ORIGINAL ARTICLE

Regulatory T cells in the blood: a new marker of surgical stress

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Abstract

Purpose The function of regulatory T cells (Foxp3⁺ CD4⁺CD25⁺ T cells: T_{reg}) after surgery remains unspecified. We investigated the potential role of T_{reg} as a new stress marker for various operations.

Methods Thirty-three patients who underwent various operations at our department were divided into the following three groups based on the invasiveness of their surgery: Group A, those who underwent massively invasive surgery; Group B, those who underwent moderately invasive surgery; and Group C, those who underwent minimally invasive surgery. We measured T_{reg} levels in the peripheral blood by flow cytometry and labeling with anti-CD4, CD25, and Foxp3 antibodies on preoperative day 1 and then on postoperative days (PODs) 1 and 6. T_{reg} subpopulations in each group on the 3 days were compared. Results Treg subpopulations were significantly higher on POD 6 than on preoperative day 1 in all patients. In Group B, T_{reg} subpopulations varied according to the operative procedures undertaken. For example, there were marked differences between open and laparoscopic abdominal surgery. In Group A, the T_{reg} subpopulations tended to be increased on POD 6, although those on POD 1 were lower than those on preoperative day 1.

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Department of Surgery, Institute of Health Biosciences, The University of Tokushima Graduate School, 3-18-15 Kuramoto-cho, Tokushima 770-8503, Japan e-mail: mshimada@clin.med.tokushima-u.ac.jp *Conclusions* These findings suggest that T_{reg} is an efficient biomarker, indicative of the degree of surgical stress and its impact on immunological status.

Keywords Foxp3 · Immune suppression · Invasiveness

Abbreviations

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Introduction

Immunosuppressive conditions following surgical stress are often associated with postoperative complications; however, the mechanism of their pathological formation, including immunoreaction, is unclear and there are no useful markers of degrees of stress in a living body.

Regulatory T cells (T_{reg}) comprise a subset of T cells with immunosuppressive function. The T_{reg} subset in both humans and rodents is defined as CD4⁺ CD25⁺ cells that express high levels of the transcription factor, Foxp3, which is required for their development in vivo [1, 2]. T_{reg} has been reported to be involved in the pathogenesis of allergic and autoimmune diseases, antigen-specific immune response, and modulation of inflammatory response [3–6]. Recently, Monneret et al. [7] reported increased numbers of CD4⁺CD25⁺ T cells in the peripheral blood of septic patients. The role of T_{reg} in various stresses, including surgery, means it could become a useful biomarker indicative of living body defense mechanisms.

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This study investigates the changes of T_{reg} in the blood after surgery, in an attempt to clarify its potential as a new biomarker of the degrees of surgical stress.

Methods

Patients

The subjects of this study were 33 patients who underwent surgery at the University of Tokushima Hospital between October 2006 and March 2007. The patients were divided into the following three groups based on the degree of surgical invasiveness. The classification was decided with reference to the physiologic ability and surgical stress (E-PASS) scoring system [8-10]. This system classifies the degree of digestive surgical stress caused by blood loss, operation time, and extent of the skin incision. Group A comprised patients undergoing massively invasive surgery, including pancreatoduodenectomy (n = 3), distal pancreatectomy (n = 2), segmentectomy or more extensive resection of the liver (n = 11), and gastrectomy (n = 3). Group B comprised patients undergoing moderately invasive surgery, including partial resection of the liver (n = 1), open colectomy (n = 3), intra-abdominal tumor extirpation for peritoneal recurrence of pancreatic cancer or gastrointestinal stromal tumor of mesocolon (n = 2), splenectomy (n = 1), and laparoscopic collectomy (n = 3). Group C comprised patients undergoing minimally invasive surgery, including laparoscopic cholecystectomy (n = 2) and inguinal hernioplasty (n = 2).

This study was authorized in advance by the Institutional Review Board of the Tokushima University Graduate School. All patients provided written informed consent to take part in our study.

Cell isolation

Peripheral blood was collected on preoperative day 1 and postoperative days (PODs) 1 and 6 from all patients in Groups A and B; and on preoperative day 1 and POD 1 from those in Group C because of their early discharge from hospital. We calculated the ratio of regulatory T cells to peripheral blood mononuclear cells (PBMCs), not the absolute number of regulatory T cells or the ratios of T^{reg} cells to CD4 positive cells, using flow cytometry, as described previously [11]. The authors of this previous clinical report also calculated T_{reg} subpopulations gating PBMCs using the anti CD4 antibody and the anti CD25 antibody and regating these subpopulations using the anti-Foxp3 antibody. We calculated T_{reg} subpopulations in the same manner. These samples were subjected to flow cytometry for multicolor analysis of T cells and stained



Fig. 1 The peripheral blood mononuclear cells (PBMCs) were subjected to flow cytometry after labeling with anti-CD4, anti-CD25, and anti-Foxp3 antibodies. In this study, we ascribed T_{reg} to CD4⁺CD25⁺Foxp3⁺ cells in CD4⁺CD25⁺cells

according to the manufacturer's guidelines. A 3-mL peripheral blood sample was taken from each subject and the PBMCs were isolated using 4 mL of Lymphoprep (AXIS-SHIELD, Oslo, Norway), centrifuged at 1800 rpm for 20 min at room temperature, and then resuspended with 4 mL of FACS buffer. After the addition of anti-CD4 (Allophycocyanin (APC)-conjugated antihuman CD4 antibody; e-Bioscience, San Diego, CA, USA) and anti-CD25 (FITC conjugated antihuman CD25 antibody; e-Bioscience) antibodies to the cell pellets, the mixture was centrifuged at 2000 rpm for 5 min at 4 °C, placed on ice for 15 min in the dark, and further incubated at 20 °C overnight. After two washes, a blocking buffer was added and the mixture was incubated in the dark at room temperature for 15 min. Next, an anti-Foxp3 antibody (Phycoerythrin (PE)-conjugated monoclonal antihuman Foxp3 antibody; e-Bioscience) was added to the cell pellet in the dark and incubated at room temperature for 30 min. The cells were then washed twice with FACS buffer, centrifuged at 1800 rpm for 5 min at 4 °C, and analyzed using a FACSCalibur (BD Biosciences, San Jose, CA, USA) (Fig. 1). The entire procedure was completed within 24 h of the blood collection.

Statistical analysis

Statistical analyses were performed using statistical software (JMP 8.0.1., SAS Campus Drive, Cary, 27513 NC, USA). T_{reg} subpopulations are expressed as mean \pm SEM and analyzed with the Mann–Whitney *U* test. A *p* value of 0.05 or less was considered significant.

Results

Based on our analysis, the T_{reg} subpopulations on POD 6 were significantly higher than those on preoperative day 1 in all patients, regardless of surgical invasiveness (1.4×

fold changes on POD 6; Fig. 2). When the T_{reg} subpopulations were analyzed according to surgical invasiveness in Group B, they increased gradually after surgery, with statistical differences between preoperative day 1 and POD 6 (Fig. 3). On the other hand, in Group A, the T_{reg} subpopulations were significantly higher on POD 6, whereas those on POD 1 were lower than those on preoperative day 1. There were no significant differences between Groups A and B on PODs 1 and 6.

We also evaluated changes in T_{reg} subpopulations within each group. Figure 4 shows the T_{reg} subpopulation changes in Group C. The T_{reg} subpopulations in patients who underwent inguinal hernioplasty tended to increase after surgery; however, the opposite change was seen in those who underwent laparoscopic cholecystectomy. In Group B,



Fig. 2 Analysis of T_{reg} subpopulations in all patients, regardless of surgical invasiveness, revealed significantly higher T_{reg} subpopulations on postoperative day 6 than on preoperative day 1 (p < 0.05)



Fig. 3 T_{reg} subpopulations were analyzed according to surgical invasiveness. In Group B, T_{reg} subpopulations gradually increased after surgical treatment. In Group A, T_{reg} subpopulations were significantly higher on postoperative day (POD) 6, although those on POD 1 were lower than those on preoperative day 1

the T_{reg} subpopulations varied according to the operative procedures the patients underwent; for example, open versus laparoscopic abdominal surgery. After open colectomy, the T_{reg} subpopulations increased gradually, whereas after laparoscopic colectomy, it normalized quickly even though it increased linearly after open colectomy (Fig. 5). There were no significant differences among preoperative day 1, PODs 1 and 6. Figure 6 shows the T_{reg} subpopulation changes in Group A. The T_{reg} subpopulations tended to be higher on POD 6, although they were lower on POD 1 than on preoperative day 1. For example, with hepatectomy, the T_{reg} subpopulations on POD 1 were significantly lower than those on preoperative day 1 (p < 0.05).

Discussion

To the best of our knowledge, this is the first report on T_{reg} subpopulations in peripheral blood during the perioperative period. We reported previously that Treg cells are regulators and suppressors of T cells, which can induce a tolerant state in transplantation [12]. Moreover, T_{reg} cells were found to be good markers for detecting metastasis in patients with pancreatic cancer, especially at advanced stages of the disease [13]. In the present study, we investigated the role of T_{reg} in evaluating surgical stress. We found that T_{reg} subpopulations on POD 6 were higher than the preoperative levels and that in Group A, the T_{reg} subpopulations on POD 1 were lower than those on preoperative day 1. Several measures of surgical stress response have been proposed including serum concentrations of IL-6, C-reactive protein (CRP), lactic acid, and cortisol [14–16]. It is thought that the concentration of a proinflammatory cytokine in the blood, such as IL-1b, TNF-a, IL-6 or IL-8 should predict the degree of surgical stress; however, the measurement of these cytokines is labor intensive and requires a long time for analysis. Furthermore, it is not common to measure pro-inflammatory cytokines during the perioperative period. On the contrary, T_{reg} subpopulations in peripheral blood can be measured easily at a low cost. We did not have to culture the blood overnight for the intranuclear staining of Foxp3. Thus, our method is faster than other surgical stress markers.

Increased percentages of T_{reg} in the blood and spleen during the early stage of experimental and human sepsis have been reported [17, 18]. Furthermore Nascimento et al. [19] proposed that the persistent expansion of T_{reg} in sepsis-surviving mice was correlated with a long-term adaptive immune dysfunction and susceptibility to secondary infection. We attributed the tendency of the T_{reg} subpopulations on POD 6 to be higher than those on preoperative day 1 in all patients to be related to immunosuppression, which was modified by anti-inflammatory cytokines such



Fig. 5 T_{reg} subpopulation changes in Group B. Treg subpopulations varied according to the operative procedures such as after open versus laparoscopic abdominal surgery





Lap-colectomy (n=3)





Fig. 6 Treg subpopulation changes in Group A. Treg subpopulations tended to be higher on postoperative day (POD) 6, although those on POD 1 were lower than those on preoperative day 1

as IL-10 and TGF-B. If homeostasis causes surgical stress, it leads to a form of SIRS, and is controlled by antiinflammatory cytokines with antagonistic actions [20]. On the other hand, there is evidence that the synergistic effects of IL-2 and TGF- β can induce naive CD4⁺ cells to become CD25⁺Foxp3⁺ suppressor cells that express the characteristic markers of natural T_{reg} cells [21].

Interestingly, after major surgery such as hepatectomy, the T_{reg} subpopulations were lower on POD 1 than on preoperative day 1 (p < 0.05). Jiao et al. [22] reported an accumulation of T_{reg} in inflammatory tissues and found significantly higher numbers of Treg cells in synovial fluid than in paired peripheral blood from patients with rheumatoid arthritis. Thus, the decreased T_{reg} subpopulations we observed in peripheral blood on POD 1 might reflect variation in the body distribution of T_{reg}. There were no significant differences between Groups A and B on PODs 1 and 6. It is likely that the T_{reg} subpopulations in Group A were lower than those in Group B. Further study on a larger number of patients is required to find a way to classify the surgically invasive groups.

Regarding the changes of T_{reg} subpopulations in Group C, we observed higher T_{reg} subpopulations after inguinal hernioplasty; however, the opposite was observed after laparoscopic cholecystectomy. Because of the small number of patients and the lack of T_{reg} subpopulations on POD 6, we could not explain why the changes of T_{reg} subpopulations after inguinal hernioplasty differed so much from those after laparoscopic cholecystectomy. Moreover, the Treg subpopulations had normalized by POD 6 after laparoscopic colectomy even though they remained elevated in the open colectomy group. Conversely, the T_{reg} subpopulations were significantly decreased on POD 1 and

increased on POD 6. Taken together, we considered that T_{reg} subpopulations in peripheral blood decreased after surgery depending on the invasiveness of the procedure, increased with recovery, and normalized after the effects of surgical stress had diminished. Further investigations are required to prove this theory, but if it is shown to be accurate, it is possible that the length of hospital stay and complication risk could be predicted from the perspective of T_{reg} subpopulation changes after surgery.

In summary, our findings suggest that T_{reg} subpopulations in peripheral blood vary, distinguishing this marker from existing stress markers. It is possible that T_{reg} will become a new surgical stress marker based on immunological status.

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