POPULATION ECOLOGY – ORIGINAL RESEARCH

Population divergence in fish elemental phenotypes associated with trophic phenotypes and lake trophic state

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Abstract Studies of ecological stoichiometry typically emphasize the role of interspecific variation in body elemental content and the effects of species or family identity. Recent work suggests substantial variation in body stoichiometry can also exist within species. The importance of this variation will depend on insights into its origins and consequences at various ecological scales, including the distribution of elemental phenotypes across landscapes and their role in nutrient recycling. We investigated whether trophic divergence can produce predictable patterns of elemental phenotypes among populations of an invasive fish, the white perch (*Morone americana*), and whether elemental phenotypes predict nutrient excretion. White perch populations exhibited a gradient of trophic phenotypes associated with landscape-scale variation in lake trophic state. Perch body chemistry varied considerably among lakes (from 0.09 for $\%$ C to 0.31-fold for $\%$ P) casting doubt on the assumption of homogenous elemental phenotypes. This variation was correlated with divergence in fish body shape and other trophic traits. Elemental phenotypes covaried (r^2) up to 0.84) with lake trophic state. This covariation likely arose in contemporary time since many of these perch populations were introduced in the last century and the trophic

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state in many of the lakes has changed in the past few decades. Nutrient excretion varied extensively among populations, but was not readily related to fish body chemistry or lake trophic state. This suggests that predictable patterns of fish body composition can arise quickly through trophic specialization to lake conditions, but such elemental phenotypes may not translate to altered nutrient recycling by fish.

Keywords Ecological stoichiometry · Nutrient recycling · Intraspecific variation · Trophic state · Trophic divergence · Contemporary evolution · Eco-evolutionary dynamics

Introduction

Emphasis on the role of animals as drivers of aquatic ecosystem dynamics is typically placed on variation in traits that aid in the capture and processing of prey, partly because trophic traits contribute to trophic cascades (Carpenter and Kitchell [1996;](#page-12-0) Post et al. [2008](#page-13-0)) and adaptive radiations (Schluter [2000\)](#page-13-1). However, animals can also influence ecosystem dynamics by altering pools and fluxes of available nutrients indirectly and directly through bioturbation, translocation, nutrient sequestration, and enhanced cycling through excretion or egestion of waste (Meyer et al. [1983](#page-12-1); Vanni [2002](#page-13-2); Sereda et al. [2008](#page-13-3); Knoll et al. [2009](#page-12-2); Flecker et al. [2010;](#page-12-3) Vanni et al. [2013](#page-13-4)). Among these, consumer-driven nutrient recycling (CDNR) from excretion is one of the most direct ways animals influence the availability and flux of nitrogen (N) and phosphorus (P) (Vanni [2002](#page-13-2)), potentially altering nutrient limitation of primary production (Elser et al. [1988](#page-12-4); Vanni et al. [2006](#page-13-5); Knoll et al. [2009\)](#page-12-2). The magnitude and chemical content of CDNR is dictated by a consumer's ability to assimilate nutrients (i.e., assimilation efficiency), the balance between

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body and food elemental content, and consumption and growth rates (Elser et al. [2000;](#page-12-5) Sterner and Elser [2002](#page-13-6); Vanni et al. [2002](#page-13-7)). Consequently, there has been considerable interest in characterizing stoichiometric variation in consumer elemental content and excretion rates and these studies have revealed considerable variation among species as a consequence of divergence in consumer body structure (Vanni et al. [2002](#page-13-7); McIntyre and Flecker [2010](#page-12-6)). Recent work has revealed considerable variation in body elemental content among populations of fishes (El-Sabaawi et al. [2012a,](#page-12-7) [b](#page-12-8), [2016](#page-12-9); Boros et al. [2015](#page-12-10)), but its origins and implications remain unclear. One important implication is that stoichiometric features of fish populations could serve as pathways for eco-evolutionary dynamics where recent evolutionary changes in elemental phenotypes might lead to a feedback on ecosystem conditions through processes like CDNR (Palkovacs et al. [2009;](#page-12-11) Jeyasingh et al. [2014](#page-12-12)). Here, we consider how consumer body elemental content varies across the landscape, how this variation may be tied to recent trophic-mediated divergence driven by the lake trophic state, and what the consequences might be for CDNR and eco-evolutionary dynamics.

The relationship between excretion and body elemental content follows from mass balance accounting where excretion is positively related to nutrient ingestion and negatively related to body N or P, which reflects demand if a consumer is growing (Sterner and Elser [2002;](#page-13-6) Vanni et al. [2002](#page-13-7)). Thus, as relative body content of an element increases, its loss through excretion should decline, assuming constant diet (Sterner and Elser [2002](#page-13-6)). Excretion is also constrained by physiological rates leading to excretion scaling with body size at approximately 3/4 following metabolic theory (Brown et al. [2004;](#page-12-13) Allgeier et al. [2015](#page-12-14)). Homeostatic regulation of carbon (C), N, and P body content is typically assumed over limited timescales. Following consumption of chemically varying food, assimilated resources are used for growth or exploited during metabolic reactions resulting in nutrient excretion, but the elemental composition of tissues remain consistent. However, recognition of the full scope of stoichiometric variation among vertebrate taxa is increasing and some findings suggest the elemental composition of aquatic consumers can be quite variable, even approaching interspecific levels (Hendrixson et al. [2007;](#page-12-15) Vrede et al. [2011](#page-13-8); El-Sabaawi et al. [2012a,](#page-12-7) [b,](#page-12-8) [2016](#page-12-9); Boros et al. [2015\)](#page-12-10).

A dichotomy has been suggested where fish body N and P are dictated by either environmental (nutrients, predation pressure) or organismal trait (body size, shape) determinants (El-Sabaawi et al. [2012a](#page-12-7)). Support for organismal trait determinants has been found, especially for the effects of body size, shape, and structural variation among taxa, which may alter investment in scales, bone and muscle tissues (Tanner et al. [2000](#page-13-9); Hendrixson et al. [2007](#page-12-15); Vrede et al. [2011](#page-13-8); Boros et al. [2015](#page-12-10); El-Sabaawi et al. [2016](#page-12-9)). These tissues may have predictable elemental signatures, where carbohydrates and lipids are high in C, protein is high in N and DNA, RNA and hydroxyapatite of bone are high in P. Other studies have found weak support for trait determinants; instead, environmental conditions such as ecosystem and resource nutrient concentration or predation environment were more important (El-Sabaawi et al. [2012a,](#page-12-7) [b,](#page-12-8) [2014\)](#page-12-16). It is unlikely that environmental and organismal determinants are mutually exclusive. Environmental variation such as available habitat and the overall input and availability of nutrients can strongly affect organismal traits such as body size, shape, growth rate and prey selectivity in fishes (Svanbäck and Eklöv [2002;](#page-13-10) Turesson and Brönmark [2007](#page-13-11); Tuckett et al. [2013](#page-13-12)), perhaps influencing consumer stoichiometry.

Trophic state is one of the most important axes of environmental variation for lentic ecosystems which integrates biotic (e.g., plankton communities) and abiotic (e.g., nutrients, lake depth, littoral area) factors ultimately affecting fish traits and ecology (Tuckett et al. [2013\)](#page-13-12). Fish also influence trophic state variables via morphological and behavioral adaptations, representing a possible reciprocal feedback between biotic trophic state factors and organismal traits (Harmon et al. [2009;](#page-12-17) Bassar et al. [2010](#page-12-18)). Animals distributed across trophic state gradients may be useful study systems for examining eco-evolutionary feedbacks, including those mediated through the relationship between environment-driven organismal trait change and stoichiometry. One such study system is the white perch (*Morone americana*) in North American lakes. White perch are native to eastern North America where their range has been expanded through deliberate and accidental introductions dating back to the 1800s (Hergenrader and Bliss [1971](#page-12-19); Busch et al. [1977;](#page-12-20) Stanley and Danie [1983;](#page-13-13) Halliwell [2005](#page-12-21)). In prior work, we showed that introduced populations of white perch exhibit contemporary divergence in a suite of trophic traits (e.g., streamlined versus robust body shape, narrow versus broad gill raker spacing, etc.) related to lake trophic state (Tuckett et al. [2013](#page-13-12)), despite recent changes in lake trophic state due to cultural eutrophication.

We used white perch to examine the stoichiometric response to invasion and adaptation along this same gradient of lake trophic state and the potential for correlated divergence in trophic/elemental phenotypes to suppress/ promote nutrient stability and eco-evolutionary feedbacks. We hypothesize that lability in trophic traits contributes to divergence in elemental phenotypes. Addressing this hypotheses requires that we build upon our previous studies showing how trophic state dictates trophic phenotypes (Tuckett et al. [2013](#page-13-12)) and the importance of body elemental content to excretion (Tuckett et al. [2015\)](#page-13-14). First, we attempt to establish a relationship between trophic state

Water body	Area (ha)	Mean depth (m)	Drainage area (ha)	TP (μ g L ⁻¹)	TN (μ g L ⁻¹)	Chl-a $(mg L^{-1})$	Secchi (m)
Cold stream	1185	12.2	4167	3	166	1.6	8.1
Brewer	388	7.9	1629	8	138	4.5	5.4
Maranacook	677	9.1	5574	4	303	3.3	5.0
North	1024	$\overline{4}$	857	14	217	4.4	3.9
East	695	5.5	1121	12	304	4.0	3.5
Pushaw	1894	3.4	19487	27	418	3.3	3.2
Umbazooksus	636	2.7	4532	17	420	1.1	2.2
Hermon	187	3	24330	14	507	20.7	1.9
Unity	1040	6.7	7410	22	685	13.1	1.6
Merrymeeting	ND	ND	1526280	37	644	ND	1.4

Table 1 Limnological and chemical characteristics of study lakes and Merrymeeting Bay

TP Total phosphorus, *TN* total nitrogen, *chl-a* chlorophyll-*a* and Secchi depth is represented by 5-year summer means taken from an online database and sampling on one date within 2 weeks of fish capture. *ND* no data

and elemental phenotypes. Second, we examine whether trophic/elemental phenotypes represent a suite of correlated traits. Finally, with respect to potential eco-evolutionary feedbacks, we examine whether feedbacks might be suppressed if eutrophic environments lead to benthic phenotypes and greater fish body N or P and, alternately, whether feedbacks might be enhanced if eutrophication leads to greater nutrient excretion regardless of stoichiometric considerations.

Methods

Study sites

We quantified morphological and elemental phenotypes of nine white perch populations inhabiting freshwater lakes spanning a range of trophic state in Maine, U.S.A (Table [1](#page-2-0); see Tuckett et al. [2013](#page-13-12) for a detailed description of fish communities and study lakes). Fish were also collected at Merrymeeting Bay, a largely freshwater tidal bay, serving as an ancestral estuarine reference condition. We used archived (2004–2007) and new data (2008) regarding lake chemistry and physical attributes to quantify environmental drivers and lake trophic status. Secchi depth, chlorophyll*a*, total phosphorus (TP) were collected under U.S. Environmental Protection Agency guidelines and archived by the Maine Department of Environmental Protection and the Volunteer Lake Management Program (lakesofmaine. org). Triplicate water samples used for analysis of chlorophyll-*a* (chl-*a*) and total nutrients were taken using a 5-cmdiameter plastic tube sampled and composited through the epilimnion. Samples were transported to the lab on ice and at least one liter of lake water was filtered onto 47 mm filters (GF/F) and subsequently frozen; filters were then ground and chl-*a* was acetone extracted and absorbance was recorded using a spectrophotometer and phaeophytin corrections (APHA [2005](#page-12-22)). Water samples for analysis of total dissolved nitrogen and total dissolved phosphorus were stored in acid-washed bottles, frozen upon laboratory arrival and dual digestion procedures were used. Unlike TP which was represented by a 5-year average, total dissolved nitrogen was unavailable for years before 2008; thus, results will represent the average of 3 replicates from a single sampling date. Merrymeeting Bay indicators are represented by a single sampling date in 2008. In most analyses we use the 5-year average Secchi depth as an integrative indicator of lake trophic state because of its correlation with TN, TP, mean lake depth, and littoral area in these lakes (Tuckett et al. [2013\)](#page-13-12).

Fish capture and processing

We collected between 27 and 32 perch at each lake and measured body morphology, elemental content, growth rate, and excretion of N and P. Fish were captured between 19:00 and 23:00 h in littoral habitats from June through August 2008 using boat electrofishing. Following capture, fish were held to measure nutrient excretion rates (described below), euthanized, and then frozen at −20 °C for 3–6 months. Fish were later photographed on the left side with a uniform background at fixed focal length (60 cm), measured for standard length $(\pm 1 \text{ mm})$, and weighed $(\pm 0.1 \text{ g})$. Fish traits typically associated with trophic-mediated divergence were analyzed from digital photographs and caliper measurements, including overall fish body shape, gill raker length and spacing, width of the closed mouth between the maxillary bones, open mouth gape size, and caudal, pectoral, pelvic fin lengths using approaches described in Tuckett et al. [\(2013](#page-13-12)).

Two-dimensional white perch shape variation was analyzed with geometric morphometric procedures using 21 homologous landmarks placed on randomly ordered digital images (8.1 mps Canon Eos Rebel) using the program tpsDig Version 2.0 (Rohlf [2004\)](#page-13-15). Landmarks represent the outline of the body and also the location of the snout, eye, opercula bone, and fin insertions. Sliding landmarks were included at the narrowest location (dorsal and ventral) of the caudal peduncle. Digitized x- and y-coordinate data from the 21 landmarks were scaled, rotated and translated using tpsRelw, version 1.42 (Rohlf [2005](#page-13-16)). Variation in white perch shape was summarized with 36 relative warp scores from orthogonal summary of the partial warps.

Raker morphology was analyzed following removal of the most anterior, left gill arch from each fish; digital images were then taken at fixed focal length using a dissecting microscope (Leica EZ4). On the digital photographs, we measured raker length and spacing on the first three rakers ventral to the apex using ImageJ software. Mouth dimensions were analyzed as the width of the closed mouth $(\pm 0.05 \text{ cm})$ between the maxillary bones and by inserting into the mouth a scaled wooden cone (36.8 mm base diameter by 75.9 height) following Nilsson and Brönmark [\(2000](#page-12-23)). Pectoral, pelvic and caudal fin length was analyzed from digital images using the Pythagorean Theorem.

We estimated fish growth using calcified scales and the Fraser–Lee method to estimate growth using back-calculated length at age with the assumption that growth in calcified scales is proportional to growth in length (Ricker [1992](#page-13-17)). Scales were photographed with a digital microscope (Leica EZ4) and distance between annuli was measured from images using ImageJ software (Abràmoff et al. [2004](#page-12-24)). Growth in length was calculated as the proportional increase in length between the two most recent annuli. A subsample of fish analyzed for trophic phenotypes by Tuckett et al. ([2013\)](#page-13-12) was also used in the present study.

Body elemental content

Following evacuation of the foregut, the digestive system was rinsed and macerated with the body in a Waring (CB15) commercial blender. Macerated tissue was dried at 65 °C, passed through a Wiley Mill with a 20 mesh screen, ground to a fine powder with a mortar and pestle and stored at −20 °C. Prior to elemental analysis, powdered material was re-dried at 65 °C. A subsample was ashed at 550 °C for 5 h, acidified with 50 % HCl, and analyzed for P and calcium (Ca) content using inductively coupled plasma atomic emission spectroscopy (ICP-AES; TJA Model 975, Waltham, MA, USA). Phosphorus was measured for all fish while Ca was measured on 33 %–100 % of each population. A second subsample was used to measure C and N content using an elemental analyzer (Leco CN-2000, St. Joseph, MI, USA). Relative percent error (among subsamples) was 1.0 % for C and N, 3.7 % for P and 5.7 % for Ca.

Nutrient excretion

Rates of N and P excretion were estimated by measuring accumulation of ammonia-N, and total dissolved N (TDN) and P (TDP) in tubs containing individual fish (Schaus et al. [1997\)](#page-13-18). Following collection, fish were held in a net pen for less than 30 min then transferred into covered, dark, plastic tubs containing 15 L of pre-filtered (Whatman GF/F) lake water. Adult fish spanning a size gradient from each site were incubated individually for 40–60 min to minimize handling and fasting effects (Whiles et al. [2009](#page-13-19)). Tubs were aerated and partially submerged in the lake to maintain ambient temperature. Water samples were collected before fish addition and 5, 15, and 40 min after adding fish, filtered (Whatman GF/F) into acid-washed bottles and frozen until analysis. Water samples were analyzed for ammonia-N, TDN and TDP using a Lachat QuikChem 8500 flow injection analyzer (Lachat Instruments, Hach Co. Loveland, CO, USA) following standard methods (APHA [2005](#page-12-22)). Individual excretion rates were estimated by regression of nutrient accumulation over incubation time. Only regressions significant at *P* < 0.100 were used in subsequent data analysis.

Statistical analyses

To account for variation among individuals and populations in body size we employed analyses of covariance (ANCOVA) using centroid size, standard length, or dry mass in initial analyses of individual body shape, proportional growth rates (g wet mass g^{-1}), and body elemental content. ANCOVA models use log (10) transformed data to meet the requirements of normality and homoscedasticity. Non-significant interaction terms ($P > 0.050$) were removed from ANCOVA models and reanalyzed. Resulting ANCOVA-derived least-squares (LS) means or allometric slopes for the respective populations were used subsequently in model selection (AIC) or Pearson correlations to assess potential associations with lake conditions (e.g., Secchi depths) at the landscape scale, as well as the relative role of trophic divergence in explaining elemental phenotypes. The Merrymeeting Bay population was not formally included in such regressions due to its estuarine nature, but is noted in figures and results for the sake of comparison. The alpha for among-population regressions was set at 0.100 in deference to the lower statistical power $(df = 7)$. The contribution of bone (% Ca used as a proxy) to body % P was evaluated using ordinary least-squares (OLS) regression with data pooled across populations (Hendrixson et al. [2007](#page-12-15)).

Individual excretion of ammonia-N, TDN, TDP and N:P among populations was similarly evaluated with ANCOVA, but with two covariates, body elemental content (N, P or

Table 2 Results from an analysis of covariance (ANCOVA) of trophic traits with the main effect of population source with size (log centroid size or log standard length) as a covariate

Parameter	\boldsymbol{n}	Whole model				Population source	Size		Source \times size	
		F_{19}	P		F_{9}	\boldsymbol{P}	F_1	P	F_{9}	\boldsymbol{P}
CA ₁	297	62.1	< 0.0001	0.81	81.1	< 0.0001	7.4	0.0070	2.2	0.0201
Raker length	244	67.9	< 0.0001	0.85	3.8	0.0002	562.6	< 0.0001	3.1	0.0015
Raker spacing	245	16.0	< 0.0001	0.58	13.0	< 0.0001	40.3	< 0.0001	3.7	0.0002
Mouth width	297	345.7	< 0.0001	0.96	18.0	< 0.0001	2359.3	< 0.0001	1.4	0.1912
Mouth gape	298	362.8	< 0.0001	0.96	15.7	< 0.0001	2777.9	< 0.0001	2.6	0.0068
Caudal fin	291	171.9	< 0.0001	0.92	15.1	< 0.0001	1201.4	< 0.0001	1.52	0.1415
Pectoral fin	294	650.5	< 0.0001	0.98	25.3	< 0.0001	5331.3	< 0.0001	7.0	< 0.0001
Pelvic fin	294	350.5	< 0.0001	0.96	21.2	< 0.0001	2776.4	< 0.0001	1.67	0.0961
Growth	286	15.3	< 0.0001	0.52	6.8	< 0.0001	166.3	< 0.0001	3.8	0.0001

N:P) and dry mass (g). Interaction terms were removed when not significant ($P \geq 0.05$). The influence of lake trophic state indicators (Secchi, TN and TP) on LS mean population excretion was again evaluated with linear regression. Mass-normalized excretion was used to account for variable size–structure among populations (Brown et al. [2002](#page-12-25), sensu Torres and Vanni [2007](#page-13-20)). Subsequent statistical analyses were performed in JMP (V. 10.0, SAS Institute Inc., Cary, NC).

Results

Lake trophic state

For the nine study lakes, 5-year summer mean Secchi depth, chl-*a*, TP, and TN varied broadly (Table [1](#page-2-0)). Secchi depth was strongly correlated with nutrient enrichment $(TN: R = -0.82, P = 0.007; TP: R = -0.76, P = 0.017).$

Organismal traits

Fish wet weight ranged from 8 to 523 g and varied among populations ($F_{9, 288} = 5.25, P < 0.001$), but was unrelated to Secchi depth ($F_{1,7} = 0.75$, $P = 0.416$). Trophic traits, gill raker length and spacing, width of the closed mouth between the maxillary bones, open mouth gape size and caudal, pectoral, pelvic fin lengths typically varied among populations, were always positively related to body size, and often exhibited significant size \times population interaction (Table [2](#page-4-0)). Proportional yearly growth ranged from 0.15 to 0.29 among populations, negatively scaled with wet weight, and exhibited a significant weight \times population interaction (Table [2](#page-4-0)). Trophic-elemental phenotype correlations are discussed below.

DFA of 36 relative warps (geometric morphometrics) produced 4 canonical axes (CA) explaining 80 % of the

variation in body shape (Wilks $\Lambda < 0.001$; Approximate $F_{324, 2218} = 9.32, P < 0.001$). CA 1 explained 31 % of total shape variation and clearly integrated many of the common trophic adaptations (pelagic/limnetic versus littoral/benthic) seen in perch and other fishes (Skulason and Smith [1995](#page-13-21); Taylor [1999](#page-13-22); Schluter [2000](#page-13-1); Tuckett et al. [2013\)](#page-13-12), and was thus the focus of further analyses. This CA describes variation in body depth ($-$ streamlined, $+$ gibbose), mouth position (− terminal, + subterminal), length and depth of the caudal peduncle (− shallow/long, + deep/short), and rotation of the opercle bone ($-$ cranial; $+$ caudal). CA 1 differed among populations, increased with fish centroid size, and exhibited a size \times population interaction (Table [2\)](#page-4-0). LS means of CA 1 strongly declined with increasing Secchi depth $(F_{1, 7} = 99.80, r^2 = 0.93, P < 0.001)$ while slopes increased $(F_{1, 7} = 5.06 \ r^2 = 0.42, P = 0.059)$. In other words, perch populations in productive lakes were more gibbose/benthic overall and became increasingly so with size whereas populations from unproductive lakes retain the more juvenile, streamlined/pelagic shape even at larger sizes.

Elemental phenotypes

Across populations, body elemental content ranged between 1.9 and 10.1 % for Ca, 34.7 and 48.4 % for C, 8.6 and 11.7 % for N, 2.5 and 5.6 % for P, and 4.0 and 8.0 for N:P. Elemental contents and molar ratios (C:N, C:P, N:P) varied among populations and typically exhibited significant size \times population interactions (Table [3](#page-5-0)). Stoichiometric variation strongly depended on population source; partial r^2 for the effect of population source varied from 0.24 to 0.47 and was greater than size or population \times size interaction terms for all elements and ratios. As lakes became more eutrophic (i.e., declining Secchi depth), body % P increased (Fig. [1](#page-5-1)a) while body % N (Fig. [1b](#page-5-1)) and N:P (Fig. [1c](#page-5-1)) declined. Body % C (Fig. [1](#page-5-1)d) and % Ca (F_1 ,

Parameter	n	Whole model		Population source		Size			Source \times size				
		F_{19}	\boldsymbol{P}		$F_{\rm o}$	\boldsymbol{P}	v	F	\boldsymbol{P}	v	$F_{\rm Q}$	\boldsymbol{P}	\cdot^2
Carbon	293	16.1	< 0.0001	0.53	22.1	< 0.0001	0.38	5.7	0.0176	0.01	8.3	< 0.0001	0.14
Nitrogen	293	13.8	< 0.0001	0.49	15.3	< 0.0001	0.31	25.1	< 0.0001	0.06	6.4	< 0.0001	0.13
Phosphorus	293	20.6	< 0.0001	0.59	31.8	< 0.0001	0.47	31.5	< 0.0001	0.05	5.1	< 0.0001	0.07
N:P	293	20.8	< 0.0001	0.60	29.3	< 0.0001	0.44	57.7	< 0.0001	0.10	4.4	< 0.0001	0.07
C: N	293	16.5	< 0.0001	0.54	21.1	< 0.0001	0.37	1.6	0.2114	< 0.01	9.3	< 0.0001	0.16
C: P	293	20.4	< 0.0001	0.59	30.7	< 0.0001	0.45	23.5	< 0.0001	0.04	6.7	< 0.0001	0.10
Calcium	122	8.0	< 0.0001	0.60	10.3	< 0.0001	0.35	28.1	< 0.0001	0.11	4.3	< 0.0001	0.15

Table 3 Results from an analysis of covariance (ANCOVA) of body elemental content with the main effect of population source with size (log dry mass, g) as a covariate

Fig. 1 Linear relationships between body elemental content and lake trophic state (Secchi depth). Values are population LS Means $(\pm 1 \text{ SE})$ of body % P (**a**), % N (**b**), molar N:P (**c**) and % C (**d**) for 10 populations of white perch. *Closed circles* represent lentic populations $(n = 9)$, the estuarine population $(n = 1)$ is the *closed triangle*. The estuarine population is not included in regressions

 $\tau_7 = 1.89, r^2 = 0.21, P = 0.211$) were not clearly related to Secchi depth but 73 % of variation in individual % P in our dataset is explained by % Ca ($F_{1, 121} = 333.85, P < 0.001$) suggesting that bone content is a major determinant.

Allometry of body chemistry also varied among populations. Six of 10 populations exhibited significant masselement scaling relationships with fish size for % C, albeit with variable slope directionality. Population-specific scaling relationships were also observed for $\%$ N (5 populations), $\%$ P (6 populations), $\%$ Ca (4 populations) and N:P (8 populations). These allometries often correlated with lake conditions. As lakes became more eutrophic (Secchi decreased), allometric slopes of body % P transitioned from more negative to more positive values; the opposite was observed for $\%$ C and N:P ratios (Fig. [2](#page-6-0)a, c, d). There was no relationship between Secchi and allometric scal-ing for % N (Fig. [2](#page-6-0)b) and % Ca ($F_{1, 2} = 2.61$, $r^2 = 0.57$; $P = 0.247$.

Significant correlations between body elemental content and both organismal trait and environmental determinants **Fig. 2** Linear relationships between allometric slopes of body stoichiometry and lake trophic state (Secchi depth). Values are ANCOVA-derived slopes of % P (**a**), % N (**b**), molar N:P (c) and $\%$ C (d) with log dry mass as a covariate for 10 populations of white perch. *Closed circles* are lake populations exhibiting statistically significant slopes (% P, $n = 5$; % N, $n = 5$; N:P, $n = 7$; % C, *n* = 5), *open circles* are lake populations with insignificant slopes and the lone estuarine population is represented by a *closed triangle*. A significant regression is indicated by a *solid line*. The estuarine population is not included in regressions

were common (Table [4\)](#page-7-0). As found above, Secchi depth and overall lake nutrient stoichiometry were often correlated with body elemental content, especially body % P and N:P. However, these environmental drivers were weaker and less consistently related to body stoichiometry than were organismal traits. Correlations between organismal traits were strongest for body % P and N:P (Table [4](#page-7-0)). Many trophic traits were especially highly correlated with body % P $(R = 0.57 - 0.78)$ and N:P ($R = -0.33$ to -0.89) wherein populations exhibiting low body N:P and high P content exhibited more robust body shape (larger CA 1; Fig. [3](#page-8-0)), longer fins, larger mouths, and higher growth rate, consistent with putative benthic phenotypes.

Nutrient excretion

Across populations, individual nutrient excretion rates ranged from 32.1 to 947.3 µmol h^{-1} (coefficient of variation = 0.73) for ammonia-N, 15.1–562.0 (0.59) for TDN, 1.8–101.5 (0.84) for TDP and 6.3–294.8 (1.01) for N:P (Fig. [4](#page-9-0)). Average ammonia-N contributed 44 % of the N to TDN but ranged broadly (7–91 %) across individuals. Excretion rate scaled positively with wet mass for ammonia-N ($F_{1, 270} = 461.98$, $r^2 = 0.63$, $P < 0.001$), TDN ($F_{1, 270} = 461.98$, $r^2 = 0.63$, $P < 0.001$), TDN $_{198}$ = 114.45, r^2 = 0.37, P < 0.001), TDP ($F_{1, 186}$ = 27.51,

 $r^2 = 0.12$, *P* < 0.001) and N:P (*F*_{1, 153} = 5.14, $r^2 = 0.03$, $P < 0.025$, but only ammonia-N scaled close to $\frac{3}{4}$ power (Fig. [4\)](#page-9-0).

ANCOVA models used to evaluate individual excretion (N, P and N:P) revealed significant variation among populations, a consistent relationship with body size and occasional relationships with body elemental content (Table [5](#page-10-0)). Model interaction terms involving population source were often significant. Population source was significant in all models but explained little variation in excretion rates or N:P ratio (partial r^2 < 0.169). Body size was significant for all excretion rates and N:P, with the exception of TDP. Size explained substantial variation in ammonia-N excretion (partial r^2 < 0.606) but little variation (partial r^2 < 0.114) in TDN or N:P. Body elemental chemistry did not explain variation in ammonia-N or TDN excretion and a small portion of variation in TDP (partial $r^2 = 0.035$) and N:P (partial $r^2 = 0.037$).

Mean mass-normalized excretion rates were related in a few cases to ecosystem stoichiometry (nutrient conditions), but not to Secchi depth $(P > 0.112)$. Both mean TDN and ammonia-N excretion increased as lake TN increased $\text{(ammonia-N: } F_{1, 8} = 5.08, r^2 = 0.39, P = 0.054; \text{TDN: } F_{1, 8} = 0.054; \text{TDN: } F_{2, 8} = 0.054; \text{TDN: } F_{3, 8} = 0.054; \text{TDN: } F_{4, 8} = 0.054; \text{TDN: } F_{5, 8} = 0.054; \text{TDN: } F_{6, 8} = 0.054; \text{TDN: } F_{7, 8} = 0.054; \text{TDN: } F_{8, 8} = 0.054; \text{TD$ $R_8 = 6.91$, $r^2 = 0.46$, $P = 0.030$). Mean TDP excretion was unrelated to lake TP ($F_{1, 8} = 0.54$, $r^2 = 0.06$, $P = 0.483$).

Table 4 The relationship between LS mean body elemental content derived from analysis of covariance (ANCOVA) and LS mean organismal and environmental drivers

Parameter	$\%$ C	$\%$ Ca	$\%$ N	$\%$ P	N:P
Organismal drivers					
Canonical axis 1	-0.43	0.33	-0.50	$0.77**$	$-0.89**$
Caudal fin length	-0.44	0.34	-0.38	$0.68**$	$-0.76**$
Pectoral fin length	$-0.60*$	$0.80**$	-0.19	$0.78**$	$-0.80**$
Pelvic fin length	-0.31	$0.65*$	-0.27	$0.68**$	$-0.74**$
Mouth width	$-0.70**$	$0.59*$	0.03	$0.78**$	$-0.76**$
Mouth gape	-0.38	0.33	-0.11	$0.61*$	$-0.59*$
Raker length	-0.25	0.31	-0.33	$0.57*$	$-0.63*$
Raker spacing	0.34	-0.11	$-0.63**$	0.15	-0.33
Growth	$0.57*$	$-0.84**$	-0.10	$-0.60**$	$0.56*$
Bone (% calcium)	$-0.68**$	1.00	0.31	$0.77**$	$-0.64**$
Environmental drivers					
Secchi depth	0.27	-0.46	$0.61*$	$-0.68**$	$0.83**$
Water total N	-0.24	0.23	-0.25	$0.60*$	$-0.64**$
Water total P	-0.47	0.33	-0.42	$0.71**$	$-0.82**$
Water N:P	0.46	-0.27	0.37	-0.46	$0.59*$
Chlorophyll-a	-0.15	0.24	$-0.58*$	0.24	-0.41
Lake mean depth	0.41	-0.39	0.18	$-0.70**$	$0.71**$

Linear significant Pearson correlations for the ten populations are indicated by $*(P < 0.10)$ and $** (<0.05)$

Excretion N:P increased with increasing lake TN (F_1) , $R_8 = 3.57, r^2 = 0.31, P = 0.095$, but was unrelated to lake N:P ($F_{1, 8}$ < 0.01, r^2 < 0.01, $P = 0.970$) or TP ($F_{1, 8} = 1.36$, $r^2 = 0.15, P = 0.277.$

Discussion

Variation in elemental phenotypes

White perch C, N, P, and Ca content was highly variable among populations, spanning a large portion of the range reported across fish species (McIntyre and Flecker [2010](#page-12-6)). For white perch, the range in % P $(2.5-5.6\%)$ was 70 % of the range reported for 100 species representing 31 fish families; the % N range for perch $(8.6-11.7 \%)$ was 48 % of the range among species. This is consistent with an emerging picture that there is substantial intraspecific variation in body elemental content, casting considerable doubt on the assumption of homeostatic elemental phenotypes within a species. The variation we found was comparable to the few other species studied to date, including among morphs of three-spine stickleback (*Gasterosteus aculeatus*; 2.5–6.1 % P) (El-Sabaawi et al. [2016\)](#page-12-9), small-bodied Trinidadian guppies (*Poecilia reticulata*; 2.1–5.4 % P) (El-Sabaawi et al. [2012b](#page-12-8)), and bluegill sunfish (*Lepomis macrochirus*; 2.7– 4.4 % P) (Hendrixson et al. 2007). In this study of white perch and also with guppies, much of the variation in elemental content was attributed to population/site differences (El-Sabaawi et al. [2014\)](#page-12-16); for white perch, the partial $r²$ for population exceeded the direct or interactive effects of body size (Table [3](#page-5-0)). Further, population means spanned 35 % of the total individual range for % P (3.43–4.50), 29 % for % N (9.90–10.80) and 39 % for N:P (4.97–6.51). The landscape variation shown here and elsewhere may shift how stoichiometry is considered, where emphasis is often placed on the ecological consequences of species or family identity (Vanni et al. [2002;](#page-13-7) McIntyre et al. [2008\)](#page-12-26).

Environmental versus trait determinants

One of our major objectives was to examine the causes of variation in elemental phenotypes and whether they can be linked to environmental or trait drivers which could promote/suppress eco-evolutionary feedbacks. To date, variation in elemental phenotypes have been tentatively linked to site-specific environmental factors such as biofilm nutrient content (El-Sabaawi et al. [2012b](#page-12-8)) and also to organismal traits such as body size, body shape, and investment in structural material (Sterner and George [2000](#page-13-23); Tanner et al. [2000;](#page-13-9) Hendrixson et al. [2007;](#page-12-15) McIntyre and Flecker [2010](#page-12-6); El-Sabaawi et al. [2016\)](#page-12-9), which may be constrained by species identity (Hendrixson et al. [2007\)](#page-12-15) and ontogenetic trajectories (Boros et al. [2015\)](#page-12-10). In perch, significant regressions between body stoichiometry and our index of lake trophic state (Secchi depth) suggests environmental factors, particularly ecosystem trophic state, are important in shaping white perch stoichiometry. However, comporting to our predictions, variation in perch body stoichiometry was often better explained by trophic traits that diverged quickly in response to lake conditions. Thus, in perch, the greatest role for environmental factors in producing consistent patterns of body stoichiometry may be via their ultimate effects on population trophic traits compared to their proximal effects on individuals. This was shown by the particularly strong correlations between a suite of trophic traits and body stoichiometry.

Trophic state is strongly related to nutrient availability and also mean depth in lakes, which influences community and ecosystem processes that govern resource availability and the distribution and profitability of prey (Fox [2007](#page-12-27)). In unproductive lakes perch were streamlined, had narrow-spaced gill rakers and shorter fins, consistent with adaptation to foraging in pelagic habitats. As lakes became

Fig. 3 Distribution of % C (**a**), % N (**b**), % P (log dry mass (g) (**c**) and N:P (log molar (**d**) originating from 10 study sites across body shape (canonical axis 1). Points $(n = 293)$ are coded by study site. Large open (study lakes; $n = 9$) and *closed* (Merrymeeting Bay; $n = 1$) *circles* are population means. Fish images on the *x*-axis are consensus representations of fish shape from geometric morphometrics and the discriminant function analysis and correspond to the axis end points $(6, -6)$

more eutrophic, perch body shape became more robust and deeper with wider spaced gill rakers and longer fins with pronounced fin rays; this is consistent with traits associated with greater benthic foraging and, possibly, predator defense (El-Sabaawi et al. [2016\)](#page-12-9). In our previous work, we showed that Secchi depth, an indicator of trophic state, was a much better predictor of these phenotypes than any one biotic or abiotic factor, including mean lake depth and littoral area (Tuckett et al. [2013\)](#page-13-12). The relationship between feeding and locomotor traits is well known and represents

a suite of correlated traits, where a streamlined body shape is more efficient at foraging in open water for spatially distributed prey and a robust body shape with larger fins is more efficient at foraging in complex habitats like the littoral zone of lakes (Webb [1984](#page-13-24); Robinson et al. [1993](#page-13-25); Blake [2004\)](#page-12-28). Ultimately, these patterns of morphology are consistent with our prior observations of these populations and support the hypothesis that populations can diverge in traits expected to shape body stoichiometry and excretion (Tuckett et al. [2013\)](#page-13-12).

Fig. 4 Allometry of log (10)-transformed ammonia-N $(n = 272)$ (a), TDN $(n = 200)$ (**b**), TDP (*n* = 188) (**c**) excretion and molar ratio of excreted N:P $(n = 155)$ (**d**) for individual white perch from 10 populations. *Closed circles* are excretion estimates which exhibited significant regressions with time (*P* < 0.10); *open circles* are nonsignificant $(P > 0.10)$ and were excluded from regression with log wet weight

Wet weight (log g)

McIntyre and Flecker ([2010\)](#page-12-6) hypothesized that elemental phenotypes in fish are also driven by tradeoffs between investment in bone (high P) and muscle (high N) due to sedentary (low N:P) versus mobile (high N:P) lifestyles. Support for this hypothesis in our data can be found not only in the strong mean trophic trait–element correlations, but also in observed patterns of allometric scaling of N, P, and N:P. Typical of lentic fishes, white perch juveniles occupy pelagic habitats, feeding on rotifers and switching to larger zooplankton and benthic invertebrates as they mature (Stanley and Danie [1983\)](#page-13-13). Allometric niche shifts are mirrored in body shape changes as perch transition from streamlined to rounded body shape, more typical of a benthic or littoral lifestyle (Webb [1978](#page-13-26), [1984](#page-13-24)).

Table 5 Relationship between excretion of ammonia-N, TDN, TDP and N:P and population source, weight and stoichiometry

Parameter	\boldsymbol{n}	F	r ²	P
Ammonia-N excretion	273	28.706	0.774	< 0.001
Population source		9.080	0.057	< 0.001
Size		222.886	0.606	< 0.001
$\%$ N		0.043	< 0.001	0.836
Source \times size		4.289	0.105	< 0.001
Source \times % N		2.699	0.066	0.005
Size \times % N		NA	NA	0.799
Source \times size \times % N		NA	NA	0.160
TDN excretion	198	6.343	0.609	< 0.001
Population source		3.366	0.143	< 0.001
Size		24.234	0.114	< 0.001
$\%$ N		2.627	0.012	0.107
Source \times size		2.288	0.097	0.019
Source \times % N		2.814	0.119	0.004
Size \times % N		1.289	0.006	0.258
Source \times size \times % N		2.782	0.118	0.004
TDP excretion	188	4.978	0.254	< 0.001
Population source		1.271	0.102	0.256
Size		3.682	0.033	0.057
$\%$ P		3.953	0.035	0.048
Source \times size		NA	NA	0.177
Source \times % P		NA	NA	0.090
Size \times % P		9.248	0.083	0.003
Source \times size \times % P		NA	NA	0.399
TDN:TDP excretion	137	3.955	0.258	< 0.001
Population source		3.193	0.169	0.002
Size		8.713	0.051	0.004
N: P		6.313	0.037	0.013
Source \times size		NA	NA	0.364
Source \times N:P		NA	NA	0.439
$Size \times N:P$		NA	NA	0.162
Source \times size \times N:P		NA	NA	0.957

Results are from an analysis of covariance (ANCOVA) with population source, wet weight (g) and body stoichiometry (% dry mass or molar ratio) as covariates. The overall model summary is shown next to the parameter. Non-significant $(P < 0.050, NS)$ interaction terms were removed and the model was reanalyzed

In more oligotrophic lakes, perch are slower to undergo this transition and retain the streamlined body shape and other trophic features (e.g., gill rakers) adapted for cruising the water column for pelagic prey (Tuckett et al. [2013](#page-13-12)), including zooplankton and eventually fish. These putative pelagic-cruising fish had higher mean body N:P and shallow or negative allometric slopes in body chemistry which might indicate neoteny (i.e., juvenile traits in adults) and retention of pelagic trophic morphology. Conversely, we would expect productive lakes to favor a more sedentary

benthic lifestyle as fish mature; this is also consistent with our results. This correlated stoichiometric and trophic divergence may have a contemporary origin because body shape closely tracks lake Secchi depth $(r^2 = 0.93)$ and several lakes in our study have undergone eutrophication with increased nutrients and large declines in Secchi depth over the past few decades (Tuckett et al. [2013\)](#page-13-12).

Although we did not directly include Merrymeeting Bay fish in many analyses due to their unique habitat, they may be an exception that proves the rule when it comes to stoichiometric associations with body shape and mobility. These fish exhibited benthic trophic morphology more extreme than fishes originating from the most productive source lakes (see also Tuckett et al. [2013\)](#page-13-12). This is expected as estuarine fish are commonly more gibbose and exhibit greater benthic feeding than their lentic counterparts (Berner et al. [2008](#page-12-29)). Here, estuarine perch exhibited high body % P and low % N and N:P, similar to fish from eutrophic lakes. However, allometric scaling for body shape and % P for these estuarine fish were opposite to those of fish from productive lakes (i.e., fish became more streamlined and lower in % P), which may be expected for estuarine fishes that transition from tidal creeks and nearshore habitats as juveniles to deep, swift estuarine channels and piscivores as adults (Able and Fahay [1998](#page-12-30)). Ultimately, we were unable to identify the exact proximate reason for lower % P in streamlined individuals and populations; yet, it could be related to all or a combination of several factors, including reduced fins, less slab-sided morphology and even reduced investment in bone and defensive traits (Tanner et al. [2000;](#page-13-9) Hendrixson et al. [2007;](#page-12-15) Vrede et al. [2011](#page-13-8); El-Sabaawi et al. [2016](#page-12-9)), as Ca, a proxy for bone, explained 73 % of the individual variation in body P.

Variation in excretion and eco‑evolutionary feedbacks

Much like body element content, excretion of N and P was highly variable across populations of white perch relative to variation among species. For example, individual ammonia-N excretion across perch populations was 95 % of the mean range reported for 47 species representing 7 trophic guilds (McIntyre et al. [2008](#page-12-26)). Population source explained 10–48 % of the variation in individual excretion rates, but excretion was not as clearly tied to environmental and organismal determinants as body stoichiometry. Previously, we showed that perch elemental phenotypes were correlated to trophic phenotypes which are dictated by landscape variation in ecosystem trophic state. Next, to examine a possible eco-evolutionary feedback, we examined the causes of variation in perch excretion and whether it was related to trophic state or elemental phenotypes. Perch N, but not P excretion increased with lake TN, somewhat consistent with a study showing fish excretion increases along a productivity

gradient; however, the increase in fish excretion was attributed to altered population biomass (Vanni et al. [2006](#page-13-5)). In particular, the increase in body % P and decrease in body % N and N:P as lakes became more eutrophic should have led to decreased relative P excretion and increased N excretion. Indeed, body elemental content explained little variation in N and P excretion rates and ratios. Interestingly, among species, excretion is well correlated with body elemental content and body N:P has been shown to be inversely related to excretion N:P (Vanni et al. [2002](#page-13-7)).

From an organismal trait perspective, metabolic and ecological stoichiometry theory suggests nutrient excretion should be driven by size, growth rate, and body and diet elemental content (Sterner and Elser [2002;](#page-13-6) Brown et al. [2004](#page-12-13); Allgeier et al. [2015](#page-12-14)). Excretion rate increased with body size as in other studies (Hall et al. [2007;](#page-12-31) McIntyre and Flecker [2010\)](#page-12-6) with scaling less than 1, but not consistently with 3/4 power scaling predicted by metabolic theory. Given the large changes in body N and P in perch, we expected to see a commensurate shift in excretion rate and ratio, but this generally did not occur. Ultimately, as has been found in other studies (Allgeier et al. [2015](#page-12-14); Tuckett et al. [2015](#page-13-14)), our data suggest, compared to body size, stoichiometric variation in body composition does not simply translate to altered recycling stoichiometry in this study system over shortened timescales. These body size considerations will be important moving forward because of its strong relationship to excretion and as climate change, sizeselective fishing, and other anthropogenic activities alter the size–structure of fishes (Shackell et al. [2010;](#page-13-27) Layman et al. [2011;](#page-12-32) Allgeier et al. [2015;](#page-12-14) Tuckett et al. [2015](#page-13-14)).

Diet shifts are also associated with trophic divergence and may offset or mask body content effects. The lack of clear results may be due to the complexity of variables influencing excretion, some of which were not measured here (e.g., assimilation efficiency leading to egestion, diet stoichiometry, etc.). For example, Trinidadian guppies alter the balance of nutrients consumed, assimilated, and excreted due to predator cues (Dalton and Flecker [2014](#page-12-33)). Further, as we would expect, diet stoichiometry is strongly related to excretion (Moody et al. [2015\)](#page-12-34). Here, we show that perch body stoichiometry is highly variable, contrary to assumptions of homeostatic regulation; thus, we cannot assume the same for dietary items which would also need to be analyzed for each lake (but see Frost et al. [2002](#page-12-35)). Ultimately, there may be difficulty in accurately quantifying diet in a way that matches the measurement of excretion where an animal is not likely to be excreting what is in the stomach, but instead excretes a metabolically averaged product of its assimilated past diet. Thus, laboratory studies may be more useful in examining the relationship between diet and excretion stoichiometry.

Implications

There is growing recognition that a diverse group of animals can influence biogeochemical processes in aquatic ecosystems via their roles in nutrient storage and CDNR (Post et al. [1998;](#page-13-28) McIntyre et al. [2008\)](#page-12-26). Typically these studies consider the effects of interspecific variation, with a few notable exceptions for intraspecific body size and life history variation (Vanni et al. [2002](#page-13-7); Hall et al. [2007;](#page-12-31) McIntyre et al. [2008](#page-12-26); Post and Palkovacs [2009](#page-13-29); Bassar et al. [2010\)](#page-12-18), and usually consider only body composition *or* excretion in relation to traits or environmental features. Our study is unique in simultaneously assessing intraspecific variation in body elemental content and excretion to one another, to trait divergence and to landscape variation in environmental conditions. Moreover, we do so for populations wherein current patterns of phenotypic divergence have originated in contemporary time. Our data suggest population divergence in response to lake trophic state leads to coherent patterns in elemental phenotypes via trophic adaptations. Theoretically, this may set the stage for eco-evolutionary feedbacks such that contemporary trait change influences ecosystem processes through stoichiometric linkages (Palkovacs et al. [2009;](#page-12-11) Bassar et al. [2010](#page-12-18)). We found limited support for a potential eco-evolutionary linkage via CDNR effects of perch, given that perch excretion was not consistently predicted by lake trophic state or trophic trait variation. However, eco-evolutionary dynamics may in many cases be cryptic and their outcomes difficult to detect (Kinnison et al. [2015\)](#page-12-36), perhaps even leading to stability as elevated trophic state leads to greater body P which suppressed P excretion. Regardless, other CDNR pathways may exist for future study, pathways that could alter the pool of available nutrients, including the potential for stoichiometrically divergent fish to act as nutrient sinks or prey. Still, we not only show that stoichiometric diversity can be large within a species, approaching interspecific scales, but also provide mechanistic support for how variation can arise quickly and predictably through the effects of landscape-scale environmental drivers on trophic divergence.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

Ethical approval All applicable institutional and/or national guidelines for the care and use of animals were followed.

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