ORIGINAL PAPER



The potential effects of silver and gold nanoparticles as molluscicides and cercaricides on *Schistosoma mansoni*

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Received: 1 May 2018 / Accepted: 21 September 2018 / Published online: 3 October 2018 © Springer-Verlag GmbH Germany, part of Springer Nature 2018

Abstract

Schistosomiasis seriously affects human health in tropical regions. Its prevention is more important than treatment, raising the need for effective control methods. Recently, the role of nanomaterials in medical science has been growing. The present study aimed to evaluate the potential effects of silver (Ag) and gold (Au) nanoparticles (NPs) on Biomphalaria alexandrina snails and Schistosoma mansoni cercariae in vitro and to assess their effects on the infectivity of cercariae in vivo. The in vitro study proved that Ag and Au NPs were effective in killing B. alexandrina snails, with 30 µg/ml Ag and 160 µg/ml Au causing 100% mortality. The LC₅₀ of 9.68 µg/ml for Ag NPs and 133.7 µg/ml for Au NPs prevented snail infection with S. mansoni miracidia. Furthermore, Ag NPs at 50 µg/ml and Au NPs at 100 µg/ml increased the mortality of S. mansoni cercariae in a dose- and time-dependent manner, reaching 100% mortality after 1 h. The in vivo study found that Ag NPs prevented the occurrence of infection when cercariae were treated before the infection by either the tail immersion (TI) or subcutaneous (SC) route, as proven by parasitological parameters and by the absence of granuloma formation in hepatic tissue. Meanwhile, infection of mice by untreated cercariae followed by treatment with NPs 1 h post-infection (PI) caused a decrease in egg count/g intestine and egg count/g liver in the TI-infected group only. The oogram patterns and granuloma formation results were similar between infection control and the SC-infected group. On the other hand, Au NPs led to a decrease in total worm burden (TWB) in all tested groups, with a decrease in egg count/g intestine and egg count/g liver in TI-infected groups with either pre-treated or post-treated cercariae, in contrast to SC-infected groups. However, the oogram patterns and granuloma formation showed similar results to infection control. Ag and Au NPs have potential as molluscicides and cercaricides in vitro and can prevent or modulate the infectivity of cercariae in vivo.

Keywords Schistosoma mansoni · Biomphalaria alexandrina · Silver- gold- nano · Miracidium · Cercaria · In vivo · In vitro

Introduction

Schistosomiasis is the second most prevalent parasitic disease in the world after malaria. It affects more than 260 million people and leads to 200,000 deaths per year (WHO 2015). There are approximately 12.7 million infected individuals in the Middle East and North Africa region, with approximately 7.2 million in Egypt (Hotez et al. 2012). The transmission has been greatly reduced in Egypt, where a non-integrated

Section Editor: Christoph G. Grevelding

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intervention strategy is still used and transmission of infection in high-risk areas in the Nile Delta remains uninterrupted. Therefore, improved, more comprehensive control strategies are still in need (Barakat et al. 2014).

Recently, nanoparticles (NPs) have been widely used in the biomedical sciences for facilitating antimicrobial drugs' administration and minimizing their side effects. The NPs' ultra-small size, large surface-area-to-mass ratio, and high reactivity add to their benefits (Zhang et al. 2010). Silver (Ag) and gold (Au) NPs are highly effective against bacteria, viruses, and eukaryotic microorganisms, as well as having the ability to destroy cancer cells (Gong et al. 2007; Kuo 2009).

Plant-synthesized Ag NPs are effective ovicides, larvicides, pupicides, adulticides, and oviposition deterrents against mosquito species of medical and veterinary importance (Jayaseelan et al. 2011; Haldar et al. 2013; Govindarajan et al. 2015; Lallawmawma et al. 2015; Dinesh et al. 2015;

	bignostic performance of Ag and Au INFS monuscicidal effect along their concentration gradients									
	Concentration Ag (3–100) µg/ml	Concentration Au (100–200) µg/ml	Control	Chi-square test	P value	P1	P2	Р3		
No. of dead	60 (75%)	38 (63.3%)	0 (0%)	22.253	< 0.001	0.192				
No. of living	20 (25%)	22 (36.7%)	10 (100%)				< 0.001	< 0.001		
Total	80 (100%)	60 (100%)	10 (100%)							

 Table 1
 Diagnostic performance of Ag and Au NPs molluscicidal effect along their concentration gradients

Chitra et al. 2015; Benelli 2016). Au NPs have also shown potential as insecticides (Teimouri et al. 2018). The effects of Au NPs on arthropod vectors, such as ticks, tsetse flies, tabanids, sandflies, and blackflies, and additive-related ecotoxicological assays are recorded (Benelli 2018).

NPs have potent antiparasitic effects by acting as a drug vehicle for conventionally used drugs, such as praziquantel in the treatment of schistosomiasis (Kolenyak-Santos et al. 2014) and doxycycline and ivermectin in the treatment of filariasis (Binnebose et al. 2015). Furthermore, NPs themselves can have antiparasitic action directly, as Ag, chitosan, and curcumin NPs have anti-giardia effects (Said et al. 2012), and Au NPs have an anti-neuroschistosomal effect in mice infected with *Schistosoma mansoni* (Dkhil et al. 2015a).

Only a few studies have investigated NPs as cercaricides or molluscicides. Ag NPs show a toxic effect on *S. mansoni* cercariae, inhibiting their penetration into linolenic acidimpregnated agar (King and Highashi 1992). They induce *Schistosoma japonicum* cercarial tail-shedding and agitated behavior and block cercarial infectivity (Cheng et al. 2013). Au NPs lead to a reduction in reproductive capacity and death of *Biomphalaria alexandrina* (El-Sayed and El-Sherbini 2006, El-Hommossany and El-Sherbibni 2011) and *Bulinus truncatus* snails (Ragheb 2009).



■ No. of Dead ■ No. of living

Fig. 1 Bar chart showing the diagnostic performance of Ag and Au NPs molluscicidal effect along their concentration gradients

In light of the above, Ag and Au NPs can have a role in preventing schistosomiasis, so this study was carried out to evaluate the potential effects of Ag and Au NPs on *B. alexandrina* snails and *S. mansoni* cercariae in vitro and to assess their effects on the infectivity of cercariae in vivo.

Materials and methods

The study was done in Medical Malacology Laboratory and Animal House at Theodor Bilharz Research Institute (TBRI) Giza, Egypt. All reagents were purchased from Sigma Aldrich (USA) except Ag and Au NPs from Nanotech Company (Giza, Egypt).

In vitro study

Snail and parasite collection

Healthy *B. alexandrina* snails were collected from different water streams at Giza Governorate, Egypt, during the spring of 2016. Snails were cultivated according to Mossalem and Elenain (2014). *S. mansoni* miracidia were obtained from cleaned eggs extracted from the intestines of infected mice by hatching them in dechlorinated tap water $(25 \pm 1 \text{ °C})$ (Eissa et al. 2011). *S. mansoni* cercariae were obtained from experimentally infected *B. alexandrina* snails which were individually placed in 24-well culture plates containing 1 ml of distilled water (dH₂O) followed by exposure to artificial light for 1 h. Emerged cercariae were observed under dissecting microscope and counted (Kiros et al. 2014).

Detection of molluscicidal rate

Serial dilutions from each drug were prepared in dH₂O (Oteifa et al. 1975). For each concentration, 10 snails were used (each snail 6–8 mm in diameter) and this was repeated in triplicates. Each snail was exposed to each concentration for 24 h then allowed to recover for the next 24 h. The result was documented on the 3rd day; all were maintained at $25^{\circ} \pm 1$ °C and pH 7.4. Control snails were maintained under the same experimental conditions in dechlorinated water (Litchfield and Wilcoxon 1949). The dose 100 µg/ml was chosen empirically

Fig. 2 Treated cercariae observed under a dissecting microscope (×10) showing; tail loss (arrows) (a) and swollen body (b)



as a starting dose; it showed a 100% lethal effect with Ag NPs while for Au NPs it was increased to 200 μ g/ml to reach a 100% lethal effect. Snails were considered dead if they did not move and showed a discolored body retracted into the shell (Vijay 2010). Percentage of snail mortality was detected and the LC₅₀ and LC₉₀ values of each drug were determined.

Effect of Ag and Au NPs on snail infection with *S. mansoni* miracidia

B. alexandrina snails were exposed to the LC_{50} (9.68 µg/ml Ag and 133.7 µg/ml Au) followed by testing its susceptibility to infection with *S. mansoni* miracidia. Infection of the snails was done by exposing 50 snails to 500 miracidia maintained at room temperature for 3 weeks in a plastic container. Infected snails were used as control group (Massoud et al. 2004). Evaluation of the infection rate of snails was checked to start from the 3rd week post-infection (PI) to assess their infection. The snails were examined for cercarial shedding by the distribution of the snails in 24-well culture plate left in light for 1 h. Then the plate was examined under dissecting microscope to determine the infection rate of snails; by counting the number of snails which acquired infection and shed cercariae (El-Assal et al. 1997).

Testing Ag and Au NPs cercaricidal potential

Previously infected *B. alexandrina* snails with *S. mansoni* miracidia were exposed to light for 1 h to shed cercariae, which were distributed in petri dishes (100 cercariae in each dish) then exposed to a range of dilutions for each drug. The same number of cercariae was placed in new petri dishes containing dechlorinated water as a control group (Kiros et al. 2014). Serial dilutions of Ag NPs (50, 25, 10, 5, and 3 μ g/ml) and of Au NPs (100, 75, 50, 25, 10, and 5 μ g/ml) were tested in duplicates. The cercariae were observed under a dissecting microscope at successive intervals of 15, 30, 45, and 60 min.

Cercariae were considered dead when they stopped movement sank down or showed a detached tail (Eissa et al. 2011). Percentage of cercarial mortality was detected and the LC_{50} and LC_{90} values of each drug were determined (Rug and Ruppel 2000). For examining the cercariae by transmission electron microscopy (TEM), a number of the treated cercariae (exposed to each of Ag and Au NPs) and control cercariae was put in eppendorfs and concentrated by centrifugation at 1400 rpm at 15 °C for 5 min, then sediments were fixed in a 10% glutaraldehyde. Ultrathin sections were examined by TEM after staining with uranyl acetate and lead citrate (Chisty et al. 2004).

In vivo study

The study included 60 male Swiss Albino mice, 6–8 weeks old, weighing 20–25 g, purchased and housed at the Animal House, TBRI, after fulfilling all animal ethical considerations.

Routes of experimental infection

The LC₉₀ for each drug (Ag-Au NPs), as estimated from the present cercaricidal study, was used. *S. mansoni* cercariae were obtained from laboratory-bred infected *B. alexandrina* snails. Two routes of experimental infection were applied where each mouse was infected with 60 ± 10 *S. mansoni* cercariae suspended in 0.2–0.3 ml diluents. Infection by tail immersion (TI) was done according to Tendler and Pinto (1981) and by subcutaneous injection (SC) according to Peters and Warren (1969). Control groups of mice were infected with cercariae suspended in dH₂O. Experimental groups included two groups of mice infected with cercariae suspended in Ag and Au NPs (pre-treated cercariae) and another two groups infected with non-treated cercariae suspended in dH₂O, then the infected mice were treated by injection of Ag and Au NPs 1 h PI targeting early stages of infection.



Fig. 3 Scattered plot illustrating the effect of different concentrations of Ag and Au NPs (µg/ml) on the mortality of *S. mansoni* cercariae (%) at different time intervals



Fig. 4 TEM showing changes in the cercariae of *S. mansoni* on exposing to Ag NPs and Au NPs. **a**, **b** Control cercariae showing intact corrugated tegument (T) with spines (arrow) and compact muscle layer (M) without cellular swelling or edema. **c**, **d** Cercariae treated with Ag NPs (10 μ g/ml) 30 min after exposure showing; thinning of tegument (T) with focal loss of spines and edematous swelling of the muscle layer and deeper parenchyma (arrow). **e–h** Cercariae treated with Au NPs (100 μ g/ml) 45 min

after exposure showing; e loss of external glycocalyx leading to thinning of the tegument (T) with focal loss of spines and edematous swelling of the muscle layer (M) and deeper parenchyma; f migration of cytoplasmic granules (cg) towards the tegument (arrow); g, h tegumental thinning (T) leading to external protrusion with focal breach of continuity of the tegument and a severe degeneration leading to focal lysis of the tegument causing expulsion of sub-tegumental materials (arrow)

Infection of mice and experimental design

Control groups (C)

Group I (infection control): mice infected with nontreated cercariae, divided into Ia: six mice infected by SC injection, and Ib: six mice infected by TI technique.

Group II (negative control): six non-infected healthy mice.

Group III (drug control): six non-infected mice treated by nano drugs divided into *IIIa*: three mice treated with Ag NPs, and *IIIb*: three mice treated with Au NPs.

Experimental groups

Group IV: 12 mice infected with treated cercariae by TI, divided into *IVa*: 6 mice infected with Ag NPs, and *IVb*: 6 mice infected with Au NPs.

 Table 2
 Evaluation of TWB in SC-infected mice with treated cercariae and with normal cercariae treated 1 h PI

TWB	SC injection with treated cercariae		Infection	F	P value	Post Hoc		
	Ag (Va)	Au (Vb)	(Ia)			P1	P2	Р3
Mean ± SD	0.00 ± 0.00	7.33 ± 1.63	15.33 ± 3.72	64.03	< 0.001	< 0.001	< 0.001	< 0.001
TWB	SC injection with ne Ag (VIa)	ormal cercariae treated 1 h PI Au (VIb)	Infection control (Ia)					
$Mean \pm SD$	7.33 ± 2.89	7.67 ± 2.08	15.33 ± 3.72	8.758	0.008	0.903	0.007	0.009

F is score for the analysis of variance test (ANOVA), P is probability value

TWB	TI with treated cercariae		Infection control (Ib)	F	P value	Post hoc		
	Ag (IVa)	Au (IVb)				P1	P2	Р3
Mean ± SD	0.00 ± 0.00	12.50 ± 3.83	22.17 ± 1.47	131.808	< 0.001	< 0.001	< 0.001	< 0.001
TWB	TI with normal cerc Ag (VIIa)	ariae treated 1 h PI Au (VIIb)	Infection control (Ib)					
Mean ± SD	12.67 ± 5.51	7.33 ± 3.06	22.17 ± 1.47	24.293	< 0.001	0.069	0.002	< 0.001

Table 3 Evaluation of TWB in TI-infected mice with treated cercariae and in TI-infected mice with normal cercariae treated 1 h PI

Table 4 Evaluation of egg count/g intestine in SC-infected mice with treated cercariae and SC-infected mice with normal cercariae treated 1 h PI

Tissue egg count/g intestine	SC injection with treated	cercariae	Infection control (Ia)	ection control (Ia) F P value Post Hoc			2	
	Ag (Va)	Au (Vb)				P1	P2	Р3
Mean ± SD	0.00 ± 0.00	3744.32 ± 795.59	3439.66 ± 1282.69	34.16	< 0.001	< 0.001	< 0.001	0.554
Tissue egg count/g intestine	SC injection with normal Ag (VIa)	cercariae treated 1 h PI Au (VIb)	Infection control (Ia)	F	P value			
Mean \pm SD	3562.14 ± 1118.09	2924.86 ± 481.57	3439.66 ± 1282.69	0.291	0.754			

Group V: 12 mice infected with treated cercariae by SC route divided into *Va*: 6 mice infected with Ag NPs, and *Vb*: 6 mice infected with Au NPs.

Group VI: six mice infected with non-treated cercariae by SC route then after 1 h PI mice were injected with the NPs, they were divided into *VIa*: three mice injected with Ag NPs, and *VIb*: three mice injected with Au NPs.

Group VII: six mice infected with non-treated cercariae by TI then after 1 h PI mice were injected with the NPs, they were divided into *VIIa*: three mice treated with Ag NPs, and *VIIb*: three mice treated with Au NPs.

Evaluation of the antischistosomal activity of Ag and Au NPs

After 6 weeks, PI mice were decapitated and each mouse was subjected to two types of assessments. Parasitological assessment by total worm burden (TWB) (Smithers and Terry 1965), tissue egg count/g intestine and liver (Cheever 1968), and lastly oogram pattern (Pellegrino

and Faria 1965) were done. Histopathological assessment by examination of liver tissue (Druray and Wallington 1980) was also performed.

Statistical analysis

Data were analyzed using Statistical Program for Social Science (SPSS) version 20.0 to calculate the LC_{50} and LC_{90} by probit analysis. Analytical statistics included the following: a one-way analysis of variance (F) (ANOVA), Chi-square test of significance, and Post Hoc test. The confidence interval was set to 95% and the margin of error accepted was set to 5%. Probability (*P* value) was calculated (*P* value ≤ 0.05 was considered significant, and *P* value ≥ 0.05 was considered as highly significant, and *P* value ≥ 0.05 was considered insignificant) (P1: comparative performance between Ag and Au, P2: comparative performance between Ag and control, and P3: comparative performance between Au and control).

Table 5 Evaluation of egg count/g intestine in TI-infected mice with treated cercariae and TI-infected mice with normal cercariae treated 1 h PI

Tissue egg count/g intestine	TI with treated cercariae		Infection control (Ib)	F	P value	Post Hoc		
	Ag (IVa)	Au (IVb)				P1	P2	P3
Mean ± SD	0.00 ± 0.00	6114.58±2684.13	8391.24±1645.07	34.2	< 0.001	< 0.001	< 0.001	0.047
Tissue egg count/g intestine	TI with normal cerca Ag (VIIa)	ariae treated 1 h PI Au (VIIb)	Infection control (Ib)	F	P value	Post Hoc P1	P2	Р3
Mean ± SD	3589.16 ± 1195.79	4276.89 ± 899.36	8391.24 ± 1645.07	15.077	< 0.001	0.566	< 0.001	0.003

Tissue egg count/g liver Mean ± SD Tissue egg count/g liver	SC injection with treated cercariae		Infection control (Ia)	F	P value	Post Hoc		
	Ag (Va)	Au (Vb)				P1	P2	Р3
Mean ± SD	0.00 ± 0.00	3080.28 ± 386.59	2228.98 ± 814.61	56.02	< 0.001	< 0.001	< 0.001	0.013
Tissue egg count/g liver	SC injection with norm Ag (VIa)	al cercariae treated 1 h PI Au (VIb)	Infection control (Ia)	F	P value			
Mean ± SD	3220.95 ± 1410.58	2213.93 ± 1041.50	2228.98 ± 814.61	1.063	0.385			

 Table 6
 Evaluation of egg count/g liver in SC-infected mice with treated cercariae and SC-infected mice with normal cercariae treated 1 h PI

Table 7 Evaluation of egg count/g liver in TI-infected mice with treated cercariae and TI-infected mice with normal cercariae treated 1 h PI

Tissue egg count/g liver	TI with treated cercariae		Infection control (Ib)	F	P value	Post Hoc		
	Ag (IVa)	Au (IVb)				P1	P2	Р3
Mean ± SD	0.00 ± 0.00	4775.56 ± 2987.82	7135.29±1797.39	19.565	< 0.001	< 0.001	< 0.001	0.06
Tissue egg count/g liver	TI with normal cercariae treated 1 h PI		Infection control (Ib)	F	P value	Post Hoc		
	Ag (VIIa)	Au (VIIb)				P1	P2	P3
Mean ± SD	2971.56 ± 1030.85	3517.37 ± 811.81	7135.29 ± 1797.39	10.532	0.004	0.661	0.003	0.007

Results

In the in vitro study, the molluscicide assay revealed that Ag NPs at concentrations of 100, 50, 40, and 30 µg/ml led to death of 100% of *B. alexandrina* snails, while at concentrations of 25, 10, 5, and 3 µg/ml, they led to 80, 60, 40, and 20% snail death, respectively. Au NPs at concentrations of 200, 180, and 160 µg/ml led to the death of 100% of snails, while at concentrations of 140, 120, and 100 µg/ml, they led to 60, 20, and 0% snail death, respectively. The LC₅₀ of Ag NPs was 9.68 µg/ml, and the LC₉₀ was 24.75 µg/ml, while for Au NPs, the LC₅₀ was 133.7 µg/ml and LC₉₀ was 150.7 µg/ml. Ag NPs showed a highly effective molluscicidal potential, even at lower concentrations than Au NPs, albeit with no statistically significant difference (P > 0.05)



Fig. 5 Bar chart showing mean of TWB in mice infected with treated cercariae and with normal cercariae treated 1 h PI by SC and TI routes for all the experimental groups. (Codes with different letters indicate significant difference)

between the effects of Ag and Au NPs (P = 0.192). Moreover, there was a highly statistically significant difference (P < 0.001) when comparing Ag or Au NPs with the control group, demonstrating their significant molluscicidal potential (Table 1, Fig. 1).

The effect of the LC_{50} of each of Ag and Au NPs on *B. alexandrina* snails revealed an absence of cercarial shedding in comparison to the control ones.

The cercariae showed changes in behavioral pattern, such as swimming and contraction, and morphological alterations, such as tail shedding (Fig. 2). Ag NPs showed a cercaricidal potency of 100% lethality at all time intervals from 30 min up to 60 min with concentrations of 50, 25, and 10 µg/ml. Au NPs showed a cercaricidal potency of 100% lethality at 60 min with concentrations 100, 75, and 50 µg/ml and at 45 min with concentrations 100 and 75 μ g/ml. The mortality rate increased with the increase in time of exposure. The dilution titer continued to 3 μ g/ml for Ag and 5 μ g/ml for Au NPs, where they reached 0% lethality (Fig. 3). The starting values of tabulation for statistical input were the concentrations showing different lethal effects. At all time intervals, Ag NPs at concentrations above 50 µg/ml were 100% lethal, whereas only Au NP concentrations above 100 µg/ml were 100% lethal at all time points. For evaluating the cercaricidal potency, the results of different concentrations at 15 min were chosen to calculate the LC_{50} and LC_{90} of each drug, as the best outcome is obtained from the shortest time with the minimal drug concentration. Ag NPs' LC50 was 19.6 µg/ml, and their LC₉₀ was 39.7 µg/ml, while Au NPs' LC₅₀ was 127 µg/ml and LC₉₀ was 244 µg/ml. Ag NPs showed a highly effective cercaricidal potency with a lower range of concentrations than Au NPs (Fig. 3).

Fig. 6 Bar chart showing mean of tissue egg count/g intestine in mice infected with treated cercariae and with normal cercariae treated 1 h PI by TI and SC routes for all the experimental groups. (Codes with different letters indicate significant difference)

Fig. 7 Bar chart showing mean of

infected with treated cercariae and with normal cercariae treated 1 h

PI by TI and SC routes for all the

experimental groups. (Codes with

different letters indicate signifi-

cant difference)

tissue egg count/g liver in mice



with Ag with Au 1h Pl with Ag 1h Pl withAu

Tissue egg count / gm intestine (SC infection)

Tissue egg count / gm intestine (TI technique)



Tissue egg count / gm liver (SC infection) Tissue egg count / gm liver (TI technique)

Table 8 Evaluation of oogram pattern in SC-infected mice with treated cercariae and with normal cercariae treated 1 h PI

Oogram pattern	SC injection with	treated cercariae	Infection control (Ia) F P value Post Hoc					
	Ag (Va)	Au (Vb)				P1	P2	Р3
Immature egg								
Mean \pm SD	0.00 ± 0.00	53.17 ± 3.49	51.00 ± 1.26	1183.81	< 0.001	< 0.001	< 0.001	0.1
Mature egg								
Mean \pm SD	0.00 ± 0.00	41.50 ± 3.45	43.17 ± 1.17	811.44	< 0.001	< 0.001	< 0.001	0.19
Dead egg	0.00 ± 0.00	5.33 ± 1.51	5.83 ± 0.75					
Mean \pm SD				66.41	< 0.001	< 0.001	< 0.001	0.387
Oogram pattern	SC injection with normal cercariae treated 1 h PI		Infection control (Ia)	F	P value	Post Hoc		
0 1	Ag (VIa)	Au (VIb)				P1	P2 <0.001 <0.001 ; P2 0.597 0.676 0.782	P3
Immature egg	51.67 ± 2.89	51.00 ± 1.00	51.00 ± 1.26					
Mean ± SD				0.169	0.847	0.647	0.597	1
Mature egg								
Mean \pm SD	42.67 ± 2.52	41.33 ± 1.53	43.17 ± 1.17	1.257	0.33	0.345	0.676	0.148
Dead egg	5.67 ± 1.15	7.67 ± 0.58	5.83 ± 0.75					
Mean \pm SD				5.899	0.023	0.016	0.782	0.012

Fig. 8 Bar chart showing oogram pattern in SC-infected mice with treated cercariae and with normal cercariae treated 1 h PI. (Codes with different letters indicate significant difference)



By TEM, cercariae exposed to Ag 10 μ g/ml and Au 100 μ g/ml showed progressive morphological (ultrastructural) changes at 30 and 45 min post-exposure, respectively, compared to negative-control cercariae. These concentrations were chosen at this time of exposure because they were 100% lethal (Fig. 4).

Assessing the effects of NPs in vivo using the parasitological parameters TWB, tissue egg count/g intestine, and tissue egg count/g liver with either route of infection, TI or SC, showed an overall stronger effect of Ag than Au NPs. The effect of Ag NPs was better when applied pre- than post-infection. There were significant outcomes among all treated groups and the control in all the experiments (Tables 2, 3, 4, 5, 6, and 7, Figs. 5, 6, and 7).

The oogram pattern in SC infection with treated cercariae showed a highly significantly different outcome when comparing Ag with Au NPs and Ag NPs with control groups (P < 0.001), though there were similar outcomes between Au

NPs and control groups (P > 0.05). At 1 h PI, there was no significantly different outcome (P > 0.05) between the treated groups and the control (Table 8, Fig. 8). The oogram pattern in TI infection with treated cercariae showed a highly significant difference ($P \le 0.001$) between all groups in the mean numbers of immature, mature, and dead eggs (Fig. 9), except for a non-significantly different outcome (P > 0.05) between Au NPs and control groups in the mean number of dead eggs. At 1 h PI, there was no significantly different outcome (P > 0.05) between both drugs, whereas there was a significantly different outcome (P > 0.05) between outcome (P > 0.05) between both drugs, whereas there was a significantly different outcome (P < 0.001) between each drug and the control, in the mean numbers of immature and mature eggs (Table 9, Fig. 10).

In histopathological examination, only in the Ag NPtreated cercariae group were normal hepatic architecture and histological patterns seen; otherwise, all other groups showed abnormal changes (Fig. 11).

Fig. 9 Oogram pattern for S. mansoni (×100) showing: a Mature ova containing fully developed miracidia, dead ova appearing granular and dark with irregular outline containing disintegrated miracidia. b Immature ova containing small embryos



Oogram pattern	TI with treated cercariae		Infection control (Ib)	F	P value	Post Hoc		
	Ag (IVa)	Au (IVb)				P1	P2	Р3
Immature egg								
$Mean \pm SD$	0.00 ± 0.00	51.17 ± 1.47	60.33 ± 3.56	1282.69	< 0.001	< 0.001	< 0.001	< 0.001
Mature egg								
$Mean \pm SD$	0.00 ± 0.00	44.00 ± 0.89	34.33 ± 3.61	694.087	< 0.001	< 0.001	< 0.001	< 0.001
Dead egg	0.00 ± 0.00	4.83 ± 0.75	5.33 ± 1.03					
Mean \pm SD				95.612	< 0.001	< 0.001	< 0.001	0.259
Oogram pattern	TI with normal co	ercariae treated 1 h PI	Infection control (Ib)	F	P value	Post Hoc		
	Ag (VIIa)	Au (VIIb)				P1	P2	P3
Immature egg								
$Mean \pm SD$	51.00 ± 1.00	51.67 ± 1.53	60.33 ± 3.56	15.664	< 0.001	0.776	< 0.001	0.002
Mature egg								
$Mean \pm SD$	43.33 ± 1.53	41.33 ± 1.15	34.33 ± 3.61	12.261	0.003	0.411	0.002	0.007
Dead egg								
$Mean\pm SD$	5.67 ± 0.58	7.00 ± 1.00	5.33 ± 1.03	3.188	0.09	0.117	0.629	0.054

 Table 9
 Evaluation of oogram pattern in TI-infected mice with treated cercariae and TI-infected mice with normal cercariae treated 1 h PI

Discussion

The in vitro study revealed that Ag and Au NPs have molluscicidal activity against *B. alexandrina* in a concentration-dependent manner. These results were in parallel to Abdel-Hamid and Mekawey (2014), who reported a dose-dependent molluscicidal effect of Ag NPs on *B. alexandrina*. Ali et al. (2014) demonstrated that Ag NPs were lethal to freshwater snail *Lymnaea luteola* which was in a dose- and time-dependent manner. Moreover, Bernot and Brandenburg (2013) and Goncalves et al. (2017), indicated that even very low concentrations of Ag NPs can affect all the vital rates of *Physa acuta* snails. In the current study, snails were examined after the 3rd week of infection to assess whether or not they had acquired infection after applying each drug individually. The study recorded the absence of cercarial shedding. This could be due to either changes induced by the drugs on snails, so that they could not acquire the infection, or that the drugs led to the death of the miracidia.

Fig. 10 Bar chart showing oogram pattern in TI-infected mice with treated cercariae and in TI-infected mice with normal cercariae treated 1 h PI. (Codes with different letters indicate significant difference)



Fig. 11 Histopathological changes in hepatic tissue in different groups 6 weeks PI (Hx and E stain ×400). a Positive control group showing severe inflammatory response in the liver indicated by inflammatory cellular infiltration, cytoplasmic vacuolation, degeneration of hepatocytes, dilated hepatic sinusoids and more Kupffer cells with intact ova in the center; b Ag NPs-treated cercariae showing normal hepatic architecture; c Au NPs-treated cercariae; d Ag NPs treated 1 h PI; e Au NPs treated 1 h PI. c-e showed the same picture as control group



Regarding the cercaricidal potency, Ag and Au NPs were effective in a dose- and time-dependent manner. Both drugs induced agitated behavior followed by death, with the appearance of swollen cercariae. The study revealed that the effect of Ag NPs was far beyond Au NPs. By the end of the 1st hour, all tested concentrations of Ag NPs were lethal to all cercariae, while Au NPs concentrations below 50 μ g/ml were not lethal. To our knowledge, this is the first study to assess Au NPs' cercaricidal activity against *S. mansoni*.

King and Highashi (1992) reported that Ag ions at concentrations above 0.09 mM were toxic to *S. mansoni* cercariae, and concentrations below that were non-toxic but inhibited their penetration into the linolenic acid-impregnated agar. Cheng et al. (2013) reported similar results to our study when testing Ag NPs on *S. japonicum* cercariae and found that it rapidly induced cercarial tail-shedding, agitated behavior and a decrease in cercarial secretion. They used higher concentrations of Ag NPs which completely blocked cercarial infectivity after 30 min of exposure. Although their concentrations were higher than that employed in the present study, they showed nearly the same impact on cercariae, and this could be attributed to the different biochemical manufacturing processes of the NPs. The potent effect of Ag NPs on cercariae was attributed to Ag ions, which are continuously released in water suspension and can bind to the papillar sites on the cercarial surface thereby, inhibiting fatty acid-induced cercarial acetabular gland secretion (Liu and Hurt 2010).

The ultrastructural changes recorded in the present study were in the form of thinning of tegument with focal loss of spines, oedematous swelling of the muscle layer and deeper parenchyma with an external protrusion and focal breach of continuity of the tegument. These findings were similar to findings by Chisty et al. (2004), who used hinokitiol, a plant that potently protects the skin, against cercarial penetration, and Eissa et al. (2011), who used miltefosine, a promising schistosomicidal compound.

The in vivo study applied different routes of infection to exclude the impact of the route on the drug administration on its cercaricidal potency. In the present work, the timing of 1 h PI was chosen to assess drug potency at preventing early infection. A higher infection rate was reported by the TI route than SC injection through the whole study. In subcutaneously infected mice with *S. mansoni* cercariae, approximately 70% of the larval forms are destroyed before reaching maturity (Holanda et al. 1974 and Tendler et al. 1985), while by TI the loss is not more than 20% of the infecting cercariae during the skin phase (Mangold and Dean 1983). In this context, it seems that experimental infection by the TI route leads to greater infection than SC (Vilar and Pinto 2005).

By either route, the in vivo study showed that Ag NPs led to the absence of adult worms in mice infected by treated cercariae, while in mice treated 1 h PI they led only to a decrease in TWB. This is in accordance with Cheng et al. (2013), who used Ag NP-treated cercariae by SC infection and proved that at high concentrations, they caused an absence of adult worms and at lower concentrations caused a decrease in TWB. Those effects were attributed to Ag ions, which at high concentrations were lethal to cercariae but at low concentration led only to tail loss. Moreover, they mentioned that cercariae without tails can still penetrate into mice and develop into adult worms, in spite of the decreased infection rate.

Within all tested groups, Au NPs showed an overall decrease in the TWB, which can be interpreted by the finding of Dkhil et al. (2015b) that Au NPs has a strong ability to scavenge free radicals.

The effect of Ag and Au NPs on tissue egg load/g intestine and tissue egg load/g liver was assessed, as an important parameter for both the magnitude of infection and efficacy of antischistosomal drugs. A higher egg load/ g intestine than that in liver was recorded in infection control mice at 6 weeks PI. Friedman (2004) stated that the highest number of eggs is deposited in the liver at the beginning of infection, and then the number of ova decreases with infection time, associated with a corresponding increase in the number of ova in the intestine. Ag NPtreated cercariae injected by either route showed an absence of eggs in the intestine and liver, which highlights the potency of the drug. This finding is in line with Cheng et al. (2013).

By TI, Au NPs showed a significant decrease in the count of eggs/g intestine and eggs/g liver compared to infection control in all groups of infected mice. The effect of NPs of decreasing the number of ova in hepatic tissue of infected mice can be attributed to being an antioxidant (Abdallahi et al. 2001). Selenium NPs increase the level of glutathione and reduce the levels of nitrite/nitrate and malondialdehyde (Dkhil et al. 2016).

In the present study by either route, no eggs were detected in mice infected with Ag NP-treated cercariae. This was due to the absence of adult worms in these mice, with a subsequent absence of eggs. However, post-treated Ag NP groups and all groups of Au NPs (pre-treated and posttreated) showed similar results to infection control. The different outcomes from previous studies may be attributed to the longer study durations or repeated dosing schedules of the tested drugs (Cancado et al. 1965; Farid et al. 2013; Abaza et al. 2013).

Histopathological examination of the liver of mice subcutaneously infected with Ag NP-treated cercariae showed no granuloma formation, confirming the earlier results in the current work and verifying the drug's potency. In contrast, the rest of the groups (Ag treated 1 h PI and all groups of Au) showed a similar histopathological picture to that of infection control as a sequel to the presence of eggs. This was contrary to Dkhil et al. (2015a, b), who used Ag NPs twice, on days 46 and 49 PI, and reported a decrease in granuloma diameter and to earlier studies that showed that Au NPs improved the histopathological changes in the spleen and brain from mice infected with *S. mansoni* (Dkhil et al. 2017). Again, this different outcome may be attributed to the different intervals in each study.

Conclusions

The present study clarified the in vitro role of Ag and Au NPs on *S. mansoni* as molluscicides and cercaricides and how they can prevent or modulate the infectivity of cercariae in vivo, eliciting their preventive role against schistosomiasis. The greater efficacy of Ag NPs than Au was highlighted. The applicability of these NPs in the field as preventive ointments or lotions is needed over longer intervals to see the potential of these NPs as chemotherapeutics or preventive drugs. More studies are required to track the different mechanisms of action of each NP, as well as using different sizes and shapes of NPs should be comparatively evaluated to assess whether different manufacturing criteria will alter the response.

Compliance with ethical standards

The study was started after being approved by the Ethical Committee of Scientific Research at TBRI after fulfilling all animal ethical considerations.

Conflict of interest Authors declare that they have no conflict of interest.

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