ORIGINAL ARTICLE



Nitrogen uptake and allocation in *Populus simonii* in different seasons supplied with isotopically labeled ammonium or nitrate

Chunxia Zhang^{1,2} · Sen Meng¹ · Yiming Li¹ · Li Su¹ · Zhong Zhao¹

Received: 18 January 2016/Accepted: 1 June 2016/Published online: 17 June 2016 © Springer-Verlag Berlin Heidelberg 2016

Abstract

Key message The total uptake of $^{15}NO_3$ -N was twofold higher than that of $^{15}NH_4$ -N when supplied with ammonium and/or nitrate in different seasons; the seedlings fertilized with NO₃-N had good growth with high photosynthetic rate and total biomass.

Abstract Appropriate fertilization is crucial for maximum plant growth and improving nitrogen use efficiency. Poplar is an important fast-growing tree species for biomass production, however, little is known about fertilizer management of poplar plantations growing on barren soil in different seasons. To understand nitrogen uptake and allocation of Populus simonii supplied with different forms of nitrogen in different seasons, we determined nitrogen uptake and allocation of P. simonii potted seedlings after a 4-day supply of ${}^{15}NH_4-N$, ${}^{15}NO_3-N$, ${}^{15}NH_4NO_3$, and NH¹⁵₄NO₃ in May, July, and September. The total ¹⁵N uptake was twofold higher when supplied with sole ¹⁵NO₃-N compared to sole ¹⁵NH₄-N in all the investigated seasons. In the presence of ammonium nitrate (¹⁵NH₄NO₃ and $NH_4^{15}NO_3$), the total ¹⁵N uptake was two times higher when supplied with NH₄¹⁵NO₃ compared to ¹⁵NH₄NO₃. Per unit biomass, the ¹⁵N-uptake ability of fine roots was higher in

Communicated by H. Rennenberg.

Zhong Zhao zhaozh@nwsuaf.edu.cn

¹ College of Forestry, Northwest A&F University, Yangling 712100, China

² State Key Laboratory of Soil Erosion and Dryland Farming on the Loess Plateau, Northwest A&F University, Yangling 712100, China May and July compared to that in September. ¹⁵N was present mainly in leaves in May and July, and was mainly stored in roots and stems in autumn. The effect of nitrogen on the growth of *P. simonii* seedlings was studied by fertilizing with NH₄-N, NO₃-N, and NH₄NO₃ for 8 weeks. The seedlings fertilized with NO₃-N had good growth with high photosynthetic rate and total biomass indicating that NO₃-N is crucial for *P. simonii* growth. These data contribute to understand the nitrogen uptake in different seasons in trees supplied with different forms of nitrogen. This provides important theoretical bases for fertilizer management of poplar plantations.

Keywords Nitrate · Ammonium · Nitrogen uptake and allocation · Seasonal variation · Growth characteristics · *Populus simonii*

Introduction

Nitrogen (N) is one of the main macronutrients for plant growth and development. Most crops grow on fertile soils, but forest trees are often planted on marginal lands where N availability is limited (Finzi et al. 2007). Thus, N fertilization is crucial to ensure high productivity and biomass of forest tree plantation. N is available in different chemical forms such as ammonium (NH₄-N), nitrate (NO₃-N), and organic N forms (McKane et al. 2002). Plants can assimilate both NH₄-N and NO₃-N, but some species perform better when fertilized with NH₄-N (Cruz et al. 1993; Britto and Kronzucker 2002; Bown et al. 2010; Metcalfe et al. 2011), while others show improved performance when grown with NO₃-N (Atkin and Cummins 1994) or a mixture of NO₃-N and NH₄-N (Öhlund and Näsholm 2001; Nicodemus 2007; Domenicano et al. 2011). The choice of

Chunxia Zhang czhang2012@163.com

nitrogen form for fertilization is very important for plant growth because plants will show best growth when supplied with their "favorite" nitrogen at appropriate times; therefore, appropriate fertilization plays a key role for maximizing plant growth and improving N use efficiency.

NH₄-N and NO₃-N have highly distinct consequences for plant growth (Haynes and Goh 1978; Britto and Kronzucker 2002; Domenicano et al. 2011). For example, NO₃-N treated hybrid poplar (*Populus maximowiczii* \times *P*. balsamifera) plants show higher ratios of fine roots: coarse roots and higher specific root lengths compared to NH₄-N treated poplar plants (Domenicano et al. 2011). Plants might be able to use different forms of nitrogen in different seasons because they have different growth characteristics in spring, summer, and autumn (McKane et al. 2002). In spring, buds and new leaves appear. Plants have rapid growth in summer while they grow slowly in autumn with decreasing temperature and short photoperiod. There is a lot of evidence that available nitrogen compounds vary in relative proportions in different seasons (Cooke and Weih 2005; Contosta et al. 2011; Brereton et al. 2013; Gilson et al. 2014). However, little information is available about NH₄-N or NO₃-N uptake of fine roots and allocation in different tissues fertilized with NH₄-N and/or NO₃-N in different seasons (Socci and Templer 2011; Brereton et al. 2013).

Poplar is an important fast-growing and high-yielding tree species with a short-rotation coppice for bioenergy production or ecological purpose (Schweier and Becker 2013; Manzone et al. 2014). Especially, Populus simonii Carr. is an important ecological and commercial breeding species in northern China; it is tolerant to drought, salinity, cold, and heat with wide distribution from Qinghai to the east coast of China in longitude and from the Heilongjiang River to the Yangtze River in latitude (Weisgerber and Han 2001; Wei et al. 2012). P. simonii is considered as an important afforestation species in China due to its large distribution range, excellent stress tolerance, rapid growth, and regeneration ability. P. simonii often exhibits poor growth with short height in the salty, or water-limited, or barren sandy soil, while P. simonii can reach a height of 30 m and a diameter of 2.5 m in the fertile and moist soil (Lu 2002). The soil in the afforestation areas, particularly the Loess Plateau, typically suffers from nutrient deficiencies, thus, fertilization is needed for afforestation species, such as P. simonii, to improve growth. However, P. simonii plantations lack management; little is known about fertilizer management in different seasons of poplar plantations growing on barren soil.

Populus simonii plants often grow on saline and alkali soil, where most available nitrogen is NO₃-N (Weisgerber and Han 2001). We hypothesized that *P. simonii* seedlings

would have higher N uptake when supplied with NO₃-N compared to NH₄-N irrespective of different N forms and seasons. To test this hypothesis, we investigated short-term nitrogen uptake dynamics in P. simonii supplied with different forms of nitrogen (NH₄-N or/and NO₃-N) in different seasons. The ¹⁵N isotope-labeled NH₄-N and NO₃-N were applied to 2-year-old P. simonii potted seedlings in three seasons: spring, summer, and autumn. Our specific objectives were to: (1) investigate N uptake of fine roots supplied with different forms of nitrogen, (2) examine the impact of season of application on N uptake, (3) understand the nitrogen allocation to different tissues after a short-time supply of nitrogen at different times in the growing season. Meanwhile, we carried out another independent relative long-term fertilizer experiment to test the effect of nitrogen form on growth of P. simonii seedlings. Based on a previous study (Rennenberg et al. 2010), we hypothesized that NO₃-N would be more important compared to NH₄-N for the growth of *P. simonii* seedlings. This study is valuable because it improves our understanding of nitrogen form uptake in different seasons. Meanwhile, the different effects of nitrogen forms on P. simonii growth provide an important theoretical base for fertilizer management of poplar plantations.

Materials and methods

Plant cultivation

Two-year-old seedlings of P. simonii were collected from Wuqi county, Shaanxi province (36°58'57"N. 108°09'34"E) in early April. Each seedling was decapitated to a stem height of about 10 cm with three to five buds and then planted in a 3 l pot filled with a 1:1 ratio of sand and vermiculite. All shoots but one were removed once buds had extended to about 5 cm. The cuttings were cultivated in a glasshouse in the condition of natural light, day/night 25/20 °C, 75 % relative humidity for 5 weeks. After that, the heat in the glasshouse was turned off, a ventilation device was used to exchange air in the greenhouse to the outside, and therefore the temperature was similar to outside before isotope treatment. We watered the cuttings every 3 days and irrigated with 250 ml modified Hoagland nutrient solution once a week (Hoagland and Arnon 1950) containing EDTA·FeNa 10 µM, MnSO₄·H₂O 5 µM, ZnSO4·7H2O 1 µM, CuSO4·5H2O 1 µM, H3BO3 30 µM, H₂MoO₄ 0.5 µM, KH₂PO₄ 1000 µM, MgSO₄·7H₂O 1000 µM, CaCl₂ 1000 µM, Na₂SO₄ 1000 µM, and NH₄. NO₃ 100 μ M; pH 6.5. To make this study more comparable to the field condition, we cultivated P. simonii seedlings with relatively low N concentration because P. simonii often grows on N-limited sites. To increase nitrogen

uptake, plants were irrigated with Hoagland nutrient solution free of nitrogen for 2 weeks before ¹⁵N treatment.

¹⁵N treatment and sampling

Plants were treated with 250 ml 99.09 atom% enriched to 0.1 mM¹⁵N isotope-labeled ammonium (¹⁵NH₄Cl-N), 99.14 atom% enriched to 0.1 mM ¹⁵N isotope-labeled nitrate (K¹⁵NO₃-N), and ¹⁵N isotope-labeled NH₄NO₃ (either ¹⁵NH₄NO₃ or NH₄¹⁵NO₃) in three seasons: spring (28 May), summer (28 July), and autumn (28 Sep). Each treatment was applied to four seedlings. The total biomass of plants in May, July, and September was 3.03 ± 0.25 , 5.31 ± 0.28 , and 7.21 ± 0.44 g, respectively. Four seedlings served as controls. They did not receive any ¹⁵Nlabeled nitrogen and thus were used to determine the natural abundance of ¹⁵N signature of plants (Templer and Dawson 2004). Plants were harvested 4 days following ¹⁵N addition and partitioned into fine roots (<2 mm in diameter), roots (coarse root and main root), stems, and leaves. The experimental period was limited to 4 days, which was sufficient to detect enriched ¹⁵N in different tissues of plants. The fine roots, roots, stems, and leaves of each seedling were dried at 65 °C for at least 48 h until the dry weight was unchanged, and then ground into powder for further analysis. The total nitrogen (N%) and ¹⁵N abundance were determined using a Thermo Electron Flash EA 1112 elemental analyzer coupled to a Thermo Electron Delta V isotope ratio mass spectrometer (EA-IRMS). The ¹⁵N abundance was expressed as a delta value (δ^{15} N, per mil, ‰) relative to atmospheric N₂: $\delta^{15}N = [(R_{sample} R_{\text{standard}}/R_{\text{standard}} \ge 1000$, where $R = {}^{15}\text{N}/{}^{14}\text{N}$ and R_{standard} d_{ard} is atmospheric N₂, and atom% of ¹⁵N is equal to $R \times 100/(1 + R)$. Reproducibility of the δ^{15} N measurements was better than 0.2 %.

We calculated the ¹⁵N content of different tissues including fine root, root, stem, and leaf as: the total ¹⁵N content = DW_{tissue} × N%_{tissue} × (atom%_{tissue} – atom%_{control}). The total ¹⁵N uptake of the plant was equal to the total ¹⁵N content of all tissues.

The ¹⁵N uptake of fine roots was calculated as the total ¹⁵N uptake of plant/dry weight of fine root reflecting the N-uptake ability of fine root per biomass.

The effect of nitrogen form on *P. simonii* plant growth

To know the effect of different nitrogen forms (NH₄-N, NH₄NO₃, NO₃-N) on plant growth, we carried out another independent experiment with 2-year-old *P. simonii* seedlings fertilized with NH₄Cl, or NH₄NO₃, or KNO₃ for 8 weeks. The *P. simonii* cuttings were also planted in the same

glasshouse described above for 5 weeks and irrigated with 250 ml modified Hoagland nutrient solution once a week (Hoagland and Arnon 1950). The fertilized experiment was started on May 28 with similar height of P. simonii seedlings described above and the temperature was not controlled manually. There was a ventilation device to exchange air in the greenhouse to the outside; therefore, the temperature in the greenhouse was close to outside. To maximize the effect of nitrogen form on plant growth, we added a high concentration of nitrogen (250 ml modified Hoagland nutrient solution including 5 mM NH₄-N, NH₄NO₃, or NO₃-N) once a week in the fertilizer experiment compared to that described above. Each fertilizer treatment was applied to ten plants. Near the harvest date, the net photosynthetic rate was measured from 9:00 to 11:00 hours with a portable photosynthesis system (Li-Cor-6400; Li-Cor, Inc, Lincoln, NE) with an attached LED light source (500 µmol photon $m^{-2} s^{-1}$). The CO₂ concentration in the chambers was 400 μ mol min⁻¹, and the air flow was 500 μ mol s⁻¹. The chlorophyll content of each plant chosen for gas exchange was measured with a portable meter (Minolta SPAD 502 Meter). Simultaneously, the height of the main shoot of each plant was measured with a ruler. Plants were harvested 8 weeks after fertilization with different forms of nitrogen and partitioned into above-ground and below-ground parts. The above-ground and below-ground samples were dried at 65 °C to constant mass, and then dry weight was recorded.

Statistical analyses

N form and time effect on N uptake were analyzed by twoway analyses of variance (ANOVA). The effect of N form on the plant growth was assessed by one-way ANOVA to test the significant differences between treatments and plant parts. When significant, Fisher's least significant difference test (LSD) was used to identify differences between treatment means. The significance level was established at p < 0.05.

Results

Nitrogen uptake of P. simonii in different seasons

To understand the total N uptake by the whole *P. simonii* plants within 4 days, we determined the total ¹⁵N uptake by summing the ¹⁵N content of all the tissues. *P. simonii* plants had different total ¹⁵N uptake when fertilized with different forms of nitrogen across the three investigated seasons (Fig. 1). The total ¹⁵N uptake of plants was significantly affected by nitrogen forms (¹⁵NO₃-N or ¹⁵NH₄-N) and the seasons (p < 0.05). The total N supplied by ¹⁵NO₃-N or ¹⁵NH₄-N was almost the same (0.375 mg/plant), yet

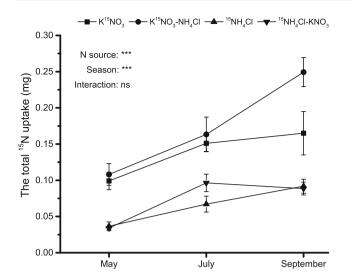


Fig. 1 The total ¹⁵N uptake of *P. simonii* in three seasons treated with different ¹⁵N forms. The data indicate the mean \pm SD (n = 4). ***Significance at p < 0.001, *ns* no significance

different total ¹⁵N uptake was observed in May, July, and September. The total uptake of ¹⁵N when supplied with sole ¹⁵NO₃-N was two times higher than that of sole ¹⁵NH₄-N in all three investigated seasons. In the ammonium nitrate treatments, the total ¹⁵N uptake was also two times higher when supplied with $NH_4^{15}NO_3$ compared to ¹⁵NH₄NO₃. Of the different nitrogen forms, the total ¹⁵N uptake was the highest when supplied with $NH_4^{15}NO_3$. It was the lowest when supplied with sole NH_4^+ . Across the three investigated seasons, the total ¹⁵N uptake was highest in September and lowest in May.

Fine root is the most important organ for nitrogen acquisition in plants; therefore, we investigated N-uptake ability of fine roots defined as the total ¹⁵N uptake of plant/dry weight of fine root. ¹⁵N uptake of fine roots was significantly affected by the nitrogen forms and season of application (Fig. 2, p < 0.05). Per unit biomass, the ¹⁵N-uptake ability of fine roots was higher in May and July compared to that in September supplied with a given nitrogen form (Fig. 2). Of the different nitrogen forms, the fine roots had the highest ¹⁵N uptake when supplied with ¹⁵NO₃-N and the lowest ¹⁵N uptake when supplied with ¹⁵NH₄-N. Per unit fine root biomass, the uptake of ¹⁵N was significantly greater when supplied with NO₃-N compared to ¹⁵NH₄-N in *P. simonii* in the three investigated seasons, especially in May.

N allocation to different tissues

The relative ¹⁵N distribution in different tissues is defined as ¹⁵N content of a given tissue relative to the total uptake of ¹⁵N by *P. simonii* plant in this study. The relative ¹⁵N distribution varied in different tissues from May to

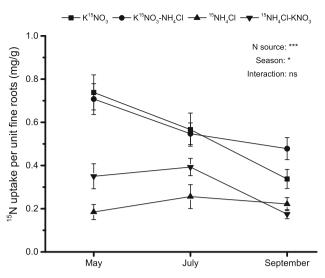


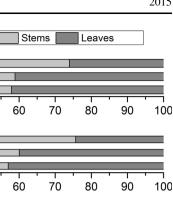
Fig. 2 ¹⁵N uptake per unit fine roots in *P. simonii* in three seasons treated with different ¹⁵N forms. The data indicate the mean \pm SD (n = 4). ***Significance at p < 0.001, *significance at p < 0.05, ns no significance

September when supplied with ammonium and/or nitrate (Fig. 3). In May and July, most of the supplied ¹⁵N was distributed in leaves, and the percent of ¹⁵N was significantly influenced by the nitrogen forms (p < 0.01). In September, the proportion of ¹⁵N allocation increased in stems or roots, while it largely decreased in leaves irrespective of the supplied nitrogen form suggesting that nitrogen assimilation was slowed in autumn.

The ¹⁵N content of different tissues was significantly affected by N form (Fig. 4, p < 0.05). Generally, different tissues of plants (fine roots, main roots, stems, and leaves) had a higher ¹⁵N content when supplied with ¹⁵NO₃-N or $NH_4^{15}NO_3$ compared to ${}^{15}NH_4-N$ or ${}^{15}NH_4NO_3$ (Fig. 4). The supply of extra non-labeled NH₄-N (NH₄¹⁵NO₃) increased the ¹⁵N content compared to sole nitrate (¹⁵NO₃-N) in all tissues except for the main roots. Leaves generally had the largest total ¹⁵N content followed by fine roots and stems. The main roots had the lowest N content. The ¹⁵N content of plants supplied with ¹⁵NO₃-N or ¹⁵NH₄-N varied from May to September in different tissues including fine roots, main roots, stems, and leaves (Fig. 4). All tissues generally had the lowest ¹⁵N content in May among the three investigated seasons (May, July, and September) (Fig. 4). ¹⁵N content of leaves decreased significantly from July to September in sole N-source treatments indicating that the translocation of N slowed down in September.

The percent of total N was also significantly affected by nitrogen forms in different tissues after a 4-day supply of different forms of nitrogen (Fig. 5). Of different nitrogen forms, nitrate could increase the total N concentration of the roots and stems compared to the other nitrogen forms (NH₄Cl or NH₄NO₃) in a given season. Leaves could have

Fig. 3 Relative ¹⁵N distribution in different tissues in three seasons (fine roots, main roots, stems, and leaves). The data indicate the mean of four replicates



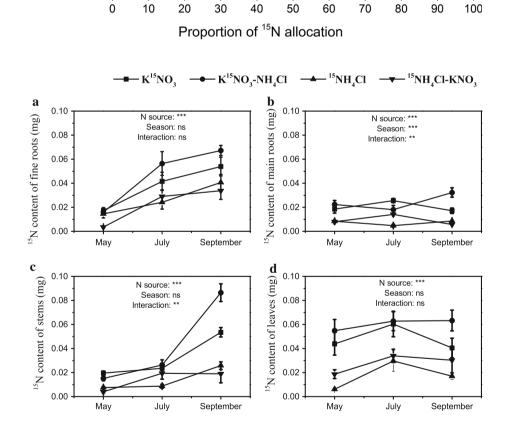
80

90

100

70

Fig. 4 The total ¹⁵N content of different tissues in three seasons (a fine roots, b main roots, c stems, d leaves) after a 4-day supply of ¹⁵N-labeled ammonium or nitrate. The data indicate the mean \pm SD (n = 4). ***Significance at p < 0.001, **significance at p < 0.01, *ns* no significance



Fine roots

20

20

20

30

30

30

Sep

July

May

Sep July

May

Sep

July May

Sep

July

May

n

0

0

10

10

10

K¹⁵NO₃

K¹⁵NO₃-

NH₄CI

¹⁵NH₄CI

KNO₂-

¹⁵NH₄CI

Main Roots

50

50

50

60

40

40

40

more total N concentration when supplied with sole ammonium compared to the other nitrogen forms (KNO3 or NH_4NO_3). The total N concentration varied in different tissues in a given season. Generally, leaves had high total N concentration followed by fine roots and stems. The main roots had the lowest total N concentration. Across different seasons, the total N concentration in most of the tissues increased from May to September which was consistent with the total ¹⁵N uptake indicating that plants could accumulate more nitrogen as they increased in size.

The effect of nitrogen forms on the growth of Populus simonii

The P. simonii seedlings showed slightly different growth characteristics when supplied with different forms of Fig. 5 The total N concentration of different tissues in three seasons (a fine roots, b main roots, c stems, d leaves) after a 4-day supply of ¹⁵N-labeled ammonium or nitrate. The data indicate the mean \pm SD (n = 4). ***Significance at p < 0.001, *significance at p < 0.05, *ns* no significance

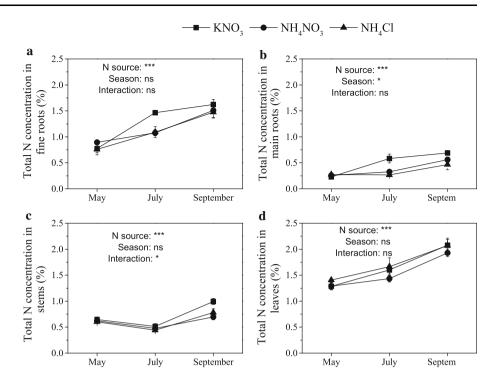


Table 1Height increment andphotosynthesis of *P. simonii*seedlings fertilized withdifferent forms of nitrogen for8 weeks

N form	Height increment (cm)	Chlorophyll content (SPAD)	Net photosynthetic rate (mmol $CO_2 m^{-2} s^{-1}$)
NO ₃ -N	$40.38 \pm 3.29a$	$36.71 \pm 3.58a$	$15.36 \pm 1.56a$
NH ₄ -N	$43.54 \pm 3.36a$	$32.62\pm3.56a$	$12.37 \pm 1.58b$
NH ₄ NO ₃	$44.99 \pm 3.88a$	$36.95\pm4.13a$	$14.64 \pm 1.43a$

Data indicate mean \pm SD (n = 10). The same letter in a column indicates that the difference is not statistically significant, while different letters in the same column indicate significant difference at p < 0.05

Table 2 Biomass (dry weight)of *P. simonii* seedlings fertilizedwith different forms of nitrogen

N form	Total biomass (g)	Aboveground (g)	Belowground (g)	Belowground/aboveground
NO ₃ –N	$9.01 \pm 0.64a$	$5.85\pm0.62a$	$3.66 \pm 0.41a$	$0.63\pm0.05a$
NH ₄ –N	$6.82\pm0.76\mathrm{b}$	$3.89\pm0.54b$	$2.99\pm0.29a$	$0.77\pm0.09\mathrm{b}$
NH ₄ NO ₃	$8.65\pm0.69ab$	$4.97\pm0.43a$	$3.58\pm0.31a$	$0.72\pm0.08ab$

Data indicate mean \pm SD (n = 10). The same letter in a column indicates that the difference is not statistically significant, while different letters in the same column indicate significant difference at p < 0.05

nitrogen (NH₄-N, NO₃-N, and NH₄NO₃). The height increment and chlorophyll content were not significantly affected by the supplied nitrogen forms (Table 1). Plants supplied with NO₃-N or NH₄NO₃ had significantly higher net photosynthetic rate than those supplied with NH₄-N.

NH₄-N and NO₃-N also had different effects on the biomass of *P. simonii* seedlings. The *P. simonii* seedlings fertilized with NO₃-N had significantly higher total biomass and aboveground biomass (dry weight) compared to those fertilized with NH₄-N (Table 2). No significant difference was observed in the biomass of *P. simonii* seedlings supplied with NO₃-N and NH₄NO₃. Of the different

nitrogen forms, NH_4NO_3 could slightly enhance the belowground biomass of plants. The ratio of belowground biomass to aboveground biomass in *P. simonii* seedlings was the highest when supplied with NH_4 -N.

Discussion

N is one of the main macronutrients for plant growth, appropriate N addition is crucial to maximize the plant growth. Excess N fertilization is both an unnecessary cost to growers and a pollution risk to groundwater, while N deficiency often results in poor growth. Thus, appropriate N fertilization is necessary to improve plant growth and avoid N pollution in the soil. N is available in different forms, such as NH_4^+ , NO_3^- , and organic N, and plants often show maximum growth with their preferred N form. It is crucial to determine the effect of different nitrogen forms on plant growth for a given species. Meanwhile, plants may need different forms of nitrogen in different seasons due to variable growth characteristics (Brockley 1995). It is therefore necessary for forest managers to understand nitrogen uptake of forest trees supplied with different forms of nitrogen in different seasons (Lea and Azevedo 2006).

N uptake of *P. simonii* with different N forms in different seasons

Fast-growing tree species such as Populus spp. are thought to be adapted to a high N supply, mainly NO_3^{-} , in the moving water table of the floodplain forests they inhabit (Rewald et al. 2014); Rennenberg et al. (2010) proposed that nitrate is much more important for nutrition of poplar species than for many other tree species. In the present study, the studied species P. simonii often grows on barren and N-deficient soils, thus we supplied low N content to make this study more close to the field conditions. Actually, under low nutrient supply net uptake of NO₃⁻ has been shown to be higher in Populus tremuloides than in Pinus contorta or Pseudotsuga menziesii (Min et al. 1999). In the present study, higher 15 N uptake was observed in *P*. simonii plants when supplied with ¹⁵NO₃-N compared to ¹⁵NH₄-N in different seasons. This result supported our initial hypothesis that P. simonii seedlings had higher N uptake when supplied with NO₃-N compared to NH₄-N irrespective of seasons. This might be due to higher concentration of nitrate when supplied with ammonium nitrate compared to sole ammonium/nitrate. A previous study reported that the nitrate reductase enzyme was active in P. tremuloides roots (Min et al. 1998). Additionally, nitrate is much more mobile in soil than ammonium, so the greater ¹⁵N uptake of plant supplied with NO₃-N could be due to its greater mobility (Vitousek et al. 1982). The high NO₃-N uptake observed in this study may be an adaptation of P. simonii to its native habitat, which is on saline and alkali soil, where, the most available nitrogen is NO₃-N (Weisgerber and Han 2001).

The different amounts of nitrogen taken up in spring, summer, and autumn suggested that plants might require different amounts of nitrogen during different seasons. The different amounts of N taken up in different seasons could relate to the size of the root system. As the plants grew, their root systems grew, and therefore, could take up more nitrogen. The amount of N might be affected by the biomass of *P. simonii* seedlings which were different in spring, summer, and autumn. The uptake of ¹⁵N increased with increasing plant biomass from May to September which suggests that the requirement of nitrogen increased as the plants increased in size (Li et al. 2009). The *P. simonii* seedlings still require nitrogen to maintain growth in autumn. Therefore, the seasonal rates of N addition should be considered to avoid N deficiency or N pollution in different seasons.

N allocation of *P. simonii* with different N forms during different seasons

For trees, N is variable in roots, stems, and leaves in different seasons (Weatherall et al. 2006; Brereton et al. 2013). In summer, most of the N was distributed in leaves because leaves constitute the dominant N sink during the summer. However, in autumn, N concentration is reduced in leaves and increased in roots and stems. This suggests that roots or stems are the main storage organs for N in autumn. High N in stems by autumn was also observed in willow (Brereton et al. 2013). Different N allocation in different tissues reflected N recycling across different seasons (Dohleman et al. 2012). This suggests that the stem is the major N storing tissue during winter (von Fircks et al. 2001).

The effect of nitrogen form on growth of P. simonii

In the present study, we studied the effect of inorganic N, NH₄-N, and/or NO₃-N, on P. simonii plant growth. P. simonii seedlings had a higher total biomass and aboveground biomass (dry weight) fertilized with NO3-N or NH₄NO₃ compared to NH₄-N (Table 2). The results of this study support our initial hypothesis that NO₃-N would be more important compared to NH₄-N for the growth of P. simonii seedlings. This agreed with Rennenberg et al. (2010) who stated that most poplar species prefer NO₃-N. It is also consistent with a previous report that 50 % NO₃-N fertilization could improve biomass productivity of poplar and willow clones (Domenicano et al. 2011). Similarly, N fertilization with 80 % NO3-N for Populus deltoides used for short-rotation coppice optimized whole-plant growth (Woolfolk and Friend 2003). Nitrogen forms also affect poplar root structure (Domenicano et al. 2011), and hence, the root biomass might also be affected by the nitrogen form.

In conclusion, NO₃-N is crucial for *P. simonii* plant growth. The amount of nitrogen required for *P. simonii* plants varied in different seasons; thus, the dose of nitrogen should be carefully considered for fertilization management in different seasons.

Author contribution statement Conceived and designed the experiments: Chunxia Zhang, and Zhong Zhao. Performed the

experiments: Chunxia Zhang, Sen Meng, Yiming Li, and Li Su. Analyzed the data: Chunxia Zhang, and Sen Meng. Wrote the paper: Chunxia Zhang, and Zhong Zhao.

Acknowledgments This research was sponsored by the State Key Basic Research Development Program (973 Program, Grant No. 2012CB416902), the National Natural Science Foundation of China (Grant No. 31500543), and the Fundamental Research Funds for the Central Universities of China (Grant No. Z109021564).

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

References

- Atkin OK, Cummins WR (1994) The effect of nitrogen source on growth, nitrogen economy and respiration of two high arctic plant species differing in relative growth rate. Funct Ecol 8:389–399
- Bown H, Watt M, Clinton P, Mason E (2010) Influence of ammonium and nitrate supply on growth, dry matter partitioning, N uptake and photosynthetic capacity of *Pinus radiata* seedlings. Trees 24:1097–1107
- Brereton NJ, Pitre FE, Shield I, Hanley SJ, Ray MJ, Murphy RJ, Karp A (2013) Insights into nitrogen allocation and recycling from nitrogen elemental analysis and ¹⁵N isotope labelling in 14 genotypes of willow. Tree Physiol 34:1252–1262
- Britto DT, Kronzucker HJ (2002) NH₄⁺ toxicity in higher plants: a critical review. J Plant Physiol 159:567–584
- Brockley RP (1995) Effects of nitrogen source and season of application on the nutrition and growth of iodgepole pine. Can J For Res 25:516–526
- Contosta AR, Frey SD, Cooper AB (2011) Seasonal dynamics of soil respiration and N mineralization in chronically warmed and fertilized soils. Ecosphere 2:art36
- Cooke JEK, Weih M (2005) Nitrogen storage and seasonal nitrogen cycling in Populus: bridging molecular physiology and ecophysiology. New Phytol 167:19–30
- Cruz C, Lips SH, Martins-Loução MA (1993) Interactions between nitrate and ammonium during uptake by carob seedlings and the effect of the form of earlier nitrogen nutrition. Physiol Plant 89:544–551
- Dohleman FG, Heaton EA, Arundale RA, Long SP (2012) Seasonal dynamics of above- and below-ground biomass and nitrogen partitioning in *Miscanthus* × giganteus and Panicum virgatum across three growing seasons. GCB Bioenergy 4:534–544
- Domenicano S, Coll L, Messier C, Berninger F (2011) Nitrogen forms affect root structure and water uptake in the hybrid poplar. New Forest 42:347–362
- Finzi AC et al (2007) Increases in nitrogen uptake rather than nitrogen-use efficiency support higher rates of temperate forest productivity under elevated CO₂. Proc Natl Acad Sci USA 104:14014–14019
- Gilson A, Barthes L, Delpierre N, Dufrêne É, Fresneau C, Bazot S (2014) Seasonal changes in carbon and nitrogen compound concentrations in a *Quercus petraea* chronosequence. Tree Physiol 34:716–729
- Haynes RJ, Goh KM (1978) Ammonium and nitrate nutrition of plants. Biol Rev 53:465–510
- Hoagland DR and Arnon DI (1950) The water-culture method for growing plants without soil. Circular California Agricultural Experiment Station 347

- Lea PJ, Azevedo RA (2006) Nitrogen use efficiency. 1. Uptake of nitrogen from the soil. Ann Appl Biol 149:243–247
- Li C, Wu C, Duan B, Korpelainen H, Luukkanen O (2009) Agerelated nutrient content and carbon isotope composition in the leaves and branches of *Quercus aquifolioides* along an altitudinal gradient. Trees 23:1109–1121
- Lu W (2002) *Populus simonii* Carr. in North China. Ningxia People's Press, Yinchuan
- Manzone M, Bergante S, Facciotto G (2014) Energy and economic evaluation of a poplar plantation for woodchips production in Italy. Biomass Bioenergy 60:164–170
- McKane RB et al (2002) Resource-based niches provide a basis for plant species diversity and dominance in arctic tundra. Nature 415:68–71
- Metcalfe RJ, Nault J, Hawkins BJ (2011) Adaptations to nitrogen form: comparing inorganic nitrogen and amino acid availability and uptake by four temperate forest plants. Can J For Res 41:1626–1637
- Min X, Siddiqi MY, Guy RD, Glass ADM, Kronzucker HJ (1998) Induction of nitrate uptake and nitrate reductase activity in trembling aspen and lodgepole pine. Plant Cell Environ 21:1039–1046
- Min X, Yaeesh Siddiqi M, Guy RD, Glass ADM, Kronzucker HJ (1999) A comparative study of fluxes and compartmentation of nitrate and ammonium in early-successional tree species. Plant Cell Environ 22:821–830
- Nicodemus MA (2007) Growth, nutrition, and photosynthetic response of black walnut to varying nitrogen sources and rates. J Plant Nutr 31:1917–1936
- Öhlund J, Näsholm T (2001) Growth of conifer seedlings on organic and inorganic nitrogen sources. Tree Physiol 21:1319–1326
- Rennenberg H, Wildhagen H, Ehlting B (2010) Nitrogen nutrition of poplar trees. Plant Biol 12:275–291
- Rewald B, Kunze ME, Godbold DL (2014) NH4:NO3 nutrition influence on biomass productivity and root respiration of poplar and willow clones. GCB Bioenergy 8:51–58
- Schweier J, Becker G (2013) Economics of poplar short rotation coppice plantations on marginal land in Germany. Biomass Bioenergy 59:494–502
- Socci AM, Templer PH (2011) Temporal patterns of inorganic nitrogen uptake by mature sugar maple (*Acer saccharum* Marsh.) and red spruce (*Picea rubens* Sarg.) trees using two common approaches. Plant Ecol Divers 4:141–152
- Templer PH, Dawson TE (2004) Nitrogen uptake by four tree species of the Catskill Mountains, New York: implications for forest N dynamics. Plant Soil 262:251–261
- Vitousek PM, Gosz JR, Grier CC, Melillo JM, Reiners WA (1982) A comparative analysis of potential nitrification and nitrate mobility in forest ecosystems. Ecol Monogr 52:155–177
- von Fircks Y, Ericsson T, Sennerby-Forsse L (2001) Seasonal variation of macronutrients in leaves, stems and roots of *Salix dasyclados* Wimm. grown at two nutrient levels. Biomass Bioenergy 21:321–334
- Weatherall A, Proe MF, Craig J, Cameron AD, Midwood AJ (2006) Internal cycling of nitrogen, potassium and magnesium in young Sitka spruce. Tree Physiol 26:673–680
- Wei Z, Du Q, Zhang J, Li B, Zhang D (2012) Genetic diversity and population structure in Chinese indigenous poplar (*Populus simonii*) populations using microsatellite markers. Plant Mol Biol Rep 31:620–632
- Weisgerber H, Han Y (2001) Diversity and breeding potential of poplar species in China. For Chron 77:227–237
- Woolfolk WTM, Friend AL (2003) Growth response of cottonwood roots to varied NH₄:NO₃ ratios in enriched patches. Tree Physiol 23:427–432