Original article

Renal tubular function in β -thalassemia

Achra Sumboonnanonda1, Prida Malasit2, Voravarn S. Tanphaichitr1, Sompong Ong-ajyooth3, Sunthorn Sunthornchart¹, Sa-nga Pattanakitsakul⁴, Siripan Petrarat¹, Amara Assateerawatt¹, and Arun Vongjirad¹

¹ Department of Pediatrics, Siriraj Hospital, Mahidol University, Prannok Road, Bangkok 10700, Thailand

² Department of Medicine, Siriraj Hospital, Mahidol University, Prannok Road, Bangkok 10700, Thailand

³ Department of Biochemistry, Siriraj Hospital, Mahidol University, Prannok Road, Bangkok 10700, Thailand

⁴ Office for Research and Development, Siriraj Hospital, Mahidol University, Prannok Road, Bangkok 10700, Thailand

Received April 5, 1997; received in revised form November 6, 1997; accepted November 12, 1997

Abstract. Studies of the renal involvement in thalassemic syndromes have been varied and few. This study was designed to define the renal abnormalities associated with b-thalassemia and to correlate the renal findings with clinical parameters. One hundred and four β -thalassemic children with various disease severity were studied. The patients were divided into three groups: 48 with severe anemia [hematocrit (Hct) $<$ 25%], 31 on a hypertransfusion program and desferrioxamine treatment, and 25 with moderate anemia (Hct $>$ 25%). The results were compared with 15 normal children. Significantly higher levels of proteinuria and low molecular weight proteinuria were found in all patients compared with normal children. Aminoaciduria was detected in one-third of patients. Thalassemic patients had significantly lower morning urine osmolarity, higher urine N -acetyl- β -D-glucoseminidase and malondialdehyde (MDA, an indicator of lipid peroxidation). Patients with severe anemia had significantly higher low-molecular weight proteinuria and MDA, and lower urine osmolarity than those with moderate anemia. Our data confirmed the high frequency of renal abnormalities in b-thalassemia patients and indicated some degree of proximal tubular dysfunction. Severity of the abnormalities correlated with the degree of anemia and were least severe in patients on hypertransfusion and desferrioxamine therapy. This suggested that the damage might be caused by anemia and increased oxidation induced by excess iron deposits.

Key words: β -Thalassemia – Hemoglobin E – Renal tubular function $-$ Low molecular weight proteinuria $-$ Malondialdehyde - Urine osmolarity

Introduction

Thalassemia hemoglobinopathies are prevalent in Thailand. The frequencies of β -thalassemia (thal) and hemoglobin (Hb) E, β -globin chain variant, are $3\% - 9\%$ and 13% $-60%$, respectively [1]. The disease forms of β -thal are homozygous b-thal and b-thal/HbE. Half of the homozygous B-thal patients die before the age of 12 years due to severe infection, anemia, and multiple organ failure $[2-4]$. The abnormal synthesis of the globin chains or the production of abnormal Hb have wide impacts on the physiological functions of virtually every major organ. Patients with thal are known to have dysfunctions of the cardiopulmonary, reticuloendothelial, and other major systems $[2-7]$. Abnormalities of renal function have also been reported, but there is no systematic study of the prevalence and type of renal involvement. Various glomerular pathologies have been sporadically reported, and it is still unknown whether those abnormalities are genuinely associated with the thalassemic syndromes [3, 6, 8, 9]. One study of ten patients with Cooley's anemia suggested some abnormalities associated with renal medulla [10]. Renal tubular acidosis has also been reported in patients with thal [10, 11]. The purpose of this study was to systemically investigate the prevalence and nature of renal abnormalities in 104 patients with β -thal.

Patients and methods

Patients aged between 3 and 15 years with homozygous β -thal or b-thal/HbE diseases attending the Department of Pediatrics, Siriraj Hospital from November 1993 through October 1994 were recruited into the study with informed consent from their parents. Patients with acute febrile illness were excluded. The diagnosis of β -thal was made by standard methods [12]. Height and weight standard deviation scores (SDS) were calculated according to Tanner et al. [13].

Patients were instructed to fast overnight before attending the clinic in the morning. Blood samples were collected from each patient for Hb and hematocrit (Hct), urea nitrogen, creatinine, and electrolytes. Fresh second-morning urine samples were collected; all samples were immediately aliquoted and frozen until further analysis. The remaining

Correspondence to: A. Sumboonnanonda

Table 1. Summary of demographic and biochemical data of three groups of patients and controls^a

SDS, Standard deviation score; HCO₃, bicarbonate; N, nitrogen; Cr, creatinine; LMW, low molecular weight; NAG, N-acetyl-β-D-glucosaminidase; MDA, malondialdehydr

 $*P < 0.05$; $*P < 0.001$

^a Values are mean \pm standard error

^b As calculated by Schwartz's formula: $0.55 \times$ height (cm)/plasma Cr (mg/dl)

^c Group A consists of patients with severe anemia; group B are on hypertransfusion with desferrioxamine treat; group C are those with mild anemia

urine was tested for osmolarity, protein, sugar (by Labstix, Bayer Diagnostics) and examined microscopically. N-Acetyl-β-_{D-glucosami}nidase (NAG) (by a spectrophotometric method [14], creatinine, (Jaffe reaction, autoanalyzer), and amino acids (by paper chromatography [15] were measured. Urine malondialdehyde (MDA) was measured by a spectrophotometric technique described by Knight et al. [16]. Urinary protein was assayed by a modified Bradford method [17] (Bio-Rad Laboratories, Richmond Calif., USA). The urine was electrophoresed in $8\% - 15\%$ polyacrylamide gels and silver stained [18]. The stained gels were scanned in a flat-bed scanner, and the amount of low molecular weight (LMW) proteins was calculated from the proportion of the protein bands smaller than 45 kilodaltons [19]. Fresh morning urine from 15 healthy children of the same age group were used as controls.

Statistical methods. All calculations were carried out using Statview statistical package (Abacus Concepts, USA). Comparison between groups was performed using unpaired Student's t-test and in categorical data using the chi-squared test. A P value of less than 0.05 was regarded as significant.

Results

One hundred and four patients comprising 27 homozygous b-thal and 77 b-thal/HbE patients were included in this study. Mean age was 9.7 years with a range of $3-15$ years. Thirty patients were splenectomized (mean age 10.7 years vs. 9.3 years of those with intact spleens). The patients were divided into three groups. Group A included 48 patients with severe anemia (Hct less than 25%), group B (31 patients) were on a hypertransfusion program (maintaining the Hct at around 30% at all times) and desferrioxamine treatment (subcutaneous injection of $20 - 40$ mg/ kg per dose, $2-5$ doses/week), and 25 patients of group C who had a Hct $>25\%$ and were not on desferrioxamine. The duration of desferrioxamine treatment 3.7 ± 2.3 years (mean \pm SD). The criteria for selection of patients for desferrioxamine and hypertransfusion treatments were not based on clinical severity, but on their financial status. Table 1 summarizes the demographic data of patients and controls. There was no significant difference in the proportion of patients with and without splenectomy amongst the three groups. None of the patients were positive for human immunodeficiency virus antibody and 2 patients were positive for hepatitis B surface antigen antigen.

Forty-one patients (39%) had a body weight \lt 3rd percentile for age. Height and weight SDS are shown in Table 1. Blood urea nitrogen and serum creatinine were within normal limits in all except 1 patient from group B, who had a serum creatinine of 1.1 mg/dl (estimated glomerular filtration rate (GFR) using Schwartz's formula [20] 69 ml/min per 1.73 m2). Using Schwartz's formula to estimate the GFR, all groups had values within the normal limit $(89 - 165 \text{ ml/min per } 1.73 \text{ m}^2)$. Serum electrolytes were all within normal limits.

Urine osmolarity in all groups except group B was lower than normal controls ($P = 0.0005$ for group A and $P = 0.02$) for group C). Group A had the lowest urine osmolarity (Table 1). The urine urea level was lower than controls in all groups. Splenectomy did not affect urine osmolarity $(P = 0.08)$. Urine protein and sugar by Labstix were negative. Microscopic hematuria (10 red blood cells per highpower field) was also found in 8 patients (5 in group A, 2 in B, and 1 in C), but no renal casts were found. Generalized aminoaciduria was found in 32 of 102 patients. No difference in the proportion of patients with aminoaciduria was found amongst the three groups (chi-squared, $P = 0.6559$).

All three groups had significantly higher levels of urine NAG than controls ($P < 0.05$), but no difference was found amongst patients. The LMW proteinuria/creatinine in groups A, B, and C was significantly higher than controls $(P = 0.0002, 0.0390, 0.0062,$ respectively). Severely anemic (A) and mildly anemic (C) groups had significantly higher total urinary protein/creatinine ratios $(P = 0.0017$ and

0.0398), but there was no difference between group B and controls ($P = 0.0595$). There was no evidence of glomerular proteinuria by electrophoresis. Urine MDA in groups A and C was significantly higher than controls $(P < 0.0001$ and 0.0003), but there was no difference between group B and controls $(P = 0.2132)$ (Table 1).

Discussion

We report a high frequency of renal abnormalities in a group of β -thal patients with disease of varying severity. The key abnormalities included: increased levels of proteinuria, especially LMW fraction, aminoaciduria, increased urinary NAG and MDA. Urine osmolarity was significantly lower in patients than controls. The data suggest some degree of proximal tubular dysfunction. Increased urine NAG [21, 22], aminoaciduria [23], and LMW proteins $[24 - 27]$ are indicators of proximal tubular damage.

When patients were divided into three groups according to their clinical severity, patients receiving multiple blood transfusions and desferrioxamine treatment (group B) had lower levels of LMW proteinuria, urinary MDA, and higher urine osmolarity. The most pronounced abnormalities were found in group A, the group with the most-severe anemia (Hct $<$ 25%). MDA, the end product of lipid peroxidation, was highest in groups A and C, and correlated well with the clinical classification. This suggested that oxidative stress might be an important factor responsible for the damage. In thal, the imbalanced synthesis of Hb leads to the presence of excess unpaired globin chains and a high intracellular non-Hb iron content. The unstable Hb subunits generate free oxygen radicals, which start a chain of oxidative events leading to disintegration to denatured globin chains, heme and iron which bind to different membrane proteins, altering their normal structure and function [28]. The excess free iron is a catalyst of lipid peroxidation via participation in the Fenton reaction [29, 30]. The high urine MDA in groups A and C, and the normal levels in group B supports the hypothesis that lipid peroxidation occurs in untreated groups and can be reduced or reversed by desferrioxamine treatment [31]. The reduction of urinary MDA and milder renal manifestations in group B may also be due to the direct suppressive effect of desferrioxamine on peroxidation [32, 33]. Group B responded favorably to desferrioxamine treatment and frequent transfusions, despite high levels of serum ferritin. The latter indicates that the treatment given to this group was not adequate, i. e., not able to significantly reduce tissue iron. The improvement in renal function in group B might be due to the direct suppressive effect of desferrioxamine on peroxidation [28]. This is significant, since a comprehensive program of desferrioxamine chelation is not usually achieved in developing countries, due to economic constraints.

We found no detrimental effects of desferrioxamine on renal function, as reported by Koren et al. [34] and Cianciulli et al. [35]. The desferrioxamine dosage in our study is lower than in others in which nephrotoxicity has been reported. A decrease in the ability to concentrate urine was correlated with clinical severity. The group with severe

anemia (A) had the lowest urine osmolarity after overnight fasting and the patients on desferrioxamine and hypertransfusion (B) had the highest urine osmolarities, which were not statistically different from controls (Table 1). These data are in accordance with those reported by others $[6, 10-12]$. The cause of this defect is not known. However, it is possible that malnutrition might play a role [36, 37], since two-thirds of the patients in group A were underweight. Paniagua et al. [38] demonstrated that malnutrition by itself did not cause the abnormalities, but other insults, such as electrolyte imbalances and infections, were responsible for the defects. All three groups had lower levels of urine urea (which generally reflects the degree of malnourishment) than controls, but patients in group B had a comparable ability to concentrate urine. This indicates that malnutrition is not the key factor leading to abnormal concentrating ability. Another possible contributory factor is the hyperperfusing effect of anemia [6]. The estimated GFR in groups A and C was high, although within normal limits; these findings support the above hypothesis. The fact that urine osmolarity was higher in patients on iron chelation suggests that the concentration defect is reversible and deposited iron and anemia might play a role in the pathogenesis of the defect.

Our findings are in accordance with previous studies in thal patients. Two studies have identified some degree of medullary fibrosis and suggested that the pathology might contribute to the abnormal concentrating ability [6, 10]. Generalized aminoaciduria was also reported by Hyman et al. [39].

In conclusion, our data indicate that there are proximal tubular dysfunctions in patients with homozygous B-thal and β -thal/HbE disease. The cause of this dysfunction is not known, but anemia and iron deposition may be key factors. The role of desferrioxamine as a protective agent for tubular damage is also suggested by our study.

Acknowledgements. This study was funded by the China Medical Board, Siriraj Hospital, Thailand (grant no. 75-348-212). P. Malasit is supported by the Senior Research Scholar Grant (no. RTA/04/2539) from the Thailand Research Fund.

References

- 1. Wasi P (1983) Hemoglobinopathies in Southeast Asia. In: Bowman JF (ed) Distribution and evolution of hemoglobin and globin loci. Elsevier, New York, pp 179-208
- 2. McDonagh KT, Neinhuis AW (1993) The thalassemias. In: Nathan DG, Oski FA (eds) Hematology of infancy and childhood, 4th edn. Saunders, Philadelphia, pp 783-879
- 3. Bhamarapravati N, Na-Nakorn S, Wasi P, Tuchinda S (1967) Pathology of abnormal hemoglobin diseases seen in Thailand. I. Pathology of b-thalassemia hemoglobin E disease. Am J Clin Pathol 47: 745-758
- 4. Issaragrisil S, Wanachiwanawin W, Bhuripango K, Benjasuratwong Y, Piankijagum A, Wasi P (1988) Infection in thalassemia: a retrospective study of 1,018 patients with β -thalassemia/HbE disease. Birth Defects 23: 505-511
- 5. Boonpucknavig S, Chiewsilp P, Isarangkura P, O'Charoen R, Akkawat R (1988) Immunologic reactivity in thalassemia. Birth Defects 23: 565 - 569
- 6. Landing BH, Gonick HC, Nadorra RL, Hyman CB, Wells TR, Villarreal-Engelhardt G, Mersch J, Agness CL (1989) Renal le-

sions and clinical findings in thalassemia major and other chronic anemias with hemosiderosis. Pediatr Pathol 9: 479-500

- 7. Wasi P (1971) Streptococcal infection leading to cardiac and renal involvement in thalassemia. Lancet I: 949-950
- 8. Ongajyooth L, Pootrakul P, Malasit P, et al. (1995) Glomerulonephritis in β-thalassemia Hb-E disease: clinical manifestation, histopathologic studies and outcome. J Med Assoc Thai 78: $119 - 126$
- 9. Sonakul D, Pacharee P, Thakerngpol K (1988) Pathological findings in 76 autopsy cases of thalassemia. Birth Defects 23: $157 - 176$
- 10. Mastrangelo F, Lopez T, Rizzelli S, Manisco G, Corliano C, Alfonso L (1975) Function of the kidney in adult patients with Cooley's disease. Nephron 14: 229-236
- 11. Shehab M, Barakat AY (1985) Thalassemia B with distal renal tubular acidosis: a previously undescribed association. Int J Pediatr Nephrol 6: 143-144
- 12. Tanphaichitr VS, Mahasandana C, Suvatte V, Yodthong S, Pungamritt P, Seeloem J (1995) Prevalence of hemoglobin E, alphathalassemia and glucose-6-phosphate dehydrogenase deficiency in 1,000 cord blood studies in Bangkok. Southeast Asian J Trop Med Public Health 26 [Suppl 1]: $271 - 274$
- 13. Tanner JM, Whitehouse RH, Cameron N, Marshall WA, Healy MJR, Goldstein H (1990) Assessment of skeletal maturity and prediction of adult height (TW2 method), 2nd edn. Alden, Oxford
- 14. Moore JC, Moris JE (1982) A simple automated colorimetric method for determination of N -acetyl- β -D-glucosaminidase. Ann Clin Biochem 19: 157-159
- 15. Efran ML, Young O, Moser HW, MacCready RA (1964) A simple chromatography screening test for the detection of disorder of amino acid metabolism. N Engl J Med 270: 1378-1380
- 16. Knight JA, Smith SE, Kinder VE, Pieper RK (1988) Urinary lipoperoxides quantified by liquid chromatography and determination of references values for adults. Clin Chem $34: 1107 - 1110$
- 17. Lim CW, Chisnall WN, Stokes YM, Debnam PM, Crooke MJ (1990) Effects of low and high relative molecular protein mass on four methods for total protein determination in urine. Pathology $22: 89 - 92$
- 18. Morrissey JH (1981) Silver stain for proteins in polyacrylamide gel. A modified procedure with enhanced uniform sensitivity. Anal Biochem 117: 307-310
- 19. Brockleband T, Cooper EH, Richmond K (1991) Sodium dodecyl sulphate polyacrylamide gel electrophoresis patterns of proteinuria in various renal diseases of childhood. Pediatr Nephrol 5: $371 - 375$
- 20. Schwartz GJ, Haycock GB, Edelmann CM Jr, Spitzer A (1976) A simple estimate of glomerular filtration rate in children derived from body length and plasma creatinine. Pediatrics $58: 259-263$
- 21. Price RG (1982) Urinary enzymes, nephrotoxicity and renal disease. Toxicology $23: 99-134$
- 22. Kunin CM, Chesney RW, Craig WA, England AC, De Angelis C (1978) Enzymuria as a marker of renal injury and disease: studies of N-acetyl-b-glucosaminidase in the general population and in patients with renal disease. Pediatrics $62: 751-760$
- 23. Jones DP, Chesney RW (1994) Tubular function. In: Holliday MA, Barratt TM, Avner ED, Kogan BA (eds) Pediatric nephrology, 3rd edn. Williams and Wilkins, Baltimore, pp $117 - 149$
- 24. Piscator M (1991) Early detection of tubular dysfunction. Kidney Int 40 [Suppl 34]: S15-S17
- 25. Jung K (1994) Urinary enzymes and low molecular weight proteins as markers of tubular dysfunction. Kidney Int 46: S29-S33
- 26. Maack T, Johnson V, Kau ST, Figueiredo J, Sigulem D (1979) Renal filtration, transport, and metabolism of low-molecularweight proteins: a review. Kidney Int $16: 251 - 270$
- 27. Tomlinson PA (1992) Low molecular weight proteins in children with renal disease. Pediatr Nephrol 6: $565 - 571$
- 28. Shinar E, Rachmilewitz EA (1990). Oxidative denaturation of red blood cells in thalassemia. Semin Hematol 27: 70-82
- 29. Hebbel RP (1985) Auto-oxidation and a membrane-associated ªFenton reagentº: a possible explanation for development of membrane lesions in sickle erythrocytes. Clin Haematol 14: $129 - 140$
- 30. Miniero R, Piya A, Luzzatto L, Gabutti V (1983) Vitamin E and beta thalassemia. Haematologica 68: 562-563
- 31. Link G, Athias P, Grynberg A, Pinson A, Hershko C (1989) Effect of iron loading on transmembrane potential, contraction, and automaticity of rat ventricular muscle cells in culture. J Lab Clin Med $113: 103 - 111$
- 32. Boyce NW, Holdsworth SR (1986) Hydroxyl radical mediation of immune renal injury by desferrioxamine. Kidney Int 30: 813-817
- 33. Gutteridge JMC, Richmond R, Halliwell B (1979) Inhibition of the iron-catalysed formation of OH \degree from O₂⁻ and of lipid peroxidation by desferrioxamine. Biochem J 184: 469-472
- 34. Koren G, Bentur Y, Strong D, Harvey E, Klein J, Baumal R, Spilberg SP, Freedman MH (1989) Acute changes in renal function associated with deferoxamine therapy. Am J Dis Child 143: 1077 ± 1080
- 35. Cianciulli P, Sollecito D, Sorrentino F, Forte L, Gilardi E, Massa A, Papa G, Carta S (1994) Early detection of nephrotoxic effects in thalassemic patients receiving desferrioxamine therapy. Kidney Int $46: 467 - 470$
- 36. Alleyne GAO (1967) The effect of severe protein calorie malnutrition on the renal function of Jamaican children. Pediatrics 39: $400 - 411$
- 37. Klahr S, Alleyne GAO (1973) Effects of chronic protein-calorie malnutrition on the kidney. Kidney Int 3: 129-141
- 38. Paniagua R, Santos D, Munoz R, Luengas J, Frenk S (1980) Renal function in protein-energy malnutrition. Pediatr Res 14: $1260 - 1262$
- 39. Hyman CB, Gonick HC, Agness C, Nadorra R, Landing B (1988) Effects of deferoxamine on renal function in thalassemia. Birth Defects 23: 135 - 140