SEQUENCE UPDATE

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## A chalcone synthase-like cDNA from rice anther

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Chalcone synthase (CHS) catalyses the first dedicated reaction in the flavanoid biosynthesis pathway, involving three rounds of condensation of malonyl-CoA onto 4-coumaroyl-CoA. Flavanoids are known to be involved in several important physiological processes in plants, such as flower pigmentation (Koes et al. 1990), UV stress protection (Wienand et al. 1986), nodulation (Recourt et el. 1992) and pollen development (Taylor and Jorgensen 1992; van der Meer et al. 1992). Chalcone synthase genes have been cloned from a number of species and are highly conserved among angiosperms and conifers (Fliegmann et al. 1992; Martin 1993).

To isolate genes expressed predominantly during anther development in rice (*Oryza sativa* var. *japonica*), a rice cDNA library (Lambda gt11, Promega) was constructed from mRNA prepared from anthers about 1.0 mm in length. Approximately 105 recombinant plaques were differentially screened with first-strand cDNA probes from developing anthers, leaves and roots, respectively. Ten clones were found to be anther probespecific (data not shown). One of these clones, designated D5, was selected for further analysis. Northern blot analysis of RNA from anthers, roots, stems, leaves and seedlings using the D5 cDNA as probe showed a hybrid-

The nucleotide sequence data reported will appear in the EMBL database under accession number X91811

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isation band of approximately 1.6 kb in the anthers but none in the other tissues (data not shown). Southern blot analysis indicated that the D5 cDNA hybridised to single fragments from rice genomic DNA digested with several restriction enzymes (data not shown), showing that the D5 cDNA was derived from a single copy gene or lowcopy number genes.

DNA sequence analysis revealed that the D5 cDNA is 1577 bp in length with a predicted open reading frame of 389 amino acid residues starting at the first ATG codon at position 53 and ending at position 1222 (Fig. 1). The 3′ end of the cDNA terminated at a poly(A) tail, preceded by a 278-bp untranslated AT-rich region containing a potential polyadenylation signal (AATACA) (Fig. 1). A database search using BLAST (Altschul et al. 1994) showed that the encoded polypeptide of the D5 cDNA is homologous to chalcone synthase sequences from plants with a minimal identity of 39% among 62 CHS sequences (including the sequence of the *Whp* gene, a maize CHS gene which plays a critical role in pollen viability) compared (data not shown). A short leucine zipper motif of four leucine residues has been found in most CHS and CHS-related proteins and is thought to play a role in protein-protein interaction (Kouzarides and Ziff 1988; Fliegmann et al. 1992). A similar motif has also been detected in the corresponding region of the predicted D5 polypeptide, although the second leucine was replaced by a phenylalanine through substitution of CTT with TTT (Fig. 1). Despite the overall similarity, the D5 polypeptide showed some interesting differences from other CHS sequences. For example, at the position 368–379 of the D5 polypeptide, a highly conserved region in other CHS proteins (WGVLFGFGPGLT), known as the CHS signature (Martin 1993), was replaced by the sequence WGLILAFGPGIT (Fig. 1). Furthermore, the D5 polypeptide diverges from another highly-conserved region around cysteine 170 thought to be an active site for substrate binding although the cysteine 170 itself is conserved (Martin 1993). These differences suggest that the D5 cDNA represents a novel member of the CHS gene family.

**Fig. 1** DNA and polypeptide sequences of the D5 gene. The start and stop codons together with the cysteine 170 and the residues forming a short leucine zipper motif are shown in bold. A possible polyadenylation signal sequence is underlined. Computer analyses of DNA and amino acids were performed using the GCG package

A GGA CTA GAA TAC TTG CTG ACT ACT GAG GCG AAA GGA TAC CGG GTG GCG AAG 52  $1R$ G **NT**  $\Omega$ GCT GAC CTT ATG GGA TTC GGC GAT  $_{\rm GCC}$ GGC AAT GGC AGC AGG AGC CAA AGG AGT 106  $\mathbf{s}$  $\overline{G}$  $\overline{\mathbf{k}}$  $\overline{G}$  $\mathcal{C}$  $\mathbf{K}$ M T. T. T. T.  $\mathbf{E}$ TGC TCC AGG GGG AAG GCG ATG CTG CTC GCC CTC GGC AAG GGC CTC CCT GAG CAA 160 Ÿ, Q 54 GTT CTT CCC CAG GAG AAG GTC GTC GAG ACC TAC CTC CAG GAC ACC ATC TGC AAC 214  $\frac{21}{72}$  $\Gamma$ K  $\overline{R}$  $\mathbf E$  $\overline{R}$ A L т R GAT CCT GCA ACA AGG GCA AAG CTG GAA AGA CTT TGC AAG ACC ACC ACA GTG AGG 268  $\mathbf T$  $\overline{R}$ v S  $\overline{K}$  $\overline{E}$  $\overline{D}$  $\overline{E}$  $\overline{P}$ Y M  $\mathbf{L}$  $H$ T L E L  $\mathbb{R}$ 90 T R Y T V M S K E L L D E H<br>ACA AGG TAC ACT GTC ATTG TAC ATTG TO T T L T P R L D T C<br>T E G T P T L T P R L D T C CCA GAG CTC AGG 322 N 108 Δ ACT GAG GGA ACT CCA ACA CTG ACG CCA CGG CTT GAC ATC TGC AAT GCT GCA GTG 376  $\mathbf F$ G  $\overline{A}$  $\mathbf{p}$ T. G E. M  $\mathbf{D}$  $126$ CTT GAG CTT GGT GCT ACT GCA GCC CGT GCC GCC CTT GGT GAA TGG GGG CGT CCA 430  $\overline{1}$  $\mathbf{H}$  $\overline{V}$  $\mathbf{v}$ S  $\mathbf{S}$  $\mathbf{E}$ 144 V  $\Gamma$  $\mathbf{L}$ S  $T_{\rm s}$ T. Þ GCA GTT GAC ATT ACC CAC CTT GTC TAC ATC TCG TCC AGT GAG CTT CGC CTC CCA 484 G  $\overline{H}$ 162 GGG GGT GAC CTT TTC CTG GCA ACT CGC CTT GGC CTC CAT CCA AAC ACC GTC CGC 538  $\overline{G}$  $\overline{c}$ S G G  $\overline{F}$ T 180 T S  $\overline{\mathbf{A}}$ A  $\overline{R}$ L L A  $\overline{A}$ ACT TCC CTT CTC TTC CTT GGC TGC  $K$  D I A E N N P TCC GGT GGC GCT GCC<br>G S R V L GCG TTC CGC ACC GCC V V A A E 592  $\frac{1}{G}$ 198 AAG GAC ATT GCT GAG AAC AAC CCA GGG AGC CGC GTC CTT GTA GTA GCC GCG GAG 646 216<br>700  $^{\mathsf{h}}$  $\mathcal{C}$ E  $\mathbb{R}$ D S P  $\overline{D}$  $\overline{R}$  $\mathbf{D}$  $\Gamma$ ACG ACG GTG CTG GGA TTC CGG CCA CCA AGT CCT GAC CGT CCT TAC GAT CTT GTT G A A L F G D G A S A A I I G A G P GGT GCC CTG TTT GGT GAC GGC GCA TCA GCT GCG ATC ATT GGA GCA GGC CCC  $234$ <br> $754$ S P  $F$ т.  $\mathbf{E}$ T.  $\circ$  $\overline{R}$  $252$ ATT GCT GCT GAG GAG AGT CCC TTC CTA GAG CTT CAG TTC TCA ACA CAG GAG TTC 808  $\overline{E}$  $\overline{v}$  $\overline{\mathbf{K}}$ Ġ K  $\overline{D}$ G CTA CCA GGG ACG GAC AAG GTA ATT GAT GGC AAG ATC ACT GAG GAA GGG ATT AAT 862  $\overline{D}$  $\overline{\mathbf{s}}$  $\overline{E}$  $\mathsf{C}$  $\overline{R}$  $\overline{E}$  $\mathbf E$ L K  $\overline{R}$ G 288 TTC AAA CTG GGG CGT GAT TTG CCC GAA AAG ATT GAA AGC CGT ATA GAA GGG TTC CCC RAT AG AGC ATA CAA AGC CGT ATA GAA GGG TTC 916 306 TGC AGG ACA CTC ATG GAT CGG GTT GGG ATA AAG GAG TTC AAT GAT GTA TTC TGG 970 N  $H$ F G G P T. N  $\mathbb{R}$ Τ. R.  $\overline{V}$ 324 GCT GTG CAT CCT GGT GGT CCA GCA ATA CTG AAC AGG CTA GAG GTT TGC TTT GAA 1024 LQREKLKISRKALMNYGN<br>CTC CAG CGA GAG AAG CTC AAG ATC AGT AGA AAG GCC CTG ATG ATAT GGT AAT<br>VRS NT VFY VLEYLRAELK  $342$ 1078 360  $\overline{R}$  $\overline{A}$ GTA CGC AGC AAC ACC GTC TTC TAT GTG TTG GAG TAT TTA AGG GCT GAG TTG AAG 1132 G W G 378 AAA GGG ATG ATA AGG GAA GAA TGG GGA CTG ATC TTG GCT TTT GGC CCA GGC ATC 1186  $\overline{F}$  $\bar{E}$  $\overline{\mathbf{V}}$  $\mathbb R$ 390  $\mathbf G$ M  $\mathbf L$  $\mathbf G$  $\mathbf N$ I ACA TTT GAA GGA ATG CTA GTT CGA GGC ATT AAC TGA GACTGAAGGGTCCAAGAAGACTT 1245 TCAGCTAGATGGAAGAATCACAAACATACCATTTCCAGGTTAAGTATATAGTATGACCTTGAACATCTCAT 1316  ${\tt TAGGAGCATAGTTGTCTTTACAGCATCTGTGCACTCCATAGTTATTTTTTACTGTTAATCTCTGTTTTATGT$ 1387  ${\tt TTCAAACAATGTGAGGTACCCTAGTTTGAATGCCAGATACTTGGGAAATTTCGGGC\underline{AATACA}TTGCATCC}$ 1458 TGTTACATTGTCATGTATAGTAATTTTCCAGAGTATCTATTT(A) $_{77}$ 1577



The D5 polypeptide showed highest homology (68% and 65% identity, respectively) to peptides encoded by two anther-specific genes in *Brassica napus,* A1 and BA42 (Scott et al. 1991; Shen and Hsu 1992). The A1 gene was found to be expressed during the tetrad and microspore release stages of pollen development (Scott et al. 1991), whereas BA42 transcripts were detected only in the tapetum, periphery cells of the vascular bundle and developing microspores (Shen and Hsu 1992). However, the spatial distribution of the D5 transcript within anthers needs further analysis. The fact that D5, A1 and BA42 share the highest homology suggests that there may be a subgroup of CHS genes which are expressed specifically in anthers; this is also supported by a dendrogram (Fig. 2) derived from a phylogenetic analysis using the PILEUP program in the GCG package (Madison, Wis.).

**Fig. 2** Dendrogram based on amino acid sequence similarity by using the PILEUP program. CHS\_Os, rice chalcone synthase (Reddy et al. 1996); CHS-Zm, chalcone synthase *Whp* in flowers of *Zea mays* (Franken et al. 1991); CHSA\_Ph and CHS\_Ph, two chalcone synthases in flowers of *Petunia hybrida* (Koes et al. 1989); CHS\_Am, chalcone synthase in flowers of *Antirrhinum*

*majus* (Sommer and Saedler 1986); CHS\_Ps, chalcone synthase in *Pinus sylvestris* (Fliegmann et al. 1992); A1\_Bn, peptide from *Brassica napus* anther-specific mRNA A1 (Scott et al. 1991); BA42\_Bn, peptide from *B. napus* anther-specific mRNA BA42 (Shen and Hsu 1992); D5\_Os, peptide from rice anther-specific mRNA D5 by the authors

Although the function of proteins encoded by this class of anther-expressed genes remains unclear, their homology to CHS and their expression patterns indicate that they play an important role in anther development. Further work is required to dissect their functions during anther development.

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