

### Stem-cell therapy for hepatobiliary pancreatic disease

TAKAHISA FUJIKAWA, SEH-HOON OH, TOM SHUPE, and BRYON E. PETERSEN

Department of Pathology, Immunology and Laboratory Medicine and Program in Stem Cell Biology and Regenerative Medicine, College of Medicine, PO Box 100275, University of Florida, Gainesville, FL, USA

#### Abstract

The transplantation of pancreatic beta cells or hepatocytes represents a potential therapeutic approach for type I diabetes and inherited liver diseases, respectively. Furthermore, acquired liver diseases, particularly acute hepatic failure due to toxic or viral injury, have been treated in limited clinical trials with fetal and adult hepatocytes. However, a major limitation is the insufficient amount of beta cells and hepatocytes available for grafts. Alternative sources of these cells have yet to be determined. During the past few years, progress has been made in the development of new strategies to produce mature beta cells and hepatocytes. In this review, we outline the current state of scientific understanding and controversy regarding the properties of embryonic and adult stem cells in the field of hepatobiliary and pancreatic diseases. Our objective is to provide a framework of understanding for the challenges behind translating fundamental stem cell biology into clinical therapies.

Key words Stem-cell therapy  $\cdot$  Hepatobiliary pancreatic disease  $\cdot$  Embryonic stem cell  $\cdot$  Bone marrow cell  $\cdot$  Diabetes mellitus

#### Introduction

Stem cells are cells that are capable of self-renewal and are multipotent, meaning that they can differentiate into many specific cell types. For a long time, "stem cell" has been a concept in mammalian biology but not a reality that could be seen, manipulated, and expanded in vitro. This has changed with the establishment of murine embryonic stem cell (ESC) culture in 1981.<sup>1,2</sup> Through progress made in stem cell research, not only ESCs but also various stem/progenitor cells derived from adult tissues have been isolated and established. These include hematopoietic stem cells,<sup>3-7</sup> neural stem cells,<sup>8.9</sup> vascular endothelial progenitor cells,<sup>10</sup> and hepatic oval cells.<sup>11-13</sup> Recent stem cell-based cell therapies have shown success in the treatment of various animal models for diseases, such as Parkinson's disease, type I diabetes, and inherited genetic liver diseases.

In this review, we survey the latest development in the study of both ESCs and adult stem cells (specifically stem cells from bone marrow (BM)) for the treatment of hepatobiliary and pancreas diseases, with an emphasis on application to clinical therapies. We summarize and discuss stem cell-based therapies for both pancreas and liver diseases, and focus on the issues related to the mechanism of stem cell adaptation, including "transdifferentiation" and "cell fusion."

#### Stem cell therapy for pancreas diseases

Type I diabetes, or insulin-dependent diabetes, represents a major pancreatic disease with tremendous appeal as a target for cell replacement therapy. This disorder, which results from the loss of insulinproducing beta islet cells due to autoimmune attack, can be reversed by pancreas or islet cell transplantation together with steroid-sparing immunosuppression.<sup>14,15</sup> Diabetes may be particularly suited to cell transplantation thereby because, unlike in Parkinson's disease where precise connections may be necessary, beta cells can function autonomously, even outside the pancreas (e.g., under the kidney capsule). The chief limitation to the wide application of this potentially curative therapy is the inadequate supply of islets from cadavers.

Offprint requests to: B.E. Petersen

Received: January 31, 2005 / Accepted: February 23, 2005

# Embryonic stem cells for the treatment of diabetes mellitus

During the past few years, various groups have tested the possibility of producing beta cells from ESCs. Soria et al.<sup>16</sup> used a cell-trapping system to select mouse ESCs able to transcribe the insulin gene, and such cells corrected hyperglycemia when implanted in the spleen of diabetic mice. Lumelsky et al.17 designed a new protocol to generate in vitro insulin containing cells from mouse ESCs. These cells secreted insulin as mature pancreatic beta cells, although they could not reverse hyperglycemia when grafted into diabetic mice. Recently, several groups have further modified in vitro protocols.<sup>16-22</sup> The addition of phosphoinositide 3-kinase inhibitors promotes differentiation of a larger number of ESCs toward functional beta cells.<sup>20</sup> With manipulation of the culture conditions and the use of pax4 or pdx-1, both of which are transcription factors associated with beta-cell lineage, more efficient and defined ways of making ESC-derived beta-like cells have been established.<sup>21,23</sup> Much of this rodent data has been adapted for use on human ESCs. The production of insulin by human ESCs has recently been demonstrated.<sup>19,24</sup> Techniques that do not require murine feeder cells have also been developed, allowing for single species propagation of ESCs, and thus avoiding possible zoonotic infection of cells intended for clinical use.25

Several researchers have argued that the insulinpositive cells derived from ESCs may not be true "insulin-producing" beta-like cells.<sup>26,27</sup> Rajagopal et al.<sup>26</sup> showed that contrary to previous reports, no message for insulin was detectable in cultured beta-like cells, which suggested that the cells may be concentrating the hormone from the medium rather than actually producing it. Sipione et al.27 demonstrated that the main producers of insulin in culture were neurons and neuronal precursors, and further demonstrated that a reporter gene under the control of the insulin I promoter was activated in cells with a neuronal phenotype. In addition, problems in the control of differentiation and teratoma formation from ESC-derived insulin-producing cells remain to be overcome.18,27 Other investigators claim that since existing ESC lines are not assumed to be identical, and none are ideal for generating islets or beta cells, additional ESC lines must be generated.<sup>28</sup> Ethical concerns about the use of ESCs and policies regarding the funding of ESC research need to be addressed before we can wholly evaluated this powerful new technology. Theoretically, ESCs may represent a cell type from which large quantities of functional insulin-producing cells may be generated. However, this remains to be definitively demonstrated.

# Bone-marrow cells for the treatment of diabetes mellitus

The BM harbors cells that can become parenchymal cells after entering the liver, intestine, skin, lung, skeletal muscle, heart muscle, and central nervous system in rodent models<sup>29</sup> and in human recipients of marrow or organ transplantation.<sup>30,31</sup> In rodents, the BM also harbors cells that can differentiate into functional pancreatic endocrine cells.32-37 One month after BM transplantation, donor-derived cells are found in pancreatic islets of recipient mice.32 These cells express insulin and genetic markers of beta cells. Although the cells secrete insulin in response to glucose in culture, only about 1% of the islet cells originate form the transplanted marrow.32 A marrow-derived cell type with pluripotential capacity to differentiate into various phenotypes has been described.<sup>38</sup> This or a similar cell type might be able to differentiate into pancreatic beta cells. Similar experiments have been done in overtly diabetic mice whose beta cells have been destroyed by streptozotocin. After BM transplantation, blood glucose and insulin concentrations were normal, and survival was better.33

Recent studies demonstrated that BM cells can differentiate in vitro under controlled conditions into insulin-expressing cells.<sup>38-40</sup> Such cells, when transplanted under the kidney capsule of diabetic rodents, reverse hyperglycemia. The removal of the grafted kidney returned the animals to a diabetic state.<sup>40</sup>

However, the discussion that cell fusion rather than true cellular differentiation might account for the detection of donor cells in regenerating tissues has muted some of the enthusiasm for genuine beta-cell differentiation.<sup>37,41</sup> Although studies involving pancreatic endocrine-cell differentiation from haemopoietic organ derivatives largely rule out cell-fusion events as a mechanism of differentiation,<sup>32–36</sup> much more work remains to be done before this issue is settled.

#### Stem cell therapy for liver diseases

The practical implications of stem cell therapy for liver diseases are that they might serve as a source of cell transplantation and bioartificial liver devices, as well as targets for gene therapies. Direct transplantation of hepatocytes has already been used to bridge patients to orthotopic liver transplant and as a therapeutic alternative to whole liver transplantation.<sup>42–46</sup> Among the diseases with the highest potential to benefit from stem cell therapy are primary liver diseases with extrahepatic manifestations arising from abnormal gene expression or defective protein production by the liver.<sup>46–48</sup> These diseases include genetic disorders such as alpha-1antitrypsin deficiency, hemochromatosis, tyrosinemia type I, and Wilson's disease. Also, metabolic deficiencies such as type I Crigler–Najjar syndrome, familial hypercholesterolemia, oxalosis, and familial amyloidotic polyneuropathy, as well as coagulation defects such as hemophilia, may be particularly suited to stem cell therapies.

To date, hepatocyte transplantation has been accomplished in a relatively small number of such patients.<sup>49,50</sup> Acquired liver disease, particularly acute hepatic failure due to toxic or viral injury, have been treated in limited clinical trials with fetal and adult hepatocytes.<sup>51–54</sup> As seen in islet cell transplantation in patients with type I diabetes, the major limiting factor is our inability to produce large quantity of hepatocytes and to keep them ready for use on demand. While bioartificial liver devices may be in clinical use for bridging patients with acute hepatic failure to survival or transplant, their use in patients with chronic liver disease may also require large numbers of hepatocytes. A stable and expandable stem cell culture is expected to provide the means to create such large numbers of cells.

### Embryonic stem cells for the treatment of liver diseases

Several recent reports from different groups mention not only in vitro differentiation mechanism of ESCderived liver cells but also in vivo differentiation after transplantation to several liver disease models. Using various culture conditions or a beta-galactosidaseactivating gene-trapping system, some cells derived from embryoid bodies cultured from mouse ESCs acquire the ability to express message for alpha-feto protein (AFP), albumin, transthyretin, and alpha-1antitrypsin.55,56 Yin et al.57 reported that AFP-positive cells isolated from cultured ESCs differentiate into hepatocytes when transplanted into livers of mice that lack the ability to express either apolipoprotein E or haptoglobin. Other reports from Yamada et al.58 and Chinzei et al.59 demonstrate that ESC-derived cells in embryoid bodies that express mRNAs for albumin, AFP, and other mature hepatocyte markers incorporate into hepatic plates, produce albumin protein, and morphologically resemble adjacent hepatocytes when transplanted into female mice. Although some promising data were recently reported, any real clinical impact awaits the clear directed differentiation of appropriate cell populations and further investigation into each targeted disease.

### Bone-marrow cells for the treatment of liver diseases

BM contains several types of stem cells, including hematopoietic stem cells (HSC); mesenchymal or stromal stem cells, which include multipotent adult progenitor cells (MAPC), and endothelial stem cells. The transplantation of unfractionated BM into the livers of lethally irradiated rats rescues animals from radiationinduced BM ablation and simultaneously yields small numbers of BM-derived hepatic oval cells (and later hepatocytes).<sup>13,60</sup> Recent results show that HSCs can transform into hepatocytes both in vivo and in vitro,<sup>61–63</sup> and that MAPCs acquire hepatocyte differentiation in cell culture<sup>38,57</sup> and give rise to hepatocytes when the cultured cells are transplanted into the liver.<sup>64</sup>

These findings have led to the speculation that HSCs (rather than endogenous liver epithelial cells) are the major source of stem cells involved in the generation of new liver parenchyma following liver damage.65 Although recent studies clearly demonstrate that cells originating in the BM contribute to the formation of a few hepatocytes, cholangiocytes, and oval cells in both healthy and diseased livers,13,60 the frequency of this transformation is low.66-69 Only the fact that HSCs circulate in the blood<sup>70</sup> and may be found in the adult liver<sup>71</sup> supports the argument that HSCs are a major source of cells for regenerating liver parenchyma. Wang et al.<sup>72</sup> demonstrated that endogenous liver epithelial cells are the most efficient type of cell at mediating liver regeneration upon transplantation to damaged liver. These studies indicate that a variety of stem-like cells from several tissues can transform into hepatocytes when transplanted into the liver in vivo or cultured in vitro.72 Which of these has the highest potential for therapeutic use remains to be determined.

The mechanism by which BM stem cells transform into hepatocytes has been hypothesized to be direct transdifferentiation, reflecting the phenotypic plasticity of stem cells within different tissue microenvironments. The most impressive generation of hepatocytes from BM cells has occurred after transplantation of BM into mice with lethal hepatic failure resulting from homozygous deletion of the fumaryl acetoacetate hydrolase (Fah) gene.<sup>61</sup> Subsequent studies in the Fah<sup>-/-</sup> model suggest that the apparent transdifferentiation of HSCs into hepatocytes results from the fusion of HSC descendant cells (possibly macrophages) with Fah-/- hepatocytes to yield heterokaryotic cells in which hepatocytic phenotypes are dominant and the Fah--- genotypes is corrected.73,74 The metabolically corrected heterokaryotic hepatocytes proliferate rapidly and replace the defective host liver parenchyma.75 Because such extensive replacement of hepatocytes has not been found in other models of damaged liver, it is possible that the accumulation of high levels of fumaryl acetoacetate and the strong stimulus for proliferation of cells with corrected metabolic defects may be responsible for this phenomenon, whether by fusion or transdifferentiation. Further research is required to reach a final conclusion.

### Discussion

The extraordinary activity and interest that currently characterizes research on stem cells afford an opportunity to reassess concepts of liver stem cells. This reevaluation may alter major paradigms of stem cell biology in fundamental ways.<sup>76,77</sup> Recent research on stem cell therapy for liver and pancreatic diseases has emphasized methods that currently dominate all stem cell research: analysis of stem cell differentiation resulting from the transplantation of freshly isolated or cultured cells into tissues in vivo and/or by subjecting these cells to defined conditions of cell culture in vitro. These studies have led to various hypotheses on the nature of stem cells and their potential clinical application regardless of the initial stem cell source.

Compared with hematopoietic cells and neuronal cells, there may be some delay in the progress of stem cell therapy for liver and pancreatic diseases. There is some difficulty in generating endodermal cells from stem cells, because attempts to apply factors characteristic of late liver and pancreas development to essentially very early-stage cells have not been fruitful. Although challenging, generating endoderm is probably the most rewarding lineage toward which stem cells may be directed. Given the fact that both islet cell and hepatocyte transplantation can restore function in patients with type I diabetes and inherited liver diseases, respectively, stem cell-based therapies for liver and pancreas diseases represent one of the most compelling opportunities in regenerative medicine. Real clinical impact will require the clear directed differentiation of appropriate cell populations. Present claims, many of which may be exaggerated, await further confirmation.

Concerning the BM stem cells such as HSCs, some of the early reports on stem cell "transdifferentiation" show the generation of functional hepatocytes or beta cells after BM transplantation. Several consequent studies showed similar results even when using highly purified HSCs or in vitro cultured cells derived from BM. However, owing to the heterogeneity of BM, the possibility that nonhematopoietic stem cells were contained within the transplanted marrow remains. Even in carefully executed studies, impurities in the enriched stem cells could have accounted for nonhematopoietic regeneration. True evidence, such as clonal analysis studies, will be required before a new understanding of fundamental embryological concepts can be accepted.

The idea that cell fusion may contribute to the endodermal differentiation of stem cells does not preclude stem cell-based therapies. Two recent papers suggest that the fusion of differentiated cells with ESCs in vitro could lead to functional cells with stem cell-like properties.78,79 This led to the idea that transplanted BM stem cells or their progeny were fusing to matured nonhematopoietic cells, resulting in cells with new phenotype and function. In the liver and pancreas, the fusion of hematopoietic cells and endogenous functional cells does appear to be at least part of the mechanism of the generation of hepatocytes or beta cells from BM cells.<sup>37,41,73,74</sup> Because both cell fusion and differentiation of stem cells have been shown to be possible mechanisms for generating mature hepatocyte and pancreatic beta cell phenotypes, further studies will be required to determine the relative frequency of differentiation and cell fusion in particular settings to define the conditions that regulate each mechanism. In addition, there is little debate that a wide range of frequencies have been reported, and the prevalence of such events is extremely low or undetectable. What the discrepancies really indicate is that we have a great deal of basic research left to do.

Acknowledgment. Grant and source of support: National Institute of Health Grants DK60015 and DK58614 awarded to BEP funded this research.

#### References

- Evans MJ, Kaufman MH (1981) Establishment in culture of pluripotential cells from mouse embryos. Nature 292:154–156
- Martin GR (1981) Isolation of a pluripotent cell line from early mouse embryos cultured in medium conditioned by teratocarcinoma stem cells. Proc Natl Acad Sci USA 78:7634–7638
- Visser JW, Bauman JG, Mulder AH, Eliason JF, de Leeuw AM (1984) Isolation of murine pluripotent hemopoietic stem cells. J Exp Med 159:1576–1590
- Muller-Sieburg CE, Whitlock CA, Weissman IL (1986) Isolation of two early B lymphocyte progenitors from mouse marrow: a committed pre-pre-B cell and a clonogenic Thy-1-lo hematopoietic stem cell. Cell 44:653–662
- Spangrude GJ, Heimfeld S, Weissman IL (1988) Purification and characterization of mouse hematopoietic stem cells. Science 241: 58–62
- Goodell MA, Brose K, Paradis G, Conner AS, Mulligan RC (1996) Isolation and functional properties of murine hematopoietic stem cells that are replicating in vivo. J Exp Med 183:1797– 1806
- Osawa M, Hanada K, Hamada H, Nakauchi H (1996) Long-term lymphohematopoietic reconstitution by a single CD34-low/negative hematopoietic stem cell. Science 273:242–245
- Bjornson CR, Rietze RL, Reynolds BA, Magli MC, Vescovi AL (1999) Turning brain into blood: a hematopoietic fate adopted by adult neural stem cells in vivo. Science 283:534–537
- Clarke DL, Johansson CB, Wilbertz J, Veress B, Nilsson E, Karlstrom H, Lendahl U, Frisen J (2000) Generalized potential of adult neural stem cells. Science 288:1660–1663

- Asahara T, Murohara T, Sullivan A, Silver M, van der Zee R, Li T, Witzenbichler B, Schatteman G, Isner JM (1997) Isolation of putative progenitor endothelial cells for angiogenesis. Science 275:964–967
- 11. Sell S (1990) Is there a liver stem cell? Cancer Res 50:3811– 3815
- Petersen BE, Zajac VF, Michalopoulos GK (1998) Hepatic oval cell activation in response to injury following chemically induced periportal or pericentral damage in rats. Hepatology 27:1030– 1038
- Petersen BE, Bowen WC, Patrene KD, Mars WM, Sullivan AK, Murase N, Boggs SS, Greenberger JS, Goff JP (1999) Bone marrow as a potential source of hepatic oval cells. Science 284:1168– 1170
- Sutherland DE, Gruessner RW, Dunn DL, Matas AJ, Humar A, Kandaswamy R, Mauer SM, Kennedy WR, Goetz FC, Robertson RP, Gruessner AC, Najarian JS (2001) Lessons learned from more than 1000 pancreas transplants at a single institution. Ann Surg 233:463–501
- Ricordi C, Strom TB (2004) Clinical islet transplantation: advances and immunological challenges. Nat Rev Immunol 4: 259–268
- Soria B, Roche E, Berna G, Leon-Quinto T, Reig JA, Martin F (2000) Insulin-secreting cells derived from embryonic stem cells normalize glycemia in streptozotocin-induced diabetic mice. Diabetes 49:157–162
- Lumelsky N, Blondel O, Laeng P, Velasco I, Ravin R, McKay R (2001) Differentiation of embryonic stem cells to insulin-secreting structures similar to pancreatic islets. Science 292:1389–1394
- Fujikawa T, Oh SH, Pi L, Hatch HM, Shupe T, Petersen BE (2005) Teratoma formation leads to failure of treatment for type I diabetes using embryonic stem cell-derived insulin-producing cells. Am J Pathol in press
- Assady S, Maor G, Amit M, Itskovitz-Eldor J, Skorecki KL, Tzukerman M (2001) Insulin production by human embryonic stem cells. Diabetes 50:1691–1697
- Hori Y, Rulifson IC, Tsai BC, Heit JJ, Cahoy JD, Kim SK (2002) Growth inhibitors promote differentiation of insulin-producing tissues from embryonic stem cells. Proc Natl Acad Sci USA 99: 16105–16110
- Blyszczuk P, Czyz J, Kania G, Wagner M, Roll U, St-Onge L, Wobus AM (2003) Expression of Pax4 in embryonic stem cells promotes differentiation of nestin-positive progenitor and insulin-producing cells. Proc Natl Acad Sci USA 100:998–1003
- 22. Kim D, Gu Y, Ishii M, Fujimiya M, Qi M, Nakamura N, Yoshikawa T, Sumi S, Inoue K (2003) In vivo functioning and transplantable mature pancreatic islet-like cell clusters differentiated from embryonic stem cells. Pancreas 27:e34–41
- Miyazaki S, Yamato E, Miyazaki J (2004) Regulated expression of pdx-1 promotes in vitro differentiation of insulin-producing cells from embryonic stem cells. Diabetes 53:1030–1037
- Segev H, Fishman B, Ziskind A, Shulman M, Itskovitz-Eldor J (2004) Differentiation of human embryonic stem cells into insulin-producing clusters. Stem Cells 22:265–274
- Moritoh Y, Yamato E, Yasui Y, Miyazaki S, Miyazaki J (2003) Analysis of insulin-producing cells during in vitro differentiation from feeder-free embryonic stem cells. Diabetes 52:1163–1168
- Rajagopal J, Anderson WJ, Kume S, Martinez OL, Melton DA (2003) Insulin staining of ES cell progeny from insulin uptake. Science 299:363
- Sipione S, Eshpeter A, Lyon JG, Korbutt GS, Bleackley RC (2004) Insulin-expressing cells from differentiated embryonic stem cells are not beta cells. Diabetologia 47:499–508
- Cowan CA, Klimanskaya I, McMahon J, Atienza J, Witmyer J, Zucker JP, Wang S, Morton CC, McMahon AP, Powers D, Melton DA (2004) Derivation of embryonic stem-cell lines from human blastocysts. N Engl J Med 350:1353–1356
- Herzog EL, Chai L, Krause DS (2003) Plasticity of marrowderived stem cells. Blood 102:3483–3493

- Theise ND, Nimmakayalu M, Gardner R, Illei PB, Morgan G, Teperman L, Henegariu O, Krause DS (2000) Liver from bone marrow in humans. Hepatology 32:11–16
- Korbling M, Katz RL, Khanna A, Ruifrok AC, Rondon G, Albitar M, Champlin RE, Estrov Z (2002) Hepatocytes and epithelial cells of donor origin in recipients of peripheral-blood stem cells. N Engl J Med 346:738–746
- 32. Ianus A, Holz GG, Theise ND, Hussain MA (2003) In vivo derivation of glucose-competent pancreatic endocrine cells from bone marrow without evidence of cells fusion. J Clin Invest 111:843–850
- Hess D, Li L, Martin M, Sakano S, Hill D, Strutt B, Thyssen S, Gray DA, Bhatia M (2003) Bone marrow-derived stem cells initiate pancreatic regeneration. Nat Biotechol 21:763–770
- 34. Zorina TD, Subbotin VM, Bertera S, Alexander AM, Haluszczak C, Gambrell B, Bottino R, Styche AJ, Trucco M (2003) Recovery of the endogenous beta cell function in the NOD model of autoimmune diabetes. Stem Cells 21:377–388
- Ryu S, Kodama S, Ryu K, Schoenfeld DA, Faustman DL (2001) Reversal of established autoimmune diabetes by restoration of endogenous beta cell function. J Clin Invest 108:63–72
- Kodama S, Kuhtreiber W, Fujimura S, Dale EA, Faustman DL (2003) Islet regeneration during the reversal of autoimmune diabetes in NOD mice. Science 302:1223–1227
- Kofman AV, Theise ND, Hussain MA (2004) Paradigms of adult stem cell therapy for type 1 diabetes in mice. Eur J Endocrinol 150:415–419
- 38. Jiang Y, Jahagirdar BN, Reinhardt RL, Schwartz RE, Keene CD, Ortiz-Gonzalez XR, Reyes M, Lenvik T, Lund T, Blackstad M, Du J, Aldrich S, Lisberg A, Low WC, Largaespada D, Verfaillie CM (2002) Pluripotency of mesenchymal stem cells derived from adult marrow. Nature 418:41–49
- Jahr H, Bretzel RG (2003) Insulin-positive cells in vitro generated from rat bone marrow stromal cells. Transplant Proc 35:2140– 2141
- 40. Oh SH, Muzzonigro TM, Bae SH, LaPlante JM, Hatch HM, Petersen BE (2004) Adult bone marrow-derived cells transdifferentiating into insulin-producing cells for the treatment of type I diabetes. Lab Invest 84:607–617
- Theise ND, Wilmut I (2003) Cell plasticity: flexible arrangement. Nature 425:21
- Stanworth SJ, Newland AC (2001) Stem cells: progress in research and edging towards the clinical setting. Clin Med 1:378– 382
- Ohashi K, Park F, Kay MA (2001) Hepatocyte transplantation: clinical and experimental application. J Mol Med 79:617–630
- 44. Susick R, Moss N, Kubota H, Lecluyse E, Hamilton G, Luntz T, Ludlow J, Fair J, Gerber D, Bergstrand K, White J, Bruce A, Drury O, Gupta S, Reid LM (2001) Hepatic progenitors and strategies for liver cell therapies. Ann NY Acad Sci 944:398– 419
- 45. Grompe M (2001) Liver repopulation for the treatment of metabolic diseases. J Inherit Metab Dis 24:231–244
- Laconi E, Laconi S (2002) Principles of hepatocyte repopulation. Semin Cell Dev Biol 13:433–438
- Min AD, Theise ND (2004) Prospects for cell-based therapies for liver disease. Panminerva Med 46:43–48
- Gupta S, Chowdhury JR (2002) Therapeutic potential of hepatocyte transplantation. Semin Cell Dev Biol 13:439–446
- Fox IJ, Chowdhury JR, Kaufman SS, Goetzen TC, Chowdhury NR, Warkentin PI, Dorko K, Sauter BV, Strom SC (1998) Treatment of the Crigler–Najjar syndrome type I with hepatocyte transplantation. N Engl J Med 338:1422–1426
- Muraca M, Gerunda G, Neri D, Vilei MT, Granato A, Feltracco P, Meroni M, Giron G, Burlina AB (2002) Hepatocyte transplantation as a treatment for glycogen storage disease type 1a. Lancet 359:317–318
- Habibullah CM, Syed IH, Qamar A, Taher-Uz Z (1994) Human fetal hepatocyte transplantation in patients with fulminant hepatic failure. Transplantation 58(8):951–952

- Strom SC, Fisher RA, Thompson MT (1997) Hepatocyte transplantation as a bridge to orthotopic liver transplantation in terminal liver failure. Transplantation 63:559–569
- Bilir BM, Guinette D, Karrer F (2000) Hepatocyte transplantation in acute liver failure. Liver Transpl 6:32–40
- Fisher RA, Bu D, Thompson M (2000) Defining hepatocellular chimerism in a liver failure patient bridged with hepatocyte infusion. Transplantation 69:303–307
- 55. Hamazaki T, Iiboshi Y, Oka M, Papst PJ, Meacham AM, Zon LI, Terada N (2001) Hepatic maturation in differentiating embryonic stem cells in vitro. FEBS Lett 497:15–19
- Jones EA, Tosh D, Wilson DI (2002) Hepatic differentiation of murine embryonic stem cells. Exp Cell Res 272:15–22
- Yin Y, Lim YK, Salto-Tellez M (2002) AFP(+), ESC-derived cells engraft and differentiate into hepatocytes in vivo. Stem Cells 20:338–346
- Yamada T, Yoshikawa M, Kanda S (2000) In vitro differentiation of embryonic stem cells into hepatocyte-like cells identified by cellular uptake of indocyanine green. Stem Cells 20:146– 154
- 59. Chinzei R, Tanaka Y, Shimizu-Saito K, Hara Y, Kakinuma S, Watanabe M, Teramoto K, Arii S, Takase K, Sato C, Terada N, Teraoka H (2002) Embryoid-body cells derived from a mouse embryonic stem cell line show differentiation into functional hepatocytes. Hepatology 36:22–29
- Theise ND, Badve S, Saxena R (2000) Derivation of hepatocytes from bone marrow cells in mice after radiation-induced myeloablation. Hepatology 31:235–240
- Lagasse E, Connors H, Al-Dhalimy M, Reitsma M, Dohse M, Osborne L, Wang X, Finegold M, Weissman IL, Grompe M (2000) Purified hematopoietic stem cells can differentiate into hepatocytes in vivo. Nat Med 6:1229–1234
- 62. Wang X, Ge S, McNamara G (2003) Albumin-expressing hepatocyte-like cells develop in the livers of immune-deficient mice that received transplants of highly purified human hematopoietic stem cells. Blood 101:4201–4208
- Danet GH, Luongo JL, Butler G (2002) C1qRp defines a new human stem cell population with hematopoietic and hepatic potential. Proc Natl Acad Sci USA 99:10441–10445
- Schwartz RE, Reyes M, Koodie L (2002) Multipotent adult progenitor cells from bone marrow differentiate into functional hepatocyte-like cells. J Clin Invest 109:1291–1302

- 65. Thorgeirsson SS, Grisham JW (2003) Overview of recent experimental studies on liver stem cells. Semin Liver Dis 23:303–312
- Gao Z, McAlister VC, Williams GM (2001) Repopulation of liver endothelium by bone-marrow-derived cells. Lancet 357:932–933
- Krause DS, Theise ND, Collector MI, Henegariu O, Hwang S, Gardner R, Neutzel S, Sharkis SJ (2001) Multi-organ, multilineage engraftment by a single bone marrow-derived stem cell. Cell 105:369–377
- Mallet VO, Mitchell C, Mezey E (2002) Bone marrow transplantation in mice leads to a minor population of hepatocytes that can be selectively amplified in vivo. Hepatology 35:799–804
- Wagers AJ, Sherwood RI, Christensen JL, Weissman IL (2002) Little evidence for developmental plasticity of adult hematopoietic stem cells. Science 297:2256–2259
- Stroncek DF, Clay ME, Smith J (1997) Composition of peripheral blood progenitor cell components collected from healthy donors. Transfusion 37:411–417
- Taniguchi H, Toyoshima T, Fukao K, Nakauchi H (1996) Presence of hematopoietic stem cells in the adult liver. Nat Med 2:198– 203
- Wang X, Montini E, Al-Dhalimy M (2002) Kinetics of liver repopulation after bone marrow transplantation. Am J Pathol 151:565–574
- Wang X, Willenbring H, Akkari Y (2003) Cell fusion is the principal source of bone-marrow-derived hepatocytes. Nature 422: 897–901
- Vassilopoulos G, Wang PR, Russell DW (2003) Transplanted bone marrow regenerates liver by cell fusion. Nature 422:901–904
- Avital I, Inderbitzin D, Aoki T (2001) Isolation, characterization, and transplantation of bone marrow-derived hepatocyte stem cells. Biochem Biophys Res Commun 288:156–164
- Blau HM, Brazelton TR, Weimann JM (2001) The evolving concept of a stem cell: entity or function? Cell 105:829–841
- Loeffler M, Roeder I (2002) Tissue stem cells: definition, plasticity, heterogeneity, self-organization and models—a conceptual approach. Cells Tissues Organs 171:8–26
- Ying QL, Nichols J, Evans EP, Smith AG (2002) Changing potency by spontaneous fusions. Nature 416:545–548
- Terada N, Hamazaki T, Oka M, Hoki M, Mastalerz DM, Nakano Y, Meyer EM, Morel L, Petersen BE, Scott EW (2002) Bone marrow cells adopt the phenotype of other cells by spontaneous cell fusion. Nature 416:542–545