REVIEW

**Matthias C. Rillig** 7 **Michael F. Allen**

# What is the role of arbuscular mycorrhizal fungi in plant-to-ecosystem responses to Elevated atmospheric  $CO<sub>2</sub>$ ?

Accepted: 12 February 1999

**Abstract** We advocate the concept of an arbuscular mycorrhiza (AM) as a temporally and spatially complex symbiosis representing a suite of hosts and fungi, as against the more traditional "dual organism" view. We use the hierarchical framework presented in Fig. 1 as a basis for organizing many unanswered questions, and several questions that have not been asked, concerning the role of AM in responses to elevated atmospheric  $CO<sub>2</sub>$ . We include the following levels: plant host, plant population, plant community, functional group and ecosystem. Measurements of the contributions of AM fungi at the various levels require the use of different response variables. For example, hyphal nutrient translocation rates or percent AM root infection may be important measures at the individual plant level, but hyphal biomass or glomalin production and turnover are more relevant at the ecosystem level. There is a discrepancy between our knowledge of the multifaceted role of AM fungi in plant and ecosystem ecology and most of the current research aimed at elucidating the importance of this symbiosis in global-change scenarios. Our framework for more integrated and multifactorial research on mycorrhizal involvement in regulating  $CO<sub>2</sub>$  responses may also serve to enhance communication between researchers working at different scales on large global-change ecosystem projects.

**Key words** Arbuscular mycorrhiza  $\cdot$  Elevated CO<sub>2</sub>  $\cdot$ Microorganisms · Rhizosphere · Soil Fungi · Global change

M.C. Rillig  $(\boxtimes)$ Carnegie Institution of Washington, Department of Plant Biology, 260 Panama Street, Stanford, CA 94305, USA e-mail: matthias@iasper.stanford.edu, Fax:  $+1-650-325-6857$ 

M.F. Allen

# Introduction

The increasing atmospheric concentration of carbon dioxide is one of the most significant factors of global change, both with respect to the empirical evidence supporting its occurrence (e.g., Keeling et al. 1995) and the number of studies aimed at describing its effect on ecosystems. As a general trend, a large proportion of the additionally fixed carbon in terrestrial ecosystems is channeled below ground, to roots (Rogers et al. 1994) and soil (e.g., Jones et al. 1998). Growth in the number of reviews on the topic of soil microbial limitations and responses to elevated  $CO<sub>2</sub>$  (e.g., Zak et al. 1993; O'Neill 1994; Díaz 1996; Hodge 1996; Sanders 1996; Sadowsky and Schortemeyer 1997; Paterson et al. 1997) is therefore not surprising.

Arbuscular mycorrhizae (AM), ubiquitous symbioses between fungi in the Glomales and roots of higher plants, constitute a crucial link at the root-soil interface. The mineral nutritional aspects of AM symbioses are widely appreciated (Smith and Read 1997). Consequently, the vast majority of studies on the role of AM fungi in global-change scenarios involving elevated atmospheric  $CO<sub>2</sub>$  concentrations have taken on this aspect of the symbiosis as their main focus (e.g., Hodge 1996).

We challenge this mineral-nutrition centered approach in this paper, and advocate a more multifaceted view of the role of AM fungi in mediating ecosystem responses to  $CO<sub>2</sub>$ . O'Neill et al. (1991) and Allen (1991, 1996) have argued that a hierarchical view (MacMahon et al. 1978; O'Neill et al. 1986) is warranted in research that tries to ascertain the importance of small-scale processes to large-scale perturbations. In this paper, we use a hierarchically structured conceptual framework to organize our discussion of research on mycorrhizae and elevated  $CO<sub>2</sub>$ .

Center for Conservation Biology, University of California, Riverside, CA 92521-0334, USA

## Arbuscular mycorrhiza: a different view

A mycorrhiza is a mutualistic symbiosis between plants and fungi localized in the root or rhizoid (e.g., Allen 1991). To most ecologists, this implies a symbiosis between a microorganism (a fungus) and a macroorganism (a plant). This view has led to the large bias that the relationship can be studied virtually exclusively at the scale of an individual plant with a response to a single fungus. However, we know that a single plant will form mycorrhizae with many fungi as the roots encounter different patches. Just as important, a single mycelium can extend across multiple plants. We do not know the limit of that spread, but labeling studies demonstrate that carbon and nutrients can be transported by AM hyphae to many surrounding plants (e.g., Chiarello et al. 1982). It is also important to note that an individual plant may die and become replaced by a new seedling (of the same or different species). The fungus will likely remain (unless, for example, there is severe disturbance) and form a mycorrhiza with the replacement. Similarly, a patch of an individual mycelium of a fungus may die and be replaced while the plant remains, again re-forming mycorrhizae. Thus, an AM is not always best represented as a "dual organism" that can be studied as an entity, but as a suite of plants and fungi whose organization is spatiotemporally complex, transient, and extensive. Therefore, we argue that research on mycorrhizal aspects of global-change biology is most appropriately hierarchically structured.

# The framework

The entities to be included are host plant, plant population, plant community, functional group, and ecosystem (Fig. 1). We use the host plant as the basic unit, because an individual organism is the basic unit of selection (MacMahon et al. 1978). From this basic unit, two hierarchical lines are pursued, population to community, and functional group to ecosystem ('population-community' versus 'process-functional' according to O'Neill et al. 1986; or 'coevolutionary' versus 'matter-energy exchange' according to MacMahon et al. 1978). Thus far, the development of conceptual models (Andersen and Rygiewicz 1991; Berntson and Bazzaz 1996) and mycorrhizal research on elevated  $CO<sub>2</sub>$  (O'Neill 1994; Sanders 1996; Klironomos et al. 1996; Díaz 1996) have concentrated on the individual plant. The ways in which mycorrhizal fungi can potentially influence responses to  $CO<sub>2</sub>$  at the various levels include: (a) influencing the homeostatic adjustment of individual host plants to elevated  $CO<sub>2</sub>$ , (b) altering the variability of responses to  $CO<sub>2</sub>$  within a plant population, (c) differentially responding and providing feedbacks to different plant species within a plant community and to different plant functional assemblages in an ecosystem,

Ecosystem C storage, nutrient cycling



**Fig. 1** Framework for the discussion of arbuscular mycorrhizal (AM) fungal contributions to responses to elevated atmospheric  $CO<sub>2</sub>$  at the levels of individual host plant, plant population, plant community and the ecosystem. 'Ecosystem' as defined here belongs to the process-functional branch, and 'community' to the 'population-community' branch of the hierarchy (see text). The *bold arrows* signify the different ways in which AM fungi can influence  $CO<sub>2</sub>$  responses at the respective levels. It will be necessary to eventually introduce the level of the functional group response group, once sufficient data have been collected to identify suitable groupings

(d) providing an increased ecosystem sink of carbon in the soil, and influencing nutrient cycling patterns. For example, because 'community' and 'ecosystem' belong to different hierarchies, it follows that changes in plant community composition mediated by AM fungi do not necessarily have to lead to ecosystem ('process') changes, and vice versa. It is necessary to consider which response variables are most meaningful at each level of organization (MacMahon et al. 1978). For example, percent root infection by AM fungi may yield desirable information for studying P uptake in an individual plant (e.g., Smith and Read 1997). However, percent infection may be a less important measure of ecosystem soil carbon storage, for which AM extraradical hyphal length, biomass or glomalin turnover may be more crucial.

## Plant homeostatic adjustment: the individual

Among the most important factors determining the extent of down-regulation of photosynthesis are availability of nutrients and sinks. Further, factors that affect growth and not necessarily photosynthesis are going to be important. We discuss mineral nutrition, water relations, physiological carbon sink, and interactions with saprophytic and parasitic fungi.

# Response variables (and mineral nutrition)

AM fungi can clearly play an important role in the mineral nutrition of their host (George et al. 1992; Smith and Read 1997), and nutrients can limit photosynthesis when not supplied at adequate rates concomitant with elevated  $CO<sub>2</sub>$ . The response variable most commonly measured to reflect AM status of plants grown in  $CO<sub>2</sub>$ (and in general) has been percent colonized root or colonized root length. While there may be a direct correlation for a few systems between colonized root length and nutrient flow to roots via the mycobionts, this is not always (or even often) true (Smith and Read 1997). This assumption becomes even more questionable when imposed treatments (like  $CO<sub>2</sub>$  or water) may cause changes in the composition of AM fungi colonizing roots (Klironomos et al. 1998), leading to different average nutrient translocation rates per colonized root length. In the shrub *Gutierrezia sarothrae*, the presence of arbuscules (structures largely responsible for carbon and nutrient exchange) increased more than 14-fold in elevated  $CO<sub>2</sub>$ , with no significant changes in percent AM infection or AM-colonized root length (Rillig and Allen 1998). In the same study, extraradical hyphal length per AM infected root length also increased in elevated  $CO<sub>2</sub>$ , again with no change in percent AM root infection. We also found an increase with elevated  $CO<sub>2</sub>$  in the number of hyphae within an individual root ('intensity' of infection) in two different species (Rillig and Allen 1998; Rillig et al. 1998), without changes in percent AM fungal colonization or changes in colonized root length.

These results clearly illustrate that AM percent infection measurements are often insufficient in capturing mycorrhizal responses to elevated  $CO<sub>2</sub>$  when used as the only indicator. Measurements of AM fungal hyphal nutrient translocation (George et al. 1992) in elevated  $CO<sub>2</sub>$  are needed to test if nutrient translocation rates change on a hyphal length basis (either as a function of changed fungal isolate composition or altered single isolate physiology in elevated  $CO<sub>2</sub>$ ).

Most studies on the individual plant level have been conducted as single-time harvest experiments. This design has the disadvantage that, potentially, plants of different phenological stages and sizes are compared, if the phytobiont responds to the treatment with increased growth. Rouhier and Read (1998), using three

sequential harvests, showed that some mycorrhizal parameters (colonized root length, arbuscular and vesicular colonization) may only respond to the  $CO<sub>2</sub>$  enrichment later in the plant's life cycle. Hence, a single-time harvest may miss potential significant responses. Staddon et al. (1998, 1999ab), in a series of experiments, showed that significant responses of percent root infection and extraradical hyphal length (Staddon et al. 1999a) to the  $CO<sub>2</sub>$  treatment can disappear when plant growth parameters are included in the analysis as covariates. Hence, Staddon and co-workers concluded that there is no direct effect of the  $CO<sub>2</sub>$  treatment on the mycorrhiza, but that the fungi respond simply to the fact that plants grow larger. Although this important effect has been demonstrated for a variety of hosts (Staddon et al. 1999b), it was only shown for a single fungal isolate in growth chamber experiments with a highly artificial growth medium. The wider relevance of the results is questionable, particularly given the known biodiversity and potential functional specialization of AM fungi (Giovannetti and Gianinazzi-Pearson 1994; Morton and Bentivenga 1994), and isolate-specific response to  $CO<sub>2</sub>$  (Klironomos et al. 1998) of the mycobionts. Following the model experiment of Staddon et al., studies containing different or several fungal species should be carried out to test the validity of this conclusion. In a growth chamber experiment with soil containing several mycobionts and involving 11 sequential harvests, we could not confirm the findings of Staddon and coworkers (Rillig, Luo, and Field; unpublished data).

# Water relations

While AM fungi can improve plant water relations (e.g. Allen and Allen 1986; George et al. 1992; but see e.g., Bryla and Duniway 1997), in water-limited ecosystems elevated atmospheric  $CO<sub>2</sub>$  often improves plant wateruse efficiency (e.g., Bazzaz 1990). Mycorrhizae appear primarily to increase water throughput, thereby presumably maximizing plant carbon uptake for subsequent translocation to the fungus itself (Allen et al. 1981). If AM fungi rarely or minimally improve wateruse efficiency, important interactions, warranting further study, could result when plants are grown in elevated  $CO<sub>2</sub>$ , particularly in arid ecosystems. Further studies on plant water relations in elevated  $CO<sub>2</sub>$  atmospheres should, therefore, not ignore mycorrhizal aspects.

# Physiological carbon sink

Plants may respond to elevated  $CO<sub>2</sub>$  initially with increased photosynthesis, but if there are sink limitations complete homeostatic adjustment of an individual plant may occur (e.g. Bazzaz 1990). AM fungi can constitute a physiologically important carbon sink (Wright et al. 1998). Roots with AM fungi receive about 4–20% more photosynthate than comparable non-mycorrhizal (NM) roots (Smith and Read 1997). Jakobsen and Rosendahl (1990) estimated that AM fungi could use up to 20% of the total fixed  ${}^{14}CO_2$  in young plants. The importance of this sink to plants grown in elevated  $CO<sub>2</sub>$  has been recognized (Jongen et al. 1996). However, upon measuring total non-structural carbon pools of mycorrhizal and NM plants in ambient and elevated  $CO<sub>2</sub>$ , no change in the C-sink was found (Jongen et al. 1996). Pools are just one representation of sinks, as turnover and respiration rates may also change. For young *Vicia faba* plants, Paul and Kucey (1981) estimated that AM fungi respired about 3% of the host photosynthate, while incorporating only ca. 1%. It is conceivable that respiration rates of AM hyphae change when plants are exposed to elevated  $CO<sub>2</sub>$ , because carbon supply may be altered. To date, hyphal respiration rates have never been measured directly for AM fungi, and consequently no data exist for rates under elevated  $CO<sub>2</sub>$  conditions.

Parasitic and facultatively parasitic/ saprophytic fungi

Specific root length often increases in elevated  $CO<sub>2</sub>$  for many plant species (Rogers et al. 1994), and hence surface area for potential fungal attack. Also, just as roots may become richer sources of carbon per root length (Rouhier et al. 1996; Paterson et al. 1997), roots may also become more attractive to NM fungi growing on or inside roots. Therefore, it appears possible that protection of roots against saprophytic or parasitic fungi could increase in importance in elevated  $CO<sub>2</sub>$ .

Mycorrhizal infection can reduce root infection by parasitic fungi probably by a range of mechanisms including direct AM fungus – pathogen interactions (St. Arnaud et al. 1995), or improvement of host nutritional status (Newsham et al. 1995). If the functioning of AM fungi with respect to these nutrient translocation or inter-fungus interaction processes increases in elevated  $CO<sub>2</sub>$ , because of increased carbon supply from the host, this protection mechanism may become increasingly important. Clearly, root-inhabiting parasitic fungi represent a carbon drain on the plant, can cause disease, promote homeostatic adjustment, and reduce fitness.

Facultatively parasitic/saprophytic fungi are typically common on plant root surfaces, but our knowledge of the biology of these fungi is still poorer than that of mycorrhizal or pathogenic fungi. Nevertheless, these fungi derive carbon from the root and represent a sink. Unlike AM fungi, this group of fungi typically does not have direct root tissue access, but depends on rhizodeposits. Therefore, AM fungi can potentially protect roots from this group simply by carbon preemption. It has been hypothesized that AM fungi can take up host extra carbon before it is rhizodeposited and available to all rhizosphere inhabitants (Diaz et al. 1993; Diaz 1996).

Runion et al. (1994) have shown a trend towards increased infection of rice by *Rhizoctonia* in elevated CO2. Klironomos et al. (1996) found increased levels of NM root infection in high-nutrient conditions in elevated  $CO<sub>2</sub>$ , but no change in low nutrient conditions, using *Artemisia*. We found decreased NM fungal infection in elevated  $CO<sub>2</sub>$  in several arbuscular mycorrhizal plant species from an annual grassland. This unexpected result was obtained when plants were grown in monoculture in pots for one growing season (Rillig et al. 1998), and when grown in the field for 6 years (Rillig, Field, and Allen, in press). Why did root infection by NM fungi decrease? Were there direct effects of roots on NM fungi (mediated by qualitative changes in rhizodeposition, or changes in plant susceptibility to NM infection)? Did competition for nutrients and/or carbon between mycorrhizal fungi and NM fungi intensify, with conditions favoring AM fungi? Did the rootinhabiting AM fungal community more aggessively protect roots against NM fungal colonization? These questions highlight the strong need for more experiments in this neglected and important area.

# Plant population: spatial heterogeneity of AM fungi

Although variability within populations in response to  $CO<sub>2</sub>$  will not have very large effects on an ecosystem's response to  $CO<sub>2</sub>$ , it may be important to a mechanistic understanding of longer-term trends. Interest in plant population level responses has increased with the realization that variability within a population in  $CO_2$ -responsiveness may be the key to potential evolutionary adaptations to elevated  $CO<sub>2</sub>$  (e.g. Curtis et al. 1996). Also, it is not experimentally obtained statistical means that respond to  $CO<sub>2</sub>$ , but the breadth represented by a population in a natural setting, as influenced for example by phenotypic plasticity (Curtis et al. 1996; Schmid et al. 1996). Can mycorrhizae contribute to this variability in  $CO<sub>2</sub>$  responses within a population, and do mycorrhizal plants have a greater breadth of  $CO<sub>2</sub>$  responses than NM plants? We hypothesize that AM fungi increase the range of plant responses to  $CO<sub>2</sub>$  in a population and that they play an important role in microevolutionary physiological adaptations to  $CO<sub>2</sub>$ .

AM fungal inoculum potential (e.g., Koide and Mooney 1987; Allen 1991) and AM fungal species (Bever et al. 1996) are non-uniformly distributed in the environment, even on a local scale of a few square meters (Bever et al. 1996). This means that plants from the same population can have quantitatively (percent infection) and qualitatively (subset of the AM fungal community) different AM fungal root colonization. AM fungal species are known to have different effects on the growth of a host plant species (e.g., Ravnskov and Jakobsen 1995). AM fungal isolates present in the same soil can also differentially influence plant clonal growth patterns (Streitwolf-Engel et al. 1997).

There is also a strong possibility that AM fungi change the amount of phenotypic plasticity in a plant population, e.g., in the case of leaf endophytic fungi (Cheplick 1997). Thus, plant populations with and without mycorrhizal fungi may differ in the amount of intrapopulation variability of response to  $CO<sub>2</sub>$ . This is a hypothesis that deserves further attention.

# Differential responses and feedbacks at the plant community level

It is known that AM fungi can influence the composition of plant communities (Allen and Allen 1990; Zobel et al. 1997; van der Heijden et al. 1998), but do AM fungi have an important role in determining  $CO<sub>2</sub>$  responses at the plant community level? The first question to be addressed is: will mycorrhizal fungi differentially respond to this ecosystem perturbation as a function of plant species? And, more importantly, will they subsequently also provide different feedbacks to plant physiology and growth as a function of plant species (by mechanisms discussed in the previous section)? Monz et al. (1994), Sanders (1996) and Rillig et al. (1998) showed that mycorrhizal percent infection of plant roots from the same communities showed differential responses among the plant species examined. These results suggest that AM fungi can respond to  $CO<sub>2</sub>$  as a function of plant species.

Possible approaches to examine the feedback of this response to plant communities could include extracting mycorrhizal spores and/ or hyphae from soils of elevated  $CO<sub>2</sub>$  plots, and comparing plant growth responses to these fungi and the fungal community extracted from 'ambient' treatment soils. Alternatively, plant communities could be grown in mesocosms with a completely factorial combination of mycorrhizal inoculum presence and  $CO<sub>2</sub>$  concentration. Because of the plant species dependence of symbiotic responses to  $CO<sub>2</sub>$ , it is important to realize that a scaling-up of responses from an observation made on an individual plant species to the plant community or ecosystem level is problematic.

# Functional groups and response groups

The previous section mentioned host plant species-specific fungus and symbiotic responses to elevated  $CO<sub>2</sub>$ . The next step needs to be the identification of suitable functional groups, because not all species from an ecosystem can be examined. Will mycorrhizal fungi be more beneficial to certain plant functional assemblages than to others in elevated  $CO<sub>2</sub>$ ? Díaz (1996) discussed how AM fungi may become important in tipping the balance between mycorrhizal and NM plant species growing in the same ecosystem when exposed to elevated  $CO<sub>2</sub>$ . It is also important to study how mycorrhizal fungi can influence nitrogen fixation in tripartite systems, and to what extent AM fungi can be responsible for frequently observed increases in N-fixation in elevated  $CO<sub>2</sub>$  (e.g., Zanetti et al. 1996; Zanetti and Hartwig 1997; Bandara et al. 1998). Other important pairs to consider are C3 versus C4 plants (see Monz et al. 1994), and early successional versus late successional plants. However, to date not nearly enough data exist to make generalizations.

With respect to the identification of response groups, i.e. groupings of organisms responding in similar ways to a disturbance (Lavorel et al. 1997), it is interesting to ask whether those plants that respond strongly to elevated  $CO<sub>2</sub>$  also typically derive a large benefit from being mycorrhizal. In other words, will plants with the highest responsiveness to the presence of mycorrhizae under ambient conditions also be the ones to which mycorrhizae confer the greatest benefit in elevated  $CO<sub>2</sub>$ ? Or will plants with little or no response to mycorrhizae under ambient  $CO<sub>2</sub>$  show the greatest benefit from mycorrhizae under elevated  $CO<sub>2</sub>$ ?

# Ecosystem level: carbon storage in the soil

Pool size of AM fungi in ecosystems

It is often overlooked that not only ectomycorrhizal fungi but also AM fungi can constitute a sizable pool of carbon in the soils of many terrestrial ecosystems (Allen 1991).

Estimates of extraradical hyphal lengths in the field vary widely. Reports range from  $0.02$  m  $g^{-1}$  soil in a poplar monoculture (Lussenhop et al. 1998), to 38 m ml<sup>-1</sup> (Allen and Allen 1986) in a shrub-steppe, to a peak length of  $111 \text{ m m}^{-1}$  of soil for a prairie community (Miller et al. 1995). For the latter estimate, an external hyphal dry weight of 457  $\mu$ g ml<sup>-1</sup> was calculated. This fungus carbon pool may also be a slow-turnover pool, because soil microarthropods seem to preferentially feed on saprophytic hyphae rather than on AM fungal hyphae (Klironomos and Kendrick 1996). Currently, we have no information concerning a potential change in the nutritional value of AM fungal hyphae in globalchange scenarios.

The far more significant pool of AM-fungus-related carbon in soil is probably the recently discovered glycoprotein glomalin (Wright and Upadhyaya 1996). Glomalin occurs in a wide variety of soils in the order of several mg per g of dry soil (Wright and Upadhyaya 1998) and is apparently only produced in significant amounts by AM fungus hyphae (Wright et al. 1996). It thus represents a pool of carbon (glomalin is approximately 20–30% carbon; S.F. Wright, personal communication) in soil that should be considered directly related to AM fungi. Very little is known about the turnover of this protein.

Potential for AM fungus carbon pool increase

Does this AM-fungi-related carbon pool have the potential for increase in elevated  $CO<sub>2</sub>$ ? Or are natural ecosystems already mycorrhiza-saturated (Allen et al. 1995)? This question must be answered in the field, after long-term  $CO<sub>2</sub>$  exposure in order to allow for all feedbacks to act upon the external hyphae and their products: grazers, litter input and quality, change in other soil microbes, change in glomalin production or decomposition rates, etc.

Lussenhop et al. (1998) found no significant difference in AM hypha biomass in high and low nitrogen soils in response to  $CO<sub>2</sub>$  exposure, whereas Klironomos et al. (1997) documented an increase in hyphal length under low-nutrient conditions with *Populus tremuloides*. Preliminary results from an experiment in an annual grassland in northern California indicate a potential for large increases in AM fungus biomass: approximately a 70% increase of AM fungus hyphal length after 6 years of  $CO<sub>2</sub>$  enrichment in the field (Rillig, Field and Allen 1999). Increases in AM fungus biomass constitute an ecosystem sink for carbon, providing a negative feedback to the atmospheric  $CO<sub>2</sub>$  concentration. In addition to carbon pool size, it is also necessary to examine carbon pool turnover of AM fungus hyphae; however, methodological limitations currently inhibit progress.

Glomalin concentration in soils has been shown to decrease when the soil and AM fungus mycelium is disturbed (Wright and Upadhyaya 1998). As more hyphae can be produced under elevated  $CO<sub>2</sub>$ , it is, therefore, possible that glomalin concentrations in soil increase after long-term  $CO<sub>2</sub>$  enrichment, or even in short-term experiments. If carbon allocation to the mycobiont increases, we speculate that AM fungi produce more of the protein (per hyphal length). Due to the unusually recalcitrant nature of the protein (it requires autoclaving for its extraction from soil; Wright and Upadhyaya 1996), it is interesting to speculate whether the carbon pathway ending in glomalin represents a bypass of the labile soil carbon pool to a slower turnover (more stable) carbon pool.

AM fungus effects on decomposition, and on carbon and nutrient cycling

AM hyphae are not involved in litter decomposition processes, but they take up nutrients (including nitrogen) and translocate them to the plant root. High AM fungus biomass may therefore, impose nutrient limitations on decomposer fungi in nutrient-limited ecosystems (Allen 1991). So far, only ectomycorrhizal fungi have been postulated to suppress decomposition by this mechanism (Gadgil and Gadgil 1971; but see Zhu and Ehrenfeld 1996). It is not known whether AM fungi can inhibit decomposition in similar ways, and whether this effect can be magnified by elevated  $CO<sub>2</sub>$ . In case AM

fungi prove to be important in this context, they would modify carbon cycling and retard the release of  $CO<sub>2</sub>$ back to the atmosphere, thereby increasing the system carbon sink.

### Conclusion

Structuring research within the framework used in this paper has heuristic value (MacMahon et al. 1978) in formulating questions for mycorrhizal contributions to ecosystem responses in elevated  $CO<sub>2</sub>$ . Because globalchange research also greatly depends on interdisciplinary collaboration because of the high investment necessary to maintain long-term  $CO<sub>2</sub>$  enrichment experiments, a structuring of research like this may also improve communication between researchers working at different scales.

**Acknowledgments** This work was supported by a US Department of Energy grant (Program for Ecosystems Research DE-FG03-93ER61669) to M. F. Allen. M. C. Rillig acknowledges a Doctoral Fellowship from the Studienstiftung des deutschen Volkes (Bonn, Germany). We thank E. G. O'Neill, G. B. Noe, J. N. Klironomos, and O. Kårén for helpful comments on an earlier version of this paper.

#### References

- Allen EB, Allen MF (1986) Water relations of xeric grasses in the field: interactions of mycorrhizae and competition. New Phytol 104:559–571
- Allen EB, Allen MF (1990) The mediation of competition by mycorrhizae in successional and patchy environments. In: Grace JR, Tilman D (eds) Perspectives on plant competition. Academic, New York, pp 367–389
- Allen MF (1991) The ecology of mycorrhizae. Cambridge University Press, Cambridge
- Allen MF (1996) The ecology of arbuscular mycorrhizas: a look back into the 20th century and a peek into the 21st. Mycol Res 100:769–782
- Allen MF, Sexton JC, Moore TS, Christensen M (1981) Comparative water relations and photosynthesis of mycorrhizal and non-mycorrhizal *Bouteloua gracilis* (J. B. K.) Lag ex Steud. New Phytol 88:683–693
- Allen MF, Morris SJ, Edwards F, Allen EB (1995) Microbe-plant interactions in Mediterranean-type habitats: shifts in fungal symbiotic and saprophytic functioning in response to globalchange. In: Moreno JM, Oechel WC (eds) Global-change and Mediterranean-type ecosystems (Ecological Studies Series 117). Springer, New York, pp 297–305
- Andersen CP, Rygiewicz PT (1991) Stress interactions and mycorrhizal plant response – understanding carbon allocation priorities. Environ Pollut 73 :217–244
- Bandara DC, Nobuyasu H, Ofosubudu KG, Ando T, Fujita K (1998) Effect of  $CO<sub>2</sub>$  enrichment on biomass production, photosynthesis, and sink activity in soybean CV Bragg and its supernodulating mutant NTS-1007. Soil Sci Plant Nutr 44:179–186
- Bazzaz FA (1990) The response of natural ecosystems to the rising global  $CO<sub>2</sub>$  levels. Annu Rev Ecol Syst  $21:167-196$
- Berntson GM, Bazzaz FA (1996) Belowground positive and negative feedbacks on  $CO<sub>2</sub>$  growth enhancement. Plant Soil 187:119–131
- Bever JD, Morton JB, Antonovics J, Schultz PA (1996) Host-dependent sporulation and species diversity of arbuscular mycorrhizal fungi in a mown grassland. J Ecol 84 :71–82
- Bryla DR, Duniway JM (1997) Water uptake by safflower and wheat roots infected with arbuscular mycorrhizal fungi. New Phytol 136: 591–601
- Cheplick GP (1997) Effects of endophytic fungi on the phenotypic plasticity of *Lolium perenne* (Poaceae). Am J Bot 84:34–40
- Chiariello NR, Hickman JC, Mooney HA (1982) Endomycorrhizal role for interspecific transfer of phosphorus in a community of annual plants. Nature 217:941–943
- Curtis PS, Klus DJ, Kalisz S, Tonsor SJ (1996) Intraspecific variation in CO2 response in *Raphanus raphanistrum* and *Plantago lanceolata*: assessing the potential of evolutionary change with rising atmospheric CO2. In: Körner C, Bazzaz FA (eds) Carbon dioxide, populations, and communities. Academic, San Diego, pp 13–22
- Díaz S (1996) Effects of elevated  $CO<sub>2</sub>$  at the community level mediated by root symbionts. Plant Soil 187: 309–320
- Díaz S, Grime JP, Harris J, McPherson E (1993) Evidence of a feedback mechanism limiting plant response to elevated carbon dioxide. Nature 364: 616–617
- Gadgil RL, Gadgil PD (1971) Mycorrhiza and litter decomposition. Nature 233:133
- George E, Häussler K, Kothari SK, Li X.-L, Marschner H (1992) Contribution of mycorrhizal hyphae to nutrient and water uptake of plants. In: Read DJ, Lewis DH, Fitter AH, Alexander IJ (eds) Mycorrhizas in ecosystems. CAB International, Wallingford, pp 42–47
- Giovannetti M, Gianinazzi-Pearson V (1994) Biodiversity in arbuscular mycorrhizal fungi. Mycol Res 98: 705–715
- Hodge A (1996) Impact of elevated  $CO<sub>2</sub>$  on mycorrhizal associations and implications for plant growth. Biol Fertil Soils 23:388–398
- Jakobsen I, Rosendahl L (1990) Carbon flow into soil and external hyphae from roots of mycorrhizal cucumber plants. New Phytol 115: 77–83
- Jones TH, Thompson LJ, Lawton JH, Bezemer TM, Bardgett RD, Blackburn TM, Bruce KD, Cannon PF, Hall GS, Hartley SE, Howson G, Jones CG, Kampichler C, Kandeler E, Ritchie DA (1998) Impacts of rising atmospheric carbon dioxide on model terrestrial ecosystems. Science 280 :441–443
- Jongen M, Fay P, Jones MB (1996) Effects of elevated carbon dioxide and arbuscular mycorrhizal infecton on *Trifolium repens*. New Phytol 132: 413–423
- Keeling CD, Whorf TP, Wahlen M, van der Plicht J (1995) Interannual extremes in the rate of rise of atmospheric carbon dioxide since 1980. Nature 375 :666–670.
- Klironomos JN, Kendrick WB (1996) Palatability of microfungi to soil arthropods in relation to the functioning of arbuscular mycorrhizae. Biol Fertil Soils 21: 43–52
- Klironomos JN, Rillig MC, Allen MF (1996) Below-ground microbial and microfaunal responses to *Artemisia tridentata* grown under elevated atmospheric  $CO<sub>2</sub>$ . Funct Ecol 10:537–534
- Klironomos JN, Rillig MC, Allen MF, Zak DR, Kubiske M, Pregitzer KS (1997) Soil fungal – arthropod responses to *Populus tremuloides* grown under enriched atmospheric CO<sub>2</sub> under field conditions. Global-change Biol 3 :473–478
- Klironomos JN, Ursic M, Rillig M, Allen MF (1998) Interspecific differences in the response of arbuscular mycorrhizal fungi to *Artemisia tridentata* grown under elevated atmospheric CO2. New Phytol 138:599–605
- Koide RT, Mooney HA (1987) Spatial variation in inoculum potential of vesicular-arbuscular mycorrhizal fungi caused by formation of gopher mounds. New Phytol 107 :173–182
- Lavorel S, McIntyre S, Landsberg J, Forbes TDA (1997) Plant functional classifications: from general groups to specific groups based on response to disturbance. Trends Ecol Evol 12:474–478
- Lussenhop J, Treonis A, Curtis PS, Teeri JA, Vogel SC (1998) Responses of soil biota to elevated atmospheric  $CO<sub>2</sub>$  in poplar model systems. Oecologia 113:247–251
- MacMahon JA, Phillips DL, Robinson JV, Schimpf DJ (1978) Levels of biological organization: an organism-centered approach. BioScience 28: 700–704
- Miller RM, Reinhardt DR, Jastrow JD (1995) External hyphal production of vesicular-arbuscular mycorrhizal fungi in pasture and tallgrass prairie communities. Oecologia 103:17–23
- Monz CA, Hunt HW, Reeves FB, Elliott ET (1994) The response of mycorrhizal colonization to elevated  $CO<sub>2</sub>$  and climate change in *Pascopyrum smithii* and *Bouteloua gracilis*. Plant Soil 165: 75–80
- Morton JB, Bentivenga SP (1994) Levels of diversity in endomycorrhizal fungi (Glomales, Zygomycetes) and their role in defining taxonomic and non-taxonomic groups. Plant Soil 159:47–59
- Newsham KK, Fitter AH, Watkinson AR (1995) Multi-functionality and biodiversity in arbuscular mycorrhizas. Trends Ecol Evol 10:407-411
- O'Neill EG (1994) Responses of soil biota to elevated atmospheric carbon dioxide. Plant Soil 165 :55–65
- O'Neill EG, O'Neill RV, Norby RJ (1991) Hierarchy theory as a guide to mycorrhizal research on large-scale problems. Environ Pollut 73 :271–284
- O'Neill RV, DeAngelis DL, Waide JB, Allen TFH (1986) A hierarchical concept of ecosystems. Princeton University Press, Princeton
- Paterson E, Hall JM, Ramsay EAS, Rattray BS, Griffiths BS, Ritz K (1997) Effect of elevated  $CO<sub>2</sub>$  on rhizosphere carbon flow and soil microbial processes. Global change Biol 3 :363–378
- Paul EA, Kucey RMN (1981) Carbon flow in plant microbial associations. Science 213 :473–474
- Ravnskov S, Jakobsen I (1995) Functional compatibility in arbuscular mycorrhizas measured as hyphal P transport to the plant. New Phytol 129:611–618
- Rillig MC, Allen MF (1998) Arbuscular mycorrhizae of *Gutierrezia sarothrae* and elevated carbon dioxide: evidence for shifts in C allocation to and within the mycobiont. Soil Biol Biochem 30: 2001–2008
- Rillig MC, Allen MF, Klironomos JN, Chiariello NR, Field CB (1998) Plant-specific changes in root-inhabiting fungi in a California annual grassland: responses to elevated  $CO<sub>2</sub>$  and nutrients. Oecologia 113 :252–259
- Rillig MC, Allen MF, Klironomos JN, Field CB (1998) Arbuscular mycorrhizal percent infection and infection intensity of *Bromus hordeaceus* grown in elevated atmospheric CO<sub>2</sub>. Mycologia 90:199–205
- Rillig MC, Field CB, Allen MF (1999) Soil biota responses to long-term atmospheric  $CO<sub>2</sub>$  enrichment in two California annual grasslands. Oecologia (in press)
- Rogers HH, Runion GB, Krupa SV (1994) Plant responses to atmospheric  $CO<sub>2</sub>$  enrichment with emphasis on roots and the rhizosphere. Environ Pollut 83: 155–189
- Rouhier H, Billes G, Bottner P (1996) Carbon fluxes in the rhizosphere of sweet chestnut seedlings (*Castanea sativa*) grown under 2 atmospheric  $CO<sub>2</sub>$  concentrations. Plant Soil 180:101–111
- Rouhier, H, Read DJ (1998) The role of mycorrhiza in determining the response of *Plantago lanceolata* to  $CO<sub>2</sub>$  enrichment. New Phytol 139:367–373
- Runion GB, Curl EA, Rogers HH, Backman PA, Rodriguez-Kabana R, Helms BE (1994) Effects of free-air  $CO<sub>2</sub>$  enrichment on microbial populations in the rhizosphere and phyllosphere of cotton. Agricultural and Forest Meteorology 70: 117–130
- Sadowsky MJ, Schortemeyer M (1997) Soil microbial responses to increased concentrations of atmospheric  $CO<sub>2</sub>$ . Globalchange Biol 3: 217–224
- Sanders IR (1996) Plant-fungal interactions in a  $CO_2$ -rich world. In: Körner C, Bazzaz FA (eds) Carbon dioxide, populations, and communities. Academic, San Diego, pp 265–272
- Schmid B, Birrer A, Lavigne C (1996) Genetic variation in the response of plant populations to elevated  $CO<sub>2</sub>$  in a nutrientpoor, calcareous grassland. In: Körner C, Bazzaz FA (eds) Carbon dioxide, populations, and communities. Academic, San Diego, pp 31–50
- Smith SE, Read DJ (1997) Myorrhizal symbiosis. Academic, San Diego
- Staddon PL, Graves JD, Fitter AH (1998) Effect of enhanced atmospheric CO<sub>2</sub> on mycorhizal colonization by *Glomus mossea* in *Plantago lanceolata* and *Trifolium repens*. New Phytol 139:571–580
- Staddon PL, Fitter AH, Graves JD (1999a) Effect of elevated atmospheric  $CO<sub>2</sub>$  on mycorrhizal colonization, external mycorrhizal hyphal production and phosphorus inflow in *Plantago lanceolata* and *Trifolium repens* in association with the arbuscular mycorrhizal fungus *Glomus mossea*. Global Change Biology 5: 347–358
- Staddon PL, Graves JD, Fitter AH (1999b) Effect of enhanced atmospheric  $CO<sub>2</sub>$  on mycorrhizal colonization and phosphorus inflow in 10 herbaceous species of contrasting growth strategies. Funct Ecol (in press)
- Streitwolf-Engel R, Boller T, Wiemken A, Sanders IR (1997) Clonal growth traits of two *Prunella* species are determined by co-occurring arbuscular mycorrhizal fungi from a calcareous grassland. J Ecol 85:181–191
- Van der Heijden MGA, Klironomos JN, Ursic M, Moutoglis P, Streitwolf-Engel R, Boller T, Wiemken T, Sanders IR (1998) Mycorrhizal fungal diversity determines plant biodiversity, ecosystem variability and productivity. Nature 396: 69–72
- Wright DP, Scholes JD, Read DJ (1998) Effects of VA mycorrhizal colonization on photosynthesis and biomass production of *Trifolium repens* L. Plant Cell Environ 21 :209–216
- Wright SF, Upadhyaya A (1996) Extraction of an abundant and unusual protein from soil and comparison with hyphal protein of arbuscular mycorrhizal fungi. Soil Sci 161:575–586
- Wright SF, Upadhyaya A (1998) A survey of soils for aggregate stability and glomalin, a glycoprotein produced by hyphae of arbuscular mycorrhizal fungi. Plant Soil 198 :97–107
- Wright SF, Franke-Snyder M, Morton JB, Updahyaya A (1996) Time-course study and partial characterization of a protein on hyphae of arbuscular-mycorrhizal fungi during active colonization of roots. Plant Soil 181:193–203
- Zak DR, Pregitzer KS, Curtis PS, Teeri JA, Fogel R, Randlett DL (1993) Elevated atmospheric  $CO<sub>2</sub>$  and feedbacks between carbon and nitrogen cycles. Plant Soil 151:105–117
- Zanetti S, Hartwig UA, Luscher A, Hebeisen T, Frehner M, Fischer BU, Hendrey GR, Blum H, Nosberger J (1996) Stimulation of symbiotic N2 fixation in *Trifolium repens* L. under elevated atmospheric  $pCO<sub>2</sub>$  in a grassland ecosystem. Plant Physiol 112: 575–583
- Zanetti S, Hartwig UA (1997) Symbiotic  $N_2$  fixation increases under elevated atmospheric  $pCO<sub>2</sub>$  in the field. Acta Oecol 18:285–290
- Zhu WX, Ehrenfeld JG (1996) The effects of mycorrhizal roots on litter decomposition, soil biota, and nutrients in a spodosolic soil. Plant Soil 179:109–118
- Zobel M, Moora M, Haukioja E (1997) Plant coexistence in the interactive environment: arbuscular mycorrhiza should not be out of mind. Oikos 78: 202–208