

Responses of symbiotic N₂ fixation in *Alnus* species to the projected elevated CO₂ environment

Hiroyuki Tobita¹ · Kenichi Yazaki¹ · Hisanori Harayama² · Mitsutoshi Kitao¹

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Abstract

Key message Nitrogen fixation in *Alnus* species in response to elevated CO₂ may depend on the presence of non-N₂-fixing tree species in addition to soil conditions.

Abstract *Alnus* is a major genus of actinorhizal plants. Symbiosis with *Frankia* allows the *Alnus* species to fix nitrogen (N) at the rate of several to 320 kg N ha⁻¹ year⁻¹ with a nodule biomass of 16–480 kg ha⁻¹. *Alnus* species ensures an effective supply of N to soils because of the high N content of leaf litter, rapid decomposition rate, and the influx of herbivorous insects. In addition, the association between regenerated endozoochorous species and *Alnus hirsuta* suggests that N₂ fixation in *Alnus* species influences the distribution patterns of regenerated plants as well as improve soil fertility. N₂ fixation by the *Alnus*–*Frankia* symbiotic relationship may be positively associated with elevated carbon dioxide (CO₂) levels. Nodule biomass increased under elevated CO₂ due to enhanced plant growth, rather than changes in biomass allocation. The inhibitory effect of high soil N on nodulation was retained under elevated CO₂, and the effects of elevated CO₂ on N₂ fixation depended on soil P availability, drought, and many other abiotic and biotic factors. Recent free-air CO₂ enrichment experiments have demonstrated

increased N₂ fixation in *A. glutinosa* exposed to elevated CO₂ in mixed-species stands containing non-N₂-fixers but not in monocultures, suggesting that N₂ fixation depends on an association with non-N₂-fixing tree species. Because elevated CO₂ can alter the N and P contents and stoichiometry of plants, it will be necessary to evaluate N allocation and accumulation of biomass when investigating the response of *Alnus* species to future global climate change.

Keywords Actinorhizal plants · *Frankia* · Nodule biomass · Soil nutrients · Stoichiometry

Introduction

Biological nitrogen (N) fixation is an important pathway to input new N into terrestrial ecosystems (Vitousek and Walker 1987). Two types of root-nodule symbioses exist between higher plants and N₂-fixing soil bacteria, such as legume–*Rhizobium* and actinorhizal plant–*Frankia* symbioses (Pawlowski and Sprent 2008). Bacteria induce the formation of nodules on plant roots during both interactions. In the case of actinorhizal symbiosis, Gram-positive actinomycetous soil bacteria in the genus *Frankia* induce the formation of root nodules in >200 dicotyledonous plants from eight families (Dawson 2008; Gtari et al. 2013). All of these host plants are perennial dicotyledons, and all except the genus *Datisca* are trees or shrubs. Although leguminous tree species are dominant symbiotic N₂-fixers in tropical regions, actinorhizal woody plants and *Frankia* play an important role in the N cycle in temperate and boreal forest ecosystems (Huss-Danell 1997). Recent research on actinorhizal plant–*Frankia* symbioses have focused on phylogenetic and genomic analyses regarding

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✉ Hiroyuki Tobita
tobi@ffpri.affrc.go.jp

¹ Department of Plant Ecology, Forestry and Forest Products Research Institute (FFPRI), Tsukuba 305-8687, Japan

² Hokkaido Research Center, Forestry and Forest Products Research Institute (FFPRI), Sapporo 062-8516, Japan

the use of transgenic actinorhizal plants (Kucho et al. 2010; Normand 2013).

Among actinorhizal plants, which are very diverse, those from the genus *Alnus* are of particular interest (Pourhassan et al. 2015). Forty-seven *Alnus* species are found worldwide (Pawlowski and Newton 2008), 16 of which grow naturally in Japan (Uemura and Sato 1975). The *Alnus*–*Frankia* symbiotic relationship has been used to revegetate deteriorated wildlife habitats and rehabilitate N₂-deficient disturbed areas (Sharma et al. 2002; Hanley et al. 2006) to enhance the growth of commercial coniferous trees (Vogel and Gower 1998; Son et al. 2007) and short-rotation plantings used for biomass energy (Eriksson and Johansson 2006; Claessens et al. 2010; Uri et al. 2011; Hytönen and Saarsalmi 2015). The range of N₂ fixation rates reported for the *Alnus*–*Frankia* symbiosis coincides with that of the legume–*Rhizobium* symbiosis (Hibbs and Cromack 1990; Lambers et al. 2008; Noh et al. 2010).

In addition, the *Alnus*–*Frankia* symbiotic relationship is attracting interest due to its responses to elevated atmospheric carbon dioxide (CO₂) concentrations (Hungate et al. 2003). Increasing CO₂ is an important factor influencing global climate change (IPCC 2007, 2013). Elevated CO₂ increases long-term forest net primary productivity (Zak et al. 2011), except under nutrient-limiting conditions (Leuzinger and Hättenschwiler 2013), and forest ecosystems are usually N₂ limited (Vitousek and Howarth 1991; Wang et al. 2010). Although carbon (C) becomes more available to plants in an elevated CO₂ environment, plants also require other resources to sustain primary production, including N, phosphorus (P), and micronutrients obtained from the soil (Pourhassan et al. 2015). Therefore, nutrient availability, particularly that of soil N, may strongly limit responses to elevated CO₂ by woody plants undergoing increased photosynthetic and growth rates (Luo et al. 2004; Hyvönen et al. 2007; Norby et al. 2010; Zak et al. 2011; Sigurdsson et al. 2013). Exogenous N input into forest ecosystems may be required to maintain enhanced growth under these conditions (Johnson 2006). Symbiotic and/or heterotrophic N₂ fixation by legumes and actinorhizal plants may be a potential source of N to sustain increased N uptake by non-N₂ fixers due to high rates of forest productivity under an elevated CO₂ environment (Vitousek et al. 2002; Finzi et al. 2007). Many studies on legumes have been conducted, and excellent reviews have been published (Ainsworth and Long 2004; Ainsworth and Rogers 2007; Rogers et al. 2009).

A number of abiotic and biotic factors affect the free-living and symbiotic properties of the *Frankia*–actinorhizal symbiosis, including moisture, aeration, temperature, pH, organic matter, inorganic chemicals, and the distributions of *Frankia*, and mycorrhizal fungi; excellent reviews have summarized these effects (Huss-Danell 1997; Dawson

2008; Valdés 2008; Pölme et al. 2014). The growth responses of legumes to elevated CO₂ are occasionally constrained by factors other than N availability, such as P availability and water conditions (Hungate et al. 2004; Reverchon et al. 2012). Understanding how *Alnus* species respond to elevated CO₂ is of great ecological and economical importance (Pourhassan et al. 2015). This review primarily focuses on the details of the interactive effects between projected elevated CO₂ in the near future and other factors such as N, P, and drought on growth and N₂ fixation in *Alnus*–*Frankia* symbiosis. The first section presents a review on determining nodule biomass and N₂ fixation rates under the present conditions.

Current *Alnus*–*Frankia* N₂ fixation ability

Nodule biomass in *Alnus* stands

Nodule biomass is an essential tool to estimate N₂ fixation at any scale (Aosaar et al. 2013). Nodules are formed on actinorhizal plants when lateral roots are infected by *Frankia*. In some hosts such as *Alnus* species, *Frankia* infects roots via root hairs (intracellular infection) or by intercellular penetration (Wall and Berry 2008; Tromas et al. 2013). Nodules can be perennial clusters of modified lateral roots and may grow to a large size (Huss-Danell 1997). The size and mean weight of *Alnus* species nodules increase with the tree diameter within a naturally established stand (Uliassi and Ruess 2002; Tobita et al. 2010a) and increase with stand age in a plantation (Aosaar et al. 2013). The nodule size affects nitrogenase activity (NA) because nodules have varying amounts of non-N₂-fixing tissue with increasing size (Sharma and Ambasht 1984; Hurd et al. 2001). This fundamental information about nodule size distribution is useful to estimate N₂ fixation in *Alnus* stands. However, NA, at least in young *Alnus* species plants, is related to nodule biomass (Gordon and Wheeler 1978) and not nodule number (Dawson and Gordon 1979).

Several studies have attempted to estimate nodule biomass in *Alnus* stands living in managed plantations and in naturally established populations. Nodule biomass varies depending on stand age, species composition, tree size, stand density, and soil nutrient concentrations (Binkley 1981, 1982; Bormann and Gordon 1984; Sharma and Ambasht 1986; Binkley et al. 1992; Uliassi and Ruess 2002; Lee and Son 2005; Son et al. 2007) and has been estimated to range from 16 to 480 kg ha⁻¹ (Table 1; Binkley 1981; Hurd et al. 2001). No clear association has been found between nodule biomass and stand age but nodule biomass tends to increase early until a stand is 10–15 years old (Fig. 1a). Variations in below-ground biomass estimates are higher than those of above-ground

Table 1 Estimates of nodule biomass in *Alnus* stands

Species	Nodule biomass (kg ha ⁻¹)	Stand age (years)	Tree density (trees ha ⁻¹)	References
<i>Alnus glutinosa</i>	454	5–20		Akkermans and van Dijk (1976)
<i>Alnus hirsuta</i>	88	17	1114	Tobita et al. (2010a)
<i>Alnus hirsuta</i>	179	27	1960	Son et al. (2007)
<i>Alnus hirsuta</i>	95	27	1130	Son et al. (2007)
<i>Alnus hirsuta</i>	220	38	700	Lee and Son (2005)
<i>Alnus incana</i> ssp. <i>incana</i>	170	5	12660	Aosaar et al. (2013)
<i>Alnus incana</i> ssp. <i>incana</i>	160	10	7400	Aosaar et al. (2013)
<i>Alnus incana</i> ssp. <i>incana</i>	310	17	5100	Aosaar et al. (2013)
<i>Alnus incana</i> ssp. <i>incana</i>	44	2		Huss-Danell and Ohlsson (1992)
<i>Alnus incana</i> ssp. <i>incana</i>	110	3	40000	Rytter (1989)
<i>Alnus incana</i> ssp. <i>incana</i>	230–480	7		Rytter (1989)
<i>Alnus incana</i> ssp. <i>incana</i>	150	30		Johnsrud (1978)
<i>Alnus incana</i> spp. <i>rugosa</i>	65		19120	Hurd et al. (2001)
<i>Alnus incana</i> spp. <i>rugosa</i>	58			Younger and Kapustka (1983)
<i>Alnus nepalensis</i>	300–420	15–20		Sharma et al. (2010)
<i>Alnus nepalensis</i>	457	7	715	Sharma and Ambasht (1986)
<i>Alnus nepalensis</i>	149	56	435	Sharma and Ambasht (1986)
<i>Alnus rubra</i>	250–325	55	3000	Binkley et al. (1992)
<i>Alnus rubra</i>	16	2		Kim (1987)
<i>Alnus rubra</i>	121	58		Kim (1987)
<i>Alnus rubra</i>	62	5	1240	Bormann and Gordon (1984)
<i>Alnus rubra</i>	146	5	10091	Bormann and Gordon (1984)
<i>Alnus rubra</i>	390	15–20		Binkley (1981)
<i>Alnus rubra</i>	30–55	2–4	5000–8000	Tripp et al. (1979)
<i>Alnus rubra</i>	117	7		Zavitkovski and Newton (1968)
<i>Alnus rubra</i>	244	30		Zavitkovski and Newton (1968)
<i>Alnus viridis</i> ssp. <i>sinuata</i>	130	5		Binkley (1982)
<i>Alnus viridis</i> ssp. <i>sinuata</i>	110	15–20		Binkley (1981)
<i>Alnus viridis</i> ssp. <i>viridis</i>	43			Moiroud and Capellano (1979)

Adapted from Binkley (1981) and Hurd et al. (2001)

estimates due to methodological difficulties (Aosaar et al. 2013). The spatial distribution patterns of nodules tend to be more homogeneous horizontally as a plantation ages (Rytter 1989) or with increasing tree size in a naturally established stand (Tobita et al. 2010a), although large variations in nodule distribution are also observed on the basis of stand age and tree size. These findings suggest that the distance from the *Alnus* tree must be considered to estimate nodule biomass in *Alnus* stands.

Seasonal variations in N₂ fixation activities and rates in *Alnus* stands

N₂ fixation begins shortly after leaf emergence in spring, remains high but variable in summer, decreases in late autumn, and ceases when all the leaves have been shed (Huss-Danell 1990; Tsutsumi et al. 1993; Sharma et al. 2010; Tobita et al. 2013a). Fluctuations in environmental

conditions such as light, soil temperature, water, mineral nutrition, and pH as well as the presence of *Frankia* strains also affect nodule N₂ fixation activity (Pawlowski and Newton 2008; Gtari et al. 2013; Tobita et al. 2013b). Declines in N₂ fixation activity caused by these factors are often related to a deficiency of carbohydrates supplied from leaves to nodules because N₂ fixation activity depends on newly formed photosynthates supplied by the host plant (Huss-Danell 1997). Nonstructural carbohydrate accretion is greater in nodules in fall, contributing to the maintenance of overall plant levels of N₂ fixation similar to those observed during summer (Kaelke and Dawson 2005). Therefore, N₂-fixing root nodules are a strong metabolic sink for photosynthates within a plant (Huss-Danell and Sellstedt 1983; Ruess et al. 2006). This character of nodules may help *Alnus* species adapt to photosynthesis under elevated CO₂ conditions (Koike et al. 1997; Tobita et al. 2010b, 2011), as will be described in detail hereinafter.

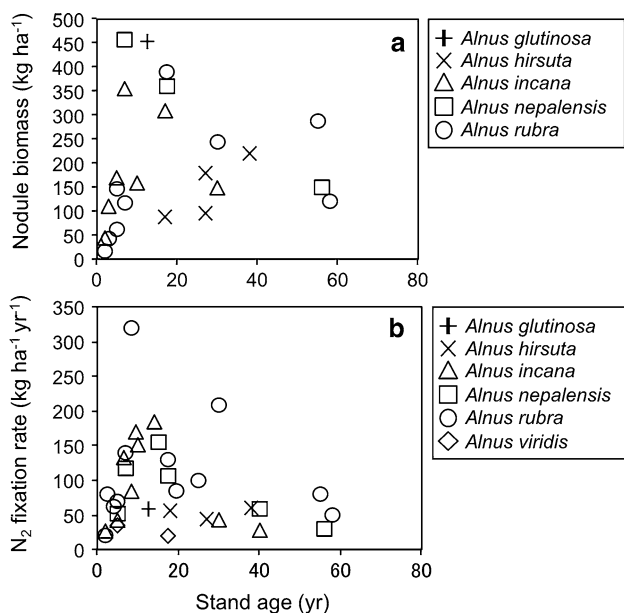


Fig. 1 Relationships between stand age and nodule biomass (a) and N_2 fixation rate (b) in *Alnus* stands. Adapted from Tables 1 and 2

Alnus species N_2 fixation rates are estimated to be several $kg\ N\ ha^{-1}\ year^{-1}$ to $320\ kg\ N\ ha^{-1}\ year^{-1}$ after several major assumptions are met (Table 2; e.g., Binkley 1981; Hibbs and Cromack 1990; Rytter et al. 1991; Cleveland et al. 1999; Hurd et al. 2001; Löhmus et al. 2002; Uri et al. 2004; Lee and Son 2005). The nitrogenase activity of each nodule and N_2 fixation rate per plant vary depending on the N demand with increasing tree age (Son et al. 2007). The N_2 fixation rate increased with stand age and was higher in 10- to 20-year-old stands than in older stands (Fig. 1b). The contribution of N_2 fixation to N economy increases with stand age, peaking in a 15- to 20-year-old *A. nepalensis* stand (Sharma et al. 2002).

While many studies listed in Table 2 adopted an acetylene reduction assay (ARA) to evaluate the N_2 fixation activity, ARA results should be interpreted cautiously because ARA has been the subject of many criticisms due to assay inconsistency (Winship and Tjepkema 1990; Silvester et al. 2008). One is the so-called C_2H_2 -induced decline in NA, which is apparent to varying degrees in actinorhizal nodules, including those of *Alnus* species (Tjepkema et al. 1988; Schwintzer and Tjepkema 1997). This decline is often followed by either partial or full recovery, which is dependent on the host species, growth conditions, and plant age (Silvester and Winship 1990). In addition, the conversion rate of C_2H_2 reduced to fixed N_2 in the ARA, which was set to 3:1 to compare the N_2 fixation data in Table 2, can also produce result errors (Winship and Tjepkema 1990). The actual ratio of acetylene reduction to N_2 fixation must be determined using ^{15}N -labeled dinitrogen concurrently (Schwintzer and Tjepkema 1997).

Facilitating effects of N_2 fixation in the *Alnus*–*Frankia* symbiotic relationship

N input into soil through N_2 fixation by *Alnus* species boosts N soil content (Wurtz 1995; Rhoades et al. 2001; Myrold and Huss-Danell 2003; Uri et al. 2014) and enhances the leaf N content and growth rates of mixed-planted trees (Vogel and Gower 1998; Brockley and Sanborn 2003; Roggy et al. 2004; Avendano-Yanez et al. 2014). However, some negative effects of N_2 -fixing *Alnus* species have been reported such as the competition for light and soil nutrients (Chapin et al. 1994; Brockley and Sanborn 2003; Simard et al. 2006; Chapin et al. 2011), N leaching, gaseous N emissions due to denitrification (Compton et al. 2002; Mander et al. 2008, 2015), and the issue of invasion (Hiltbrunner et al. 2014).

Alnus species usually exhibit lower N resorption rates than those of non- N_2 -fixers (Uliassi and Ruesch 2002). These lower N_2 resorption rates can cause relatively high photosynthetic rates in autumn, which may help retain relatively high N_2 fixation activity (Tateno 2003; Tobita et al. 2013a). In addition, because a low N resorption rate will produce fallen leaves with higher N content, the *Alnus* species leaf litter decomposition rate is usually faster than that of other non- N_2 -fixers (Sharma et al. 2008). The initial C/N ratio of leaf litter in non- N_2 -fixers is usually higher, and their decomposition rate is slower than those of N_2 -fixing species (Tateno et al. 2007). The leaf litter C/N ratio decreases rapidly from 20 to 12 in *A. hirsuta* (Tobita et al. 2013a) and from 20.5 to 15 in *A. japonica* (Yoon et al. 2014), suggesting that litter decomposition immediately moves into the mineralization stage (Takeda 1998). Another feature of *Alnus* species is the high susceptibility of leaves to herbivore damage (Kikuzawa et al. 1979; Tadaki et al. 1987; Tobita et al. 2013a), and their feces are a N input pathway to soils (Meehan and Lindroth 2007). In addition, symbiotic N_2 fixation in *Alnus* species may affect the distribution patterns of regenerated tree species (Tobita et al. 2015) and diversity (Hanley et al. 2006) as well as improve soil fertility. In early successional stages, *Alnus* species are used as nurse trees and may have a mothering role with these regenerated endozoochorous tree species.

Alnus species often regenerate easily during the early stages of succession (Bormann and Sidle 1990). However, it is occasionally difficult for *Alnus* species to recruit and expand their distribution in areas where *Frankia* densities are low (Seeds and Bishop 2009). The availability of infective *Frankia* and their compatibility with the host may limit the successful formation of root nodules capable of N_2 fixation (Markham and Chanway 1999). Symbiotic *Frankia* assemblages can differ widely between sympatric *Alnus* spp. and between successional habitats occupied by a given host species (Anderson et al. 2009). Phylogenetic

Table 2 Estimates of annual N₂ fixation in *Alnus* stands

Species	N ₂ fixation (kgN ha ⁻¹ year ⁻¹)	Stand age (years)	References	Methods ^a	Conversion rate ^b
<i>Alnus glutinosa</i>	58	5–20	Akkermans and van Dijk (1976)	AR	
<i>Alnus hirsuta</i>	56	18	Tobita et al. (2013a)	AR	3
<i>Alnus hirsuta</i>	41–47	27	Son et al. (2007)	AR	3
<i>Alnus hirsuta</i>	60	38	Lee and Son (2005)	AR	3
<i>Alnus incana</i> ssp. <i>incana</i>	152	10	Uri et al. (2011)	ACC	
<i>Alnus incana</i> ssp. <i>incana</i>	42	5	Uri et al. (2004)	ACC	
<i>Alnus incana</i> ssp. <i>incana</i>	185	14	Löhmus et al. (2002)	ACC	
<i>Alnus incana</i> ssp. <i>incana</i>	28	40	Löhmus et al. (2002)	ACC	
<i>Alnus incana</i> ssp. <i>incana</i>	27 (20)	2	Huss-Danell et al. (1992)	AR	3 (4)
<i>Alnus incana</i> ssp. <i>incana</i>	113–153 (85–115)	6–7	Rytter et al. (1991)	AR	3 (4)
<i>Alnus incana</i> ssp. <i>incana</i>	43 (32)	30	Johnsrud (1978)	AR	3 (4)
<i>Alnus incana</i> ssp. <i>rugosa</i>	49 (37)		Hurd et al. (2001)	AR	3 (4)
<i>Alnus incana</i> ssp. <i>rugosa</i>	1–5 (1–4)		Younger and Kapustka (1983)	AR	3 (4)
<i>Alnus incana</i> ssp. <i>rugosa</i>	85	1–16	Voigt and Steucek (1969)	ACC	
<i>Alnus incana</i> ssp. <i>rugosa</i>	170	1–18	Daly (1966)	ACC	
<i>Alnus nepalensis</i>	57–155	15–20	Sharma et al. (2010)	AR	3
<i>Alnus nepalensis</i>	52	5	Sharma et al. (2002)	AR	3
<i>Alnus nepalensis</i>	155	15	Sharma et al. (2002)	AR	3
<i>Alnus nepalensis</i>	59	40	Sharma et al. (2002)	AR	3
<i>Alnus nepalensis</i>	117	7	Sharma and Ambasht (1988)	AR	3
<i>Alnus nepalensis</i>	29	56	Sharma and Ambasht (1988)	AR	3
<i>Alnus rubra</i>	140	7	Zavitkovski and Newton (1968)	GAN	
<i>Alnus rubra</i>	209	30	Zavitkovski and Newton (1968)	GAN	
<i>Alnus rubra</i>	62	4	Tripp et al. (1979)	AR	3
<i>Alnus rubra</i>	75–85	55	Binkley et al. (1992)	AR	3
<i>Alnus rubra</i>	18–23	2	Kim (1987)	AR	3
<i>Alnus rubra</i>	50	58	Kim (1987)	AR	3
<i>Alnus rubra</i>	70	5	Bormann and Gordon (1984)	AR	3
<i>Alnus rubra</i>	130	15–20	Binkley (1981)	AR	3
<i>Alnus rubra</i>	100	10–40	Bormann and DeBell (1981)	ACC	
<i>Alnus rubra</i>	80	1–4	DeBell and Radwan (1979)	ACC	
<i>Alnus rubra</i>	85	1–38	Cole et al. (1978)	ACC	
<i>Alnus rubra</i>	320	2–15	Newton et al. (1968)	ACC	
<i>Alnus tenuifolia</i>	59		Uliassi and Ruess (2002)	AR	3
<i>Alnus viridis</i> ssp. <i>sinuata</i>	10–15		Sanborn et al. (2002)	15N	
<i>Alnus viridis</i> ssp. <i>sinuata</i>	35	5	Binkley (1982)	AR	3
<i>Alnus viridis</i> ssp. <i>sinuata</i>	20 (15)	15–20	Binkley (1981)	AR	3 (4)
<i>Alnus viridis</i> ssp. <i>viridis</i>	8 (6)		Moiroud and Capellano (1979)	AR	3 (4)

Adapted from Binkley (1981), Hibbs and Cromack (1990), and Hurd et al. (2001)

^a N₂ fixation estimate method; AR acetylene reduction assay; ACC accretion study; GAN greenhouse accretion per gram of nodule × nodule biomass in the field; 15N ¹⁵N isotope dilution method

^b Molar conversion ratio of acetylene reduction to N₂ fixation

specificity is a significant factor in the *Alnus tenuifolia*–*Frankia* interaction, and significant habitat-based differentiation may exist among *A. tenuifolia*-infective genotypes (Anderson et al. 2013). The global biogeographic

community of *Alnus*-associated *Frankia* (Benson and Dawson 2007; Pölme et al. 2014) and the genetic diversity of *Frankia* populations in the soil and root nodules (Pokharel et al. 2011) have also been evaluated.

Predicted effects of elevated CO₂ on *Alnus*–*Frankia* N₂ fixation

Increasing CO₂ is an important factor influencing global climate change (IPCC 2013) and nutrient availability, particularly that of soil N, may strongly limit the growth response of woody plants to elevated CO₂ (Norby et al. 2010; Zak et al. 2011; Sigurdsson et al. 2013) because forest ecosystems are usually N limited (Wang et al. 2010). Symbiotic N₂ fixation may play an important role as exogenous N input to sustain the enhanced growth of non-N₂-fixers under an elevated CO₂ environment (Finzi et al. 2007). However, because N₂ fixation is influenced by several abiotic and biotic factors, it is predicted that N₂-fixers, such as *Alnus* species, do not always enhance their N₂ fixation ability under elevated CO₂ (Tobita et al. 2010b). We will discuss the probable responses of *Alnus* species to elevated CO₂, considering other factors, such as N, P, and water conditions, by reviewing the results from chamber experiments (Tobita et al. 2011) and recent free-air CO₂ enrichment (FACE) experiments (Millett et al. 2012). In addition, we will review the understanding of the effects of elevated ozone (O₃) (Wittig et al. 2009) and leaf chemistry in relation to herbivores (Koike et al. 2006), which can decrease the growth of *Alnus* species.

Photosynthetic and growth responses to elevated CO₂ in *Alnus* species

As N₂ fixers in legumes and actinorhizal plants are largely independent of soil N content, they may respond to elevated CO₂ more directly than non-N₂-fixers by increasing the photosynthetic and growth rates (Temperton et al. 2003a; Reverchon et al. 2012). Excellent reviews have summarized these responses in legumes (Ainsworth and Long 2004; Ainsworth and Rogers 2007; Rogers et al. 2009). *Alnus* species exhibit a photosynthetic acclimation response to elevated CO₂ (Vogel and Curtis 1995), which means they increase their photosynthetic rates under elevated CO₂ compared to those under ambient CO₂ even in N₂-deficient soil (Koike et al. 1997; Tobita et al. 2010b, 2011), rather than downregulate photosynthesis (Long et al. 2004; Ainsworth and Rogers 2007). *Alnus hirsuta* saplings used in FACE experiments in Japan also did not downregulate photosynthesis in infertile soil, whereas photosynthesis was downregulated in two *Betula* species under elevated CO₂, regardless of the soil fertility (Eguchi et al. 2008a). As mentioned in the previous section, the N₂-fixing root nodules of *Alnus* species act as a strong metabolic sink for photosynthates to avoid photosynthetic downregulation under elevated CO₂ conditions. Biomass production by *Alnus* species is significantly stimulated by increasing CO₂ in the presence of *Frankia* species, whereas they show no

response to elevated CO₂ in the absence of *Frankia* species (Pourhassan et al. 2015).

Interactive effects of soil N and elevated CO₂ on N₂ fixation in *Alnus* species

The positive photosynthetic response to elevated CO₂ by N₂-fixing plants increases the C supply to root nodules (Tissue et al. 1997), which may stimulate N₂ fixation in trees. N₂ fixation may be adjusted in response to environmental change, either through variations in nodule biomass or NA (Valverde et al. 2002). Elevated CO₂ increases the total amount of N₂ fixed per *Alnus* species plant because of increased nodule mass (Hibbs et al. 1995; Tobita et al. 2010b) and NA (Temperton et al. 2003a), or both (Norby 1987; Arnone and Gordon 1990; Vogel et al. 1997), as reported by several growth chamber and open-top chamber experiments. One important level of plant control during actinorhizal symbiosis may be the regulation of the proportion of symbiotic tissue in the plant relative to plant biomass allocation (Wall and Berry 2008). Moreover, elevated CO₂ has no effect on the relationship between plant mass and nodule mass, even when nodule biomass increases under elevated CO₂ conditions (Hibbs et al. 1995; Tobita et al. 2005, 2010b). These results suggest that elevated CO₂ enhances nodule mass as a function of the increasing total plant mass, rather than by enhancing the allocation of biomass to roots and nodules.

Soil mineral N content often limits nodule formation and NA because larger quantities of photosynthates are needed for N₂ fixation compared with N, which can be absorbed from the soil (Ekblad and Huss-Danell 1995; Vogel et al. 1997; Lambers et al. 2008; Wall and Berry 2008; Chapin et al. 2011). Thomas et al. (2000) suggested that elevated CO₂ mitigates these inhibitory effects of substrate N in leguminous tree species, either through increased allocation of C to nodules or through increased N demand by the plant. However, increased soil N availability has a negative effect on nodule production and biomass allocation to nodules in *Alnus* species, regardless of CO₂ treatment (Koike et al. 1997; Bucher et al. 1998; Temperton et al. 2003b; Tobita et al. 2005). These results indicate that the inhibitory effect of high soil N availability on nodulation in *Alnus* species is retained even under elevated CO₂ levels.

N₂ fixation response in *Alnus* species subjected to FACE experiments

Only two FACE experiments have been reported on *Alnus* species. One was conducted in Japan (Hokkaido), as introduced in the previous section, on the responses of *A. hirsuta* to elevated CO₂ in fertile and infertile soils compared to those of non-N₂-fixing deciduous tree

species, including *Betula platyphylla*, *Betula maximowicziana*, *Quercus mongolica*, and *Fagus crenata* (Agari et al. 2007; Eguchi et al. 2008a, 2008b; Watanabe et al. 2010). The other was the UK Bangor FACE experiment in which the effects of elevated CO₂ on *A. glutinosa* performance were compared between monocultures and mixed plantings of *Betula pendula*, *Fagus sylvatica*, and *Populus tremula* × *tremuloides* (Hoosbeek et al. 2011; Millett et al. 2012; Smith et al. 2013a, 2013b; Godbold et al. 2014; Scullion et al. 2014). N₂ fixation in *A. glutinosa* increases under elevated CO₂ despite the absence of significant growth stimulation in a mixed-species stand after 4 years (Millett et al. 2012). However, the fraction of N₂ derived from N₂ fixation, calculated using the ¹⁵N natural abundance method (Chaia and Myrold 2010; Zhang et al. 2014), was unaffected by the elevated CO₂ in an *A. glutinosa* monoculture stand, indicating no increase in N₂ fixation under elevated CO₂ in a monoculture although plant biomass increased significantly (Hoosbeek et al. 2011). These differences in responses to elevated CO₂ may be related to enhanced growth rate, N uptake, and N₂ fixation of *A. glutinosa* in a mixed stand compared to those in a monoculture due to increased ecosystem resource utilization through below-ground niche differentiation among trees (Smith et al. 2013a). In contrast, N₂ fixation in *Lupinus* species legumes increases under elevated CO₂ in both a monoculture and a mixed grassland system in a FACE experiment (Lee et al. 2003). Plants rarely grow in isolation, and their response to elevated CO₂ can be affected by the extent and type of plant–plant interactions (Poorter and Navas 2003). Understanding how mixed-species forests respond to elevated CO₂ will be essential to assess forest growth dynamics including the response of N₂ fixation in *Alnus* species and improving the parameterization of global change cycle models (Norby and Zak 2011).

N allocation in *Alnus* species under elevated CO₂

N concentrations generally decline in plant tissues under elevated CO₂ (Ainsworth and Rogers 2007; Sardans and Peñuelas 2012). N uptake is not affected as much as C uptake, whereas increased CO₂ alters the plant C/N balance (Kallarackal and Roby 2012). The increase in the total *Alnus* species plant N mass under elevated CO₂ is smaller than that predicted by the response of the total biomass to elevated CO₂ in phytotron experiments (Temperton et al. 2003b; Tobita et al. 2011). The same phenomenon was observed in FACE studies, which showed increased N use efficiency in an *A. glutinosa* monoculture stand under elevated CO₂ (Millett et al. 2012; Pourhassan et al. 2015). These results suggest that it is necessary to evaluate biomass accumulation as well as total N content and its

allocation when considering the N₂-fixing ability of *Alnus* species under elevated CO₂.

Interactive effects of soil P and elevated CO₂ on N₂ fixation in *Alnus* species

N availability limits plant responses to elevated CO₂ (Norby et al. 2010). However, higher soil N availability under elevated CO₂ does not necessarily lead to higher plant biomass production (Körner et al. 2005; Schleppei et al. 2012) because stoichiometric constraints extend to elements other than N, such as P, or some micronutrients. More generally, any biomass response to elevated CO₂ is controlled by the stoichiometric balance among many elements required to construct new tissues and used for active metabolism (Hungate et al. 2004; Sardans and Peñuelas 2012; Leuzinger and Härrenschwiler 2013). Many studies have focused on N but P limitations are also common in many terrestrial ecosystems (Nord and Lynch 2009; Wang et al. 2010). In addition, P is unlikely to increase in the future because it is a non-renewable resource (Pandey et al. 2015). N₂ fixation in actinorhizal plants as well as legumes is a P-consuming activity that accompanies the synthesis of DNA and plasma membranes for cell division during nodule development and ATP synthesis to reduce N (Gentili et al. 2006). Therefore, P is often the most growth-limiting nutrient for actinorhizal plants because of the relatively high demand for P compared to that of non-N₂-fixers (Ingestad 1981; Uliassi et al. 2000; Brown et al. 2011). P deficiency limits nodule formation and N₂ fixation in *A. incana*, even under ambient CO₂ conditions (Gentili and Huss-Danell 2003; Ruess et al. 2013). Although studies on the combined effects of elevated CO₂ and P deficiency on N₂ fixation are scarce, N₂ fixation per plant in two *Alnus* species does not increase under elevated CO₂ and P-deficient conditions because plant growth is strongly suppressed and nodule formation is inhibited without a marked change in NA (Tobita et al. 2010b). In the BangorFACE experiment, Smith et al. (2013a) suggested that soil P availability, rather than N, have been a limiting factor compared to that at other FACE sites because the experimental site was established on former agricultural soils. Legumes exhibit a particularly strong increase in the biomass production under combined elevated CO₂ and P fertilization conditions in grassland communities (Stöcklin and Körner 1999), indicating a stoichiometric control of the CO₂ effects (Leuzinger and Hättenschwiler 2013). The increased demand for P to support an increased growth rate may be another key constraint on plant responses to elevated CO₂ (Kogawara et al. 2006). Tripartite symbiosis with mycorrhiza (Yamanaka et al. 2003; Urgiles et al. 2014) may play an important role to meet the increased P demand and to

effectively utilize P under projected elevated CO₂ conditions.

Interactive effects of drought and elevated CO₂ on N₂ fixation in *Alnus* species

Increasing temperatures have been predicted to decrease precipitation during summer (Calfapietra et al. 2010). Drought conditions have the potential to decrease the photosynthetic rates and limit growth through stomatal closure (Flexas and Medrano 2002), which may also affect symbiotic N₂ fixation. Drought limits the N₂-fixing capacity by limiting metabolic capacity of bacteria and by generating oxidative damage in legumes (Aranjuelo et al. 2011). Several *Alnus* species such as *A. rubra* are sensitive to water stress (Pezeshki and Hinckley 1988), and NA is sensitive to short-term drought (Huss-Danell 1997). In contrast, NA in *A. glutinosa* shows no marked decrease after adaptation to moderately dry soil (Seiler and Johnson 1984). Most studies that have been conducted on elevated CO₂ and drought indicate that elevated CO₂ tends to ameliorate the negative effects of drought on the net photosynthetic rates and biomass accumulation (Kitao et al. 2007; Sicher and Barnaby 2012; Feng et al. 2014). Plant growth is stimulated by elevated CO₂ in C₃ species, even under moderate drought conditions (Xu et al. 2013). The light-saturated net photosynthetic rate and growth of *A. hirsuta* and *A. maximowiczii* are also enhanced by elevated CO₂ under sufficient P conditions, even in dry soil (Tobita et al. 2010b). However, when soil P is limiting, these two *Alnus* species have no positive responses to elevated CO₂. Elevated CO₂ alters leaf water potential of these two *Alnus* species in wet soil (Uemura et al. 2009) and increases the susceptibility to photoinhibition (Tobita et al. 2008). These results suggest that the risk of an occasional severe drought increases under elevated CO₂, particularly when these *Alnus* species are grown in wet soil.

Sensitivity to tropospheric ozone under elevated CO₂

Tropospheric O₃ levels have increased globally since pre-industrial times (IPCC 2007, 2013) and continue to rise, particularly in East Asia (Fowler et al. 2008). Ozone and CO₂ are two major anthropogenic air pollutants with opposing impacts on plant growth (Lindroth 2010; Leisner and Ainsworth 2012) because increased O₃ reduces net photosynthesis. *Alnus* species are relatively sensitive to O₃, and *A. incana* have been used as O₃ bioindicators in Europe (Manning et al. 2002; Manning and Godzik 2004). Surface ambient background O₃ over land in the northern hemisphere has already increased to levels that have decreased growth in several tree species (Matyssek et al. 2007),

including *Alnus viridis* (VanderHeyden et al. 2001). In addition, increased O₃ decreases shoot and root dry weights and enhances leaf senescence in *A. incana* (Mortensen and Skre 1990; Wittig et al. 2009). Although the enhanced growth of *Alnus* species will be expected to increase the N input in ecosystems under future elevated CO₂ conditions, the projected increase in future O₃ level may decrease the growth of *Alnus* species.

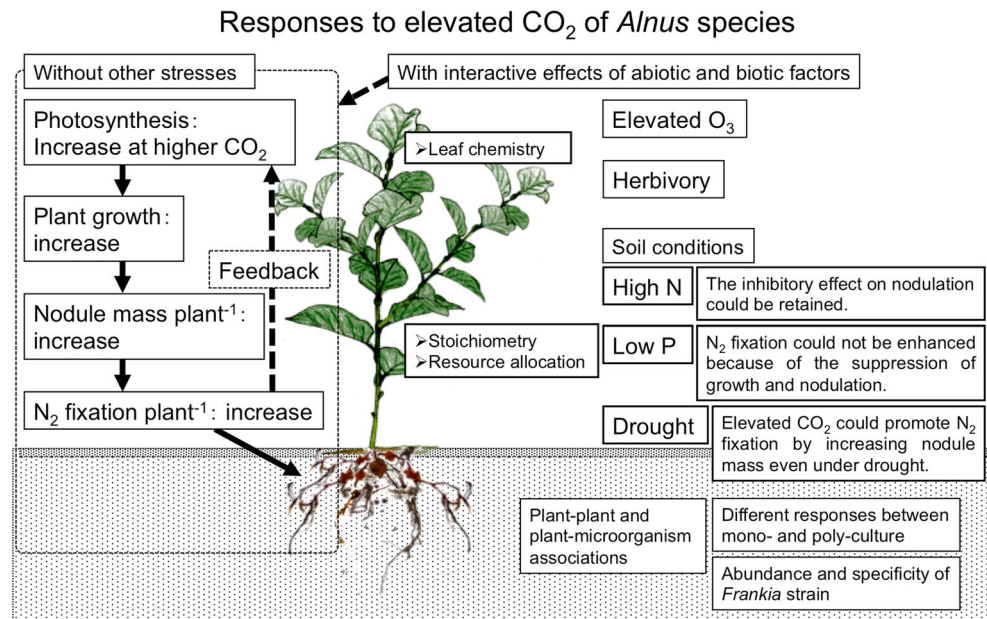
The legume soybean is an O₃ sensitive crop (Mills et al. 2007). Rising O₃ decreases yield (Morgan et al. 2003; Long et al. 2005) and alters the gene expression in the reproductive tissues of soybean (Leisner et al. 2014). Root biomass and the number of root nodules decreased in two clover species in response to O₃, and one revealed a reduced N₂ fixation rate under elevated O₃ (Hewitt et al. 2014). In contrast, intact subalpine grassland communities that include legumes (clover) show low sensitivity to O₃, despite the high O₃ sensitivity found in earlier experiments using pot-grown plants (Bassin et al. 2013).

Considering the interactive effects of elevated CO₂ and O₃, a key question is whether elevated CO₂ will ameliorate the negative effect of O₃, which is an oxidative stressor in plants, or whether O₃ will offset the positive effect of elevated CO₂ on plant growth (Feng et al. 2014). A FACE experiment on soybean (SoyFACE) was performed to investigate the interactive effects of elevated CO₂ and O₃ (Gillespie et al. 2012) and revealed that growth under elevated CO₂ conditions could decrease many of the negative effects of elevated O₃ on plant physiology. However, a clover FACE experiment in the forest understory (AspenFACE) showed that enriched CO₂ and O₃ have large direct and indirect effects on colonization, establishment, and performance (Awmack et al. 2007). Thus, it will become important to also determine the interactive effects of elevated CO₂ and O₃ on N₂ fixation by *Alnus* species to predict N supply in future forest ecosystems.

Effect of elevated CO₂ on *Alnus* species leaf chemistry

Elevated CO₂ and O₃ can change leaf chemistry, such as C, N, P, lignin, and secondary metabolites (Lindroth 2012). In addition, these changes in leaves can alter leaf litter quality, which may affect palatability to detritivores, decomposition, and nutrient turnover (Dray et al. 2014). The defense capacity of broadleaf trees usually increases under elevated CO₂ (Lindroth 2010, 2012). However, the survival rates and longevity of silkworm fed *A. hirsuta* leaves are independent of CO₂ level, unlike what occurs with the non-N₂-fixers *Betula platyphylla*, *Quercus mongolica*, and *Acer mono*. In addition, the survival rates and longevity of silkworms are enhanced by infertile soil (Koike et al. 2006). *Alnus hirsuta* leaves do not have increased levels of

Fig. 2 Schematic diagram of the responses of growth and N_2 fixation in *Alnus* species to elevated CO_2 with and without the interactive effects of many abiotic and biotic factors



defense chemicals in plants held in an elevated CO_2 environment (Koike et al. 2006; Agari et al. 2007), whereas the concentrations of condensed tannins in the leaves of *A. maximowiczii*, which show limited leaf production than that of *A. hirsuta*, are much higher than those of *A. hirsuta*, even under ambient CO_2 and increased under elevated CO_2 (Agari et al. 2007). These results indicate that some variations in the defense strategy may occur under elevated CO_2 conditions, even within *Alnus* species. The litter chemistry of *A. glutinosa* was largely unaffected by elevated CO_2 in a FACE experiment, unlike that of *Betula pendula* (Dray et al. 2014). The feeding behavior of invertebrates on *Alnus* leaves shows large species-specific variations (Dray et al. 2014; Scullion et al. 2014), and only two invertebrate species revealed compensatory feeding when consuming more elevated- CO_2 litter than litter produced under ambient- CO_2 . A few studies have evaluated the impacts of elevated O_3 on soil invertebrate performance and litter decomposition (Lindroth 2012). Therefore, it is necessary to conduct multi-factorial FACE experiments under both elevated CO_2 and O_3 using *Alnus* species (Kawaguchi et al. 2012; Lindroth 2012; Kostianinen et al. 2014) to better understand whether N_2 fixation by *Alnus* species is a potential source of N in forest ecosystem under elevated CO_2 conditions.

Conclusion

The *Alnus*–*Frankia* symbiotic relationship fixes as much N as that of the legume–*Rhizobium* symbiotic relationship and has been utilized to revegetate and rehabilitate

N-deficient disturbed areas. In addition, N_2 fixation by *Alnus* species may affect the distribution pattern of regenerated plants while improving soil fertility. N_2 fixation because of *Alnus*–*Frankia* symbiosis could supply an important source of N needed to sustain increased N uptake due to high rates of forest productivity in the face of global climate change under elevated CO_2 . However, recent findings including those from FACE experiments, suggest that the response of N_2 fixation to elevated CO_2 in *Alnus* species depends on the composition of mixed non- N_2 -fixing species and that soil N and P availability as well as many other abiotic and biotic factors also have interactive effects on N_2 fixation (Fig. 2). Because elevated CO_2 can alter plant N and P contents and stoichiometry, it will be necessary to evaluate N mass allocation as well as biomass accumulation when investigating the N_2 fixing ability of *Alnus* species. In addition, because *Alnus* species are relatively sensitive to O_3 , determining the responses of *Alnus* species to increased CO_2 and O_3 levels will be important to predict N supply in future forest ecosystems.

Author contribution statement H. Tobita wrote the manuscript. H. Tobita, K. Yazaki, H. Harayama, and M. Kitao compiled the review. K. Yazaki, H. Harayama, and M. Kitao revised the manuscript.

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Compliance with ethical standards

Conflict of interest We have no conflict of interest.

References

- Agari T, Matsuki S, Tobita H et al (2007) The effects of elevated CO₂ and soil fertility on the defense capacity against herbivore in two species of alder seedlings. *Trans Mtg Hokkaido Br For Soc* 55:56–58 (in Japanese)
- Ainsworth EA, Long SP (2004) What have we learned from 15 years of free-air CO₂ enrichment (FACE)? A meta-analytic review of the responses of photosynthesis, canopy properties and plant production to rising CO₂. *New Phytol* 165:351–372
- Ainsworth EA, Rogers A (2007) The response of photosynthesis and stomatal conductance to rising [CO₂]: mechanisms and environmental interactions. *Plant Cell Environ* 30:258–270
- Akkermans ADL, van Dijk C (1976) The formation and nitrogen-fixing activity of the root nodules of *Alnus glutinosa* under field conditions. In: Nutman PS (ed) *Symbiotic nitrogen fixation in plants*. Cambridge University Press, London, pp 511–520
- Anderson MD, Ruess RW, Myrold DD, Taylor DL (2009) Host species and habitat affect nodulation by specific *Frankia* genotypes in two species of *Alnus* in interior Alaska. *Oecologia* 160:619–630
- Anderson MD, Taylor DL, Ruess RW (2013) Phylogeny and assemblage composition of *Frankia* in *Alnus tenuifolia* nodules across a primary successional sere in interior Alaska. *Molecular Ecol* 22:3864–3877
- Aosaar J, Varik M, Lõhmus K et al (2013) Long-term study of above- and below-ground biomass production in relation to nitrogen and carbon accumulation dynamics in a grey alder (*Alnus incana* (L.) Moench) plantation on former agricultural land. *Europ J For Res* 132(5–6):737–749
- Aranjuelo I, Molero G, Erice G et al (2011) Plant physiology and proteomics reveals the leaf response to drought in alfalfa (*Medicago sativa* L.). *J Exp Bot* 62:111–123
- Amone JA III, Gordon JC (1990) Effect of nodulation, nitrogen fixation and CO₂ enrichment on the physiology, growth and dry mass allocation of seedlings of *Alnus rubra* Bong. *New Phytol* 116:55–66
- Avendano-Yanez MD, Sanchez-Velasquez LR, Meave JA, Pineda-Lopez MD (2014) Is facilitation a promising strategy for cloud forest restoration? *For Ecol Manage* 329:328–333
- Awmack CS, Mondor EB, Lindroth RL (2007) Forest understory clover populations in enriched CO₂ and O₃ atmosphere: interspecific, intraspecific, and indirect effects. *Environ Exp Bot* 59:340–346
- Bassin S, Volk M, Fuhrer J (2013) Species composition of subalpine grassland is sensitive to nitrogen deposition, but not to ozone, after seven years of treatment. *Ecosystems* 16:1105–1117
- Benson D, Dawson JO (2007) Recent advances in biogeography and genecology of symbiotic *Frankia* and its host plants. *Physiol Plant* 130:318–330
- Binkley D (1981) Nodule biomass and acetylene reduction rates of red alder and Sitka alder on Vancouver Island, B.C. *Can J For Res* 11:181–286
- Binkley D (1982) Nitrogen fixation and net primary production in a young Sitka alder stand. *Can J Bot* 60:281–284
- Binkley D, Sollins P, Bell R et al (1992) Biogeochemistry of adjacent conifer and alder-conifer stands. *Ecology* 73:2022–2033
- Bormann BT, DeBell DS (1981) Nitrogen content and other soil properties related to age of red alder stands. *Soil Sci Soc Am* 45:428–432
- Bormann BT, Gordon JC (1984) Stand density effects in young red alder plantations: productivity, photosynthate partitioning, and nitrogen fixation. *Ecology* 65:394–402
- Bormann BT, Sidle RC (1990) Changes in productivity and distribution of nutrients in a chronosequence at Glacier Bay national park, Alaska. *J Ecol* 78:561–578
- Brockley RP, Sanborn P (2003) Effects of Sitka alder on the growth and foliar nutrition of young lodgepole pine in the central interior of British Columbia. *Can J For Res* 33:1761–1771
- Brown KR, Courtin PJ, Negrave RW (2011) Growth, foliar nutrition and δ¹³C responses of red alder (*Alnus rubra*) to phosphorus additions soon after planting on moist sites. *For Ecol Manage* 262:791–802
- Bucher JB, Tarjan DP, Siegwolf RTW et al (1998) Growth of a deciduous tree seedlings community in response to elevated CO₂ and nutrient supply. *Chemosphere* 36:777–782
- Calfapietra C, Ainsworth EA, Beier C et al (2010) Challenges in elevated CO₂ experiments on forests. *Trends Plant Sci* 15:5–10
- Chaia EE, Myrold DD (2010) Variation of ¹⁵N natural abundance in leaves and nodules of actinorhizal shrubs in Northwest Patagonia. *Symbiosis* 50:97–105
- Chapin FSIII, Walker LR, Fastie CL, Sharman LC (1994) Mechanisms of primary succession following deglaciation at Glacier bay, Alaska. *Ecol Monog* 64(2):149–175
- Chapin FSIII, Matson PA, Vitousek PM (2011) *Principles of terrestrial ecosystem ecology*, 2nd edn. Springer, New York
- Claessens H, Oosterbaan A, Savill P, Rondeux J (2010) A review of the characteristics of black alder (*Alnus glutinosa* (L.) Gaertn.) and their implications for silvicultural practices. *Forestry* 83:164–175
- Cleveland CC, Townsend AR, Schimel DS et al (1999) Global patterns of terrestrial biological nitrogen (N₂) fixation in natural ecosystems. *Global Biochem Cycle* 13:623–645
- Cole DW, Gessel SP, Turner J (1978) Comparative mineral cycling in red alder and Douglas-fir. In: Briggs DG, DeBell DS, Atkinson WA (eds) *Utilization and management of alder*. USFS Pacific Northwest Forest and Range Experiment Station, Portland, OR, pp 327–336
- Compton JE, Church MR, Larned ST, Hogsett WE (2002) Nitrogen export from forested watersheds in the Oregon coast range: the role of N₂-fixing red alder. *Ecosystems* 6:773–785
- Daly GT (1966) Nitrogen fixation by nodulated *Alnus rugosa*. *Can J Bot* 44:1607–1621
- Dawson JO (2008) Ecology of actinorhizal plants. In: Pawlowski K, Newton WE (eds) *Nitrogen-fixing actinorhizal symbioses*. Springer, Dordrecht, pp 119–234
- Dawson JO, Gordon JC (1979) Nitrogen fixation in relation to photosynthesis in *Alnus glutinosa*. *Bot Gaz* 140:S70–S75
- DeBell DS, Radwan MA (1979) Growth and nitrogen relations of coppiced black cottonwood and red alder in pure and mixed plantings. *Bot Gaz* 140:S97–S101
- Dray MW, Crowther TW, Thomas SM et al (2014) Effects of elevated CO₂ on litter chemistry and subsequent invertebrate detritivores feeding responses. *PLoS One* 9(1):e86246
- Eguchi N, Karatsu K, Ueda T et al (2008a) Photosynthetic responses of birch and alder saplings grown in a free air CO₂ enrichment system in northern Japan. *Trees* 22:437–447
- Eguchi N, Morii N, Ueda T et al (2008b) Changes in petiole hydraulic properties and leaf water flow in birch and oak saplings in a CO₂-enriched atmosphere. *Tree Physiol* 28:287–295
- Eklblad A, Huss-Danell K (1995) Nitrogen fixation by *Alnus incana* and nitrogen transfer from *A. incana* to *Pinus sylvestris* influenced by macronutrients and ectomycorrhiza. *New Phytol* 131:453–459
- Eriksson E, Johansson T (2006) Effects of rotation period on biomass production and atmospheric CO₂ emissions from broadleaved stands growing on abandoned farmland. *Silva Fennica* 40:603–613

- Feng GQ, Li Y, Cheng ZM (2014) Plant molecular and genomic responses to stresses in projected future CO₂ environment. *Crit Rev Plant Sci* 33:238–249
- Finzi AC, Norby RJ, Calfapietra C et al (2007) Increases in nitrogen uptake rather than nitrogen-use efficiency support higher rates of temperate forest productivity under elevated CO₂. *Proc Natl Acad Sci USA* 104:14014–14019
- Flexas J, Medrano H (2002) Drought-inhibition of photosynthesis in C₃ plants: stomatal and non-stomatal limitations revisited. *Ann Bot* 89:183–189
- Fowler D, Amann M, Anderson R et al (2008) Ground-level ozone in the 21st century: future trends, impacts and policy implications. R Soc Polic Doc, London
- Gentili F, Huss-Danell K (2003) Local and systemic effects of phosphorus and nitrogen on nodulation and nodule function in *Alnus incana*. *J Exp Bot* 54:2757–2767
- Gentili F, Wall LG, Huss-Danell K (2006) Effects of phosphorus and nitrogen on nodulation are seen already at the stage of early cell divisions in *Alnus incana*. *Ann Bot* 98:309–315
- Gillespie KM, Xu F, Richter KT et al (2012) Greater antioxidant and respiratory metabolism in field-grown soybean exposed to elevated O₃ under both ambient and elevated CO₂. *Plant Cell Environ* 35:169–184
- Godbold D, Tullus A, Kupper P et al (2014) Elevated atmospheric CO₂ and humidity delay leaf fall in *Betula pendula*, but not in *Alnus glutinosa* or *Populus tremula* x *tremuloides*. *Ann For Sci* 71:831–842
- Gordon JC, Wheeler CT (1978) Whole plant studies on photosynthesis and acetylene reduction in *Alnus glutinosa*. *New Phytol* 80:179–186
- Gtari M, Tisa LS, Normand P (2013) Diversity of *Frankia* Strains, actinobacterial symbionts of actinorhizal plants. In: Ricardo A (ed) *Symbiotic Endophytes*. Springer, Berlin Heidelberg, pp 123–148
- Hanley TA, Deal RL, Orlikowska EH (2006) Relationships between red alder composition and understory vegetation in young mixed forests of southeast Alaska. *Can J For Res* 36:738–748
- Hewitt DKL, Mills G, Hayes F et al (2014) Highlighting the threat from current and near-future ozone pollution to clover in pasture. *Environ Pollut* 189:111–117
- Hibbs DE, Cromack CJR (1990) Actinorhizal plants in Pacific Northwest forests. In: Schwintzer CR, Tjepkema JD (eds) *The biology of Frankia and actinorhizal plants*. Academic Press Inc, San Diego, pp 343–363
- Hibbs DE, Chan SS, Castellano M, Niu C-H (1995) Response of red alder seedlings to CO₂ enrichment and water stress. *New Phytol* 129:569–577
- Hiltbrunner E, Aerts R, Bühlmann T et al (2014) Ecological consequences of the expansion of N₂-fixing plants in cold biomes. *Oecologia* 176:11–24
- Hoosbeek MR, Lukae M, Velthorst E et al (2011) Free atmospheric CO₂ enrichment increased above ground biomass but did not affect symbiotic N₂-fixation and soil carbon dynamics in a mixed deciduous stand in Wales. *Biogeoscience* 8:353–364
- Hungate BA, Dukes JT, Shaw MR et al (2003) Nitrogen and climate change. *Science* 302:1512–1513
- Hungate BA, Stiling PD, Dijkstra P et al (2004) CO₂ elicits long-term decline in nitrogen fixation. *Science* 304:1291
- Hurd TM, Raynal DJ, Schwintzer CR (2001) Symbiotic N₂ fixation of *Alnus incana* ssp. *rugosa* in shrub wetlands of the Adirondack Mountains, New York, USA. *Oecologia* 126:94–103
- Huss-Danell K (1990) The physiology of actinorhizal nodules. In: Schwintzer CR, Tjepkema JD (eds) *The biology of Frankia and actinorhizal plants*. Academic Press Inc, Tokyo, pp 129–156
- Huss-Danell K (1997) Actinorhizal symbioses and their N₂ fixation. *New Phytol* 136:375–405
- Huss-Danell K, Ohlsson H (1992) Distribution of biomass and nitrogen among plant parts and soil nitrogen in a young *Alnus incana* stand. *Can J Bot* 70:1545–1549
- Huss-Danell K, Sellstedt A (1983) Nitrogenase activity in response to restricted shoot growth in *Alnus incana*. *Can J Bot* 61:2949–2955
- Huss-Danell K, Lundquist PO, Ohlsson H (1992) N₂ fixation in a young *Alnus incana* stand, based on seasonal and diurnal variation in whole plant nitrogenase activity. *Can J Bot* 70:1537–1544
- Hytönen J, Saarsalmi A (2015) Biomass production of coppiced grey alder and the effects of fertilization. *Silva Fennica* 49 no. 1 article id 1260. <http://dx.doi.org/10.14214/sf.1260>
- Hyvönen R, Ågren GI, Linder S et al (2007) The likely impact of elevated [CO₂], nitrogen deposition, increased temperature and management on carbon sequestration in temperate and boreal forest ecosystems: a literature review. *New Phytol* 173:463–480
- Ingestad T (1981) Nutrition and growth of birch and grey alder seedlings in low conductivity solutions and at varied relative rates of nutrient addition. *Physiol Plant* 52:454–466
- IPCC (2007) *Climate change 2007: impacts, adaptation and vulnerability. contribution of working group II to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change*. Parry ML, Canziani OF, Palutikof JP, van der Linden PJ, Hanson CE (eds) Cambridge University Press, Cambridge, UK
- IPCC (2013) *Climate Change 2013: the physical science basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*. Stocker TF, Qin D, Plattner G-K, Tignor M, Allen SK, Boschung J, Nauels A, Xia Y, Bex B, Midgley PM (eds) Cambridge University Press, Cambridge, UK
- Johnson DW (2006) Progressive N limitation in forest: review and implications for long-term responses to elevated CO₂. *Ecology* 87(1):64–75
- Johnsrud SC (1978) Nitrogen fixation by root nodules of *Alnus incana* in a Norwegian forest ecosystem. *Oikos* 30:475–479
- Kaelke CM, Dawson JO (2005) The accretion of nonstructural carbohydrates changes seasonally in *Alnus incana* ssp. *rugosa* in accord with tissue type, growth, N allocation, and root hypoxia. *Symbiosis* 39:61–66
- Kallarackal J, Roby TJ (2012) Responses of trees to elevated carbon dioxide and climate change. *Biodivers Conserv* 21:1327–1342
- Kawaguchi K, Hoshika Y, Watanabe M, Koike T (2012) Ecophysiological responses of northern birch forests to the changing atmospheric CO₂ and O₃ concentration. *J Atmospheric Environ* 6:192–205
- Kikuzawa K, Asai T, Higashiura Y (1979) Leaf production and the effect of defoliation by the larval population of the winter moth, *Operophtera brumata* L. in an alder (*Alnus inokumae* MURAI et KUSAKA) stand. *J J Ecol* 29:111–120
- Kim DY (1987) Seasonal estimates of nitrogen fixation by *Alnus rubra* and *Ceanothus* species in western Oregon forest ecosystems. Dissertation, Oregon State University
- Kitao M, Lei TT, Koike T et al (2007) Interaction of drought and elevated CO₂ on photosynthetic down-regulation and susceptibility to photoinhibition in Japanese white birch (*Betula platyphylla* var. *japonica*) seedlings grown under limited N availability. *Tree Physiol* 27(5):727–735
- Kogawara S, Norisada M, Tange T et al (2006) Elevated atmospheric CO₂ concentration alters the effects of phosphate supply on growth of Japanese red pine (*Pinus densiflora*) seedlings. *Tree Physiol* 26:25–33
- Koike T, Izuta T, Lei TT et al (1997) Effects of high CO₂ on nodule formation in roots of Japanese mountain alder seedlings grown under two nutrient levels. In: Ando T, Fujita K, Mae T, Matsumoto H, Mori S, Sekiya J (eds) *Plant nutrition—for*

- sustainable food production and environment. Kluwer Academic Publishers, Japan, pp 887–888
- Koike T, Tobita H, Shibata T et al (2006) Defense characteristics of deciduous broad-leaved tree seedlings grown under factorial combination of two levels of CO₂ and nutrients. *Popul Ecol* 48:23–29
- Körner C, Asshoff R, Bignucolo O et al (2005) Carbon flux and growth in mature deciduous forest tree exposed to elevated CO₂. *Science* 309:1360–1362
- Kostiainen K, Saranpää P, Lundqvist SO et al (2014) Wood properties of *Populus* and *Betula* in long-term exposure to elevated CO₂ and O₃. *Plant Cell Environ* 37:1452–1463
- Kucho K, Hay AE, Normand P (2010) The determinants of the actinorhizal symbiosis. *Microbes Environ* 25:241–252D
- Lambers H, Chapin III FS, Pons T (2008) *Plant physiological ecology*, 2nd edn. Springer, Berlin
- Lee YY, Son Y (2005) Diurnal and seasonal patterns of nitrogen fixation in an *Alnus hirsuta* plantation of central Korea. *J Plant Biol* 48(3):332–337
- Lee TD, Reich PB, Tjoelker MG (2003) Legume presence increases photosynthesis and N concentrations of co-occurring non-fixers but does not modulate their responsiveness to carbon dioxide enrichment. *Oecologia* 137:22–31
- Leisner CP, Ainsworth EA (2012) Quantifying the effects of ozone on plant reproductive growth and development. *Global Change Biol* 18:606–616
- Leisner CP, Ming R, Ainsworth EA (2014) Distinct transcriptional profiles of ozone stress in soybean (*Glycine max*) flowers and pods. *BMC Plant Biol* 14:335–347
- Leuzinger S, Hättenschwiler S (2013) Beyond global change: lessons from 25 years of CO₂ research. *Oecologia* 171:639–651
- Lindroth RL (2010) Impacts of elevated atmospheric CO₂ and O₃ on forests: phytochemistry, trophic interactions, and ecosystem dynamics. *J Chem Ecol* 36:2–21
- Lindroth RL (2012) Atmospheric change, plant secondary metabolites, and ecological interactions. In: Iason GR, Dicke M, Hartley S (eds) *The ecology of plant secondary metabolites: from genes to global processes*. Cambridge University Press, Cambridge, pp 120–153
- Löhmus K, Kuusemets V, Ivask M et al (2002) Budgets of nitrogen fluxes in riparian gray alder forests. *Archiv fur Hydrobiol* 13:321–332
- Long SP, Ainsworth EA, Rogers A, Ort DR (2004) Riding atmospheric carbon dioxide: plants FACE the future. *Annu Rev Plant Biol* 55:591–628
- Long SP, Ainsworth EA, Leakey ADB, Morgan PB (2005) Global food insecurity. Treatment of major food crops with elevated carbon dioxide or ozone under large-scale fully open-air conditions suggests recent models may have overestimated future yields. *Phil Trans R Soc B* 360:2011–2022
- Luo Y, Su B, Currie WS et al (2004) Progressive nitrogen limitation of ecosystem responses to rising atmospheric carbon dioxide. *Bioscience* 54(8):731–739
- Mander Ü, Löhmus K, Teiter S et al (2008) Gaseous nitrogen and carbon fluxes in riparian alder stands. *Boreal Env Res* 13:231–241
- Mander Ü, Maddison M, Soosaar K et al (2015) The impact of a pulsing groundwater table on greenhouse gas emissions in riparian grey alder stands. *Environ Sci Pollut Res* 22:2360–2371
- Manning WJ, Godzik B (2004) Bioindicator plants for ambient ozone in central and Eastern Europe. *Environ Pollut* 130:33–39
- Manning WJ, Godzik B, Musselman RM (2002) Potential bioindicator plant species for ambient ozone in forested mountain areas of central Europe. *Environ Pollut* 119:283–290
- Markham JH, Chanway CP (1999) Does past contact reduce the degree of mutualism in the *Alnus rubra*-*Frankia* symbiosis? *Can J Bot* 77:434–441
- Matyssek R, Bytnerowicz A, Karlsson P-E et al (2007) Promoting the O₃ flux concept for European forest trees. *Environ Pollut* 146:587–607
- Meehan TD, Lindroth RL (2007) Modeling nitrogen flux by larval insect herbivores from a temperate hardwood forest. *Oecologia* 153:833–843
- Millett J, Godbold D, Smith AR, Grant H (2012) N₂ fixation and cycling in *Alnus glutinosa*, *Betula pendula* and *Fagus sylvatica* woodland exposed to free air CO₂ enrichment. *Oecologia* 169:541–552
- Mills G, Buse A, Gimeno B et al (2007) A synthesis of AOT40-based response functions and critical level of ozone for agricultural and horticultural crops. *Atmos Environ* 41:2630–2643
- Moiroud A, Capellano A (1979) Etude de la dynamique de l'azote à haute altitude. I. Fixation d'azote (réduction de l'acétylène) par *Alnus viridis*. *Can J Bot* 57:1979–1985
- Morgan PB, Ainsworth EA, Long SP (2003) How does elevated ozone impact soybean? A meta-analysis of photosynthesis, growth and yield. *Plant Cell Environ* 26:1317–1328
- Mortensen LM, Skre O (1990) Effects of low ozone concentrations on growth of *Betula pubescens* Ehrh., *Betula verrucosa* Ehrh. and *Alnus incana* (L.) Moench. *New Phytol* 115:165–170
- Myrold DD, Huss-Danell K (2003) Alder and lupine enhance nitrogen cycling in a degraded forest soil in Northern Sweden. *Plant Soil* 254:47–56
- Newton M, Hassen BAE, Zavitkovski J (1968) Role of red alder in western forest succession. In: Trappe JM, Franklin JF, Tarrant RF, Hansen GH (eds) *Biology of alder*. USFS Pacific Northwest Forest and Range Experiment Station, Portland, OR, pp 73–83
- Noh NJ, Son Y, Koo JW et al (2010) Comparison of nitrogen fixation for north- and south-facing *Robinia pseudoacacia* stands in central Korea. *J Plant Biol* 53:61–69
- Norby RJ (1987) Nodulation and nitrogenase activity in nitrogen-fixing woody plants stimulated by CO₂ enrichment of the atmosphere. *Physiol Plant* 71:77–82
- Norby R, Zak DR (2011) Ecological lessons from free-air CO₂ Enrichment (FACE) experiments. *Annu Rev Ecol Evol Syst* 42:181–203
- Norby RJ, Warren JM, Iversen CM et al (2010) CO₂ enhancement of forest productivity constrained by limited nitrogen availability. *PNAS* 107(45):19368–19373
- Nord EA, Lynch JP (2009) Plant phenology: a critical controller of soil resource acquisition. *J Experiment Bot* 60(7):1927–1937
- Normand P (2013) A brief history of *Frankia* and actinorhizal plants meetings. *J Bioscience* 38:677–684
- Pandey R, Zinta G, Abdelgawad H et al (2015) Physiological and molecular alterations in plants exposed to high [CO₂] under phosphorus stress. *Biotech Advances* 33:303–316
- Pawlowski N, Newton WE (2008) *Nitrogen-fixing actinorhizal symbioses*. Springer, Dordrecht
- Pawlowski N, Sprent JI (2008) Comparison between actinorhizal and legume symbiosis. In: Pawlowski K, Newton WE (eds) *Nitrogen-fixing actinorhizal symbioses*. Springer, Dordrecht, pp 261–288
- Pezeshki SR, Hinkley TM (1988) The water relations characteristics of *Alnus rubra* and *Populus trichocarpa*: responses to field drought. *Can J For Res* 18:1159–1166
- Pokharel A, Mirza BS, Dawson JO, Hahn D (2011) *Frankia* populations in soil and root nodules of sympatrically grown *Alnus* taxa. *Microb Ecol* 61:92–100
- Pöhlme S, Bahram M, Köljalg U, Tedersoo L (2014) Global biogeography of *Alnus*-associated *Frankia* actinobacterial. *New Phytol* 204:979–988

- Poorter H, Navas ML (2003) Plant growth and competition at elevated CO₂: on winners, losers and functional groups. *New Phytol* 157:175–198
- Pourhassan N, Wichard T, Roy S, Bellenger JP (2015) Impact of elevated CO₂ on metal homeostasis and the actinorhizal symbiosis in early successional alder shrubs. *Environ Exp Bot* 109:168–176
- Reverchon F, Xu Z, Blumfield TJ et al (2012) Impact of global change and fire on the occurrence and function of understory legumes in forest ecosystems. *J Soil Sediments* 12:150–160
- Rhoades C, Oskarsson H, Binkley D, Stottleyer B (2001) Alder (*Alnus crispa*) effects on soils in ecosystems of the Agashashok River valley, northwest Alaska. *Ecoscience* 8:89–95
- Rogers A, Ainsworth EA, Leakey ADB (2009) Will elevated carbon dioxide concentration amplify the benefits of nitrogen fixation in legumes? *Plant Physiol* 131:1009–1016
- Roggy JC, Moiroud A, Lensi R, Domenach AM (2004) Estimating N transfers between N₂-fixing actinorhizal species and the non-N₂-fixing *Prunus avium* under partially controlled conditions. *Biol Fertil Soils* 39:312–319
- Ruess RW, Anderson MD, Mitchell JS, McFarland JW (2006) Effects of defoliation on growth and N fixation in *Alnus tenuifolia*: consequences for changing disturbance regimes at high latitudes. *Ecoscience* 13:404–412
- Ruess RW, Anderson MD, McFarland JM et al (2013) Ecosystem-level consequences of symbionts partnerships in an N-fixing shrub from interior Alaskan floodplains. *Ecol Monog* 83:177–194
- Rytter L (1989) Distribution of roots and root nodules and biomass allocation in young intensively managed gray alder stands on a peat bog. *Plant Soil* 119:71–79
- Rytter L, Arveby AS, Granhall U (1991) Dinitrogen (C₂H₂) fixation in relation to nitrogen fertilization of grey alder [*Alnus incana* (L.) Moench.] plantations in a peat bog. *Biol Fertil Soils* 10:233–240
- Sanborn P, Preston C, Brockley R (2002) N₂-fixation by Sitka alder in a young lodgepole pine stand in central interior British Columbia, Canada. *For Ecol Manage* 167: 223–231
- Sardans J, Peñuelas J (2012) The role of plants in the effects of global change on nutrient availability and stoichiometry in the plant-soil system. *Plant Physiol* 160:1741–1761
- Schleppi P, Bucher-Wallin I, Hagedorn F, Körner C (2012) Increased nitrate availability in the soil of mixed mature temperate forest subjected to elevated CO₂ concentration (canopy FACE). *Global Change Biol* 18:757–768
- Schwintzer CR, Tjepkema JD (1997) Field nodules of *Alnus incana* ssp. *rugosa* and *Myrica gale* exhibit pronounced acetylene-induced declines in nitrogenase activity. *Can J Bot* 75:1415–1423
- Scullion J, Smith AR, Gwynn-Jones D et al (2014) Deciduous woodland exposed to elevated atmospheric CO₂ has species-specific impact on anecic earthworms. *Appl Soil Ecol* 80: 84–92
- Seeds JD, Bishop JG (2009) Low *Frankia* inoculation potentials in primary successional sites at Mount St. Helens, Washington, USA. *Plant Soil* 323:225–233
- Seiler JR, Johnson JD (1984) Growth and acetylene reduction of black alder seedlings in response to water stress. *Can J For Res* 14:477–480
- Sharma E, Ambasht RS (1984) Seasonal variation in nitrogen fixation by different ages of root nodules of *Alnus nepalensis* plantation, in the eastern Himalayas. *J Appl Ecol* 21:265–270
- Sharma E, Ambasht RS (1986) Root nodule age-class transition, production and decomposition in an age sequence of *Alnus nepalensis* plantation stands in the eastern Himalayas. *J Appl Ecol* 23:689–701
- Sharma E, Ambasht RS (1988) Nitrogen accretion and its energetics in the Himalayan alder. *Funct Ecol* 2:229–235
- Sharma G, Sharma R, Sharma E, Singh KK (2002) Performance of age series of *Alnus*-cardamom plantation in the Sikkim Himalaya: nutrient dynamics. *Ann Bot* 89:273–282
- Sharma G, Sharma R, Sharma E (2008) Influence of stand age on nutrient and energy release through decomposition in alder-cardamom agroforestry systems of the Eastern Himalayas. *Ecol Res* 23:99–106
- Sharma G, Sharma R, Sharma E (2010) Impact of altitudinal gradients on energetics and efficiencies of N₂-fixation in alder-cardamom agroforestry systems of the eastern Himalayas. *Ecol Res* 25:1–12
- Sicher RC, Barnaby JY (2012) Impact of carbon dioxide enrichment on the responses of maize leaf transcripts and metabolites to water stress. *Physiol Plant* 144:238–253
- Sigurdsson BD, Medhurst JL, Wallin G et al (2013) Growth of mature boreal Norway spruce was not affected by elevated [CO₂] and/or air temperature unless nutrient availability was improved. *Tree Physiol* 33:1192–1205
- Silvester WB, Winship LJ (1990) Transient responses of nitrogenase to acetylene and oxygen by actinorhizal nodules and cultured *Frankia*. *Plant Physiol* 91:480–486
- Silvester WB, Berg RH, Schwintzer CR, Tjepkema JD (2008) Oxygen responses, hemoglobin, and the structure and function of vesicles. In: Pawlowski K, Newton WE (eds) *Nitrogen-fixing Actinorhizal symbioses*. Springer, Dordrecht, pp 105–146
- Simard SW, Radosevich SR, Sachs DL, Hagerman SM (2006) Evidence for competition and facilitation trade-offs: effects of Sitka alder density on pine regeneration and soil productivity. *Can J For Res* 36:1286–1298
- Smith AR, Lukac M, Bambrick M et al (2013a) Tree species diversity interacts with elevated CO₂ to induce a greater root system response. *Global Change Biol* 19:217–228
- Smith AR, Lukac M, Hood R et al (2013b) Elevated CO₂ enrichment induces a differential biomass response in a mixed species temperate forest plantation. *New Phytol* 198:156–168
- Son Y, Lee YY, Lee CY, Yi MJ (2007) Nitrogen fixation, soil nitrogen availability, and biomass in pure and mixed plantations of alder and pine in central Korea. *J Plant Nutri* 30:1841–1853
- Stöcklin J, Körner CH (1999) Interactive effects of elevated CO₂, P availability and legume presence on calcareous grassland: results of a glasshouse experiment. *Funct Ecol* 13:200–209
- Tadaki Y, Mori H, Mori S (1987) Studies on the production structure of forests (XX) Primary productivity of a young alder stand. *J J For Soc* 69:207–214 (in Japanese)
- Takeda H (1998) Decomposition processes of litter along a latitudinal gradient. In: Sassa K (ed) *Environmental forest science*. Kluwer, Dordrecht, pp 197–206
- Tateno M (2003) Benefit to N₂-fixing alder of extending growth period at the cost of leaf nitrogen loss without resorption. *Oecologia* 137:338–343
- Tateno R, Tokuchi N, Yamanaka N et al (2007) Comparison of litterfall production and leaf litter decomposition between an exotic black locust plantation and an indigenous oak forest near Yan'an on the Loess Plateau, China. *For Ecol Manage* 241:84–90
- Temperton VM, Grayston SJ, Jackson G et al (2003a) Effects of elevated carbon dioxide concentration on growth and nitrogen fixation in *Alnus glutinosa* in a long-term field experiment. *Tree Physiol* 23:1051–1059
- Temperton VM, Millard P, Jarvis PG (2003b) Does elevated atmospheric carbon dioxide affect internal nitrogen allocation in the temperate trees *Alnus glutinosa* and *Pinus sylvestris*. *Global Change Biol* 9:286–294
- Thomas RB, Bashkin MA, Richter DD (2000) Nitrogen inhibition of nodulation and N₂ fixation of a tropical N₂-fixing tree (*Gliricidia*

- sepium*) grown in elevated atmospheric CO₂. *New Phytol* 145:233–243
- Tissue DT, Megonigal JP, Thomas RB (1997) Nitrogenase activities and N₂ fixation are stimulated by elevated CO₂ in a tropical N₂-fixing tree. *Oecologia* 109:28–33
- Tjepkema JD, Schwintzer CR, Monz CA (1988) Time course of acetylene reduction in nodules of five actinorhizal genera. *Plant Physiol* 86:581–583
- Tobita H, Kitao M, Koike T, Maruyama Y (2005) Effects of elevated CO₂ and nitrogen availability on nodulation of *Alnus hirsuta* Turcz. *Phyton* 45:125–131
- Tobita H, Uemura A, Kitao M et al (2008) The effects of elevated CO₂, low phosphorus supply, and drought on photosynthetic activity of *Alnus hirsuta* (Turcz.). *Trans Mtg Hokkaido Br Jpn For Soc* 56:43–45 (in Japanese)
- Tobita H, Hasegawa SF, Tian X et al (2010a) Spatial distribution and biomass of root nodules in a naturally regenerated stand of *Alnus hirsuta* (Turcz.) var. *sibirica*. *Symbiosis* 50:77–86
- Tobita H, Uemura A, Kitao M et al (2010b) Interactive effects of elevated CO₂, phosphorus deficiency, and soil drought on nodulation and nitrogenase activity in *Alnus hirsuta* and *Alnus maximowiczii*. *Symbiosis* 50:59–69
- Tobita H, Uemura A, Kitao M et al (2011) Effects of elevated [CO₂] and soil nutrients and water conditions on photosynthetic and growth responses of *Alnus hirsuta*. *Funct Plant Biol* 38:702–710
- Tobita H, Hasegawa SF, Yazaki K et al (2013a) Growth and N₂ fixation in an *Alnus hirsuta* (Turcz.) var. *sibirica* stand in Japan. *J Biosci* 38(4):761–776
- Tobita H, Kucho K, Yamanaka T (2013b) Abiotic factors influencing nitrogen-fixing actinorhizal symbioses. In: Ricardo A (ed) *Symbiotic endophytes*. Springer, Berlin Heidelberg, pp 103–122
- Tobita H, Nanami S, Hasegawa SF et al (2015) Spatial distribution of regenerated woody plants in *Alnus hirsuta* (Turcz.) var. *sibirica* stand in Japan. *Open J For* 5:210–220
- Tripp LN, Bezdicsek DF, Heilman PE (1979) Seasonal and diurnal patterns and rates of nitrogen fixation by young red alder. *Forest Sci* 25: 371–380
- Tromas A, Diagne N, Diedhiou I et al (2013) Establishment of actinorhizal symbioses. In: Ricardo A (ed) *Symbiotic Endophytes*. Springer, Berlin Heidelberg, pp 89–101
- Tsutsumi H, Nakatsubo T, Ino Y (1993) Field measurements of nitrogen-fixing activity of intact saplings of *Alnus maximowiczii* in the subalpine zone of Mt Fuji. *Ecol Res* 8:85–92
- Uemura S, Sato T (1975) Non-leguminous root nodules in Japan (a supplementary report). In: Takahashi H (ed) *Nitrogen fixation and nitrogen cycle*. JIBP Synthesis. Univ of Tokyo Press, Tokyo, pp 17–24
- Uemura A, Tobita H, Kitaoka S, Utsugi H (2009) Effects of high CO₂ concentration on water relations of two *Alnus* species. *Trans Mtg Hokkaido Br Jpn For Soc* 57:195–197 (in Japanese)
- Uliassi DD, Ruess RW (2002) Limitation to symbiotic nitrogen fixation in primary succession on the Tanana river floodplain. *Ecology* 83:88–103
- Uliassi DD, Huss-Danell K, Ruess RW, Doran K (2000) Biomass allocation and nitrogenase activity in *Alnus tenuifolia*: responses to successional soil type and phosphorus availability. *Ecoscience* 7:73–79
- Urgiles N, Strauss A, Lojan P, Schussler A (2014) Cultured arbuscular mycorrhizal fungi and native soil inocula improve seedling development of two pioneer trees in the Andean region. *New For* 45:859–874
- Uri V, Löhmus K, Tullus H (2004) The budget of demand for nitrogen in grey alder (*Alnus incana* (L.) Moench) plantation on abandoned agricultural land in Estonia. *Balt For* 10:12–18
- Uri V, Löhmus K, Mander Ü et al (2011) Long-term effects on the nitrogen budget of a short-rotation grey alder (*Alnus incana* (L.) Moench) forest on abandoned agricultural land. *Ecol Eng* 37:920–930
- Uri V, Aosaar J, Varik M et al (2014) The dynamics of biomass production, carbon and nitrogen accumulation in grey alder (*Alnus incana* (L.) Moench) chronosequence stands in Estonia. *For Ecol Manage* 327:106–117
- Valdés M (2008) *Frankia* ecology. In: Pawlowski K, Newton WE (eds) *Nitrogen-fixing actinorhizal symbioses*. Springer, Dordrecht, pp 49–72
- Valverde C, Ferrari A, Wall LG (2002) Phosphorous and the regulation of nodulation in the actinorhizal symbiosis between *Discaria trinervis* (Rhamnaceae) and *Frankia* BCU110501. *New Phytol* 153:43–52
- VanderHeyden D, Skelly J, Innes J et al (2001) Ozone exposure thresholds and foliar injury on forest plants in Switzerland. *Environ Pollut* 111:321–331
- Vitousek PM, Howarth RW (1991) Nitrogen limitation on land and in the sea: how can it occur? *Biogeochemistry* 13:87–115
- Vitousek PM, Walker LR (1987) Colonization, succession and resource availability: ecosystem-level interactions. In: Gray AJ, Crawley MJ, Edwards PJ (eds) *Colonization, succession and stability*. Blackwell Scientific, Oxford, pp 207–223
- Vitousek PM, Cassman K, Cleveland C et al (2002) Towards and ecological understanding of biological nitrogen fixation. *Biogeochemistry* 57:1–45
- Vogel CS, Curtis PS (1995) Leaf gas exchange and nitrogen dynamics of N₂-fixing, field-grown *Alnus glutinosa* under elevated atmospheric CO₂. *Global Change Biol* 1:55–61
- Vogel JG, Gower ST (1998) Carbon and nitrogen dynamics of boreal jack pine stands with and without a green alder understory. *Ecosystems* 1:386–400
- Vogel CS, Curtis PS, Thomas RB (1997) Growth and nitrogen accretion of dinitrogen-fixing *Alnus glutinosa* (L.) Gaertn. under elevated carbon dioxide. *Plant Ecol* 130:63–70
- Voigt GK, Steucek GL (1969) Nitrogen distribution and accretion in an alder ecosystem. *Soil Sci Soc Am* 33:946–949
- Wall LG, Berry AM (2008) Early interactions, infection and nodulation in actinorhizal symbiosis. In: Pawlowski K, Newton WE (eds) *Nitrogen-fixing actinorhizal symbioses*. Springer, Dordrecht, pp 147–166
- Wang YP, Law RM, Pak B (2010) A global model of carbon, nitrogen and phosphorus cycles for the terrestrial biosphere. *Biogeoscience* 7:2261–2282
- Watanabe Y, Satomura T, Sasa K et al (2010) Differential anatomical responses to elevated CO₂ in saplings of four hardwood species. *Plant Cell Environ* 33:1101–1111
- Winship LJ, Tjepkema JD (1990) Techniques for measuring nitrogenase activity in *Frankia* and actinorhizal plants. In: Schwintzer CR, Tjepkema JD (eds) *The biology of Frankia and actinorhizal plants*. Academic Press Inc, Tokyo, pp 264–280
- Wittig VE, Ainsworth EA, Naidu SL et al (2009) Quantifying the impact of current and future tropospheric ozone on tree biomass, growth, physiology and biochemistry: a quantitative meta-analysis. *Global Change Biol* 15:396–424
- Wurtz TL (1995) Understory alder in three boreal forests of Alaska: local distribution and effects on soil fertility. *Can J For Res* 25:987–996
- Xu Z, Shimizu H, Yagasaki Y et al (2013) Interactive effects of elevated CO₂, drought, and warming on plants. *J Plant Growth Regul* 32:692–707
- Yamanaka T, Li CY, Bormann BT, Okabe H (2003) Tripartite associations in an alder: effects of *Frankia* and *Alpova*

- diplophloeus* on the growth, nitrogen fixation and mineral acquisition of *Alnus tenuifolia*. Plant Soil 254:179–186
- Yoon TK, Noh NJ, Han S et al. (2014) Soil moisture effects on leaf litter decomposition and soil carbon dioxide efflux in wetland and upland forests. Soil Sci Soc Am J 78:1804–1816
- Younger PD, Kapustka LA (1983) N₂ (C₂H₂) ase activity by *Alnus incana* ssp. *rugosa* (Betulaceae) in the northern hardwood forest. Am J Bot 70:30–39
- Zak DR, Pregitzer KS, Kubiske ME, Burton AJ (2011) Forest productivity under elevated CO₂ and O₃: positive feedbacks to soil N cycling sustain decade-long net primary productivity enhancement by CO₂. Ecol Lett 14:1220–1226
- Zavitkovski J, Newton M (1968) Effect of organic matter and combined nitrogen on nodulation and nitrogen fixation in red alder. In: Trappe JM, Franklin JF, Tarrant RF, Hansen GH (eds) Biology of alder. USFS Pacific Northwest Forest and Range Experiment Station, Portland, OR, pp 209–223
- Zhang X, Sigman DM, Morel FMM, Kraepiel AML (2014) Nitrogen isotope fractionation by alternative nitrogenases and past ocean anoxia. PNAS 111:4782–4787