for use in further studies.

From the pH study, it was found that the adsorption of Cu²⁺ on Fe₃O₄/IRMOF-3 at lower solution pH was negligible. This



Fig. 4 Effect of sample solution pH on the sorption efficiency of Cu²⁺ on Fe₃O₄/IRMOF-3. Conditions: 10 mL 1.0 μ g L⁻¹ Cu²⁺ solution, extraction time: 15 min, amount of sorbent: 15 mg. Desorption conditions: 1 mL of 0.1 mol L⁻¹ NaCl, pH=2, 15 min. Instrument conditions: as show in Fig. 3

suggested that desorption of Cu²⁺ on Fe₃O₄/IRMOF-3 was possible at lower solution pH. In this work, eluents (H₂SO₄, HCl, HNO₃, NaCl) were chosen as the desorption solvent. Results show that H₂SO₄, HCl and HNO₃ will decompose the structure of Fe₃O₄/IRMOF-3, but at different pH levels, NaCl increases recovery without any decomposition of Fe₃O₄/ IRMOF-3 structure. Therefore, 0.1 mol L^{-1} of NaCl solutions with different pH (1, 2 and 3) and volumes were chosen for further study. Based on measurements, 1 mL of 0.1 mol L^{-1} NaCl solution (pH=2) was sufficient to elute the analyte, and was further employed throughout this work.

Desorption time was evaluated in the range of 5-25 min. The experimental results indicated that a quantitative recovery of all analytes in 10 mL of sample solution was achieved when the stirring time was greater than 15 min for adsorption. Therefore, a desorption time of 15 min was used in subsequent experiments.

In the analysis of real samples, the maximum sample volume is an important factor in achieving a high enrichment factor. Under optimum conditions, the effect of sample volume on quantitative adsorption of Cu²⁺ was investigated in the range of 10-250 mL, and the total amount of Cu²⁺ was kept constant in 1.0 µg. Based on measurements, the recovery of Cu²⁺ was more than 95 % in the case of sample volumes up to 50 mL, and a decrease was observed with further increases in sample volume. With an elution volume of 1 mL, a theoretical enrichment factor of 50 was achieved by this method. However, for convenience, all the experiments were carried out with 10 mL of the aqueous phase.

Adsorption capacity is an important factor corresponding to the performance of the absorbent. In order to determine the adsorption capacity, 10 mg of Fe₃O₄/IRMOF-3 and 50 mL of 1 mg L^{-1} Cu²⁺ were equilibrated for 30 min. The maximum adsorption capacity has been found to be 2.4 mg g^{-1} . Under the same conditions, the adsorption capacity of Fe₃O₄ for

Table 1 Determination of Cu^{2+} in real water samples

Sample	Determined by present method ($\mu g L^{-1}$)	Added $(\mu g L^{-1})$	Found $(\mu g L^{-1})$	Recovery (%)
Tap water	3.85±0.27	1.00	4.83±0.31	102.0
		5.00	$8.78 {\pm} 0.37$	100.6
Lake water	2.08 ± 0.14	1.00	$3.02{\pm}0.18$	98.0
		5.00	6.98±0.33	101.8

 Cu^{2+} was 0.36 mg g⁻¹. The results showed that the adsorption on copper was the synergistic effect of IRMOF-3 and Fe₃O₄.

The interferences

For demonstrating the selectivity of the procedure developed, the interference of alkali, alkaline earth and heavy metals, which are common elements in environmental samples, was investigated under the above optimized conditions. For this purpose, a solution of 1.0 μ g L⁻¹ Cu²⁺ containing the corresponding interfering ion alone was prepared, and an ion was considered to have interfered when its presence produced a variation of more than ±5 %. The results showed that 300-fold concentrations of Cd^{2+} , Pb^{2+} , 200-fold concentrations of Cr^{2+} , Ag⁺, 100-fold concentrations of Co²⁺, Mn²⁺, Sn²⁺, 50-fold concentration of Ni²⁺, Zn²⁺ have no influence on the signals of Cu²⁺. The influence of some inorganic ions such as Na⁺, K⁺, Ca²⁺, Mg²⁺, Cl⁻, NO₃⁻, SO₄²⁻, CO₃²⁻, PO₄³⁻ could exist at concentrations of at least 10,000-fold. The above results indicated that the method developed is suitable for the analysis of Cu²⁺ in real samples and no further treatment or masking reagents are needed.

Performance and analytical applications

The analysis of a series of solutions containing 0.1, 0.5, 1.0, 5.0, and 10.0 μ g L⁻¹ Cu²⁺, respectively, gave a calibration function of A=0.0768C (µg L⁻¹)+0.0188, with a correlation coefficient of 0.9996. According to the IUPAC definition, the instrumental detection limit (3 σ) for Cu²⁺ was 0.073 µg L⁻¹. Relative standard deviation of the procedure determined by 10 replicates of samples containing 0.1 μ g L⁻¹ Cu²⁺ was calculated to be 0.4 %. Determination of Cu²⁺ using the magnetic solid-phase extraction procedure developed was compared with the other procedures reported in the literature [34–37] (Table S1, ESM). The results indicated that the adsorption capacity, limit of detection, and enrichment factor of the method developed is better than those of the other methods used for the separation and preconcentration of copper. Other advantages of this method are: (1) Magnetic IRMOF-3 composite could be easily produced with a larger surface area and highly active surface sites. (2) The presence of the free amine functionality in the linker enhances the selectivity of the new solid phase towards copper. (3) The separation of copperadsorbed magnetic adsorbent from the solution can easily be achieved via an external magnetic field. Therefore, satisfactory results can be achieved using lower volumes of sorbent. Regeneration is one of the key factors in evaluating the performance of the sorbent. It was found that the prepared Fe₃O₄/IRMOF-3 composite was relatively stable up to at least 10 adsorption-elution cycles without any obvious decrease in recovery. In order to demonstrate the performance of the method, tap water from laboratory and natural water from Slender West Lake were analyzed for the presence of copper. The results are given in Table 1 and indicate that the recoveries were reasonable for trace analysis, in a range of 98.0–102.0 %. To further establish the validity of the procedure, the method was applied to the determination of the content of Cu^{2+} in standard reference materials: GBW07303a (stream sediment) and GBW07405 (soil). The experimental results demonstrated good agreement with the certified values (Table 2). In conclusion, the analytical procedure is accurate and precise for copper determination in real samples.

Conclusion

In this research, a novel superparamagnetic Fe_3O_4 /IRMOF-3 sorbent was successfully synthesized and applied for separation and preconcentration of heavy metal ions before determination by ETAAS. Compared with traditional SPE methods, this SPE method has the following merits: (a) the abundant amine groups in IRMOF-3 demonstrate high adsorption capacity and extraction efficiency for the target analyte. (b) superparamagnetic Fe_3O_4 /IRMOF-3 sorbents can be easily collected and eluted with the help of a magnet. This research has not only developed a rapid and convenient MSPE method for the extraction of heavy metal ions from complicated samples, but has also extended the scope of applications of metalorganic framework.

Table 2 Validation of the method for the determination of Cu^{2^+} in certified reference materials

Sample	Certified value (µg g^{-1})	Found value (µg g^{-1})	RSD(%)
GBW07405	166±9	162±6	3.7
GBW07303a	202±7	205±5	2.4

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