

Developmental morphology of a lyriform organ in the Western black widow (*Latrodectus hesperus*)

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Received: 5 July 2016/Revised: 30 August 2016/Accepted: 6 September 2016/Published online: 16 September 2016
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Abstract For most spiders, their sensory world is dominated by their ability to detect vibrational stimuli. The organs responsible for detecting substrate vibrations are located on the animals' extremities and known as lyriform organs, close aggregations of membrane-covered slits in the cuticular exoskeleton. The morphology and geometry of the lyriform organ is an important determinant of how it functions and the range of stimuli it can detect. Most work on the morphology, mechanics and physiology of lyriform organs has been conducted on adult wandering spiders, *Cupiennius salei*, and little is known about the morphology in other species or juveniles. We examine the morphology of the HS10 lyriform organ in both adult and juvenile Western black widows (*Latrodectus hesperus*). We find hypoallometric scaling of the lyriform organ and the size of individual slits when compared to body size. However, the cuticular pad distal to HS10 scales isometrically across successive instars. We also find an increase in the number of slits within the lyriform organ with each moult. Future work should address physiological responses of the organ across development, which could lead to a better understanding of the function of the cuticular pad and stimuli pertinent to the survival of little-studied juvenile spiders.

Keywords *Latrodectus hesperus* · Lyriform organ · Slit sensilla · Development · Allometry · Vibration detection

Introduction

Spiders are exquisitely sensitive to vibration. From species that dwell and hunt on plants and the ground to those that use silk webs and traps, vibration is commonly the stimulus modality used to detect prey (Masters and Markl 1981; Klärner and Barth 1982; Landolfi and Barth 1996), predators (Uetz et al. 2002; Lohrey et al. 2009; Uma and Weiss 2012) and potential mates (Schüch and Barth 1985; Maklakov et al. 2003; Elias 2003; Elias et al. 2005; Vibert et al. 2014). Vibrational signals are detected with mechanoreceptive strain sensors embedded within the cuticular exoskeleton (see Barth 2004, 2012 for reviews). Known as slit sensilla, these sensors detect strain in the cuticle and can be found singly, in loose clusters, or as distinct groupings known as lyriform organs. Their function spans proprioception of self-generated strains from muscle contraction and haemolymph pressure changes, to detection of externally generated vibrations (Barth 2012). However, it is the lyriform organs that are primarily concerned with detecting vibrational displacement from the external environment, stimuli that can be in the order of mere nanometres (Barth and Geethabali 1982; Hössl et al. 2009).

Spiders have many lyriform organs which are exclusively found on their extremities, most frequently at joints on distal sections of the legs (Pringle 1955; Barth 1971; Barth and Stagl 1976). The organs are composed of an array of membrane-covered slits in the cuticular exoskeleton. When a vibrational stimulus is transmitted to the lyriform organ, the compliant membranes are

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compressed by the surrounding hard cuticle which stimulates neural dendrites (Walcott 1969; Barth et al. 1984; Molina et al. 2009). Each slit within an organ has its own sensory cells and responds independently to a stimulus (Barth and Geethabali 1982). The attachment of the dendrite to the outer membrane is visible externally and is known as the coupling cylinder, the position of which does not necessarily correspond to the position of maximal compression of the slit (Hössl et al. 2007).

The main lyriform organ involved in detecting substrate vibrations is the HS10 organ, found dorsally on the distal end of the metatarsus (MT), just proximal of the metatarsal-tarsal (MT-T) joint (Barth and Libera 1970). Substrate vibrations are transmitted up the tarsus (T) causing it to strike the MT, at the MT-T joint. At the distal end of the MT, between the HS10 organ and MT-T joint, resides a cuticular pad. In the wandering spider, *Cupiennius salei*, this pad is heterogeneous in composition, being soft and compliant at its distal end, but hard and sclerotised at the proximal end, adjacent to the organ (Young et al. 2014; Erko et al. 2015). The pad acts as a high-pass filter, reducing the transmission of biologically irrelevant stimuli below 40 Hz, but also protecting the organ from high-amplitude stimuli that are potentially damaging (McConney et al. 2007; Young et al. 2014). The location of the slit within the lyriform organ, its length and its aspect ratio are all important factors in determining sensitivity to stimuli (Hössl et al. 2006, 2007, 2009), and with its own filter, the spider has evolved to detect a specific range of stimuli relevant to its survival and reproduction. Morphological aspects of the lyriform organ are crucial to its function, raising the question of how lyriform organs retain their function throughout development where the size of the animal changes significantly.

Much of what has been learned about the physiology, mechanics and morphology of lyriform organs has come from work on the wandering spider, *C. salei* (see Barth 2012 for review), although some work has also been conducted on web-dwelling species (Finck 1981; Klärner and Barth 1982; Barth 2002). The slit arrangement in HS10 in

four spider species shares a ‘basic pattern’ but variable numbers of slits, from 21 slits in the *C. salei* to just 8–10 in the American house spider *Achaearanea tepidariorum*. In addition to this, the length of slits 2 and 11 in the HS10 organ has been measured for nine spider species (Barth 2002). Although there are comparisons of slit numbers in lyriform organs between different instars in *C. salei* (Barth and Libera 1970), little is known about the development of this organ throughout a spider’s life span, or indeed to vibrational senses in juveniles in general.

We examine the HS10 metatarsal lyriform organ of the Western black widow, *Latrodectus hesperus*, throughout developing instars to adult spiders (Fig. 1). *L. hesperus* is a web-dwelling spider that builds cob webs in arid environments (Kaston 1970). After emerging from the egg sac, spiderlings spend up to 14 days in the natal web, before dispersing and remaining on solitary webs for the majority of their lives (Kaston 1970). Mature males abandon their solitary webs to seek out female mates where they perform courtship displays with a vibratory component (Ross and Smith 1979; Kasumovic and Andrade 2004; Vibert et al. 2014). These courtship vibrations are not a stimulus found in juvenile environments. However, the need to detect prey and predators will persist throughout all life stages. Because the sensitivity of lyriform organs depends on their morphology, it is imperative to understand how they change throughout spider development. Developmental changes in morphology are likely to impact the range of environmental stimuli each instar is able to perceive; we would therefore expect minimal changes to overall organ morphology if the biologically relevant stimuli within the environment remain consistent throughout different stages of growth.

Materials and methods

L. hesperus egg sacs were collected and incubated in plastic containers (length \times width \times height cm³ = 8.73 \times 8.73 \times 11.27; Amac Plastics) at 25 °C until spiderlings

Table 1 Measurements from the MT and HS10 across all instars and both adult sexes measured

Instar	MT length (mm)	Pad width (μ m)	HS10 width (μ m)	Max. slits seen
1st	0.51 \pm 0.02 (n = 10)	5.4 \pm 0.25 (n = 6)	14.7 \pm 1.56 (n = 6)	13 (n = 6)
2nd	0.71 \pm 0.05 (n = 10)	5.4 \pm 2.12 (n = 9)	13.7 \pm 2.12 (n = 9)	14 (n = 10)
3rd	1.12 \pm 0.03 (n = 10)	8.9 \pm 1.25 (n = 7)	18.1 \pm 1.39 (n = 7)	15 (n = 7)
4th	1.30 \pm 0.17 (n = 4)	11.0 \pm 2.80 (n = 4)	17.3 \pm 2.15 (n = 3)	16 (n = 3)
5th	2.09 \pm 0.08 (n = 2)	15.9 (n = 1)	22.9 (n = 1)	16 (n = 1)
Adult (m)	4.02 \pm 0.08 (n = 9)	37.2 \pm 5.64 (n = 9)	26.19 \pm 3.30 (n = 9)	20 (n = 9)
Adult (f)	6.44 \pm 0.32 (n = 10)	57.4 \pm 11.16 (n = 10)	47.17 \pm 6.63 (n = 10)	23 (n = 9)

Width of pad was measured between its distal margin and slit 1. Width of HS10 is measured between slit 1 and 12. MT length is measured along its dorsal edge

emerged. In keeping with previous terminology, the first instar was defined as the instar that emerged from the egg sac (Kaston 1970). Shortly after emergence, spiderlings were transferred to individual containers (length \times width \times height (cm) = $4.13 \times 4.13 \times 5.56$) where they were allowed to moult into successive instars at 24 °C. Up to 20 spiderlings were taken from each instar group and euthanised in 70 % ethanol where they were kept until imaging.

Images of the HS10 metatarsal lyriform organ were taken using scanning electron microscopy (SEM). To prepare samples for imaging, spiders were removed from ethanol and affixed to an SEM stub using double-sided tape. For early instars, the entire animal was positioned on the stub, but for adults and later instars, the first pair of legs was removed using fine scissors and positioned on the stub. Photographs were taken with a Nikon DXM 1200 camera mounted on a Zeiss Stemi 2000-C dissecting microscope. Images of the first pair of legs were taken, using Act-1 (Nikon Corp. 2000; Fig. 1c), to allow measurement of dimensions of the MT. Where necessary, hairs were shaved from around the HS10 organ; however, this was often not possible on smaller animals without causing damage to the surrounding tissue or leaving behind significant debris. Samples were subsequently sputter-coated with gold (PS3, Polaron, Watford, UK) before SEM imaging (Hitachi S530 SEM, Hitachi, Tokyo, Japan). Images were acquired and

digitised using Quartz PCI imaging software (Quartz PCI, Quartz Imaging Corporation, Vancouver, Canada), and morphological measurements taken using Corel Draw (Corel Corporation, Ottawa, Canada) and the JH Curve-Length macro.

Results

The HS10 organ is present from the first instar through to adulthood in *L. hesperus*. There is a basic set of 13 slits that are present across all instars examined (Figs. 2, 3; Table 1). At each moult between instars 1 and 5, HS10 gains an additional slit, added to the proximal end of the HS10 organ. The maximum number of slits seen in any metatarsal HS10 organ was 23, and this number was only seen on the largest spiders, the adult females. Males had a maximum of 20 slits. It was only possible to measure the first five instars due to mortality so it is not possible to tell whether the addition of a single slit at each moult remains the case after the fifth instar into the final moult to adulthood. The positions of the slits are largely conserved between individuals, although there is a small degree of variation (Fig. 3).

The coupling cylinder was visible in the larger HS10s of the adults due to the greater slit width. It was also possible to see it in the SEM images of at least some juveniles in

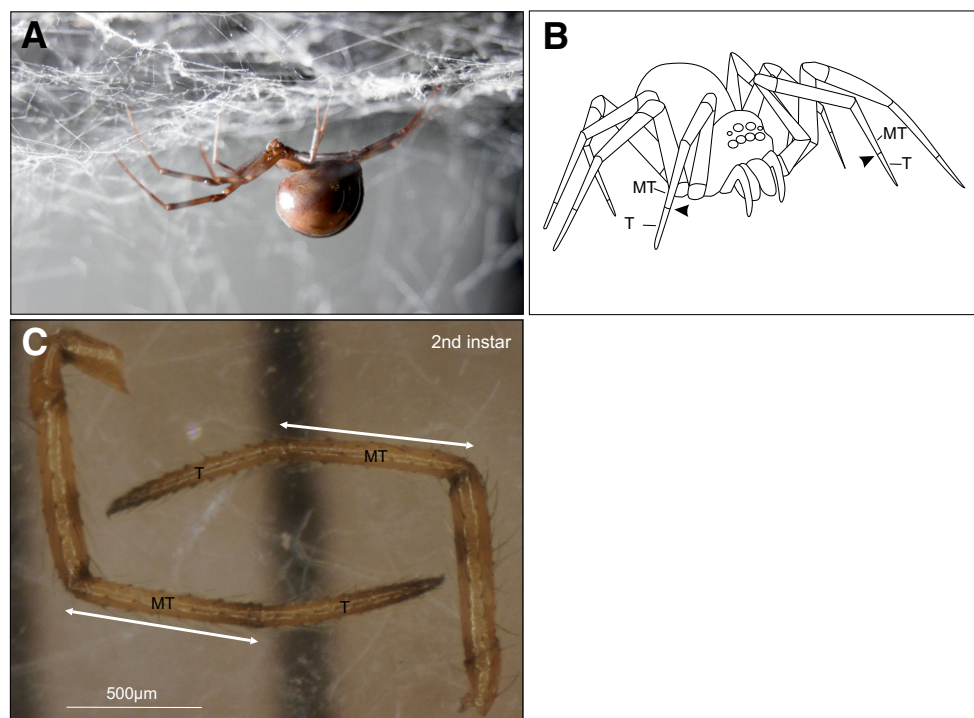
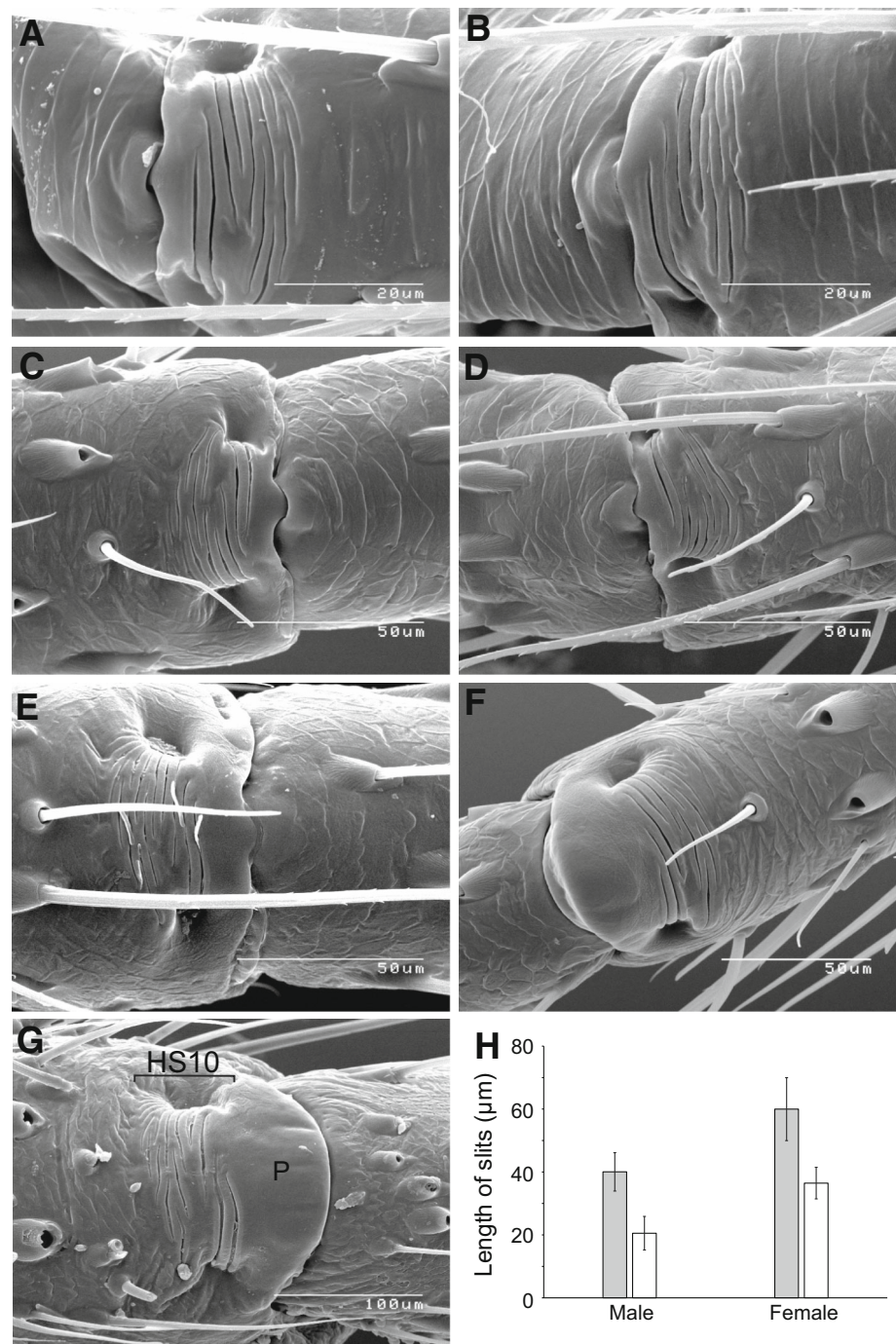


Fig. 1 **a** Adult female *L. hesperus* on her web. **b** Metatarsal lyriform organ (HS10) measured is located on the dorsal side of the metatarsus at the metatarsus–tarsus joint (arrows). **c** First pair of legs of a second-instar *L. hesperus*. Arrows indicate metatarsus length. MT metatarsus, T tarsus

Fig. 2 SEM images of the black widow (*L. hesperus*) HS10 lyriform organ. **a** First instar; **b** second instar; **c** third instar; **d** fourth instar; **e** fifth instar; **f** adult male; **g** adult female; **h** slit length for slit 2 (grey) and 11 (white) in adult males and females. $N = 7$



instars 1–4 (Fig. 4). The position of the coupling cylinder in adults of both sexes is somewhat offset from the centre of the longitudinal length of the slit, corresponding with previous findings in the wandering spider *C. salei* (Barth and Pickelmann 1975).

Relative to the growth of the MT, the HS10 organ shows little change in size across development and has a hypoallometric relationship with MT length. The allometric coefficient (α), or slope of a straight line fit to the log data, is less than 1 ($\alpha = 0.45$, Fig. 5a). Slit length is also

hypoallometric throughout development, with a slightly smaller allometric coefficient ($\alpha = 0.31$ – 0.36 , Fig. 5b). To reach mean adult male size, MT increases 7.9 times. This is a far greater rate than the 2.6–2.9 times increase in slit length to reach adult female size (1.4- to 1.9-fold increase to full adult male size) and HS10 width which grows slightly more than slit length to 3.2 times the size of the first instar in adult females and 1.8 times for adult males. In contrast, the cuticular pad at the distal end of the MT develops isometrically with MT length ($\alpha = 0.99$, Fig. 5).

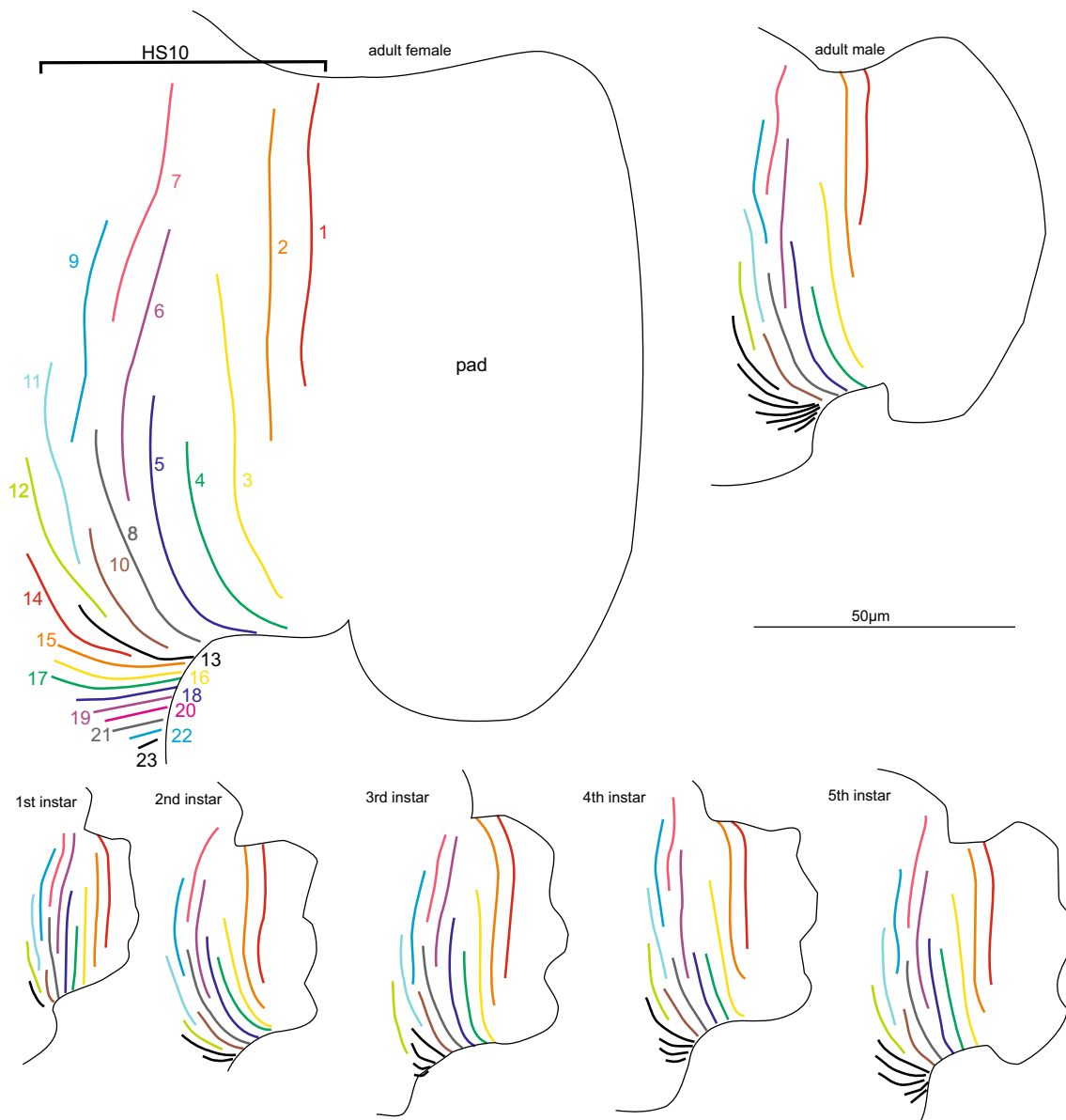


Fig. 3 External morphology of the lyriform organ across all instars illustrated at the same scale. Adult female slits are numbered

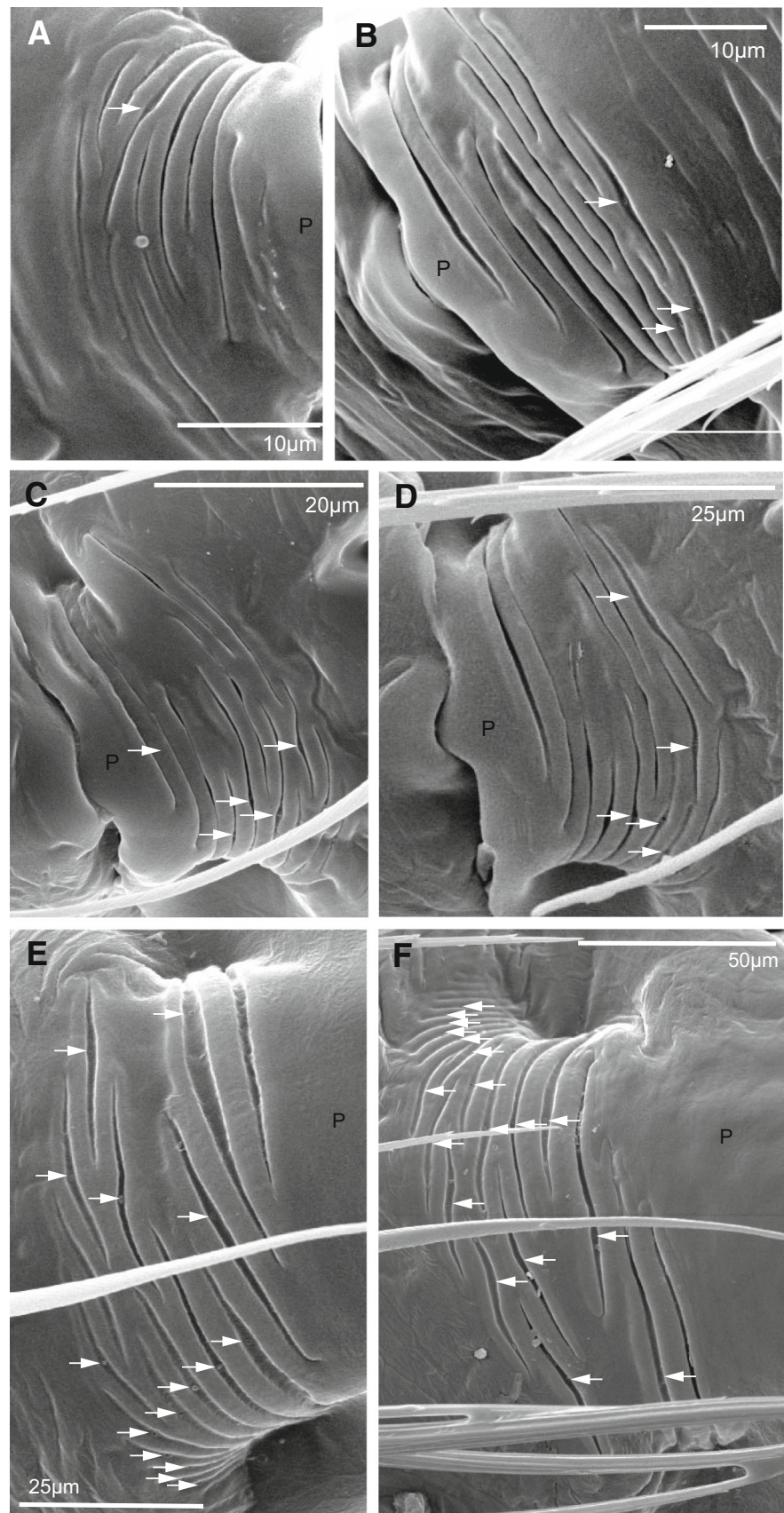
Using MT length as a proxy, first-instar spiders increase in size 12.7 times to reach the mean size of an adult female. The pad is pronounced in adults of both sexes, but in the 5 early instars measured, it is small in size (Fig. 3).

Discussion

The slit arrangement in the HS10 organ of *L. hesperus* resembles those in other web-building species, in particular *Zygiella x-notata* (Barth 2002). Similarly, the location of dendritic attachment to the outer membrane, visible externally as the coupling cylinder (Fig. 4), does not significantly deviate from the basic pattern found in

other species previously investigated (Barth 2002). It is possible that the number of slits present in the HS10 organ of *L. hesperus* reflects the number of moults that the animal has undergone. Kaston (1970) observed female *L. hesperus* moulting 9 times, occasionally up to 10, while males can mature twice as fast moulting up to 7 times. We observed the addition of a single slit per moult up to the fifth instar. If this trend continues to the final moult, then these females would have undergone a maximum of 10 moults (23 slits, with first instars having the basic 13 slits), and males 7 (20 slits), which is in fact what we observed. Ecological conditions can influence the rate of development in spiders and the number of instars before the final moult. If it is the case that the

Fig. 4 Position of coupling cylinder in HS10. **a–d** Are first to fourth instars, respectively. **e** Adult male. **f** Adult female. No specimens had all coupling cylinders visible simultaneously, especially in younger instars. *Arrows* indicate position of coupling cylinder. *P* indicates position of pad



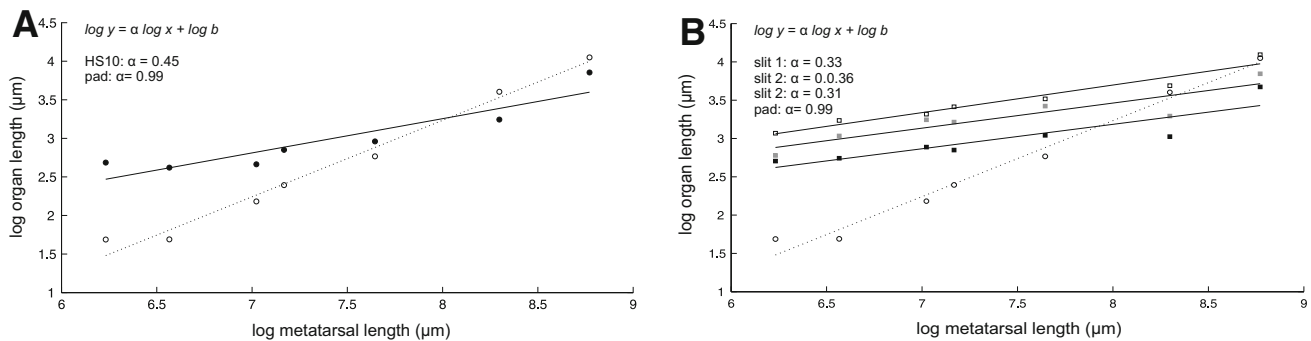


Fig. 5 **a** Log MT length against log HS10 width (black) and pad width (hollow). Straight line fits to the data show isometric growth of the pad with MT (dashed) and hypoallometric growth of the HS10 (black). **b** Log MT length against log slit length and pad width

(hollow circles). Straight line fits to the data show isometric growth of the pad with MT (dashed) and hypoallometric growth of the slit 1 (grey squares), slit 2 (white squares) and slit 9 (black squares)

number of slits in the HS10 lyriform organ corresponds to the number of moults, as seems likely from our results, it may be possible to use this as a tool in further studies where adults are available and the number of moults is of interest in developmental studies.

The aspect ratio of the slits within a lyriform organ determines its sensitivity to vibrational stimuli, with smaller slits requiring higher loads to generate enough deformation for a nervous response (Hössl et al. 2006, 2007, 2009). Allometry is defined as differential growth rates of body parts (Huxley and Teissier 1936). With the importance of morphology on the function of the lyriform organ, it is perhaps unsurprising that HS10 develops hypoallometrically (slower rate than body size) compared with body size. However, it is somewhat surprising that the cuticular pad, an important filter physically protecting the lyriform organ from high-load stimuli, in contrast to HS10, grows isometrically (same rate as body size). If the need to filter stimuli remains consistent from juvenile to adult, then it is curious that the pad should not scale hypoallometrically, like the HS10 organ, to minimise changes in its filter properties.

The metatarsal pad distal of the HS10 lyriform organ in *L. hesperus* is small relative to that found in the wandering spider *C. salei*, the subject of previous studies on this structure (McConney et al. 2007; Young et al. 2014; Erko et al. 2015). In all but adult *L. hesperus*, there is a very small area of cuticle distal to HS10. In males and females, this becomes a larger structure that resembles the ‘appendix’ area found in *Cupiennius* (Erko et al. 2015). In *Cupiennius*, the metatarsal pad acts as a high-pass filter, removing biologically irrelevant stimuli below 40 Hz (McConney et al. 2007), its material properties being complex and heterogeneous to perform this task (Young et al. 2014; Erko et al. 2015). Unlike the distal portion of the pad which is soft and compliant, the ‘appendix’ adjacent to slit 1 of HS10 is hard and sclerotised (Erko et al. 2015). Without analysis of the material properties of the

pad in adults, or indeed juvenile *L. hesperus*, it is not possible to infer whether their pad performs the same function as in *Cupiennius* or whether it has a different structure specific to the web substrate that transmits vibrational signals in *L. hesperus*. The angle of the joint on which the lyriform organ is located also affects responses to vibrational stimuli (Finck 1981). The T of *Cupiennius* has a wider range of angular motion before cuticular structures of the MT and T come into contact than that of the web-dwelling species, *Nephila clavipes* (Barth 2002; Schaber et al. 2012). *L. hesperus* and *Cupiennius* inhabit different substrate types and show considerable differences in the angle of MT-T joint superficially appearing similar to those observed in *Nephila*, which could reflect differences in pad structure and function.

The lack of any significant pad in juvenile spiders may imply that they do not require much signal filtering in order to respond to relevant stimuli, but there are also other explanations that are not mutually exclusive. As well as acting as a filter, it is thought that the pad serves to protect the HS10 organ from high-amplitude, potentially damaging stimuli (Erko et al. 2015). This function is perhaps more relevant to adults than to juveniles, as in juveniles a damaged structure will be replaced with a fully functioning new structure at the next moult. Cumulative damage to HS10 should therefore be more of a problem in adults and could be an explanation for the greater change in morphology over development of the pad relative to the lyriform organ itself.

Small spiders need to be able to detect prey and potential predators as much as far larger adult spiders, and although behaviour and the overall composition of prey and predators may differ somewhat between small spiders and large adults, there is overlap and detection of similar stimuli will be required to survive (see Uma and Weiss 2012). However, very little is known about the ecology of juvenile *L. hesperus* or indeed juvenile spiders in general. Further work should include electrophysiological measurements of spiderling lyriform organs in response to

vibrational stimuli in order to determine whether the sensitivity of the organ changes throughout the development of the animal. This would also give insight into the function of the cuticular pad through comparison of responses between juveniles and adults whose pad sizes differ far more than the size of the lyriform organ.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

Human and animal rights All applicable international, national and/or institutional guidelines for the care and use of animals were followed.

Informed consent Informed consent was obtained from all individual participants included in the study.

References

- Barth FG (1971) Der sensorische Apparat der Spaltsinnesorgane (*Cupiennius salei* Keys., Araneae). *Z Zellforsch* 112:212–246
- Barth FG (2002) A spider's world senses and behavior. Springer, Berlin
- Barth FG (2004) Spider mechanoreceptors. *Curr Opin Neurobiol* 14:415–422. doi:10.1016/j.conb.2004.07.005
- Barth FG (2012) Spider strain detection. *Frontiers in sensing*. Springer, Wien, pp 251–273
- Barth FG, Geethabali (1982) Spider vibration receptors: threshold curves of individual slits in the metatarsal lyriform organ. *J Comp Physiol A* 148:175–185. doi:10.1007/BF00619124
- Barth FG, Libera W (1970) Ein Atlas der Spaltsinnesorgane von *Cupiennius salei* Keys. Chelicerata (Araneae). *Z Morph Tiere* 68:343–369
- Barth F, Pickelmann P (1975) Lyriform slit sense organs. *J Comp Physiol* 103:39–54. doi:10.1007/BF01380043
- Barth FG, Stagl J (1976) The slit sense organs of arachnids. *Zoomorphologie* 86:1–23. doi:10.1007/BF01006710
- Barth FG, Ficker E, Federle H-U (1984) Model studies on the mechanical significance of compound spider slit sensilla. *Zoomorphologie* 104:204–215
- Elias DO (2003) Seismic signals in a courting male jumping spider (Araneae: Salticidae). *J Exp Biol* 206:4029–4039. doi:10.1242/jeb.00634
- Elias DO, Hebets EA, Hoy RR, Mason AC (2005) Seismic signals are crucial for male mating success in a visual specialist jumping spider (Araneae: Salticidae). *Anim Behav* 69:931–938. doi:10.1016/j.anbehav.2004.06.024
- Erko M, Younes-Metzler O, Rack A et al (2015) Micro- and nano-structural details of a spider's filter for substrate vibrations: relevance for low-frequency signal transmission. *J R Soc Interface* 12:20141111. doi:10.1098/rsif.2014.1111
- Finck A (1981) The lyriform organ of the orb-weaving spider *Araneus sericatus*: vibration sensitivity is altered by bending the leg. *J Acoust Soc Am* 70:70231–70233
- Hössl B, Böhm HJ, Rammerstorfer FG et al (2006) Studying the deformation of arachnid slit sensilla by a fracture mechanical approach. *J Biomech* 39:1761–1768. doi:10.1016/j.jbiomech.2005.05.031
- Hössl B, Böhm HJ, Rammerstorfer FG, Barth FG (2007) Finite element modeling of arachnid slit sensilla-I: the mechanical significance of different slit arrays. *J Comp Physiol A* 193:445–459. doi:10.1007/s00359-006-0201-y
- Hössl B, Böhm HJ, Schaber CF et al (2009) Finite element modeling of arachnid slit sensilla: II—actual lyriform organs and the face deformations of the individual slits. *J Comp Physiol A* 195:881–894. doi:10.1007/s00359-009-0467-y
- Huxley JS, Teissier G (1936) Terminology of relative growth. *Nature* 137:780–781
- Kaston BJ (1970) Comparative biology of American black widow spiders. *San Diego Soc Nat Hist* 16:33–82
- Kasumovic MM, Andrade MCB (2004) Discrimination of airborne pheromones by mate-searching male western black widow spiders (*Latrodectus hesperus*): species- and population-specific responses. *Can J Zool* 82:1027–1034. doi:10.1139/z04-081
- Klärner D, Barth FG (1982) Vibratory signals and prey capture in orb-weaving spiders. *J Comp Physiol A* 148:445–455
- Landolf MA, Barth FG (1996) Vibrations in the orb web of the spider *Nephila clavipes*: cues for discrimination and orientation. *J Comp Physiol A*. doi:10.1007/BF00192316
- Lohrey AK, Clark DL, Gordon SD, Uetz GW (2009) Antipredator responses of wolf spiders (Araneae: Lycosidae) to sensory cues representing an avian predator. *Anim Behav* 77:813–821. doi:10.1016/j.anbehav.2008.12.025
- Maklakov AA, Bilde T, Lubin Y (2003) Vibratory courtship in a web-building spider: signalling quality or stimulating the female? *Anim Behav* 66:623–630. doi:10.1006/anbe.2003.2245
- Masters WM, Markl H (1981) Signal transmission in spider orb webs. *Science* 213(4505):363–365
- McConney ME, Schaber CF, Julian MD et al (2007) Viscoelastic nanoscale properties of cuticle contribute to the high-pass properties of spider vibration receptor (*Cupiennius salei* Keys). *J R Soc Interface* 4:1135–1143. doi:10.1098/rsif.2007.1000
- Molina J, Schaber CF, Barth FG (2009) In search of differences between the two types of sensory cells innervating spider slit sensilla (*Cupiennius salei* Keys.). *J Comp Physiol A* 195:1031–1041. doi:10.1007/s00359-009-0477-9
- Pringle JWS (1955) The function of the lyriform organs of arachnids. *J Exp Biol* 32:270–278
- Ross K, Smith R (1979) Aspects of the courtship behavior of the black widow spider, *Latrodectus hesperus* (Araneae: Theridiidae), with evidence for the existence of a contact sex pheromone. *J Arachnol* 7:69–77
- Schaber CF, Gorb SN, Barth FG (2012) Force transformation in spider strain sensors: white light interferometry. *J R Soc Interface* 9:1254–1264. doi:10.1098/rsif.2011.0565
- Schüch W, Barth FG (1985) Temporal patterns in the vibratory courtship signals of the wandering spider *Cupiennius salei* keys. *Behav Ecol Sociobiol* 16:263–271. doi:10.1007/BF00310990
- Uetz GW, Boyle J, Hieber CS, Wilcox RS (2002) Antipredator benefits of group living in colonial web-building spiders: the “early warning” effect. *Anim Behav* 63:445–452. doi:10.1006/anbe.2001.1918
- Uma DB, Weiss MR (2012) Flee or fight: ontogenetic changes in the behavior of cobweb spiders in encounters with spider-hunting wasps. *Environ Entomol* 41:1474–1480. doi:10.1603/EN12126
- Vibert S, Scott C, Gries G (2014) A meal or a male: the “whispers” of black widow males do not trigger a predatory response in females. *Front Zool* 11:4. doi:10.1186/1742-9994-11-4
- Walcott C (1969) A spider's vibration receptor: its anatomy and physiology. *Integr Comp Biol* 9:133–144. doi:10.1093/icb/9.1.133
- Young SL, Chyasnachyus M, Erko M et al (2014) A spider's biological vibration filter: micromechanical characteristics of a biomaterial surface. *Acta Biomater* 10:4832–4842. doi:10.1016/j.actbio.2014.07.023