REVIEW ARTICLE

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The Finnish disease heritage III: the individual diseases

Received: 20 June 2002 / Accepted: 30 October 2002 / Published online: 8 March 2003 © Springer-Verlag 2003

Abstract This article is the third and last in a series entitled The Finnish Disease Heritage I–III. All the 36 rare hereditary diseases belonging to this entity are described for clinical and molecular genetic purposes, based on the Finnish experience gathered over a period of half a century. In addition, five other diseases are mentioned. They may be included in the list of the "Finnish diseases" after adequate complementary studies.

Abbreviations AFP alpha-fetoprotein $\cdot CT$ computed tomography $\cdot ECG$ electrocardiography $\cdot EEG$ electroencephalography $\cdot ENMG$ electroneuromyography \cdot ERG electroretinography $\cdot HUCH$ Helsinki University Central Hospital $\cdot MBD$ minimal brain dysfunction \cdot MIM Mendelian Inheritance of Man (McKusick) $\cdot MRI$ magnetic resonance imaging $\cdot OUH$ Oulu University Hospital $\cdot SEP$ somatosensory evoked potential $\cdot SPECT$ single photon emission computed tomography $\cdot TUH$ Turku University Hospital $\cdot VEP$ visual evoked potential

Introduction

This article series will acquaint the international readership with the Finnish Disease Heritage, a collection of nearly 40 rare hereditary diseases over-represented in Finland. In the first article, investigations of the characteristics, causes and background of this phenomenon were reported. The second part dealt with the population prehistory and genetic roots of the Finns.

In this third article, the individual diseases are described so as to present the relevant details for practical purposes to medical geneticists and other physicians. Aspects of each disease are presented in sub-sections, facili-

R. Norio ([∞]) Department of Medical Genetics, The Family Federation of Finland, P.O. Box 849, FIN 00101 Helsinki, Finland Fax: +358-9-645018, e-mail: reijo.norio@kolumbus.fi tating the easy finding of desired details. While these descriptions must be limited to a certain extent, further detailed information is available from the selected articles given in the references section at the end, which is divided into separate sub-sections for each disease. In most cases, these articles are not separately referred to in the text. In choosing the references preference is given to the research based on the numerous patients and gathered experience in Finland. Also the "historical" development of the growing knowledge and the cornerstones of investigation are included. Most references in the Finnish language have been excluded. They are, however, mentioned in my book Suomi-neidon geenit (The Genes of Maiden Finland), Otava, Helsinki, 2000, upon which the presentations of this article are based. Those who can read Finnish texts may also have access to that book.

Out of the 36 diseases described, the most (32), are transmitted in an autosomal recessive manner. This has not been mentioned separately. The mode of inheritance of the two autosomal dominant and two X-linked disorders is *italicised* in the introduction of the disease.

The rare diseases have many — not to say too many names. Among them, the name used mostly in Finland has been chosen as the first name. Unfortunately, no system prevails in this; practical circumstances and habit have often decided the name used. It can also be an abbreviation if the proper name is desperately long or otherwise complicated (e.g. AGU instead of aspartylglucosaminuria). Eponyms, though not recommended in general, may be handy in use (Herva disease instead of lethal congenital contracture syndrome). Important synonyms, practical abbreviations and old names are mentioned for clarity, but every possible name has not been included. As regards them, *McKusick's* (*Online*) *Mendelian Inheritance in Man* is referred to.

The most important clinical features are presented to such an extent as to give an impression and preliminary diagnostic data for the disease. The pictures drawn by my colleague Dr. Mari Markkanen-Leppänen also serve to give an impression and memory aid. As concerns diagnostic investigations and prenatal diagnosis, the Finnish physicians are especially privileged in regard to gene di-

Table 1	The diseases	of the Finnish Disea	se Heritage in orde	er of their incidence	e in Finland

Name of disease	Clinical abbreviation*	Incidence in Finland	Patients known		Data known about the gene		
			in Finland	Elsewhere	Name	Locus	Character
Autosomal recessive							
Congenital nephrosis	CNF (NPHS1)	1:8,000	>300	<300	NPHS1	+	+
Infantile neuronal ceroid lipofuscinosis	INCL CLN1	1:14,000	>160	>200	CLN1 PPT1	+	+
Meckel syndrome		1:15,000	>100	Not very rare	MKS1 MES	+	_
Unverricht-Lundborg disease	<i>PME</i> PME-UL	1:17,000	~200	Not very rare	EPM1 CSTB	+	+
Aspartylglucosaminuria	AGU	1:18,000	>260	>30	AGA	+	+
Cartilage-hair hypoplasia	СНН	1:18,000	>170	>200	RMRP (CHH)	+	+
Spielmeyer-Sjögren disease	JNCL SS	1:19,000	~200	Not very rare	CLN3	+	+
Hydrolethalus syndrome		1:22,000	>80	~10		+	_
Diastrophic dysplasia	DD DTD	1:22,000	~200	>250	SLC26A2 (DTDST)	+	+
Autoimmune polyendocrinopathy — candidiasis — ectodermal dystrophy	APECED APS I	1:27,000	>90	~300	AIRE	+	+
Lethal congenital contracture syndrome (Herva)	LCCS	1:29,000	>40	Some	LCCS	+	_
Congenital chloride diarrhea	CCD? CLD?	1:33,000	>50	>200	DRA SLC26A3	+	+
Mulibrey nanism	MUL	1:37,000	>80	<20	TRIM37 (MUL)	+	+
Usher syndrome type 3	USH3	1:42,000	~100	Some	USH3	+	+
Salla disease		1:42,000	>100	>20	SLC17A5 (SIASD)	+	+
Cornea plana congenita	CPC CNA2	1:46,000	~90	>50	KERA	+	+
Congenital lactase deficiency	CLD	1:48,000	>40	<40	CLD?	+	_
Muscle-eye-brain disease	MEB	1:52,000	~30	>10	POMGnT1	+	+
Nonketotic hyperglycinemia	NKH	1:52,000	~50	~100	GCSP	+	+
Lethal arthrogryposis with anterior horn cell disease (Vuopala)	LAAHD	1:53,000?	~20	Some?		_	_
Jansky-Bielschowsky disease variant	vLINCL _{Fin} CLN5 JB	1:59,000	>30	Some	CLN5	+	+
Hyperornithinemia with gyrate atrophy of choroid and retina	HOGA (GA)	1:63,000	>80	Not very rare	OAT	+	+
GRACILE syndrome (Fellman)		1:64,000?	>20	_	BCS1L	+	+
Selective malabsorption of vitamin B_{12}	SMB ₁₂ (MGA1)	1:68,000	~40	~150	CUBN MGA1	+	+
Nasu-Hakola disease	PLO-SL	1:71,000	>30	>120	TYROBP (DAP12)	+	+
Lysinuric protein intolerance	LPI	1:76,000	~50	>100	SLC7A7 (LPI)	+	+

Table 1 (continued)

Name of disease	Clinical abbreviation*	Incidence in Finland	Patients known		Data known about the gene		
			in Finland	Elsewhere	Name	Locus	Character
PEHO syndrome	РЕНО	1:78,000	~30	~10		_	_
IOSCA syndrome	<i>IOSCA</i> (OHAHA)	1:90,000	>20	_	SCA8	+	_
Cohen syndrome		1:105,000	>30	>100	COH1 (CHS1)	+	_
Rapadilino syndrome		1:105,000	>10	Some?		-	-
Follicle stimulating hormone- resistant ovaries (Aittomäki)	FSH-RO	1:127,000	>20**	?	ODG1 FSHR	+	+
Northern epilepsy	CLN8	1:176,000	>20	-	EPMR CLN8	+	+
Autosomal dominant							
Meretoja disease	FAF	~1:6,000 (prevalence)	>400	9 kindreds	GSN	+	+
Tibial muscular dystrophy	TMD	3/year?	>300	6 kindreds	TTN	+	+
X-chromosomal							
Choroideremia		~2/year	>100	<400	CHM	+	+
Retinoschisis	RS	~1:17,000 (prevalence)	>300	Not very rare	XLRS1 (RS)	+	+

*Italicised abbreviations are used as the practical name of the disease.

**Incidence figures include only the patients with a demonstrated gene mutation. In reality, the disease is much more common

agnosis; in general, diagnostic investigations that are useful in Finland are not necessarily suitable or reliable in other countries. For pathogenesis and molecular genetics, the biochemical and molecular genetic details are not presented here but can be found in the list of references. The same is true for the management. Of the historical aspects, the first descriptions of the disease in medical literature and in Finland are given. The Finnish researchers who have been active with their diseases recently are not mentioned, but the list of references will disclose them effectively. The sub-sections on epidemiology give data on the number of known patients in Finland and elsewhere. The map of Finland showing the birth places of patients' grandparents not only displays the geographical distribution of the genes of a particular disease, but it also visualises the different types of distributions dealt with in Part I (DOI s00439-002-0875-3). The naming of some Finnish experts gives the reader a possibility for solving problems that can be encountered with their individual patients. The experts can be contacted the easiest by email. In case the e-mail address should fail — e.g. after some years - postal addresses of the institutions concerned given in the references may bring help. An expert on most of the gene tests is Dr. Irma Järvelä, HUCH (email: irma.jarvela@hus.fi).

Heterozygous manifestations are not mentioned under a separate subtitle in the descriptions of individual diseases, because their practical significance today is mostly small (see Vogel 1984, Clin Genet 25:381–415). Three diseases, however, deserve a special mention. Autosomal dominant tibial muscular dystrophy (TMD) is, in fact, a heterozygous manifestation of the gene of a severe limb girdle dystrophy. In X-linked recessive choroideremia, the heterozygous females have fundus changes resembling retinitis pigmentosa. This can lead to a tragic misdiagnosis. In congenital nephrotic syndrome of the Finnish type, heterozygous mid-trimester fetuses may show very high, but later decreasing, concentrations of alpha-fetoprotein in the amniotic fluid. Without a control amniocentesis, the consequence could be the abortion of a healthy fetus.

In the nationwide clinical investigations of ten diseases, heterozygous affections have been sought but the findings have been incidental and/or insignificant. Details can be found in the following references given in connection with the respective diseases: AGU, Autio (1972); APECED, Ahonen (1985); CNF, Norio (1966); INCL, Santavuori, personal comm.; LPI, Norio et al. (1971); Meckel, Salonen and Norio (1984); NKH, von Wendt et al. (1981); PLO-SL, Hakola (1972); PME, Norio and Koskiniemi (1979); rapadilino, Kääriäinen et al. (1989). In addition, obligate heterozygotes have shown intermediate values of enzymes or metabolites, respectively, in AGU (Aula et al. 1974, 1976; Mononen and Aronson 1997), HOGA (Takki and Simell 1974; Valle and Simell 2001), NKH (von Wendt et al. 1979), and Salla disease (Aula and Gahl 2001). AGU heterozygotes have shown proneness to chronic arthritis (Arvio et al. 2002). A statement of having demonstrated no heterozygous affections was found in the investigations of 12 diseases. In 11 diseases, no mention of heterozygous manifestations could be found.

Before the disease descriptions, a compendium of the diseases in the order of their incidences in Finland is presented in Table 1. The incidences are based on lists of patients received from the expert doctors for each particular disorder. The birth incidence figures are calculated from the number of patients born in the decade when the number was the greatest. Most often this fell on the period when a doctor had made his or her nationwide thesis study on the disease. In many earlier texts, even widely differing estimates of the disease incidences have been presented. The incidence figures in Table 1 are calculated in the same way for all the diseases and are thus comparable. Over-estimations can hardly be obtained in this manner.

AGU

Aspartylglucosaminuria MIM 208400



Fig.1

AGU is a lysosomal storage disorder caused by defective activity of the enzyme aspartylglucosaminidase (glycoasparaginase) leading to progressive mental retardation.

Clinical features

After an uneventful infancy the symptoms manifest at preschool age. Proneness to respiratory infections occurs. Speech development and learning abilities slow down. Restlessness, irritability, clumsiness and floppiness may cause suspicion of the so-called MBD syndrome. Attending normal school will not succeed. Attacks of agitation and confusion appear. By adult age, mental retardation is severe and speech is defective. Epileptic seizures are common.

The facial features become coarse with age, showing i.a. thick lips and big tongue. Facial angiofibromas as in tuberous sclerosis appear at adult age. Tendency to fractures is due to osteoporosis. Survival over 50 years is seldom, bacterial infections being the usual cause of death.

The growth has characteristic features. The height increases rapidly during the first year of life. The pubertal height spurt begins early but ends soon, resulting in a considerably low adult height. Menarche appears early, at about 11 years.

Diagnostic investigations

In the urine, aspartylglucosamine can be found. Vacuoles are seen in lymphocytes and monocytes. The deficiency of the AGA enzyme can be verified in fibroblasts or lymphocytes. Gene diagnosis is accurate and easy in Finland.

Pathogenesis

The activity of glycoprotein degrading enzyme aspartylglucosamine (N-aspartyl- β -D-glucosaminidase = AGA) is deficient. Aspartylglucosamine and other abnormal degradation products are stored in the lysosomes. This damages especially the brain cells.

Molecular genetics

The gene AGA is in chromosome 4q32-q33. Amongst the Finnish AGU genes, in 98% the point mutation AGU Fin major (488G>C) causes a change of cysteine into serine (C163S) in the AGA enzyme. Mutation AGU Fin minor, a 2-bp deletion in the second exon of the AGA gene (199–200delGA), is found as compound heterozygote in seven families. In AGU patients of the neighbouring countries of Finland, Sweden and Norway, the mutation AGU Fin major is found. In other countries different mutations occur in the same locus.

Management

There is no curative treatment. Adult patients often need institutional care. Carbamazepine is the first choice for epileptic seizures. Attempts at gene therapy are ongoing, also by aid of mouse models. Already, the AGA enzyme functions successfully in cultivated cells of the AGU patient.

Prenatal diagnosis

Prenatal diagnosis is possible by showing defective activity of AGA in cultivated amniotic cells or, in Finland, by a gene test.

Historical aspects

AGU was detected in 1967 as a by-product in Jorma Palo's investigation on the incidence of PKU in Finland.

He found an unknown amino acid spot in some of the tested urine samples. In the same year, Jenner and Pollitt described the same finding in two mentally retarded British siblings.

Epidemiology

In Finland, more than 260 patients are known from almost all over the country. From elsewhere, over 30 patients have been published.

Finnish experts

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Molecular genetics

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APECED

Autoimmune polyendocrinopathy — candidiasis — ectodermal dystrophy Autoimmune polyglandular disease type I APS I MIM 240300

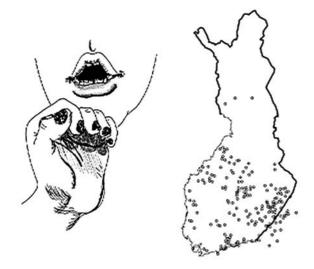


Fig. 2

APECED is a very polymorphic autoimmune disease, developing new manifestations over tens of years. Main findings are persistent candidial infections, chronic ectodermal alterations and many endocrinologic disturbances, one after another.

Clinical features

The long name of the disease describes the three groups of manifestations. The predilection for mucocutaneous candidial infections is the mouth. Cheilosis (rhagades at the corners of the mouth) is a typical finding. Other sites are nails, skin, esophagus, intestine, and genitals. Ectodermal lesions are enamel hypoplasia of the permanent teeth, punctate nail pits, corneal irritation and opacities, calcifications of eardrums, alopecia, and vitiligo. Different endocrinological defects appearing in childhood, and after that during subsequent decades, are hypoparathyroidism, adrenocortical insufficiency, atrophy of ovaries and testes, hypothyroidism, and diabetes mellitus. Also megaloblastic anemia, steatorrhea, and hepatitis may be autoimmune manifestations.

Diagnostic investigations

Laboratory investigations are needed, depending on the suspected manifestations. Autoimmune antibodies against different target organs may be detectable. A diagnostic gene test is in use in Finland, but its limitations must be taken into consideration as concerns uncommon mutations.

Pathogenesis

The immune defence mechanisms have gone astray, destroying the tissues of the organism. The details are unknown, as are the interconnections of ectodermal and autoimmune phenomena.

Molecular genetics

The gene, *AIRE* (autoimmune regulator), is in chromosome 21q22.3. In nearly 90% of the Finnish disease genes the mutation is 889C>T = R257X. Over 40 different mutations have been detected in the same locus in different populations. The predicted structure of the proline-rich AIRE protein includes two zinc finger motifs. It is a DNA-binding protein and probably participates in the regulation of transcription. A mouse model exists.

Management

No treatment is available to stop the autoimmune process. Supplementary hormone therapy and accurate follow-up are essential for the endocrine deficiencies. Alertness for new endocrinopathies is important, but it must not be allowed to invalidate the daily life of the patient too much. A permanent endocrinologist is of utmost importance to the patient. Support for the mental health of the patient should not be forgotten.

Candidial infections must be treated often with systemic fungicides like ketoconazole, because continuous candida infection may expose the oral mucosa to malignant transformation. Effective local treatment is important for corneal symptoms. Cosmetic nail, skin and hair problems demand both sympathy and concrete means.

Prenatal diagnosis

A gene test gives possibilities for prenatal diagnosis in risk families, at least in Finland, although the diagnosis of the first affected sib often comes too late for that.

Historical aspects

The disease was described by Thorpe and Handley of the USA as early as 1929. Many names have been given to the disease. In Finland, Visakorpi and Gerber published the first description in 1963.

Epidemiology

In Finland, over 90 patients are known; elsewhere about 300. The incidence in Finland is about 1:25,000, but in Sardinia 1:15,000 and in an Iranian Jewish population greater than 1:10,000.

Finnish experts

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Cartilage-hair hypoplasia

Metaphyseal chondrodysplasia, type McKusick CHH MIM 250250

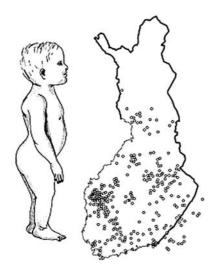


Fig. 3

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Clinical features

Dwarfism is of intrauterine onset. The femora and humeri are particularly short. The fingers are short and hypermobile. The laxity of articular ligaments, limited extension at the elbows, bow legs, and accentuated lumbar lordosis are the usual findings. The structure and function of joints is nearly normal. The adult height is 100–140 cm.

The hair, beard, eyebrows, eyelashes, etc. are sparse and thin. In some patients, banal infections may be overwhelming, even life-threatening, due to immunodeficiency. Childhood anemia may be frequent and even severe. Colonic aganglionosis and impairment of spermatogenesis are often associated with CHH. Disposition for malignancies, particularly for lymphomas, is increased.

Diagnostic investigations

The metaphyses of the long bones of the extremities show splaying and irregular growth plates in X-ray during childhood. Laboratory investigations for detecting cellmediated immunodeficiency are needed. A gene test is available for the majority of patients.

Pathogenesis

The pathogenetic mechanism is unknown. Some kind of disturbance in cell proliferation would explain many affections of the disease.

Molecular genetics

The gene (*RMRP*, formerly *CHH*) is in chromosome 9p13. Instead of a protein, it encodes an RNA molecule that combines with several proteins to form a complex functioning as an endoribonuclease. The most common mutation is 70A>G, found in 92% of the Finnish, 40% of other European and in all Amish patients investigated so far. About 40 other mutations have been found in the *RMRP* gene. The details of the disturbance caused by the mutated gene are not known.

Management

With the exception of short stature, other skeletal problems are few, at least if being overweight can be avoided and physical exercise is not forgotten. Possible orthopedic problems must be treated symptomatically. Growth cannot be enhanced by medication. Inconveniences due to shortness can be alleviated by structural amendments in the dwelling surroundings.

Severe hypoplastic anemia may require repeated blood transfusions. Precautions for a fulminant course of infections should be taken. Alleviation for varicella can be tried at the detection of the first vesicles by antiviral medication such as acyclovir. Vaccinations with live or attenuated viral vaccines should be avoided.

Cartilage-hair hypoplasia is a short stature syndrome combined with sparse growth of hair and immunodeficiency.

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Prenatal diagnosis

Intrauterine ultrasound investigation of shortlimbedness is uncertain. A gene test gives the diagnosis in a majority of at risk families.

Historical aspects

Victor McKusick described CHH from the Amish community in the USA in 1964. Jaakko Perheentupa identified CHH as belonging to the Finnish Disease Heritage in 1972.

Epidemiology

In Finland over 170 patients are known; in the Amish community about as many. Some dozens of cases are known elsewhere.

Finnish experts

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Choroideremia

MIM 303100

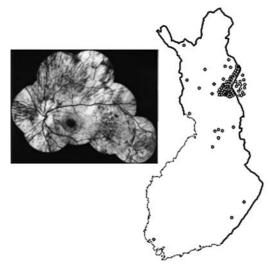


Fig. 4

Choroideremia is an *X*-linked dystrophy of ocular choroidea and retina, which leads to visual handicap at adult age.

Clinical features

The features of choroideremia resemble those of retinitis pigmentosa, with symptoms such as night blindness and

narrowing of visual fields. Visual acuity decreases slowly. Macular visus is maintained for a long time; reading ability may be preserved beyond the age of normal retirement. The severity varies greatly, also intrafamilially.

Diagnostic investigations

The eye fundi show whitish, slowly increasing atrophy of the retinal pigment epithelium and choriocapillaris. It resembles that seen in HOGA, but is not sharply demarcated. ERG will be abolished. The pedigree is compatible with X-linked transmission.

Also carrier females are recognizable through slowly progressive pigmentary fundus changes resembling retinitis pigmentosa. They, however, cause no or only insignificant symptoms. It is a tragic mistake if the doctor, without knowing the exact diagnosis, frightens the carrier female with a proclamation of blindness.

Pathogenesis

The disease process is linked in a complicated way to the so-called rab escort protein (REP1), which regulates the transport of proteins. Many theories of the details have been proposed.

Molecular genetics

The gene, *CHM*, is in the X chromosome at q21.1. Dozens of point mutations and several deletions are known. The main mutation in Finland, splice mutation CHM*SAL (1639+2insT), has not been found elsewhere. Contrary to expectations, there are three different mutations in Northern Finland, and three different deletions have been found in three kindreds in Southern Finland.

Management

The degeneration process cannot be corrected nor delayed. The visual handicap can be aided by several rehabilitative procedures and auxiliary devices. The prognosis is often much better than could be predicted from the ocular fundi. Accurate genetic counselling is important.

Prenatal diagnosis

Prenatal diagnosis has been possible for a long time by a gene test, but its requirement and ethical principles are unclear. The carriers can be detected by the ophthalmoscopy.

Historical aspects

The first published description is that of Mauthner of Austria in 1872. In Finland, extensive investigations have been done by Forsius and his collaborators since the 1960s. The first Finnish publication is by Takki in 1974.

Epidemiology

Choroideremia also often appears outside Finland. About 500 cases have been reported, of which one fourth is from northern Finland.

Finnish experts

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Cohen syndrome

MIM 216550

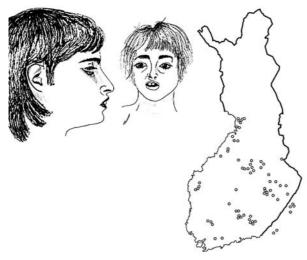


Fig. 5

Cohen syndrome is characterised by mental retardation, typical structural features, slowly progressive visual handicap, granulocytopenia, and cheerful disposition.

Clinical features

Psychomotor retardation is noticed during the first year of life. Muscular hypotonia and hypermobile joints are among the first symptoms. Microcephaly, motor clumsiness and normal or brisk tendon reflexes despite hypotonia are characteristic. The patients learn to walk at two to five years of age and to speak at different ages and in varying amounts. Many patients understand speech better than they can speak themselves. Mental retardation is not progressive. The psychical character is positive, cooperative and sunny. The lifespan may not be shortened.

Typical facial features appear at pre-school age, whereas facial diagnosis in infants is difficult. The patients have

beautiful, high-arched or flame-shaped eyelids, long and thick eyelashes and eyebrows, thick hair and low hairline. The root of the nose is prominent. A short philtrum that is unable to cover the upper central incisors properly is a very typical sign.

The hands and feet, fingers and toes are narrow but short. The height proceeds along the lower limits of the normal curves. Contrary to some textbooks, truncal obesity is not a very typical finding. Genu valgum, pes planovalgus and thoracic kyphosis develop with age.

The main ophthalmologic findings are myopia of the refractive type and chorioretinal degeneration, which proceeds slowly and does not lead to total blindness. Cataracts appear early. Symptoms of retinitis pigmentosa become manifest usually at school age.

Diagnostic investigations

No laboratory finding is diagnostic. However, granulocytopenia is typical but intermittent. It does not harm the patients. Accurate ophthalmological investigations, including ERG, are essential, even if not very easy to do on the retarded patients. MRI of the brain often reveals an enlarged corpus callosum, a very rare finding in mentally retarded patients. Low voltage EEG is found frequently.

Pathogenesis

The pathogenesis of this pleiotropic syndrome is unknown. Whether the enlarged corpus callosum has something to do with the cheerful and social disposition is an interesting but unsolved question.

Molecular genetics

The gene (*COH1*, formerly *CHS1*) is mapped to chromosome 8q22-q23, but not characterised as yet. Concluded from the haplotypes, one main mutation and at least two others will be found in Finland.

Management

Because of their social character, these patients often manage at home and in supervised workplaces. Myopia must be corrected by glasses. Bright lighting is important because of night blindness. The patients benefit from speech therapy.

Prenatal diagnosis

For the time being, prenatal diagnosis may be possible only by aid of linkage analysis in some of the risk families.

Historical aspects

The syndrome was described by Michael Cohen Jr. and his collaborators in 1973. In Finland, similar patients were studied and followed since the 1960s, but the first Finnish report by Norio et al. appeared only in 1984.

Epidemiology

In Finland, over 30 patients are known, the ancestry being mostly from the area of late settlement. The incidence is not greater than 1:100,000, but due to the probably normal survival the prevalence appears greater than that. From other parts of the world more than 100 patients have been described with this diagnosis, but heterogeneity among them certainly exists. Apart from Finland, Cohen syndrome seems to be particularly common in Israel. The haplotypes of Finnish and Israeli patients resemble each other slightly.

Finnish experts

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Congenital chloride diarrhea

CCD (CLD) (Congenital alkalosis with diarrhea) MIM 214700



Fig. 6

Congenital chloride diarrhea is a prenatally manifesting, life-threatening watery diarrhea due to impaired absorption of chloride. After the correct diagnosis, treatment is easy.

Clinical features

The first and constant symptom is polyhydramnios during pregnancy. It is due to intrauterine watery diarrhea, which also washes away the meconium from the bowel. The AFP content of the amniotic fluid may be elevated.

The baby is delivered prematurely, shows excessive weight loss during the first days of life, symptoms of dehydration, and hyperbilirubinemia. The abdomen is protuberant, mimicking ileus. The baby wets the diapers incessantly. When looking for the cause for the excessive voiding it is noticed that the fluid is not urine but it comes from the anus.

If correct diagnosis is not achieved, the infants usually die of severe electrolyte disturbances during the first weeks of life.

Those few who survive fail to thrive and may develop renal and brain damage. After the correct diagnosis, in turn, the treatment is simple and the life of the "patient" is normal, although the stools are permanently slightly looser than normal.

Diagnostic investigations

The chloride content of the fluid coming from the intestine is over 90 mmol/l. In the blood investigations, early changes are hyponatremia, hypochloridemia, and acidosis. Later, the patients develop severe hypokalemic alkalosis associated with hypochloridemia. No chloride is secreted in the urine.

Pathogenesis

The absorption of chloride from the colon and ileum is disturbed because the exchange of intestinal chloride ion into bicarbonate ion of the blood is impaired. The excess of chloride in the intestine causes osmotic diarrhea, the excess of bicarbonate in the blood causes alkalosis. Severe disturbances of fluid balance develop through complicated regulation mechanisms. Over-production of renin and aldosterone causes alterations in renal microanatomy and function.

Molecular genetics

The gene (*CLD*, *DRA*, *SLC26A3*) regulating the anion transport in the intestine is in chromosome 7q31. All the investigated Finnish families have a GGT deletion in nucleotides 951–953, causing a deficiency of one valine molecule (V317del) in the SLC26A3 protein. Elsewhere, over 20 mutations are known in the same locus.

There is some confusion concerning the clinical and genetic abbreviations of this disease. The early clinical

abbreviation was CCD, which I prefer still. The disease and the gene were then named CLD (Cl = chloride), until the gene was noticed to be identical with *DRA* (down-regulated in adenoma) found earlier. Recently, the gene causing CCD was recognised to be a member of *SLC26* anion exchanger gene family and renamed *SLC26A3* (SLC = solute liquid carrier). Thus, it might now be appropriate to leave the abbreviation CLD for another Finnish disorder, congenital lactase deficiency.

Management

After diagnosing CCD, the neonatal fluid balance disturbances are treated conventionally. After that, treatment is a life-long peroral use of sodium chloride and potassium chloride and surplus of water. These supplementary doses must be adjusted so that the blood electrolyte values and pH remain within normal limits and chloride is secreted in the urine. The life-long substitution therapy is important also because interrupting it will lead to renal lesion, even to end stage renal failure. During acute infectious diseases, one must be prepared to avoid fluid balance disturbances. The transport defect proper of the intestine cannot be corrected.

Prenatal diagnosis

In Finland, prenatal diagnosis can be achieved by a gene test, but the need of it is questionable because the treatment of CCD is so easy. If a fetal ultrasound scan is done because of polyhydramnios, distended intestinal loops should lead the thoughts to the diagnosis of CCD immediately after birth.

Historical aspects

First descriptions of CCD were published by Gamble et al. (1945) and by Darrow, also in 1945. The first Finnish publication was by Perheentupa et al. in 1965.

Epidemiology

From Finland, over 50 patients are known. The majority of ancestors come from the late settlement area of the 1500s. From other parts of the world, more than a hundred solitary families are known. In addition, accumulations of CCD like the Finnish one have been reported from Poland (40 patients, main mutation 2025_{2026} insATC = $I675_{676}$ ins) and from Kuwait and Saudi Arabia (total of 50 patients, main mutation 559G>T = G187X).

Finnish experts

Clinical

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Molecular genetics

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Congenital lactase deficiency

CLD MIM 223000





Because the enzyme lactase does not function properly, the newborn baby suffers from severe diarrhea and failure to thrive, both of which are corrected by a lactose-free diet.

Clinical features

The newborn baby manifests with severe watery diarrhea, usually after the first meal. Despite that, the patient is vigorous, hungry, and does not vomit. The weight gain is nonexistent; the weight on admission is below the birth weight. During the lactose-free diet the symptoms disappear rapidly; the prognosis is excellent ().

Diagnostic investigations

The stools are acid and contain reducing sugars demonstrable by Clinitest. The diarrhea ceases by lactose-free (milkless) diet. When the condition of the baby has improved, a peroral lactose loading test can be done. During the test, the blood glucose concentration rises less than 1 mmol/l and diarrhea reappears. A definite diagnosis is achieved by intestinal biopsy: the activity of lactase is minimal or lacking, whereas the structure of intestinal mucosa is normal. Untreated patients may show hypercalcemia of unknown cause and renal calcifications by ultrasound. During the treatment, hypercalcemia disappears but nephrocalcinosis may persist.

Pathogenesis

The function of lactase phlorizin hydrolase is lacking in the epithelial cells of the small intestine (cf. Molecular genetics).

The common lactose intolerance, where the function of lactase enzyme is diminished with age, must not be confused with CLD.

Molecular genetics

The gene locus of CLD is in chromosome 2q21. It is not identical with the lactase enzyme gene (which is already known), but is situated in its vicinity (2 Mb) in the same chromosome. Apparently the gene regulates the lactase gene, and thus the mutated gene makes the ordinary lactase gene inactive. The structure of the mutated gene is not known as yet. According to the haplotype investigations, a major gene is represented in more than 90% of the Finnish families.

Management

The acute disturbances of the fluid balance must be corrected in the conventional manner. The lactose-free diet makes the patients symptomless. At advanced age some patients may tolerate low lactose milk products in which the lactose is hydrolyzed.

Prenatal diagnosis

Prenatal diagnosis may succeed by linkage analysis in some risk families. Its need is questionable because of the benign character and easy management of the disease.

Historical aspects

The first description is by Holzel et al. in 1959, the first Finnish publication by Launiala et al. in 1966. I may have treated the first known Finnish patient in 1962.

Besides congenital lactase deficiency, the abbreviation CLD has been used for congenital chloride diarrhea. About the settling of this confusion, see the section preceding this one on congenital chloride diarrhea.

Epidemiology

Over 40 patients are known in Finland, which is more than from other countries altogether. The patient series of 16 cases published in 1983 by Savilahti et al. was overwhelmigly the largest published so far.

Finnish experts

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Molecular genetics

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Congenital nephrosis of the Finnish type

CNF, (NPHS1) MIM 256300

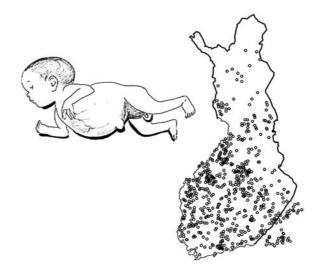


Fig. 8

Congenital nephrosis of the Finnish type is a nephrosis which manifests in newborns and without renal transplantation is lethal.

Clinical features

As sign of the intrauterine origin of the disease, the placenta is always large, more than 25% of the birth weight. The baby is often born prematurely. Proteinuria is seen in the first specimen investigated. Edema manifests in the first days or weeks of life. The patient fails to thrive, abdomen is protuberant, hernias are common, infectious diseases occur frequently. The patients die during the first or second year of life without learning to walk. The cause of death is not renal insufficiency but infection; often the immediate cause remains unknown.

The course and prognosis, however, have changed totally from the description above because of renal transplantation.

Diagnostic investigations

A large placenta and proteinuria in the first urinary specimen are important diagnostic findings. Serum protein changes are those typical of nephrosis. Serum creatinine is normal. Renal histopathology reveals tubular dilatations after the first three months of life. In electron microscopy, a fusion of foot processes of the glomerular epithelial cells, podocytes, is seen.

Pathogenesis

Protein leakage through the glomerular filter begins in utero, as soon as the urinary secretion begins. The damaged cell adhesion protein nephrin located at the slit diaphragm between glomerular podocytes has a central role in this leakage. The basement membrane is intact. The damage of the slit diaphragm also explains the electron microscopic finding of fusion of the foot processes of the podocytes. The lethal failure to thrive of the infants was caused by starvation due to a severe protein leakage into the urine.

Molecular genetics

The gene *NPHS1* is in chromosome 19q13.1. Mutation Fin major, a deletion of two nucleotides (121-122delCT), is found in 78% of the Finnish disease chromosomes. Mutation Fin minor, a point mutation (3325C>T = R1009X) is represented in 16% of the Finnish disease chromosomes. Dozens of other mutations have been found in other populations and some others also in Finland. The clinical features caused by different mutations and their combinations may cause different clinical courses. Although even most of them are severe, some milder CN cases as published from different populations may appear and even respond to conventional treatment. Thus, a nephrotic infant with a compound heterozygous genotype Fin major/2227C>T expressed nephrin and responded to enalapril and indomethacin therapy.

Management

The traditional treatment of nephrosis with corticosteroids and immunosuppressive drugs is without effect in CNF. Renal transplantation has changed the prognosis completely and allowed a normal life, though with continuous immunosuppressive treatment. Before transplantation the infants must gain weight up to 9 kg. This is possible by forced, partly parenteral nutrition, albumine substitution, peritoneal dialysis and often by removal of their own kidneys. Some patients homozygous for the Fin major gene develop nephrosis in the grafted kidney, apparently due to autoimmune reaction against nephrin.

Prenatal diagnosis

In risk families prenatal diagnosis has been possible for years by aid of a hugely (>10 SD) elevated AFP concentration in the amniotic fluid. The massive overproduction of this fetal protein into amniotic fluid is due to heavy fetal proteinuria. In Finland, AFP investigation from maternal serum has been used as a regional screening study.

Recently, it has been found that the CNF gene is able to cause a massive AFP concentration in the amniotic fluid also in heterozygotes. This fetal proteinuria vanishes during pregnancy and the baby is born healthy. Thus, elevated AFP concentration in the amniotic fluid must be controlled. A lower value in the second sample probably points to a healthy fetus.

Today, gene tests solve the problems of prenatal diagnosis in Finland. In other countries this matter is not so straightforward. Moreover, many cases of more or less congenital nephrosis may not present with elevated AFP, nor even with large placenta — but then the prognosis may also be better than in CNF.

Historical aspects

The role of the first report of CN has been given to the description by Gautier and Miville in 1942. The first Finnish article is by Hallman et al. in 1956. CNF is considered to be the prototype of Finnish disorders (see Part I, DOI s00439-002-0875-3), which has given rise to at least 14 thesis investigations, among others.

Epidemiology

More than 300 Finnish patients are known, from elsewhere less than that. The incidence in Finland is 1:10,000 at least. The bulk of ancestors come from the area of late settlement, but several ancestors even come from other parts of Finland. An accumulation of patients is known in Minnesota, USA, a region of active Finnish immigration.

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Cornea plana congenita

CPC, CNA2 MIM 217300

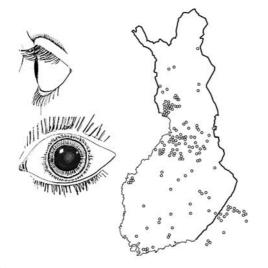


Fig. 9

Cornea plana congenita is an ocular disease causing moderate visual handicap.

Clinical features

The cornea is congenitally "flat", which means that its radius is longer and thus its curvature is slighter than normal. The cornea is also thin and its small refractive power appears as hyperopia. The central opacities of the cornea disturb the visual acuity and a peripheral arcus senilis develops early. The risk for closed angle glaucoma is increased. Systemic findings do not exist.

Diagnostic investigations

An accurate ophthalmologic investigation, including slit lamp inspection, discloses the diagnosis.

Pathogenesis

The detailed pathogenesis is not known. The structure of the gene mutation indicates that the cooperation of certain collagens with small protein-rich proteoglycans is disturbed.

Molecular genetics

The gene, *KERA*, is in chromosome 12q22. The Finnish mutation common to all the studied patients is a point mutation AAC>AGC at codon 247, causing an alteration, N247S, in a leucine-rich proteoglycan called keratocan. At least two other point mutations have been documented.

Management

Hyperopia must be corrected by glasses. At school, the patients usually manage in a normal class.

Prenatal diagnosis

Prenatal diagnosis is possible by a genetic test at least in Finland, but selective abortion may not be ethically justi-fied.

Historical aspects

Rübel of Germany described the disease in 1912. In Finland, extensive investigations have been carried out by Henrik Forsius and his collaborators since the 1950s. His short congress report from 1957 is the first Finnish publication.

Epidemiology

In Finland, about 90 patients are known; from elsewhere considerably less have been published. The disease is concentrated in northern Finland.

Also a dominantly inherited form of CPC is known. Its gene is situated in the vicinity of the *KERA* gene but is not identical with it.

Finnish experts

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Diastrophic dysplasia

DD, DTD MIM 222600

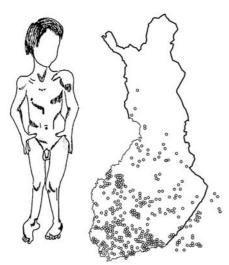


Fig. 10

Diastrophic dysplasia is a congenital short-limbed bone dysplasia, causing deformations and dysfunctions of the joints.

Clinical features

The limbs are found to be short at birth. Other signs visible in newborns are stiffness of the big joints, partial flexion limitations of the finger joints, club-foot-like malformation, cleft palate, and swollen ear lobes that later develop cartilagineous deformities. The often mentioned "hitchhiker's thumb" may not easily be visible in newborns.

With age, deformed knee and hip joints lead to motility restrictions, malpositions, and early arthrosis. Scolioses and kyphoses may be severe. Adult height is 100–160 cm. The severity of the deformations shows great variability, even intrafamilially.

Diagnostic investigations

Diagnosis by inspection is easy to those who know the disease. In X-ray, tubular bones are short and thick, knee and hip joints are broad and flattened, and metatarsal bones spread out forming a metatarsus adductus deformation.

Pathogenesis

The pathogenetic background has been revealed along with the characterisation of the gene. The transport of sulphate into cartilagineous cells is hampered, proteoglycans suffer from lack of sulphate and the growth of cartilage and bone is disturbed.

Molecular genetics

The gene is in chromosome 5q32-q33. It was originally named diastrophic dysplasia sulphate transporter, *DTDST*. In Finland, the main mutation DTDST_{Fin} (c.–26+2T>C) was found in over 90% of the disease chromosomes. Also, two other mutations (262C>T = R279W and 1045– 1047delGTT = V340del) are known in Finland, whereas in 4% the mutation is unknown. Elsewhere, about 20% of disease chromosomes have the DTDST_{Fin} mutation, but dozens of other mutations are also known. Recently, it has been recognised that the *DTDST* gene belongs to an anion exchanger gene family called *SLC26*. Thus, the gene has been renamed *SLC26A2*.

Different mutations in the *SLC26A2* gene cause two more difficult skeletal diseases, atelosteogenesis type II and achondrogenesis type IB, and one less severe disorder, viz. multiple epiphyseal dysplasia.

Management

The quality of life of the diastrophic patients depends not so much on short-limbedness as on restricted mobility, deformations and early arthrosis of the joints. Almost all patients need orthopedic operations, including hip prostheses. Physiotherapy is important in order to maintain mobility and to prevent malpositions. Deformations of the upper cervical spine may be life-threatening if the neck is moved into abnormal positions, e.g., in general anaesthesia. The structure of cervical spine must be investigated preoperatively by X-ray.

Prenatal diagnosis

Shortness of the limbs may be seen by ultrasound by the 16th–18th week of pregnancy. At least in Finland, prenatal diagnosis can be achieved earlier and reliably by a gene test from the chorionic villi.

Historical aspects

The first published description of a DD patient is that by Arraga of Argentina in 1897 (see Lapunzina et al. 1998). Lamy and Maroteaux in 1960 separated DD as an entity of its own. In Finland, Perheentupa identified and introduced DD in 1972.

Epidemiology

In Finland, nearly 200 patients are known. DD is not very rare even elsewhere: at least 250 patients have been pub-

lished outside Finland. This is in concordance with the exceptional, southwestern distribution of DD ancestors in Finland. Apparently the gene was brought to Finland thousands of years ago by western immigrants (Part I, DOI s00439-002-0875-3).

Finnish experts

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FSH-RO

Follicle stimulating hormone-resistant ovaries Hereditary hypergonadotrophic ovarian failure Aittomäki disease MIM 233300



Fig. 11

FSH-RO is a deficiency of the female sex hormones presenting with primary amenorrhea and sterility.

Clinical features

Female patients are symptomless until the appearance of primary amenorrhea and sterility. Secondary sexual characteristics are normal or subnormal. The ovaries are small and fibrotic.

(Homozygous men are mostly subfertile: the amount and quality of sperm cells may be subnormal and the testicles are small.)

Diagnostic investigations

The concentration of follicle stimulating hormone (FSH) is high, >40 IU/l, whereas the values of female sex hormones are low. Karyotype is normal XX.

Pathogenesis

The ovaries contain oocytes, but they do not mature despite the overproduction of FSH, because the FSH receptor in the ovarian follicles is defective. Because the follicles do not develop normally, their production of female hormones is not functioning.

Molecular genetics

The gene (*FSHR*) was mapped to chromosome 2p16-p21. It proved to be the already known gene of the FSH receptor, *ODG1*. The Finnish patients have the point mutation 566C>T, which changes alanine to valine in position 189 of the receptor protein. In one Finnish chromosome, a gene with the mutation 1255G>A produces a change A419T in the receptor protein. Apparently several mutations, even in several loci, may cause primary hypergonadotropic amenorrhea with the XX karyotype.

Management

As yet, no treatment is able to normalise the patient's own egg cells nor the production of sex hormones. Menstruation can be started and secondary sex characteristics improved by sex hormone supplementation. This is important also in order to prevent osteoporosis and coronary heart disease. Patients can even become pregnant by aid of ovum donation.

When there are problems with becoming pregnant in the families of the female FSH-RO patient's brothers, this diagnosis of these brothers can be verified or excluded. The fertilising ability of FSH-resistant men can be aided by means used in fertility therapy.

The patients may have considered the sterility problem so delicate that they have not discussed it, even with their siblings who possibly suffer from the same problem. The patients should be encouraged to deal with the problem openly and without shame.

Prenatal diagnosis

Because of the nature of the problem there may be no place for prenatal diagnosis, although a diagnostic gene test is available at least for some patients.

Historical aspects

Kristiina Aittomäki of Finland worked out the whole problem from patient series and genealogical investigations to the gene locus and structure of the mutation in the 1990s.

Epidemiology

Among the over 70 Finnish patients with primary hypergonadotropic amenorrhea and XX karyotype, one third has the Finnish *ODG1* mutation, one third does not have it, and one third has not been investigated as yet. For the time being, this mutation has not been found in other countries. In Finland, the ancestors are mainly from northern Finland.

Finnish expert

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GRACILE syndrome

(<u>G</u>rowth <u>r</u>etardation, <u>a</u>minoaciduria, <u>c</u>holestasis, <u>i</u>ron overload, <u>l</u>actic acidosis, <u>e</u>arly death) Fellman syndrome Lethal lactic acidosis with hemosiderosis MIM 603358

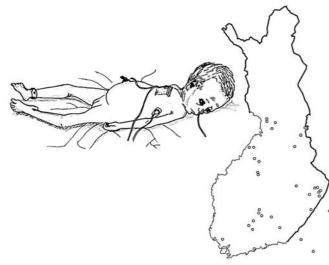


Fig. 12

This neonatally lethal disease causes lactic acidosis resistant to all treatment and accumulation of iron in the liver.

Clinical features

The patients are born extremely small for date. In the first day of life a severe lactic acidosis develops that usually cannot be corrected, even with huge amounts of bicarbonate. The facial expression seems "worried". No neurological or other specific symptoms are revealed. No weight gain appears. The cachectic and acidotic patients succumb at the age of some days or weeks.

Diagnostic investigations

In addition to a fully exceptional acid-base balance the patients show aminoaciduria. The investigations of iron metabolism give abnormal results: serum ferritine concentration is elevated ten-fold, transferrine is mostly saturated but its concentration is low, and free iron appears in the serum.

At autopsy, severe pathological changes with cholestasis are seen in the liver. Hemosiderin is stored in the hepatocytes and in Kupffer cells of sinusoids. Iron deposits can be seen also in the macrophages of the spleen and pulmonary alveoli. Interstitial fibrosis and exocrine atrophy of pancreas and calcifications of the renal medulla are usual findings.

Pathogenesis

How the lactic acidosis and disturbed iron metabolism are associated is not known yet.

Molecular genetics

The gene is in chromosome 2q33-q37. All Finnish patients are homozygous for a missense mutation 232A>G (S78G) in the *BCS1L* gene. The mitochondrial innermembrane protein BCS1L is known as a chaperone necessary for the assembly of mitochondrial respiratory chain complex III. In the GRACILE syndrome, the mutation apparently disturbs some other function of BCS1L whereas complex III activity is within the normal range.

Management

At present, no treatment has been able to keep the patients alive. Attempts were made to diminish the iron load of the organism by apoferritine and repeated exchange transfusions. The values of iron metabolism shifted towards normal, but the patients died nevertheless.

Prenatal diagnosis

Prenatal diagnosis has been done in one Finnish risk family by linkage and haplotype analysis. Now the diagnosis is possible by gene test in Finland, whereas sequence analysis of the *BCS1L* gene offers a diagnostic option for non-Finnish families.

Historical aspects

The first patients astonished the doctors at the Children's Hospital, University of Helsinki in the 1960s. However, they escaped sufficient attention until Vineta Fellman and her collaborators began to penetrate this problem in the

1980s. The storage of iron was then noticed, and the disease was described as a novel entity in 1998.

Epidemiology

In Finland over 20 patients are known; in Sweden one family with Finnish ancestry. The roots of the Finnish families are mainly in the area of late settlement. In three British patients four other mutations and in four Turkish families also four mutations of the *BCS1L* gene have been found. The phenotypes of these patients differ considerably from the Finnish ones.

Finnish experts

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Herva disease

Lethal congenital contracture syndrome LCCS Multiple contracture syndrome, Finnish type (Cf. Lethal arthrogryposis with anterior horn cell disease = *Vuopala disease*) MIM 253310

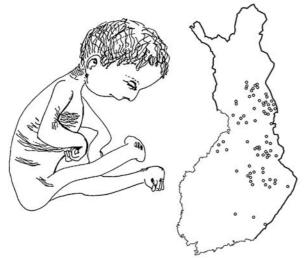


Fig. 13

Due to a fetal deficiency of the anterior spinal cord, severe intrauterine contractures have developed in the stillborn patient.

Clinical features

The fetus grows slowly and moves poorly or not at all. Profuse hydrops develops in the first half of pregnancy. Severe contractures and malpositions of the big joints appear, usually flexion contractures in the elbows and extension in the knees. On the flexor side of the elbow, a skin fold, pterygium, is often formed. The muscles are almost nonexistent. The long bones and ribs are extremely thin and may be fractured in utero or at delivery. The facial features are abnormal with hypertelorism, low set ears, micrognathia, and flat nose. The lungs are hypoplastic. The umbilical cord is often short. The pregnancy ends in the stillbirth of a small-for-date fetus.

Diagnostic investigations

The sick fetus cannot be helped, but achieving the right diagnosis is essential for the future pregnancies of the family. Photos and autopsy are truly important. At autopsy, spinal cord and muscle samples must also be investigated.

Pathogenesis

The syndrome described above is called fetal akinesia deformation sequence or Pena-Shokeir phenotype. It is an outcome of the immobility of the fetus due to different causes. In LCCS, motor neuron cells in the spinal cord are lacking almost entirely and those that exist are calcified remnants. The whole anterior spinal cord is atrophic. The pathogenesis of the spinal damage and the mechanism of intrauterine death are not known.

Molecular genetics

The gene *LCCS* is situated in chromosome 9q34, at a locus different to that of the gene for spinal muscular atrophy (Werdnig-Hoffman disease). The structure of the gene is not known yet, but haplotype analysis speaks in favour of one Finnish major mutation.

Management

The disease cannot be treated nor prevented. Prenatal diagnosis is an important aid to the families.

Prenatal diagnosis

Ultrasound may reveal the disease as early as in the 14th week of pregnancy.

Historical aspects

The disease was first documented by Riitta Herva and coworkers of Oulu, northern Finland in 1985.

Epidemiology

In Finland, over 60 patients are known in Kainuu and northern Savo. In other countries some patients resembling LCCS have been published, but differential diagnosis outside Finland is difficult. In Finland a similar, slightly less severe disease exists, with possible survival of some days or weeks; this is called Vuopala disease or LAAHD.

Finnish expert

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HOGA

Hyperornithinemia with gyrate atrophy of the choroid and retina GA

MIM 258870

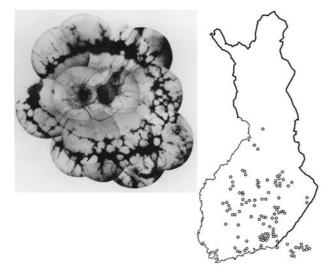


Fig. 14

HOGA is an ophthalmological disease with a disturbance of the amino acid metabolism, leading gradually to blindness.

Clinical features

Myopia and symptoms resembling retinitis pigmentosa often begin before ten years of age and progress into blindness by 40 years of age. Cataract may develop before 20 years of age. Mild muscular weakness can occur.

Diagnostic investigations

In ophthalmoscopy, retinochoroidal atrophy appears as whitish, sharp-edged patches, beginning in the periphery and spreading and confluing over the whole eye fundus. ERG abolishes early. The ornithine concentration in plasma is increased $(10-20 \times \text{normal values})$ and ornithine is also secreted abundantly in the urine. Enzyme and gene diagnosis are also possible, but because of the pathognomonic eye finding and hyperornithinemia they are seldom needed in practice.

Pathogenesis

The activity of the enzyme ornithine aminotransferase is almost totally lacking. Because of failure of the degradation of ornithine its concentration in tissues, blood and urine is increased. How the enzyme defect affects the eye is not known. In the muscle biopsy, atrophy of type 2 muscle cells and tubular aggregates in electron microscopy are seen.

Molecular genetics

The gene *OAT* is situated in chromosome 10q26. Its structure and about 80 different mutations are known. Among the Finnish patients, 90% have the same mutation 1205T>C= L402P not found in other countries. The remaining 10% carry mutation R180T or some unknown mutation. The mode of function of the mutated gene is not known.

Management

Different treatments affecting the ornithine metabolism have been tried. Results as to vision have been scanty, but biochemical abnormalities tend to diminish. In transgenic mice some prevention of eye fundus alterations has been achieved. Muscular symptoms can be removed with creatine.

Preliminary data suggest that arginine-deficient diet normalises plasma ornithine concentration and may prevent the progression of the fundus changes, at least when started early in life. Thus, low-arginine diet is recommended for the patients, although its maintenance is not easy.

Early cataract operation, often in the third decade of life, improves the vision considerably.

Prenatal diagnosis

Prenatal diagnosis is possible with enzyme determination and, at least in Finland, by a gene test. However, the family size is usually final, as the diagnosis of first affected child is reached. The ethical principles of prenatal diagnosis in diseases causing blindness in adulthood have not been established as yet.

Historical aspects

The ophthalmological findings of the disease have been known since the 1800s (Jacobsohn 1888; Cutler 1895). The

association of hyperornithinemia was found by Simell and Takki of Finland in 1973.

Epidemiology

In Finland, over 80 patients are known, most of them descending from the late settlement area. The disease is not very uncommon elsewhere, either.

Finnish experts

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Hydrolethalus syndrome

MIM 236680

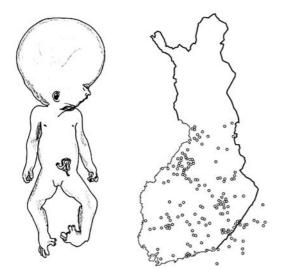


Fig. 15

Hydrolethalus syndrome is a combination of several severe congenital malformations leading to perinatal death.

Clinical features

The pregnancy is complicated by polyhydramnios. The baby is stillborn or dies in the first day of life. The most prominent abnormality is a huge hydrocephaly. Other findings are listed in the Diagnostic investigations.

The syndrome is usually revealed in the intrauterine ultrasound scan due to the atypical hydrocephaly. In such case most families choose an abortion, and that is why only a few full-term newborns are currently born.

Diagnostic investigations

Autopsy with accurate photos is of utmost importance for the genetic counselling. The lateral brain ventricles are wide open, and the remnants of the brain lacking midline structures are surrounded by cerebral fluid. The foramen occipitale magnum is not roundish but keyhole shaped. The eyes are deep-set and small. The nose is small or bifid, the chin is small or nonexistent, cleft lip and/or palate may occur. The tongue, larynx, trachea and bronchi may be deformed. The lobulation of the lungs is abnormal. Heart anomaly is often of canalis atrioventricularis communis type. The abdominal organs are usually normal and there is no cystic liver nor kidneys in contrast to the Meckel syndrome. The genitals may show slight abnormalities such as a double uterus, but there is no confusion about the sex. Polydactyly (postaxial in fingers, preaxial in toes) and club-foot-like deformity are common.

As is usual in syndromes, all the described abnormalities may not appear in every patient.

Karyotype analysis is important, because especially trisomy 13 may resemble the hydrolethalus syndrome.

Pathogenesis

Nothing is known about the pathogenesis of this pleiotropic syndrome. However, it has many features in common with midline field defects.

Molecular genetics

The gene is situated in chromosome 11q23-q25. Its structure is not known as yet. Judging by the haplotype in common, all the Finnish patients have the same mutation.

Management

Achieving the right diagnosis is the best aid for the future children of the family.

Prenatal diagnosis

Prenatal diagnosis may succeed in risk families, even in the 12th week of pregnancy, by ultrasound scan. Abnormal structure of the brain is noted at the 18th week at the latest, although the overgrowth of the skull may not become visible before the 20th week.

Historical aspects

The syndrome was separated from the Meckel syndrome as late as in 1981 by Salonen et al. in Finland. The name of the syndrome was put together from its three most prominent features: (poly)hydramnios, hydrocephaly, and lethality.

Epidemiology

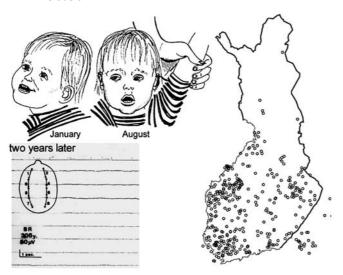
In Finland over 80 patients are known, most with the ancestry from the late settlement area. From other parts of the world about ten cases have been reported with this diagnosis, but most of them show atypical features. This may mean that the disease is really rare elsewhere, especially as it has been described frequently and well in the literature.

Finnish expert

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INCL

Infantile neuronal ceroid lipofuscinosis Santavuori-Haltia disease CLN1 MIM 256730





See also *Neuronal ceroid lipofuscinoses*

INCL manifests itself soon after the first year of life and rapidly leads to an extraordinarily profound mental and motor retardation.

Clinical features

The early development is normal. Around its first birthday the infant gradually begins to lose the gained psychomotor skills. The rapid deterioration is accompanied by hyperexcitability, crying, sleeping disturbances, myoclonic jerks, and knitting hand movements. In the eye fundi, signs of progressing atrophy are seen. Microcephaly is an early finding. Blindness and spasticity belong to the end state. The average survival is ten years.

Diagnostic investigations

EEG shows an exceptional reaction to opening and closing of the eyes, loss of sleep spindles and isoelectricity by three years of age. Also ERG, VEP and SEP become extinguished. MRI of the brain shows hypointense signals in the thalami and hyperintense signals in the periventricular white matter, already present by 7–8 months of age, viz. before the appearance of the clinical symptoms. SPECT shows hypoperfusion especially in the cerebrum and cerebellum.

Electron microscopic investigation from rectal mucosal biopsy shows granular osmiophilic deposits (GROD) in the cytosomes. The autopsy shows extraordinary atrophy of the brain: neurons, axons and myelin sheats have disappeared, whereas astrocytic glial tissue is left with fluorescent lipopigment deposits in the brain and other organs.

Pathogenesis

The activity of the lysosomal enzyme palmitoyl-protein thioesterase (PPT) is missing. Details of the destruction of the brain cells are not known. The abnormal deposit material consists mainly of lysosomal sphingolipid activator proteins called saposins A and D.

Molecular genetics

The gene (*CLN1* or *PPT1*) is in chromosome 1p32. Of the Finnish patients 98% has mutation 364A>T changing arginine to tryptophan in the PPT enzyme, position 122. Over 30 other mutations are known elsewhere.

Management

Curative treatment does not exist, but hopes are raised by the advancement of the molecular genetic investigations. Trials with bone marrow transplantation with normally functioning PPT in the monocytes may have delayed the progression of the disease.

The patients need active physiotherapy, alleviation of pain and other symptomatic therapy.

Prenatal diagnosis

In no disease is prenatal diagnosis needed more urgently than in INCL. This has been possible for a long time through GROD alterations in the chorionic villus cells in the first trimester of the pregnancy. In Finland, gene test is the primary choice, whereas PPT enzyme assay is suitable for all patients irrespective of the mutation.

Historical aspects

The disease was described the first time by Hagberg et al. of Sweden in 1968. The first report from Finland was published in 1973 by Santavuori, Haltia, Rapola and Raitta. If in the past the diagnosis of Tay-Sachs disease (which is extremely rare in Finland) had been set in Finland, the correct diagnosis would probably have been INCL.

Epidemiology

In Finland over 160 patients are known, elsewhere more than 200. The ancestors have three accumulation regions, two in the area of late settlement and one, exceptionally, in southwestern Finland.

Finnish experts

Clinical

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IOSCA syndrome

Infantile <u>onset spinoc</u>erebellar <u>a</u>taxia Former name OHAHA syndrome = <u>o</u>phthalmoplegia-<u>h</u>ypoacusis-<u>a</u>taxia-<u>h</u>ypotonia-<u>a</u>thetosis MIM 271245

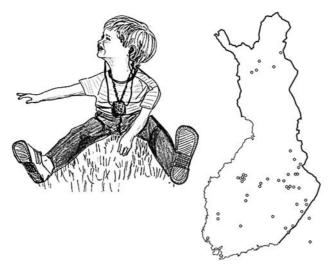


Fig. 17

IOSCA syndrome is a multisymptomatic neurological disease with infantile onset and severe, progressive handicap.

Clinical features

The neonatal period and development of the first year of life are normal. Soon after that, clumsiness, disturbances of gait, hypotonia, ataxia and athetosis appear. Tendon reflexes become abolished. Deterioration of hearing and ophthalmoplegia begin by school age. Variations of vigilance appear. By puberty, symptoms from peripheral nerves develop: tactile, proprioceptive and kinesthetic sensations become impaired and Babinski's sign becomes positive as a manifestation of damage of long corticospinal tracts. Optic atrophy develops. Injury of autonomous nervous system presents as sweating and functional disturbances of the bowel and urinary bladder. The pubertal development of girls is deficient, showing amenorrhea and underdevelopment of secondary sex characteristics.

New symptoms often appear in association with common infectious diseases. Young adults are wheelchair bound. They also may get severe epileptic crises with abdominal pains and vomiting. These attacks can be lifethreatening. The oldest Finnish patients are in their forties.

Diagnostic investigations

No investigations are pathognomonic. Neurological tests may reveal a slowing of conduction velocity of the sensory nerves and abnormal SEP. With age cerebellar and brain stem atrophy develop.

Pathogenesis

The pathogenesis is unknown. Although the clinical features resemble those of mitochondrial diseases, no investigations are in favour of such etiology so far.

Molecular genetics

The gene *SCA8* is situated in chromosome 10q24. It is not characterised as yet, but haplotype analysis reveals that all Finnish patients have one and the same mutation.

Management

There is no cure. The right diagnosis helps to direct symptomatic treatment and rehabilitation properly. These patients with hearing deficit and athetosis may be misdiagnosed as mentally retarded.

Prenatal diagnosis

Prenatal diagnosis may be possible in Finland by linkage analysis.

Historical aspects

The disease was first described in Finland by Kallio and Jauhiainen in 1985 as OHAHA syndrome (see former name above). As the patients grew older, the cerebellar and sensory symptoms became apparent. The disease was renamed as IOSCA in order to point out that it is a clearly defined entity in the entangled group of spinocerebellar ataxias.

Epidemiology

In Finland over 20 patients are known, most of them from eastern Finland. The disease is not described from other countries.

Finnish expert

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Jansky-Bielschowsky disease, Finnish variant

Variant late infantile neuronal ceroid lipofuscinosis JB, vLINCL_{Fin}, CLN5 MIM 256731



Fig. 18

See also Neuronal ceroid lipofuscinoses

This disease of the central nervous system begins at preschool age and leads to blindness and profound psychomotor retardation.

Clinical features

The development in the first few years of life is normal. At the age of 4–7 years, clumsiness, visuomotor problems and mild mental retardation appear. The first misdiagnosis at that age may be MBD syndrome. At school age, the

mentioned symptoms become worse, and epileptic seizures, myoclonic jerks, ataxic gait, and difficulties in speech appear. Problems in the activities of daily living develop. Eye fundi show signs of atrophy of the retina and optic nerves. All patients become blind by ten years of age. They lose the ability to speak and move, become spastic and deeply retarded. The life expectancy is 20 (14–40) years.

The classical LINCL differs from the Finnish variant by more rapid progression: beginning at 2–4 years, continuation at 5–6 years, death at 10–20 years.

Diagnostic investigations

From imaging investigations of the brain, CT is useless, as in all NCL disorders. In MRI, hypointense signals are seen in the thalamic region, hyperintense signals in the white matter, and very early appearing atrophy in the cerebellum. SPECT shows cerebellar hypoperfusion. In EEG, basal activity is disturbed and low-frequency flash causes occipital spikes. Typical neurophysiological findings of giant flash-VEP and SEP appear at 7–9 years of age. ERG becomes isoelectric by seven years. Thus, in contrast to the early appearing cerebellar atrophy, most of the electrophysiological investigations do not reveal the disease until several years after its beginning.

In the chorionic villi, the rectal mucosal biopsy, and in the brain specimen at autopsy, s.c. curvilinear bodies and fingerprint profiles are seen in the cytosomes. Vacuoles in the leukocytes, such as in Spielmeyer-Sjögren disease, are not seen. In classical LINCL curvilinear bodies, but no fingerprint patterns, are found. The gene test is an early, easy, and reliable diagnostic tool in Finland.

Pathogenesis

The basic mechanism may be associated with the defective lysosomal trafficking.

Molecular genetics

The gene, *CLN5*, is in chromosome 13q22. It has been characterised. In Finland, all the families except one have mutation 2467delAT = Y392X. The remaining family has mutation 1517G>A = W75X.

Management

Curative treatment does not exist. Symptomatic treatment and rehabilitation are important, not least in order to offer the patient a good quality of life for as long as possible.

Prenatal diagnosis

Prenatal diagnosis can be done by electron microscopy from chorionic villi in the 14th–15th week of pregnancy. Gene diagnosis is safer and can be done earlier in Finland.

Historical aspects

The first publication was from Finland, by Santavuori et al. in 1982.

Epidemiology

From Finland over 30 patients are known. All ancestors come from a small area in western Finland, where the incidence is 1:1,500 and carrier frequency according to gene tests 1:25. The Finnish variant is known in one Dutch and one Swedish family. Other variants of LINCL have been described outside Finland in about 100 patients.

Finnish experts

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Lysinuric protein intolerance

LPI MIM 222700



Fig. 19

In this metabolic disorder tolerance for proteins is poor, but the disease can be treated and tolerated reasonably well.

Clinical features

The infants tolerate mother's milk well. The shift to formulae containing bigger amounts of proteins brings on vague intolerance symptoms: poor appetite, failure to thrive, vomiting, diarrhea. If protein feeding continues, the symptoms become worse. Gain of weight and height is poor, muscles are thin, fractures appear easily due to osteoporosis; proneness to infections and hepato- and splenomegalia are found. Meals rich in protein may cause ammonium intoxication, manifesting itself as comatous central nervous symptoms, and even as mild mental retardation. On the other hand, the patients develop an aversion to protein, which protects them against worsening of the symptoms, if they are not forced to eat protein-rich meals.

Recently it has been noticed that some adult patients develop odd pulmonary complications: acute or chronic respiratory insufficiency, pulmonary fibrosis, proteinosis or hemorrhages. Also renal damage simulating glomerulonephritis may appear and sometimes lead to renal insufficiency.

Diagnostic investigations

The urinary excretion of dibasic amino acids lysine, arginine and ornithine is increased and their concentration in the plasma is diminished. As accessory findings, anemia, leukopenia, thrombocytopenia and increased serum concentrations of lactate dehydrogenase and ferritine may appear. A reliable diagnosis can be achieved in Finland by a gene test.

Pathogenesis

The basic disturbance is in the transportation of dibasic amino acids through the basolateral membrane of intestinal epithelial cells. In renal tubuli the reabsorption of these amino acids is also deficient. Thus, the organism suffers from the deficiency of lysine, arginine, and ornithine. The lack of ornithine disturbs the function of the urea cycle, which should metabolize ammonium, toxic for the organism, to harmless urea. The lack of lysine probably causes growth retardation.

Molecular genetics

The gene, *LPI* or *SLC7A7*, is in chromosome 14q11.2. It encodes a protein called y+LAT-1 needed for the transporting of dibasic amino acids. All the Finnish patients studied have had the splice site mutation $LPI_{Fin} = 1181-2A>T$. This has not been found in other populations, where more than ten other mutations in the same locus have been detected.

Management

Peroral administration of neutral amino acid citrulline supports the function of the urea cycle. Then, ammonium becomes degraded, ornithine concentration in plasma rises and symptoms are alleviated. Even so, a diet moderately low in protein is needed. Treated in this way, the patients remain symptomless, at least if pulmonary or renal complications do not appear later. Psychic development is normal, although hyperammonemia may cause difficulties in mathematics and other kind of abstract thinking for some patients.

Prenatal diagnosis

Prenatal diagnosis, easily done in Finland by a gene test, is hardly needed because of easy treatment and good prognosis. Whether the threat of pulmonary and renal complications alters this consideration, remains to be seen.

Historical aspects

The first description was given by Perheentupa and Visakorpi of Finland in 1965. In the beginning, the name of the disorder was familial protein intolerance with deficient transport of basic amino acids.

Epidemiology

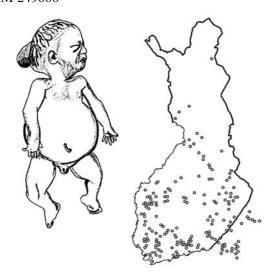
In Finland about 50 patients are known, elsewhere over 100. The chain of Finnish ancestors reaches from southern Savo through eastern Finland to Lapland.

Finnish expert

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Meckel syndrome

Former names: Gruber syndrome; Dysencephalia splanchnocystica MIM 249000



Clinical features

Typical components of the neural tube defect are microcephalus and encephalocele, whereas hydrocephaly is rare (cf. *Hydrolethalus syndrome*). Polydactyly is postaxial. Kidneys and liver are dysplastic, cystic and so large that they may make a vaginal delivery impossible. Among other possible malformations are cleft lip and palate, congenital heart defects, ambiguous genitalia, and club feet. The placenta may be large. The patients are stillborn or survive for some hours.

Today, at least in Finland, very few patients are born after a full-term pregnancy because the malformation is verified in the second trimester ultrasound scan screening and most families choose an abortion.

From other countries, patients claimed to have Meckel syndrome have been reported with more or less different manifestations and longer survival. The problems of heterogeneity cannot be solved before gene diagnosis is available.

Diagnostic investigations

An accurate description of the malformations, photos, and autopsy are important for the correct diagnosis. Contrary to polycystic diseases of the kidney, normal nephrons are lacking almost totally (multicystic dysplasia). In the liver, microscopical fibrosis, in the fetus ductal plate malformation is always seen. Chromosome anomalies should be exluded.

Pathogenesis

The pathway from the mutated gene to the phenotype is not known.

Molecular genetics

Most of the Finnish patients are homozygous for the gene MKS1 in the chromosome 17q22-q23. It is not characterised as yet. According to haplotypes, about 75% of the Finnish patients have one and the same mutation and possibly five other mutations exist. In other populations, the Meckel locus is not always the same. Another known locus MKS2 is at 11q13 in patients (fetuses) from Northern Africa and the Middle East. A third locus (MKS3) is reported recently at 8q24 in Indian and Pakistani populations.

Management

The patients cannot be helped. The delivery must happen without endangering the mother. It is imperative to distinguish Meckel syndrome from other, incidental malformation complexes because of the recessive recurrence risk and possibility for prenatal diagnosis.

Prenatal diagnosis

In case of an open neural tube defect, AFP concentration in the amniotic fluid may be increased. In an accurate ultrasound scan, the cranial malformations and even enlargement of the kidneys may be distinguishable already in the 12th week of pregnancy. Later, the amount of amniotic fluid decreases and may totally disappear by the 18th week. This makes diagnosis by ultrasound difficult.

Historical aspects

Meckel described "his" syndrome as early as in 1822. In Finland, the first publication was by Aula et al. in 1977.

Epidemiology

In Finland, more than 100 patients are known, and incidence is about 1:15,000. Exceptionally, the map of ancestors is congruent with the density of population, thus being densest in western Finland). The disease is not very rare elsewhere either: Beduins in Kuwait, Hindues in Gujarat, Tatars, and Belgians show greater incidence figures than the Finns. There has been a discussion whether or not Meckel syndrome should at all be included in the Finnish Disease Heritage. On the other hand, in many Meckel cases outside Finland, the Finnish gene locus is excluded. Thus, the Finns may have not only a mutation of their own but even their own gene locus.

Finnish expert

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Meretoja disease

Corneal lattice dystrophy — amyloidosis Gelsolin-related amyloidosis Familial amyloidosis, Finnish type (FAF) Familial amyloidotic polyneuropathy, type IV Amyloidosis V (so numbered by OMIM) MIM 105120



Fig. 21

This *autosomal dominant* disease with a confusing multitude of different names is an amyloidosis characterised by corneal lattice dystrophy in adult age.

Clinical features

The patients are symptomless until the third, often the fourth decade of life. Then a corneal lattice dystrophy appears with symptoms of irritation and, often, dysesthesia. Visual acuity decreases only slowly. Glaucoma is common. From the fifth decade of life onwards other signs of amyloid storage appear: cranial nerve palsies, symptoms from peripheral nerves, and renal damage. "Hanging" facial skin is the most typical finding. Skin elsewhere can be brittle and atrophic. Tendency to bruising or postoperative bleeding and sleep apnea are recent findings. The lifespan is nearly normal.

Diagnostic investigations

The ophthalmological findings can be verified by biomicroscopic investigation of the cornea. ENMG may show signs of cranial and peripheral neuropathy. A histopathological demonstration of amyloidosis is seldom needed. In Finland, a gene test is the best method, which also makes predictive diagnosis possible.

Pathogenesis

The structure of the actin-modulating protein gelsoline (gel>sol) is abnormal. Its altered degradation produces amyloid, which accumulates in certain tissues. The abnormality of the gelsolin may also cause disturbances independent of amyloid such as deficient plasticity of the thrombocytes.

Molecular genetics

The gelsoline gene (*GSN*) is in chromosome 9q34. The Finnish point mutation 654G>A, common to all Finnish patients, causes a change of aspartic acid to asparagine in the gelsoline protein, position 187 (D187N). Another mutation (654G>T = D187Y) in the same locus and position has been found in a Danish and a Czech family.

Management

Corneal symptoms are treated by moisturing drops and ointments. Symptomatic treatment of corneal inflammations must be aggressive enough. Corneal transplantation is needed only exceptionally. Hanging facial skin is treated by plastic surgery. Carpal tunnel syndrome is not infrequent and may need operative treatment. The development of amyloid cannot be delayed.

Prenatal diagnosis

Prenatal diagnosis is possible by a gene test, but its need and ethical guidelines are not established.

Historical aspects

The disease was described by the Finnish ophthalmologist Jouko Meretoja in 1969.

Epidemiology

In Finland over 400 patients are known. The affected grandparents of today's patients come from two areas, which have historical population connections. Most probably this extremely rare disorder has originated in Finland from one point mutation several centuries ago. Very few patients have been described outside Finland. The Finnish mutation has been found in one kindred from Holland and Great Britain, five in USA and four in Japan.

Finnish experts

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Mulibrey nanism

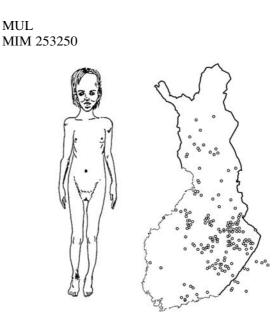


Fig. 22

Mulibrey nanism is a growth disturbance disease associated with a singular combination of abnormalities from different organs.

Clinical features

Growth failure starts prenatally. The proportion between trunk and extremities is normal. Adult height is 130–150 cm.

In infancy, feeding difficulties are common. Triangular face, high and broad forehead, thin extremities, gracile and weak muscles and high-pitched voice are characteristic. Naevi flammei appear in the ankles or soles. Liver is enlarged due to cardial congestion. Myocardial insufficiency may endanger life in infancy. More characteristic still is fibrotic thickening of the pericardium, which hampers the contractions of the heart. Fibrous dysplasia of tibiae and hypogonadism are common findings. Mental retardation does not belong to the disease, but psychosocial deprivation is common as in many chronic abnormalities.

Diagnostic investigations

No diagnostic laboratory investigation exists. The skull is dolichocephalic, sella turcica narrow and J-shaped. The cerebral ventricles are often wide. In eye fundi, yellowish spots are typical. Pertinent cardiological investigations are important. Endocrinological studies may be needed.

Pathogenesis

How one gene brings about these pleiotropic manifestations, is totally open.

Molecular genetics

The gene (*TRIM37*, formerly *MUL*) is in chromosome 17q22-q23, in a region overlapping with the Meckel syndrome gene. It encodes the TRIM37 protein, which belongs to the group of RING-B-box-coiled-coil (!) family of zinc finger proteins and is located in the peroxisomes. The detailed function of TRIM37 protein is not known. Of the Finnish *TRIM37* genes, 98% show one and the same mutation, Fin_{major} = c.493–2A>G. Mutation Fin_{minor}, found in two Finnish disease chromosomes, is c.2212delG. Two further mutations are known, from the USA and the Czech Republic. All these are frameshift mutations causing different truncations of TRIM37 protein.

Management

Cardiological monitoring and follow-up throughout life are imperative. Pericardiectomy becomes often necessary. The family of the affected infant needs support because of feeding difficulties and problems of growth and development. A close contact with an expert doctor is important also later.

Prenatal diagnosis

A gene test is possible at least in the Finnish risk families.

Historical aspects

The disease is one of the many detected by Jaakko Perheentupa (1970) in Finland. The name mulibrey is an acronym compiled of <u>muscle</u>, <u>liver</u>, <u>brain</u> and <u>eye</u>. Today this selection would not be the same, but the name is well established and deserves not to be changed at least until the pathogenesis joining together the different manifestations is solved.

Epidemiology

In Finland, over 80 patients are known, elsewhere less than 20. The map corresponds typically with the late settlement area.

Finnish experts

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Muscle-eye-brain disease

MEB MIM 253280

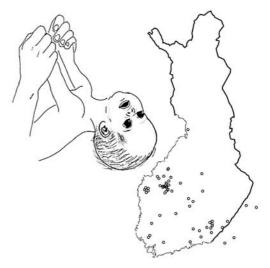


Fig. 23

The triad of MEB comprises congenital muscular dystrophy, mental retardation, and severe ocular findings.

Clinical features

The babies are born at term. Muscular weakness manifests in the early infantile period, causes among others sucking difficulties and is severe by six months of age. Motor development is greatly disturbed: only few patients learn to walk even aided. In childhood, spasticity and joint contractures may develop in the lower extremities. Tendon reflexes are brisk or weak according to age.

The head is often large. Four patients have needed surgical treatment because of hydrocephaly. The forehead is high and temporally narrowed. The midface is flat and expressionless, the palate is narrow. Epileptic seizures are common. Psychic development is usually severely retarded and may deteriorate further with age.

The ocular findings are varied, often severe. Congenital myopia, glaucoma due to structural abnormalities of the anterior chamber, dystrophy of the retina, optic atrophy and cataract show varying combinations. The visual handicap is severe, as is predictable from the nystagmus or wandering eye movements. Lifespan is often shortened although the disease in itself may not be lethal.

Diagnostic investigations

Serum creatine kinase concentration rises moderately during the first year of life, but may become normal in adult age. EMG shows a myopathic pattern by two years of age. The findings in muscular biopsy vary from nearly normal to severe dystrophic alterations. Different EEG changes appear after one year of age. MRI of the brain reveals migration disturbance of pachygyria with a nodular cortical surface called cobblestone cortex. Further findings are large ventricles, flat pons, and cerebellar cysts. The midline structures may be defective. The ERG is low or extinguished, flash-VEP shows delayed giant responses.

The features of MEB vary considerably as concerns composition, severity and time scale. Many syndromes resemble MEB greatly, such as Fukuyama congenital muscular dystrophy (FCMD), cerebro-ocular dysplasia-muscular dystrophy (COD-MD), and Walker-Warburg syndrome (W-W, HARD+-E). Their nosology is disputed. At least the gene loci of MEB and Fukuyama disease are not identical.

Pathogenesis

The pathogenetic defect is a loss of the enzymatic activity of the protein POMGnT1 (O-mannose beta-1,2-N-acetylglucosaminyltransferase) needed in O-mannosyl glycosylation and leading to a deficiency of alpha-dystroglycan. This defect may offer a sensible pathogenetic connection between muscular dystrophy and migration disorder of the brain. Molecular genetics

The gene *POMGnT1* (cf. above) is in chromosome 1p32p34. It has been characterised by a Japanese group. They identified six different mutations in one French and five Turkish patients. The mutation of Finnish patients is not reported as yet.

Management

Myopia and raised intraocular pressure should be treated properly. Physiotherapy is beneficial.

Prenatal diagnosis

Prenatal diagnosis may be or become possible in risk families, if the mutation is known.

Historical aspects

Santavuori, Leisti and Kruus of Finland described MEB in 1977.

Epidemiology

About 30 patients are known in Finland. Over 20 more or less similar cases may be described from elsewhere.

Finnish experts

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Abbre- viation	Names	Age at onset	Gene	Pathoanatomy
INCL	Infantile neuronal ceroid lipofuscinosis, Santavuori-Haltia disease	Infancy	CLN1 PPT	Granular osmiophilic deposits=GROD
vLINCL _{Fin}	Variant late infantile NCL, Finnish type Jansky-Bielschowsky variant	Preschool age	CLN5	Curvilinear bodies, fingerprint profiles
JNCL	Juvenile NCL Spielmeyer-Sjögren disease Spielmeyer-Vogt disease Batten disease	School age	CLN3	Fingerprint profiles
EPMR	Northern epilepsy	School age	CLN8 EPMR	Curvilinear bodies (GROD)

Table 2The neuronal ceroidlipofuscinoses (CLNs) in Fin-land*

*The genes of NCL diseases rare or absent in Finland are: *NCL2* classical LINCL; *NCL4* adult NCL = Kufs disease; *NCL6*, *NCL7* rare LINCL vari-

ants

Neuronal ceroid lipofuscinoses

NCLs

Neuronal ceroid lipofuscinoses are a group of diseases in which ceroid- and lipofuscin-like material is accumulated in neural and other tissues. Common features are recessive transmission, progressive mental retardation after a normal initial development, retinal degeneration and early death. Distinguishing factors are age at onset, additional neurological symptoms, neurophysiologic and neuropathological findings and survival time.

Earlier these diseases were thought to be somehow genetically related to each other. However, for each of them separate gene loci have been detected. Despite that, their pathogenesis might have some factor(s) in common, especially as exceptional, interesting overlappings have been noticed between genes, clinical pictures and neuropathological findings. Peculiarly enough, outside Finland more than ten patients have been described with a clinical picture resembling Spielmeyer-Sjögren disease (JNCL) but caused by mutations of the INCL gene. In these patients, the electron microscopic storage material also resembles that of INCL, and the patients have no vacuolated lymphocytes. Additionally, several other overlappings of similar character have been observed.

Four NCL diseases belong to the Finnish Disease Heritage (Table 2).

Confusion in the polyform nomenclature is caused by the fact that some authors use Batten disease as a collective name for all NCL diseases, while others as synonym for JNCL.

In this article, different NCL diseases are described separately according to the alphabetical site determined by the name most commonly used in Finland: INCL, Jansky-Bielschowsky variant, Northern epilepsy, and Spielmeyer-Sjögren disease.

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Nonketotic hyperglycinemia

NKH

MIM 238300



Fig. 24

NKH is a metabolic disease manifesting in the first few days of life and leading to either neonatal death or profound mental retardation.

Clinical features

Among many forms of nonketotic hyperglycinemia, the Finnish one is of the most severe neonatal type. In the first days of life, the patients develop a picture of serious brain damage: floppiness, lethargy, muscle jerks, diminishing reaction to pain, and respiratory distress. The patients often die of respiratory insufficiency. If not, a most profound psychomotor deterioration develops rapidly. Survival in a decerebrated state may last for more than ten years.

Diagnostic investigations

The glycine concentrations of plasma, urine and especially of cerebrospinal fluid are increased in the absence of ketosis and metabolic acidosis. The proportion between the glycine concentrations of the cerebrospinal fluid and plasma is greater than normal. EEG shows burst suppression pattern in the first days of life, later hypsarrhythmia. In MRI of the brain, a diminished amount of cerebral parenchyma, thin corpus callosum, and delayed myelinization may be seen. Quantitative proton magnetic resonance spectroscopy may disclose high glycine concentration in the neonatal brain. Gene diagnosis is successful in most Finnish cases.

Pathogenesis

In the glycine cleavage, four enzyme proteins, P, H, T and L are needed. In most of the Finnish patients, P-protein, viz. pyridoxal phosphate-regulated glycine decarboxylase, is inactive. Undegraded glycine cumulates and damages the brain.

Molecular genetics

The gene *GCSP*, (glycine cleavage system P) is in chromosome 9p13 or 9p22-p24 (?). Several mutations are known. Of the genes of Finnish patients, 70% present with the point mutation 1691G>T, which alters serine to isoleucine in position 564 of the P-protein. This mutation is not known in other populations. In Finland, two additional point mutations and one deletion are known.

Management

Curative treatment is not available, although trials with many substances affecting glycine metabolism, such as strychnine and dextrometorphane, have been made. It is most important to suspect and diagnose NKH in the first days of life. If an evident diagnosis of NKH can be made, the newborn should probably not be put in a respirator, because surmounting the respiratory distress would mean a futile struggle of many years against a hopeless disease.

Prenatal diagnosis

The determination of glycine/serine proportion in the amniotic fluid is unreliable. The activity of glycine degradation can be measured in the chorionic villi. A gene test is available for most of the Finnish risk families.

Historical aspects

The disease was described in the same year by Gerritsen et al. 1965 of the USA and Visakorpi with his collaborators 1965 from Finland.

Epidemiology

In Finland nearly 50 patients are known, elsewhere about 100. The majority of the Finnish patients are from northern Finland.

Finnish experts

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Northern epilepsy

Progressive epilepsy with mental deterioration CLN8 MIM 600143



Fig. 25

See also Neuronal ceroid lipofuscinoses

This disease is a combination of epilepsy and later manifesting mental deterioration.

Clinical features

Epileptic seizures manifest at five to ten years of age, become more frequent at puberty and diminish in adult age. Mental development begins to deteriorate two to five years after manifestation of the seizures and proceeds to mental retardation. Yet most patients get through their elementary school. Behavioural problems, clumsiness, and disturbances of balance may add to the picture. The duration of life is not known as yet.

Diagnostic investigations

EEG shows slowing of background activity and disappearance of sleep patterns. In MRI, signs of cerebellar, brain stem, and later also cerebral atrophy can be seen.

Pathogenesis

Surprisingly, in autopsy, curvilinear bodies and granular osmiophilic deposits typical of neuronal ceroid lipofuscinoses have been found. That is why Northern epilepsy has been added to NCL diseases, although no ophthalmologic findings belong to this disease. This classification, however, does not explain the pathogenetic mechanism.

Molecular genetics

The gene (*EPMR*, *CLN8*) is in chromosome 8p23. All the Finnish patients have a point mutation 70C>G = R24G. It

may cause disturbance in an endoplasmic reticulum resident membrane protein, whose structure and mode of action is not known.

Management

The most effective antiepileptic is clonazepam. The careful treatment of epilepsy tends to retard the progression of the disease. It is important to understand the nature of the disease in order to deal with the behavioural problems in the right way, to seek individual solutions at school and to find a purposeful job.

Prenatal diagnosis

Prenatal diagnosis might be needed in the last pregnancies of the family. It is uncertain whether this will succeed from chorionic villus biopsy by pathoanatomic means. A gene test, however, is easy and reliable.

Historical aspects

The disease was first reported by Hirvasniemi and Leisti in 1991, but had probably already been described in the 1920s and 1930s by Ilmari Kianto in his novels about the life of rural people in Kainuu.

Epidemiology

Until now, the disease has been found only in one rural district in Kainuu, northeastern Finland. Almost all the patients have been shown by church records to be remotely related to each other via one ancestor.

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PEHO syndrome

<u>Progressive encephalopathy with edema, hypsarrhythmia, and optic atrophy</u> MIM 260565



Fig. 26

Peho syndrome is an early manifesting brain disease leading to profound mental retardation and blindness.

Clinical features

Floppiness and feeding difficulties appear during the first few days of life. Edema of hands and feet, narrow forehead, short nose, out-turned ear lobuli and tapered fingers are typical. Already in early infancy the eyes are wandering or upturned as a sign of severe visual failure. The patients do not learn to sit, walk or speak. Severe microcephaly develops. Infantile spasms begin during the first year of life and change later to epileptic seizures of the Gastaut-Lennox type. Despite the floppiness, tendon reflexes are brisk. With time spasticity and contractures develop. Data on survival time are still lacking.

Diagnostic investigations

There are no specific laboratory investigations. Many patients with severe mental retardation show some solitary features typical of PEHO. The most important diagnostic finding is cerebellar and brain stem atrophy. In MRI, myelinization is delayed. EEG is hypsarrhythmic, SEP and VEP often abnormal. Optic atrophy is seen in the ocular fundi, but ERG is normal.

Pathogenesis

The disease process is not known.

Molecular genetics

The gene is not mapped as yet.

Management

Physiotherapy in order to prevent contractures, stimuli through hearing and touch, and antiepileptic medication are needed throughout life.

Prenatal diagnosis

Prenatal diagnosis does not exist.

Historical aspects

Riitta Salonen with her colleagues detected the disease among her Finnish counselling patients and described it in 1991.

Epidemiology

About 30 evident patients are known in Finland. In addition, there are several retarded patients resembling PEHO, yet without verifiable cerebellar atrophy. These are called PEHO-like patients. In other populations about ten cases are published, four of them from Japan.

Finnish expert

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PLO-SL

Polycystic lipomembranous osteodysplasia — sclerosing leukoencephalopathy Frontal lobe dementia with bone cysts (Nasu)-Hakola disease MIM 221770



PLO-SL is a disease of the central nervous system and cystic osseous lesions, beginning in middle age and leading to presenile dementia and early death.

Clinical features

The patients are symptomless until 20 years of age. After that, pain, swelling and possible fractures may appear in the ankles or wrists, due to juxta-articular bone cysts. At over 30 years of age, neuropsychiatric symptoms begin: disturbances of memory, deteriorating of personality and other characteristics of frontal lobe syndrome. The patients become euphoric, without feeling of illness. Later, agnostic-apractic-aphasic symptoms, myoclonic twitchings, and epileptic seizures appear. The progression continues into deep dementia and death by 50 years of age.

Diagnostic investigations

Bone X-ray reveals juxta-articular cysts. They contain necrotic fat tissue with fatty acid crystals and PAS-positive lipomembranes. In CT and MRI of the brain, atrophy of the white matter especially in the frontal area, enlarged ventricles and signs of demyelinization are seen. EEG shows synchronous, episodic and diffuse 6–8 Hz activity and the alpha rhythm is replaced by diffuse theta and delta activities. On autopsy, brain alterations include demyelinization and gliosis of the white matter, calcifications in the basal nuclei, sudanophilic granules in the perivascular macrophages, microangiopathy, and axonal changes.

Pathogenesis

Despite the characterisation of the gene, the basic mechanism is not known. Proposed "macroscopic" pathogenetic theories include microangiopathy and disturbance of fat metabolism.

Molecular genetics

The gene is in chromosome 19q13.1, in the vicinity of the gene for congenital nephrosis. All the 26 Finnish patients investigated have the mutation $PLOSL_{Fin}$: a 5.3-kb deletion covering exons 1–4 in *TYROBP*, which is the same as the *DAP12* gene known beforehand. The only Japanese patient investigated so far had a different mutation in this gene. The transmembrane protein TYROBP participates in the tyrosine metabolism and is a membrane receptor component in myeloid and natural killer cells. Their function, however, is shown to be normal in the PLO-SL patients.

Surprisingly, other, non-Finnish patients with definitely similar phenotype do not have any mutations in *TYROBP*, but in *TREM2* located in chromosome 6p21-p22 and belonging to the immunoglobulin superfamily. These two genes probably encode different subunits of one receptor signalling complex, the function of which behind the phenotype is not known.

Management

The disease cannot be cured nor delayed. The family needs support and thorough knowledge about the disease in order to avoid bad emotional and other crises. Fractures and epilepsy are treated in the usual manner.

Prenatal diagnosis

The sibship has reached its final size as the first affected member manifests the disease.

Historical aspects

The skeletal findings were described the first time by Terayama of Japan in 1961. They had been observed also in Finland (Järvi et al. 1964). Järvi and Hakola with their collaborators combined the orthopedic and psychiatric features to one entity in 1968. After that, Hakola has studied this disease extensively and in Finland it is mostly called Hakola disease. The Japanese scientists have named it Nasu disease after their own principal investigator. In international literature the combination name Nasu-Hakola disease is common.

Epidemiology

In Finland more than 30 patients have been diagnosed. A greater accumulation of about 100 patients is known in Japan. This combination has urged Hakola to study a possible linguistic interrelationship between Finnish and Japanese (Part II, DOI s00439-002-0876-2, Classical Finnish studies). In Finland, the map of ancestors is most typical for the late settlement area; in nearly all the known families both parents were born in the same commune.

Except in Finland and Japan, *TYROBP* mutations have been found in a few patients from Sweden, Norway, and Brazil. Patients with *TREM2* mutations are known in Sweden, Norway, Italy, USA and Bolivia.

Finnish expert

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Progressive myoclonus epilepsy, Unverricht-Lundborg type

PME, PME-UL, EPM1 MIM 254800

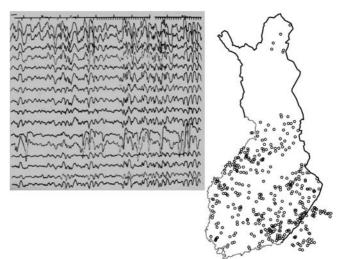


Fig. 28

PME is an epileptic disorder with myotonic jerks and motility difficulties.

Clinical features

The first years of life are normal. At the age of 6–15 years epileptic seizures begin and also involuntary myoclonic jerks provoked by different stimuli, such as moving the limbs or flashlight. The jerks had tendency to extend to series or to epileptic seizures, especially in the mornings. Dysarthric speech, lability of mood and slight decline of psychic capacity were common. The exaggerated jerks prevented moving about and the use of hands. The patients became bedridden and died in early adult age.

Modern treatment has totally changed the prognosis and the disease hardly affects the life span.

Diagnostic investigations

Typical EEG changes are spike and polyspike slow wave discharges (3–5/sec), with marked photosensivity. VEP and SEP may show giant reactions. In the imaging studies of the brain nothing pathological appears. Increased amounts of indican are secreted in the urine.

Pathogenesis

The basic disturbance may be connected with the metabolism of nerve impulse transfer.

Molecular genetics

The gene (*EPM1*, *CSTB*) is in chromosome 21q22.3. It encodes the formation of cystatin B, an inhibitor of cystathione proteinase. Some different mutations are known. The most common Finnish mutation is of a rare type of minisatellite dodecamer expansion in the promoter of the *CSTB* gene resulting in reduced cystatin B expression.

Management

PME is a prime example of diseases in which new treatment has totally changed the course and prognosis of a severe disorder. The prevention of jerks and seizures must be done with sodium valproate alone or in combination with clonazepam. Most of the other epilepsy medicines are ineffective or even cause serious side effects. As addon treatment, especially against myoclonus, piracetam and zonisamide have shown promising results. With appropriate treatment the jerks appear only seldom and do not invalidate the patients. They must tenaciously hang on to normal life and they must be given effective physical and psychic rehabilitation.

Prenatal diagnosis

At the time when PME is diagnosed in the oldest affected sib, the family often already is of planned size. If not, gene diagnosis is available at least in Finland.

Historical aspects

The disease has been known since the 1890s because of Unverricht's thorough description from Estonia and Lundborg's report from Sweden. In Finland, Harenko and Toivakka published their observations on PME in 1961.

The nomenclature of this and related diseases is manifold and confusing. The Finnish form of PME is exactly similar with the descriptions by Unverricht and Lundborg, or PME-UL. Lafora disease (gene *EPM2A*) is totally different with its intracellular inclusions, later manifestation, excessively degenerative psychic component and shorter survival. Juvenile myoclonus epilepsy is milder than PME-UL, does not progress, but morning attacks are typical, too. Actually, the abbreviation PME is used today as a group term for diseases with epileptic fits and myoclonus. In Finland, however, this abbreviation is established to refer to the Unverricht-Lundborg disease, and there may be no reason to alter this practice. With today's knowledge, Baltic myoclonus is a misnomer (cf. Epidemiology).

Epidemiology

In Finland, about 200 patients are known. PME or at least a disease resembling it has been described from different populations, but especially from Mediterranean countries, where it has been called Mediterranean myoclonus, viz. Ramsay-Hunt disease. However, the disease, the gene and its main mutation seem to be identical in Finland and in the Mediterranean countries. That is why Baltic myoclonus is no longer an appropriate name for the Finnish-Estonian-Swedish disease. In Finland, the gene has spread into most areas of the country, but the southeastern part (Karjala) shows the greatest concentration of ancestors. It is quite possible, that the Finns have got the PME gene from the Mediterranean areas brought by foreign traders thousands of years ago (Part I, DOI s00439-002-0875-3, Age of gene mutations).

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Rapadilino syndrome

MIM 266280

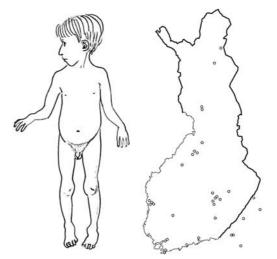


Fig. 29

Rapadilino syndrome comprises several skeletal abnormalities, typical face, growth disturbance and inexplicable diarrhea in infancy.

Clinical features

Skeletal malformations consist of aplasia or hypoplasia of the radii, thumbs and patellae and dislocations or malpositions of the joints of the lower extremities. Because of the longish face, slender and long nose, cleft or narrow palate and micrognathia, the patients resemble each other more than their unaffected sibs. The voice is high-pitched. The stools are loose by unknown mechanism, but become normal without any treatment at the age of 2–3 years. The proportionate growth disturbance is of prenatal onset. The growth curves are below the normal curves, parallel with them. The intelligence is normal.

Diagnostic investigations

The skeletal X-ray investigations are important. If the diagnosis is evident, special investigations of the diarrhea are of no use. Differentiation from other syndromes with radial aplasia, such as TAR, Holt-Oram syndrome, and Fanconi anemia, is important.

Pathogenesis

The way of function of the gene is not known.

Molecular genetics

The localisation of the gene is not known because the patient series has been too small for study.

Management

The malpositions of the lower extremities may demand orthopedic correction even in newborns. The cleft palate is corrected in the usual way. The diarrhea is treated symptomatically. Corrective operations of the upper extremities are considered in due time. The patients learn to use their hands very cleverly. The face is beautiful in a way, although unusual. The psychic burden caused by the singularity must be alleviated by encouragement.

Prenatal diagnosis

The abnormalities of the upper limbs may be visible in accurate ultrasound scan.

Historical aspects

The "mother" of the syndrome is Helena Kääriäinen who found these patients among her Finnish counselling patients in the 1980s. The name rapadilino is not Italian language but an acronym: ra =<u>ra</u>dial aplasia, pa = absent <u>patellae</u> and cleft <u>palate</u>, di = <u>diarrhea</u> and <u>dislocated</u> joints, li = <u>little</u> size and <u>limb</u> malformations, no = long, slender <u>no</u>se and <u>no</u>rmal intelligence.

Epidemiology

More than ten patients are known in Finland. Three case reports have been published from elsewhere.

Finnish expert

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Retinoschisis

X-linked juvenile retinoschisis RS, XLRS MIM 312700

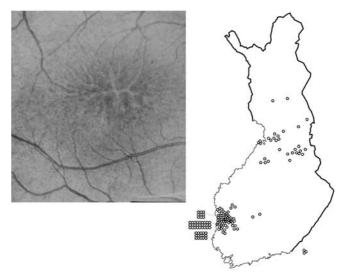


Fig. 30

This type of retinoschisis is an *X*-chromosomal ocular disease causing progressive visual handicap of varying degree in boys.

Clinical features

The disease is found usually in the first eyesight investigation at school. The grade of the visual handicap is different in different patients, but usually remains considerably constant until late middle age. Then, the visual acuity decreases, causing difficulties especially in reading.

In the ocular fundi, macular degeneration with a wheel spoke figure is the first, mild manifestation. In one third of the cases, the superficial layer of the retina is loosened, especially in the temporal periphery, and large holes may be seen, sometimes also folds called vitreous veils. In the rare, grave cases, a large area of the retina may be destroyed and become rolled.

Nothing other than ocular features belong to this disease.

Diagnostic investigations

The ophthalmoscopic diagnosis is easy for the ophthalmologist acquainted with the disease. The green light in the ophthalmoscope helps to perceive the wheel spoke figure. In Finland, gene tests are available for diagnosis.

Opposite to choroideremia, female carriers cannot be verified by ophthalmoscopy but certainly by gene tests. The pedigree compatible with X-linked inheritance affirms the diagnosis for its part.

Pathogenesis

Microcystic degeneration in the deep nerve fiber layer of the retina causes loosening of the retinal layers from each other.

Molecular genetics

The gene, *XLRS1* or *RS1*, is in chromosome Xp22.2. Its structure and over 100 different mutations are known. It encodes a secreted protein, retinoschisin, which is expressed in photoreceptors of the outer retina and bipolar cells of the inner retina and may function as a cell adhesion protein. In Finland three mutations prevail: in Satakunta, southwestern Finland, the mutation 214G>A (E72K) represents 70%, eastwards from there the mutation 221G>T (G74 V) stands for 6%, and in the district of Oulu, northern Finland, the mutation 325G>C (G109R) covers 20% of the Finnish mutations. All these also have their own basic haplotypes. As solitary findings, four other mutations have been found.

Management

The disease cannot be cured or delayed. Precautionary measures such as avoiding sport are not indicated. "Preventive" treatments cause more harm than benefit.

In rare cases, retinoschisis patients may get a true detachment of the retina, which must, of course, be treated by an immediate operation.

Prenatal diagnosis

Prenatal diagnosis is possible by a gene test, if the mutation is known. The need and ethical principles of it are not established.

Historical aspects

Haas of Germany described the disesase in 1898. In Finland, Henrik Forsius with his collaborators have studied retinoschisis comprehensively since the 1960s.

Epidemiology

In Finland over 300 patients are known, among them one female homozygote. The disease is concentrated in the regions of Satakunta and Oulu, where retinoschisis is the commonest cause of bilateral amblyopia of schoolboys. The disease is not very rare in other populations, either.

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Salla disease

Sialic acid storage disease, Finnish type MIM 269920

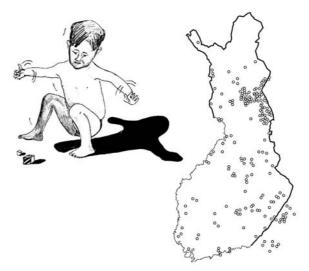


Fig. 31

Salla disease is a slowly progressive storage disease causing mental retardation and motility disturbances.

Clinical features

The first weeks of life are normal. At the age of 4–12 months, floppiness, clumsiness, ataxia, and later, stiffness appear. Transient horizontal nystagmus is often seen. The milestones of psychomotor development are delayed. The speech ability may comprise single words, but many pa-

tients understand speech better than they can speak themselves. A third of the patients do not learn to walk. The development makes slow progress until the third decade of life and begins to go downhill after that. Cheerfulness is a typical characteristic. The degree of severity varies considerably between patients. The age of the oldest known patient is over 70 years.

Diagnostic investigations

In neuroradiological studies, loss of the white matter and deficient myelinization are seen. In difficult cases and in time, cerebral and cerebellar atrophy appear. Corpus callosum is thin. Signs of dysmyelination may be found also in the peripheral nervous system. The concentration of free sialic acid (N-acetyl neuraminic acid) is increased in the urine and in several tissues. Vacuoles appear often in the blood lymphocytes. Lysosomes are expanded by stored sialic acid in skin biopsies.

Pathogenesis

The membrane transport of sialic acid from the lysosomes is impaired, causing a storage of this material in the lysosomes.

Molecular genetics

The gene *SLC17A5* (formerly also *SIASD*, *SLD*, *AST*) is in chromosome 6q14-q15. The gene has been characterised. In Finland, mutation 115C>T (R39C) causes 95% of the Finnish cases in a homozygous form and the rest as compound heterozygous with four other mutations.

In the same locus, other mutations are known that in a homozygous state cause a more severe, infantile sialic acid disease (ISSD). It seems that different mutations in different combinations of compound heterozygotes form a clinical continuum of different grades of severity.

Some principles are understood of the protein coded by the gene, sialine, but the structural details are not known.

Management

Curative treatment does not exist. Communication therapy is important.

Prenatal diagnosis

Prenatal diagnosis is possible by determining the concentration of free sialic acid in the chorionic villi. A gene test is available, if the mutation is known, as it is in Finland.

Historical aspects

Salla disease was found as a byproduct in 1979 by Aula et al., when AGU patients were sought for in northern Finland. Salla is a commune in Lapland.

Epidemiology

Over 100 patients are known in Finland, most of them from northern Finland. Elsewhere, over 20 patients have been published. Half of them are from Sweden; the Finnish mutation has been found in most of them.

Finnish experts

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Selective malabsorption of vitamin B₁₂

Megaloblastic anemia 1 Gräsbeck-Imerslund disease SMB₁₂, MGA1 MIM 261100

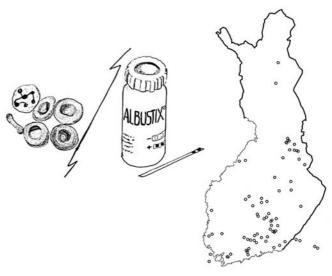


Fig. 32

This disease is a megaloblastic anemia caused by impaired absorption of vitamin B_{12} from the small intestine.

Clinical features

The first symptoms are vague: fatigue, failure to thrive, pallor, poor appetite, vomiting. They appear around 12 months of age. The anemia behind these symptoms would without proper treatment lead to death. Most patients have proteinuria that apparently causes no harm. Untreated patients also develop neurological symptoms.

Diagnostic investigations

Anemia is macrocytic. The concentration of vitamin B_{12} in the blood is very low, but the intrinsic factor of gastric juice needed for the absorption of vitamin B_{12} is normal. The activity of the urinary receptor for the intrinsic factor — cobalamin complex can be measured by a radioisotope-binding assay. In Finland the gene test gives a reliable diagnosis.

Pathogenesis

Vitamin B_{12} , needed for the maturation of the red blood cells, is not absorbed from the alimentary canal. The activity of the intrinsic factor — cobalamin receptor in the mucosa of the small intestine and probably also in the kidney is low.

Molecular genetics

The gene *MGA1*, *CUBN*, is in chromosome 10p12.1. Its mutations lead to an impaired recognition and loss of affinity of the intrinsic factor — cobalamin-binding region of the protein cubilin in the cells of intestinal mucosa and renal tubuli. In more than 90% of the Finnish disease genes, mutation FM1 (3916C>T = P1297L) is found. Also two other mutations (FM2 and FM3) are known in Finland. No data are available on the mutations in other populations.

Management

Parenteral administration of vitamin B_{12} cures the anemia totally but does not affect the proteinuria. Diagnosed properly and treated regularly, the "disease" is fully harmless.

Prenatal diagnosis

Prenatal diagnosis is hardly needed because the treatment is curative.

Historical aspects

The disease was described simultaneously by Olga Imerslund of Norway (1959, 1960) and Ralph Gräsbeck with his collaborators in Finland (1960).

Epidemiology

In Finland about 40 patients are known; in Norway about 15, in the Near East about 20 and in other populations more than 100.

The ancestors of the Norwegian patients come mainly from a small area in southeastern Norway (Valdres) where the "Forest Finns", first having immigrated from Savo to Sweden, moved in the 1500–1600s. It would be tempting to assume that the genes of the Norwegians and the Finns had a common origin. This seems, however, not to be the case: three Norwegian patients investigated so far do not have any of the Finnish mutations.

The appearance of the disease in Finland has been erratic. As 16 patients born in the 1950s were diagnosed, only three patients born in the 1970s, one in the 1980s and three in the 1990s have been seen. Some doctors have even doubted the existence of this whole disease, but this doubt has been proved wrong because of the many studies in Finland and elsewhere. True, this disease has not been a favourite disease of any Finnish pediatrician. It is not probable that an anemia, lethal without proper treatment, could in modern times escape diagnosing. It could have been treated outside the university hospitals in a "natural" way by parenteral administration of vitamin B₁₂. The incidence of no other Finnish disease has diminished so drastically because of internal migration into cities. It is even not impossible that some changes in environmental, say dietary factors, would have contributed in some way to the resorption of vitamin B_{12} during the last 20 years.

Finnish expert

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Spielmeyer-Sjögren disease

Juvenile neuronal ceroid lipofuscinosis Spielmeyer-Vogt disease, (Late onset) Batten disease SS, JNCL, CLN3 MIM 204200

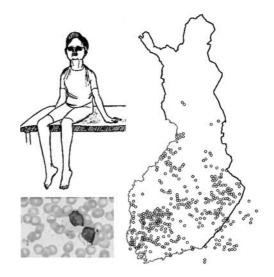


Fig. 33

See also Neuronal ceroid lipofuscinoses

Spielmeyer-Sjögren disease begins before or at school age with impairing of the visus and leads to neuropsychological deterioration and death.

Clinical features

The first feature is the gradual impairing of vision at the age of 4-8 years. Severe visual handicap and even blindness follow at 8-15 years of age. Epileptic seizures, extrapyramidal speech and motor problems, as well as the deterioration of psychic abilities appear at 10-15 years. The lifespan is 18-35 years. The final, bedridden period lasts for only some months. Great inter- and intra-familial variability exists in the succession, severity, and age at manifestation of the different components of the disease.

Diagnostic investigations

In the eye fundi, macular degeneration, and later, attenuated vessels, optic atrophy and "salt and pepper" alterations develop. Abolishing ERG and decreasing and delaying flash-VEP appear. EEG alterations are unspecific. MRI of the brain is normal at the early stage, but hypointense signals in the thalami and hyperintense signals in the white matter appear in the second decade of age. SPECT shows local hypoperfusion. The late appearance of these findings is important for differentiating this disease from the Finnish variant of Jansky-Bielschowsky disease.

Blood lymphocytes show vacuoles. Thus, a combination of deteriorating visus, degenerative alterations of eye fundi and vacuolated lymphocytes is almost pathognomonic for Spielmeyer-Sjögren disease.

In brain samples at autopsy, fingerprint patterns are seen in the neurons in electron microscopy. In chorionic villus biopsy of the fetus and rectal biopsy of the patient, a mixed picture of fingerprint patterns and curvilinear bodies are seen. The gene test confirms the diagnosis, if the mutation is known.

Pathogenesis

Also this disease can be called a lysosomal storage disorder. Many theories, a.o. peroxidation disturbance of fatty acids, have been proposed, but none of them has been confirmed as yet. The fault in some lysosomal membrane protein possibly hampers the function of the nerve synapses.

Molecular genetics

The gene *CLN3* is in chromosome 16p12. Over 30 different mutations are known. The most common is a deletion of 1.02 kb (461–677del), which is found in 90% of the Finnish and 80% of other populations' disease genes. In Finland, four additional mutations are known, all of them as compound heterozygotes with the main mutation. The lysosomal protein CLN3 is expressed in neuronal synapses but not in synaptic vesicles. The mutated protein seems to be arrested in the neuronal cell soma and cannot reach the axons and synapses. The details of the function of the CLN3 protein are not known.

Management

Since the 1970s the patients have been treated with antioxidants, viz. vitamins B_2 , B_6 , E, and selenium. This treatment seems to retard the progression of other features except the deterioration of the visus. Additionally, many kinds of symptomatic treatment are needed for the epilepsy, extrapyramidal symptoms, and psychiatric problems. In the physical and psychic rehabilitation, the visual handicap has a central role, but also many kinds of other rehabilitation, such as physiotherapy, riding etc., are beneficial.

Prenatal diagnosis

Prenatal diagnosis succeeds from the chorionic villi with both electron microscopic investigation and gene test. The problem is that the final size of the sibship has often been reached before the diagnosis of the eldest affected child is made.

Historical aspects

The first description is from Norway by Stengel in 1826. In Finland, the pioneer both in diagnosis and treatment has been Pirkko Santavuori for this disorder, as also in other NCL diseases. The Finnish doctors von Bagh and Hortling were the first to describe the vacuolated lymphocytes in this disease in 1948.

Epidemiology

About 200 patients are known in Finland, where the incidence is 1:20,000 and carrier frequency about 1:70. The disease is not very rare elsewhere either. As an overall incidence even a figure of 1:25,000 has been proposed. It may be an overestimate rather than vice versa.

Finnish experts

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Molecular genetics

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Tibial muscular dystrophy

TMD MIM 600334

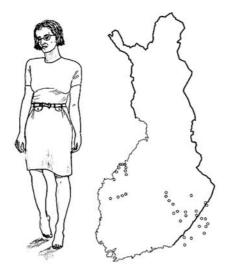


Fig. 34

This *autosomal dominant* myopathy presents with a mild weakness of the anterior tibial muscles in adult age.

Clinical features

TMD is a very mild and limited muscular disease. The "patient" notices that the feet flap in walking (impaired dorsiflexion of the ankle) and walking on heels is impossible. Later, a mild foot drop develops. This dystrophy does not cause marked invalidisation even after years.

Diagnostic investigations

In CT or MRI of the leg, the tibialis anterior muscle shows fatty degeneration. Similar focal alterations may be seen even in other, asymptomatic muscles. In the biopsy from the affected muscle, common dystrophic changes and, possibly, rimmed vacuoles are found. In ENMG, signs of muscular dystrophy, but not those of neural damage, are seen. Serum creatine kinase values are normal or slightly raised. A gene test is available for the Finnish patients.

Pathogenesis

Muscular damage is caused by a fault in the titanic skeletal muscle protein titin (formerly connectin). Titin keeps the contractile elements of the sarcomere in place and provides multiple ligand binding sites for several other muscle proteins. One of these, calpain-3 shows secondary deficiency, which may be an important part in the pathogenesis of TMD.

Molecular genetics

The gene *TTN* is in chromosome 2q31. It is a giant gene with 363 exons. The Finnish mutation in Mex6 is an 11-bp change in the 363rd and last exon, changing four amino acids in the titin protein. All Finnish patients investigated have this unique mutation and the LGMD patients of the TMD kindreds are homozygous for it (cf. Historical aspects).

Management

The symptoms are so mild that usually no treatment is needed. It is important to inform the "patients" that they do not need to fear a severe worsening of the disease. However, in some patients the foot drop may become so difficult that tibial posterior tendon transposition may be indicated.

Prenatal diagnosis

Prenatal diagnosis is not needed. "Screening" by the gene test among the spouse candidates of near relatives might be sensible in order to avoid homozygosity of the TMD gene on its core areas (cf. Historical aspects).

Historical aspects

TMD was revealed when the Finnish neurologist Bjarne Udd investigated a severe muscular dystrophy of the limb girdle type in a coastal isolate Larsmo in Ostrobothnia, western Finland. Then, mild weakness of the legs was noticed in close relatives of the patients. It turned out that the homozygosity of the gene causes the severe disorder (LGMD2J), whereas the same gene in heterozygous individuals causes TMD. Among the confusing group of distal muscular myopathies, TMD is a distinctly delineated entity.

Epidemiology

Except in Ostrobothnia, TMD was soon found also in eastern Finland and nowadays even all over the country. According to the haplotypes and genotypes, all occurrences have a common source. In Finland, more than 300 affected individuals are known. Elsewhere, in Belgium, France, and the USA, one kindred in each has been found so far. In addition, affected descendants of the Finnish immigrants have been found in Sweden, Germany and Canada.

Finnish expert

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Usher syndrome type 3

USH3

(The old Finnish name is dystrophia retinae pigmentosa — dysacusis = Nuutila disease) MIM 276902

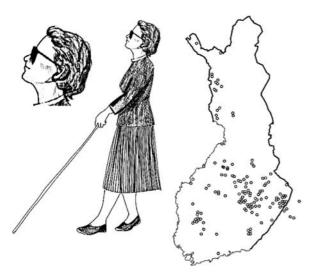


Fig. 35

Usher syndrome is a combination of hearing loss and visual failure of the retinitis pigmentosa type.

Clinical features

Three main types of Usher syndrome exist. In all of them, the visual findings of retinitis pigmentosa type manifest in childhood, show night blindness, photophobia, narrowing of visual fields, decreasing visual acuity and early cataract, and end in a severe visual handicap if not in blindness. The hearing loss is congenital and severe in type 1 and congenital and moderate in type 2. In type 3, the hearing loss develops gradually but becomes at least moderately severe. The age at manifestation and speed of the progress vary considerably. In type 1, a disturbance of balance is also common.

In Finland, type 1 may be as prevalent as elsewhere and patients with type 2 are also seen. However, type 3 belongs to the Finnish Disease Heritage. It is described in the following.

Diagnostic investigations

The hearing investigations show inner ear damage. The audiogram is declining or U-shaped. Audiograms from different ages should be compared in order to be assured of the existence of type 3. The responses of the vestibular organ are normal or slightly decreased.

The ocular investigations are made along lines common in retinitis pigmentosa. Often, hyperopia and astigmatism are also found.

Pathogenesis

Despite active investigations on both retinitis pigmentosa and inner ear, the pathogenesis and common denominator of these components are only fragmentarily known.

Molecular genetics

Several gene loci are responsible for Usher 1 and 2. The gene for the Finnish Usher 3 (*USH3A*) is in chromosome 3q21-q25. The gene encodes the transmembrane protein clarin-1, which may have a role in cochlear hair cell and retinal photoreceptor cell synapses. The main mutation (Fin major) in 56 Finnish patients investigated is c.300T>G = Y100X. Four patients in two kindreds were compound heterozygotes between Finmajor and Finminor = c.131T>A = M44K. In one consanguineous Italian family a third mutation, c.231–233 delATT, has been found.

Management

Any component of the Usher syndromes cannot be cured or retarded. However, both hearing and seeing can be aided with many kinds of devices and rehabilitative measures. Finally, the patients become both auditorily and visually impaired, and thus communication may be extremely difficult. Fortunately, most patients have learned normal speech before the loss of hearing, in contrast to Usher 1.

Prenatal diagnosis

The characterisation of the gene has made prenatal diagnosis possible. However, its demand and the ethical principles have not been established in diseases like this.

Historical aspects

Usher syndrome has been known since the 1800s by different names after the classical investigations by von Graefe, Lindenow and Hallgren. In Finland, the neurologist Arto Nuutila carried out a nationwide investigation of "dystrophia retinae pigmentosa — dysacusis" in the 1960s. The existence of type 3 was considered questionable for a long time, until the Finnish doctors Leenamaija Pakarinen and Eeva-Marja Sankila delineated it and mapped its gene.

Epidemiology

In Finland, nearly 300 Usher patients are known, out of whom about 100 belong to type 3. Their region of predilection is in Eastern Finland, whereas type 1 is prevalent in Lapland. Type 1 is considerably common in several populations and thus Usher syndrome should not be included in the Finnish Disease Heritage without the exceptional abundant occurrence of type 3.

Finnish experts

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Ophthalmology and molecular genetics

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Vuopala disease

Lethal arthrogryposis with anterior horn cell disease LAAHD

Cf. Lethal congenital contracture syndrome = *Herva disease* MIM —

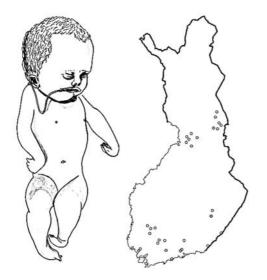


Fig. 36

This fetal immobility syndrome is due to intrauterine spinal anterior horn cell damage and causes perinatal death.

Clinical features

The findings resemble those of Herva disease. However, more than half the patients are born full-term. Compared with Herva disease, these newborns have more muscles, but do not have edema or pterygia. The upper limbs are often rotated in pronation. The patients are stillborn or die soon after birth. The longest survival age has been one month.

Diagnostic investigations

Although all newborns die, the correct diagnosis is imperative for the genetic counselling of the family. Photos and autopsy are of utmost importance. At autopsy, the spinal cord and muscle samples must also be investigated.

Pathogenesis

The clinical findings are due to immobility of the fetus, while the immobility is due to degeneration and diminished number of the spinal anterior horn motor neurons. The muscles show a microscopic picture of neurogenic damage as in spinal muscular atrophies. The disease cannot be called spinal muscular atrophy (SMA) as long as, by definition, congenital contractures are not allowed in SMA.

Molecular genetics

The gene is not mapped, still less characterised. The locus of either Herva disease in chromosome 9q34 or that of SMAs in 5q is not excluded as yet.

Management

The patients are beyond reach of effective help. If the diagnosis is certain, respirator treatment can hardly produce a sensible prolongation of life.

Prenatal diagnosis

Prenatal diagnosis may be possible by ultrasound in risk families.

Historical aspects

The Finnish pediatric pathologists Katri Vuopala and Riitta Herva separated this disease from Herva disease to a distinct entity in the 1990s.

Epidemiology

In Finland about 20 evident and as many probable patients are known from two areas in Häme and Kainuu. Some solitary cases with similar phenotype have been described from elsewhere.

Finnish experts

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New candidates for addition to the Finnish Disease Heritage

At least five diseases are already waiting to be added to "Perheentupa's steps" (see Part I, DOI s00439-002-0875-3, Description and definition of FDH, Fig. 2).

Two of them have been known in Finland for a long time. Tyrosinemia type I appears in a concise area resembling that of Jansky-Bielschowsky disease. About 20 patients are known in Finland and 200 in the world, most of them in Saguenay-Lac-Saint-Jean, Canada, settled by French immigrants. The gene FAH is in chromosome 15q23-q25 and the Finnish mutation is 786G>A =W262X. Curiously enough, Visakorpi in 1972 described tyrosinemia type I, together with lysinuric protein intolerance and nonketotic hyperglycinemia, in an article of the Finnish medical journal Duodecim, in the issue in which the Finnish Disease Heritage was introduced to the Finnish doctors. Because tyrosinemia has been considered a classical disease, it has not been included in the Finnish Disease Heritage. In fact, it is globally much rarer than assumed and thus its "classical" character seems questionable.

The other classical disease is the recessively inherited *nephronophthisis*. The preliminary investigations were done in Finland in the 1970s, but detailed studies were not carried out until the 1990s, by Sirpa Ala-Mello. According to these studies, over 40 patients have been found in Finland and over 100 elsewhere. The gene is mapped to chromosome 2q13 and the commonest mutation in Finland, as well as elsewhere, is a 250-kb deletion. Detailed gene studies are in progress.

The three other candidate diseases are not classical. *Bernard-Soulier syndrome* is a disorder of the hemostasis. Platelets are diminished, large, short-lived and do not agglutinate by ristocetin. The fault is in one of the four glycoproteins of the cell membranes: GPIb alfa, GPIb beta, GPV, and GPIX, each determined by a different gene locus. Genetically speaking, Bernard-Soulier syndrome is not one disease but a group of diseases like retinitis pigmentosa. According to the studies by Riitta Kekomäki and Satu Koskela, 30 patients in Finland are known, elsewhere about 70. Four mutations of two genes are known in Finland. One of them, mutation N45S in the gene of GPIX might be a Finnish speciality especially as its clini-

cal picture seems to differentiate more or less from the complex. Detailed studies are in progress.

LCHAD-deficiency is a severe metabolic disorder, a disturbance of the mitochondrial oxygenation of longchain fatty acids causing severe chronic failure to thrive, acute metabolic crises and marked ocular findings in infants. LCHAD stands for long-chain 3-hydroxyacyl-CoA dehydrogenase deficiency. The disease was described ten years ago. According to the recent investigations by Tiina Tyni, Helena Pihko and others, over 30 patients in Finland are known, elsewhere about 50. The gene locus and structure are known. All the Finnish patients and most of the others show the same mutation G1528C (E510Q) in the chromosome 2p23. The epidemiological studies, among others, are not finished as yet.

The *recessive incisive hypodontia* (*RIH*) is a new odontological syndrome. The most typical features are missing of all four lower incisors and lateral upper incisors. Atopic eczema, childhood asthma, symptoms of nails and some facial features may accompany the syndrome. In the investigations by Sinikka Pirinen, Sirpa Arte, and their collaborators, more than 40 patients are known in Finland and new patients seem to present themselves continuously. Two similar patients have been described from Egypt and one family from Lebanon.

Structural anomalies of the teeth are usually dominantly inherited and defects of the lower incisors are rare indeed. In both respects, RIH makes an exception. When the investigations are finished, this disease most probably will be included in the Finnish Disease Heritage.

Acknowledgements My warmest thanks to all the expert doctors for the patient data and for checking the manuscript on their respective section; to Markku Löytönen, Mari Markkanen-Leppänen and Publishing Company Otava for the figures; to Kari Markovaara, Jonna Mervelä, Taina Miikkulainen, Liisa Savolainen, and Leena Toivanen for their everlasting patience in different kinds of support, and to the Finnish Cultural Foundation for the grants.

References

References for AGU

- Arvio M (1993) Life with aspartylglucosaminuria. Thesis, University of Helsinki
- Arvio P, Arvio M (2002) Progressive nature of aspartylglucosaminuria. Acta Paediatr 91:255–257
- Arvio M, Sauna-aho O, Peippo M (2001) Bone marrow transplantation for aspartylglucosaminuria: follow-up study of transplanted and non-transplanted patients. J Pediatr 138:288–290
- Arvio M, Laiho K, Kauppi M, Peippo M, Leino P, Kautiainen H, Kaipiainen-Seppänen O, Mononen I (2002) Carriers of the aspartylglucosaminuria genetic mutation and chronic arthritis. Ann Rheum Dis 61:180–181
- Aula P, Autio S, Raivio K, Näntö V (1974) Detection of heterozygotes for aspartylglucosaminuria (AGU) in cultured fibroblasts. Humangenetik 25:307–314
- Aula P, Raivio K, Autio S (1976) Enzymatic diagnosis and carrier detection of aspartylglucosaminuria using blood samples. Pediat Res 10:625–629
- Autio S (1972) Aspartylglucosaminuria. Analysis of thirty-four patients. J Ment Defic Res Monogr Ser I

- Dunder U, Kaartinen V, Valtonen P, Väänänen E, Kosma VM, Heisterkamp N, Groffen J, Mononen I (2000) Enzyme replacement therapy in a mouse model of aspartylglycosaminuria. FASEB J 14:361–367
- Enomaa N, Danos O, Peltonen L, Jalanko A (1995) Correction of deficient enzyme activity in a lysosomal storage disease, aspartylglucosaminuria, by enzyme replacement and retroviral gene transfer. Hum Gene Ther 6:723–731
- Haltia M, Palo J, Autio S (1975) Aspartylglucosaminuria: a generalised storage disease. Morphological and histochemical studies. Acta Neuropath (Berl) 31:243–255
- Ikonen E, Aula P, Grön K, Tollersrud O, Halila R, Manninen T, Syvänen AC, Peltonen L (1991) Spectrum of mutations in aspartylglucosaminuria. Proc Natl Acad Sci USA 88:11222– 11226
- Jalanko A, Tenhunen K, McKinney C, LaMarca M, Rapola J, Autti T, Joensuu R, Manninen T, Sipilä I, Ikonen S, Riekkinen PJr, Ginns E, Peltonen L (1998) Mice with an aspartylglucosaminuria mutation similar to humans replicate the pathophysiology in patients. Hum Mol Genet 7:265–272
- Jenner FA, Pollitt RJ (1967) Large quantities of 2-acetamido-1-(β-L-aspartamido)1,2-dideoxyglucose in the urine of mentally retarded siblings. Biochem J 103:48p-49p
- Kaartinen V, Mononen I, Voncken JW, Noronkoski T, Gonzales-Gomez I, Heisterkamp N, Groffen J (1996) A mouse model for the human lysosomal disease aspartylglycosaminuria. Nat Med 2:1375–1378
- Mononen I, Aronson NN Jr (1997) Lysosomal storage disease: aspartylglycosaminuria. Springer, Heidelberg
- Morris C, Heisterkamp N, Groffen J, Williams JC, Mononen I (1992) Chromosomal localization of the human glycoasparaginase gene to 4q32-q33. Hum Genet 88:295–297
- Palo J (1967) Prevalence of phenylketonuria and some other metabolic disorders among mentally retarded patients in Finland. Acta Neurol Scand 43:573–579
- Peltola M, Kyttälä A, Heinonen O, Rapola J, Paunio T, Revah F, Peltonen L, Jalanko A (1998) Adenovirus-mediated gene transfer results in decreased lysosomal storage in brain and total correction in liver of aspartylglucosaminuria (AGU) mouse. Gene Ther 5:1314–1321
- Saarela J, Laine M, Oinonen C, Schantz C, Jalanko A, Rouvinen J, Peltonen L (2001) Molecular pathogenesis of a disease: structural consequences of aspartylglucosaminuria mutations. Hum Mol Genet 10:983–995
- Syvänen AC, Ikonen E, Manninen T, Bengtström M, Söderlund H, Aula P, Peltonen L (1992) Convenient and quantitative determination of the frequency of a mutant allele using solid-phase minisequencing: application to aspartylglucosaminuria in Finland. Genomics 12:590–595

References for APECED

- Aaltonen J, Björses P (1999) Cloning of the APECED gene provides new insight into human autoimmunity. Ann Med 31:111– 116
- Aaltonen J, Björses P, Sandkuijl L, Perheentupa J, Peltonen L (1994) An autosomal locus causing autoimmune disease: autoimmune polyglandular disease type I assigned to chromosome 21. Nat Genet 8:83–87
- Ahonen P (1985) Autoimmune polyendocrinopathy–candidosis– ectodermal dystrophy (APECED): autosomal recessive inheritance. Clin Genet 27:535–542
- Ahonen P, Myllärniemi S, Kahanpää A, Perheentupa J (1986) Ketokonazole is effective against the chronic mucocutaneous candidosis of autoimmune polyendocrinopathy – candidosis – ectodermal dystrophy (APECED). Acta Med Scand 220:333–339
- Ahonen P, Myllärniemi S, Sipilä I, Perheentupa J (1990) Clinical variation of autoimmune polyendocrinopathy – candidiasis – ectodermal dystrophy (APECED) in a series of 68 patients. N Engl J Med 322:1829–1836

- Björses P, Aaltonen J, Vikman A, Perheentupa J, Ben-Zion G, Chiumello G, Dahl N, Heideman P, Hoorwe-Nijman JJG, Mathivon L, Mullis PE, Pohl M, Ritzen M, Romeo G, Shapiro MS, Smith CS, Solyom J, Zlotogora J, Peltonen L (1996) Genetic homogeneity of autoimmune polyglandular disease type I. Am J Hum Genet 59:879–886
- Björses P, Aaltonen J, Horelli-Kuitunen N, Yaspo ML, Peltonen L (1998) Gene defect behind APECED: a new glue to autoimmunity. Hum Mol Genet 7:1547–1553
- Björses P, Halonen M, Palvimo JJ, Kolmer M, Aaltonen J, Ellonen P, Perheentupa J, Ulmanen I, Peltonen L (2000) Mutations in the AIRE gene: effects on subcellular location and transactivation function of the autoimmune polyendocrinopathy – candidiasis – ectodermal dystrophy protein. Am J Hum Genet 66: 378–392
- Halonen M, Eskelin P, Myhre AG, Perheentupa J, Husebye ES, Kampe O, Rorsman F, Peltonen L, Ulmanen I, Partanen J (2002) AIRE mutations and human leukocyte antigen genotypes as determinants of the autoimmmune polyendocrinopathy–candidiasis–ectodermal dystrophy phenotype. J Clin Endocrinol Metab 87:2568–2574
- Heino M, Peterson P, Kudoh J, Shimizu N, Antonarakis SE, Scott HS, Krohn K (2001) APECED mutations in the autoimmune regulator (AIRE) gene. Hum Mutat 18:205–211
- Kumar PG, Laloraya M, Wang CY, Ruan QG, Davoodi-Semiromi A, Kao KJ, She JX (2001) The autoimuune regulator (AIRE) is a DNA-binding protein. J Biol Chem 276:41357–41364
- Nagamine K, Peterson P, Scott HS, Kudoh J, Minoshima S, Heino M, Krohn KJE, Lalioti MD, Mullis PE, Antonarakis SE, Kawasaki K, Asakawa S, Ito F, Shimizu N (1997) Positional cloning of the APECED gene. Nat Genet 17:393–398
- Perheentupa J (2002) APS-I/APECED: the clinical disease and therapy. Endocrinol Metab Clin North Am 31:295–320
- Pitkänen J, Vähämurto P, Krohn K, Peterson P (2001) Subcellular localization of the autoimmune regulator protein, characterization of nuclear targeting and transcriptional activation domain. J Biol Chem 276:19597–19602
- Ramsey C, Winqvist O, Puhakka L, Halonen M, Moro A, Kampe O, Eskelin P, Pelto-Huikko M, Peltonen L (2002) Aire deficient mice develop multiple features of APECED phenotype and show altered immune response. Hum Mol Genet 11:397– 409
- Tarkkanen A, Merenmies L (2001) Corneal pathology and outcome of keratoplasty in autoimmune polyendocrinopathy–candidiasis–ectodermal dystrophy (APECED). Acta Ophthalmol Scand 79:204–207
- The Finnish-German APECED Consortium (1997) An autoimmune disease, APECED, caused by mutations in a novel gene featuring two PHD-type zinc-finger domains. Nat Genet 17: 399–403
- Thorpe ES Jr, Handley HE (1929) Chronic tetany and chronic mycelial stomatitis in a child aged four and one half years. Am J Dis Childh 38:328–338
- Visakorpi J, Gerber M (1963) Hypoparathyroidism with steatorrhoea and some features of pernicious anaemia in a 5-year-old girl. Ann Paediat Fenn 9:129–137
- Vogel A, Strassburg CP, Obermayer-Straub P, Brabant G, Manns MP (2002) The genetic background of autoimmune polyendocrinopathy – candidiasis – ectodermal dystrophy and its autoimmune disease components. J Mol Med 80:201–211
- Zlotogora J, Shapiro MS (1992) Polyglandular autoimmune syndrome type I among Iranian Jews. J Med Genet 29:824–826

References for cartilage-hair hypoplasia

- McKusick VA (1964) Metaphyseal dysostosis and thin hair: a "new" recessively inherited syndrome? Lancet I:832–833
- McKusick VA, Eldridge R, Hostetler JA, Ruangwit U, Egeland JA (1965) Dwarfism in the Amish. II. Cartilage-hair hypoplasia. Bull Johns Hopkins Hosp 116:285–326 and (1978) in: McKusick V (ed) Medical genetic studies of the Amish. Johns Hopkins University Press, Baltimore and London, pp 231–272

- Mäkitie O (1992) Cartilage-hair hypoplasia in Finland epidemiologic and genetic aspects in 107 patients. J Med Genet 29: 652–655
- Mäkitie O, Kaitila I (1993) Cartilage-hair hypoplasia; clinical manifestations in 108 Finnish patients. Eur J Pediatr 152:211– 217
- Mäkitie O, Sulisalo T, de la Chapelle A, Kaitila I (1995) Syndrome of the month: cartilage-hair hypoplasia. J Med Genet 32:39–43
- Mäkitie O, Pukkala E, Teppo L, Kaitila I (1999) Increased incidence of cancer in patients with cartilage-hair hypoplasia. J Pediatr 134:315–318
- Mäkitie O, Kaitila I, Savilahti E (2000) Deficiency of humoral immunity in cartilage-hair hypoplasia. J Pediatr 137:487–492
- Mäkitie O, Kaitila I, Rintala R (2001) Hirschsprung disease associated with severe cartilage-hair hypoplasia. J Pediatr 138:929– 931
- Mäkitie O, Tapanainen PJ, Dunkel L, Siimes MA (2001) Impaired spermatogenesis: an unrecognized feature of cartilage-hair hypoplasia. Ann Med 33:201–205
- Perheentupa J (1972) Kolme periytyvää kasvuhäiriötä (Cartilagehair hypoplasia, diastrophic nanism, and mulibrey nanism, English summary). Duodecim 88:60–71
- Ridanpää M, van Eenennaam H, Pelin K, Chadwick R, Johnson C, Yuan B, van Venrooij W, Pruijn G, Salmela R, Rockas S, Mäkitie O, Kaitila I, de la Chapelle A (2001) Mutations in the RNA component of RNase MRP cause a pleiotropic human disease, cartilage-hair hypoplasia. Cell 104:195–203
- Ridanpää M, Sistonen P, Rockas S, Rimoin DL, Mäkitie O, Kaitila I (2002) Worldwide mutation spectrum in cartilage hair hypoplasia: ancient founder origin of the major 70A>G mutation of the untranslated *RMRP*. Eur J Hum Genet 10:439–447
- Sulisalo T, Sistonen P, Hästbacka J, Wadelius C, Mäkitie O, de la Chapelle A, Kaitila I (1993) Cartilage-hair hypoplasia gene assigned to chromosome 9 by linkage analysis. Nat Genet 3:338– 341
- Sulisalo T, van den Burgt I, Rimoin DL, Bonaventure J, Sillence D, Campbell JB, Chitayat D, Scott CI, de la Chapelle A, Sistonen P, Kaitila I (1995) Genetic homogeneity of cartilage-hair hypoplasia. Hum Genet 95:157–160

References for choroideremia

- Cremers FPM, van de Pol DJR, van Kerkhoff LPM, Wieringa B, Ropers HH (1990) Cloning of a gene that is rearranged in patients with choroideremia. Nature 347:674–677
- Forsius H, Eriksson AW (1978) Choroideremia. In: Francois J (ed) 5. Kongress der Europäischen Gesellschaft für Ophthalmologie, Hamburg 5–9 April 1976, Ferdinand Enke Verlag, Stuttgart
- Furu T, Kääriäinen H, Sankila EM, Norio R (1993) Attitudes towards prenatal diagnosis and selective abortion among patients with retinitis pigmentosa or choroideremia as well as among their relatives. Clin Genet 43:160–165
- Kärnä J (1986) Choroideremia; a clinical and genetic study of 84 Finnish patients and 126 female carriers. Thesis, University of Oulu
- Mauthner H (1872) Ein Fall von Chorioideremia. Ber Naturmed Ver Innsbruck 2:191
- McTaggart KE, Tran M, Mah DY, Lai SW, Nesslinger NJ, MacDonald IM (2002) Mutational analysis of patients with the diagnosis of choroideremia. Hum Mutat 20:189–196
- Nussbaum RL, Lewis RA, Lesko JG, Ferrell R (1985) Choroideremia is linked to the restriction fragment length polymorphism DXYS1 at Xq13–21. Am J Hum Genet 37:473–481
- Sankila EM, Sistonen P, Cremers FPM, de la Chapelle A (1991) Choroideremia: linkage analysis with physically mapped close DNA-markers. Hum Genet 87:348–352
- Sankila EM, Tolvanen R, van den Hurk JAJM, Cremers FPM, de la Chapelle A (1992) Aberrant splicing of the CHM gene is a significant cause of choroideremia. Nat Genet 1:109–113

- Seabra MC, Brown MS, Goldstein JL (1993) Retinal degeneration in choroideremia: deficiency of Rab geranylgeranyl transferase. Science 259:377–381
- Takki K (1974) Differential diagnosis between the primary total choroidal vascular atrophies. Brit J Ophthalmol 58:24–35

References for Cohen syndrome

- Chandler KE, Clayton-Smith J (2002) Does a Jewish type of Cohen syndrome truly exist? Am J Med Genet 111:453–454
- Cohen MMJr, Hall B, Smith D, Graham B, Lampert K (1973) A new syndrome with hypotonia, obesity, mental deficiency, and facial, oral, ocular and limb anomalies. J Pediatr 83:280–284
- Kivitie-Kallio S, Norio R (2001) Cohen syndrome: essential features, natural history, and heterogeneity. Am J Med Genet 102: 125–135
- Kivitie-Kallio S, Rajantie J, Juvonen E, Norio R (1997) Granulocytopenia in Cohen syndrome. Br J Haematol 98:308–311
- Kivitie-Kallio S, Autti T, Salonen O, Norio R (1998) MRI of the brain in the Cohen syndrome: a relatively large corpus callosum in patients with mental retardation and microcephaly. Neuropediatrics 29:298–301
- Kivitie-Kallio S, Larsen A, Kajasto K, Norio R (1999) Neurological and psychological findings in patients with Cohen syndrome: a study of 18 patients aged 11 months to 57 years. Neuropediatrics 30:181–189
- Kivitie-Kallio S, Summanen P, Raitta C, Norio R (2000) Ophthalmologic findings in Cohen syndrome; a long-term follow-up. Ophthalmology 107:1737–1745
- Kolehmainen J, Norio R, Kivitie-Kallio S, Tahvanainen E, de la Chapelle A, Lehesjoki AE (1997) Refined mapping of the Cohen syndrome by gene linkage disequilibrium. Eur J Hum Genet 5:206–213
- Norio R, Raitta C, Lindahl E (1984) Further delineation of the Cohen syndrome; report on chorioretinal dystrophy, leukopenia and consanguinity. Clin Genet 25:1–14
- Summanen P, Kivitie-Kallio S, Norio R, Raitta C, Kivelä T (2002) Mechanisms of myopia in Cohen syndrome mapped to chromosome 8q22. Invest Ophthalmol Vis Sci 43:1686–1693
- Tahvanainen E, Norio R, Karila E, Ranta S, Weissenbach J, Sistonen P, de la Chapelle A (1994) Cohen syndrome gene assigned to the long arm of chromosome 8 by linkage analysis. Nat Genet 7:201–204

References for congenital chloride diarrhea

- Darrow DC (1945) Congenital alkalosis with diarrhea. J Pediatr 26:519–532
- Gamble JL, Fahey KR, Appleton J, MacLachlan E (1945) Congenital alkalosis with diarrhea. J Pediatr 26:509–518
- Haila S, Saarialho-Kere U, Karjalainen-Lindsberg ML, Lohi H, Airola K, Holmberg C, Hästbacka J, Kere J, Höglund P (2000) The congenital chloride diarrhea gene is expressed in seminal vesicle, sweat gland, inflammatory colon epithelium, and in some dysplastic colon cells. Histochem Cell Biol 113:279–286
- Holmberg C (1986) Congenital chloride diarrhoea. Clin Gastroenterol 15:583–602
- Holmberg C, Perheentupa J, Launiala K, Hallman N (1977) Congenital chloride diarrhea. A clinical analysis of 21 Finnish patients. Arch Dis Childh 52:255–267
- Höglund P, Haila S, Socha J, Tomaszewski L, Saarialho-Kere U, Karjalainen-Lindsberg ML, Airola K, Holmberg C, de la Chapelle A, Kere J (1996) Mutations of the down-regulated in adenoma (DRA) gene cause congenital chloride diarrhoea. Nat Genet 14:316–319
- Höglund P, Auranen M, Socha J, Popinska K, Nazer H, Rajaram U, Al Sanie A, Al-Ghanim M, Holmberg C, de la Chapelle A, Kere J (1998) Genetic background of congenital chloride diarrhea in two high frequency populations: Poland and Arabic countries. Am J Hum Genet 63:760–768

- Höglund P, Haila S, Gustavson KH, Taipale M, Hannula K, Popinska K, Holmberg C, Socha J, de la Chapelle A, Kere J (1998)
 Clustering of private mutations in the congenital chloride diarrhea / down-regulated in adenoma gene. Hum Mutat 11:321–327
- Höglund P, Holmberg C, Sherman P, Kere J (2001) Distinct outcomes of chloride diarrhoea in two siblings with identical genetic background of the disease: implications for early diagnosis and treatment. Gut 48:724–727
- Höglund P, Sormaala M, Haila S, Socha J, Rajaram U, Scheurlen W, Sinaasappel M, de Jonge H, Holmberg C, Yoshikawa H, Kere J (2001) Identification of seven novel mutations including the first two genomic rearrangements in SLC26A3 mutated in congenital chloride diarrhea. Hum Mutat 18:233–242
- Kagalwalla AF (1994) Congenital chloride diarrhea; a study in Arab children. J Clin Gastroenterol 19:36–40
- Kere J, Sistonen P, Holmberg C, de la Chapelle A (1993) The gene for congenital chloride diarrhea maps close to but is distinct from the gene for cystic fibrosis transmembrane conductance regulator. Proc Natl Acad Sci USA 90:10686–10689
- Kirkinen P, Jouppila P (1984) Prenatal ultrasonic findings in congenital chloride diarrhoea. Prenat Diagn 4:457–461
- Lubani MM, Doudin KI, Sharda DC, Shaltout AA, Al-Shab TS, Abdul Al YK, Said MA, Salhi MM, Ahmed SA (1989) Congenital chloride diarrhoea in Kuwaiti children. Eur J Pediatr 148:333–336
- Norio R, Perheentupa J, Launiala K, Hallman N (1971) Congenital chloride diarrhea, an autosomal recessive disease. Genetic study of 14 Finnish and 12 other families. Clin Genet 2:182– 192
- Perheentupa J, Eklund J, Kojo N (1965) Familial chloride diarrhea ("congenital alkalosis with diarrhea"). Acta Paediat Scand Suppl 159:119–120
- Tomaszewski L, Kulesza E, Socha J (1987) Congenital chloride diarrhoea in Poland. Materia Medica Polona 64:271–277

References for congenital lactase deficiency

- Holzel A, Schwarz V, Sutcliffe KW (1959) Defective lactose absorption causing malnutrition in infancy. Lancet I:1126–1128
- Järvelä I, Sabri Enattah N, Kokkonen J, Varilo T, Savilahti E, Peltonen L (1998) Assignment of the locus for congenital lactose deficiency to 2q21, in the vicinity of but separate from the lactase-phlorizin hydrolase gene. Am J Hum Genet 63:1078–1085
- Launiala K, Kuitunen P, Visakorpi JK (1966) Disaccharidases and histology of duodenal mucosa in congenital lactose malabsorption. Acta Paediatr Scand 55:257–263
- Saarela T, Similä S, Koivisto M (1995) Hypercalcemia and nephrocalcinosis in patients with congenital lactase deficiency. J Pediatr 127:920–923
- Savilahti E, Launiala K, Kuitunen P (1983) Congenital lactase deficiency; a clinical study on 16 patients. Arch Dis Childh 58: 246–252

References for congenital nephrosis of the Finnish type

- Beltcheva O, Martin P, Lenkkeri U, Tryggvason K (2001) Mutation spectrum in the nephrin gene (NPHS1) in congenital nephrotic syndrome. Hum Mutat 17:368–373
- Gautier P, Miville D (1942) Syndrome de néphrose lipóidique congénitale. Rev Med Suisse Romande 62:740–747
- Hallman N, Hjelt L, Ahvenainen EK (1956) Nephrotic syndrome in newborn and young infants. Ann Paediatr Fenn 2:227–241
- Haltia A, Solin ML, Muramatsu T, Jalanko H, Holmberg C, Miettinen A, Holthöfder H (1996) Mechanisms of proteinuria: Vascular permeability factor in congenital nephrotic syndrome of the Finnish type. Pediatr Res 40:652–657
- Holmberg C, Antikainen M, Rönnholm K, Ala-Houhala M, Jalanko H (1995) Management of the congenital nephrotic syndrome of the Finnish type. Pediatr Nephrol 9:87–93

- Holmberg C, Jalanko H, Tryggvason K, Rapola J (1999) Congenital nephrotic syndrome. In: Barratt TM, Avner ET, Harmon WE (eds) Pediatric nephrology, 4th edn. Lippincott Williams & Wilkins, Baltimore, pp 765–777
- Huttunen NP (1976) Congenital nephrotic syndrome of Finnish type. A study of 75 cases. Arch Dis Childh 51:344–348
- Jalanko H, Kääriäinen H, Norio R (2002) Nephrotic disorders. In: Rimoin DL, Connor JM, Pyeritz RE, Korf BR (eds) Emery & Rimoin's Principles and practice of medical genetics, 4th edn. Churchill Livingstone, London New York, pp 1708–1719
- Kestilä M, Männikkö M, Holmberg C, Gyapay G, Weissenbach J, Savolainen ER, Peltonen L, Tryggvason K (1994) Congenital nephrotic syndrome of the Finnish type maps to the long arm of chromosome 19. Am J Hum Genet 54:757–764
- Kestilä M, Lenkkeri U, Männikkö M, Lamerdin J, McCready P, Putaala H, Ruotsalainen V, Morita T, Nissinen M, Herva R, Kashtan CE, Peltonen L, Holmberg C, Olsen A, Tryggvason K (1998) Positionally cloned gene for a novel glomerular protein – nephrin – is mutated in congenital nephrotic syndrome. Molec Cell 1:575–582
- Laine J, Jalanko H, Holthöfer H, Krogerus L, Rapola J, von Willebrand E, Lautenschlager I, Salmela K, Holmberg C (1993) Post-transplantational nephrosis in congenital nephrotic syndrome of Finnish type. Kidney Int 44:867–874
- Lenkkeri U, Männikkö M, McCready P, Lamerdin J, Gribouval O, Niaudet P, Antignac C, Kashtan CE, Holmberg C, Olsen A, Kestilä M, Tryggvason K (1999) Structure of the gene for congenital nephrotic syndrome of the Finnish type (NPHS1) and characterization of mutations. Am J Hum Genet 64:51–61
- Liu L, Done SC, Knoshnoodi J, Bertorello A, Wartiovaara J, Berggren PO, Tryggvason K (2001) Defective nephrin trafficking caused by missense mutations in the *NPHS1* gene: insight into the mechanism of congenital nephrotic syndrome. Hum Mol Genet 10:2637–2644
- Männikkö M, Kestilä M, Holmberg C, Norio R, Ryynänen M, Olsen A, Peltonen L, Tryggvason K (1995) Fine mapping and haplotype analysis of the locus for congenital nephrotic syndrome on chromosome 19q13.1. Am J Hum Genet 57:1377– 1383
- Norio R (1966) Heredity in the congenital nephrotic syndrome; a genetic study of 57 Finnish families with a review of reported cases. Ann Paediatr Fenn Suppl 27
- Patrakka J, Kestilä M, Wartiovaara J, Ruotsalainen V, Tissari P, Lenkkeri U, Männnikkö M, Visapää I, Holmberg C, Rapola J, Tryggvason K, Jalanko H (2000) Congenital nephrotic syndrome (NPHS1): features resulting from different mutations in Finnish patients. Kidney Int 58:972–980
- Patrakka J, Martin P, Salonen R, Ruotsalainen V, Kestilä M, Männikkö M, Ryynänen M, Rapola J, Holmberg C, Tryggvason K, Jalanko H (2002) Proteinuria and prenatal diagnosis of congenital nephrosis in fetal carriers of nephrin gene mutations. Lancet 359:1575–1577
- Patrakka J, Ruotsalainen V, Reponen P, Qvist E, Laine J, Holmberg C, Tryggvason K, Jalanko H (2002) Recurrence of nephrotic syndrome in kidney grafts of patients with congenital nephrotic syndrome of the Finnish type: role of nephrin. Transplantation 73:394–403
- Rapola J, Huttunen NP, Hallman N (1992) Congenital and infantile nephrotic syndrome. In: Edelmann CM Jr (ed) Pediatric kidney disease, 2nd edn. Little Brown, Boston, pp 1291–1305
- Ruotsalainen V, Patrakka J, Tissari P, Hess M, Kestilä M, Holmberg C, Salonen R, Heikinheimo M, Wartiovaara J, Tryggvason K, Jalanko H (2000) Role of nephrin in cell junction formation in human nephrogenesis. Am J Pathol 157:1905–1916

References for cornea plana congenita

- Eriksson A, Lehmann W, Forsius H (1973) Congenital cornea plana in Finland. Clin Genet 4:301–310
- Forsius H (1957) Cornea plana and embryotoxon corneae posterius. Acta Ophthalmol 35:65

- Forsius H (1961) Studien über Cornea plana congenita bei 19 Kranken in 9 Familien. Acta Ophthalmol 39:203–221
- Forsius H, Damsten M, Eriksson AW, Fellman J, Lindh S, Tahvanainen E (1998) Autosomal recessive cornea plana; a clinical and genetic study of 78 cases in Finland. Acta Ophthalmol Scand 76:196–203
- Pellegata NS, Dieguez-Lucena JL, Joensuu T, Lau S, Montgomery KT, Krahe R, Kivelä T, Kucherlapati R, Forsius H, de la Chapelle A (2000) Mutations in KERA, encoding keratocan, cause cornea plana. Nat Genet 25:91–95
- Rübel E (1912) Kongenitale familiäre Flachheit der Kornea (Cornea plana). Klin Monatsbl Augenheilk 50:427–433
- Sigler-Villanueva A, Tahvanainen E, Lindh S, Dieguez-Lucena J, Forsius H (1997) Autosomal dominant cornea plana: clinical findings in a Cuban family and a review of the literature. Ophthalmol Genet 18:55–62
- Tahvanainen E, Forsius H, Karila E, Ranta S, Eerola M, Weissenbach J, Sistonen P, de la Chapelle A (1995) Cornea plana congenita gene assigned to the long arm of chromosome 12 by linkage analysis. Genomics 26:290–293
- Tahvanainen E, Forsius H, Kolehmainen J, Damsten M, Fellman J, de la Chapelle A (1996) The genetics of cornea plana congenita. J Med Genet 33:116–119
- Tahvanainen E, Villanueva AS, Forsius H, Salo P, de la Chapelle A (1996) Dominantly and recessively inherited cornea plana congenita map to the same small region of chromosome 12. Genome Res 6:249–254
- Tasheva ES, Pettenati M, Von Kap-Her C, Conrad GW (2000) Assignment of keratocan gene (KERA) to human chromosome band 12q22 by in situ hybridization. Cytogenet Cell Genet 88: 244–245
- Vesaluoma MH, Sankila EM, Gallar J, Muller LJ, Petroll WM, Moilanen JA, Forsius H, Tervo TM (2000) Autosomal recessive cornea plana: in vivo corneal morphology and corneal sensitivity. Invest Ophthalmol Vis Sci 41:2120–2126

References for diastrophic dysplasia

- Czarny-Ratajzak M, Lohiniva J, Rogala P, Kozlowski K, Perälä M, Carter L, Spector TD, Kolodziej L, Seppänen U, Glazar R, Krolewski J, Latos-Bielenska A, Ala-Kokko L (2001) A mutation in COL9A1 causes multiple epiphyseal dysplasia: further evidence for locus heterogeneity. Am J Hum Genet 69:969– 980
- Haila S, Hästbacka J, Böhling T, Karjalainen-Lindsberg ML, Kere J, Saarialho-Kere U (2001) SLC26A2 (Diastrophic dysplasia sulphate transporter) is expressed in developing and mature cartilage but also in other tissues and cell types. J Histochem Cytochem 49:973–982
- Horton WA, Rimoin DL, Lachman RS, Skovby F, Hollister DW, Spranger J, Scott CI, Hall JG (1978) The phenotypic variability of diastrophic dysplasia. J Pediatr 93:609–613
- Hästbacka J, Kaitila I, Sistonen P, de la Chapelle A (1990) Diastrophic dysplasia gene maps to the distal long arm of chromosome 5. Proc Natl Acad Sci USA 87:8056–8059
- Hästbacka J, de la Chapelle A, Kaitila I, Sistonen P, Weaver A, Lander E (1992) Linkage disequilibrium mapping in isolated founder populations: diastrophic dysplasia in Finland. Nat Genet 2:204–211
- Hästbacka J, de la Chapelle A, Mahtani M, Clines G, Reeve-Daly MP, Daly M, Hamilton BA, Kusumi K, Trivedi B, Weaver A, Coloma A, Lovett M, Buckler A, Kaitila I, Lander ES (1994) The diastrophic dysplasia gene encodes a novel sulfate transporter: positional cloning by fine-structure linkage disequilibrium mapping. Cell 78:1073–1087
- Hästbacka J, Superti-Furga A, Wilcox WR, Rimoin DL, Cohn DH, Lander ES (1996) Atelosteogenesis type II is caused by mutations in the diastrophic dysplasia sulfate-transporter gene (DTDST): evidence for a phenotypic series involving three chondrodysplasias. Am J Hum Genet 58:255–262

- Hästbacka J, Kerrebrock A, Mokkala K, Clines G, Lovett M, Kaitila I, de la Chapelle A, Lander ES (1999) Identification of the Finnish founder mutation for diastrophic dysplasia (DTD). Eur J Hum Genet 7:664–670
- Karniski LP (2001) Mutations in the diastrophic dysplasia sulfate transporter (DTDST) gene: correlation between sulfate transport activity and chondrodysplasia phenotype. Hum Mol Genet 10:1485–1490
- Lamy M, Maroteaux P (1960) Le nanisme diastrophique. Presse Méd 68:1977–1980
- Lapunzina P, Arberas C, del Carmen Fernandez M, Tello AM (1998) Diastrophic dysplasia diagnosed in a case published 100 years ago. Am J Med Genet 77:334–336
- Mäkitie O, Kaitila I (1997) Growth in diastrophic dysplasia. J Pediatr 130:641–646
- Peltonen J, Vaara P, Marttinen E, Poussa M, Ryöppy S (1999) Knee joint in diastrophic dysplasia. A clinical and radiographical study. J Bone Joint Surg (Br) 81:625–631
- Perheentupa J (1972) Kolme periytyvää kasvuhäiriötä (Cartilagehair hypoplasia, diastrophic nanism, mulibrey nanism; English summary) Duodecim 88:60–71
- Poussa M, Merikanto J, Ryöppy S, Marttinen E, Kaitila I (1991) The spine in diastrophic dysplasia. Spine 16:881–887
- Remes V, Poussa M, Peltonen J (2001) Scoliosis in patients with diastrophic dysplasia: a new classification. Spine 26:1689–1697
- Remes V, Helenius I, Peltonen J, Poussa M, Sovijärvi A (2002) Lung function in diastrophic dysplasia. Pediatr Pulmonol 33: 277–282
- Remes VM, Marttinen EJ, Poussa MS, Helenius IJ, Peltonen JI (2002) Cervical spine in patients with diastrophic dysplasia–radiographic findings in 122 patients. Pediatr Radiol 32:621–628
- Remes V, Tervahartiala P, Helenius I, Peltonen J (2002) Magnetic resonance imaging analysis of hip joint development in patients with diastrophic dysplasia. J Pediatr Orthop 22:212–216
- Rintala A, Marttinen E, Rantala SL, Kaitila I (1986) Cleft palate in diastrophic dysplasia; morphology, results of treatment and complications. Scand J Plast Reconstr Surg 20:45–49
- Ryöppy S, Poussa M, Merikanto J, Marttinen E, Kaitila I (1990) Deformities of the lower extremities in diastrophic dysplasia. Acta Orthop Scand 61 Suppl 237:49
- Ryöppy S, Poussa M, Merikanto J, Marttinen E, Kaitila I (1992) Foot deformities in diastrophic dysplasia; an analysis of 102 patients. J Bone Joint Surg (Br) 74:441–444
- Vaara P, Peltonen J, Poussa M, Merikanto J, Nurminen M, Kaitila I, Ryöppy S (1998) Development of the hip in diastrophic dysplasia. J Bone Joint Surg (Br) 80:315–320
- Vaara P, Sintonen H, Peltonen J, Hokkanen H, Pousssa M, Ryöppy S (1999) Health-related quality of life in patients with diastrophic dysplasia. Scand J Public Health 27:38–42
- Walker BA, Scott CI, Hall JG, Murdoch JL, McKusick VA (1972) Diastrophic dwarfism. Medicine 51:41–59

References for FSH-RO

- Aittomäki K (1994) The genetics of XX gonadal dysgenesis. Am J Hum Genet 54:844–851
- Aittomäki K, Dieguez Lucena JL, Pakarinen P, Sistonen P, Tapaninen J, Gromoll J, Kaskikari R, Sankila EM, Lehväslaiho H, Engel AR, Nieschlag E, Huhtaniemi I, de la Chapelle A (1995) Mutation in the follicle-stimulating hormone receptor gene causes hereditary hypergonadotropic ovarian failure. Cell 82: 959–968
- Aittomäki K, Herva R, Stenman UH, Juntunen K, Ylöstalo P, Hovatta O, de la Chapelle (1996) Clinical features of primary ovarian failure caused by a point mutation in the follicle-stimulating hormone receptor gene. J Clin Endocrinol Metabol 81: 3722–3726

- Doherty E, Pakarinen P, Tiitinen A, Kiilavuori A, Huhtaniemi I, Forrest S, Aittomäki K (2002) A novel mutation in the folliclestimulating hormone receptor inhibing signal transduction and causing primary ovarian failure. J Clin Endocrinol Metabol 87: 1151–1155
- Rannikko A, Pakarinen P, Manna P, Beau I, Milrom E, Misrahi M, Aittomäki K, Huhtaniemi I (2002) Functional characterization of the human follicle-stimulating hormone receptor with inactivating Ala189Val mutation. Molec Human Reprod 4:311–317
- Simpson JL, Christakos AC, Horwith M, Silverman FS (1971) Gonadal dysgenesis in individuals with apparently normal chromosomal complements: tabulation of cases and compilation of genetic data. Birth Defects Original Article Series Vol 7 Part X:215–228
- Vaskivuo TE, Aittomäki K, Anttonen M, Ruokonen A, Herva R, Osawa Y, Heikinheimo M, Huhtaniemi I, Tapanainen JS (2002) Effects of follicle-stimulating hormone (FSH) and human chorionic gonadotropin in individuals with an inactivating mutation of the FSH receptor. Fertil Steril 78:108–113

References for GRACILE syndrome

- Fellman V, Rapola J, Pihko H, Varilo T, Raivio K (1998) Ironoverload disease in infants involving fetal growth retardation, lactic acidosis, liver hemosiderosis, and aminoaciduria. Lancet 351:490–493
- Fellman V, von Bonsdorff L, Parkkinen J (2000) Exogenous apotransferrin and exchange transfusions in hereditary iron overload disease. Pediatrics 105:398–401
- Fellman V, Visapää I, Vujic M, Wennerholm UB, Peltonen L (2002) Antenatal diagnosis of hereditary fetal growth retardation with aminoaciduria, cholestasis, iron overload, and lactic acidosis in the newborn infant. Acta Obst Gynecol Scand 81:398–402
- Norio R, Nevanlinna HR, Perheentupa J (1973) Hereditary diseases in Finland; rare flora in rare soil. Ann Clin Res 5:109– 141
- Rapola J, Heikkilä P, Fellman V (2002) Pathology of lethal fetal growth retardation syndrome with aminoaciduria, iron overload, and lactic acidosis (GRACILE). Pediatr Pathol Mol Med 21:183–193
- Visapää I, Fellman V, Varilo T, Palotie A, Raivio KO, Peltonen L (1998) Assignment of the locus for a new lethal neonatal metabolic syndrome to 2q33–37. Am J Hum Genet 63:1396–1403
- Visapää I, Fellman V, Vesa J, Dasvarma A, Hutton JL, Kumar V, Payne GS, Makarow M, Van Coster R, Taylor RW, Turnbull DM, Suomalainen A, Peltonen L (2002) GRACILE syndrome, a lethal metabolic disorder with iron overload, is caused by a point mutation in *BCS1L*. Am J Hum Genet 71:863–876

References for Herva disease

- Herva R, Leisti J, Kirkinen P, Seppänen U (1985) A lethal autosomal recessive syndrome of multiple congenital contractures. Am J Med Genet 20:431–439
- Herva R, Conradi NG, Kalimo H, Leisti J, Sourander P (1988) A syndrome of multiple congenital contractures: neuropathological analysis on five fetal cases. Am J Med Genet 29:67–76
- Kirkinen P, Herva R, Leisti J (1987) Early prenatal diagnosis of a lethal syndrome of multiple congenital contractures. Prenat Diagn 7:189–196
- Mäkelä-Bengs P, Järvinen N, Vuopala K, Suomalainen A, Ignatius J, Sipilä M, Herva R, Palotie A, Peltonen L (1998) Assignment of the disease locus for lethal congenital contracture syndrome to a restricted region of chromosome 9q34, by genome scan using five affected individuals. Am J Hum Genet 63:506–516
- Vuopala K, Herva R (1994) Lethal congenital contracture syndrome–further delineation with genetic aspects. J Med Genet 31:521–527

Vuopala K, Mäkelä-Bengs P, Suomalainen A, Herva R, Leisti J, Peltonen L (1995) Lethal congenital contracture syndrome – a fetal SMA-like disease is not linked to SMA-locus 5q. J Med Genet 32:36–38

References for HOGA

- Cutler CW (1895) Drei ungewöhnliche Fälle von Retino-Chorioideal-Degeneration. Arch Augenheilk 30:117
- Heinänen K, Näntö-Salonen K, Komu M, Erkintalo M, Alanen A, Heinonen OJ, Pulkki K, Nikoskelainen E. Sipilä I, Simell O (1999) Creatine corrects muscle ³¹P spectrum in gyrate atrophy with hyperornithinemia. Eur J Clin Invest 29:1060–1065
- Jacobsohn E (1888) Ein Fall von Retinitis pigmentosa atypica. Klin Monatsbl Augenheilk 26:202–206
- Kaiser-Kupfer MI, Caruso RC, Valle D (2002) Gyrate atrophy of the choroid and retina: further experience with long-term reduction of ornithine levels in children. Arch Ophthalmol 129: 146–153
- Peltola K, Heinonen OJ, Näntö-Salonen K, Pulkki K, Simell O (2000) Oral lysine feeding in gyrate atrophy with hyperornithinemia – a pilot study. J Inherit Metab Dis 23:305–307
- Peltola KE, Näntö-Salonen K, Heinonen OJ, Heinänen K, Jääskeläinen S, Simell O, Nikoskelainen E (2001) Ophthalmological heterogeneity in patients with gyrate atrophy of choroid and retina harboring the L402P mutation of ornithine aminotransferase. Ophthalmology 108:721–729
- Simell O, Takki K (1973) Raised plasma-ornithine and gyrate atrophy of the choroid and retina. Lancet I:1031
- Sipilä I, Simell O, Rapola J, Sainio K, Tuuteri L (1979) Gyrate atrophy of the choroid and retina with hyperornithinemia: tubular aggregates and type 2 fiber atrophy in muscle. Neurology 29: 996–1005
- Takki K (1975) Gyrate atrophy of the choroid and retina associated with hyperornithinaemia. Brit J Ophthalmol 58:3–23
- Takki K, Simell O (1974) Genetic aspects in gyrate atrophy of the choroid and retina with hyperornithinaemia. Brit J Ophthalmol 58:907–916
- Valle D, Simell O (2001) The hyperornithinemias. In: Scriver CR, Beaudet AL, Valle D, Sly WS (eds) The metabolic & molecular bases of inherited disease, 8th edn. McGraw-Hill, New York, pp 1857–1895

References for hydrolethalus syndrome

- Herva R, Seppänen U (1984) Roentgenologic findings of the hydrolethalus syndrome. Pediatr Radiol 14:41–43
- Salonen R, Herva R (1990) Syndrome of the month: hydrolethalus syndrome. J Med Genet 27:756–759
- Salonen R, Herva R, Norio R (1981) The hydrolethalus syndrome: delineation of a "new", lethal malformation syndrome based on 28 patients. Clin Genet 19:321–330
- Visapää I, Salonen R, Varilo T, Paavola P, Peltonen L (1999) Assignment of the locus for hydrolethalus syndrome to a highly restricted region on 11q23–25. Am J Hum Genet 65:1086– 1095
- Ämmälä P, Salonen R (1995) First-trimester diagnosis of hydrolethalus syndrome. Ultrasound Obstet Gynecol 5:60–62

References for INCL

- Cho S, Dawson PE, Dawson G (2001) Role of palmitoyl-protein thioesterase in cell death: implications for infantile neuronal ceroid lipofuscinosis. Eur J Paediatr Neurol 5 Suppl A:53–55
- Hagberg B, Sourander P, Svennerholm L (1968) Late infantile progressive encephalopathy with disturbed polyunsaturated fat metabolism. Acta Paediat Scand 57:495–499

- Haltia M, Rapola J, Santavuori P (1973) Infantile type of so-called neuronal ceroid-lipofuscinosis. Histological and electron-microscopic studies. Acta Neuropathol (Berl) 26:157–170
- Haltia M, Rapola J, Santavuori P, Keränen P (1973) Infantile type of so-called neuronal ceroid-lipofuscinosis. Part 2. Morphological and biochemical studies. J Neurol Sci 18:269–285
- Hellsten E, Vesa J, Speer MC, Mäkelä TP, Järvelä I, Alitalo K, Ott J, Peltonen L (1993) Refined assignment of the infantile neuronal ceroid lipofuscinosis (INCL, CLN1) locus at 1p32: incorporation of linkage disequilibrium in multipoint analysis. Genomics 16, 720–725
- Hofmann SL, Das AK, Lu JY, Wisniewski KE, Gupta P (2001) Infantile neuronal ceroid lipofuscinosis: no longer just a 'Finnish' disease. Eur J Paediatr Neurol 5 Suppl A:47–51
- Järvelä I, Schleutker J, Haataja L, Santavuori P, Puhakka L, Manninen T, Palotie A, Sandkuijl LA, Renlund M, White R, Aula P, Peltonen L (1991) Infantile neuronal ceroid lipofuscinosis (INCL, CLN1) maps to the short arm of chromosome 1. Genomics 8:170–173
- Lehtovirta M, Kyttälä A, Eskelinen EL, Hess M, Heinonen O, Jalanko A (2001) Palmitoyl protein thioesterase (PPT) localizes into synaptosomes and synaptic vesicles in neurons: implications for infantile neuronal ceroid lipofuscinosis (INCL). Hum Mol Genet 10:69–75
- Lu JY, Verkruyse LA, Hofmann SL (2002) The effects of lysosomotropic agents on normal and INCL cells provide further evidence for the lysosomal nature of palmitoyl-protein thioesterase function. Biochim Biophys Acta 1583:35–44
- Lönnqvist T, Vanhanen SL, Vettenranta K, Autti T, Rapola J, Santavuori P, Saarinen-Pihkala UM (2001) Hematopoietic stem cell transplantation in infantile neuronal ceroid lipofuscinosis. Neurology 57:1411–1416
- Mannerkoski MK, Heiskala HJ, Santavuori PR, Pouttu JA (2001) Transdermal fentanyl therapy for pains in children with infantile neuronal ceroid lipofuscinosis. Eur J Paediatr Neurol 5 Suppl A:175–177
- Raitta C, Santavuori P (1973) Ophthalmological findings in infantile type of so-called neuronal ceroid lipofuscinosis. Acta Ophthalmol 51:755–763
- Riikonen R, Vanhanen SL, Tyynelä J, Santavuori P, Turpeinen U (2000) CSF insulin-like growth factor-1 in infantile neuronal ceroid lipofuscinosis. Neurology 54:1828–1832
- Salonen T, Heinonen-Kopra O, Vesa J, Jalanko A (2001) Neuronal trafficking of palmitoyl protein thioesterase provides an excellent model to study the effects of different mutations which cause infantile neuronal ceroid lipofuscinosis. Mol Cell Neurosci 18:131–140
- Salonen T, Järvelä I, Peltonen L, Jalanko A (2001) Detection of eight novel palmitoyl protein thioesterase (PPT) mutations underlying infantile neuronal ceroid lipofuscinosis (INCL; CLN1). Hum Mutat 15:273–279
- Santavuori P, Haltia M, Rapola J, Raitta C (1973) Infantile type of so-called neuronal ceroid-lipofuscinosis. Part 1. A clinical study of 15 patients. J Neurol Sci 18:257–267
- Van Diggelen OP, Keulemans JL, Kleijer WJ, Thobois S, Tilikete C, Voznyi YV (2001) Pre- amd postnatal enzyme analysis for infantile, late infantile and adult neuronal ceroid lipofuscinosis (CLN1 and CLN2). Eur J Paediatr Neurol 5 Suppl A:189–192
- Vanhanen SL, Raininko R, Autti T, Santavuori P (1995) MRI evaluation of the brain in infantile neuronal ceroid- lipofuscinosis. Part 2: MRI findings in 21 patients. J Child Neurol 10:444–450
- Vanhanen SL, Sainio K, Lappi M, Santavuori P (1997) EEG and evoked potentials in infantile neuronal ceroid-lipofuscinosis. Develop Med Child Neurol 39:456–463
- Vesa J, Hellsten E, Verkruyse LA, Camp LA, Rapola J, Santavuori P, Hofmann SL, Peltonen L (1995) Mutations in the palmitoyl protein thioesterase gene causing infantile neuronal ceroid lipofuscinosis. Nature 376:584–587

References for IOSCA syndrome

- Kallio AK, Jauhiainen T (1985) A new syndrome of ophthalmoplegia, hypoacusis, ataxia, hypotonia and athetosis (OHAHA). Adv Audiol 3:84–90
- Koskinen T, Sainio K, Rapola J, Pihko H, Paetau A (1994) Sensory neuropathy in infantile onset spinocerebellar ataxia (IOSCA). Muscle & Nerve 17:509–515
- Koskinen T, Santavuori P, Sainio K, Lappi M, Kallio AK, Pihko H (1994) Infantile onset spinocerebellar ataxia with sensory neuropathy: a new inherited disease. J Neurol Sci 121:50–56
- Lönnqvist T, Paetau A, Nikali K, von Boguslawski K, Pihko H (1998) Infant onset spinocerebellar ataxia with sensory neuropathy (IOSCA): neuropathological features. J Neurol Sci 161: 57–65
- Nikali K, Suomalainen A, Terwilliger J, Koskinen T, Weissenbach J, Peltonen L (1995) Random search for shared chromosomal regions in four affected individuals: the assignment of a new hereditary ataxia locus. Am J Hum Genet 56:1088–1095
- Nikali K, Isosomppi J, Lönnqvist T, Mao JI, Suomalainen A, Peltonen L (1997) Toward cloning of a novel ataxia gene: refined assignment and physical map of the IOSCA locus (SCA8) on 10q24. Genomics 39:185–191
- Varilo T, Nikali K, Suomalainen A, Lönnqvist T, Peltonen L (1996) Tracing an ancestral mutation: genealogical and haplotype analysis of the infantile onset spinocerebellar ataxia locus. Genome Res 6:870–875

References for Jansky-Bielschowsky disease, Finnish variant

- Autti T, Raininko R, Launes J, Nuutila A, Santavuori P (1992) Jansky-Bielschowsky variant disease: CT, MRI and SPECT findings. Pediatr Neurol 8:121–126
- Holmberg V, Lauronen L, Autti T, Santavuori P, Savukoski M, Uvebrant P, Hofman I, Peltonen L, Järvelä I (2000) Phenotypegenotype correlation in eight patients with Finnish variant late infantile NCL (CLN5). Neurology 55:579–581
- Isosomppi J, Vesa J, Jalanko A, Peltonen L (2002) Lysosomal localization of the neuronal lipofuscinosis CLN5 protein. Hum Mol Genet 11:885–891
- Kirveskari E, Partinen M, Santavuori P (2001) Sleep and its disturbance in a variant form of late infantile neuronal ceroid lipofuscinosis (CLN5). J Child Neurol 16:707–713
- Klockars T, Savukoski M, Isosomppi J, Laan M, Järvelä I, Petrukhin K, Palotie A, Peltonen L (1996) Efficient construction of a physical map by fiber-fish of the CLN5 region: refined assignment and long-range contig covering the critical region on 13q22. Genomics 35:71–78
- Lauronen L, Huttunen J, Kirveskari E, Wikström H, Sainio K, Autti T, Santavuori P (2002) Enlarged SI and SII somatosensory evoked responses in the CLN5 form of neuronal ceroid lipofuscinosis. Clin Neurophysiol 113:1491–1500
- Santavuori P, Rapola J, Sainio K, Raitta C (1982) A variant of Jansky-Bielschowsky disease. Neuropediatrics 13:135–141
- Santavuori P, Rapola J, Nuutila A, Raininko R, Lappi M, Launes J, Herva R, Sainio K (1991) The spectrum of Jansky-Bielschowsky disease. Neuropediatrics 22:92–96
- Savukoski M, Kestilä M, Williams R, Järvelä I, Sharp J, Harris J, Santavuori P, Gardiner M, Peltonen L (1994) Defined chromosomal assignment of CLN5 demonstrates that at least four genetic loci are involved in the pathogenesis of human ceroid lipofuscinoses. Am J Hum Genet 55:695–701
- Savukoski M, Klockars T, Holmberg V, Santavuori P, Lander ES, Peltonen L (1998) CLN5, a novel gene encoding a putative transmembrane protein mutated in Finnish variant late infantile neuronal ceroid lipofuscinosis. Nat Genet 19:286–288
- Sharp JD, Wheeler RB, Lake BD, Savukoski M, Järvelä IE, Peltonen L, Gardiner RM, Williams RE (1997) Loci for classical and a variant late infantile neuronal ceroid lipofuscinosis map to chromosomes 11p15 and 15q21–23. Hum Mol Genet 6:591– 595

Varilo T, Savukoski M, Norio R, Santavuori P, Peltonen L, Järvelä I (1996) The age of human mutation: genealogical and linkage disequilibrium analysis of the CLN5 mutation in the Finnish population. Am J Hum Genet 58:506–512

References for lysinuric protein intolerance

- DiRocco M, Garibotto G, Rossi GA, Caruso U, Taccone A, Picco P, Borrone C (1993) Role of haematological, pulmonary and renal complications in the long-term prognosis of patients with lysinuric protein intolerance. Eur J Pediatr 152:437–440
- Kekomäki M, Visakorpi JK, Perheentupa J, Saxén L (1967) Familial protein intolerance with deficient transport of basic amino acids; an analysis of 10 patients. Acta Paediat Scand 56: 617–630
- Koizumi A, Shoji Y, Nozaki J, Noguchi A, E X, Dakeishi M, Ohura T, Tsuyoshi K, Yasuhiko W, Manabe M, Takasago Y, Takada G (2000) A cluster of lysinuric protein intolerance (LPI) patients in a northern part of Iwate, Japan due to a founder effect. The Mass Screening Group. Hum Mutat 16: 270–271
- Lauteala T, Sistonen P, Savontaus ML, Mykkänen J, Simell J, Lukkarinen M, Simell O, Aula P (1997) Lysinuric protein intolerance (LPI) gene maps to the long arm of chromosome 14. Am J Hum Genet 60:1479–1486
- Lauteala T, Mykkänen J, Sperandeo MP, Gasparini P, Savontaus ML, Simell O, Andria G, Sebastio G, Aula P (1998) Genetic homogeneity of lysinuric protein intolerance. Eur J Hum Genet 6:612–615
- Lukkarinen M, Näntö-Salonen K, Pulkki K, Mattila K, Simell O (2000) Effect of lysine infusion on urea cycle in lysinuric protein intolerance. Metabolism 49:621–625
- Mykkänen J, Torrents D, Pineda M, Camps M, Yoldi ME, Horelli-Kuitunen N, Huoponen K, Heinonen M, Oksanen J, Simell O, Savontaus ML, Zorzano A, Palacín M, Aula P (2000) Functional analysis of novel mutations in y+LAT-1 amino acid transporter gene causing lysinuric protein intolerance. Hum Mol Genet 9:431–438
- Noguchi A, Shoji Y, Koizumi A, Takahashi T, Matsumori M, Kayo T, Ohata T, Wada Y, Yoshimura I, Maisawa S, Konishi M, Takasago Y, Takada G (2000) SLC7A7 genomic structure and novel variants in three Japanese lysinuric protein intolerance families. Hum Mutat 15:367–372
- Norio R, Perheentupa J, Kekomäki M, Visakorpi JK (1971) Lysinuric protein intolerance, an autosomal recessive disease; a genetic study of 10 Finnish families. Clin Genet 2:214–222
- Parto K, Svedström E, Majurin ML, Härkönen R, Simell O (1993) Pulmonary manifestations in lysinuric protein intolerance. Chest 104:1176–1182
- Perheentupa J, Visakorpi JK (1965) Protein intolerance with deficient transport of basic amino acids: another inborn error of metabolism. Lancet II:813–816
- Rajantie J, Simell O, Rapola J, Perheentupa J (1980) Lysinuric protein intolerance: a two-year trial of dietary supplementation therapy with citrulline and lysine. J Pediatr 97:927–932
- Simell O (2001) Lysinuric protein intolerance and other kationic aminoacidurias. In: Scriver CR, Beaudet AL, Valle D, Sly WS (eds) The metabolic & molecular bases of inherited disease, 8th edn. McGraw-Hill, New York, pp 4933–4956
- Simell O, Perheentupa J, Rapola J, Visakorpi JK; Eskelin LE (1975) Lysinuric protein intolerance. Am J Med 59:229–239
- Sperandeo MP, Bassi MT, Riboni M, Parentio G, Buoninconti A, Manzoni M, Incertio B, Larocca MR, Di Rocco M, Strisciuglio P, Dianzani I, Parini R, Candito M, Endo F, Ballabio A, Andria G, Sebastio G, Borsani G (2000) Structure of the SLC7A7 gene and mutational analysis of patients affected by lysinuric protein intolerance. Am J Hum Genet 66:92–99
- Toivonen M, Mykkänen J, Aula P, Simell O, Savontaus ML, Huoponen K (2002) Expression of normal and mutant GFP-tagged y(+)L amino acid transporter-1 in mammalian cells. Biochem Biophys Res Commun 29:1173–1179

Torrents D, Mykkänen J, Pineda M, Feliubadaló L, Estévez R, de Cid R, Sanjurjo P, Zorzano A, Nunes V, Huoponen K, Reinikainen A, Simell O, Savontaus ML, Aula P, Palacín (1999) Identification of SLC7A7, encoding y+LAT-1, as the lysinuric protein intolerance gene. Nat Genet 21:293–296

References for Meckel syndrome

- Aula P, Karjalainen O, Rapola J, Lindgren J, Seppälä M (1977) Prenatal diagnosis of the Meckel syndrome. Am J Obstet Gynecol 129:700–702
- Gruber GB (1934) Beiträge zur Frage "gekoppelter" Missbildungen (Akrocephalo-Syndactylie und Dysencephalia splanchnocystica). Beitr Pathol Anat 93:459–476
- Johnson VP, Holzwarth DR (1984) Prenatal diagnosis of Meckel syndrome: case reports and literature review. Am J Med Genet 18:699–711
- Lurie IW, Prytkov AN, Meldere LV (1984) Meckel syndrome in different populations. Am J Med Genet 18:661–669
- Meckel JF (1822) Beschreibung zweier, durch sehr ähnliche Bildungsabweichungen entstellter Geschwister. Dtsch Arch Physiol 7:99–172
- Moerman P, Verbeken E, Fryns JP, Goddeeris P, Lauweryns JM (1982) The Meckel syndrome. Pathological and cytogenetic observations in eight cases. Hum Genet 62:240–245
- Morgan NV, Gissen P, Malik Sharif S, Baumber L, Sutherland J, Kelly DA, Aminu K, Bennett CP, Woods CG, Mueller RF, Trembath RC, Maher ER, Johnson CA (2002) A novel locus for Meckel-Gruber syndrome, *MKS3*, maps to chromosome 8q24. Hum Genet 111:456-461
- Paavola P. Salonen R, Weissenbach J, Peltonen L (1995) The locus for Meckel syndrome with multiple congenital anomalies maps to chromosome 17q21-q24. Nat Genet 11:213–215
- Paavola P, Salonen R, Baumer A, Schinzel A, Boyd PA, Gould S, Meusburger H, Tenconi R, Barnicoat A, Winter R, Peltonen L (1997) Clinical and genetic heterogeneity in Meckel syndrome. Hum Genet 101:88–92
- Paavola P, Avela K, Horelli-Kuitunen N, Bärlund M, Kallioniemi A, Idänheimo N, Kyttälä M, de la Chapelle A, Palotie A, Lehesjoki AE, Peltonen L (1999) High-resolution physical and genetic mapping of the critical region for Meckel syndrome and mulibrey nanism on chromosome 17q22-q23. Genome Res 9:267–276
- Paetau A, Salonen R, Haltia M (1985) Brain pathology in the Meckel syndrome, a study of 59 cases. Clin Neuropathol 4:56– 62
- Rapola J, Salonen R (1985) Visceral anomalies in the Meckel syndrome. Teratology 31:193–201
- Roume J, Genin E, Cormier-Daire V, Ma HW, Mehaye B, Attie T, Razavi-Encha F, Munnich A, Le Merrer M (1998) A gene for Meckel syndrome maps to chromosome 11q13. Am J Hum Genet 63:1095–1101
- Salonen R (1984) The Meckel syndrome: clinicopathological findings in 67 patients. Am J Med Genet 18:671–689
- Salonen R, Norio R (1984) The Meckel syndrome in Finland: epidemiological and genetic aspects. Am J Med Genet 18:691– 698
- Salonen R, Paavola P (1998) Syndrome of the month: Meckel syndrome. J Med Genet 35:497–501
- Seppänen U, Herva R (1983) Roentgenologic features of the Meckel syndrome. Pediatr Radiol 13:329–331
- Sergi C, Adam S, Kahl P, Otto HF (2000) Study of the malformation of ductal plate of the liver in Meckel syndrome and review of other syndromes presenting with this anomaly. Pediatr Dev Pathol 3:568–583
- Teebi AS, al Saleh QA, Odeh H (1992) Meckel syndrome and neural tube defects in Kuwait. J Med Genet 29:140
- Young ID, Rickett AB, Clarke M (1985) High incidence of Meckel's syndrome in Gujarati Indians. J Med Genet 22:301– 304

References for Meretoja disease

- Chen CD, Huff ME, Matteson J, Page L, Phillips R, Kelly JW, Balch WE (2001) Furin initiates gelsolin familial amyloidosis in the Golgi through a defect in Ca(2+) stabilization. EMBO J 20:6277–6287
- Haltia M, Ghiso J, Prelli F, Gallo G, Kiuru S, Somer H, Palo J, Frangione B (1990) Amyloid in familial amyloidosis, Finnish type, is antigenically and structurally related to gelsolin. Am J Pathol 136:1223–1228
- Kangas H, Seidah NG, Paunio T (2002) Role of proprotein convertases in the pathogenic processing of the amyloidosis-associated form of secretory gelsolin. Amyloid 9:83–87
- Kazmirski SL, Isaacson RL, An C, Buckle A, Johnson CM, Daggett V, Fersht AR (2002) Loss of metal-binding site in gelsolin leads to familial amyloidosis Finnish type. Nat Struct Biol 9:112–116
- Kiuru S (1992) Familial amyloidosis of the Finnish type (FAF): a clinical study of 30 patients. Acta Neurol Scand 86:346–353
- Kiuru S (1998) Gelsolin-related familial amyloidosis, Finnish type (FAF), and its variants found worldwide. Amyloid Int J Exp Clin Invest 5:55–66
- Kiuru S,Seppäläinen AM (1994) Neuropathy in familial amyloidosis, Finnish type (FAF): electrophysiological studies. Muscle Nerve 17:299–304
- Kiuru S, Nieminen T, Partinen M (1999) Obstructive sleep apnea syndrome in hereditary gelsolin-related amyloidosis. J Sleep Res 81:143–149
- Kiuru S, Salonen O, Haltia M (1999) Gelsolin-related spinal and cerebral amyloid angiopathy. Ann Neurol 45:305–311
- Kiuru S, Javela K, Somer H, Kekomäki R (2000) Altered platelet shape change in hereditary gelsolin Asp187Asn amyloidosis. Thromb Haemost 83:491–495
- Kiuru-Enari S, Somer H, Seppäläinen AM, Notkola IL, Haltia M (2002) Neuromuscular pathology in hereditary gelsolin amyloidosis. J Neuropathol Exp Neurol 61:565–571
- Kwiatkowski DJ, Westbrook CA, Bruns GAP, Morton CC (1988) Localization of gelsolin proximal to ABL on chromosome 9. Am J Hum Genet 42:565–572
- Maury CPJ, Alli K, Baumann M (1990) Finnish hereditary amyloidosis. Amino acid sequence homology between the amyloid fibril protein and human plasma gelsoline. FEBS Lett 260:85– 87
- Maury CPJ, Kere J, Tolvanen R, de la Chapelle A (1990) Finnish hereditary amyloidosis is caused by a single nucleotide substitution in the gelsolin gene. FEBS Lett 276:75–77
- Maury CP, Liljeström M, Boysen G, Törnroth T, de la Chapelle A, Nurmiaho-Lassila EL (2000) Danish type gelsolin related amyloidosis: 654G>T mutation is associated with a disease pathogenetically and clinically similar to that caused by the 654G>A mutation (familial amyloidosis of the Finnish type). J Clin Pathol 53:95–99
- Meretoja J (1969) Familial systemic paramyloidosis with lattice dystrophy of the cornea, progressive cranial neuropathy, skin change 324
- Meretoja J (1973) Genetic aspects of familial amyloidosis with corneal lattice dystrophy and cranial neuropathy. Clin Genet 4: 173–185
- Paunio T, Kangas H, Kalkkinen N, Haltia M, Palo J, Peltonen L (1994) Towards understanding the pathogenetic mechanism in gelsolin-related amyloidosis: in vitro expression reveals an abnormal gelsolin fragment. Hum Mol Genet 3:2223–2229
- Rintala AE, Alanko A, Mäkinen J, Nordström R, Salo H (1988) Primary hereditary systemic amyloidosis (Meretoja's syndrome): clinical features and treatment by plastic surgery. Scand J Plast Reconstr Surg 22:141–145
- Rosenberg ME, Tervo TM, Gallar J, Acosta MC, Muller LJ, Moilanen JA, Tarkkanen AH, Vesaluoma MH (2001) Corneal morphology and sensitivity in lattice dystrophy type II (familial amyloidosis, Finnish type). Invest Ophthalmol Vis Sci 42: 634–641

References for mulibrey nanism

- Avela K, Lipsanen-Nyman M, Perheentupa J, Wallgren-Pettersson C, Marchand S, Fauré S, Sistonen P, de la Chapelle A, Lehesjoki AE (1997) Assignment of the mulibrey nanism gene to 17q by linkage and linkage-disequilibrium analysis. Am J Hum Genet 60:896–902
- Avela K, Lipsanen-Nyman M, Idänheimo N, Seemanová E, Rosengren S, Mäkelä TP, Perheentupa J, de la Chapelle A, Lehesjoki AE (2000) Gene encoding a new RING-B-box-coiled-coil protein is mutated in mulibrey nanism. Nat Genet 25:298–301
- Kallijärvi J, Avela K, Lipsanen-Nyman M, Ulmanen I, Lehesjoki AE (2002) The *TRIM37* gene encodes a peroxisomal RING-Bbox-coiled-coil protein: classification of mulibrey nanism as a new peroxisomal disorder. Am J Hum Genet 70:1215–1228
- Lapunzina P, Rodríguez JI, de Matteo E, Gracia R, Moreno F (1995) Mulibrey nanism: three additional patients and a review of 39 patients. Am J Med Genet 55:349–355
- Lipsanen-Nyman M (1986) Mulibrey-nanismi (in Finnish, English summary). Thesis, University of Helsinki
- Paavola P, Avela K, Horelli-Kuitunen N, Bärlund M, Kallioniemi A, Idänheimo N, Kyttälä M, de la Chapelle A, Palotie A, Lehesjoki AE, Peltonen L (1999) High resolution physical and genetic mapping of the critical region for Meckel syndrome and mulibrey nanism on chromosome 17q22-q23. Genome Res 9:267–276
- Perheentupa J, Autio S, Leisti S, Raitta C (1970) Mulibreynanism: dwarfism with muscle, liver, brain and eye involvement. Acta Paediatr Scand 59 Suppl 206:74–75
- Perheentupa J, Autio S, Leisti S, Raitta C, Tuuteri L (1973) Mulibrey nanism, an autosomal recessive syndrome with pericardial constriction. Lancet II:351–355
- Perheentupa J, Autio S, Leisti S, Raitta C, Tuuteri L (1975) Mulibrey nanism: review of 23 cases of a new autosomal recessive syndrome. Birth Defects Original Article Series Vol XI No 2: 3–17
- Raitta C, Perheentupa J (1974) Mulibrey nanism; an inherited dysmorphic syndrome with characteristic ocular findings. Acta Ophthalmol Suppl 123:162–171

References for muscle-eye-brain disease

- Auranen M, Rapola J, Pihko H, Haltia M, Leivo I, Soinila S, Virtanen I, Kalimo H, Anderson LV, Santavuori P, Somer H (2000) Muscle membrane-skeleton protein changes and histopathological characterization of muscle-eye-brain disease. Neuromuscul Disord 10:16–23
- Cormand B, Avela K, Pihko H, Santavuori P, Talim B, Topaloglu H, de la Chapelle A, Lehesjoki AE (1999) Assignment of the muscle-eye-brain disease gene to 1p32-p34 by linkage analysis and homozygosity mapping. Am J Hum Genet 64:126–135
- Cormand B, Pihko H, Bayes M, Valanne L, Santavuori P, Talim B, Gershoni-Baruch R, Ahmad A, van Bokhoven H, Brunner HG, Voit T, Topaloglu H, Dobyns WB, Lehesjoki AE (2001) Clinical and genetic distinction between Walker-Warburg syndrome and muscle-eye-brain disease. Neurology 56:1059–1069
- Dobyns WB, Pagon RA, Curry CJR, Greenberg F (1990) Response to Santavuori et al. regarding Walker-Warburg syndrome and muscle-eye-brain disease. Am J Med Genet 36:373–374
- Haltia M, Leivo I, Somer H, Pihko H, Paetau A, Kivelä T, Tarkkanen A, Tomé F, Engvall E, Santavuori P (1997) Muscle-eyebrain disease: a neuropathological study. Ann Neurol 41:173– 180
- Kano H, Kobayashi K, Herrmann R, Tachikawa M, Manya H, Nishino I, Nonaka I, Straub V, Talim B, Voit T, Topaloglu H, Endo T, Youshikawa H, Toda T (2002) Deficiency of alphadystroglycan in muscle-eye-brain disease. Biochem Biophys Res Commun 291:1283–1286

- Michele DE, Barresi R, Kanagawa M, Saito F, Cohn RD, Satz JS, dollar J, Nishino I, Kelley RI, Somer H, Straub V, Mathews KD, Moore SA, Campbell KP (2002) Post-translational disruption of dystroglycan-ligand interactions in congenital musular dystrophies. Nature 418:417–422
- Moore SA, Saito F, Chen J, Michele DE, Henry MD, Messing A, Cohn RD, Ross-Barta SE, Westra S, Williamson RA, Hoshi T, Campbell KP (2002) Deletion of brain dystroglycan recapitulates aspects of congenital muscular dystrophy. Nature 418: 422–425
- Pihko H, Lappi M, Raitta C, Sainio K, Valanne L, Somer H, Santavuori P (1995) Ocular findings in muscle-eye-brain (MEB) disease: a follow-up study. Brain Dev 17:57–61
- Raitta C, Lamminen M, Santavuori P, Leisti J (1978) Ophthalmological findings in a new syndrome with muscle, eye and brain involvement. Acta Ophthalmol 56:465–472
- Ranta S, Pihko H, Santavuori P, Tahvanainen E, de la Chapelle A (1995) Muscle-eye-brain disease and Fukuyama type congenital muscular dystrophy are not allelic. Neuromusc Disord 5: 221–225
- Santavuori P, Leisti J, Kruus S (1977) Muscle, eye and brain disease: a new syndrome. Neuropädiatrie 8 (Suppl):553–558
- Santavuori P, Somer H, Sainio K, Rapola J, Kruus S, Nikitin T, Ketonen L, Leisti J (1989) Muscle-eye-brain disease (MEB). Brain Dev 11:147–153
- Santavuori P, Pihko H, Sainio K, Lappi M, Somer H, Haltia M, Raitta C, Ketonen L, Leisti J (1990) Muscle-eye-brain disease and Walker-Warburg syndrome. Am J Med Genet 36:371–372
- Santavuori P, Valanne L, Autti T, Haltia M, Pihko H, Sainio K (1998) Muscle-eye-brain disease; clinical features, visual evoked potentials and brain imaging in 20 patients. Eur J Paediatr Neurol 1:41–47
- Valanne L, Pihko H, Katevuo K, Karttunen P, Somer H, Santavuori P (1994) MRI of the brain in muscle-eye-brain (MEB) disease. Neuroradiology 36:473–476
- Yoshida A, Kobayashi K, Manya H, Taniguchi K, Kano H, Mizuno M, Inazu T, Mizuhashi H, Takahashi S, Takeuchi M, Herrman R, Straub V, Talim B, Voit T, Topaloglu H, Toda T, Endo T (2001) Muscular dystrophy and neuronal migration disorder caused by mutations in a glycosyltransferase, POMGnT1. Develop Cell 1:717–724

References for neuronal ceroid lipofuscinoses

- Åberg L, Järvelä I, Rapola J, Autti T, Kirveskari E, Lappi M, Sipilä L, Santavuori P (1998) Atypical juvenile neuronal ceroid lipofuscinosis with granular osmiophilic deposit -like inclusions in the autonomic nerve cells of the gut wall. Acta Neuropathol 95:306–312
- Das AK, Becerra CHR, Yi W, Lu JY, Siakotos AN, Wisniewski KE (1998) Molecular genetics of palmitoyl-protein thioesterase deficiency in the U.S. J Clin Invest 102:361–370
- Goebel HH, Mole SE, Lake BD (eds) (1999) The neuronal ceroid lipofuscinoses (Batten disease). IOS Press, Amsterdam
- Haltia M (2003) The neuronal ceroid-lipofuscinoses. J Neuropathol Exp Neurol 62:1–13
- Haltia M, Herva R, Suopanki J, Baumann M, Tyynelä J (2001) Hippocampal lesions in the neuronal ceroid lipofuscinoses. Eur J Paediatr Neurol 5 (Suppl A):209–211
- Heinonen O, Salonen T, Jalanko A, Peltonen L, Copp A (2000) CLN-1 and CLN-5, genes for infantile and variant late infantile neuronal ceroid lipofuscinoses, are expressed in the embryonic human brain. J Comp Neurol 426:406–412
- Mitchell WA, Wheeler RB, Sharp JD, Bate SL, Gardiner RM, Ranta US, Lonka L, Williams RE, Lehesjoki AE, Mole SE (2001) Turkish variant late infantile neuronal ceroid lipofuscinosis (CLN7) may be allelic to CLN8. Eur J Paediatr Neurol 5 (Suppl A):21–27
- Mitchison HM, Mole SE (eds) (2001) Recent advances in the neuronal ceroid lipofuscinoses. Eur J Paediatr Neurol 5 (Suppl A)

- Mitchison HM, Hofmann SL, Becerra CHR, Munroe P, Lake BD, Crow YJ, Stephenson JBP, Williams RE, Hofman IL, Taschner PEM, Martin JJ, Philippart M, Andermann E, Andermann F, Mole SE, Gardiner RM, O'Rawe AM (1998) Mutations in the palmitoyl-protein thioesterase gene (PPT; CLN1) causing juvenile neuronal ceroid lipofuscinosis with granular osmiophilic deposits. Hum Mol Genet 7:291–297
- Ranta S, Savukoski M, Santavuori P, Haltia M (2001) Studies of homogenous populations: CLN5 and CLN8. Adv Genet 45: 123–140
- Rapola J, Santavuori P, Savilahti E (1984) Suction biopsy of rectal mucosa in the diagnosis of infantile and juvenile types of neuronal ceroid-lipofuscinoses. Hum Pathol 15:352–360
- Santavuori P, Vanhanen SL, Autti T (2001) Clinical and neuroradiological diagnostic aspects of neuronal ceroid lipofuscinosis disorders. Eur J Paediatr Neurol 5 (Suppl A):157–161
- Van Diggelen OP, Thobois S, Tilikete C, Zabot MT, Keulemans JLM, van Bunderen PA, Taschner PEM, Losekoot M, Voznyi YV (2001) Adult neuronal ceroid lipofuscinosis with palmitoyl-protein thioesterase deficiency: first adult-onset patients of a childhood disease. Ann Neurol 50:269–272
- Vesa J, Chin MH, Oelgeschlager K, Isosomppi J, DellAngelica EC, Jalanko A, Peltonen L (2002) Neuronal ceroid lipofuscinoses are connected at molecular level: interaction of CLN5 protein with CLN2 and CLN3. Mol Biol Cell 13:2410–2420
- Wisniewski KE (2001) Pheno/genotypic correlations of neuronal ceroid lipofuscinoses. Neurology 57:576–581
- Wisniewski KE, Zhong N (eds) (2001) Batten disease. Adv Genet 45: (several articles on different NCL diseases)

References for nonketotic hyperglycinemia

- Choi CG, Lee HK, Yoon JH (2001) Localized proton MR spectroscopic detection of nonketotic hyperglycinemia in an infant. Korean J Radiol 2:239–242
- Gerritsen T, Kaveggia E, Waisman HA (1965) A new type of idiopathic hyperglycinemia with hypo-oxaluria. Pediatrics 36:882– 891
- Hamosh A, Johnston MV (2001) Nonketotic hyperglycinemia. In: Scriver CR, Beaudet AL, Valle D, Sly WS (eds) The metabolic & molecular bases of inherited disease, 8th edn. McGraw-Hill, New York, pp 2065–2078
- Huisman TA, Thiel T, Steinmann B, Zeilinger G, Martin E (2002) Proton magnetic resonance spectroscopy of the brain of a neonate with nonketotic hyperglycinemia: in vivo–in vitro (ex vivo) correlation. Eur Radiol 12:858–861
- Kure S, Takayanagi M, Narisawa K, Tada K, Leisti J (1992) Identification of a common mutation in Finnish patients with nonketotic hyperglycinemia. J Clin Invest 90:160–164
- Press GA, Barshop BA, Haas RH, Nyhan WL, Glass RF, Hesselink JR (1989) Abnormalities of the brain in nonketotic hyperglycinemia: MR manifestations. Am J Neuroradiol 10:315– 321
- Tada K, Kure S (1993) Non-ketotic hyperglycinaemia: molecular lesion, diagnosis and pathophysiology. J Inher Metab Dis 16: 691–703
- Visakorpi JK, Donner M, Norio R (1965) Hyperglycinuria with severe neurological manifestations. Ann Paediatr Fenn 11:114– 117
- von Wendt L, Similä S, Hirvasniemi A, Suvanto E (1978) Nonketotic hyperglycinaemia. A clinical analysis of 19 Finnish patients. Monogr Hum Genet 9:58–64
- von Wendt L, Similä S, Hirvasniemi A, Suvanto E (1978) Altered levels of various amino acids in blood plasma and cerebrospinal fluid of patients with nonketotic hyperglycinemia. Neuropädiatrie 9:360–368
- von Wendt L, Hirvasniemi A, Similä S (1979) Nonketotic hyperglycinemia. A genetic study of 13 Finnish families. Clin Genet 15:411–417

von Wendt L, Alanko H, Sorri M, Toivakka E, Saukkonen AL, Similä S (1981) Clinical and neurophysiological findings in heterozygotes for nonketotic hyperglycinemia. Clin Genet 19:94–100

References for northern epilepsy

- Herva R, Tyynelä J, Hirvasniemi A, Syrjäkallio-Ylitalo M, Haltia M (2000) Northern epilepsy: a novel form of neuronal ceroidlipofuscinosis. Brain Pathol 10:215–222 (also in Abstracts, 12. Scandinavian Congress of Neurology, June 10–13, 1998, Oulu, Finland)
- Hirvasniemi A, Leisti J (1991) An inherited form of childhood epilepsy associated with mental retardation. Am J Hum Genet 49 (Suppl, without number):147
- Hirvasniemi A, Lang H, Lehesjoki AE, Leisti J (1994) Northern epilepsy syndrome: an inherited childhood onset epilepsy with associated mental deterioration. J Med Genet 31:177–182
- Lauronen L, Santavuori P, Hirvasniemi A, Kirveskari E, Huttunen J, Autti T (2001) Northern epilepsy syndrome (NES, CLN8) – MRI and electrophysiological studies. Eur J Paediat Neurol 5:167–173
- Lonka L, Kyttälä A, Ranta S, Jalanko A, Lehesjoki AE (2000) The neuronal ceroid lipofuscinosis CLN8 membrane protein is a resident of the endoplasmic reticulum. Hum Mol Genet 9: 1691–1697
- Ranta S, Lehesjoki AE, de Fatima Bonaldo M, Knowles JA, Hirvasniemi A, Ross B, de Jong PJ, Bento Soares M, de la Chapelle A, Gilliam TC (1997) High-resolution mapping and transcript identification at the progressive epilepsy with mental retardation locus on chromosome 8p. Genome Res 7:887–896
- Ranta S, Zhang Y, Ross B, Lonka L, Takkunen E, Messer A, Sharp J, Wheeler R, Kusumi K, Mole S, Liu W, Bento Soares M, de Fatima Bonaldo M, Hirvasniemi A, de la Chapelle A, Gilliam TC, Lehesjoki AE (1999) The neuronal ceroid lipofuscinoses in human EPMR and mnd mutant mice are associated with mutations in CLN8. Nat Genet 23:233–236
- Tahvanainen E, Ranta S, Hirvasniemi A, Karila E, Leisti J, Sistonen P, Weissenbach J, Lehesjoki AE, de la Chapelle A (1994) The gene for a recessively inherited human childhood progressive epilepsy with mental retardation maps to the distal short arm of chromosome 8. Proc Natl Acad Sci USA 91:7267–7270

References for PEHO syndrome

- Chitty LS, Robb S, Berry C, Silver D, Baraitser M (1996) PEHO or PEHO-like syndrome? Clin Dysmorphol 5:143–152
- Fujimoto S, Yokochi K, Nakano M, Wada Y (1995) Progressive encephalopathy with edema, hypsarrhythmia, and optic atrophy (PEHO syndrome) in two Japanese siblings. Neuropediatrics 26:270–272
- Haltia M, Somer M (1993) Infantile cerebello-optic atrophy; neuropathology of the progressive encephalopathy syndrome with edema, hypsarrhythmia and optic atrophy (the PEHO syndrome). Acta Neuropathol 85:241–247
- Salonen R, Somer M, Haltia M, Lorenz M, Norio R (1991) Progressive encephalopathy with edema, hypsarrhythmia, and optic atrophy (PEHO syndrome). Clin Genet 39:287–293
- Somer M (1993) Diagnostic criteria and genetics of the PEHO syndrome. J Med Genet 30:932–936
- Somer M, Salonen O, Pihko H, Norio R (1993) PEHO syndrome (progressive encephalopathy with edema, hypsarrhythmia, and optic atrophy): neuroradiological findings. Am J Neuroradiol 14:861–867
- Somer M, Setälä K, Kivelä T, Haltia M, Norio R (1993) The PEHO syndrome (progressive encephalopathy with oedema, hypsarrhythmia and optic atrophy); ophthalmological findings and differential diagnosis. Neuro-ophthalmology 13:65–72

Vanhatalo S, Somer M, Barth PG (2002) Dutch patients with progressive encephalopathy with edema, hypsarrhythmia, and optic atrophy (PEHO) syndrome. Neuropediatrics 33:100–104

References for PLO-SL

- Hakola HPA (1972) Neuropsychiatric and genetic aspects of a new hereditary disease characterized by progressive dementia and lipomembranous polycystic osteodysplasia. Acta Psychiatr Scand Suppl 232
- Hakola HPA (1990) Polycystic lipomembranous osteodysplasia with sclerosing encephalopathy (membranous lipodystrophy); a neuropsychiatric follow-up study with an appendix by PEJ Virtama, MT Hakola and HPA Hakola: Bone radiography of PLO-SL cases. Monographs of Psychiatria Fennica 17
- Järvi OH, Lauttamus LL, Solonen KA (1964) Membranous reticulin dysplasia of bones. Probably a new disease entity. (Only title). In: Proceedings of the 14th Scandinavian congress of pathology and microbiology. Universitetsforlaget, Oslo, p 51
- Järvi OH, Hakola HPA, Lauttamus LL, Solonen KA, Vilppula AH (1968) Cystic capillary-necrotic osteodysplasia, a systemic bone disease probably caused by arteriolar and capillary necroses. Relation to brain affections. Abstracts, Seventh International Congress of International Academy of Pathology, Milan 1968, pp 291–292
- Kalimo H, Sourander P, Järvi O, Hakola P (1994) Vascular changes and blood-brain barrier damage in the pathogenesis of polycystic lipomembranous osteodysplasia with sclerosing leukoencephalopathy (membranous lipodystrophy). Acta Neurol Scand 89:353–361
- Nasu T, Tsukahara Y, Terayama K (1973) A lipid metabolic disease – membranous lipodystrophy – an autopsy case demonstrating numerous peculiar membrane-structures composed of compound lipid in bone and bone marrow and various adipose tissues. Acta Pathol Jap 23:539–558
- Nylander PO, Drugge U, Holmgren G, Adolfsson R (1996) Polycystic lipomembranous osteodysplasia with sclerosing leukoencephalopathy (PLO-SL): a genealogic study of Swedish families of probable Finnish background. Clin Genet 50:353–357
- Paloneva J, Kestilä M, Wu J, Salminen A, Böhling T, Ruotsalainen V, Hakola P, Bakker ABH, Phillips JH, Pekkarinen P, Lanier LL, Timonen T, Peltonen L (2000) Loss-of-function mutations in TYROBP (DAP12) result in a presenile dementia with bone cysts. Nat Genet 25:357–361
- Paloneva J, Autti T, Raininko R, Partanen J, Salonen O, Puranen M, Hakola P, Haltia M (2001) CNS manifestations of Nasu-Hakola disease: a frontal dementia with bone cysts. Neurology 56:1552–1558
- Paloneva J, Manninen T, Christman G, Hovanes K, Mandelin J, Adolfsson R, Bianchin M, Bird T, Miranda R, Salmaggi A, Tranebjaerg L, Konttinen Y, Peltonen L (2002) Mutations in two genes encoding different subunits of a receptor signaling complex result in an identical disease phenotype. Am J Hum Genet 71:656–662
- Pekkarinen P, Hovatta I, Hakola P, Järvi O, Kestilä M, Lenkkeri U, Adolfsson R, Holmgren G, Nylander PO, Tranebjaerg L, Terwilliger JD, Lönnqvist J, Peltonen L (1998) Assignment of the locus for PLO-SL, a frontal lobe dementia with bone cysts, to 19q13. Am J Hum Genet 62:362–372
- Pekkarinen P, Kestilä M, Paloneva J, Terwilliger J, Varilo T, Järvi O, Hakola P, Peltonen L (1998) Fine-scale mapping of a novel dementia gene, PLO-SL, by linkage disequilibrium. Genomics 54:307–315
- Terayama K (1961) Two cases of cystic bone disease showing peculiar features. Nippon Seikeigeka Gakkai Zasshi 35:626 (in Japanese)
- Verloes A, Maquet P, Sadzot B, Vivario M, Thiry A, Franck G (1997) Syndrome of the month: Nasu-Hakola syndrome: polycystic lipomembranous osteodysplasia with sclerosing leukoencephalopathy and presenile dementia. J Med Genet 34:753– 757

References for progressive myoclonus epilepsy, Unverricht-Lundborg type

- Berkovic SF, So NK, Andermann F (1991) Progressive myoclonus epilepsies: clinical and neurophysiological diagnosis. J Clin Neurophysiol 8:261–274
- Fedi M, Reutens D, Dubeau F, Andermann E, D'Agostino D, Andermann F (2001) Long-term efficacy and safety of piracetam in the treatment of progressive myoclonus epilepsy. Arch Neurol 58:781–786
- Forss N, Silen T, Karjalainen T (2001) Lack of activation of human secondary somatosensory cortex in Unverricht-Lundborg type of progressive myoclonus epilepsy. Ann Neurol 49:90–97
- Haltia M, Kristensson K, Sourander P (1969) Neuropathological studies in three Scandinavian cases of progressive myoclonus epilepsy. Acta Neurol Scand 45:63–77
- Harenko A, Toivakka E (1961) Myoclonus epilepsy (Unverricht-Lundborg) in Finland. Acta Neurol Scand 37:282–296
- Koskiniemi M, Donner M, Majuri H, Haltia M, Norio R (1974) Progressive myoclonus epilepsy. A clinical and histopathological study. Acta Neurol Scand 50:307–332
- Lafrenière RG, Rochefort DL, Chrétien N, Rommens JM, Cochius JI, Kälviäinen R, Nousiainen U, Patry G, Farrell K, Söderfeldt B, Federico A, Hale BR, Hernandez Cossio O, Sörensen T, Pouliot MA, Kmiec T, Uldall P, Janszky J, Pranzatelli MR, Andermann F, Andermann E, Rouleau GA (1997) Unstable insertion in the 5' flanking region of the cystatin B gene is the most common mutation in progressive myoclonus epilepsy type 1, EPM1. Nat Genet 15:298–302
- Lalioti MD, Scott HS, Buresi C, Rossier C, Bottani A, Morris MA, Malafosse A, Antonarakis SE (1997) Dodecamer repeat expansion in cystatin B gene in progressive myoclonus epilepsy. Nature 386:847–851
- Lehesjoki AE, Koskiniemi M (1998) Clinical features and genetics of progressive myoclonus epilepsy of the Unverricht-Lundborg type. Ann Med 30:474–480
- Lehesjoki AE, Koskiniemi M, Sistonen P, Miao J, Hästbacka J, Norio R, de la Chapelle A (1991) Localization of a gene for progressive myoclonus epilepsy to chromosome 21q22. Proc Natl Acad Sci USA 88:3696–3699
- Lehesjoki AE, Koskiniemi M, Pandolfo M, Antonelli A, Kyllerman M, Wahlström J, Nergårdh A, Burmeister M, Sistonen P, Norio R, de la Chapelle A (1992) Linkage studies in progressive myoclonus epilepsy: Unverricht-Lundborg and Lafora's diseases. Neurology 42:1545–1550
- Lundborg H (1901) Über Degeneration und degenerierte Geschlechter in Schweden. I. Klinische Studien und Erfahrungen hinsichtlich der familiären Myoklonie und damit verwandter Krankheiten. Isaac Marcus' Boktr.-Aktiebolag, Stockholm
- Lundborg H (1903) Die progressive Myoklonus-Épilepsie (Unverrichts Myoklonie). Almqvist & Wiksell, Uppsala
- Malafosse A, Lehesjoki AE, Genton P, Labauge P, Durand G, Tassinari CA, Dravet C, Michelucci R, de la Chapelle A (1992) Identical genetic locus for Baltic and Mediterranean myoclonus. Lancet 339:1080–1081
- Marseille Consensus Group (1990) Classification of progessive myoclonus epilepsies and related disorders. Ann Neurol 28: 113–116
- Moulard B, Genton P, Grid D, Jeanpierre M, Ouazzani R, Mrabet A, Morris M, LeGuern E, Dravet C, Mauguiere F, Utermann B, Baldy-Moulinier M, Belaidi H, Bertran F, Biraben A, Ali Cherif A, Chkili T, Crespel A, Darcel F, Dulac O, Geny C, Humbert-Claude V, Kassiotis P, Buresi C, Malafosse A (2002) Haplotype study of West European and North African Unverricht-Lundborg chromosomes: evidence for a few founder mutations. Hum Genet 111:255–262
- Norio R, Koskiniemi M (1979) Progressive myoclonus epilepsy; genetic and nosological aspects with a special reference to 107 Finnish patients. Clin Genet 15:382–398

- Pennacchio LA, Lehesjoki AE, Stone NE, Willour VL, Virtaneva K, Miao J, D'Amato E, Ramirez L, Faham M, Koskiniemi M, Warrington JA, Norio R, de la Chapelle A, Cox DR, Myers RM (1996) Mutations in the gene encoding cystatin B in progressive myoclonmus epilepsy (EPM1). Science 271:1731–1734
- Pennacchio LA, Bouley DM, Higgins KM, Scott MP, Noebels JL, Myers RM (1998) Progressive ataxia, myoclonus epilepsy and cerebellar apoptosis in cystatin B-deficient mice. Nat Genet 20:251–258
- Unverricht H (1891) Die Myoclonie. Deuticke, Leipzig und Wien Unverricht H (1895) Über familiäre Myoclonie. Dtsch Z Nervenheilkd 7:32–67
- Uthman BM, Reichl A (2002) Progressive myoclonus epilepsies. Curr Treat Options Neurol 4:3–17 (data on zonisamide and piracetam medication)
- Virtaneva K, D'Amato E, Miao J, Koskiniemi M, Norio R, Avanzini G, Franchescetti S, Michelucci R, Tassinari CA, Omer S, Pennacchio LA, Myers RM, Dieguez-Lucena JL, Krahe R, de la Chapelle A, Lehesjoki AE (1997) Unstable minisatellite expansion causing recessively inherited myoclonus epilepsy, EPM1. Nat Genet 15:393–396

References for Rapadilino syndrome

- Jam K, Fox M, Crandall BF (1999) RAPADILINO syndrome: a multiple malformation syndrome with radial and patellar aplasia. Teratology 60:37–38
- Kant SG, Baraitser M, Milla PJ, Winter (1998) Rapadilino syndrome–a non-Finnish case. Clin Dysmorphol 7:135–138
- Kääriäinen H, Ryöppy S, Norio R (1989) RAPADILINO syndrome with radial and patellar aplasia/hypoplasia as main manifestations. Am J Med Genet 33:346–351
- Regla Vargas F, Cabral de Almeida JC, Clinton Llerena JJr, Fagundes Reis D (1992) RAPADILINO syndrome. Am J Med Genet 44:716–719

References for retinoschisis

- Alitalo T, Kruse TA, de la Chapelle A (1991) Refined localization of the gene causing X-linked juvenile retinoschisis. Genomics 9:505–510
- de la Chapelle A, Alitalo T, Forsius H (1994) X-linked juvenile retinoschisis. In: Wright AW, Jay B (eds) Molecular genetics of inherited eye disorders (Modern genetics vol.2). Harwood Academic Publications, Great Britain, pp 339–357
- Forsius H, Vainio-Mattila B, Eriksson A (1962) X-linked hereditary retinoschisis. Brit J Ophthalmol 46:678–681
- Forsius H, Krause U, Helve J, Vuopala V, Mustonen E, Vainio-Mattila B, Fellman J, Eriksson AW (1973) Visual acuity in 183 cases of X-chromosomal retinoschisis. Can J Ophthalmol 8: 385–393
- Forsius HR, Eriksson AW, Damsten M (1990) Progression in juvenile X-chromosomal retinoschisis. Acta Ophthalmol 68 Suppl 195: 113–119
- Grayson C, Reid SN, Ellis JA, Rutherford A, Sowden JC, Yates JR, Farber DB, Trump D (2000) Retinoschisin, the X-linked retinoschisis protein, is a secreted photoreceptor protein, and is expressed and released by Weri-Rb1 cells. Hum Mol Genet 9: 1873–1879
- Haas J (1898) Über das Zusammenvorkommen von Veränderungen der Retina und Chorioidea. Arch Augenheilkd 37:343–348
- Huopaniemi L, Rantala A, Tahvanainen E, de la Chapelle A, Alitalo T (1997) Linkage disequilibrium and physical mapping of X-linked juvenile retinoschisis. Am J Hum Genet 60:1139– 1149
- Huopaniemi L, Rantala A, Forsius H, Somer M, de la Chapelle A, Alitalo T (1999) Three widespread founder mutations contribute to high incidence of X-linked juvenile retinoschisis in Finland. Eur J Hum Genet 7:368–376

- Molday LL, Hicks D, Sauer CG, Weber BH, Molday RS (2001) Expression of X-linked retinoschisis protein RS1 in photoreceptor and bipolar cells. Invest Ophthalmol Vis Sci 42:816– 825
- The Retinoschisis Consortium (1998) Functional implications of the spectrum of mutations found in 234 cases with X-linked juvenile retinoschisis (XLRS). Hum Mol Genet 7:1185–1192
- Sauer CG, Gehrig A, Warneke-Wittstock R, Marquardt A, Ewing CC, Gibson A, Lorenz B, Jurklies B, Weber BHF (1997) Positional cloning of the gene associated with X-linked juvenile retinoschisis. Nat Genet 17:164–170
- Vainio-Mattila B, Eriksson AW, Forsius H (1969) X-chromosomal recessive retinoschisis in the region of Pori; an ophthalmo-genetical analysis of 103 cases. Acta Ophthalmol 47:1135–1148

References for Salla disease

- Aula N, Salomäki P, Timonen R, Verheijen F, Mancini G, Mansson JE, Aula P, Peltonen L (2000) The spectrum of SLC17A5gene mutations resulting in free sialic acid storage diseases indicates some genotype-phenotype correlation. Am J Hum Genet 67:832–840
- Aula P, Gahl WA (2001) Disorders of free sialic acid storage. In: Scriver CR, Beaudet AL, Valle D, Sly WS (eds) The metabolic & molecular bases of inherited disease, 8th edn. McGraw-Hill, New York, pp 5109–5120
- Aula P, Autio S, Raivio K, Rapola J, Thodén CJ, Koskela SL, Yamashina I (1979) "Salla disease". A new lysosomal storage disorder. Arch Neurol 36:88–94
- Haataja L, Parkkola R, Sonninen P, Vanhanen SL, Schleutker J, Äärimaa T, Turpeinen V, Renlund M, Aula P (1994) Phenotypic variation and magnetic resonance imaging (MRI) in Salla disease, a free sialic acid storage disorder. Neuropediatrics 25:1–7
- Haataja L, Schleutker J, Laine AP, Renlund M, Savontaus ML, Dib C, Weissenbach J, Peltonen L, Aula P (1994) The genetic locus for free sialic acid storage disease maps to the long arm of chromosome 6. Am J Hum Genet 54:1042–1049
- Havelaar AC, Beerens CEMT, Mancini GMS, Verheijen FW (1999) Transport of organic anions by the lysosomal sialic acid transporter: a functional approach towards the gene for sialic acid storage disease. FEBS Lett 446:65–68
- Leppänen P, Isosomppi J, Schleutker J, Aula P, Peltonen L (1996) A physical map of the 6q14-q15 region harboring the locus for the lysosomal membrane sialic acid transport defect. Genomics 37:62–67
- Renlund M, Aula P, Raivio K, Autio S, Sainio K, Rapola J, Koskela SL (1983) Salla disease: a new lysosomal storage disorder with disturbed sialic acid metabolism. Neurology 33:57–66
- Renlund M, Kovanen PT, Raivio KO, Aula P, Gahmberg CG, Ehnholm C (1986) Studies on the defect underlying the lysosomal storage of sialic acid in Salla disease. J Clin Invest 77: 568–574
- Salomäki P, Aula N, Juvonen V, Renlund M, Aula P (2001) Prenatal detection of free sialic acid storage disease: genetic and biochemical studies in nine families. Prenat Diagn 21:354–358
- Schleutker J, Laine AP, Haataja L, Renlund M, Weissenbach J, Aula P, Peltonen L (1995) Linkage disequilibrium utilized to establish a refined genetic position of the Salla disease locus on 6q14-q15. Genomics 27:286–292
- Schleutker J, Leppänen P, Månsson JE, Erikson A, Weissenbach J, Peltonen L, Aula P (1995) Lysosomal free sialic acid storage disorders with different phenotypic presentations – infantileform sialic acid storage disease and Salla disease – represent allelic disorders on 6q14–15. Am J Hum Genet 57:893–901
- Varho T, Jääskeläinen S, Tolonen U, Sonninen P, Vainionpää L, Aula P, Sillanpää M (2000) Central and peripheral nervous system dysfunction in the clinical variation of Salla disease. Neurology 55:99–104

- Varho TT, Alajoki LE, Posti KM, Korhonen TT, Renlund MG, Nyman SRG, Sillanpää ML, Aula PP (2002) Phenotypic spectrum of Salla disease, a free sialic acid storage disorder. Pediat Neurol 26:267–273
- Verheijen FW, Verbeek E, Aula N, Beerens CEMT, Havelaar AC, Joosse M, Peltonen L, Aula P, Galjaard H, van der Spek PJ, Mancini GMS (1999) A new gene, encoding an anion transporter, is mutated in sialic acid storage diseases. Nat Genet 23:462–465

References for selective malabsorption of vitamin B₁₂

- Aminoff M, Tahvanainen E, Gräsbeck R, Weissenbach J, Broch H, de la Chapelle A (1995) Selective intestinal malabsorption of vitamin B12 displays recessive Mendelian inheritance: assignment of a locus to chromosome 10 by linkage. Am J Hum Genet 57:824–831
- Aminoff M, Carter JE, Chadwick RB, Johnson C, Gräsbeck R, Abdelaal MA, Broch H, Jenner LB, Verroust PJ, Moestrup SK, de la Chapelle A, Krahe R (1999) Mutations in CUBN, encoding the intrinsic factor – vitamin B12 receptor, cubilin, cause hereditary megaloblastic anaemia 1. Nat Genet 21:309–313
- Dugue B, Aminoff M, Aimone-Gastin I, Leppänen E, Gräsbeck R, Gueant JL (1998) A urinary radioisotope-binding assay to diagnose Gräsbeck-Imerslund disease. J Pediatr Gastroenterol Nutr 26:21–25
- Furuhjelm U, Nevanlinna HR (1973) Inheritance of selective malabsorption of vitamin B₁₂. Scand J Haematol 11:27–34
- Fyfe JC, Giger U, Hall CA, Jezyk PF, Klompp SA, Levine JS, Patterson DF (1991) Inherited selective intestinal cobalamin malabsorption and cobalamin deficiency in dogs. Pediat Res 29: 24–31
- Gräsbeck R, Gordin R, Kantero I, Kuhlbäck B (1960) Selective vitamin B₁₂ malabsorption and proteinuria in young people: a syndrome. Acta Med Scand 167:289–296
- Guéant JL, Saunier M, Gastin I, Safi A, Lamireau T, Duclos B Bigard MA, Gräsbeck R (1995) Decreased activity of intestinal and urinary intrinsic factor in Gräsbeck-Imerslund disease. Gastroenterology 108:1622–1628
- Imerslund O (1959, 1960) Idiopathic chronic megaloblastic anemia in children. Oslo University Press, Oslo 1959 and Acta Paediat 1960:49:Suppl 119; Summary of supplement = Acta Paediat 1960:49:208–209
- Kozyraki R, Kristiansen M, Silahtaroglu A, Hansen C, Jacobsen C, Tommerup N, Verroust PJ, Moestrup SK (1998) The human intrinsic factor – vitamin B12 receptor, cubilin: molecular characterization and chromosomal mapping of the gene to 10p within the autosomal recessive megaloblastic anemia (MGA1) region. Blood 91:3593–3600
- Kristiansen M, Aminoff M, Jacobsen C, de la Chapelle A, Krahe R, Verroust PJ, Moestrup SK (2001) Cubilin P1297L mutation associated with hereditary megaloblastic anemia 1 causes impaired recognition of intrinsic factor – vitamin B(12) by cubilin. Blood 97:3316–3317
- Visakorpi JK, Furuhjelm U (1968) Selective malabsorption of vitamin B₁₂. Mod Probl Pediatr 11:150–160

References for Spielmeyer-Sjögren disease

- Åberg LE, Bäckman M, Kirveskari E, Santavuori P (2000) Epilepsy and antiepileptic drug therapy in juvenile neuronal ceroid lipofuscinosis. Epilepsia 41:1296–1302
- Autti T, Raininko R, Vanhanen SL, Santavuori P (1996) Cranial MRI of 30 patients with juvenile neuronal ceroid lipofuscinosis. Neuroradiology 38:476–482
- von Bagh K, Hortling H (1948) Blodfynd vid juvenil amaurotisk idioti. Finska Läkaresällsk Handl 38:1072–1076

- Callen DF, Baker E, Lane S, Nancarrow J, Thompson A, Whitmore SA, MacLennan DH, Berger R, Cherif D, Järvelä I, Peltonen L, Sutherland GR, Gardiner RM (1991) Regional mapping of the Batten disease locus (CLN3) to human chromosome 16p12. Am J Hum Genet 49:1372–1377
- Gardiner RM, Sandford A, Deadman M, Poulton J, Reeders S, Jokiaho I, Peltonen L, Julier C (1990) Batten disease (Spielmeyer-Vogt; juvenile onset neuronal ceroid lipofuscinosis) maps to human chromosome 16. Genomics 8:387–390
- Järvelä I, Mitchison HM, Munroe PB, O'Rawe AM, Mole SE, Syvänen AC (1996) Rapid diagnostic test for the major mutation underlying Batten disease. J Med Genet 33:1041–1042
- Järvelä I, Sainio M, Rantamäki T, Olkkonen VM, Carpén O, Peltonen L, Jalanko A (1998) Biosynthesis and intracellular targeting of the CLN3 protein defective in Batten disease. Hum Mol Genet 7:85–90
- Järvelä I, Lehtovirta M, Tikkanen R, Kyttälä A, Jalanko A (1999) Defective intracellular transport of CLN3 is the molecular basis of Batten disease (JNCL). Hum Mol Genet 8:1091–1098
- Lamminranta S, Åberg LE, Autti T, Moren R, Laine T, Kaukoranta J, Santavuori P (2001) Neuropsychological test battery in the follow-up of patients with juvenile neuronal ceroid lipofuscinosis. J Intellect Disabil Res 45:8–17
- Launes J, Heiskala H, Nikkinen P, Santavuori P (1996) Brain perfusion SPECT in juvenile neuronal ceroid lipofuscinosis. Neuropediatrics 27:84–87
- Lauronen L, Heikkilä E, Autti T, Sainio K, Huttunen J, Aronen HJ, Korvenoja A, Ilmoniemi RJ, Santavuori P (1997) Somatosensory evoked magnetic fields from primary sensorimotor cortex in juvenile neuronal ceroid lipofuscinosis. J Child Neurol 12: 355–360
- Lauronen L, Munroe PB, Järvelä I, Autti T, Mitchison HM, O'Rawe AM, Gardiner RM, Mole SE, Puranen J, Häkkinen AM, Kirveskari E, Santavuori P (1999) Delayed classic and protracted phenotypes of compound heterozygous juvenile neuronal ceroid lipofuscinosis. Neurology 52:360–365
- Luiro K, Kopra O, Lehtovirta M, Jalanko A (2001) CLN3 protein is targeted to neuronal synapses but excluded from synaptic vesicles: new clues to Batten disease. Hum Mol Genet 10: 2123–2131
- Munroe PB, Rapola J, Mitchison HM, Mustonen A, Mole SE, Gardiner RM, Järvelä I (1996) Prenatal diagnosis of Batten's disease. Lancet 347:1014–1015
- Munroe PB, Mitchison HM, O'Rawe AM, Anderson JW, Boustany RM, Lerner TJ, Taschner PEM, de Vos N, Breuning MH, Gardiner RM, Mole SE (1997) Spectrum of mutations in the Batten disease gene. Am J Hum Genet 61:310–316
- The International Batten Disease Consortium (1995) Isolation of a novel gene underlying Batten disease, CLN3. Cell 82:949–957
- Stengel S (1826) Account of a singular illness among four siblings in the vicinity of Roraas. Eyr (Christiania) 1:347-352
- Vesa J, Peltonen L (2002) Mutated genes in juvenile and variant late infantile neuronal ceroid lipofuscinoses encode lysosomal proteins. Curr Mol Med 2:439–444

References for tibial muscular dystrophy

- Hackman P, Vihola A, Haravuori H, Marchand S, Sarparanta J, De Seze J, Labeit S, Witt C, Peltonen L, Richard I, Udd B (2002) Tibial muscular dystrophy is a titinopathy caused by mutations in *TTN*, the gene encoding the giant skeletal-muscle protein titin. Am J Hum Genet 71:492–500
- Haravuori H, Mäkelä-Bengs P, Udd B, Partanen J, Pulkkinen L, Somer H, Peltonen L (1998) Assignment of the tibial muscular dystrophy locus to chromosome 2q31. Am J Hum Genet 62:620–626
- Haravuori H, Vihola A, Straub V, Auranen M, Richard I, Marchand S, Voit T, Labeit S, Somer H, Peltonen L, Beckmann JS, Udd B (2001) Secondary calpain3 deficiency in 2q-linked muscular dystrophy: titin is the candidate gene. Neurology 56:869– 877

- Udd B (1992) Limb-girdle type muscular dystrophy in a large family with distal myopathy: homozygous manifestation of a dominant gene? J Med Genet 29:383–389
- Udd B, Kääriäinen H, Somer H (1991) Muscular dystrophy with separate clinical phenotypes in a large family. Muscle & Nerve 14:1050–1058
- Udd B, Partanen J, Halonen P, Falck B, Hakamies L, Heikkilä H, Ingo S, Kalimo H, Kääriäinen H, Laulumaa V, Paljärvi L, Rapola J, Reunanen M, Sonninen V, Somer H (1993) Tibial muscular dystrophy–late adult onset distal myopathy in 66 Finnish patients. Arch Neurol 50:604–608

References for Usher syndrome type 3

- Adato A, Vreugde S, Joensuu T, Avidan N, Hämäläinen R, Belenkiy O, Olender T, Bonne-Tamir B, Ben-Asher E, Espinos C, Millan JM, Lehesjoki AE, Flannery JG, Avraham KB, Pietrokowski S, Sankila EM, Beckmann JS, Lancet D (2002) Ush3A transcripts encode clarin-1, a four-transmembrane-domain protein with a possible role in sensory synapses. Eur J Hum Genet 10:339–350
- Fields RR, Zhou G, Huang D, Davis JR, Moller C, Jacobson SG, Kimberling WJ, Sumegi J (2002) Usher syndrome type III: revised genomic structure of the USH3 gene and identification of novel mutations. Am J Hum Genet 71:607–617
- Gorlin RJ, Tilsner TJ, Feinstein S, Duvall AJ (1979) Usher's syndrome type III. Arch Otolaryngol 105:353–354
- von Graefe A (1858) Exceptionelles Verhalten des Gesichtsfeldes bei Pigmententartung der Netzhaut. Albrecht von Graefes Arch Ophthalmol 4:250–253
- Hallgren B (1959) Retinitis pigmentosa combined with congenital deafness; with vestibulo-cerebellar ataxia and mental abnormality in a proportion of cases. A clinical and genetico-statistical study. Acta Psychiatr Neurol Scand 34: Suppl 138
- Joensuu T, Blanco G, Pakarinen L, Sistonen P, Kääriäinen H, Brown S, de la Chapelle A, Sankila EM (1996) Refined mapping of the Usher syndrome type III locus on chromosome 3, exclusion of candidate genes, and identification of the putative mouse homologous region. Genomics 38:255–263
- Joensuu T, Hämäläinen R, Lehesjoki AE, de la Chapelle A, Sankila EM (2000) A sequence-ready map of the Usher syndrome type III critical region on chromosome 3q. Genomics 63:409–416
- Joensuu T, Hämäläinen R, Yuan B, Johnson C, Tegelberg S, Gasparini P, Zelante L, Pirvola U, Pakarinen L, Lehesjoki AE, de la Chapelle A, Sankila EM (2001) Mutations in a novel gene with transmembrane domains underlie Usher syndrome type 3. Am J Hum Genet 69:673–684
- Karjalainen S, Pakarinen L, Teräsvirta M, Kääriäinen H, Vartiainen E (1989) Progressive hearing loss in Usher's syndome. Ann Otol Rhinol Laryngol 98:863–866
- Lindenov H (1945) The aetiology of deaf-mutism with special reference to heredity. Thesis, Copenhagen, Opera ex Domo Univ. Hafniensis No 8
- Nuutila A (1968) Neuropsychiatric and genetic aspects of the dystrophia retinae pigmentosa-dysacusis syndrome. Thesis, University of Helsinki
- Nuutila A (1970) Dystrophia retinae pigmentosa-dysacusis syndrome (DRD): a study of the Usher- or Hallgren syndrome. J Génét Hum 18:57–88
- Pakarinen L, Sankila EM, Tuppurainen K, Karjalainen S, Kääriäinen H (1995) Usher syndrome type III (USH3): the clinical manifestations in 42 patients. Scand J Log Phon 20:141–150
- Pakarinen L, Tuppurainen K, Laippala P, Mäntyjärvi M, Puhakka H (1996) The ophthalmological course of Usher syndrome type III. Int Ophthalmol 19:307–311
- Sankila EM, Pakarinen L, Kääriäinen H, Aittomäki K, Karjalainen S, Sistonen P, de la Chapelle A (1995) Assignment of an Usher syndrome type III (USH3) gene to chromosome 3q. Hum Mol Genet 4:93–98

Smith RJH, Berlin CI, Hejtmancik JF, Keats BJB, Kimberling WJ, Lewis RA, Möller CG, Pelias MSZ, Tranebjaerg L (1994) Clinical diagnosis of the Usher syndromes. Am J Med Genet 50:32–38

Usher CH (1913–1914) On the inheritance of retinitis pigmentosa with notes of cases. Royal London Ophthalmol Hosp Rep 19: 130–236

References for Vuopala disease

- Vuopala K, Herva R, Leisti J (1994) Lethal arthrogryposis in Finland – a clinicopathological study of 83 cases during thirteen years. Neuropediatrics 25:308–315
- Vuopala K, Ignatius J, Herva R (1995) Lethal arthrogryposis with anterior horn cell disease. Human Pathol 26:12–19

References for new candidates for the Finnish Disease Heritage

Ala-Mello S, Koskimies O, Rapola J, Kääriäinen H (1999) Nephronophthisis in Finland: epidemiology and comparison of genetically classified subgroups. Eur J Hum Genet 7:205–211

- Koskela S, Javela K, Jouppila J, Juvonen E, Nyblom O, Partanen J, Kekomäki R (1999) Variant Bernard-Soulier syndrome due to homozygous Asn45Ser mutation in the platelet glycoprotein (GP) IX in eleven patients of five unrelated Finnish families. Eur J Haematol 62:256–264
- Pirinen S, Kentala A, Nieminen P, Varilo T, Thesleff I, Arte S (2001) Recessively inherited low incisor hypodontia. J Med Genet 38:551–556
- St-Louis M, Leclerc B, Laine J, Salo MK, Holmberg C, Tanguay RM (1994) Identification of a stop mutation in five Finnish patients suffering from hereditary tyrosinemia type I. Hum Mol Genet 3:69–72
- Tyni T, Pihko H (1999) Long-chain 3-hydroxyacyl-CoA dehydrogenase deficiency. Acta Paediatr 88:237–245
- Tyni T, Palotie A, Viinikka L, Valanne L, Salo M, Von Döbeln U, Jackson S, Wanders R, Venizelos N, Pihko H (1997) Longchain 3-hydroxyacyl-coenzyme A dehydrogenase deficiency with the G1528C mutation: clinical presentation of thirteen patients. J Pediatr 130:67–76
- Visakorpi JK (1972) Aminohappojen aineenvaihduntahäiriöt (Inborn errors of amino acid metabolism, English summary). Duodecim 88:72–85