

β -Adrenergic modulation of cancer cell proliferation: available evidence and clinical perspectives

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Abstract

Purpose In this review, we aimed to present and discuss the available preclinical and epidemiological evidences regarding the modulation of cancer cell proliferation by β -adrenoceptors (β -AR), with a specific focus on the putative effects of β -blockers according to their pharmacological properties.

Methods A comprehensive review of the published literature was conducted, and the evidences concerning the involvement of β -AR in cancer as well as the possible role of β -blockers were selected and discussed.

Results The majority of reviewed studies show that: (1) All the cancer types express both β 1- and β 2-AR, with the exception of neuroblastoma only seeming to express β 2-AR; (2) adrenergic agonists are able to increase proliferation of several types of cancers; (3) the proliferative effect seems to be mediated by both β 1- and β 2-AR; (4) binding to β -AR results in a cAMP transient flux which activates two major downstream effector systems: protein kinase A and EPAC and (5) β -blockers might be putative adjuvants for cancer treatment.

Conclusions Overall, the reviewed studies show strong evidences that β -AR activation, through several intracellular

mechanisms, modulate tumor cell proliferation suggesting β -blockers can be a feasible therapeutic approach to antagonize β -adrenergic response or have a protective effect per se. This review highlight the need for intensifying the research not only on the molecular mechanisms underlying the β -adrenergic influence in cancer, but also on the implications of biased agonism of β -blockers as potential antitumor agents.

Keywords Adrenergic system · Catecholamines · Cancer · Proliferation · β -Blockers

Abbreviations

| | |
|---------------|---------------------------------------|
| SAM | Sympathoadrenomedullary |
| AD | Adrenaline |
| NA | Noradrenaline |
| ISO | Isoprenaline |
| PRO | Propranolol |
| CAS | Catecholamines |
| ATE | Atenolol |
| ICI | ICI-118,551 |
| MET | Metoprolol |
| NEB | Nebivolol |
| LAB | Labetalol |
| BUT | Butoxamine |
| SALB | Salbutamol |
| BIS | Bisoprolol |
| CAR | Carvedilol |
| TERB | Terbutraline |
| MMP | Matrix metalloproteinase |
| VEGF | Vascular endothelial growth factor |
| PKA | Protein kinase A |
| cAMP | Cyclic adenosine monophosphate |
| ERK | Extracellular signal-regulated kinase |
| NF κ B | Nuclear factor κ B |

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| | |
|------|---------------------------------------|
| AP-1 | Activator protein 1 |
| CREB | CAMP response element binding protein |
| AA | Arachidonic acid |
| GPCR | G-protein-coupled receptor |
| EGF | Epidermal growth factor |

Introduction

Cancer figures among the leading causes of death worldwide, accounting for 8.2 million deaths and 14 million new cases in 2012 (Ferlay et al. 2015). The burden of cancer is increasing in economically developed countries, and the number of new cases is expected to rise by about 70 % over the next 20 years, as a result of population aging and growth as well as of the adoption of cancer-associated lifestyle choices (Torre et al. 2015).

Over the last three decades, clinical and epidemiological studies have identified psychosocial factors including stress, chronic depression and lack of social support as risk factors for cancer progression (Moreno-Smith 2010; Spiegel 1994; Spiegel and Giese-Davis 2003). A meta-analysis by Chida et al. (2008) showed that stress-related psychosocial factors are associated with higher cancer incidence even in healthy populations (Chida et al. 2008). Others reported that stressful life experiences are related to poor cancer survival and higher mortality, despite not affecting incidence (Chida et al. 2008).

Stress response is a key mechanism for the constant adaptation to changes in social and physical environments (Goldstein 2003). Multicellular organisms cope with stress through the activation of two main systems, the hypothalamic–pituitary–adrenal axis and the sympathoadrenomedullary (SAM) system, and the release of cortisol and the catecholamines (CAs), adrenaline (AD) and noradrenaline (NA), respectively. The effects of CA are mediated through interactions with α - and β -adrenoceptors (AR) (Guimaraes and Moura 2001).

A growing number of studies suggest that stress-related persistent stimulus can result in CA overproduction, which might impact cancer prognosis and mortality (Tang et al. 2013). Over the last years, chronic stress effects on cancer progression have been focused on tumor cell proliferation, resistance to apoptosis, invasion, metastasis, angiogenesis, stroma cells microenvironment and cellular immune responses (Cole and Sood 2012; Moreno-Smith 2010). Studies addressing the link between stress-activated pathways and cancer progression suggested that CA, besides affecting the antitumor immune response (Marino and Cosentino 2013), may also display direct tumor-promoting effects in, but not limited to, breast, ovary, colorectal, esophagus, lung, prostate, nasopharynx, melanoma,

leukemia, hemangioendothelium and angiosarcoma (Tang et al. 2013).

Among therapeutic drugs acting on β -AR, β -AR antagonists, commonly known as β -blockers, are widely used to treat cardiac ailments, such as hypertension and arrhythmia and other ailments. Recently, antitumor effects involving the inhibition of multiple pro-survival pathways in tumor cells have been demonstrated for many of these drugs (Baker et al. 2011). Growing epidemiological evidences have revealed strong correlations between both progression-free and long-term survival and β -blockers usage in cancer patients (Eng et al. 2014). A meta-analysis assessing 12 studies using β -blockers in cancer patients showed a positive association with overall and disease-free survival (Choi et al. 2014). Overall, the impact on survival was more pronounced for patients submitted to surgery, meaning that the perioperative period might be an opportunity to arrest tumor progression or to promote its eradication (Choi et al. 2014). Tumor cells may express β -AR, which are associated with multiple intracellular signal transduction pathways involved in cellular replication, inflammation, angiogenesis, apoptosis/anokis, cell motility and trafficking, activation of tumor-associated viruses, DNA damage repair, cellular immune response and epithelial–mesenchymal transition (Cole and Sood 2012; Eng et al. 2014; Lutgendorf et al. 2010; Marino and Cosentino 2013). Although the involvement of β -adrenergic signaling in the progression of malignant diseases has been increasingly recognized, the underlying detailed cellular mechanisms remain elusive so far (Tang et al. 2013).

The aim of this review is to present and discuss available preclinical and clinical evidence regarding β -AR-mediated regulation of cancer cell proliferation, a crucial step in cancer development and progression, with a specific focus on the pharmacological properties and on the possible effects of β -blockers.

Physiology and pharmacology of β -AR

NA is a neurotransmitter in the central and peripheral nervous systems. AD, synthesized from NA through demethylation, is produced by chromaffin cells in the adrenal medulla and released in the bloodstream upon stimulation by the sympathetic nervous system. In the central nervous system, NA is involved in attention, arousal and vigilance, while in peripheral tissues, NA is the main transmitter of sympathetic postganglionic fibers.

NA and AD act on 7-transmembrane, G-protein-coupled receptors named “adrenoceptors” (AR), which control blood pressure, heart rate and force, airway reactivity, glucose metabolism and many central nervous system

functions. AR include α_1 , α_2 and β -AR types, each further divided into three subtypes. In particular, β -AR are expressed in heart (β_1 and a few β_2 , mediating contraction), in smooth muscle (β_2 , inducing relaxation) and in skeletal muscle (β_2 , inducing hypertrophy). β_2 -AR are possibly expressed in all normal human cell types. Usually, β_1 -AR are located close to sympathetic terminals and are targeted mainly by NA released from nerves, while β_2 -AR are often extrajunctional receptors and may be preferentially acted upon by circulating NA and AD. β_3 -AR are mainly expressed in adipose tissue, where they control lipolysis, and in skeletal muscle, where they contribute to thermogenesis. Extensive information about physiology and pharmacology of AR can be found in Perez et al. (2016) (Dianne Perez).

AR agonists and antagonists are used as therapeutics for several indications, including cardiovascular disease, asthma, benign prostatic hypertrophy and glaucoma. In particular, β -blockers are used in cardiac arrhythmias, in the secondary prevention of myocardial infarction, and as second choice antihypertensives (reviewed in López-Sendón et al. 2004).

β -AR influence proliferation of several cancer cell lines

In 1989, Schuller and Cole (1989) provided the first evidence that β -AR activation promotes the proliferation of lung adenocarcinoma cells. Indeed, they demonstrated that isoprenaline (ISO), a synthetic agonist, was able to increase the proliferation of these cells and that β -AR blockade with propranolol (PRO) reverted this effect (Schuller and Cole 1989). Afterward, several *in vitro* and *in vivo* studies have shown that both CA promote cell proliferation in different types of cancer (Bernabé et al. 2011; Lin et al. 2013; Liu et al. 2008; Wong et al. 2011).

Stimulation of β -AR change intracellular cAMP levels which, in turn, can affect cell proliferation, differentiation and quiescence (Perez-Sayans et al. 2010). However, low receptor density might indicate deficient production of cAMP, resulting in downregulation of cell growth and differentiation (Cole and Sood 2012). *In vitro* studies have shown that exogenous cAMP can inhibit or stimulate cell proliferation depending on the cell type, the oncogene controlling growth or the amount of cAMP (Perez-Sayans et al. 2010). A large number of studies have also suggested that hormones that stimulate G α s-coupled receptors and cAMP/PKA activity, such as CA, regulate a diverse array of cellular processes in cancer cell biology. In a widely range of cancer cell lines, these mediators lead to the activation of signals and proteases that are active drivers of tumor behavior such as PI3 K/Akt, Ras-ERK1/2, AP-1, Stat3, NF- κ B, and CREB, and increased expression of VEGF, IL-6, IL-8 and metalloproteases (MMP) (McCarty 2014).

Tables 1 and 2, respectively, summarize the variety of cancer cell lines from different types of tumors expressing β -AR, as the effect of adrenergic agonists upon cellular proliferation.

A study using the esophageal squamous cell carcinoma cell line, HKESC-1, demonstrated that AD, via β_1/β_2 -AR/ERK/COX-2 signaling pathways, stimulates cellular proliferation, an effect abolished by the selective blockade of both β_1 - and β_2 -AR (Liu et al. 2008). Nonetheless, the authors did not advance the mechanisms by which β -AR activation leads to ERK1/2 phosphorylation and cyclooxygenase-2 (COX-2) induction. In the same cell line, they also showed that AD was able to increase protein levels of the cell cycle regulators, CDK-4, CDK-6, cyclin D1 and cyclin E2, an effect mainly reversed by β_2 -AR blockade (Liu et al. 2008). The same authors showed that β -AR are functionally activated by EGF (epidermal growth factor) which increases HKESC-1 cellular proliferation via up-regulation of PKA. They also suggest that EGF can indirectly affect proliferation through the increase in TH (tyrosine hydroxylase) expression and subsequently AD production, which then will increase proliferation after binding to β -AR. Interesting enough, the blockade of β -AR with ATE and ICI almost completely reversed the proliferative effect of EGF (Liu et al. 2008).

Evidences have shown that there is an overexpression of β_2 -AR in human gastric cancer tissues (Shan et al. 2014). In a study using the gastric cancer cell lines, BGC-823 and SGC-7901, ISO-enhanced cellular proliferation (Liao et al. 2010). PRO was able to decrease cell proliferation in a concentration-dependent manner by reducing NF- κ B DNA binding activity and concomitantly inhibiting the expression of COX-2, MMP-2/9 and VEGF at both mRNA and protein levels (Liao et al. 2010).

Oral squamous carcinoma cell proliferation also seems to be affected by adrenergic activation. A retrospective clinical study showed that β_2 -AR expression is a favorable prognostic factor for oral squamous carcinoma patients and could be a target for new antineoplastic pharmacological strategies (Bravo-Calderon et al. 2011). Bernabé et al. (2011) showed that NA induces the proliferation of different oral squamous carcinoma cell lines (SCC-9, SCC-15 and SCC-25) through activation of both β_1 - and β_2 -AR, an action inhibited by PRO (Bernabé et al. 2011). In another study, NA was a potent mitogen for TCa8113 and ACC cell lines, an effect again abolished by PRO (Shang et al. 2009). In these cells, only β_2 -AR were expressed and, given their correlation with age, tumor size, clinical stage and with cervical lymph node metastasis, they putatively related with tumor development and clinical outcomes (Shang et al. 2009).

Zhang and colleagues (Zhang et al. 2010) demonstrated that the proliferation increase in the pancreatic cancer cells,

Table 1 β -Adrenoceptors expression in several types of cancer cells

| Type of cancer cell | Adrenoceptors | | | | References |
|--------------------------------|--------------------|--------------------|--------------------|--------------------|---|
| | β_1 | | β_2 | | |
| | mRNA | Protein | mRNA | Protein | |
| <i>Esophageal cancer</i> | | | | | |
| HKESC-1 | Yes ^{1,2} | Yes ^{1,2} | Yes ^{1,2} | Yes ^{1,2} | |
| HKESC-3 | Yes ² | – | Yes ² | – | Liu et al. (2008) ¹ |
| KYSE-150 | Yes ² | – | Yes ² | – | Liu et al. (2008) ² |
| <i>Gastric cancer</i> | | | | | |
| SGC-7901 | Yes ¹ | Yes ¹ | Yes ¹ | Yes ¹ | Liao et al. (2010) ¹ |
| BGC-823 | Yes ¹ | Yes ¹ | Yes ¹ | Yes ^{1,2} | Shi et al. (2013) ² |
| NCI-N87 | – | – | – | Yes ² | |
| MGC-803 | – | – | – | Yes ² | |
| HGC-27 | – | – | – | Yes ² | |
| <i>Oral squamous carcinoma</i> | | | | | |
| SCC9 | Yes ¹ | – | Yes ¹ | – | |
| SCC15 | Yes ¹ | – | Yes ¹ | – | Bernabé et al. (2011) ¹ |
| SCC25 | Yes ¹ | – | Yes ¹ | – | Shang et al. (2009) ² |
| TCa8113 | – | – | Yes ² | Yes ² | |
| ACC | – | – | Yes ² | No ² | |
| <i>Pancreatic cancer</i> | | | | | |
| MIA PaCa-2 | Yes ¹ | Yes ¹ | Yes ¹ | Yes ¹ | |
| BxPC-3 | Yes ¹ | Yes ¹ | Yes ¹ | Yes ¹ | Zhang et al. (2010) ¹ |
| PC-2 | Yes ² | – | Yes ² | – | Shen et al. (2008) ² |
| PC-3 | Yes ² | – | Yes ² | – | Lin et al. (2012) ³ |
| Panc-1 | – | Yes ³ | – | Yes ³ | |
| <i>Colon cancer</i> | | | | | |
| HT-29 | Yes ^{1,2} | Yes ² | Yes ^{1,2} | Yes ^{1,2} | |
| SW116 | Yes ¹ | Yes ² | Yes ^{1,2} | Yes ^{1,2} | Wu et al. (2005) ¹ |
| SW480 | Yes ¹ | – | Yes ¹ | – | Lin et al. (2013) ² |
| LS174T | Yes ² | Yes ² | Yes ² | Yes ² | |
| <i>Melanoma</i> | | | | | |
| C8161 | Yes ¹ | Yes ¹ | Yes ¹ | Yes ¹ | Yang et al. (2009) ¹ |
| 1174MEL | Yes ¹ | Yes ¹ | Yes ¹ | Yes ¹ | Moretti et al. (2013) ² |
| Me18105 | Yes ¹ | Yes ¹ | Yes ¹ | Yes ¹ | |
| A375 | Yes ² | Yes ² | Yes ² | Yes ² | |
| HS29-47 | Yes ² | Yes ² | Yes ² | Yes ² | |
| <i>Ovarian cancer</i> | | | | | |
| SKOV-3 | Yes ¹ | – | Yes ¹ | – | Lutgendorf et al. (2003) ¹ |
| EG | Yes ¹ | – | Yes ¹ | – | Rangarajan et al. (2003) ² |
| 222 | Yes ¹ | – | Yes ¹ | – | |
| <i>Breast cancer</i> | | | | | |
| MCF-7 | Yes ¹ | – | Yes ^{1,3} | Yes ³ | |
| ZR-75 | Yes ¹ | – | Yes ¹ | – | |
| MDA-MB-361 | Yes ¹ | – | Yes ¹ | – | |
| MDA-MB-435 | No ¹ | – | Yes ¹ | – | Cakir et al. (2002) ¹ |
| MDA-MB-453 | No ¹ | – | Yes ^{1,3} | Yes ³ | Pérez Piñero et al. (2012) ² |
| MDA-MB-468 | Yes ¹ | – | Yes ¹ | – | Shi et al. (2011) ³ |
| BT-474 | – | – | Yes ³ | Yes ³ | |
| MDA-MB-231 | – | – | Yes ² | – | |
| IBH-4 | – | – | Yes ² | – | |

Table 1 continued

| Type of cancer cell | Adrenoceptors | | | | References |
|------------------------|------------------|------------------|------------------|------------------|------------------------------------|
| | β_1 | | β_2 | | |
| | mRNA | Protein | mRNA | Protein | |
| IBH-6 | – | – | Yes ² | | |
| <i>Prostate cancer</i> | | | | | |
| LNCap | Yes ² | – | Yes ² | – | (Penn et al. 1996) ¹ |
| PC3 | – | Yes ¹ | – | Yes ¹ | Ramberg et al. (2008) ² |
| <i>Lung cancer</i> | | | | | |
| H322 | Yes | – | Yes | – | Schuller et al. (1999) |
| H441 | Yes | – | Yes | – | |
| <i>Nasopharyngeal</i> | | | | | |
| HONE-1 | Yes | Yes | Yes | Yes | Yang et al. (2006) |
| <i>Neuroblastoma</i> | | | | | |
| IMR-32 | – | – | Yes | – | |
| LAN-5 | – | – | Yes | – | |
| LAN-6 | – | – | Yes | – | |
| CHLA-15 | – | – | Yes | – | |
| CHLA-20 | – | – | Yes | – | |
| CHLA-90 | – | – | Yes | – | |
| SK-N-SH | – | – | Yes | – | |
| SH-EP | – | – | Yes | – | Wolter et al. (2014) |
| SK-N-Be1 | – | – | Yes | – | |
| SK-N-Be2 | – | – | Yes | – | |
| SK-N-Be(2)c | – | – | Yes | – | |
| SK-N-FI | – | – | Yes | – | |
| KELLY | – | – | Yes | – | |
| SK-N-AS | – | – | Yes | – | |
| SK-N-DZ | – | – | Yes | – | |

Expression of both β_1 - and β_2 -AR (mRNA and protein levels) in several human cancer cell lines

MIA PaCa-2 and BxPC-3, probably occurred through the activation of β_2 -AR, given that both PRO and ICI were significantly more effective than metoprolol (MET), a β_1 -AR selective antagonist (Zhang et al. 2010). Furthermore, they showed that β_2 -AR blockade suppressed proliferation by inhibition of both cAMP/PKA and Ras, which regulate activation of the MAPK pathway and transcription factors, such as NF κ B, AP-1 and CREB, as well as expression of its target genes, MMP-9, MMP-2 and VEGF. However, the β_1 -adrenergic antagonists suppressed invasion by solely inhibiting the cAMP/PKA pathway, suggesting these drugs as novel preventive and therapeutic approaches for pancreatic cancer (Zhang et al. 2010).

A wide variety of studies have assessed the effect of stress hormones, and other adrenergic agonists, upon colon tumor biology. In fact, in this context, colon cancer seems to be the most well-studied tumor. In 2005, Wu et al. (2005) demonstrated that the activation of β -AR by ISO and NA results in an increase in colon cancer cellular proliferation accompanied by the up-regulation of arachidonic

acid (AA) cascade. Wong et al. (2011) also showed that AD was able to increase HT-29 cell proliferation probably by binding to both β_1 - and β_2 -AR (Wu et al. 2005). Results from our group were also consistent with these previous reports, by showing that stress hormones and ISO were able to increase HT-29 colon cancer cell proliferation, most likely through the involvement of both β -AR (Coelho et al. 2015). Lin et al. (2013) demonstrated that AD, NA and ISO enhanced cell proliferation through β -AR-dependent pathways in three human colon cancer cell lines (HT-29, SW116 and LS174T). These findings were similar to an in vivo study where a chronic restraint stress model was used to show the effect of stress upon tumor growth (Lin et al. 2013). All together, these results strongly support the role of stress hormones in the promotion of colon cancer cell proliferation through β -AR activation.

A study of Yang et al. (2009) in melanoma tumor cell lines showed that NA can stimulate the aggressive potential of C8161, 1174MEL and Me18105 cells, not only via the promotion of cellular proliferation, but also by evolving

Table 2 Effect of β -adrenergic modulation upon common human cancers proliferation and underlying intracellular mechanisms

| Cell line | AR agonist | β -Blocker | β -Blocker + AR agonist | β -AR | Signaling pathways | Proliferation assay | References |
|-----------------------------|--|--|--|---|--|--|-----------------------|
| <i>Esophageal cancer</i> | | | | | | | |
| HKESC-1 | AD (0, 1, 10 μ M) \uparrow | ATE (100 μ M) \leftrightarrow ICI (50 μ M) \leftrightarrow | AD (10 μ M) + ATE (100 μ M) \downarrow AD (10 μ M) + ICI (50 μ M) \downarrow | β 1 and β 2 (predominantly β 2) | – | [3 H]-thymidine incorporation and MTT | Liu et al. (2008) |
| | | – | – | – | Via AR transactivation of ERK1/2/COX-2 by AD and reversed by PRO | [3 H]-thymidine incorporation | Liu et al. (2008) |
| <i>Gastric cancer</i> | | | | | | | |
| SGC-7901 | ISO (20–150 μ M) \uparrow | PRO (75–300 μ M) \downarrow | – | – | Via β -AR activation by ISO and inactivation by PRO of NF- κ B-VEGF/MMP-2/9/COX-2 | MTT | Liao et al. (2010) |
| BGC-823 | ISO (40–150 μ M) \uparrow | PRO (100–300 μ M) \downarrow | – | – | – | – | – |
| <i>Oral squamous cancer</i> | | | | | | | |
| SCC9 | NA (10 μ M) \uparrow | – | NA (10 μ M) + PRO (1 μ M) \downarrow | – | – | MTT | Bernabé et al. (2011) |
| SCC15 | – | – | – | β 2 | – | Trypan blue exclusion | Shang et al. (2009) |
| TCa8113 | NA \uparrow | PRO \downarrow | – | – | – | – | – |
| ACC | – | – | – | – | – | – | – |
| <i>Pancreatic cancer</i> | | | | | | | |
| MIA PaCa-2 | – | PRO (100; 200 μ M) \downarrow ICI (100; 200 μ M) \downarrow MET (25–200 μ M) \leftrightarrow | – | β 2 | Inhibition of cAMP/PKA and Ras pathways by PRO and ICI | MTT | Zhang et al. (2010) |
| | NA (10 μ M) \downarrow NA (1 μ M; 0.01 μ M) \leftrightarrow | – | NA (10 ⁻⁸ M) + BUT (10 ⁻⁸ M) \leftrightarrow NA (10 ⁻⁶ M) + BUT (10 ⁻⁶ M) \leftrightarrow | – | – | – | Wang et al. (2012) |
| BxPC-3 | – | PRO (100; 200 μ M) \downarrow ICI (100; 200 μ M) \downarrow MET (25–200 μ M) \leftrightarrow | – | β 2 | Inhibition of cAMP/PKA and Ras pathways by PRO and ICI | MTT | Zhang et al. (2010) |
| | NA (10 μ M) \downarrow NA (1 μ M; 0.01 μ M) \leftrightarrow | – | NA (10 ⁻⁸ M) + BUT (10 ⁻⁸ M) \leftrightarrow NA (10 ⁻⁶ M) + BUT (10 ⁻⁶ M) \leftrightarrow | – | – | – | Wang et al. (2012) |
| <i>Colon cancer</i> | | | | | | | |
| HT-29 | NA (0.1; 1; 10 μ M) \uparrow AD (0.1; 1; 10 μ M) \uparrow ISO (1, 10 μ M) \uparrow | PRO (50 μ M) \leftrightarrow ATE (50 μ M) \leftrightarrow ICI (50 μ M) \leftrightarrow | NA (10 μ M) + PRO (50 μ M) \downarrow AD (10 μ M) + PRO (50 μ M) \downarrow AD (10 μ M) + ATE (50 μ M) \downarrow AD (10 μ M) + ICI (50 μ M) \downarrow ISO (1 μ M) + ICI (50 μ M) \downarrow ISO (1 μ M) + ATE (50 μ M) \downarrow ISO (1 μ M) + PRO (50 μ M) \downarrow | β 1 and β 2 | β -AR-mediated ERK1/2 activation by agonists (AD, NA and ISO) | BrdU incorporation | Lin et al. (2013) |
| | AD (0.1; 1; 10 μ M) \uparrow | ICI (50 μ M) \downarrow ATE (100 μ M) \leftrightarrow | AD (10 μ M) + ATE (100 μ M) \downarrow AD (10 μ M) + ICI (50 μ M) \downarrow | β 1 and β 2 | Via β -AR activation of COX-2-dependent pathway by AD | [3 H]-thymidine incorporation | Wong et al. (2011) |
| | ISO (1; 10; 100 μ M) \uparrow NA (100 μ M) \uparrow | ICI (50 μ M) \downarrow ATE (100 μ M) \leftrightarrow | – | β 1 and β 2 | – | [3 H]-thymidine incorporation | Wu et al. (2005) |
| | NA (0.1; 1; 10; 100 μ M) \uparrow AD (1; 10 μ M) \uparrow ISO (1, 10 μ M) \uparrow | PRO (50 μ M) \downarrow ATE (50 μ M) \downarrow CAR (5 μ M) \leftrightarrow ICI (5 μ M) \leftrightarrow | AD (1; 10 μ M) + PRO (50 μ M) \downarrow ISO (1; 10 μ M) + PRO (50 μ M) \downarrow AD (1; 10 μ M) + ATE (50 μ M) \downarrow ISO (1; 10 μ M) + ATE (50 μ M) \downarrow AD (1; 10 μ M) + CAR (5 μ M) \downarrow ISO (1; 10 μ M) + CAR (5 μ M) \downarrow AD (1; 10 μ M) + ICI (5 μ M) \downarrow ISO (1; 10 μ M) + ICI (5 μ M) \downarrow | β 1 and β 2 | – | [3 H]-thymidine incorporation and BrdU incorporation | Coelho et al. (2015) |

Table 2 continued

| Cell line | AR agonist | β-Blocker | β-Blocker + AR agonist | β-AR | Signaling pathways | Proliferation assay | References |
|------------------------|--|---|--|-----------|--|--|--|
| SW116 | NA (0.1; 1; 10 μM) ↑ AD (0.1; 1; 10 μM) ↑ ISO (1, 10, 100 μM) ↑ | PRO (50 μM) ↔ ATE (50 μM) ↔ ICI (50 μM) ↔ | NA (10 μM) + PRO (50 μM) ↓ AD (10 μM) + PRO (50 μM) ↓ ISO (1 μM) + PRO (50 μM) ↔ ISO (1 μM) + ATE (50 μM) ↔ ISO (1 μM) + ICI (50 μM) ↓ | β1 and β2 | β-AR-mediated ERK1/2 activation | BrdU incorporation | Lin et al. (2013) |
| LS174T | NA (10 μM) ↑ AD (0.1; 1; 10 μM) ↑ | - | - | - | - | - | - |
| <i>Melanoma</i> | | | | | | | |
| C8161 | NA (10 μM) ↓ | - | - | - | - | [³ H]-thymidine incorporation | Yang et al. (2009) |
| 1174MEL | - | - | - | - | - | - | - |
| Me18105 | NA (0.1; 1; 10 μM) ↔ | - | - | - | - | - | - |
| <i>Breast cancer</i> | | | | | | | |
| IBH-4 | AD (1 nM; 0.1 μM) ↑ | PRO (1 μM) ↔ | AD (0.1 μM) + PRO (1 μM) ↑ | - | ERK1/2 inhibited phosphorylation | [³ H]-thymidine incorporation | Pérez Piñero et al. (2012) |
| IBH-6 | AD (0.1 μM) ↑ | PRO (1 μM) ↔ | AD (0.1 μM) + PRO (1 μM) ↔ | - | - | - | - |
| MDA-MB-231 | AD (1 nM) ↑ ISO (0.1; 1; 10 μM) ↔ | PRO (1 μM; 1 mM) ↔ - | AD (0.1 μM) + PRO (1 μM) ↑ - | - | - | Fluorescent DNA binding dye | Madden et al. (2011) |
| MCF-7 | ISO (0.1 μM) ↔ AD (1 nM) ↔ AD (1 μM) ↑ ISO (1 μM) ↓ | PRO (10; 25; 50 μM) ↔ - - - | - - - - | - b | - - | - [³ H]-thymidine incorporation | Wilson et al. (2015) Gargiulo et al. (2014) |
| ZR-75 | ISO (1 μM) ↔ | - | - | - | - | MTT | Shi et al. (2011) |
| MDA-MB-361 | ISO (1 μM) ↑ | - | - | β1 and β2 | - | [³ H]-thymidine incorporation | Cakir et al. (2002) |
| MDA-MB-435 | - | - | - | - | - | - | - |
| MDA-MB-453 | ISO (1 μM) ↔ | - | - | - | - | - | - |
| MDA-MB-468 | ISO (1 μM) ↔ | - | - | - | - | - | - |
| <i>Prostate cancer</i> | | | | | | | |
| LNCaP | ISO (1, 2, 5, 10 μM) ↑ ISO (1, 2, 5 μM) ↑ ISO (10 μM) ↔ ISO (1 μM) ↔ ISO (2, 5, 10 μM) ↑ ISO (1, 2, 5 μM) ↑ ISO (10 μM) ↔ | - - - - - - - | ISO (5 μM) + PRO (1 μM) ↔ ^a | β2 | Via β2-AR-activated ERK1/2 by ISO and modulated by β-arrestin2/c-Src complex formation | Trypan blue exclusion MTT | Zhang et al. (2011) |
| PC3 | ISO (1 μM) ↔ ISO (2, 5, 10 μM) ↑ ISO (1, 2, 5 μM) ↑ ISO (10 μM) ↔ ISO (1 μM) ↔ ISO (2, 5, 10 μM) ↑ ISO (1 μM) ↔ ISO (2, 5, 10 μM) ↑ | - - - - - - - - | - - - - - - - - | - | - | BrdU incorporation Trypan blue exclusion MTT | - |

Table 2 continued

| Cell line | AR agonist | β -Blocker | β -Blocker + AR agonist | β -AR | Signaling pathways | Proliferation assay | References |
|--------------------|---|-----------------------------------|--|-------------|--------------------|-----------------------------------|-------------------------|
| <i>Lung cancer</i> | | | | | | | |
| H322 | Shang et al. (2009) AD (1 μ M) \uparrow | PRO (1 μ M) \leftrightarrow | AD (1 μ M) + PRO (1 μ M) \leftrightarrow^a | - | - | [3 H]-thymidine incorporation | Al-Wadei et al. (2012a) |
| H441 | | | | | | | |

\uparrow , \downarrow and \leftrightarrow represent a significant increase, decrease or a non-effect upon cancer cell proliferation, respectively

AD adrenaline, NA noradrenaline, ISO isoprenaline, ATE atenolol, ICI/ICI-118,551, PRO propranolol, CAR carvedilol, MET metoprolol

^a Compared to the untreated group

^b The authors suggest that actions are mediated by α_2

the release of pro-angiogenic factors, such as VEGF, IL-8 and IL-6. Besides the solid data about the role of β 1- and β 2-AR on melanoma progression, recent works have shown that β 3-AR as well play a key role in this tumor. In 2014, Dal Monte et al. (2014) using B16F10 cells, a murine melanoma cell line, demonstrated that β 3-AR modulate melanoma cell proliferation and survival through nitric oxide signaling. The iNOS-produced NO acted as a downstream effector of β 3-AR proving that the beneficial effects of β 3-AR blockade on cell proliferation and apoptosis were functionally linked to reduced iNOS expression and NO production (Dal Monte et al. 2014). Studies by Chiarugi P. and colleagues (Calvani et al. 2015; Moretti et al. 2013) reported that β 3-AR expression in human melanoma is correlated with tumor aggressiveness, being up-regulated in malignant and advanced lesions when compared with melanocytic lesions. Moreover, they also showed that NA, through β 3-AR, stimulates the activation of cancer-associated macrophages, the recruitment of monocytes as well as their polarization into M2 macrophages (pro-tumorigenic type), and sustains the secretion of pro-inflammatory cytokines (Calvani et al. 2015; Moretti et al. 2013). β 3-AR are also involved in the recruitment of bone marrow-derived precursors to tumor cells and promote their differentiation into mature cancer-associated fibroblast and endothelial cells, sustaining tumor inflammation, angiogenesis and ultimately promoting melanoma malignancy (Calvani et al. 2015). These findings suggest that β 3-AR exert an extensive influence on the tumor microenvironment and open new and promising perspectives for the role of β 3-AR in cancer biology.

The paradoxical nature of AR action in breast cancer cells was reviewed by Luthy et al. (2009). In breast cancer cells, the proliferative effect of adrenergic drugs seems to be dependent on the cellular experimental model and the activated AR-subtype (Luthy et al. 2009). Cakir et al. (2002) investigated the adrenergic influence upon breast cancer cell proliferation in six different cell lines (estrogen-responsive and non-responsive). These authors showed that ISO was able to enhance proliferation of two estrogen non-responsive cell lines (MDA-MB-435 and MDA-MB-453), but did not affect any of the estrogen-responsive cell lines (MCF-7, ZR-75, MDA-MB-361). Interestingly enough, PRO was able to significantly inhibit cell proliferation, regardless the estrogen-responsiveness. In addition, ATE and ICI also inhibited the proliferation of all the above mentioned cancer cell lines, with ICI having the greater effect. These authors also suggested that AA cascade is directly triggered by β -AR activation, as MD-MB-435 cells after exposure to ISO released high levels of AA. Shi et al. (2011) showed that ISO, in a concentration-dependent manner, markedly increased the proliferation of the breast cancer cells, MCF-7. In this study, the overexpression of

Her2 increased AD release from these cells, through the activation of ERK by phosphorylation, resulting in the up-regulation of β 2-AR expression. A positive feedback loop is afterward established when, after stimulation of these receptors with different agonists (including AD), there is an increase in Her2 expression (Shi et al. 2011). Pérez Piñero et al. (2012) clearly showed that AD significantly enhanced proliferation of the human cell lines, IBH-4, IBH-6 and MDA-MB-231. Nevertheless, AD seems to increase or decrease breast cancer cell proliferation, depending on its binding to α 2 or β -AR, respectively (Pérez Piñero et al. 2012). In addition, PRO did not completely abolish AD-induced proliferation, also suggesting the involvement of α -AR. These authors also showed that both ISO and salbutamol (SALB, β 2-AR agonist) repressed cell proliferation, probably by the inhibition of ERK1/2 phosphorylation mediated by PKA, but not by EPAC. Madden et al. (2011) concluded that β -AR activation with ISO and terbutaline (TER, β 2-AR agonist) did not alter MDA-MB-231 breast cancer cell proliferation. However, these authors reported that in cells with high β -AR density, stimulation of these receptors regulates VEGF production through the classical β /AR/cAMP/PKA pathway. Gargiulo et al. (2014) showed that both AD and ISO are able to decrease cell proliferation of the non-tumorigenic cell lines, MCF-10A and HBL-100, mainly via β -AR, and to increase proliferation of the tumor cell lines, MCF-7 and MDA-231, through α 2-AR.

The majority of studies performed in lung cancer mention that the nicotine-derived nitrosamine NNK induces its development in experimental *in vitro* and *in vivo* animal models, thereby indicating a direct causative association between smoking and lung cancer incidence (Al-Wadei et al. 2012b; Hoffmann et al. 1991; Schuller 2013; Schuller et al. 1990). Interestingly enough, NNK seems to act as a high affinity agonist for both β 1- and β 2-AR leading to the development and progression of lung cancer through the activation of AA cascade and related cellular events (Schuller et al. 1999). Al-Wadei et al. (2012a) showed that AD increased NCI-H322 and NCI-H441 lung cancer cell proliferation, and PRO was able to revert this effect. Indeed, proliferation of these cells seems to be regulated by both nicotinic and β -AR (Al-Wadei et al. 2012b; Hoffmann et al. 1991; Schuller 2013; Schuller et al. 1990). The activation of nicotinic receptors by nicotine increases NA production, that, by interacting with β -AR, ultimately leads to cellular proliferation through p-ERK and p-CREB overexpression (Al-Wadei et al. 2012a). Once smoking is the most lethal risk factor associated with lung cancer cell proliferation, current literature has shed light upon the multiple molecular mechanisms by which components of tobacco smoke can initiate tumor development, induce cell cycle progression and proliferation in multiple cancer types (Al-Wadei et al. 2012b;

Hoffmann et al. 1991; Schuller 2013; Schuller et al. 1990, 1999).

In 2011, Zang et al. (2011) showed that β 2-AR activation by ISO-enhanced proliferation of the prostatic cancer cells, LNCaP and PC3, and that PRO reverted this effect. Furthermore, the scaffold protein β -arrestin2 was found to be involved in both β 2-AR-mediated activation of ERK1/2 and proliferation increase in LNCaP cells overexpressing this protein (LNCaP- β Arr2). Besides, the activation of β 2-AR in these cells leads to the formation of the complex β -arrestin2/c-Src, an effect that disappears with the inhibition of c-Src (Zhang et al. 2011).

Overall, the above studies show that adrenergic agonists mainly through β -AR are able to increase cancer cell proliferation. However, a few studies showed that these drugs have antiproliferative effects upon some cancer types. For instance, NA decreases proliferation of pancreatic (Zhang et al. 2010) and melanoma cancer cells (Yang et al. 2009) and ISO of breast cancer cells (Pérez Piñero et al. 2012). In this type of cancer, proliferation increase seems to be mainly mediated by α 2-AR (Pérez Piñero et al. 2012). On the other hand, the finding by our (Coelho et al. 2015) and other groups (Cakir et al. 2002; Coelho et al. 2015; Liao et al. 2010; Shang et al. 2009; Wang et al. 2012; Wong et al. 2011; Wu et al. 2005; Zhang et al. 2010) that β -blockers per se are able to decrease cellular proliferation suggests that they act as inverse agonists.

β -AR signaling on tumor microenvironment

Despite the clear importance of the cell proliferation on cancer progression, it is well established that tumor cells do not proliferate and progress as isolated entities. During carcinogenesis, the acquisition of malignant traits is influenced by the surrounding microenvironment (Hanahan and Weinberg 2011). Activation of β -AR through physiological or pharmacological stimuli induces a pro-metastatic gene expression signature in the tumor microenvironment (Cole and Sood 2012). These alterations remodel the primary tumor architecture and increase the possibility of cell dissemination through several ways, including: recruitment of macrophages into the tumor microenvironment (Lamkin et al. 2016), influences on immune response (Eng et al. 2014) and remodeling of blood vessels (Chakraborty et al. 2009) and lymph vessels (Le et al. 2016). Cancer-related molecular pathways can be influenced not only by β -AR expressed on tumor cells (which is the focus of the present review) but also by activation of β -AR expressed on other cell types present in the tumor microenvironment (Cole et al. 2015). For instance, macrophages play a key role in mediating inflammation, modulating the tumor microenvironment and promoting metastasis (Pimentel et al. 2012), and β -AR signaling strongly enhances macrophage

recruitment into tumor parenchyma through at least two different mechanisms: by stimulating production of chemotactic factors from tumor cells and by promoting myelopoietic development of monocyte precursors in the bone marrow and spleen (Armaiz-Pena et al. 2015; Scanzano and Cosentino 2015). In addition, in macrophages, the expression of several genes related to tumor progression such as TGF- β , VEGF, IL-6, MMP9 and PTGS2 is substantially increased in response to β -AR stimulation, suggesting a shift toward an immunosuppressive M2-like myeloid phenotype (Lamkin et al. 2016; Sloan et al. 2010). β -AR signaling is also involved in the reduction in lymphocyte proliferation, decrease in NK cell cytotoxicity and reduction in T cell response to mitogen stimulation, which strongly contribute to cancer progression (Inbar et al. 2011; Marino and Cosentino 2013).

Several authors also showed that AD and NA may upregulate the expression of important pro-angiogenic factors such as VEGF, interleukin 6 (IL-6), IL-8, MMP-2 and MMP-9 in several types of cancer cells through β -AR signaling (Chakraborty et al. 2009; Cole et al. 2015; Moretti et al. 2013; Thaker et al. 2006; Yang et al. 2009). These factors are crucial catalysts of angiogenesis, which is essential in the support of tumor growth and metastasis. Recently, Le et al. (2016) demonstrated that the activation of β -AR promotes tumor dissemination also by increasing the density of intratumoral lymph vessels in a process mediated by PGE2 and VEGFC.

Another interesting point is in the tumor microenvironment immune cells themselves may also be a source of CAs, thus possibly contributing to trigger local β -AR-dependent mechanisms involved in tumor progression. Indeed, results from non-tumor models show that the synthesis of CAs occurs in both macrophages (Flierl et al. 2007; Nguyen 2011) and T lymphocytes (Cosentino et al. 2000, 2015). No evidences of CA synthesis by these cells have been shown so far within tumor microenvironment, and the possible contribution of immune cells-derived CAs to tumor progression and the antitumor immune response deserves consideration.

β -blockers: a novel class of antitumor agents?

β -blockers, one of the most currently widely prescribed classes of drugs, represent a heterogeneous group of agents with distinct pharmacological properties (Baker et al. 2011; Poirier and Tobe 2014). These drugs differ in β -AR specificity, intrinsic sympathomimetic activity, vasodilatory effects and ability to cross the blood–brain barrier (Frishman and Saunders 2011). β -blockers are usually divided into three categories according to their β_1/β_2 -AR selectivity. Non-selective β -blockers show equal antagonistic activity at both β_1 - and β_2 -AR; selective β -blockers display higher

affinity for β_1 -AR. In addition, α/β -AR blocking agents differ from the previous categories as they are also antagonists at α -AR (Table 3).

Given the high expression of β -AR in tumor cells, and the tight relationship between stress response and cancer progression, a significant amount of epidemiological studies have emerged to clarify the association between β -blockers use and mortality (Monami et al. 2013; Thiele et al. 2015; Watkins et al. 2015). The majority of these studies showed that oncological patients, treated with β -blockers for other clinical conditions, present lower mortality rates comparing to their counterparts (Diaz et al. 2012; Johannesdottir et al. 2013b). Since β -adrenergic signaling can modulate multiple intracellular pathways underlying tumor progression and metastasis, β -blockers may be highly desirable for therapeutic intervention (Ji et al. 2012; Vaklavas et al. 2011).

Grytli et al. (2014) investigated the potential association between β -blockers use and prostate cancer-specific mortality. These authors observed a reduction in prostate cancer mortality among patients with high-risk or metastatic disease taking β -blockers, regardless the clinical characteristics at diagnosis and the use of statins or acetylsalicylic acid.

Wang et al. (2012, 2013) tested the hypothesis of β -blockers use for reducing the rates of disease progression and improving overall survival in locally advanced non-small cell lung cancer (NSCLC). This study concluded that the incidental use of β -blockers in patients with NSCLC was associated with improved distant metastasis-free survival, disease-free survival and overall survival, but not with locoregional progression-free survival, after radiotherapy. These findings were in accordance with those of previous preclinical studies, suggesting that β -blockers have specific effects on the metastatic cascade (Armaiz-Pena et al. 2009; Moreno-Smith 2010; Schuller 2010; Thaker et al. 2007).

A recent large population-based cohort study aimed to investigate whether the use of β -blockers increase survival in patients with malignant melanoma (Lemeshow et al. 2011). This study showed an association between β -blockers use and reduced mortality risk in patients diagnosed with malignant melanoma, a finding supporting the hypothesis that CA affects melanoma progression and that β -blockers may have unrecognized therapeutic applications (Colucci and Moretti 2016; Lemeshow et al. 2011).

In a cohort of patients with epithelial ovarian cancer, the potential correlation between β -blockers use and survival in woman with advanced stage disease was investigated (Diaz et al. 2012). An association between β -blockers use and progression-free and overall survival was identified, suggesting that these drugs may improve clinical outcomes in advanced epithelial ovarian carcinoma (Diaz et al. 2012).

Table 3 Pharmacological characteristics of β -blockers. *Main sources* http://www.guidetopharmacology.org/PSDP/Ki_database <http://kiddbdev.med.unc.edu/databases/kiddb.php/> Westfall (2011)

| Name | pKi (Perez et al. 2016) | MSA | ISA | VD | Lypophilicity | Main current therapeutic indications (Perez et al. 2016; Westfall 2011) | Comments | | |
|--|-------------------------|---------------|---------------|----|---------------|---|-----------------|---|---|
| | β_1 -AR | β_2 -AR | β_3 -AR | | | | | | |
| <i>β Blocking agents, non-selective</i> | | | | | | | | | |
| Alprenolol | 8.2 | 8.9 | 7.4 | 0 | + | 0 | NA | Angina pectoris | SHT1A antagonist |
| Oxprenolol | NA | NA | NA | + | + | 0 | Moderate | Angina, hypertension and cardiac arrhythmias | NA |
| Pindolol | 8.6–9.3 | 8.3–9.4 | 7.4 | + | ++ | 0 | High | Angina pectoris and hypertension | Antagonist for 5HT1A (pKi = 8.1), 5HT2B (pKi = 5.7) and 5HT2A (pKi = 5.0); full agonist for 5HT2B (pKi = 5.7) and 5HT2A (pKi = 5.0) |
| Propranolol | 8.6–10.7 | 9.2–11.0 | 6.3–7.2 | ++ | 0 | 0 | High | Hypertension and angina; supraventricular arrhythmias/tachycardias, ventricular arrhythmias/tachycardias, premature ventricular contractions, digitalis-induced tachyarrhythmias myocardial infarction, pheochromocytoma, essential tremor and the prophylaxis of migraine. From 2014, it is also approved as a treatment for infantile hemangiomas | 5HT1A antagonist (pKi = 7.5); large volume of distribution (4 L/kg) and readily enters the CNS |
| Timolol | NA | 9.7 | NA | 0 | 0 | 0 | Low to moderate | Hypertension, congestive heart failure, acute MI and migraine prophylaxis. In ophthalmology, timolol has been used in the treatment of open-angle glaucoma and intraocular hypertension | NA |
| Sotalol | 6.1 | 6.5 | NA | 0 | 0 | 0 | Low | Cardiac arrhythmias and life-threatening ventricular arrhythmias | K+ channels blocker |
| Nadolol | 6.9 | 7.0–8.6 | 6.3 | 0 | 0 | 0 | Low | Hypertension and angina pectoris | Long half-life (12–24 h) |
| Mepindolol | NA | NA | NA | NA | NA | NA | NA | NA | NA |
| Carteolol | NA | NA | NA | 0 | + | 0 | Low | Hypertension and to reduce intraocular pressure | NA |
| Tertatolol | NA | NA | 8.6 | NA | NA | NA | NA | NA | NA |
| Bopindolol | NA | NA | NA | NA | NA | NA | NA | NA | NA |
| Bupranolol | 7.3–9.0 | 8.3–9.1 | 6.8–7.3 | NA | NA | NA | NA | Hypertension and tachycardia and in the management of glaucoma | NA |
| Penbutolol | NA | NA | NA | 0 | + | NA | High | Hypertension | NA |
| Cloranolol | NA | NA | NA | NA | NA | NA | NA | NA | NA |
| <i>β Blocking agents, selective</i> | | | | | | | | | |
| Practolol | 6.1–6.8 | NA | NA | 0 | ++ | 0 | Low | Emergency treatment of cardiac arrhythmias | NA |
| Metoprolol | 7.0–7.6 | 6.3 | NA | + | 0 | 0 | High | Essential hypertension, angina pectoris, tachycardia, heart failure, vasovagal syncope, and as secondary prevention after myocardial infarction, an adjunct in treatment of hyperten-roidism and for migraine prophylaxis | NA |
| Atenolol | 6.4–7.6 | 5.1–6.0 | 5.0 | 0 | 0 | 0 | Low | Hypertension, coronary heart disease, arrhythmias and angina pectoris, and to treat or reduce the risk of heart complications following myocardial infarction. It is also used to treat Graves disease until antithyroid medication can take effect | NA |

Table 3 continued

| Name | pKi (Perez et al. 2016) | | MSA | ISA | VD | Lypophilicity | Main current therapeutic indications (Perez et al. 2016; Westfall 2011) | Comments |
|---|-------------------------|---------------|---------|-----|----|---------------|--|---|
| | β_1 -AR | β_2 -AR | | | | | | |
| Acebutolol | 6.4 | NA | + | + | 0 | Moderate | Hypertension, ventricular and atrial cardiac arrhythmias, acute myocardial infarction in high-risk patients and Smith–Magenis syndrome | NA |
| Betaxolol | 8.8 | 7.2 | + | 0 | 0 | Moderate | Treat hypertension, angina pectoris and glaucoma | NA |
| Bevantolol | NA | NA | NA | NA | NA | NA | NA | NA |
| Bisoprolol | 7.8–7.6 | 5.9–6.7 | 0 | 0 | 0 | Moderate | Hypertension | NA |
| Celiprolol | NA | NA | 0 | + | + | Moderate | Hypertension and angina | NA |
| Esmolol | 6.9 | NA | 0 | 0 | 0 | Low | Commonly used in patients during surgery to prevent or treat tachycardia and in the treatment of supraventricular tachycardia | Short acting IV injection: t1/2 of ~8 min |
| Epanolol | NA | NA | NA | NA | NA | NA | NA | NA |
| s-Atenolol | NA | NA | NA | NA | NA | NA | NA | NA |
| Nebivolol | 8.2 | 7.8 | 0 | 0 | ++ | Moderate | Hypertension | NO-mediated vasodilation |
| Talimolol | NA | NA | NA | NA | NA | NA | NA | NA |
| <i>α and β blocking agents</i> | | | | | | | | |
| Labetalol | 8.2 | 8.0 | + | + | ++ | Low | Oral form for therapy of chronic hypertension and as an intravenous formulation for use in hypertensive emergencies | α 1D-AR antagonist (pKi = 6.6); each isomer displays different relative activities; R,R isomer is about four times more potent as a β receptor antagonist than is racemic labetalol and accounts for much of the β blockade produced by the mixture of isomers |
| Carvedilol | 8.0–9.5 | 9.0–9.9 | 6.6–9.4 | ++ | 0 | ++ | Hypertension, congestive heart failure and left ventricular dysfunction following MI. | Antioxidant and anti-inflammatory effects |

ISA intrinsic sympathomimetic activity, MSA membrane-stabilizing activity, VD vasodilatory, + low, ++ moderate, +++ high, 0 absence of effect, NA not available

Nonetheless, another population-based studies cohort examined whether β -blockers affect mortality, following ovarian cancer diagnosis and found no association (Heitz et al. 2013; Johannesdottir et al. 2013a).

Powe et al. (2010) hypothesized whether patients started on, and maintained with, antihypertensive β -blocker therapy, prior to a breast cancer diagnosis, would show reduced distant metastasis compared to both non-hypertensive breast cancer patients and those treated with other antihypertensive drugs. These authors evaluated 466 women with invasive breast cancer and found that those taking β -blockers showed a significant reduction in tumor recurrence and longer disease-free interval. Furthermore, this study reported 57 % reduced risk of metastasis and 71 % reduction in breast cancer mortality after 10 years. Barron et al. (2011) evaluated 5801 women with stages I–IV breast cancer and matched those taking PRO or ATE to those not taking β -blockers. They found that PRO users were significantly less likely to present metastatic disease and had significantly lower cumulative probability of breast cancer-specific mortality, compared with nonusers. Surprisingly, there was no difference between ATE users and nonusers. These data suggest improved outcomes in patients with breast cancer under non-selective β -blockers therapy. In a very recent meta-analysis, Childers et al. (2015) showed, for the first time, that the use of β -blockers significantly reduced the risk of breast cancer death (Childers et al. 2015).

Recently, Choi et al. (2014) published a very interesting meta-analysis about the association between β -blockers and survival of cancer patients. Twelve studies published between 1993 and 2013 were included in this meta-analysis, that showed that β -blockers use was associated with prolonged survival, especially in patients with early-stage cancer who had been primarily submitted to surgery. The authors argued that β -blockers can be considered a standard approach for adjuvant therapy in various types of cancer.

The observation that β -blockers exert multiple anticancer effects and improve survival opens a novel way of research about the role of β -adrenergic signaling in cancer. Nevertheless, the majority of studies do not distinguish the affinity of the β -blockers more often associated with positive outcomes, and the signaling pathways implicated in these responses remain poorly understood.

Indeed, biased agonism may be relevant for β -blockers therapeutic use in cancer, since distinct signaling through several pathways is considered to have specific functional consequences (Galandrin and Bouvier 2006; Rajagopal et al. 2010). In fact, β -blockers are not solely antagonists for the G-protein pathways, but may independently regulate more than one pathway, behaving as partial agonists, inverse agonists or pure antagonists in each pathway (Galandrin and Bouvier 2006; Rajagopal et al. 2010).

Inverse agonists means that rather than just occupying the binding site and blocking the action of the agonists, they are associated with conformations of the receptor that turn off signaling leading to a suppression of basal receptor activity, while partial agonists can block one effector pathway but stimulate one or more alternative pathways (Baker et al. 2011; Evans et al. 2010; Luttrell et al. 2015). Actually, among β -blockers described in Table 3, the most part of them are already described as inverse agonists or partial agonists (Galandrin and Bouvier 2006; Grazia Perrone and Scilimati 2010). For example, ICI-118,551 and propranolol, which act as inverse agonists on the β 2-AR toward the adenylyl cyclase signaling pathway, were shown to be partial agonists when tested on the extracellular signal-regulated kinase (ERK) activity which display a great complexity in terms of biased agonism (Fig. 1). In this context, compounds can be agonist for the two pathways, inverse agonist for the two pathways or have opposite efficacies on each of the pathways. (Azzi et al. 2003; Baker et al. 2003; Galandrin and Bouvier 2006).

In addition, several studies, aiming to characterize the properties of clinically relevant β -blockers at β 1- and β 2-AR level, have shown that these drugs have divergent effects on $G\alpha_s$ - and β -arrestin-mediated signaling (Evans et al. 2010). For instance, β -blockers per se are able to diminish the proliferation of oral, gastric, pancreatic, colon and breast cancer cell lines (Cakir et al. 2002; Coelho et al. 2015; Liao et al. 2010; Shang et al. 2009; Wong et al. 2011; Wu et al. 2005; Zhang et al. 2010). However, the intracellular ramifications of these findings and the implications of either $G\alpha_s$ or β -arrestin-mediated signaling via β -AR in general, and specifically in cancer, are mostly unknown.

Various authors have suggested the use of β -blockers as chemotherapy adjuvants (Ji et al. 2012; Nagaraja et al. 2013). In fact, several clinical trials are currently studying the effect of β -blockers and chemotherapy agents in lung, breast and ovarian cancers (Nagaraja et al. 2013). In addition, future prospective trials are needed to confirm the retrospective findings and establish whether the timing and extent of β -blocker use impact cancer survival outcomes.

On the other hand, knowledge about the expression of β -AR according to tumor type could help to fit therapy, maximizing benefits and decreasing side effects (Nagaraja et al. 2013).

The majority of studies reviewed here show that: (1) All the cancer types express both β 1- and β 2-AR, with the possible exception of neuroblastoma which only seems to express β 2-AR; (2) adrenergic agonists are able to increase proliferation of several types of cancers; (3) the proliferative effect seems to be mediated by both β 1- and β 2-AR and (4) binding to β -AR results in a cAMP transient flux which activates two major downstream effector systems: protein kinase A (PKA) and EPAC. More recently,

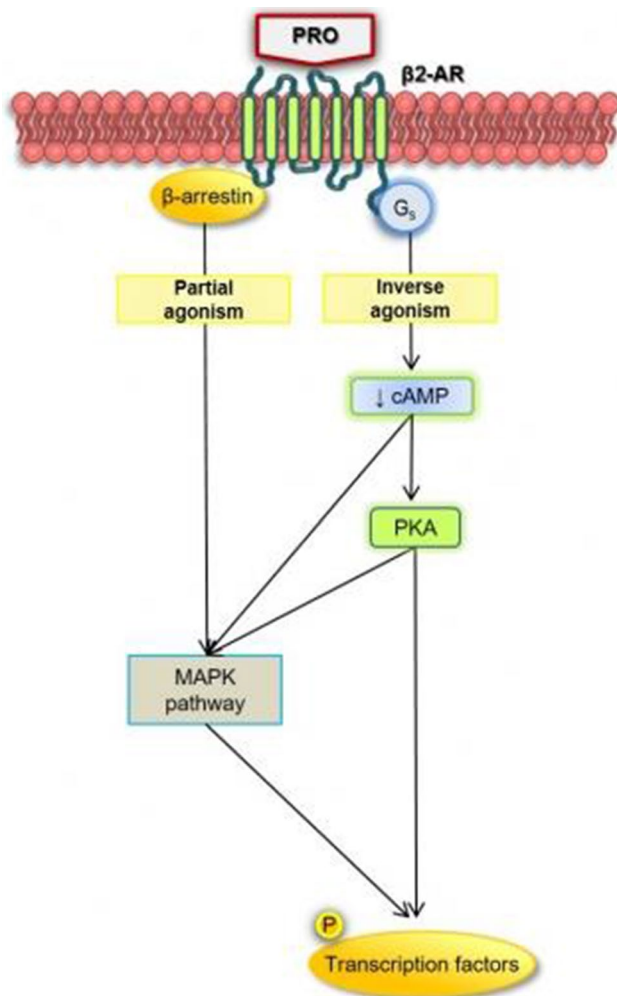


Fig. 1 Schematic representation of biased signaling of propranolol on β 2-AR. Propranolol acts on β 2-AR as a partial agonist by β -arrestin signaling activating MAPK pathway and as inverse agonist on the canonical G_s pathway decreasing the levels of cAMP. In the latter case is suggested the existence of two pathways downstream from cAMP: one involving a cAMP-dependent activation of MAPK signaling and another involving the PKA activation. Both pathways seem to increase the phosphorylation of transcription factors such as CREB. These evidences show that drugs acting on a unique receptor can have different efficacies depending on the signaling pathway activated, which add a new level of complexity in the study of β -blockers effects not only in cancer, in which these drugs have been gaining momentum, but also in other diseases. PRO—propranolol; cAMP—3′–5′-cyclic adenosine monophosphate; CREB—cAMP response element transcription factor; PKA—protein kinase A; MAPK—mitogen-activated protein kinase; β 2-AR— β 2-adrenoceptor

signaling mediated by β -arrestin is known to have distinct functional and physiological consequences from that mediated by G-proteins. “In vitro” studies exploring the underlying mechanisms involved in cellular response to CA through β -AR are lacking.

Concluding remarks

The association between stress and cancer has been described and well-studied over time, and evidences seem to support that chronic stress increases cancer progression (Cole and Sood 2012; Tang et al. 2013). In the last years, many clinical and epidemiological studies were performed in order to clarify this association (Chida et al. 2008; Lutgendorf et al. 2010; Tang et al. 2013). In the present review, we discuss the role of β -adrenergic signaling on proliferation of several human cancer types. Overall, the scrutinized studies display strong evidence that adrenergic agonists, through multiple intracellular mechanisms, enhance tumor cell proliferation and that β -blockers can be used to reverse this effect.

Unraveling the β -signaling pathways involved in cancer cell proliferation eventually will repurpose β -blockers currently in use as novel adjuvants drugs for cancer. Some open issues that should be addressed by future research are: (1) evaluation of β -AR expression on human tumor tissues which may be an useful tool to select which patients can benefit with β -blockers treatment; (2) to characterize the β -blockers more often associated with positive outcomes and which are more likely to benefit cancer patients; (3) To characterize the properties of clinically relevant β -blockers in terms of their action on $G_{\alpha s}$ and β -arrestin-mediated signaling, given that their implications in vivo are mostly unknown.

Compliance with ethical standards

Conflict of interest Author Marisa Coelho declares that she has no conflict of interest. Author Cátia Soares-Silva declares that she has no conflict of interest. Author Daniela Brandão declares that she has no conflict of interest. Author Franca Marino declares that she has no conflict of interest. Author Marco Cosentino declares that he has no conflict of interest. Author Laura Ribeiro declares that she has no conflict of interest.

Ethical approval This article does not contain any studies with human participants or animals performed by any of the authors.

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