## REVIEW

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# An update on the role of free radicals and antioxidant defense in human disease

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Abstract Mounting clinical and experimental evidence indicates that free radicals play important roles in many physiological and pathological conditions. The wider application of free radical measurement has increased awareness of functional implications of radical-induced impairment of the oxidative/antioxidative balance. In the following review, the role of oxygen free radicals in some human and experimental pathological conditions is described, with particular emphasis on the mechanisms by which they produce oxidative damage to lipids, proteins, and nucleic bases. The role of free radicals and the activation of the antioxidant systems in arteriosclerosis and ageing, diabetes, ischemia/reperfusion injury, ethanol intoxication, and liver steatosis is discussed. Therapeutic approaches to the use of antioxidants have been described and prospects for clinical use have been considered.

**Key words** Antioxidants · Free radicals · Glutathione · Oxidized proteins · Oxidative stress · Vitamin E

## Introduction

In the last few years there has been growing interest in the role played by oxidative reactions in human disease. A number of cell functions appear to be upregulated by the release of oxygen free radicals, such as DNA expression [1] and mitochondrial energy production [2]. Several experimental [3–5] and human pathological [6–8] conditions have been closely related to an overproduction of free radicals or to an impairment of the oxidative/antioxidative balance, which seems to be involved in the cell differentiation process, activation of specific metabolic pathways, and liver regeneration [9–11].

Oxygen free radicals are unstable compounds which exert their toxic effect by reacting with lipids, proteins,

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and nucleotides to produce oxidized compounds. Under particular conditions, some cofactors are involved in the mediation and amplification of these oxidative events. In the case of ethanol intoxication, for example, transition metals (Cu<sup>2+</sup>, Fe<sup>2+</sup>) are mobilized by free radicals from the cell storage pool and actively participate in the genesis of lipid and protein oxidation [12]. Other clinical conditions characterized by intracellular accumulation of iron (hemochromatosis) or copper (Wilson's disease) show pathogenic mechanisms of damage involving lipid and protein oxidation and also mitochondrial structures [13, 14]. Under similar experimental conditions, concomitant treatment with antioxidants (vitamin E) delays and lowers the extent of hepatic injury [15]. Most oxidative products are also intrinsically toxic and can cause damage to sites distant from the site of production after diffusion [16].

#### Mechanisms of oxidative injury

Lipid peroxidation was the first oxidative phenomenon to be investigated. It was initially documented during drug and ethanol intoxication [17, 18]. Peroxidation of lipids occurs when a pro-oxidant compound reacts with unsaturated fatty acids of biological membranes; their oxidative modification causes changes in the physical and chemical properties of the membranes, thus altering their fluidity and permeability, with swelling of intracellular organelles and increased risk of membrane rupture. This is the mechanism responsible for the hemolysis in alcoholics [19, 20]. Modification of the lipid redox state may also affect specific properties of the membranes, such as signal transduction and ion exchange [16]. Moreover, lipid peroxides are reported to be mediators of fibroblastic cell activation in the presence of inflammation [21] and remodelling after tissue damage [22].

More recently, the protein oxidation process has been extensively investigated and has been assigned an important role in ageing [23], cataractogenesis [24], and radiation cell injury [25]. In all these conditions an increased

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accumulation of carbonyl proteins (characterized by the oxidation of the carboxylic group into the carbonyl group) has been demonstrated. But, while in the case of ageing, the intracellular accumulation of carbonyl proteins seems to depend on a reduced capacity of the cell to degrade the oxidatively damaged proteins, in other conditions the accumulation of damaged proteins appears to be the consequence of an increased free radical production and/or decreased antioxidant protection. The sulfhydryl (-SH) group of the proteins, which is generally the first to be attacked and sacrificed when an increased production of pro-oxidant molecules occurs, is also involved in protein oxidation. The radical S-proteins react immediately with free glutathione (GSH) to form GS-S-proteins, which, in turn, are susceptible to be reconverted to the reduced form. Proteins containing an elevated number of -SH groups, namely sulfhydryl proteins (P-SH), play important roles in the cell (carriers, respiratory complexes, enzymes, histones, cytoskeleton) [26]. Thus, their oxidation may dramatically compromise cell function and survival.

The most recent to be investigated among the oxidative processes has been that of nucleic bases, and in particular of DNA. Increased levels of 8-hydroxy-deoxyguanosine have been found in aged animals [27] and in rats with chronic ethanol and iron intoxication [28]. These findings seem to have particular importance because of the association of the above conditions with DNA mutations and the high risk of cancer development. Oxidative alterations of mitochondria DNA bases and RNA bases are likely to be even more important, because of the lack of repair systems at these two levels. Damage occurring to these molecules may result in irreversible transcription or translation errors.

#### Arteriosclerosis and ageing

It is well known that high cholesterol levels associated with smoking and hypertension represent primary risk factors for cardiovascular disease, and that the pathogeneses of arteriosclerosis and ageing show several similarities. A major aspect is the oxidative modification of lowdensity lipoproteins (LDL), which promote arterial wall alterations [29]. Endothelial cells are able to oxidize LDL which successively accumulate in macrophages (foam cells) in the form of cholesterol and cholesterol esters [30]. Oxidized LDL can exert a strong chemotactic effect on monocytes and may also promote the activation of myofibroblasts [31]. Oxidized LDL may also activate circulating platelets with consequent thrombophilia effect [32]. The combination of these events can lead to partial or even complete occlusion of vessels, with a consequent dramatic reduction of blood supply to the relative tissues.

Under normal conditions, the reduced state of LDL is maintained by vitamin E [33], a circulating lipophilic compound which is also contained within the lipoprotein complex. Conditions such as smoking, diabetes, and hyperlipidemia, which alter the balance between unsaturated fatty acids and the vitamin E content within lipoproteins, expose LDL to the risk of oxidation.

A new interesting aspect of the free radical hypothesis of ageing is the oxidation of proteins. Increased levels of oxidized proteins have been detected in glial cells obtained from the brains of aged animals [34], and increased oxidation of the protein component of LDL has also been observed in the eldery [35]. The oxidation of cell and mitochondrial proteins in aged animals has been directly related to mitochondrial DNA oxidation and inversely related to the cell antioxidant capacity [36]. These observations clearly support the hypothesis of free radical-mediated intracellular oxidative damage in the ageing process. Furthermore, the increased oxidative damage of DNA bases observed in several cell lines derived from old people may correlate with the increased risk of malignancies in the elderly.

Finally, diet seems to play an important role in the protection against oxidative alterations. A Mediterranean diet, and especially its component olive oil, which contains large amounts of vitamins and antioxidant substances in an appropriate ratio with unsaturated fatty acids, has been shown to be protective against coronary heart disease [37], mainly by lowering the level of circulating oxidized LDL. Based on these observations, supplementation of diet with vitamin E and antioxidants may represent a preventive approach to degenerative diseases.

## Diabetes

Diabetes mellitus represents a typical chronic degenerative disease associated with an early onset of atherosclerotic alterations. The high incidence of arteriosclerosis in diabetics has been associated with increased intracellular oxidative stress: an increased accumulation of protein and lipid oxidative products has been noted in the tissues of diabetic subjects [38, 39]. This is related to the impaired antioxidant capacity of both serum and cells of diabetic patients [40, 41]. This imbalance is thought to be involved in the genesis of diabetic complications [42], and originates from impairment of the pentose phosphate pathway, which is due to the deficiency of insulin. This metabolic impairment results in a decreased availability of reduced substrates, such as reduced nicotinamide adenine dinucleotide phosphate (NADPH), and consequently of GSH, because of the decreased oxidized glutathione (GSSG) reductase activity, which utilizes NADPH [43]. The consequence is an increased susceptibility to oxidation of proteins and membrane lipids. All these events are of particular importance to blood vessels, where diabetics show a pattern of alterations very similar to those described in arteriosclerotic patients and the eldery [44].

The eye is also affected in diabetic subjects [45]. It is hypothesized that the retina, which is very rich in polyunsaturated fatty acids, is repeatedly exposed to hypoxic G. Vendemiale et al.: The role of free radicals and antioxidant defense in human disease

conditions and produces a large amount of lipid peroxides [46]. These, in turn, are able to migrate and extend the damage to protein structures, thus producing alterations of the protein redox status, with depletion of -SH groups and accumulation of carbonyl derivatives which are less soluble and consequently precipitate. In this way the damage extends from the retina to other ocular compartments, such as the vitreous humor and the lens, with the formation of opacities and cataract [47]. Based on recent investigations performed both in humans and animals by our group and others [46-48], it has been noted that the oxidative eye alterations in diabetic subjects are preceded by depletion of antioxidant agents, in particular GSH, vitamin E, and ascorbic acid. The antioxidant depletion and the oxidative alterations are related to the age of onset of diabetes and to the degree of glycometabolic control. Moreover, the presence of proliferative retinopathy seems to increase the ocular oxidative damage, as evidenced by measurements performed in the subretinal fluid from diabetic patients with retinal detachment in the presence or absence of proliferative retinopathy [49]. All these structural modifications seem to be consequential to the original oxidative alteration of the retina.

Other ocular diseases also show an impairment of the redox balance. Senile cataract, for example, is formed as a consequence of the accumulation of oxidative products, thus resembling myopic cataract [50]. Uveitis and episcleritis are typical inflammatory processes in which activated macrophages release oxygen free radicals with consequent oxidative damage to ocular structures [51].

Experimental treatment of the oxidative eye complications in diabetic patients by systemic administration or local application of antioxidants, such as GSH and vitamin E, did not produce appreciable results, probably because of the difficulties in reaching the inner eye compartments. However, the essential role of GSH in the maintenance of lens transparency is confirmed by the efficacy of GSH ester in the prevention of cataract in GSH-deprived new born mice [52], even if, at present, the best means of treating or avoiding the onset of diabetic oxidative eye complications in humans remains good metabolic control.

#### Ischemia/reperfusion

Reperfusion injury of post-ischemic tissue is the classic model for investigating the mechanisms of free radicalinduced oxidative cell damage [53]. Ischemia and reperfusion occur when the blood supply to an organ, or part of it, is suddenly interrupted due to an intrinsic (thrombotic occlusion) or extrinsic (surgical clamp) cause. Every tissue may theoretically be subjected to ischemia and reperfusion injury and the pathophysiological mechanisms of cell damage are substantially identical. Because of the social impact of coronary heart disease, myocardial ischemia was the first condition to be investigated and represents a typical clinical example. Reperfusion injury of the liver, which occurs during major surgery of this organ or after liver transplantation, is another. However, it shows some pathogenic differences compared with the heart.

It is well known that during the anoxic period of ischemia important metabolic changes occur in the cell. Firstly, anaerobiosis lowers the intracellular pH (acidosis) which, if particularly prolonged, may favor irreversible damage during reperfusion [54]. The longer the period of ischemia, the more serious the damage during reperfusion. Reperfusion is characterized by the release of molecules (i.e., nitric oxide), probably by endothelial cells [55], which are responsible for the activation of pro-inflammatory cells, which subsequently deliver cytokines (interleukins, tumor necrosis factor, etc.) and adhesion molecules (intercellular adhesion molecule-1) to the vascular compartment [56]. These cytokines are involved in the propagation of inflammation and are directly responsible for cell damage. During the early phase of reperfusion after warm ischemia, endothelial cells of the hepatic sinusoids release factors which activate the resident macrophages (Kupffer cells) which, when activated, are able to produce an enormous amount of oxygen free radicals [57]. At this time conspicuous amounts of GSH are oxidized and consumed in the vascular compartment to conteract free radical attack [58]. The intracellular GSH is consumed because it is used as a source of extracellular GSH [59]. After a few hours (second or late phase), circulating neutrophils are attracted and activated with consequent release of toxic and necrotizing factors [60]. Most of these events have also been described when livers are subjected to cold ischemia and subsequent reperfusion, as in the case of organ storage for transplantation [61].

#### Alcohol

Ethanol intoxication is one of the most widely investigated pathologies characterized by increased free radical production and oxidative alterations. Ethanol is an exogenous substance which is mainly metabolized in the liver by a cytosolic dehydrogenase and a microsomal enzymatic complex (P-450) [62]. These metabolic pathways transform ethanol into acetaldehyde, which is responsible for the formation of acetaldehyde-protein adducts [63] and for the main toxic effects attributed to ethanol [64]. Within hepatic cells, during ethanol metabolism, ethyl radicals [65] are also released, with the promotion of oxidative damage and GSH consumption [66].

There are several ways in which ethanol and its metabolic products damage the cell. A major one is the interference with some metabolic pathways (i.e., transulfuration) and the competition with other exogenous and endogenous compounds for the P-450 metabolic site, with enhancement of the toxic effects and production of metabolic disturbances and cell alterations. One of these is the fat infiltration of hepatocytes which results in increased lipid peroxidation because of the relative decrease in the antioxidant protection [67]. Acetaldehyde interferes with some mitochondrial functions, thus reducing the capacity for respiration and the production of reduced substrates (reduced nicotinamide adenine dinucleotide, NADH) and energy compounds (ATP) [68]. Mitochondria are selective targets of ethanol toxicity and are subjected to swelling with loss of cristae [69]. Recently, in both chronic and acute ethanol intoxication in the rat, a significant increase of protein and DNA oxidation has been demonstrated [28]. The former is certainly mediated by the Fenton reaction in which a metabolic activation of the iron storage pool represents a crucial mechanism for the amplification of the intracellular free radical damage [12]. The latter may be responsible for the increased risk of DNA mutation and cancer observed in alcoholics [70]. Peroxidative products of lipids have recently been indicated as promoters of Kupffer and Ito cell activation during chronic ethanol consumption, with consequent stimulation of hepatic fibrosis [21, 22]. Thus, interventions directed at conteracting the increased lipid peroxidation may also represent a protective measure against the onset of liver fibrosis.

Other areas are oxidatively damaged by ethanol. The stomach is the first organ to come into contact with ethanol after ingestion. The activity of alcohol dehydrogenase (ADH) in the stomach, an enzyme which is present in other tissues (pancreas, testis, brain) exposes these tissues to oxidative damage [71]. However, since the ADH content is lower in these organs than liver, this should expose them to less oxidative injury. The capacity to produce fatty acid ethyl esters by some kind of cells (pancreas, heart) represents an additional way by which ethanol induces cell injury [72]. Finally, ethanol also exerts its toxicity by stimulating the conversion of hypoxanthine to xanthine oxidase which, in turn, is able to release free radicals into the vasculature with consequent oxidative damage to circulating lipids and proteins [20]. The oxidative damage to erythrocyte membranes, where acetaldehyde-protein adducts have also been detected [73], results in a loss of fluidity and an increased risk of hemolysis.

## **Liver steatosis**

Steatosis of the liver is considered an innocent and fully reversible condition [74] characterized by intracellular accumulation of triglycerides in vesicles. Causes of liver steatosis include metabolic disorders, hypothyroidism, alcoholism, drug intoxication, and viral infections. The recent observation that transplantation of fatty liver is associated with a high degree of primary non-function, caused by an increased susceptibility to ischemia/reperfusion injury [75], has prompted a series of studies to investigate the pathological mechanisms of reduced cell resistance to ischemic insult and to verify the hepatocyte antioxidant capacity.

By the use of animal models (choline-deficient diet, choline/methionine-deficient diet, drug or ethanol intoxication), it has been shown that steatosis of the liver is characterized by an increased peroxidation of unsaturated lipids, regardless of the etiology of fat accumulation, with the formation of diffusible products. Fatty livers are reported to have small amounts of antioxidant compounds, in particular vitamin E, ascorbic acid, and GSH, which could render cell structures more susceptible to free radical attack.

Of even greater importance seems the observation that mitochondria isolated from fat-infiltrating hepatocytes show impaired energy production, serious depletion of antioxidants, protein oxidation, and morphological changes such as mitochondrial swelling. These morphological/functional alterations of mitochondria are exaggerated by fasting [76], which per se represents a prooxidant condition because the cell is starved of metabolic nutrients. Thus, fatty livers from fasted rats have smaller amounts of ATP and are further damaged by ischemia/reperfusion than normal livers from fasted animals. Pre-treatment with infusions of glucose seems to attenuate this oxidative damage, probably by replenishing the cellular glycogen, which is also decreased in fasted rats.

#### Therapeutic approaches with antioxidants

Several lines of evidence suggest that free radical scavengers may represent a preventive measure or a cure for a large number of human conditions. Free radical scavengers may be used in all pathological conditions characterized by an increased formation of pro-oxidant compounds and/or by depletion of antioxidative capacity. The former typically occurs during ischemia/reperfusion, the latter in prolonged food deprivation. Moreover, high-risk situations occur when both mechanisms operate, such as when paracetamol overdose follows alcohol intoxication [77], in which paracetamol metabolites find hepatic cells depleted of sulfhydryl groups, which have previously been consumed by ethanol free radicals.

In most of the cases reported above, when associated with standard treatments, replenishment of antioxidant reserve may represent an important strategy to conteract early and late complications of diabetes and arteriosclerosis, as well as to delay the onset of degenerative phenomena which accompany ageing and cancer. However, despite the large number of compounds showing antioxidant properties in vitro, only a few may be of interest for humans. An antioxidant to be used in man should fulfil the following criteria: (1) it should be a biological compound naturally present in animal tissues; (2) it should be active in the protection of both lipid and protein molecules; (3) it should have good bioavailability after oral and parenteral administration; (4) it should have a long half-life; (5) it should act in both the extracellular and intracellular spaces; (6) it should be able to cross intact cell membranes; (7) it shouldn't have a high cost.

Antioxidant substances which satisfy all these criteria are few. Vitamin E is a lipid-soluble molecule contained

in natural foods (olive oil, fish,...), however it does not act directly in the protection of proteins, and no studies have been carried out with parenteral administration. Vitamin E shows a certain efficacy when administered per os in supplemented diets [15], and is reported to have some modulating activity on the immune system. Exogenous GSH shows a poor absorption after oral administration [78], has a very short half life (approximately 2 min), and is not able to cross intact cell membranes. Superoxide dismutase is active against oxygen free radicals (hydrogen peroxide), but not against free radicals generated during xenobiotic metabolism, and has never been tested in human with pharmacokinetic evaluations. Some other compounds are very costly or are not biological substances. In order to improve the characteristics of the most-interesting antioxidants, they have been conjugated to carriers or administered in the form of prodrug. Some have been used under experimental conditions: N-acetylcysteine (NAC), glutathione mono- (GSHE) and diethylester, y-glutamylcysteinylethylester, S-acetyl and S-phenylacetyl-GSH.

Substrate molecules which may increase the synthesis of antioxidants have also been used. S-Adenosylmethionine, which is transformed within the cell in methionine, the amino acid precursor of cysteine, has been widely studied in many conditions of GSH depletion [79, 80]. NAC, which delivers free cysteine within the cell, is the universal antidote against paracetamol overdose [81].

Cysteine availability is the limiting step in the ex novo synthesis of GSH; however, administration of cysteine itself is not recommended, because it is rapidly transformed to cystine which is toxic and follows alternative routes. To overcome the block of GSH synthesis in conditions of poor ATP availability, GSH esters have been tested with promising results. GSHE has been reported to be a slow-release form of extracellular GSH under normal conditions [82]. Little is known about the ability of GSHE to cross membranes in conditions of oxidative stress, even if a protective effect of GSHE has been observed in livers subjected to ischemia/reperfusion and in acute pancreatitis [83, 84]. Diethyl esters of GSH disappear very soon from the circulation and release two molecules of ethanol per molecule of GSH, which shouldn't guarantee safety [85]. Hence at present, a cocktail of antioxidants, such as vitamin E, GSHE, and vitamin C, is the most-reasonable solution to the prevention or treatment of oxidative stress.

## References

- Suthanthiran M, Anderson ME, Sharma VK, Meister A. GSH regulates activation-dependent DNA synthesis in highly purified normal human T lymphocytes stimulated via the CD<sub>2</sub> and CD<sub>3</sub> antigens. Proc Natl Acad Sci USA 1990; 87:9943.
- Chance B, Sies H, Boveris A. Hydroperoxide metabolism in mammalian organ. Physiol Rev 1979; 59:527.
- Altomare E, Grattagliano I, Vendemiale G, Palmieri V, Palasciano G. Acute ethanol administration induces oxidative modifications in rat pancreatic tissue. Gut 1996; 38:742.

- Grattagliano I, Vendemiale G, Errico F, Bolognino A, Lilo F, Salerno MT, Altomare E. Chronic ethanol intake induces oxidative alterations in rat testis. J Appl Toxicol 1997; 17:307.
- Guerrieri F, Vendemiale G, Grattagliano I, Cocco T, Pellecchia G, Altomare E. Mitochondrial oxidative alterations following partial hepatectomy. Free Radic Biol Med 1999; 26:34.
- Fink R, Marjot DH, Cawood P. Increased free-radical activity in alcoholics. Lancet 1985; II:291.
- Altomare E, Vendemiale G, Albano O. Hepatic glutathione content in patients with alcoholic and non-alcoholic liver disease. Life Sci 1988; 43:991.
- 8. Gut A, Shiel N, Kay PM, Segal I, Braganza JM. Heightened free radical activity in blacks with chronic pancreatitis at Johannesburg, South Africa. Clin Chim Acta 1994; 230:189.
- De Capoa A, Ferraro M, Lavia P, Pelliccia F, Finazzi-Agro A. Silver staining of the nucleolus organizer regions (NOR) requires clusters of sulfhydryl groups. J Histochem Cytochem 1982; 30:908.
- Pascale R, Pirisi L, Daino L, Zanetti S, Satta A, Bartoli E, Feo F. Role of phosphatidylethanolamine methylation in the synthesis of phosphatidylcholine by hepatocytes isolated from choline-deficient rats. FEBS Lett 1982; 145:293.
- Vendemiale G, Guerrieri F, Grattagliano I, Didonna D, Muolo L, Altomare E. Mitochondrial oxidative phosphorylation and intracellular glutathione compartmentation during rat liver regeneration. Hepatology 1995; 21:1450.
- Shaw S, Jayatilleke E, Lieber CS. Lipid peroxidation as a mechanism of alcoholic liver injury: role of iron mobilization and microsomal induction. Alcohol 1988; 5:135.
- Britton RS, O'Neill R, Bacon BR. Hepatic mitochondrial malondialdehyde metabolism in rats with chronic iron overload. Hepatology 1990; 11:93.
- 14. Sokol RJ, Twedt D, McKim JM, Devereaux MW, Karrer FM, Kam I, Steigman G von, Narkewicz MR, Bacon BR, Britton RS, Neuschwander-Tetri BA. Oxidant injury to hepatic mitochondria in patients with Wilson's disease and Bedlington terriers with copper toxicosis. Gastroenterology 1994; 107:1788.
- Sokol RJ, Devereaux M, Mierau GW, Hambige KM, Shikes RH. Oxidant injury to hepatic mitochondrial lipids in rats with dietary copper overload. Modification by vitamin E deficiency. Gastroenterology 1990; 99:1061.
- Esterbauer H, Schaur KG, Zollner H. Chemistry and biochemistry of 4-hydroxynonenal, malondialdehyde and related aldehydes. Free Radic Biol Med 1991; 11:81.
- Fairhurst S, Barber DJ, Clark B, Horton AA. Studies on paracetamol-induced lipid peroxidation. Toxicology 1982; 23:249.
- Shaw S, Jayatilleke E, Ross WA, Gordon ER, Lieber CS. Ethanol-induced lipid peroxidation: potentiation by long-term alcohol feeding and attenuation by methionine. J Lab Clin Med 1981; 98:417.
- Fujii S, Dale GL, Beutler E. GSH-dependent protection against oxidative damage of the human red cell membrane. Blood 1984; 63:1096.
- Grattagliano I, Vendemiale G, Sabbà C, Buonamico P, Altomare E. Oxidation of circulating proteins in alcoholics: role of acetaldehyde and xanthine oxidase. J Hepatol 1996; 25:28.
- 21. Casini A, Ceni E, Salzano R, Biondi P, Parola M, Galli A, Foschi M, Caligiuri A, Pinzani M, Surrenti C. Neutrophil-derived superoxide anion induces lipid peroxidation and stimulates collagen synthesis in human hepatic stellate cells: role of nitric oxide. Hepatology 1997; 25:361.
- 22. Pinzani M, Marra F, Čarloni V. Signal transduction in hepatic stellate cells. Liver 1998; 18:2.
- Oliver CN, Ahn BW, Moerman EJ, Goldstein S, Stadtman ER. Age-related changes in oxidized proteins. J Biol Chem 1987; 262:5488.
- Augusteyn RC. Protein modification in cataract: possible mechanisms. In: Duncan G, ed. Mechanisms of cataract formation in the human lens. New York: Academic Press; 1981:71–115.
- 25. Guerrieri F, Vendemiale G, Turturro N, Fratello A, Furio A, Muolo L, Grattagliano I, Papa S. Alteration of mitochondrial

F0F1 ATP synthase during aging. Possible involvement of oxygen free radicals. Ann N Y Acad Sci 1996; 786:62.

- 26. Ajiboye R, Harding JJ. The non enzymatic glycosylation of bovine lens proteins by glucosamine and its inhibition by aspirin, ibuprofen and glutathione. Exp Eye Res 1989; 49:31.
- 27. Shigenaga HK, Hagen TM, Ames HN. Oxidative damage and mitochondrial decay in aging. Proc Natl Acad Sci USA 1994; 91:10771.
- 28. Wieland P, Lauterburg BH. Oxidation of mitochondrial proteins and DNA following administration of ethanol. Biochem Biophys Res Commun 1995; 213:815.
- 29. Rengstrom J, Nilsson J, Tornvall P, Landou C, Hamsten A. Susceptibility to low-density lipoprotein oxidation and coronary atherosclerosis in man. Lancet 1992; 339:1183.
- 30. Steinbrecher UP, Parathasarathy S, Leake DS, Witztum JL, Steinberg D. Modification of low density lipoprotein by endothelial cells involves lipid peroxidation and degradation of low density lipoprotein phospholipids. Proc Natl Acad Sci USA 1984; 81:3883.
- 31. Berliner JA, Territo MC, Sevanian A, Ramin S, Kim JA, Bamshad B, Esterson M, Fogelman AM. Minimally modified low density lipoprotein stimulates monocyte endothelial interactions. J Clin Invest 1990; 85:1260.
- 32. Wanatabe J, Umeda F, Wakasugi H, Ibayashi H. Effect of vitamin E on platelet aggregation in diabetes mellitus. Thromb Haemost 1984; 51:313.
- 33. Jessup W, Rankin SM, De Whalley CV, Hoult JRS, Scott J, Leake DS. Alpha-tocopherol consumption during low-density lipoprotein oxidation. Biochem J 1990; 265:399.
- 34. Carney JM, Starke-Reed PE, Oliver CN, Landum RW, Cheng MS, Wu JF, Floyd RA. Reversal of age-related increase in brain protein oxidation, decrease in enzyme activity, and loss in temporal and spatial memory by chronic administration of the spin-trapping compound N-tert-butyl-α-phenylnitrone. Proc Natl Acad Sci USA 1991;88:3633.
- 35. Wiztum JL, Steinberg D. Role of oxidized low density lipoprotein in atherogenesis. J Clin Invest 1991; 88:1785.
- 36. De La Asuncion JG, Millan A, Pla R, Bruseghini L, Esteras A, Pallardo FV, Sastre J, Vina J. Mitochondrial glutathione oxidation correlates with age-associated oxidative damage to mitochondrial DNA. FASEB J 1996; 10:333.
- 37. Gey F, Puska P, Jordan P, Moser UK. Inverse correlation between plasma vitamin E and mortality from ischemic heart disease in cross-cultural epidemiology. Am J Clin Nutr 1991; 53:326S.
- 38. Altomare E, Vendemiale G, Procacci V, Chicco D, Cirelli F. Increased lipid peroxidation in type-2 poorly controlled diabetic patients. Diabetes Metab 1992; 18:264.
- 39. Altomare E, Vendemiale G, Grattagliano I, Angelini P, Micelli-Ferrari T, Cardia L. Human diabetic cataract: role of lipid peroxidation. Diabetes Metab 1995; 21:173.
- 40. Maxwell SRJ, Thomason H, Sandler D, Legneu C, Baxter MA, Thorpe GHG, Jones AF, Barnett AH. Antioxidant status in patients with uncomplicated insulin-dependent and non-insulin dependent diabetes mellitus. Eur J Clin Invest 1997; 27:484.
- 41. Lyon TJ, Li W, Wells-Knecht MC, Jokl R. Toxicity of mildly modified low density lipoproteins to cultured retinal capillary endothelial cells and pericytes. Diabetes 1994; 43:1090.
- 42. Baynes JW. Role of oxidative stress in development of complications in diabetes. Diabetes 1991; 40:405.
- 43. Cheng M, Gonzales G. The effect of high glucose and oxidative stress on lens metabolism, aldose reductase and senile cataractogenesis. Metabolism 1986; 35:10.
- 44. Ceriello A, Pirisi M. Is oxidative stress the missing link between insulin resistance and atherosclerosis? Diabetologia 1996; 39:357.
- 45. Altomare E, Grattagliano I, Vendemiale G, Micelli-Ferrari, Signorile A, Cardia L. Oxidative protein damage in human diabetic eye: evidence for a retinal participation. Eur J Clin Invest 1997; 27:141.
- 46. Williamson JR, Chang K, Frangos M, Hasan KS, Ido Y, Kawamura T, Nyengaard JR, Van Den Enden M, Kilo C, Tilton

RG. Hyperglycaemic pseudohypoxia and diabetic complications. Diabetes 1993; 42:801.

- 47. Vendemiale G, Grattagliano I, Micelli-Ferrari T, Cardia L, Altomare E. Abnormal redox status in the lens and vitreous of diabetic subjects. Diabetologia 1996; 39:1239.
- 48. Babizhaev MA, Deev AJ. Lens opacity induced by lipid peroxidation products as a model of cataract associated with retinal disease. Biochem Biophys Acta 1989; 1004:124.
- 49. Grattagliano I, Vendemiale G, Boscia F, Micelli-Ferrari T, Cardia L, Altomare E. Oxidative retinal products and ocular damages in diabetic patients. Free Radic Biol Med 1998; 25:369
- 50. Micelli-Ferrari T, Vendemiale G, Grattagliano I, Boscia F, Arnese L, Altomare E, Cardia L. Role of lipid peroxidation in the pathogenesis of myopic and senile cataract. Br J Ophthalmol 1996; 80:840.
- 51. Rao NA, Sevanian A, Fernandez MAS, Romero JL, Faure JP, Kozak Y de, Till GO, Marak GE. Role of oxygen radicals in experimental allergic uveitis. Invest Ophthalmol Vis Sci 1987; 28:886.
- 52. Martensson J, Steinherz R, Jain A, Meister A. GSH ester prevents buthionine sulfoximine-induced cataracts and lens epithelial cell damage. Proc Natl Acad Sci USA 1989; 86:8728.
- 53. McCord JM. Oxygen-derived free radicals in postischemic tissue injury. N Engl J Med 1985; 312:159.
- 54. Kobayashi H, Nonami T, Kurokawa T, Marada A, Nakao A, Sugiyama S, Ozawa T, Takagi H. Changes in the glutathione redox system during ischemia and reperfusion in rat liver. Scand J Gastroenterol 1992; 27:711.
- 55. Samarasinghe DA, Farrell GC. The central role of sinusoidal endothelial cells in hepatic hypoxia-reoxygenation injury in the rat. Hepatology 1996; 24:1230.
- 56. Kuzume M, Nakano H, Yamaguchi M, Matsumiya A, Shimokohbe G, Kitamura N, Nagasaki H, Kumada K. A monoclonal antibody against ICAM-1 suppresses hepatic ischemia-reperfusion injury in rats. Eur Surg Res 1997; 27:93.
- 57. Jaeschke H, Bautista AP, Spolarics Z, Spitzer JJ. Superoxide generation by Kupffer cells and priming of neutrophils during reperfusion after hepatic ischemia. Free Radic Res Commun 1991; 15:277.
- 58. Jaeschke H, Farood A. Neutrophils and Kupffer cell-induced oxidant stress and ischemia-reperfusion injury in rat liver. Am J Physiol 1991; 260:G355.
- 59. Jaeschke H, Smith CV, Mitchell JR. Reactive oxygen species during ischemia-reflow injury in isolated perfused rat liver. J Clin Invest 1988; 81:1240.
- 60. Bilzer M, Lauterburg BH. Oxidant stress and potentiation of ischemia/reperfusion injury to the perfused rat liver by human polymorphonuclear leukocytes. J Hepatol 1994; 20:473
- 61. Shibuya H, Ohkohchi N, Seya K, Satomi S. Kupffer cells generate superoxide anions and modulate reperfusion injury in rat livers after cold preservation. Hepatology 1997; 25:356.
- 62. Lieber CS. Alcohol, protein metabolism and liver injury. Gastroenterology 1980; 79:373. 63. Stevens VJ, Fantl WJ, Newman BC. Acetaldehyde adducts
- with hemoglobin. J Clin Invest 1981; 67:361.
- 64. Kera Y, Ohbora Y, Komura S. The metabolism of acetaldehyde and not acetaldehyde itself is responsible for in vivo ethanol-induced lipid peroxidation in rats. Biochem Pharmacol 1988; 37:363.
- 65. Albano E, Tomasi A, Goria-Gatti L, Dianzani MU. Spin trapping of free radical species produced during the microsomal metabolism of ethanol. Chem Biol Interact 1988; 65:223
- Vendemiale G, Grattagliano I, Signorile A, Altomare E. Ethanol-induced changes of intracellular thiol compartmentation and protein redox status in the rat liver: effect of tauroursodeoxycholate. J Hepatol 1998; 28:46.
- 67. Videla LA, Valenzuela A. Alcohol ingestion, liver GSH and lipoperoxidation: metabolic interrelations and pathological implications. Life Sci 1982; 31:2395.
- 68. Lauterburg BH, Bilzer M. Mechanisms of acetaldehyde hepatotoxicity. J Hepatol 1988; 7:384.

- G. Vendemiale et al.: The role of free radicals and antioxidant defense in human disease
- 69. Uchida T, Kronborg I, Peters RL. Giant mitochondria in the alcoholic liver diseases. Their identification, frequency and pathologic significance. Liver 1984; 4:29.
- Mufti SI, Eskelson CD, Odeleye OE, Nachiappan V. Alcoholassociated generation of oxygen free radicals and tumor promotion. Alcohol Alcohol 1993; 28:621.
- 71. Altomare E, Grattagliano I, Didonna D, Gentile A, Vendemiale G. Gastric and intestinal ethanol toxicity in the rat. Effect on glutathione level and the role of alcohol and acetaldehyde metabolisms. Ital J Gastroenterol Hepatol 1998; 90:86.
- Bora PS, Bora NS, Wu XL, Lange LG. Molecular cloning, sequencing, and expression of human myocardial fatty acid ethyl ester synthase-III cDNA. J Biol Chem 1991; 26:16774.
- Clot P, Tabone M, Aricò S, Albano E. Monitoring oxidative damage in patients with liver cirrhosis and different daily alcohol intake. Gut 1994; 35:1637.
- 74. Bellentani S, Tiribelli C, Saccoccio G, Sodde M, Fratti N, De Martin C, Cristianini G and the Dionysos Study Group. Prevalence of chronic liver disease in the general population of Northern Italy: the Dionysos Study. Hepatology 1994; 20: 1442.
- 75. Strasberg SM, Howard TK, Molmenti EP, Hertl M. Selecting the donor liver: risk factors for poor function after orthotopic liver transplantation. Hepatology 1994; 20:829.
- 76. Caraceni P, Grattagliano I, Domenicali M, Nardo B, Dall'Agata M, Simoncini M, Vendemiale G, Altomare E, Cavallari A, Trevisani F, Bernardi M. Effect of fasting on mitochondrial injury in a rat model of fatty liver (abstract). Hepatology 1998; 28:324A

- 77. Lauterburg BH, Velez MA. GSH deficiency in alcoholics: risk factor for paracetamol hepatotoxicity. Gut 1988; 29:1153.
- Grattagliano I, Wieland P, Schranz C, Lauterburg BH. Effect of oral GSH monoethyl ester and GSH on circulating and hepatic sulfhydryls in the rat. Pharmacol Toxicol 1994; 75:343.
- 79. Lieber CS, Casini A, DeCarli LM, Kim CL, Lowe K, Sasaki R, Leo MA. S-Adenosyl-methionine attenuates alcohol-induced liver injury in the baboon. Hepatology 1990; 11:165.
- Frezza M, Surrenti C, Mantillo G, Fiaccadori F, Bortolini M, DiPadova C. Oral S-Adenosylmethionine in the symptomatic treatment of intrahepatic cholestasis: a doubleblind, placebocontrolled study. Gastroenterology 1990; 99:211.
- Prescott LF, Ballantyne A, Park J, Adriaenssens P, Proudfoot AT. Treatment of paracetamol (acetaminophen) poisoning with N-acetylcysteine. Lancet 1977; II:432.
- 82. Grattagliano I, Wieland P, Schranz C, Lauterburg BH. Disposition of glutathione monoethyl ester in the rat: glutathione ester is a slow release form of extracellular glutathione. J Pharmacol Exp Ther 1995; 272:484.
- Grattagliano I, Lauterburg BH. Reperfusion injury of the liver: role of mitochondria and protection by glutathione ester. J Surg Res 1999; in press.
- Neuschwander-Tyetri B, Ferrell LD, Sukhabote RJ, Grendell JH. Glutathione monoethyl ester ameliorates caerulein-induced pancreatitis in the mouse. J Clin Invest 1992; 89:109.
- Levy ÉJ, Anderson ME, Meister A. Transport of glutathione diethyl ester into human cells. Proc Natl Acad Sci USA 1993; 90:9171.