#### **REVIEW ARTICLE**



# Hepatokines: unveiling the molecular and cellular mechanisms connecting hepatic tissue to insulin resistance and inflammation

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#### Abstract

Insulin resistance arising from Non-Alcoholic Fatty Liver Disease (NAFLD) stands as a prevalent global ailment, a manifestation within societies stemming from individuals' suboptimal dietary habits and lifestyles. This form of insulin resistance emerges as a pivotal factor in the development of type 2 diabetes mellitus (T2DM). Emerging evidence underscores the significant role of hepatokines, as hepatic-secreted hormone-like entities, in the genesis of insulin resistance and eventual onset of type 2 diabetes. Hepatokines exert influence over extrahepatic metabolism regulation. Their principal functions encompass impacting adipocytes, pancreatic cells, muscles, and the brain, thereby playing a crucial role in shaping body metabolism through signaling to target tissues. This review explores the most important hepatokines, each with distinct influences. Our review shows that Fetuin-A promotes lipid-induced insulin resistance by acting as an endogenous ligand for Toll-like receptor 4 (TLR-4). FGF21 reduces inflammation in diabetes by blocking the nuclear translocation of nuclear factor-κB (NF-κB) in adipocytes and adipose tissue, while also improving glucose metabolism. ANGPTL6 enhances AMPK and insulin signaling in muscle, and suppresses gluconeogenesis. Follistatin can influence insulin resistance and inflammation by interacting with members of the TGF- $\beta$  family. Adropin show a positive correlation with phosphoenolpyruvate carboxykinase 1 (PCK1), a key regulator of gluconeogenesis. This article delves into hepatokines' impact on NAFLD, inflammation, and T2DM, with a specific focus on insulin resistance. The aim is to comprehend the influence of these recently identified hormones on disease development and their underlying physiological and pathological mechanisms.

Keywords Diabetes · Hepatokines · Inflammation · Insulin resistance · Liver · Obesity

#### Abbreviations

Abbieviatio	113				
NAFLD	Non-alcoholic fatty liver disease				
T2DM	Type 2 diabetes mellitus				
TLR-4	Toll-like receptor 4				
FGF-21	Fibroblast growth factor 21				
ANGPTLs	Angiopoietin-like proteins				
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GDF15	Growth/Differentiation Factor-15
AMPK	Adenosine monophosphate-activated protein
	kinase
GLUT4	Glucose transporter type 4
MAPKs	Mitogen-activated protein kinases
JNK	c-Jun N-terminal kinases
ERK5	Extracellular signal-regulated kinases 5
IRS1	insulin receptor substrate 1
mTOR	Mammalian target of rapamycin
Fet A	Fetuin A
NF-κB	Nuclear factor-kB
WAT	white adipose tissue
LPLs	Lipoprotein lipase gene
FAK	Focal adhesion kinase
IL-6	Interleukin 6
ICAM-1	Intercellular adhesion molecule 1
VLDLs	Very-low-density lipoproteins

LXR	Liver X receptor
PPAR	Peroxisome proliferator-activated receptor
FFA	Free fatty acid
CVD	Cardiovascular diseases
G6P	glucose-6-phosphatase
GH	Growth hormone
HDL	High-density lipoprotein
ACC	AMPK-acetyl coenzyme A carboxylase
NASH	non-alcoholic steatohepatitis
MIC-1	Macrophage inhibitory cytokine-1
GDNF	Glial cell-derived neurotrophic factor
UPR	unfolded protein response
MCP-1	Monocyte chemoattractant protein-1
RTK	Receptor tyrosine kinase
HMGB1	High mobility group box protein 1
PAMPs	Pathogen-associated molecular patterns
TGF-β	Transforming growth factor-β
IGF-1	Insulin-like growth factor 1
FSH	Follicle-stimulating hormone
TNF-α	Tumor Necrosis Factora
CHM2	Chondromodulin II
BMI	Body mass index
HFREP1	Hepatocyte-derived fibrinogen-related pro-
	tein 1
IL	Interleukin
EC	Extracellular calcium
TY	Thyroglobulin-like
CRP	C-reactive protein
NO	Nitric oxide
GHR	GH receptor
JAK2	Janus Kinase 2
BAT	Brown Adipose Tissue
SNS	Sympathetic nervous system
HFD	High-fat diet
CPT1a	Carnitine palmitoyltransferase-1a
ChREBP	carbohydrate response element binding
	protein
TSK	Tsukushi
LCN-13	Lipocalin-13
HSL	Hormone-sensitive lipase
РКА	Protein Kinase A
HUVEC	Human Umbilical Vein Endothe-
	lial CellsActR: Activin A receptors,
	PCK1:Phosphoenolpyruvate carboxykinase 1
LBP	Lipopolysaccharide binding protein
PEPCK	Phosphoenolpyruvate carboxykinase
LPS	Lipopolysaccharide
G6PC	Glucose-6-phosphatase catalytic subunit

# Introduction

The extensive impact of insulin resistance caused by nonalcoholic fatty liver disease (NAFLD) is highlighted by its global prevalence. This condition is a result of people's lessthan-ideal food choices and lifestyle trends [1]. Among the many risk factors for developing type 2 diabetes mellitus (T2DM), insulin resistance due to NAFLD stands out [2]. It has been established that hepatokines are liver-secreted hormone-like substances. Despite their role in metabolism; recent findings suggest the participation of some these hepatokines, as endocrine messengers in the development of insulin resistance and ultimately, type 2 diabetes. [3]. Hepatokines constitute a category of substances released by the liver that function in the regulation of metabolism beyond hepatic tissues. Their principal roles include influencing adipocytes, pancreatic cells, muscles, and the brain, thereby exerting a substantial impact on body metabolism through signaling to target tissues [4, 5].

Within this comprehensive review, we delve into the intriguing realm of the most important hepatokines, each wielding a distinctive influence. These remarkable substances wield their effects by modulating the quantity of liver insulin receptors, exemplified by activin E [6], orchestrating a surge in glucose overload within liver or muscle cells, as demonstrated by Fibroblast growth factor 21 (FGF21) [7]. Furthermore, hepatokines such as Angiopoietin-like proteins (ANGPTLs) [8] orchestrate a metabolic shift towards fat.

As well as, hepatokines could potentially influence the molecular mechanisms that result in inflammation within the liver, adipose tissues, or muscles, for instance like Growth/ Differentiation Factor-15 (GDF15), intricately navigate inflammatory pathways. This exploration illuminates the captivating interplay of hepatokines, unraveling their multifaceted roles in pathophysiological mechanisms [9]. In this regards, this article endeavors to unravel the intricacies of hepatokines, delving into their impact on NAFLD, inflammation, and ultimately, T2DM, with a specific emphasis on the intricate dynamics of insulin resistance. Through this exploration, we aim to elucidate the influence of these recently unveiled hormones on the pathogenesis of the mentioned diseases, offering valuable insights into their physiological mechanisms.

# Insulin resistance, inflammation and hepatokines

As mentioned before, new research indicates that hepatokines may play a role as endocrine messengers in the progression of insulin resistance and, ultimately, the development of T2DM [3]. Insulin resistance denotes a diminution in the responsiveness of tissues to insulin, a hormone pivotal in facilitating glucose movement to muscles, adipose tissues, and the liver [10]. The impediment of insulin signal transduction, primarily attributed to mutations and polymorphisms in insulin receptors [11], stands as a predominant cause of insulin resistance. Another mechanism that might lead to insulin resistance is disruption of lipid storage in the skeletal muscles and liver. This insulin resistance has been linked to a number of metabolic disorders, including T2DM, atherosclerosis, and NAFLD [12].

Insulin resistance may manifest at multiple junctures along the intricate insulin signaling pathway, exerting a direct influence on the modulation of glucose levels, protein synthesis, and the onset of T2DM along with its associated complications. The intricacies of insulin resistance are elucidated through diverse molecular mechanisms [2, 13, 14].

On the other hand, hepatokines are acknowledged as a category of hormones instrumental in the physiological modulation of inflammation. The ensuing instances elucidate three scenarios wherein hepatokines influence the molecular pathways underpinning inflammation and insulin resistance. Illustratively, fetuin-A (FetA) facilitates the development of lipid-induced insulin resistance through its role as an endogenous ligand for TLR-4 [15-18]. Moreover, this protein exerts a pronounced impact on expediting the assimilation of exogenous fatty acids into cellular triglycerides [19]. Functioning as an upstream modulator, FetA plays a regulatory role in the polarization of M1 macrophages and the activation of TLR4 induced by free fatty acids in adipocytes. Consequently, FetA emerges as a potential novel therapeutic target for addressing T2DM mellitus and inflammation associated with obesity [20, 21]. Recent investigations have associated FGF21 with a spectrum of anti-inflammatory effects, contributing to the expanding body of research elucidating its advantageous implications for metabolic processes [7, 21]. Notably, FGF21 mitigates inflammation in diabetes by impeding the nuclear translocation of nuclear factor- $\kappa B$  (NF- $\kappa B$ ) in adjpocytes and adjpose tissue under insulin-resistant conditions, concurrently augmenting glucose metabolism [7, 22].

# Hepatic function and secretion: insights into essential dynamics

The energy metabolism of mammals intricately hinges upon nutrient intake, wherein the liver assumes a pivotal responsibility for assessing and orchestrating the utilization of these nutrients across various tissues. Glucose, along with other monosaccharides, stands as the predominant substrate for metabolism in living organisms. The liver, in its regulatory capacity, modulates blood glucose levels through dynamic interactions with the pancreas, which releases insulin to sustain a consistent blood glucose concentration. Furthermore, during phases of satiety, the liver actively assimilates blood glucose, converting and storing it in the form of glycogen [23].

Findings from proteomic studies involving both human subjects and rodents indicate that around 40% of the proteins originate from hepatic sources [24, 25]. With the help of hormones like hepatokines, myokines, adipokines, and neurokines, the liver is able to interact with other organs. These hormones elicit physiological effects, contributing to conditions such as atherosclerosis and insulin resistance [26]. The dynamic modulation of hepatokine levels during the evolution and manifestation of metabolic disorders significantly influences metabolic processes. Understanding the dynamic interactions between hepatokines and other organs is essential not only for unraveling their diverse physiological roles and potential implications in systemic health, but also for revealing metabolic disorders significantly influences metabolic processes [5, 27]. In this regard, the following information, summarized in Table 1, furnishes a comprehensive overview of diverse hepatokines and their influence on critical physiological aspects such as insulin resistance, liver function, lipid metabolism, metabolic conditions, and lifestyle factors.

#### Angiopoietin-like proteins (ANGPTL)

They derive their nomenclature from the resemblance of their protein domains to the angiopoietin family. These glycoproteins, secreted by hepatocytes, operate as vascular growth factors. Notably, a coiled-coil domain in their N-terminal facilitates subunit binding and the assembly of homo-oligomers. Furthermore, they feature a fibrinogen-like domain in their C-terminal, which binds to the Tie2 receptor, endowing them with a structural similarity to angiopoietin proteins [28, 29]. As a physiological function, these hepatokines assume a pivotal role in the modulation of white adipose tissue (WAT) function and the regulation of the lipoprotein lipase gene (LPLs) expression [30]. Located on the inner surface of capillaries, lipoprotein lipase is accountable for the hydrolysis of triglycerides into fatty acids, subsequently releasing them into the tissues that lipoproteins aim to reach [31]. The Golgi system experiences regulation by ANG-PTL1-8, exerting inhibitory effects on LPLs expression at the post-translational level [30]. Within the ANGPTL protein family, ANGPTL2, 4, and 8 predominantly participate in inflammatory processes, while other members can also contribute to the overall inflammatory milieu [32].

As mentioned before, the ANGPTL protein family is involved in inflammatory processes in various bodily

Hepatokines	Insulin resistance	Metabolic conditions	Adipose tissue and lipid profile	Liver tissue	Life style
Activin E	Exacerbation of insulin resis- tance associated with obesity.		Facilitation of ther- mogenesis in brown adipose tissue	The control of insulin membrane receptor den- sity in hepatocytes	Elevated lev- els in mice undergo- ing obesity induced by a high-fat diet.
ANGPTL3	Insulin, at the mRNA expres- sion level, inhibits ANGPTL3 production downregulation of genes associated with the gluconeogenesis reduction in the efficiency of insulin-mediated VLDL secretion	elevated serum levels in T2DM	Correlation of elevated ANGPTL3 expression with higher plasma triglyceride levels and reduced free fatty acids. the accumulation of triglycerides in white adipose tissue cells Suppression of VLDL Breakdown	Regulatory influence of LXR on ANGPTL3 concentrations	
ANGPTL4	An increase in insulin sensi- tivity via its glucose-lowering effect and enhanced glucose processing	Lower serum level in individual with obesity, diabetes, and atherosclerosis	As fasting-induced adi- pose factor, is secreted by adipose tissue. Increased serum triglyceride levels both by inhibiting lipoprotein lipases and triglyceride clearance Facilitation of lipid accumulation in adipo- cytes, contributing to steatosis	As Fasting-induced adi- pose factor, is secreted by hepatocytes Development of T2DM by enhancing lipid buildup in liver cells	the release of fatty acids in response to catechamine- induced short-term fasting and gluco- corticoids- induced long-term fasting
ANGPTL6	Enhancement of AMPK and insulin signaling in muscle			Suppression of gluconeogenesis, catabolic shift in sugar metabolism induced by inhibiting the G6Pase gene	Direct correlation with fasting insulin and fasting blood sugar levels
ANGPTL8	Potential enhancement of insulin sensitivity by directly affecting Akt phosphorylation	Polymorphism in NAFLD Development	Its inhibitory effect on LPL influenced by ANGPTL3 and ANGPTL4		
Adropin	Therapeutic potential to increase glucose tolerance and insulin sensitivity	Lower serum levels in Obesity and T2DM		Lower levels in response to more liver availability of lipid Impact of hepatic ste- atosis on ENHO gene expression encoding adropin A positive correlation between adropin and PCK1 in hepatic steatosis	Upregulation in hepatic steatosis induced by lipid-rich diet
Selenoprotein P	SELENOP synthesis trig- gered by AMPK activation, achievable through the use of anti-inflammatory or insulin- sensitizing medications. Dysregulation of Glucose Metabolism	Positive correlation with NAFL Elevation of its mRNA level in T2DM			

Table 1
 Comprehensive overview of hepatokines: effects on insulin resistance, liver function, l,ipid metabolism, metabolic conditions, and life-style

Table 1 (continued)

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Hepatokines	Insulin resistance	Metabolic conditions	Adipose tissue and lipid profile	Liver tissue	Life style
Fibroblast growth factor 21	As an insulin-dependent hormone in humans Improved insulin and leptin sensitivity Enhanced glucose uptake and utilization Glucose Homeostasis and β-Cell Protection	Protective effects against T2DM, hyper- glycemia, dyslipid- emia, Non-Alcoholic Steatohepatitis, NAFLD	Stimulating adipose tissue lipolysis Independent glucose uptake in adipocytes and systemic insulin resistance reduction	Decreased hepatic steatosis Direct promotion of liver fatty acid oxidation Improving Hepatic Insu- lin Sensitivity	Fasting- induced metabolic changes regulated by FGF21 in liver
Growth differen- tiation factor 15	Preventive effect against obe- sity and insulin resistance	Efficacy of GDF15 in ameliorating NAFLD and mitigating obesity	Metabolic shift from glucose to fat gen- eration in response to stress p53-mediated expres- sion of GDF15 in adipose tissue for influencing lipid catabolism, heat gen- eration, and oxidative metabolism		Central appetite- suppressing effects in response to stress conditions, Severe Ail- ments and GDF15- Induced Anorexia- Cachexia Syndrome
Fetuin-A	Positive Correlation with Insulin Resistance Antagonistic effect on insulin- stimulated insulin receptor tyrosine kinase	Positive Correla- tion with T2DM and NAFLD Dualistic Inflamma- tory Properties Protective role against HMGB1 release in severe systemic inflammation	Modulation of adipo- nectin levels via Wnt- PPARγ pathway in lipid-induced inflamed adipocytes		
Follistatin	Association between FST, insulin resistance, glu- cose intolerance and mild inflammation	Moderate increase in plasma FST levels in individuals with T2DM	An increase in white adipose tissue fat breakdown and the blood levels of glycerol and non-esterified fatty acids		
Leukocyte cell-derived chemotaxin-2	Insulin sensitivity enhance- ment in Lect2–/– mice A causative factor in skeletal muscle insulin resistance Contribution of hepatic LECT2 to JNK phosphoryla- tion and insulin resistance	Obesity-related insulin resistance		liver regeneration A direct correlation between hepatic LECT2 mRNA levels and obesity	
Hepassocin	The lower hepatic hepas- socin level, the more insulin sensitivity Insulin resistance in skel- etal muscle mediated by hepassocin	Association with obesity Key role in NAFLD development Insulin resistance and diabetes may increase with high hepassocin levels.		Mitogenic effects on liver cells Hepatic lipid accumula- tion via a ERK1/2-depen- dent mechanism	
Secreted modu- lar calcium- binding protein 1	Insulin sensitivity enhancement	Lower circulating levels in obesity Potential therapeutic for glycemic control and insulin sensitivity in T2DM		Downregulation of gluco- neogenic gene expression Suppression of hepatic glucose production	

Hepatokines	Insulin resistance	Metabolic conditions	Adipose tissue and lipid profile	Liver tissue	Life style
Insulin-like growth factor 1	Structurally similar to insulin The more IGF1, the better insulin sensitivity in both individuals with and without T2DM Overexpression of IGFBP1 mitigates hypoglycemic impact, but induces insulin resistance in skeletal muscle	Reduced plasma IGF1 levels as a prognostic indicator for T2DM onset Association of lower IGF Levels with NAFLD and Obesity	Role of IGF1 in energy homeostasis regulation in brown adipose tissue through sympathetic nervous system activity		Modulates glucose metabolism in mice subjected to regular or high-fat diets The regulation of appetite
Lipocalin-13		An inverse correlation with obesity Therapeutic candi- date for T2DM and NAFLD	Enhancement of adipocytes' insulin sensitivity facilitating glucose absorption in both insulin-independent and insulin-dependent manners	Inhibition of hepatic gluconeogenesis Prevention of obesity- related hepatic steatosis by enhancing fatty acid β-oxidation and reducing lipogenesis Insulin-independent regulation of glucose metabolism in liver regulation of lipid metabolism via CPT1α	A reduction in circulat- ing LCN13 levels after a high-fat diet
Tsukushi		Induction by obesity and cold exposure Interplay between TSK and inflammation Prolonged TSK elevation in obesity, Atherosclerosis, and NAFLD	Deletion of TSK induces thermogenesis in brown adipose tissue	Liver Steatosis Liver regulation of TSK expression by inflamma- tion and ER stress Hepatocytes respond to proinflammatory cyto- kines by promoting the transcription of TSK Protective role of initial TSK surge in stressed liver cells	Deletion of TSK pro- vides protec- tion against diet-induced obesity
Hepcidin	Affect iron metabolism in adi- pose tissue, liver ferroptosis and development of diabetes and insulin resistance	Dysregulation of hepcidin leads to vari- ous metabolic diseases such as obesity, type 2 diabetes, and insulin resistance		Dysregulation of hepcidin leads to liver ferroptosis inactivation of the hepcidin-coding gene in liver cells results in the activation of Akt and gly- cogen synthase kinase-3β (GSK3β)	
Lipopolysacc- haride binding protein	By changing the lipid metabolism can affect insulin sensitivity		Participating in the metabolism of triglyc- erides and high-density lipoprotein (HDL) cholesterol		The serum level of LPS increases with a high- fat diet

conditions. Although the first member of this family, ANG-PTL1, has an unclear role in inflammation, it does interfere with inflammatory pathways in cancer. ANGPTL1 inhibits the integrin a1ß1/focal adhesion kinase (FAK)-Src/JAK/ STAT3 signaling pathway by binding to integrin a1ß1 in liver cancer cells, thus suppressing metastasis and angiogenesis. Furthermore, ANGPTL1's binding to integrin a1ß1 inhibits the expression of zinc finger protein SLUG, which

has an inhibitory effect on lung cancer [33–36]. Moreover, ANGPTL2 plays a role in inflammatory responses and various metabolic disorders. This hepatokine, which is also released from adipose tissue, exerts a favorable impact on inflammation, obesity, and systemic insulin resistance in both humans and mice. Figure 2 shows, in a research study, recombinant ANGPTL2 was utilized to treat human umbilical vein endothelial cells, leading to the activation of Rac1 and NF- $\kappa$ B via integrin  $\alpha$ 5 $\beta$ 1, thereby enhancing the inflammatory pathway [33, 37]. Additionally, this treatment was applied to endothelial cells in atherosclerosis, where the interaction of this hepatokine with integrin  $\alpha$ 5 $\beta$ 1 resulted in increased degradation of I $\kappa$ B $\alpha$ and elevated secretion of pro-inflammatory cytokines and

Table 1 (continued)



Fig. 1 The impact of hepatokines on insulin signaling pathways. GLUT4: Glucose transporter type 4, IRS: insulin receptor substrate, JAK2: Janus Kinase 2, IGF-1: Insulin-like Growth Factor 1, FGF-21: fibroblast growth factor 21, ANGPTL: Angiopoietin-like protein, SELENOP: Selenoprotein P, PPARs: Peroxisome proliferatoractivated receptors, LCN13: Lipocalin 13, PI3K: Phosphoinositide

adhesive molecules such as TNF- $\alpha$ , interleukin 6 (IL-6), and intercellular adhesion molecule 1 (ICAM-1), as presented [33, 38, 39].

Within its structural framework, ANGPTL3 incorporates a distinctive epitope associated with lipoprotein lipase, thereby influencing the activity of lipoprotein lipase and modulating plasma triglyceride levels. The influence of ANGPTL3 on obesity, insulin resistance, and hyperlipidemia has been substantiated through rodent studies [40]. Subsequent investigations involving rodents elucidated that elevated ANGPTL3 expression correlates with increased plasma triglyceride levels and decreased levels of free fatty acids [41]. The ANGPTL3 protein was identified as a suppressor of LPLs in an in-vitro setting, resulting in consequential alterations to free fatty acid and triglyceride levels in the bloodstream [42]. Human studies have revealed that the inhibition of ANGPTL3 leads to a reduction in triglyceride levels [43]. Moreover, diabetic patients exhibit elevated serum levels of ANGPTL3 compared to non-diabetic individuals [8]. Investigations on mice have demonstrated the involvement of ANGPTL3 in the accumulation of triglycerides in white adipose tissue cells. By supplying glucose to accumulated adipocytes, ANGPTL3 KO mice have

3-kinases, LECT2: Leukocyte cell-derived chemotaxin-2, FST: Follistatin, FSTL1: Follistatin-like protein 1, ACC: Acetyl-coA carboxylase, AMPK: Adenosine monophosphate-activated protein kinase, GDF-15: Growth differentiation factor 15, SMOC1: Secreted modular calcium-binding protein 1, mTORC1: Mammalian target of rapamycin complex 1

compensated for their tissue mass weight, which stayed similar to control mice [44]. Evidence suggests that the coiledcoil region of ANGPTL3 plays a pivotal role in suppressing the breakdown of very-low-density lipoproteins (VLDLs) [45]. The regulatory influence of liver X receptor (LXR) activity on ANGPLT3 concentrations has been documented as well [46]. Leptin and insulin, at the mRNA expression level, inhibit the production of ANGPTL3. Hormones like these cause hyperlipidemias and hyperfattyacidemia in those who are overweight, diabetic, or insulin-resistant [47]. Inhibiting ANGPTL3 leads to a downregulation of genes associated with the gluconeogenesis pathway and a reduction in the efficiency of insulin-mediated VLDL secretion [48]. Apart from the aforementioned functions, when ANGPTL3 binds to aVB3 integrin, it enhances the movement and attachment of endothelial cells, promoting angiogenesis and indicating the potential pro-inflammatory role of ANGPTL3. Additionally, a separate animal study demonstrated that upregulation of ANGPTL3 in mice with diabetic retinopathy led to increased levels of Bax, p53, VEGF, and inflammatory cytokines (IL-6 and IL-1B) [33, 49].

Regarding ANGPTL4, this protein displays a reduced molecular size in comparison to ANGPTL3, while yet

maintaining its structural arrangement for LPL binding [50]. Its ability to increase liver cell lipid buildup contributes to T2DM [5]. This particular protein is alternatively recognized as Fasting-induced adipose factor (Fiaf) [4]. Predominantly secreted by adipose tissue and hepatocytes, ANGPTL4 levels exhibit an increase during periods of fasting. Catechamine-induced short-term fasting appears to trigger a signaling cascade that releases fatty acids and shifts metabolism toward fat use. In response to glucocorticoids, ANGPTL4 expression is stimulated during extended fasting, leading to fatty acid release. Consequently, the impact of this protein is more pronounced during extended fasting durations [51]. As depicted in Fig. 1, augmenting ANGPTL4 expression triggers the activation of the peroxisome proliferator-activated receptor (PPAR) signaling pathway, subsequently suppressed upon refeeding [52]. When studying mice with obesity, diabetes, and atherosclerosis, researchers found that their ANGPTL4 expression was much lower. [53]. Increased serum triglyceride levels are caused by the N-terminal of ANGPTL4, which inhibits LPLs. At the same time, this protein hinders triglyceride clearance, further increasing blood lipid levels. In contrast to its effects on adipocytes, an augmentation in free fatty acid (FFA) concentration in the blood is observed, emanating from ectopic sources such as the liver or muscle cells [5, 54]. ANGPTL4 exhibits a glucose-lowering effect and enhances glucose processing capabilities, as evidenced by numerous studies. It may be possible to increase insulin sensitivity using this [4, 27, 55]. Human studies involving individuals with T2DM have reported a decrease in the serum concentration of ANGPTL4 [56]. Additionally, ANGPTL4 facilitates lipid accumulation in adipocytes, contributing to steatosis [57]. Conversely, the genetic inactivation of ANGPTL4 has shown promise in substantially reducing the risk of developing coronary artery disease and diabetes [58]. Is there a correlation between insulin resistance and serum concentrations of ANGPTL4 or not? The contradictory results of the research on this protein make it impossible to draw any firm conclusions. It seems that it has a dual role even in inflammation [58]. Stem cells have a wide range of applications in treating cardiovascular diseases (CVD). One example is the use of ANGPTL4 derived from mesenchymal stem cells. which have been found to suppress  $\beta 1/\alpha V\beta 3$  integrin signaling by inhibiting NF- $\kappa$ B (Fig. 2). This contributes to the polarization of inflammatory macrophages as an anti-inflammatory molecule [33, 59]. Moreover, in other study involving mice, it was observed that ANGPTL4 levels increased in cases of acute lung injury. Furthermore, the reduction of this



**Fig. 2** The impact of hepatokines on inflammation pathways. ANG-PTL: Angiopoietin-like protein, TNF- $\alpha$ : Tumor necrosis factor  $\alpha$ , FSTL1: Follistatin-like protein 1, P38 MAPK: P38 mitogen-activated protein kinase, ROS: Reactive oxygen species, LECT2: Leukocyte

cell-derived chemotaxin-2, NF- $\kappa$ B: Nuclear factor- $\kappa$ B, JNK: Jun N-terminal kinase, GDF-15: Growth differentiation factor 15, IGF-1: Insulin-like Growth Factor 1

hepatokine led to an increase in sirtuin1 (SIRT1), which has anti-inflammatory and antioxidant properties [33].

Previous research findings suggest that the levels of ANGPTL5 may be linked to inflammation in both obese individuals and those with type 2 diabetes. In addition, the rise in this hepatokine is correlated with level of hs-CRP [33, 60].

ANGPTL6, on the other hand, lacks a binding domain for LPLs, distinguishing it from the preceding two variants of ANGPTL [28]. Its liver expression is predominate, with little in other tissues [61]. This protein enhances AMPK and boosts insulin signaling in muscle (Fig. 1). Additionally, it suppresses gluconeogenesis by inhibiting the expression of the glucose-6-phosphatase (G6P) gene, leading to a catabolic shift in sugar metabolism [29, 62]. Fasting insulin and blood sugar levels are two indicators that are directly correlated with ANGPTL6 [63]. As well as, this hepatokine is involved in the inflammatory processes associated with metabolic syndromes. For instance, mice that were induced with psoriasis displayed elevated levels of epidermal ANG-PTL6, resulting in more intense skin inflammation [33, 64].

Regarding ANGPTL8, this protein possesses an LPL binding domain, albeit lacking a coiled-coil or fibrinogen C-terminal [28]. The ANGPTL8 protein is primarily secreted by adipose tissue and liver cells [65]. The inhibitory effect of this protein on LPL is greatly influenced by the concentration of ANPGTL3 and 4. It emerges as a potential biomarker for the identification and treatment of metabolic irregularities on a broad scale [4]. The impact of ANGPTL8 on both NAFLD and insulin resistance remains a subject of discourse. Arguments posit that ANGPTL8 may enhance insulin sensitivity by directly affecting Akt phosphorylation [66]. Decreasing ANGPTL8 levels is associated with a reduction in insulin resistance [67]. However, polymorphism of this protein is implicated in the development of NAFLD [68]. It has been reported that overexpression of ANGPTL8 improved insulin sensitivity and glucose tolerance and decreased fasting blood glucose levels in high-fat diet/streptozotocin-induced diabetic and db/db mice. In fact, ANGPTL8 promoted glucose metabolism via the inhibition of expression of gluconeogenesis-related genes [phosphoenolpyruvate carboxykinase (PEPCK) and glucose-6-phosphatase catalytic subunit (G6PC)] by activating the AKT signaling pathway [69].

It is important to note that the final member of this family also plays a role in acute inflammatory conditions by acting as an anti-inflammatory agent. For instance, in individuals with acute infections or in laboratory mice with stimulated hepatocytes, ANGPTL8 controls the NF- $\kappa$ B signaling pathway by promoting the targeted autophagic breakdown of IKK $\gamma$  [33, 70]. Zhang et al. [71] 's study expanded on the function of selective autophagy in fine-tuned inflammatory responses and proposed the ANGPTL8/p62-IKKγ axis as a negative feedback loop that controlled NF-κB activation.

Despite various potential explanations and connections between the members of this family and inflammatory pathways, the involvement of the ANGPTL family in inflammation remains uncertain and requires additional research.

### Fibroblast growth factor 21 (FGF21)

Various tissues, such as the liver (hepatocytes), adipose tissue, pancreas, heart, and brain, release this protein [7, 72]. Fasting-induced metabolic changes in the liver, such as resistance to growth hormone (GH), fatty acid oxidation, ketogenesis, and gluconeogenesis, are tightly regulated by FGF21, as a physiological function [73]. Mice treated with FGF21 have improved insulin and leptin sensitivity, decreased hepatic steatosis, increased energy expenditure, and decreased sugar and alcohol consumption [4, 74, 75].

FGF21 primarily promotes liver fatty acid oxidation while blocking lipogenesis and gluconeogenesis [76]. Second, by reducing the severity of hyperlipidemia and hyperglycemia, FGF21 indirectly affects the liver. To do this, the following mechanisms are activated: mitochondrial activity is enhanced, fatty acid oxidation is facilitated, adipose tissue lipolysis is stimulated, energy is dissipated as heat, and glucose uptake and utilization are improved. Additionally, it influences several areas of the hypothalamus and hindbrain to encourage thermogenesis and energy expenditure as well as an aversion to sweets and alcohol intake [76]. Furthermore, there is a conjecture that FGF21 resistance in both rats and humans is attributed to obesity and insulin resistance [18]. Clinical trials involving FGF21 analogues indicate an elevation in high density cholesterol (HDL) and a reduction in plasma triglyceride concentrations, although they do not exert any discernible impact on insulin sensitivity or glucose levels [77].

The metabolic impacts attributed to FGF21 are acknowledged to arise from its interactions with various tissues [78]. FGF21 directly communicates with adipose tissue, enhancing insulin sensitivity [4]. Simultaneously, it stimulates the central nervous system to elevate calorie expenditure and facilitate weight loss [79, 80].

FGF21, recognized as an insulin-dependent hormone in humans [72], also plays a pivotal role in enhancing glucose balance by safeguarding  $\beta$  cells. FGF21 is believed to prevent  $\beta$ -cell dysfunction and cell death in vivo and in vitro through its inhibition of lipid accumulation in islet cells. As illustrated in Fig. 1, this likely occurs through the activation of the PPAR $\delta/\gamma$  and AMPK-acetyl coenzyme A carboxylase (ACC) signaling pathways [81]. Beyond its reliance on insulin-dependent mechanisms for improving glucose homeostasis, FGF21 independently facilitates glucose uptake in adipocytes [72]. In addressing systemic insulin resistance, FGF21 demonstrates the capacity to reduce serum insulin levels and enhance insulin sensitivity [72]. In-depth investigations into the precise mechanisms underlying FGF21's enhancement of hepatic insulin sensitivity involved administering FGF21 to mice. They discovered that FGF21 can enhance insulin sensitivity via blocking liver mTORC1 (Fig. 1). FGF21-deficient mice consistently exhibit heightened hepatic insulin resistance and activation of mTORC1 [82]. Furthermore, FGF21 targets subcutaneous fat tissue as a crucial component to control systemic insulin sensitivity. To prevent systemic insulin resistance in vivo, FGF21 can encourage the growth of subcutaneous fat and raise adiponectin levels in subcutaneous fat [72].

In summary, FGF21 provides protection against T2DM by improving glucose regulation [83]. Noteworthy is its demonstrated anti-inflammatory prowess [22]. Elevated FGF21 in insulin-sensitive obese individuals worsens the polarization of M2 macrophages and controls adiponectin levels in subcutaneous adipose tissue. As well as, elevating the concentration of this growth factor impacts the transforming growth factor- $\beta$  (TGF- $\beta$ ) signaling pathway and inhibits adipogenesis by influencing the Smad3 pathway [84, 85]. As a result, because of its effects on hepatocytes, the brain-liver axis, and adiponectin synthesis in adipose tissue, FGF21 protects against hyperglycemia, dyslipidemia, non-alcoholic steatohepatitis (NASH), and other comorbidities [86]. FGF21 emerges as a promising therapeutic avenue for the management of T2DM and NAFLD [87].

# Fetuin-A

Classified as a2-Heremans-Schmid-glycoprotein, Fetuin-A is a complex glycoprotein belonging to the cystatin protease inhibitor superfamily. Predominantly synthesized by hepatocytes and adipose tissue, this protein establishes a novel nexus linking obesity, inflammation, and insulin resistance [88, 89]. Fetuin-A, a plasma protein, regulates multiple physiological processes, including cellular protein metabolism, acute inflammatory responses, neutrophil and platelet release, lymphocyte activation, and binding to fatty acids, thyroid hormones, and calcium ions [89]. It serves as a natural antagonist to the insulin-stimulated insulin receptor tyrosine kinase, impeding the autophosphorylation of the insulin receptor and subsequent downstream signaling, as evidenced in laboratory investigations [90, 91]. Notably, as showed in Fig. 1, this hepatokine has been observed to bind to the receptor tyrosine kinase (RTK) at a distinct site from the insulin binding region; however, the precise domain of fetuin-A implicated in this interaction remains unclear [92]. Numerous investigations suggest a positive correlation between heightened levels of circulating fetuin-A and insulin resistance in human subjects [93]. Consequently, this protein emerges as a prospective independent risk factor in the predisposition to T2DM [94-96]. Through its modulation of adiponectin levels in lipid-induced inflamed adipocytes via the Wingless-related integration site (Wnt)-PPARy pathway, Fetuin-A potentially instigates insulin resistance [88, 97]. This cascade of events underscores the plausible role of Fetuin-A in the pathogenesis of type 2 diabetes. Notably, there exists a consensus within the scientific community that NAFLD can be a precursor to insulin resistance and the subsequent onset of T2DM [18, 98]. Regardless of adiposity, Fetuin-A is elevated in NAFLD, suggesting a link between fatty liver and insulin resistance. [93, 99]. The pro-inflammatory or anti-inflammatory effects of this hepatokine are dependent on the particular clinical setting in which it is activated, thereby displaying dualistic inflammatory properties [89, 92, 100]. This molecule, renowned for its proinflammatory attributes, assumes a role in the development of insulin resistance. Additionally, Fetuin-A demonstrates neuroprotective properties and plays a significant part in mitigating inflammation in conditions such as sepsis and autoimmune diseases [100]. In instances of severe systemic inflammation, the protective role of Fetuin-A is ascribed to its capacity to impede the release of high mobility group box protein 1 (HMGB1), induced by pathogenassociated molecular patterns (PAMPs) [89]. As well as, Fig. 2 displays this hepatokine enhances the polarization of M1 macrophages and boosts the release of pro-inflammatory cytokines like interleukin-1β, Tumor Necrosis Factor α (TNF- α), and IL-6 [101].

# Follistatin (FST)

This glucosylated plasma protein can bind and neutralize TGF- $\beta$  family members [102]. Nowadays, 5 types of this family are identified [103]. The gene responsible for its expression is discernible in diverse tissues, including the pituitary gland, placenta, ovary, testis, brain, and skeletal muscle [104, 105]. In the human system, the predominant source of circulating FST is the hepatocytes, with its production and release being augmented in response to an elevated ratio of glucagon to insulin [106]. FST primarily operates through autocrine and paracrine signaling pathways [107, 108]. The manifold physiological roles ascribed to FST encompass the regulation of follicle-stimulating hormone (FSH) production in the pituitary, facilitation of ovarian follicle maturation, control of spermatogenesis, maintenance of liver homeostasis, facilitation of wound repair, and responsiveness to inflammatory stimuli [107]. Emergent evidence suggests a potential involvement of this hepatokine in insulin resistance and mild inflammation. However, it appears that plasma FST levels exhibit a closer association with metabolic disruptions than with low-grade inflammation [104].

Individuals with T2DM show a moderate increase in plasma FST levels, yet further investigations are imperative to elucidate whether the elevated plasma levels constitutes a causative factor or a consequence of metabolic irregularities in T2DM [104, 109]. A recent study has presented findings indicating that heightened circulating FST levels are linked to an increased susceptibility to developing type 2 diabetes, owing to their contribution to insulin resistance in adipose tissue [105]. Here, FST secretion by the liver causes insulin resistance and white adipose tissue fat breakdown. Increased levels of glycerol and non-esterified fatty acids in the bloodstream cause the liver to produce glucose uncontrollably, which ultimately results in glucose intolerance [105, 110]. Follistatin possesses the potential to influence insulin resistance and inflammation by virtue of its interaction with members of the TGF- $\beta$  family. Thus, Fig. 2 presents the expression of FSTL1 is related to the activity of the NF-kB signaling pathway. Also, FSTL1 leads to the expression of various chemokines and inflammatory cytokines that are related to the NF-kB signaling pathway. Among them, CCL-2/MCP-1, TNF-α, CXCL8/IL-8, IL-1β and IL-6 can be mentioned [103]. Moreover, FSTL is attributed to its recognized capability to bind and counteract both myostatin and activin A within the circulatory system [111]. Notably, activin A, emanating from epicardial adipose tissue (in addition to liver) in individuals with T2DM, impedes insulin function by instigating the production of miR-143 in cardiomyocytes. This specific miRNA, in turn, suppresses the Akt pathway by diminishing the levels of oxysterol-binding protein-related protein 8 (ORP8), a recently identified regulator of insulin activity (Fig. 1) [112]. Myostatin, commonly acknowledged for its potent control over muscle growth and size, is implicated in metabolic processes. Although research indicates a potential correlation between myostatin and insulin resistance, the intricate details of this association remain under investigation [113]. Overall, Serum levels of FST are generally higher in those with T2DM. The underlying process, on the other hand, is still a mystery.

#### Leukocyte cell-derived chemotaxin-2 (LECT2)

This protein exhibits hormone-like properties and was initially identified as a chemokine influencing the regulation of neutrophil movement [114]. Subsequently, it garnered further characterization as chondromodulin II (CHM2), attributed to its capacity to stimulate proteoglycan synthesis by chondrocytes and facilitate cartilage growth [115]. Substantial progress has been achieved in comprehending its multifaceted roles, spanning liver regeneration, immune system modulation, bone development, neuronal growth, glucose regulation, metabolic syndrome, cancer, and amyloidosis [116]. While hepatocytes constitute the primary source of production and release of LECT2 into the bloodstream, its presence is also discernible in diverse cell types, encompassing vascular endothelial cells, smooth muscle cells, cerebral nerve cells, and adipocytes [117, 118]. LECT2, identified as a hepatokine, has been the subject of research establishing a direct correlation between hepatic LECT2 mRNA levels and body mass index (BMI). This association implies that elevated mRNA levels of hepatic LECT2 are associated with the severity of obesity in humans, potentially leading to insulin resistance in skeletal muscle [119, 120]. Glucose and insulin loading tests on Lect2-/- mice revealed reduced blood glucose levels following glucose or insulin administration. Furthermore, Lect2-/- mice exhibited heightened insulin-stimulated Akt phosphorylation specifically in skeletal muscle, with no discernible impact in the liver or adipose tissue. These observations suggest that the absence of the hepatokine LECT2 enhances insulin sensitivity in rodent skeletal muscle [119]. Moreover, as displayed in Fig. 1, the overproduction of LECT2 in the liver may contribute to JNK phosphorylation, resulting in insulin resistance in the skeletal muscle of obese individuals. However, the precise mechanism underlying how LECT2 facilitates JNK phosphorylation remains incompletely elucidated [120]. Additionally, LECT2 is involved in the body's inflammation response by attaching to cell surface receptors like CD209a, Met, and Tie1. Additionally, in NAFLD, this hepatokine enhances the polarization of M2 macrophages and influences liver inflammation by impacting the JNK signaling pathway (Fig. 2) [121]. In summary, LECT2 emerges as a causative factor in inducing insulin resistance in skeletal muscles.

#### Hepassocin

Hepassocin, also recognized as hepatocyte-derived fibrinogen-related protein 1 (HFREP1) or fibrinogen-like protein 1, stands out as a specialized factor exerting mitogenic effects, particularly fostering the proliferation of liver cells (in an autocrine manner) [122–124]. Its primary source of secretion is the liver, but noteworthy findings indicate its expression within brown adipose tissues as well. Following liver damage, some signals stimulate an upregulation of its expression in brown adipose tissues [125]. While an investigation observed elevated levels of hepassocin in overweight or obese individuals compared to those with normal weight, the precise nature of the association between hepassocin and obesity remains elusive [126, 127]. As outlined in a study, this hepatokine assumes a pivotal role in the development of NAFLD, contributing to hepatic lipid accumulation through the activation of an ERK1/2-dependent mechanism

[128]. Conversely, a reduction in hepassocin levels correlates with heightened insulin sensitivity, achieved through the modulation of ERK1/2 activity within the liver. This implies that elevated hepassocin concentrations may escalate the susceptibility to insulin resistance and diabetes [129]. Furthermore, hepassocin instigates insulin resistance in skeletal muscle cells through a 396 EGFR/JNK-mediated mechanism [130]. Nevertheless, conflicting perspectives arise from certain studies, challenging the earlier postulates. One investigation suggests a potentially beneficial impact of hepassocin on liver fat accumulation, coupled with a partial mitigation of hepatic apoptosis, fibrosis, and inflammation through the inhibition of oxidative stress. If the liver is injured, an increase in hepassocin in the liver can protect the liver against inflammation, steatosis, fibrosis and cell death. Therefore, the production of inflammatory cytokines such as IL-1 $\beta$ , IL-6, and TNF $\alpha$  increased in the mice where the expression of hepasosin was knocked down (Fig. 2) [121]. This, in turn, proved instrumental in preventing the development of steatohepatitis in mice [131].

# Secreted modular calcium-binding protein 1 (SMOC1)

SMOC-1, secreted by hepatocytes, encompasses one N-terminal follistatin-like (FS), one extracellular calcium (EC), two thyroglobulin-like (TY) domains, and a distinctive domain lacking known homologs. Its expression is widespread across diverse tissues, frequently positioning it in proximity to the cell's basement membrane [132, 133]. This hepatokine demonstrates interaction capabilities with laminins [132], C-reactive protein (CRP), fibulin-1, vitronectin [134], transglutaminase 2 [135], and tenascin-C [132, 136]. These interactions substantiate its involvement in integrin-matrix interactions and cell adhesion [137]. Through an acute intraperitoneal injection in mice, SMOC1 demonstrated the capacity to enhance insulin sensitivity and regulate glucose without influencing insulin secretion. The mechanism underlying its favorable glycemic effects involves the inhibition of adenosine 3',5'-cyclic monophosphate (cAMP)-cAMP-dependent protein kinase (PKA)cAMP response element-binding protein (CREB) signaling in the liver. Figure 1 depicts, this, in turn, led to a downregulation of gluconeogenic gene expression and a suppression of hepatic glucose production [138]. Additionally, individuals characterized by obesity and insulin resistance exhibited lower circulating levels of SMOC1, with a discernible association with both hepatic and systemic insulin sensitivity [138]. Overall, SMOC1 seems to manifest beneficial effects in the context of T2DM. In addition, SMOC1 is involved in the inflammatory processes of kidney diseases. A prior study showed that interleukin 1 beta reduces the expression of SMOC1 in animal mesangial cells by stimulating the production of nitric oxide (NO). Consequently, blocking the function of nitric oxide synthases results in an elevation of SMOC1 expression, exacerbating inflammation and fibrin deposition in the animal's glomerulus. Furthermore, decreasing SMOC1 expression reduces the activity of the TGF- $\beta$  signaling pathway, subsequently leading to a decline in the activity and expression of the Smad gene [139].

# Insulin-like growth factor 1 (IGF-1)

This hepatokine, referred to as somatomedin C, is a hormone exhibiting structural similarities to insulin and holds pivotal significance in the growth processes during childhood. The liver (hepatocytes) serves as the primary site for IGF-1 synthesis, a response elicited by the stimulation of GH. The majority of IGF-1 is bound to one of six binding proteins (IGF-BP) [140]. IGF-1 exerts a comprehensive influence on systemic body growth, displaying growth-promoting effects across diverse cell types such as skeletal muscle, cartilage, bone, liver, kidney, nerve, skin, hematopoietic, and lung cells. In addition to its insulin-like effects, IGF-1 plays a regulatory role in cellular DNA synthesis [141]. GH engages with the cell-surface GH receptor (GHR), a homodimeric transmembrane protein. As showed in Fig. 1, this interaction initiates signal transduction by recruiting and activating cytosolic Janus Kinase 2 (JAK2) [142].

In individuals with T2DM, IGFBP1 exhibits an elevation and possesses the capacity to counteract insulin's hypoglycemic effects by inhibiting IGF1 [143]. There exists compelling evidence indicating that insulin resistance, an increased susceptibility to metabolic syndrome and cardiovascular disease, along with diminished plasma levels of IGF1, collectively serve as prognostic indicators for the onset of type 2 diabetes [144–146]. Furthermore, certain studies propose that the modulation of IGF1 significantly influences glucose metabolism in mice subjected to a regular or high-fat diet. For instance, the systemic overexpression of IGFBP1 led to heightened plasma insulin levels, effectively mitigating the hypoglycemic impact of endogenous IGF1. However, this intervention also induced a reduction in insulin-stimulated glucose transport and glycogen synthesis in skeletal muscle, indicative of a subtle degree of insulin resistance [147]. Investigations have also unveiled a reduction in IGF levels among individuals afflicted with NAFLD and obesity [148]. A noteworthy observation arises from mice lacking Stat5a/b specifically in the central nervous system, denoted as Stat5NKO mice. These mice exhibit pronounced obesity, accompanied by insulin resistance, hyperphagia, hyperleptinemia, and a diminished capacity to respond thermally to cold stimuli [149]. In the realm of energy homeostasis regulation, a prominent player is UCP1 within Brown Adipose Tissue (BAT) activity. Comparative analyses between BAT from AAV-Igf1 mice (subjected to adeno-associated virus (AAV)-mediated delivery of IGF-I cDNA) and control mice (those injected with empty AAV) elucidated a marked augmentation in UCP1 expression, as discerned through Western blotting assays [150].

Robust neuroanatomical and functional evidence substantiates the involvement of the sympathetic nervous system (SNS) in the innervation and regulation of both WAT and BAT, orchestrating processes essential for thermoregulation and the generation of beige adipocytes [151]. The pivotal protein UCP1 in BAT, crucial for initiating thermogenesis in response to dietary and cold stimuli, is subject to control by SNS activity [152]. Intriguingly, the aforementioned findings suggest that the central activity of IGF1 can enhance central sympathetic outflow, consequently influencing UCP1 expression [150]. Beyond its role in improving insulin sensitivity and glucose tolerance, as well as fostering energy expenditure through thermogenesis, central IGF1 is implicated in the regulation of appetite [150]. Notably, administration of IGF1 has been associated with a boosted insulin sensitivity and lowered blood glucose levels in both individuals with T2DM and those without diabetes [150, 153, 154]. Moreover, Obese individuals have enlarged fat cells that, in collaboration with immune cells, elevate the production of pro-inflammatory cytokines like IL-6 and TNF- $\alpha$ . In such cases, the rise in serum IGF1 levels can serve as an anti-inflammatory agent and help regulate the levels of inflammatory factors (Fig. 2) [155].

#### Growth differentiation factor 15 (GDF15)

The protein secreted by hepatocytes, also known as macrophage inhibitory cytokine-1 (MIC-1), is a constituent of the TGF- $\beta$  superfamily [9, 156]. In physiological condition, its regulatory influence extends to hematopoietic growth, energy equilibrium, adipose tissue metabolism, somatic growth, bone remodeling, and responsiveness to stress signals [9]. GDF15 assumes a multifaceted role in various pathological conditions, encompassing cancer, cardiometabolic disorders, etc. [9]. While GDF15 is typically absent in reproductive organs, it can be induced in numerous cell types under stress conditions [156]. A critical aspect of its operation is the regulation of energy balance, which is accomplished by increasing energy expenditure and decreasing body weight gain [4].

In mice with MIC-1/GDF15-induced anorexia/cachexia syndrome, glucose, insulin, and IGF-1 levels were lower [157]. In the context of NAFLD and other chronic liver disorders, there is a proportional elevation of GDF15 levels corresponding to disease severity and the presence of steatotic hepatitis. The augmentation of GDF15, either through

overexpression or the introduction of recombinant GDF15, proves efficacious in ameliorating NAFLD and mitigating obesity [158]. In the milieu of obesity, the activation of p53 in adipose tissue contributes to the secretion of proinflammatory cytokines, insulin resistance, and the onset of diabetes. Concurrently, it mediates the expression of GDF15 within adipose tissue [159]. Also, in cases of insulin resistance or obesity, the integrative stress pathway and mitochondrial unfolded protein response (UPR) become active, resulting in the attachment of CHOP and ATF4 to the promoter region, this in turn triggers the transcription of GDF15 [160]. As well as, GDF-15 is intricately involved in lipid catabolism, heat generation, and oxidative metabolism. Its presence demonstrates a preventive effect against obesity and insulin resistance arising from dietary or genetic factors. In humans, there is a direct correlation between the expression of the GDF15 gene and the expression of genes associated with inflammation. It is also plausible that GDF15 may be linked to chronic low-grade inflammation, although the precise molecular mechanism for this connection has not yet been determined. When obese mice were given GDF15 antibody, there was an increase in inflammatory markers such as IL-6, CD-68, Tnf, monocyte chemoattractant protein-1 (Mcp1), and F480 in adipose tissue (Fig. 2). However, more research is needed to fully understand the relationship between GDF15 and inflammatory factors. Consequently, GDF15 emerges as a promising therapeutic target for addressing obesity and insulin resistance [161].

## Lipocalin-13 (LCN13)

The lipocalin family encompasses a diverse array of small secreted proteins, typically comprising 160-180 amino acids and featuring an N-terminal signal peptide. LCN13 is produced in various tissues such as the hepatocytes, epididymis, pancreas, and skeletal muscle, and it is released into the blood circulation [162]. The distinctive biological activity exhibited by LCN family members is intricately linked to their capacity to selectively bind to diminutive hydrophobic molecules, including phospholipids, fatty acids, steroids, retinol, and pheromones. Research has illuminated the expansive role of LCN family members in governing chemical communication, reproduction, immunological responses, and the progression of cancer in rats. Recent revelations underscore the newfound significance of LCNs in modulating food metabolism and insulin sensitivity among individuals grappling with obesity. Among these, LCN13 stands out, being released into the circulation of mice and expressed by various organs such as the liver, pancreas, epididymis, and skeletal muscle [163]. Notably, LCN13 levels exhibit an inverse correlation with obesity,

and interventions involving LCN13 in mice afflicted with hereditary or diet-induced obesity have demonstrated a reversal of LCN13 deficiency. This reversal, in turn, leads to improvements in insulin resistance, glucose intolerance, hepatic steatosis, hyperglycemia, and hyperinsulinemia. Mechanistically, LCN13 directly impedes hepatic lipogenesis and gluconeogenesis in hepatocytes, while concurrently promoting fatty acid  $\beta$ -oxidation. LCN13 also enhances adipocytes' responsiveness to insulin. A thorough exploration of the underlying processes contributing to LCN13's antidiabetic and anti-steatotic effects is currently a subject of discourse [163].

Concerning glucose metabolism, the expression and secretion patterns of LCN13 in mice exhibit variability in response to alterations in metabolic conditions. There is an inconsistency in LCN13 expression, which diminishes in the context of obesity. Mice subject to genetic (db/db) or high-fat diet (HFD) conditions display a substantial reduction in circulating LCN13 levels, contributing to the progression of obesity. Remarkably, hepatic LCN13 operates in an autocrine/paracrine manner, exerting inhibitory effects on glucose synthesis within the liver. Even in the absence of insulin, LCN13 facilitates glucose absorption in adipocytes while concurrently suppressing glucose synthesis and the expression of key gluconeogenic genes, such as phosphoenolpyruvate carboxykinase and glucose-6-phosphatase, in primary hepatocytes. These observations suggest an alternate, insulin-independent mechanism through which LCN13 modulates glucose metabolism [164].

Shifting focus to lipid metabolism, there is a plausible mechanism suggesting that LCN13 plays a preventive role in obesity-related hepatic steatosis. This preventive action is primarily achieved through the enhancement of fatty acid β-oxidation and the concurrent reduction of lipogenesis within the liver (Fig. 1). LCN13 and its analogs exhibit therapeutic potential for the treatment of T2DM and NAFLD, owing to their dual properties of anti-steatosis and anti-diabetic effects [163]. It is hypothesized that LCN13 orchestrates lipid metabolism by upregulating the expression of carnitine palmitoyltransferase-1 $\alpha$  (CPT1 $\alpha$ ) in the liver, while concurrently downregulating the expression of key lipogenic genes such as PPARy and carbohydrate response element binding protein (ChREBP) (Fig. 1). This dual regulation serves to mitigate lipogenesis and augment fatty acid  $\beta$ -oxidation [165]. In the context of obese mice, LCN13 demonstrates a notable reduction in hepatic steatosis without a concomitant impact on adiposity, suggesting a preferential inhibition of lipogenesis and promotion of fatty acid oxidation specifically within hepatocytes, as opposed to adipocytes. Additionally, LCN13 directly facilitates glucose absorption by adipocytes through pathways that are both insulin-dependent and insulin-independent [164]. Although

the level of LCN13 expression through the effect of Akt S473 phosphorylation and Glut4 expression can affect the insulin signaling pathway in the liver and skeletal muscles, its effect on the expression of inflammatory cytokines has not been observed [166].

# Tsukushi (TSK)

TSK, secreted by hepatocytes, stands as an atypical member within the small leucine-rich proteoglycan family, exerting regulatory influence over developmental processes in diverse organisms [167]. TSK has been associated with the process of neurogenesis, the formation of the anterior commissure, the development of the eyes and inner ear, as well as bone growth and maintaining liver homeostasis [168]. This hepatokine, induced by both obesity and exposure to cold conditions, emerges as a significant player in the orchestration of physiological responses. The prevalence of liver steatosis, commonly associated with obesity and acute cold exposure, exhibits a high correlation with circulating TSK levels in various mouse models. Intriguingly, both inflammation and endoplasmic reticulum stress, implicated in excessive hepatic lipid accumulation, serve as stimulants for the expression of TSK in the liver [169]. A study by Wang et al. has demonstrated that the deletion of TSK amplifies sympathetic innervation and thermogenesis in BAT, contributing to a protective effect against diet-induced obesity and an enhancement in glucose regulation in mice [170]. TSK deficiency also heightens adrenergic action and augments thermogenic stimulation in brown fat by promoting the phosphorylation of hormone-sensitive lipase (HSL) and Protein Kinase A (PKA) substrates [171].

The interplay between Tsk and inflammation has been elucidated, with Lipopolysaccharide (LPS), a component of gram-negative bacteria, demonstrating an ability to elevate TSK levels in mice. This observation suggests a direct link between inflammation and the induction of Tsk expression [172]. Notably, deficiency in TSK has been associated with an increase in protein levels of UCP1 [169]. Liver cells respond to proinflammatory cytokines, such as TNF-a, IL-1 $\beta$ , and IFN- $\gamma$ , collectively promoting the transcription of TSK [169]. The initial surge in TSK levels may serve a protective role in stressed liver cells by mitigating cholesterol influx into the liver. However, the prolonged elevation of TSK observed in conditions like obesity and NAFLD raises concerns about its potential contribution to atherosclerosis, potentially attributed to the chronic constriction of reverse cholesterol transport [172].

# Adropin

Hepatocytes and brain are two of the many tissues that express adropin, a small peptide with 76 amino acids. Its presence has been noted in proteins of the pancreas, heart, blood vessels, and kidneys as well [173-175]. This peptide exhibits a decrease in response to heightened lipid availability in the liver. Furthermore, individuals with obesity and T2DM manifest a decline in serum adropin levels [5]. Mouse hepatic steatosis reduces energy homeostasis-associated gene (ENHO) gene expression, which encodes adropin [176]. Conversely, in hepatic steatosis induced by a lipidrich diet, an upregulation of adropin expression is observed [177]. An intriguing observation reveals a positive correlation between adropin and phosphoenolpyruvate carboxykinase 1 (PCK1), a pivotal regulatory point in gluconeogenesis [178]. In addition, Adropin is used as a therapeutic stimulus to increase glucose tolerance and insulin sensitivity in rats induced by high-fat diet [179]. Furthermore, aside from its involvement in the body's metabolic processes, adropin also has a function in reducing inflammatory factors in diabetes. Adropin is able to prevent the attachment of THP1 monocytes to Human Umbilical Vein Endothelial Cells (HUVEC) cells by affecting the TNF- $\alpha$  signaling pathway, thus exhibiting its anti-inflammatory properties (Fig. 2) [180].

#### Activins

Activins, members of the TGF $\beta$  superfamily, constitute a diverse group of proteins with multifaceted effects on various tissues. Traditionally perceived as heterogeneous, these proteins consist of multiple BA and BB subunits interconnected by disulfide bridges. Their pivotal role in the formation of gonadal follicles was initially discerned. Recent investigations have unveiled two additional subunits, namely  $\beta E$  and  $\beta C$ , characterized by sequences akin to their classical counterparts. Intriguingly, despite their initial association with gonads, it has been established that the hepatocytes serves as the primary source of activins [181, 182]. This protein instigates a cascade of signaling events by binding to the Activin A receptors type I (ActRI) and type II (ActRII), characterized as serine-threonine kinases. These signals subsequently culminate in the activation of genes with diverse functionalities [183].

In 2023, Liu and colleagues uncovered that the expression of Activin A in the liver led to a decrease in inflammation, expansion of hematopoietic stem cells, reduction of liver steatosis, decreased fat accumulation and lower levels of circulating cholesterol. These findings suggested that these effects collectively contribute to the observed protection against atherosclerosis and metabolic related diseases [184]. Moreover, in murine models, the analysis of activin E mRNA levels within hepatocytes' cytosol suggests a plausible involvement in both exacerbating insulin resistance associated with obesity and facilitating thermogenesis in brown adipose tissue [185]. In a study, it has been shown that overexpression of hepatic activin E led to the activation of thermogenesis by upregulating Ucp1 in both white and brown adipose tissues. Furthermore, hepatic activin E-transgenic mice showed enhanced insulin sensitivity. Additionally, experiments conducted in vitro indicated that activin E directly promoted the expression of Ucp1 and FGF21, potentially through the mediation of transforming growth factor-b or activin type I receptors [186]. Within this gene cohort, UCP1 holds significance, encoding uncoupler proteins that induce the leakage of hydrogen ions into the mitochondrial space, consequently generating heat within the targeted cell and tissue. Another noteworthy gene in this category is FGF21, contributing to the growth and specialization of brown adipocytes [187]. During fasting, the protein level of FGF21 is elevated, with its suppression attributed to the absence of an insulin receptor [188]. In contrast, there is an observed elevation in the level of activin E in mice experiencing obesity due to a high-fat diet [6].

Additionally, a prior study explored the inflammatory function of Activin in umbilical vein endothelial cells. As depicted in Fig. 2, the findings indicated that this hepatokine impacts the expression of chemokines and inflammatory cytokines by influencing NF- $\kappa$ B and MAPK signaling pathways. As a result, it exerts an anti-inflammatory effect against TNF- $\alpha$  in these cells [189].

#### Selenoproteins

Selenocysteine, an amino acid implicated in oxidationreduction processes, is found in selenoprotein P [190, 191], which is primarily secreted from the hepatocytes in mammals, selenoprotein P may also be released into the bloodstream by the testis and brain tissue [192]. In addition, Selenoprotein S is involved in different metabolic conditions in the body, including insulin resistance, diabetes, and obesity. When selenoprotein S is not expressed in liver cells, it can worsen obesity and lead to reduced insulin sensitivity and glucose tolerance [193]. Human studies suggest a positive correlation between SELENOP and NAFLD [194]. The expression of this protein escalates in response to endoplasmic reticulum stress and the activation of the JNK protein. Furthermore, SELENOP synthesis is triggered by the activation of AMPK, which can be achieved by the use of antiinflammatory or insulin-sensitizing medications (Fig. 1) [195, 196]. In the context of T2DM, there is an observed elevation in the mRNA level of SELENOP. In-vitro investigations indicate that SELENOP dysregulates glucose metabolism [197, 198]. While selenoprotein-S is involved in controlling the release of inflammatory cytokines, selenoprotein-P functions as a regulator of homeostasis and a transporter of Se from the liver to other tissues in the body [199, 200].

# Hepcidin

Hepcidin, the primary regulator of iron homeostasis in the body, is a polypeptide hormone that primarily originates from liver hepatocytes. Hepcidin acts as an iron regulator by affecting the transfer of iron to the plasma through the action of ferroportin, reducing the amount of iron absorption from the diet and increasing cellular iron storage [201, 202]. Iron metabolism is one of the factors that influences the functioning of the endocrine system. Ferroptosis, a form of iron-dependent cell death, leads to insulin resistance in fat, liver, and muscle cells, and also reduces insulin secretion in pancreatic  $\beta$  cells [203]. Dysregulation of hepcidin leads to various metabolic diseases such as obesity, type 2 diabetes, and insulin resistance. Additionally, the plasma concentration of hepcidin changes in acute and chronic inflammatory conditions, and by affecting macrophages, it leads to the release of inflammatory cytokines [201, 202].

Iron metabolism plays a crucial role in blood glucose homeostasis through its effects on fat metabolism, insulin secretion, and liver metabolism [203]. Uncontrolled iron metabolism in adipose tissue and increased liver ferroptosis can lead to the development of diabetes and insulin resistance, respectively [203]. Previous studies have shown that individuals with T2DM tend to have higher levels of hepcidin compared to those without the condition [201, 204]. Additionally, inactivation of the hepcidin-coding gene in liver cells results in the activation of Akt and glycogen synthase kinase-3 $\beta$  (GSK3 $\beta$ ) in the insulin signaling pathway [205]. Protein kinase Akt, by activating GSK3β and increasing the expression of GLUT4, leads to enhanced glucose uptake by cells and a reduction in insulin resistance [206, 207]. Increased iron levels can also lead to elevated blood glucose, fasting insulin, and hepatic ferroptosis. Through the JAK2/STAT3 signaling pathway, this can then result in the activation of SOCS-1 and the development of insulin resistance [208, 209]. Considering that hepcidin is an acutephase reactant, the inflammation associated with T2DM can lead to increased secretion of IL-6 and activin-B (Act-B), which in turn stimulate the synthesis of hepcidin in the liver via the STAT3 signaling pathway [204].

### Lipopolysaccharide binding protein

Lipopolysaccharide binding protein (LBP) is an acute-phase protein that plays a crucial role in the lipopolysaccharide (LPS) transport cascade. LBP is primarily produced by hepatocytes and, to a lesser extent, in non-hepatic tissues such as intestinal epithelial cells, lungs, and gum tissue, also, it is present in low amounts in the blood circulation. During the acute-phase response to infection and inflammation, LBP's concentration increases 10 to 20 times compared to its baseline level [210, 211].

Lipopolysaccharide is a pathogenic factor found in the outer membrane of Gram-negative bacteria, commonly known as endotoxin [211]. Since lipopolysaccharide primarily exerts its endotoxic effects by activating the innate immune system, it is not inherently toxic [212]. However, the entry and increased level of lipopolysaccharide in the bloodstream can lead to a condition called endotoxemia, which can occur due to unhealthy eating habits or digestive disorders (derived from the intestinal microbiome). This endotoxemia can cause disorders such as metabolic endotoxemia (even in healthy individuals) and chronically contribute to inflammation-related disorders, including cardiovascular diseases, liver diseases, insulin resistance, metabolic syndromes, obesity, and diabetes [213].

LBP plays a crucial role in the LPS-to-TLR4 signaling cascade. LBP can form high-affinity complexes with LPS and then bind to CD14. LPS interacts with the LRR13-LRR15 region of TLR4, gets transferred from CD14 to MD2, and leads to signal generation, thereby greatly enhancing the host's response to extracellular LPS [214]. Activation of the TLR4 pathway triggers the production of pro-inflammatory cytokines, such as TNF- $\alpha$  and IL-6. These pro-inflammatory cytokines can contribute to insulin resistance by interfering with insulin signaling pathways [215].

LBP is involved in the metabolism of lipids, particularly triglycerides and high-density lipoprotein (HDL) cholesterol. Clinical studies on metabolic endotoxemia in obese patients have shown that the serum level of LPS increases with a high-fat diet. As a result, obesity is considered a state of low-grade chronic inflammation [216, 217]. The signaling cascade initiated by the binding of LBP to LPS leads to an inflammatory response and modulation of lipid metabolism. These changes in lipid metabolism can affect insulin sensitivity and contribute to insulin resistance [87, 210, 218].

# Conclusion

Recent investigations posit that the disorder in hepatokines significantly contributes to the genesis of insulin resistance and inflammation culminating in the onset of T2DM. In the context of this study, we scrutinized various hepatokines and explored their impacts on insulin resistance and inflammation. It is evident that the involvement of hepatokines stems from their modulation of key metabolic mechanisms, including lipogenesis, lipolysis, and the transfer of lipoproteins, insulin-dependent sugar metabolism and the modulation of inflammatory pathways. Briefly, fetuin-A facilitates the development of lipid-induced insulin resistance through its role as an endogenous ligand for TLR-4. FGF21 mitigates inflammation in diabetes by impeding the nuclear translocation of NF-kB in adipocytes and adipose tissue under insulin-resistant conditions, concurrently augmenting glucose metabolism. Moreover, the overproduction of LECT2 in the liver may contribute to JNK phosphorylation, resulting in insulin resistance in the skeletal muscle of obese individuals. ANGPTL6 enhances AMPK and boosts insulin signaling in muscle. Additionally, it suppresses gluconeogenesis by inhibiting the expression of the G6P gene. Furthermore, ANGPTL8 may enhance insulin sensitivity by directly affecting Akt phosphorylation. More importantly, Follistatin possesses the potential to influence insulin resistance and inflammation by virtue of its interaction with members of the TGF-β family. Also, Hepassocin assumes a pivotal role in the development of NAFLD, contributing to hepatic lipid accumulation through the activation of an ERK1/2-dependent mechanism. Furthermore, hepassocin instigates insulin resistance in skeletal muscle cells through a 396 EGFR/ JNK-mediated mechanism. SMOC1 demonstrated the capacity to enhance insulin sensitivity and regulate glucose, as well. The mechanism underlying its favorable glycemic effects involves the inhibition of adenosine 3',5'-cyclic monophosphate (cAMP)-cAMP-dependent protein kinase (PKA)-cAMP response element-binding protein (CREB) signaling in the liver. This, in turn, led to a downregulation of gluconeogenic gene expression and a suppression of hepatic glucose production. Intriguingly, the central activity of IGF1 can enhance central sympathetic outflow, consequently influencing UCP1 expression. Beyond its role in improving insulin sensitivity and glucose tolerance, as well as fostering energy expenditure through thermogenesis, central IGF1 is implicated in the regulation of appetite. In addition, LCN13 orchestrates lipid metabolism by upregulating the expression of CPT1 $\alpha$  in the liver, while concurrently downregulating the expression of key lipogenic genes such as PPARy and ChREBP. Liver cells respond to proinflammatory cytokines, such as TNF- $\alpha$ , IL-1 $\beta$ , and IFN- $\gamma$ , collectively promoting the transcription of TSK. The initial surge in TSK levels may serve a protective role in stressed liver cells by mitigating cholesterol influx into the liver. An intriguing observation reveals a positive correlation between adropin and phosphoenolpyruvate carboxykinase 1 (PCK1), a pivotal regulatory point in gluconeogenesis. The prospect is optimistic that harnessing the potential of hepatokines and manipulating the pertinent pathways through the use of agonists and antagonists could pave the way for enhancing human health and advancing the treatment of T2DM and

fatty liver disease. In addition, research in the field of hepatokines may help for the pathomechanism-based clustering of insulin resistance in some diseases to better implement precision medicine in clinical practice [18]. However, more complementary studies are needed in this regard.

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#### Declarations

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# References

- 1. Meigs JB (2003) Epidemiology of the insulin resistance syndrome. Curr Diab Rep 3(1):73–79
- Roden M, Shulman GI (2019) The integrative biology of type 2 diabetes. Nature 576(7785):51–60
- Lebovitz HE (2001) Insulin resistance: definition and consequences. Exp Clin Endocrinol Diabetes 109(Suppl 2):S135–S148
- Jensen-Cody SO, Potthoff MJ (2021) Hepatokines and metabolism: deciphering communication from the liver. Mol Metab 44:101138
- Watt MJ, Miotto PM, De Nardo W, Montgomery MK (2019) The liver as an endocrine organ-linking NAFLD and insulin resistance. Endocr Rev 40(5):1367–1393
- Adam RC, Pryce DS, Lee JS, Zhao Y, Mintah IJ, Min S et al (2023) Activin E-ACVR1C cross talk controls energy storage via suppression of adipose lipolysis in mice. Proc Natl Acad Sci U S A 120(32):e2309967120
- Kliewer SA, Mangelsdorf DJ (2019) A dozen years of discovery: insights into the physiology and pharmacology of FGF21. Cell Metabol 29(2):246–253
- Min L, Xiang J, Wang B, Ye C, Su X (2023) Novel insights of ANGPTL-3 on modulating cholesterol efflux Capacity Induced by HDL particle. Curr Mol Med
- Siddiqui JA, Pothuraju R, Khan P, Sharma G, Muniyan S, Seshacharyulu P et al (2022) Pathophysiological role of growth differentiation factor 15 (GDF15) in obesity, cancer, and cachexia. Cytokine Growth Factor Rev 64:71–83

- Govender N, Khaliq OP, Moodley J, Naicker T (2021) Insulin resistance in COVID-19 and diabetes. Prim Care Diabetes 15(4):629–634
- 11. Pei J, Wang B, Wang D (2022) Current studies on molecular mechanisms of insulin resistance. Journal of Diabetes Research. ;2022
- Lee S-H, Park S-Y, Choi CS (2022) Insulin resistance: from mechanisms to therapeutic strategies. Diabetes Metabolism J 46(1):15–37
- Saltiel AR, Kahn CR (2001) Insulin signalling and the regulation of glucose and lipid metabolism. Nature 414(6865):799–806
- Czech MP (2017) Insulin action and resistance in obesity and type 2 diabetes. Nat Med 23(7):804–814
- Pal D, Dasgupta S, Kundu R, Maitra S, Das G, Mukhopadhyay S et al (2012) Fetuin-A acts as an endogenous ligand of TLR4 to promote lipid-induced insulin resistance. Nat Med 18(8):1279–1285
- Stefan N, Häring H-U (2013) Circulating fetuin-A and free fatty acids interact to predict insulin resistance in humans. Nat Med 19(4):394–395
- 17. Shulman GI (2014) Ectopic fat in insulin resistance, dyslipidemia, and cardiometabolic disease. N Engl J Med 371(12):1131–1141
- Stefan N, Schick F, Birkenfeld AL, Häring H-U, White MF (2023) The role of hepatokines in NAFLD. Cell Metabol 35(2):236–252
- Cayatte AJ, Kumbla L, Subbiah MT (1990) Marked acceleration of exogenous fatty acid incorporation into cellular triglycerides by fetuin. J Biol Chem 265(10):5883–5888
- Chattopadhyay D, Das S, Guria S, Basu S, Mukherjee S (2021) Fetuin-A regulates adipose tissue macrophage content and activation in insulin resistant mice through MCP-1 and iNOS: involvement of IFNγ-JAK2-STAT1 pathway. Biochem J 478(22):4027–4043
- Afrisham R, Sadegh-Nejadi S, Meshkani R, Emamgholipour S, Paknejad M (2020) Effect of circulating exosomes derived from normal-weight and obese women on gluconeogenesis, glycogenesis, lipogenesis and secretion of FGF21 and fetuin A in HepG2 cells. Diabetol Metab Syndr 12:1–11
- Wang N, Xu TY, Zhang X, Li JY, Wang YX, Guo XC et al (2018) Improving hyperglycemic effect of FGF-21 is associated with alleviating inflammatory state in diabetes. Int Immunopharmacol 56:301–309
- 23. Rui L (2014) Energy metabolism in the liver. Compr Physiol 4(1):177–197
- Azimifar SB, Nagaraj N, Cox J, Mann M (2014) Cell-typeresolved quantitative proteomics of murine liver. Cell Metab 20(6):1076–1087
- Kim MS, Pinto SM, Getnet D, Nirujogi RS, Manda SS, Chaerkady R et al (2014) A draft map of the human proteome. Nature 509(7502):575–581
- Choi KM (2016) The impact of Organokines on insulin resistance, inflammation, and atherosclerosis. Endocrinol Metab (Seoul) 31(1):1–6
- Meex RCR, Watt MJ (2017) Hepatokines: linking nonalcoholic fatty liver disease and insulin resistance. Nat Rev Endocrinol 13(9):509–520
- Carbone C, Piro G, Merz V, Simionato F, Santoro R, Zecchetto C et al (2018) Angiopoietin-like proteins in angiogenesis, inflammation and Cancer. Int J Mol Sci. ;19(2)
- Hato T, Tabata M, Oike Y (2008) The role of angiopoietin-like proteins in angiogenesis and metabolism. Trends Cardiovase Med 18(1):6–14
- Wu SA, Kersten S, Qi L (2021) Lipoprotein Lipase and its regulators: an Unfolding Story. Trends Endocrinol Metab 32(1):48–61
- Wang H, Eckel RH (2009) Lipoprotein lipase: from gene to obesity. Am J Physiol Endocrinol Metab 297(2):E271–E288
- 32. Yang J, Song QY, Niu SX, Chen HJ, Petersen RB, Zhang Y et al (2022) Emerging roles of angiopoietin-like proteins in

inflammation: mechanisms and potential as pharmacological targets. J Cell Physiol 237(1):98–117

- 33. Yang J, Song Qy N, Sx C, Hj, Petersen RB, Zhang Y et al (2022) Emerging roles of angiopoietin-like proteins in inflammation: mechanisms and potential as pharmacological targets. J Cell Physiol 237(1):98–117
- Carbone C, Piro G, Merz V, Simionato F, Santoro R, Zecchetto C et al (2018) Angiopoietin-like proteins in angiogenesis, inflammation and cancer. Int J Mol Sci 19(2):431
- 35. Yan Q, Jiang L, Liu M, Yu D, Zhang Y, Li Y et al (2017) ANG-PTL1 interacts with integrin α1β1 to suppress HCC angiogenesis and metastasis by inhibiting JAK2/STAT3 signaling. Cancer Res 77(21):5831–5845
- 36. Chen HA, Kuo TC, Tseng CF, Ma JT, Yang ST, Yen CJ et al (2016) Angiopoietin-like protein 1 antagonizes MET receptor activity to repress sorafenib resistance and cancer stemness in hepatocellular carcinoma. Hepatology 64(5):1637–1651
- 37. Tabata M, Kadomatsu T, Fukuhara S, Miyata K, Ito Y, Endo M et al (2009) Angiopoietin-like protein 2 promotes chronic adipose tissue inflammation and obesity-related systemic insulin resistance. Cell Metabol 10(3):178–188
- Horio E, Kadomatsu T, Miyata K, Arai Y, Hosokawa K, Doi Y et al (2014) Role of endothelial cell–derived Angptl2 in vascular inflammation leading to endothelial dysfunction and atherosclerosis progression. Arteriosclerosis, thrombosis, and vascular biology. 34(4):790–800
- Farhat N, Thorin-Trescases N, Mamarbachi M, Villeneuve L, Yu C, Martel C et al (2013) Angiopoietin-like 2 promotes atherogenesis in mice. J Am Heart Association 2(3):e000201
- 40. Nakamura M, Yamada K (1967) Studies on a diabetic (KK) strain of the mouse. Diabetologia 3(2):212–221
- Koishi R, Ando Y, Ono M, Shimamura M, Yasumo H, Fujiwara T et al (2002) Angptl3 regulates lipid metabolism in mice. Nat Genet 30(2):151–157
- 42. Shimizugawa T, Ono M, Shimamura M, Yoshida K, Ando Y, Koishi R et al (2002) ANGPTL3 decreases very low density lipoprotein triglyceride clearance by inhibition of lipoprotein lipase. J Biol Chem 277(37):33742–33748
- 43. Minicocci I, Montali A, Robciuc MR, Quagliarini F, Censi V, Labbadia G et al (2012) Mutations in the ANGPTL3 gene and familial combined hypolipidemia: a clinical and biochemical characterization. J Clin Endocrinol Metab 97(7):E1266–E1275
- 44. Wang Y, McNutt MC, Banfi S, Levin MG, Holland WL, Gusarova V et al (2015) Hepatic ANGPTL3 regulates adipose tissue energy homeostasis. Proc Natl Acad Sci U S A 112(37):11630–11635
- 45. Ono M, Shimizugawa T, Shimamura M, Yoshida K, Noji-Sakikawa C, Ando Y et al (2003) Protein region important for regulation of lipid metabolism in angiopoietin-like 3 (ANG-PTL3): ANGPTL3 is cleaved and activated in vivo. J Biol Chem 278(43):41804–41809
- Kaplan R, Zhang T, Hernandez M, Gan FX, Wright SD, Waters MG et al (2003) Regulation of the angiopoietin-like protein 3 gene by LXR. J Lipid Res 44(1):136–143
- 47. Shimamura M, Matsuda M, Ando Y, Koishi R, Yasumo H, Furukawa H et al (2004) Leptin and insulin down-regulate angiopoietin-like protein 3, a plasma triglyceride-increasing factor. Biochem Biophys Res Commun 322(3):1080–1085
- 48. Tikka A, Soronen J, Laurila PP, Metso J, Ehnholm C, Jauhiainen M (2014) Silencing of ANGPTL 3 (angiopoietin-like protein 3) in human hepatocytes results in decreased expression of gluco-neogenic genes and reduced triacylglycerol-rich VLDL secretion upon insulin stimulation. Biosci Rep 34(6):e00160
- 49. Wang C, Tong Y, Wen Y, Cai J, Guo H, Huang L et al (2018) Hepatocellular carcinoma-associated protein TD26 interacts and enhances sterol regulatory element-binding protein 1

activity to promote tumor cell proliferation and growth. Hepatology 68(5):1833–1850

- Biterova E, Esmaeeli M, Alanen HI, Saaranen M, Ruddock LW (2018) Structures of Angptl3 and Angptl4, modulators of triglyceride levels and coronary artery disease. Sci Rep 8(1):6752
- Koliwad SK, Gray NE, Wang JC (2012) Angiopoietin-like 4 (Angptl4): a glucocorticoid-dependent gatekeeper of fatty acid flux during fasting. Adipocyte 1(3):182–187
- 52. La Paglia L, Listì A, Caruso S, Amodeo V, Passiglia F, Bazan V et al (2017) Potential role of ANGPTL4 in the Cross talk between metabolism and Cancer through PPAR signaling pathway. PPAR Res 2017:8187235
- 53. Singh AK, Chaube B, Zhang X, Sun J, Citrin KM, Canfrán-Duque A et al (2021) Hepatocyte-specific suppression of ANGPTL4 improves obesity-associated diabetes and mitigates atherosclerosis in mice. J Clin Invest. ;131(17)
- 54. Yoshida K, Shimizugawa T, Ono M, Furukawa H (2002) Angiopoietin-like protein 4 is a potent hyperlipidemia-inducing factor in mice and inhibitor of lipoprotein lipase. J Lipid Res 43(11):1770–1772
- 55. Wang Y, Liu LM, Wei L, Ye WW, Meng XY, Chen F et al (2016) Angiopoietin-like protein 4 improves glucose tolerance and insulin resistance but induces liver steatosis in high-fat-diet mice. Mol Med Rep 14(4):3293–3300
- 56. Xu A, Lam MC, Chan KW, Wang Y, Zhang J, Hoo RL et al (2005) Angiopoietin-like protein 4 decreases blood glucose and improves glucose tolerance but induces hyperlipidemia and hepatic steatosis in mice. Proc Natl Acad Sci U S A 102(17):6086–6091
- 57. Altun Ö, Dikker O, Arman Y, Ugurlukisi B, Kutlu O, Ozgun Cil E et al (2018) Serum angiopoietin-like peptide 4 levels in patients with hepatic steatosis. Cytokine 111:496–499
- 58. Zuo Y, He Z, Chen Y, Dai L (2023) Dual role of ANGPTL4 in inflammation. Inflamm Res 72(6):1303–1313
- Im Cho D, Kang H-j, Jeon JH, Eom GH, Cho HH, Kim MR et al (2019) Antiinflammatory activity of ANGPTL4 facilitates macrophage polarization to induce cardiac repair. JCI Insight. ;4(16)
- 60. Alghanim G, Qaddoumi MG, Alhasawi N, Cherian P, Al-Khairi I, Nizam R et al (2019) Higher levels of ANGPTL5 in the circulation of subjects with obesity and type 2 diabetes are associated with insulin resistance. Front Endocrinol 10:461900
- 61. Fan KC, Wu HT, Wei JN, Chuang LM, Hsu CY, Yen IW et al (2020) Serum angiopoietin-like protein 6, risk of type 2 diabetes, and response to hyperglycemia: a prospective cohort study. J Clin Endocrinol Metab. ;105(5)
- 62. Kitazawa M, Ohizumi Y, Oike Y, Hishinuma T, Hashimoto S (2007) Angiopoietin-related growth factor suppresses gluconeogenesis through the Akt/forkhead box class O1-dependent pathway in hepatocytes. J Pharmacol Exp Ther 323(3):787–793
- Erkan G, Muratoglu S, Ercin U, Bilgihan A (2018) Angiopoietinlike protein 2 and angiopoietin-like protein 6 levels in patients with nonalcoholic fatty liver disease. Arch Med Sci 14(4):781–787
- 64. Tanigawa H, Miyata K, Tian Z, Aoi J, Kadomatsu T, Fukushima S et al (2016) Upregulation of ANGPTL6 in mouse keratinocytes enhances susceptibility to psoriasis. Sci Rep 6(1):34690
- 65. Zhang L, Shannon CE, Bakewell TM, Abdul-Ghani MA, Fourcaudot M, Norton L (2020) Regulation of ANGPTL8 in liver and adipose tissue by nutritional and hormonal signals and its effect on glucose homeostasis in mice. Am J Physiol Endocrinol Metab 318(5):E613–e24
- 66. Zhang Z, Wu H, Dai L, Yuan Y, Zhu Y, Ma Z et al (2020) ANG-PTL8 enhances insulin sensitivity by directly activating insulinmediated AKT phosphorylation. Gene 749:144707
- Bai Y, Du Q, Zhang L, Li L, Wang N, Wu B et al (2021) Silencing of ANGPTL8 alleviates insulin resistance in trophoblast cells. Front Endocrinol (Lausanne) 12:635321

- 68. Saghafi S, Chamani E, Salmani F, Fadaei R, Shafiei E, Moradi N et al (2023) Genetic predisposition to nonalcoholic fatty liver disease: insights from ANGPTL8 gene variants in Iranian adults. Lipids Health Dis 22(1):147
- Zhao Z, Deng X, Jia J, Zhao L, Wang C, Cai Z et al (2022) Angiopoietin-like protein 8 (betatrophin) inhibits hepatic gluconeogenesis through PI3K/Akt signaling pathway in diabetic mice. Metabolism 126:154921
- Zhang Y, Zheng L, Huang K (2018) A new way to regulate inflammation: selective autophagic degradation of IKKγ mediated by ANGPTL8. Cell Stress 2(3):66
- Zhang Y, Guo X, Yan W, Chen Y, Ke M, Cheng C et al (2017) ANGPTL8 negatively regulates NF-κB activation by facilitating selective autophagic degradation of IKKγ. Nat Commun 8(1):2164
- 72. Tan H, Yue T, Chen Z, Wu W, Xu S, Weng J (2023) Targeting FGF21 in cardiovascular and metabolic diseases: from mechanism to medicine. Int J Biol Sci 19(1):66–88
- 73. Ge X, Wang Y, Lam KS, Xu A (2012) Metabolic actions of FGF21: molecular mechanisms and therapeutic implications. Acta Pharm Sinica B 2(4):350–357
- 74. von Holstein-Rathlou S, BonDurant LD, Peltekian L, Naber MC, Yin TC, Claflin KE et al (2016) FGF21 Mediates Endocrine Control of Simple Sugar Intake and Sweet taste preference by the liver. Cell Metab 23(2):335–343
- 75. Hotta Y, Nakamura H, Konishi M, Murata Y, Takagi H, Matsumura S et al (2009) Fibroblast growth factor 21 regulates Lipolysis in White Adipose tissue but is not required for ketogenesis and triglyceride clearance in liver. Endocrinology 150(10):4625–4633
- 76. Lu W, Li X, Luo Y (2021) FGF21 in obesity and cancer: new insights. Cancer Lett 499:5–13
- Kliewer SA, Mangelsdorf DJ (2019) A Dozen years of Discovery: insights into the physiology and pharmacology of FGF21. Cell Metab 29(2):246–253
- BonDurant LD, Potthoff MJ (2018) Fibroblast growth factor 21: a Versatile Regulator of metabolic homeostasis. Annu Rev Nutr 38(1):173–196
- BonDurant LD, Ameka M, Naber MC, Markan KR, Idiga SO, Acevedo MR et al (2017) FGF21 regulates metabolism through adipose-dependent and -independent mechanisms. Cell Metab 25(4):935–44e4
- Owen BM, Ding X, Morgan DA, Coate KC, Bookout AL, Rahmouni K et al (2014) FGF21 acts centrally to induce sympathetic nerve activity, energy expenditure, and weight loss. Cell Metab 20(4):670–677
- Xie T, So WY, Li XY, Leung PS (2019) Fibroblast growth factor 21 protects against lipotoxicity-induced pancreatic β-cell dysfunction via regulation of AMPK signaling and lipid metabolism. Clin Sci (Lond) 133(19):2029–2044
- 82. Gong Q, Hu Z, Zhang F, Cui A, Chen X, Jiang H et al (2016) Fibroblast growth factor 21 improves hepatic insulin sensitivity by inhibiting mammalian target of rapamycin complex 1 in mice. Hepatology 64(2):425–438
- Goto T, Hirata M, Aoki Y, Iwase M, Takahashi H, Kim M et al (2017) The hepatokine FGF21 is crucial for peroxisome proliferator-activated receptor-α agonist-induced amelioration of metabolic disorders in obese mice. J Biol Chem 292(22):9175–9190
- Al-Mansoori L, Al-Jaber H, Prince MS, Elrayess MA (2022) Role of inflammatory cytokines, growth factors and adipokines in adipogenesis and insulin resistance. Inflammation. :1–14
- ZhuGe D-L, Javaid HMA, Sahar NE, Zhao Y-Z, Huh JY (2020) Fibroblast growth factor 2 exacerbates inflammation in adipocytes through NLRP3 inflammasome activation. Arch Pharm Res 43:1311–1324

- Jin L, Yang R, Geng L, Xu A (2023) Fibroblast growth factorbased pharmacotherapies for the treatment of obesity-related metabolic complications. Annu Rev Pharmacol Toxicol 63:359–382
- Jensen-Cody SO, Potthoff MJ (2021) Hepatokines and metabolism: deciphering communication from the liver. Mol Metabolism 44:101138
- Agarwal S, Chattopadhyay M, Mukherjee S, Dasgupta S, Mukhopadhyay S, Bhattacharya S (2017) Fetuin-A downregulates adiponectin through Wnt-PPARγ pathway in lipid induced inflamed adipocyte. Biochim Biophys Acta Mol Basis Dis 1863(1):174–181
- Komsa-Penkova RS, Golemanov GM, Radionova ZV, Tonchev PT, Iliev SD, Penkov VV (2017) Fetuin-A–alpha2-heremansschmid glycoprotein: from structure to a novel marker of chronic diseases part 1. Fetuin-A as a calcium chaperone and inflammatory marker. J Biomedical Clin Res 10(2):90–97
- Mathews ST, Chellam N, Srinivas PR, Cintron VJ, Leon MA, Goustin AS et al (2000) Alpha2-HSG, a specific inhibitor of insulin receptor autophosphorylation, interacts with the insulin receptor. Mol Cell Endocrinol 164(1–2):87–98
- Auberger P, Falquerho L, Contreres JO, Pages G, Le Cam G, Rossi B et al (1989) Characterization of a natural inhibitor of the insulin receptor tyrosine kinase: cDNA cloning, purification, and anti-mitogenic activity. Cell 58(4):631–640
- 92. Chekol Abebe E, Tilahun Muche Z, Behaile TMA, Mengie Ayele T, Mekonnen Agidew M, Teshome Azezew M et al (2022) The structure, biosynthesis, and biological roles of fetuin-A: a review. Front Cell Dev Biol 10:945287
- 93. Stefan N, Hennige AM, Staiger H, Machann J, Schick F, Kröber SM et al (2006) Alpha2-Heremans-Schmid glycoprotein/fetuin-A is associated with insulin resistance and fat accumulation in the liver in humans. Diabetes Care 29(4):853–857
- 94. Stefan N, Fritsche A, Weikert C, Boeing H, Joost H-G, Häring H-U et al (2008) Plasma fetuin-A levels and the risk of type 2 diabetes. Diabetes 57(10):2762–2767
- 95. Kroeger J, Meidtner K, Stefan N, Guevara M, Kerrison ND, Ardanaz E et al (2018) Circulating fetuin-A and risk of type 2 diabetes: a mendelian randomization analysis. Diabetes 67(6):1200–1205
- Ix JH, Wassel CL, Kanaya AM, Vittinghoff E, Johnson KC, Koster A et al (2008) Fetuin-A and incident diabetes mellitus in older persons. JAMA 300(2):182–188
- Hennige AM, Staiger H, Wicke C, Machicao F, Fritsche A, Häring HU et al (2008) Fetuin-A induces cytokine expression and suppresses adiponectin production. PLoS ONE 3(3):e1765
- Meex RC, Watt MJ (2017) Hepatokines: linking nonalcoholic fatty liver disease and insulin resistance. Nat Reviews Endocrinol 13(9):509–520
- Pan X, Kaminga AC, Chen J, Luo M, Luo J (2020) Fetuin-A and Fetuin-B in non-alcoholic fatty liver disease: a Meta-analysis and Meta-regression. Int J Environ Res Public Health. ;17(8)
- 100. Mukhopadhyay S, Mondal SA, Kumar M, Dutta D (2014) Proinflammatory and antiinflammatory attributes of fetuin-a: a novel hepatokine modulating cardiovascular and glycemic outcomes in metabolic syndrome. Endocr Pract 20(12):1345–1351
- 101. Mukhuty A, Fouzder C, Kundu R (2021) Fetuin-A secretion from β-cells leads to accumulation of macrophages in islets, aggravates inflammation and impairs insulin secretion. J Cell Sci 134(21):jcs258507
- 102. Jones KL, Mansell A, Patella S, Scott BJ, Hedger MP, de Kretser DM et al (2007) Activin A is a critical component of the inflammatory response, and its binding protein, follistatin, reduces mortality in endotoxemia. Proc Natl Acad Sci U S A 104(41):16239–16244

- 103. Parfenova OK, Kukes VG, Grishin DV (2021) Follistatin-like proteins: structure, functions and biomedical importance. Biomedicines 9(8):999
- 104. Hansen J, Rinnov A, Krogh-Madsen R, Fischer CP, Andreasen AS, Berg RM et al (2013) Plasma follistatin is elevated in patients with type 2 diabetes: relationship to hyperglycemia, hyperinsulinemia, and systemic low-grade inflammation. Diabetes Metab Res Rev 29(6):463–472
- 105. Wu C, Borné Y, Gao R, López Rodriguez M, Roell WC, Wilson JM et al (2021) Elevated circulating follistatin associates with an increased risk of type 2 diabetes. Nat Commun 12(1):6486
- 106. Hansen JS, Rutti S, Arous C, Clemmesen JO, Secher NH, Drescher A et al (2016) Circulating follistatin is liver-derived and regulated by the glucagon-to-insulin ratio. J Clin Endocrinol Metab 101(2):550–560
- 107. Sidis Y, Mukherjee A, Keutmann H, Delbaere A, Sadatsuki M, Schneyer A (2006) Biological activity of follistatin isoforms and follistatin-like-3 is dependent on differential cell surface binding and specificity for activin, myostatin, and bone morphogenetic proteins. Endocrinology 147(7):3586–3597
- 108. Welt C, Sidis Y, Keutmann H, Schneyer A (2002) Activins, inhibins, and follistatins: from endocrinology to signaling. A paradigm for the new millennium. Experimental Biology Med 227(9):724–752
- Hansen JS, Plomgaard P (2016) Circulating follistatin in relation to energy metabolism. Mol Cell Endocrinol 433:87–93
- 110. Tao R, Stöhr O, Wang C, Qiu W, Copps KD, White MF (2023) Hepatic follistatin increases basal metabolic rate and attenuates diet-induced obesity during hepatic insulin resistance. Mol Metabolism 71:101703
- 111. Lee S-J, McPherron AC (2001) Regulation of myostatin activity and muscle growth. Proc Natl Acad Sci 98(16):9306–9311
- 112. Blumensatt M, Greulich S, Herzfeld de Wiza D, Mueller H, Maxhera B, Rabelink MJ et al (2013) Activin a impairs insulin action in cardiomyocytes via up-regulation of miR-143. Cardiovasc Res 100(2):201–210
- 113. Yang J (2014) Enhanced skeletal muscle for effective glucose homeostasis. Prog Mol Biol Transl Sci 121:133–163
- 114. Yamagoe S, Yamakawa Y, Matsuo Y, Minowada J, Mizuno S, Suzuki K (1996) Purification and primary amino acid sequence of a novel neutrophil chemotactic factor LECT2. Immunol Lett 52(1):9–13
- 115. Hiraki Y, Inoue H, Kondo J, Kamizono A, Yoshitake Y, Shukunami C et al (1996) A novel growth-promoting factor derived from fetal bovine cartilage, chondromodulin II: purification and amino acid sequence. J Biol Chem 271(37):22657–22662
- 116. Slowik V, Apte U (2017) Leukocyte cell-derived Chemotaxin-2: it's role in pathophysiology and future in Clinical Medicine. Clin Transl Sci 10(4):249–259
- 117. Nagal H, Hamada T, Uchida T, Yamagoe S, Suzuki K (1998) Systemic expression of a newly recognized protein, LECT2, in the human body. Pathol Int 48(11):882–886
- 118. Yamagoe S, Mizuno S, Suzuki K (1998) Molecular cloning of human and bovine LECT2 having a neutrophil chemotactic activity and its specific expression in the liver. Biochim Biophys Acta 1396(1):105–113
- 119. Misu H (2019) Identification of hepatokines involved in pathology of type 2 diabetes and obesity. Endocr J 66(8):659–662
- 120. Lan F, Misu H, Chikamoto K, Takayama H, Kikuchi A, Mohri K et al (2014) LECT2 functions as a hepatokine that links obesity to skeletal muscle insulin resistance. Diabetes 63(5):1649–1664
- 121. Zhu M-H, Liu Y-J, Yang G-J, Chen J (2023) The emerging roles of leukocyte cell-derived chemotaxin-2 in immune diseases: from mechanisms to therapeutic potential. Front Immunol 14:1158083
- 122. Hara H, Uchida S, Yoshimura H, Aoki M, Toyoda Y, Sakai Y et al (2000) Isolation and characterization of a novel liver-specific

gene, hepassocin, upregulated during liver regeneration. Biochim Biophys Acta 1492(1):31-44

- 123. Hara H, Yoshimura H, Uchida S, Toyoda Y, Aoki M, Sakai Y et al (2001) Molecular cloning and functional expression analysis of a cDNA for human hepassocin, a liver-specific protein with hepatocyte mitogenic activity. Biochim Biophys Acta 1520(1):45–53
- 124. Cao MM, Xu WX, Li CY, Cao CZ, Wang ZD, Yao JW et al (2011) Hepassocin regulates cell proliferation of the human hepatic cells L02 and hepatocarcinoma cells through different mechanisms. J Cell Biochem 112(10):2882–2890
- 125. Demchev V, Malana G, Vangala D, Stoll J, Desai A, Kang HW et al (2013) Targeted deletion of fibrinogen like protein 1 reveals a novel role in energy substrate utilization. PLoS ONE 8(3):e58084
- 126. Huang RL, Li CH, Du YF, Cheng KP, Lin CH, Hu CY et al (2020) Discovery of a role of the novel hepatokine, hepassocin, in obesity. BioFactors 46(1):100–105
- 127. Wu HT, Chen SC, Fan KC, Kuo CH, Lin SY, Wang SH et al (2020) Targeting fibrinogen-like protein 1 is a novel therapeutic strategy to combat obesity. Faseb j 34(2):2958–2967
- 128. Wu HT, Lu FH, Ou HY, Su YC, Hung HC, Wu JS et al (2013) The role of hepassocin in the development of non-alcoholic fatty liver disease. J Hepatol 59(5):1065–1072
- 129. Wu HT, Ou HY, Hung HC, Su YC, Lu FH, Wu JS et al (2016) A novel hepatokine, HFREP1, plays a crucial role in the development of insulin resistance and type 2 diabetes. Diabetologia 59(8):1732–1742
- 130. Jung TW, Chung YH, Kim HC, Abd El-Aty AM, Jeong JH (2018) Hyperlipidemia-induced hepassocin in the liver contributes to insulin resistance in skeletal muscle. Mol Cell Endocrinol 470:26–33
- 131. Yang Y, Liu X, Chen H, Wang P, Yao S, Zhou B et al (2022) HPS protects the liver against steatosis, cell death, inflammation, and fibrosis in mice with steatohepatitis. Febs j 289(17):5279–5304
- 132. Vannahme C, Smyth N, Miosge N, Gösling S, Frie C, Paulsson M et al (2002) Characterization of SMOC-1, a novel modular calcium-binding protein in basement membranes. J Biol Chem 277(41):37977–37986
- Bornstein P, Sage EH (2002) Matricellular proteins: extracellular modulators of cell function. Curr Opin Cell Biol 14(5):608–616
- 134. Novinec M, Kovacic L, Skrlj N, Turk V, Lenarcic B (2008) Recombinant human SMOCs produced by in vitro refolding: calcium-binding properties and interactions with serum proteins. Protein Expr Purif 62(1):75–82
- 135. Di Niro R, Sulic AM, Mignone F, D'Angelo S, Bordoni R, Iacono M et al (2010) Rapid interactome profiling by massive sequencing. Nucleic Acids Res 38(9):e110
- 136. Brellier F, Ruggiero S, Zwolanek D, Martina E, Hess D, Brown-Luedi M et al (2011) SMOC1 is a tenascin-C interacting protein over-expressed in brain tumors. Matrix Biol 30(3):225–233
- 137. Klemenčič M, Novinec M, Maier S, Hartmann U, Lenarčič B (2013) The heparin-binding activity of secreted modular calciumbinding protein 1 (SMOC-1) modulates its cell adhesion properties. PLoS ONE 8(2):e56839
- 138. Montgomery MK, Bayliss J, Devereux C, Bezawork-Geleta A, Roberts D, Huang C et al (2020) SMOC1 is a glucose-responsive hepatokine and therapeutic target for glycemic control. Sci Transl Med. ;12(559)
- 139. Gao Q, Mok H-P, Zhuang J (2019) Secreted modular calciumbinding proteins in pathophysiological processes and embryonic development. Chin Med J 132(20):2476–2484
- 140. Decourtye L, Mire E, Clemessy M, Heurtier V, Ledent T, Robinson IC et al (2017) IGF-1 induces GHRH Neuronal Axon Elongation during early postnatal life in mice. PLoS ONE 12(1):e0170083
- 141. Yakar S, Rosen CJ, Beamer WG, Ackert-Bicknell CL, Wu Y, Liu JL et al (2002) Circulating levels of IGF-1 directly regulate bone growth and density. J Clin Invest 110(6):771–781

- 142. Kofoed EM, Hwa V, Little B, Woods KA, Buckway CK, Tsubaki J et al (2003) Growth hormone insensitivity associated with a STAT5b mutation. N Engl J Med 349(12):1139–1147
- 143. Rajkumar K, Krsek M, Dheen ST, Murphy LJ (1996) Impaired glucose homeostasis in insulin-like growth factor binding protein-1 transgenic mice. J Clin Investig 98(8):1818–1825
- 144. Sandhu MS, Heald AH, Gibson JM, Cruickshank JK, Dunger DB, Wareham NJ (2002) Circulating concentrations of insulinlike growth factor-I and development of glucose intolerance: a prospective observational study. Lancet 359(9319):1740–1745
- 145. Succurro E, Andreozzi F, Marini MA, Lauro R, Hribal ML, Perticone F et al (2009) Low plasma insulin-like growth factor-1 levels are associated with reduced insulin sensitivity and increased insulin secretion in nondiabetic subjects. Nutr Metab Cardiovasc Dis 19(10):713–719
- 146. Boger RH, Frystyk J, Ledet T, Moller N, Flyvbjerg A, Orskov H (2003) Low serum insulin-like growth factor I is associated with increased risk of ischemic heart disease. Circulation 107(20):e193 author reply e
- 147. Rajkumar K, Krsek M, Dheen ST, Murphy LJ (1996) Impaired glucose homeostasis in insulin-like growth factor binding protein-1 transgenic mice. J Clin Invest 98(8):1818–1825
- 148. Bach MA, Shen-Orr Z, Lowe WL Jr., Roberts CT Jr., LeRoith D (1991) Insulin-like growth factor I mRNA levels are developmentally regulated in specific regions of the rat brain. Brain Res Mol Brain Res 10(1):43–48
- 149. Lee JY, Muenzberg H, Gavrilova O, Reed JA, Berryman D, Villanueva EC et al (2008) Loss of cytokine-STAT5 signaling in the CNS and pituitary gland alters energy balance and leads to obesity. PLoS ONE 3(2):e1639
- 150. Hong H, Cui ZZ, Zhu L, Fu SP, Rossi M, Cui YH et al (2017) Central IGF1 improves glucose tolerance and insulin sensitivity in mice. Nutr Diabetes 7(12):2
- 151. Nguyen NL, Barr CL, Ryu V, Cao Q, Xue B, Bartness TJ (2017) Separate and shared sympathetic outflow to white and brown fat coordinately regulates thermoregulation and beige adipocyte recruitment. Am J Physiol Regul Integr Comp Physiol 312(1):R132–r45
- 152. Bonet ML, Mercader J, Palou A (2017) A nutritional perspective on UCP1-dependent thermogenesis. Biochimie 134:99–117
- 153. Laron Z (1999) The essential role of IGF-I: lessons from the longterm study and treatment of children and adults with Laron syndrome. J Clin Endocrinol Metab 84(12):4397–4404
- 154. Laron Z, Avitzur Y, Klinger B (1995) Carbohydrate metabolism in primary growth hormone resistance (Laron syndrome) before and during insulin-like growth factor-I treatment. Metabolism 44(10 Suppl 4):113–118
- 155. Wang X, Sun H, Ma B, Gao J, Yin J, Qu S (2020) Insulin-like growth factor 1 related to chronic low-grade inflammation in patients with obesity and early change of its levels after laparoscopic sleeve gastrectomy. Obes Surg 30(9):3326–3332
- 156. Wischhusen J, Melero I, Fridman WH (2020) Growth/Differentiation Factor-15 (GDF-15): from biomarker to Novel Targetable Immune Checkpoint. Front Immunol 11:951
- 157. Johnen H, Lin S, Kuffner T, Brown DA, Tsai VW, Bauskin AR et al (2007) Tumor-induced anorexia and weight loss are mediated by the TGF-beta superfamily cytokine MIC-1. Nat Med 13(11):1333–1340
- 158. Breit SN, Brown DA, Tsai VW (2021) The GDF15-GFRAL pathway in Health and Metabolic Disease: friend or foe? Annu Rev Physiol 83:127–151
- 159. Kelly JA, Lucia MS, Lambert JR (2009) p53 controls prostatederived factor/macrophage inhibitory cytokine/NSAID-activated gene expression in response to cell density, DNA damage and hypoxia through diverse mechanisms. Cancer Lett 277(1):38–47

- 160. Asrih M, Wei S, Nguyen TT, Yi HS, Ryu D, Gariani K (2023) Overview of growth differentiation factor 15 in metabolic syndrome. J Cell Mol Med 27(9):1157–1167
- 161. Chrysovergis K, Wang X, Kosak J, Lee SH, Kim JS, Foley JF et al (2014) NAG-1/GDF-15 prevents obesity by increasing thermogenesis, lipolysis and oxidative metabolism. Int J Obes (Lond) 38(12):1555–1564
- 162. Zhou Y, Rui L (2013) Lipocalin 13 regulation of glucose and lipid metabolism in obesity. Vitamins Horm 91:369–383
- 163. Zhou Y, Rui L (2013) Lipocalin 13 regulation of glucose and lipid metabolism in obesity. Vitam Horm 91:369–383
- 164. Cho KW, Zhou Y, Sheng L, Rui L (2011) Lipocalin-13 regulates glucose metabolism by both insulin-dependent and insulin-independent mechanisms. Mol Cell Biol 31(3):450–457
- 165. Sheng L, Cho KW, Zhou Y, Shen H, Rui L (2011) Lipocalin 13 protein protects against hepatic steatosis by both inhibiting lipogenesis and stimulating fatty acid β-oxidation. J Biol Chem 286(44):38128–38135
- 166. Ekim Üstünel B, Friedrich K, Maida A, Wang X, Krones-Herzig A, Seibert O et al (2016) Control of diabetic hyperglycaemia and insulin resistance through TSC22D4. Nat Commun 7(1):13267
- 167. Ahmad SAI, Anam MB, Ito N, Ohta K (2018) Involvement of Tsukushi in diverse developmental processes. J Cell Communication Signal 12(1):205–210
- Istiaq A, Ohta K (2022) A review on Tsukushi: mammalian development, disorders, and therapy. J Cell Communication Signal 16(4):505–513
- 169. Mouchiroud M, Camiré É, Aldow M, Caron A, Jubinville É, Turcotte L et al (2019) The hepatokine Tsukushi is released in response to NAFLD and impacts cholesterol homeostasis. JCI Insight. ;4(15)
- 170. Wang Q, Sharma V, Shen H, Xiao Y, Zhu Q, Xiong X et al (2019) The hepatokine Tsukushi gates energy expenditure via brown fat sympathetic innervation. Nat Metab 1:251–260
- 171. Wang Q, Sharma VP, Shen H, Xiao Y, Zhu Q, Xiong X et al (2019) The hepatokine Tsukushi gates energy expenditure via brown fat sympathetic innervation. Nat Metab 1(2):251–260
- 172. Mouchiroud M, Camiré É, Aldow M, Caron A, Jubinville É, Turcotte L et al (2019) The Hepatokine TSK does not affect brown fat thermogenic capacity, body weight gain, and glucose homeostasis. Mol Metab 30:184–191
- 173. Aydin S (2014) Three new players in energy regulation: preptin, adropin and irisin. Peptides 56:94–110
- 174. Ali II, D'Souza C, Singh J, Adeghate E (2022) Adropin's Role in Energy Homeostasis and Metabolic Disorders. Int J Mol Sci. ;23(15)
- 175. Kumar KG, Trevaskis JL, Lam DD, Sutton GM, Koza RA, Chouljenko VN et al (2008) Identification of adropin as a secreted factor linking dietary macronutrient intake with energy homeostasis and lipid metabolism. Cell Metab 8(6):468–481
- 176. Jasaszwili M, Billert M, Strowski MZ, Nowak KW, Skrzypski M (2020) Adropin as a Fat-Burning hormone with multiple functions-review of a Decade of Research. Molecules. ;25(3)
- 177. Ganesh Kumar K, Zhang J, Gao S, Rossi J, McGuinness OP, Halem HH et al (2012) Adropin deficiency is associated with increased adiposity and insulin resistance. Obes (Silver Spring) 20(7):1394–1402
- 178. Banerjee S, Ghoshal S, Stevens JR, McCommis KS, Gao S, Castro-Sepulveda M et al (2020) Hepatocyte expression of the micropeptide adropin regulates the liver fasting response and is enhanced by caloric restriction. J Biol Chem 295(40):13753–13768
- 179. Gao S, McMillan RP, Zhu Q, Lopaschuk GD, Hulver MW, Butler AA (2015) Therapeutic effects of adropin on glucose tolerance and substrate utilization in diet-induced obese mice with insulin resistance. Mol Metab 4(4):310–324

- 180. Jasaszwili M, Billert M, Strowski MZ, Nowak KW, Skrzypski M (2020) Adropin as a fat-burning hormone with multiple functions—review of a decade of research. Molecules 25(3):549
- 181. Takanori Nakamura MA, Yuzuru EI, Takio K, Uchiyama H, Moriya N, Ariizumi T (1992) Takayuki Yashiro, Kishiko Sugino, Koiti Titani, and Hiromu Sugino. Isolation and Characterization of Native Activin B
- 182. Hashimoto O, Ushiro Y, Sekiyama K, Yamaguchi O, Yoshioka K, Mutoh K et al (2006) Impaired growth of pancreatic exocrine cells in transgenic mice expressing human activin betaE subunit. Biochem Biophys Res Commun 341(2):416–424
- 183. Abe Y, Minegishi T, Leung PC (2004) Activin receptor signaling. Growth Factors 22(2):105–110
- 184. Liu H, Hallauer Hastings M, Kitchen R, Xiao C, Baldovino Guerra JR, Kuznetsov A, Arteriosclerosis et al (2023) Thromb Vascular Biology 43(2):330–349
- 185. Hashimoto O, Tsuchida K, Ushiro Y, Hosoi Y, Hoshi N, Sugino H et al (2002) cDNA cloning and expression of human activin betaE subunit. Mol Cell Endocrinol 194(1–2):117–122
- 186. Hashimoto O, Funaba M, Sekiyama K, Doi S, Shindo D, Satoh R et al (2018) Activin E controls energy homeostasis in both brown and white adipose tissues as a hepatokine. Cell Rep 25(5):1193–1203
- 187. Sekiyama K, Ushiro Y, Kurisaki A, Funaba M, Hashimoto O (2019) Activin E enhances insulin sensitivity and thermogenesis by activating brown/beige adipocytes. J Vet Med Sci 81(5):646–652
- 188. Smati S, Régnier M, Fougeray T, Polizzi A, Fougerat A, Lasserre F et al (2020) Regulation of hepatokine gene expression in response to fasting and feeding: influence of PPAR-α and insulin-dependent signalling in hepatocytes. Diabetes Metab 46(2):129–136
- 189. Ko H, Il Kim Y, Ahn HJ (2022) Activin suppresses the inflammatory response of TNF-α-stimulated human umbilical vein endothelial cells. Die Pharmazie-An Int J Pharm Sci 77(5):152–156
- 190. Schmidt RL, Simonović M (2012) Synthesis and decoding of selenocysteine and human health. Croat Med J 53(6):535–550
- 191. Yu S-s, Du J-1 (2017) Selenoprotein S: a therapeutic target for diabetes and macroangiopathy? Cardiovasc Diabetol 16(1):101
- 192. Arteel GE, Klotz L-O, Buchczyk DP, Sies H (2002) [11] selenoprotein P. In: Sies H, Packer L (eds) Methods in Enzymology, vol 347. Academic, pp 121–125
- 193. Yu S-s, Du J-l (2024) Current views on selenoprotein S in the pathophysiological processes of diabetes-induced atherosclerosis: potential therapeutics and underlying biomarkers. Diabetol Metab Syndr 16(1):5
- 194. Polyzos SA, Kountouras J, Goulas A, Duntas L (2020) Selenium and selenoprotein P in nonalcoholic fatty liver disease. Horm (Athens) 19(1):61–72
- 195. Takayama H, Misu H, Iwama H, Chikamoto K, Saito Y, Murao K et al (2014) Metformin suppresses expression of the selenoprotein P gene via an AMP-activated kinase (AMPK)/FoxO3a pathway in H4IIEC3 hepatocytes. J Biol Chem 289(1):335–345
- 196. Ye R, Huang J, Wang Z, Chen Y, Dong Y (2022) The role and mechanism of essential selenoproteins for Homeostasis. Antioxid (Basel). ;11(5)
- 197. Misu H, Takamura T, Takayama H, Hayashi H, Matsuzawa-Nagata N, Kurita S et al (2010) A liver-derived secretory protein, selenoprotein P, causes insulin resistance. Cell Metab 12(5):483–495
- 198. Yang SJ, Hwang SY, Choi HY, Yoo HJ, Seo JA, Kim SG et al (2011) Serum selenoprotein P levels in patients with type 2 diabetes and prediabetes: implications for insulin resistance, inflammation, and atherosclerosis. J Clin Endocrinol Metab 96(8):E1325–E1329

- 199. Hariharan S, Dharmaraj S (2020) Selenium and selenoproteins: it's role in regulation of inflammation. Inflammopharmacology 28:667–695
- 200. Polyzos SA, Kountouras J, Goulas A, Duntas L (2020) Selenium and selenoprotein P in nonalcoholic fatty liver disease. Hormones 19:61–72
- 201. Alfawaz HA, Alfaifi AA, Yakout SM, Khattak MNK, Alnaami AM, Al-Thayidi A et al (2022) Circulating hepcidin and its associations with low-grade inflammation and iron indices among arab adults with and without T2DM. Am J Translational Res 14(10):7520
- 202. Chambers K, Ashraf MA, Sharma S (2019) Physiology, hepcidin
- 203. Miao R, Fang X, Zhang Y, Wei J, Zhang Y, Tian J (2023) Iron metabolism and ferroptosis in type 2 diabetes mellitus and complications: mechanisms and therapeutic opportunities. Cell Death Dis 14(3):186
- 204. Maherdika M, Samsuria IK, Hendrianingtyas M, Widyastiti NS, Rahayu M (2022) The difference levels of hepcidin and interleukin-6 between obese and non-obese type 2 diabetes mellitus. Indonesian Biomedical J 14(1):98–103
- 205. Zhang X, Zhang L, Tan Y-m, Liu Y-p, Li J-j, Deng Q-m et al (2021) Hepcidin gene silencing ameliorated inflammation and insulin resistance in adipose tissue of db/db mice via inhibiting METs formation. Mol Immunol 133:110–121
- 206. Zhang Z, Liu H, Liu J (2019) Akt activation: a potential strategy to ameliorate insulin resistance. Diabetes Res Clin Pract 156:107092
- 207. Zhang X-g, Liu A-x, Zhang Y-x, Zhou M-y, Li X-y, Fu M-h et al (2022) A diarylheptanoid compound from alpinia officinarum hance ameliorates high glucose-induced insulin resistance by regulating pi3k/akt-nrf2-gsk3β signaling pathways in hepg2 cells. J Ethnopharmacol 295:115397
- 208. Mo M, Pan L, Deng L, Liang M, Xia N, Liang Y (2024) Iron Overload induces hepatic ferroptosis and insulin resistance by inhibiting the Jak2/stat3/slc7a11 signaling pathway. Cell Biochem Biophys. :1–16
- 209. Lu L, Ye X, Yao Q, Lu A, Zhao Z, Ding Y et al (2018) Egr2 enhances insulin resistance via JAK2/STAT3/SOCS-1 pathway in HepG2 cells treated with palmitate. Gen Comp Endocrinol 260:25–31

- 210. Page M, Kell D, Pretorius E (2022) The role of lipopolysaccharide-induced cell signalling in chronic inflammation. Chronic Stress (Thousand Oaks) 6:24705470221076390
- 211. Meng L, Song Z, Liu A, Dahmen U, Yang X, Fang H (2021) Effects of lipopolysaccharide-binding protein (LBP) single nucleotide polymorphism (SNP) in infections, inflammatory diseases, metabolic disorders and cancers. Front Immunol 12:681810
- 212. Page MJ, Kell DB, Pretorius E (2022) The role of lipopolysaccharide-induced cell signalling in chronic inflammation. Chronic Stress 6:24705470221076390
- 213. Pussinen PJ, Kopra E, Pietiäinen M, Lehto M, Zaric S, Paju S et al (2022) Periodontitis and cardiometabolic disorders: the role of lipopolysaccharide and endotoxemia. Periodontol 2000 89(1):19–40
- 214. Liu J, Kang R, Tang D (2024) Lipopolysaccharide delivery systems in innate immunity. Trends Immunol
- Mazgaeen L, Gurung P (2020) Recent advances in lipopolysaccharide recognition systems. Int J Mol Sci 21(2):379
- 216. Frances L, Tavernier G, Viguerie N (2021) Adipose-derived lipidbinding proteins: the good, the bad and the metabolic diseases. Int J Mol Sci 22(19):10460
- 217. Won Y, Yang JI, Park S, Chun JS (2021) Lipopolysaccharide binding protein and CD14, cofactors of toll-like receptors, are essential for low-Grade inflammation–Induced exacerbation of cartilage damage in mouse models of Posttraumatic Osteoarthritis. Arthritis Rheumatol 73(8):1451–1460
- 218. Wu H, Ballantyne CM (2020) Metabolic inflammation and insulin resistance in obesity. Circul Res 126(11):1549–1564

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