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Morphological classifications of enteric neurons – 100 years after Dogiel

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Abstract The first differentiation of enteric neurons into three morphological types was done by the Russian histologist A. S. Dogiel on the basis of the different shapes and lengths of their dendrites. Although a number of authors considered his results during the following decades, only a division into two types withstood time: type I neurons had one long and several short processes, whereas type II neurons were characterized by several long processes. Some further structural features were discussed but substantial progress was not made until the late 1970s. This stagnation was due to some inaccuracies in Dogiel's descriptions, to the fact that most histologists in this field followed the reticular concept of the nervous system, to the idea that enteric neurons represent no more than a vegetative, postganglionic relay station between the central nervous system and the periphery, and to methodological difficulties. With the application of modern neuroanatomical techniques it was realized that the enteric nervous system contains a considerable number of neuronal subpopulations. The search for morphological correlates of the chemical diversity of enteric neurons was done mainly in the pig and the guinea-pig. In the pig, additional structural features such as axonal projection, distribution of neurons within ganglia, within different plexuses and along the length of the gut, blood supply etc. were included as criteria for further refining neuronal classification. Most of our knowledge about functional features of enteric neurons, e.g. chemical coding, neuronal connectivity, electrophysiological behaviour, was derived from studies in the guinea-pig small intestine. In light of interspecies differences, comparison of findings from different species is mandatory. The search for morphological and functional peculiarities of human enteric neuronal circuitry has to consider all

methodological and conceptual advances made within the past 100 years since the pioneering work of Dogiel.

Key words Enteric nervous system · Morphology · Innervation · Intestine · Neuron

Introduction

In the late 19th and early 20th century, some fundamental principles of the autonomous gastrointestinal functions were realized (Bayliss and Starling 1899; Trendelenburg 1917). The term “gastroenterology”, which was coined by the German physician Hemmeter – one of the founders of an American gastroenterological society in 1897 (Martini 1996), entered general usage (Chen and Chen 1995). Langley (1900, 1921) introduced the name “enteric nervous system” (ENS) to characterize the peculiarity of the neuronal elements within the gut wall. Some years before, based on the works of Ramón y Cajal and formulated by Waldeyer (1891), the neuron doctrine (“Neuronenlehre”) was founded in contrast to the reticular concept of the arrangement of the nervous tissue (reviewed Kirsche 1984; Peters et al. 1991; Clarke and O'Malley 1996). The dispute between the proponents of the two rival theories would remarkably influence the recognition of the enteric neuroarchitecture for more than half a century.

Criteria for classifications since Dogiel

In 1899, Dogiel (Fig. 1) published his last work dealing with the histological differentiation of enteric neuron populations. His attempts were the first to suggest a correlation between structural and functional features of enteric neurons. Although he favoured the reticular concept, his classification based on the possibility of distinguishing between neuronal processes as dendrites (“Protoplasmafortsätze”) and axons (“Axencylinderfortsätze”, “Nervenfortsätze”). As dendrites he regarded processes

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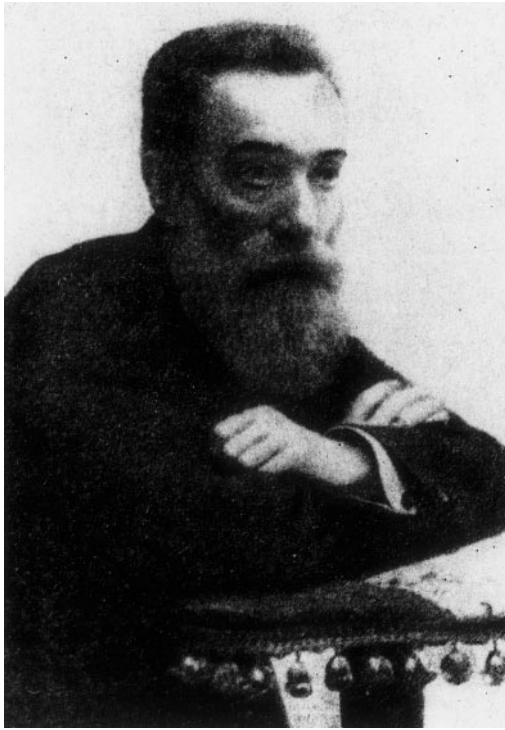


Fig. 1 Aleksandr Stanislavovic Dogiel (1852–1922; from Schierhorn 1981, with permission)

that ramify within the ganglion of origin whereas axons do not (Dogiel 1895b, 1896, 1899), although he reported on dendrites emanating from the axon as well as collaterals of axons (Dogiel 1895a). Dogiel's main criteria for distinguishing different neuron types were the lengths and shapes of dendrites. Furthermore, he mentioned sizes of nerve cell bodies, their locations within ganglia and the positions of nuclei. Very important for his functional conclusions (Table 1) were the observations that in some

although rare instances (“einer oder der andere von ihnen”), axons of myenteric type I neurons could be followed into fibres enmeshing bundles of the external musculature and, that dendritic branches of type II neurons could be occasionally traced to the mucosal plexus. Axons of myenteric type III neurons could be followed over considerable distances through other ganglia without leaving the plexus. No functional suggestion about this type was made by Dogiel. However, his concept was that different shapes of somata and dendrites correlate with different functions, one link being the projection of the axon.

The following decades saw less striking progress in this field. This was partly due to some vagueness in Dogiel's original descriptions concerning the distinctions between the dendritic tree patterns of the three types. Prior to Dogiel, Ramón y Cajal (1893) suggested that enteric neurons have several long processes that cannot be distinguished into axons and dendrites. A number of later authors gave up Dogiel's distinction between dendrites and axons in type II neurons (La Villa 1898; Ramón y Cajal 1911; Müller E. 1921; van Esveld 1928; Oshima 1929; Harting 1931; Sokolowa 1931; Stöhr 1931; Okamura 1934) whereas others maintained this distinction (Koelliker 1896; Hill 1927; Kolossow and Sabussow 1928; Rossi 1929; Iwanowa 1958). A further dispute arose as to the general possibility and functional significance of a morphological classification of enteric neurons at all. This was denied by researchers until the recent past (Kuntz 1913, 1923; Johnson 1925; Michail and Karamanlidis 1967; Christensen 1988). The most important argument for the proponents of the possibility of a morphological classification became the presence of *several long processes* in type II neurons and of one long and *several short processes* in other neurons. The validity of this distinction was sufficient for many authors (Hill 1927; Tiegs 1927; van Esveld 1928; Oshima 1929; Iwanow 1930; Harting 1931; Lawrentjew 1931;

Table 1 Some important features of the three enteric neuron types as described by Dogiel (1896, 1899)

	Types and numbers of processes	Dendrites	Axonal course	Suggested function
I	4–20 Dendrites 1 Axon	Branch and end within the ganglion of origin Short, thick, flat, with varicosities, lamellar	Through neighbouring ganglia With collaterals Sporadically to the musculature	Motor
II	1–16 Dendrites 1 Axon	Leave ganglion of origin Structural resemblance to axons Ramify into long, thin, smoothly contoured branches Much longer than type I dendrites Some dendritic branches run to the submucosa and mucosa	To other ganglia	Sensor (secretomotor ?)
III	2–10 (and more) Dendrites 1 Axon	Ramify and end within ganglion of origin Longer than type I dendrites Endings of branches: tapering (“dünner und dünner”)	Through other ganglia Traceable over considerable distances	?

Kolossow and Sabussow 1932; Reiser 1932; Murat 1933; Cavazzana and Borsetto 1948; de Biscop 1949; Jabonero 1951, 1958, 1960; Stöhr 1949, 1952; Greving 1951b; Temesrékási 1955; Kolossow and Milochin 1963) and resulted in “collapsing of the classifications” (Furness and Costa 1987), which furthermore frequently referred to only two types of neurons. The characteristic flat and lamellar shape of the type I dendrites – “lamellenförmig” (Dogiel 1899; “Dendritenlamellen”; Lawrentjew 1929) became a facultative feature for the differentiation of type I neurons. Based on the shapes and lengths of their short processes, occasional subdivisions of the type I neurons had been proposed, but these merely underlined the general grouping into two types of enteric neurons (Hill 1927; Lawrentjew 1931; Greving 1951a, b; Temesrékási 1955; Jabonero 1958, 1960).

Further reasons for the reduction of Dogiel’s original classification were manifold. Many of the authors cited above were followers of the reticular concept of the nervous system. In an extreme view, the neuronal perikarya were regarded as cytoplasmatic and nucleated junctions of a complicate neural syncytium extending between the central nervous system and the target tissues. Consequently, some authors avoided the terms “dendrite” and “axon” since they would anyhow communicate with each other and described them rather as long and short “processes” (Stöhr 1931; Reiser 1932). Others restricted their results to the “proof” of anastomoses between dendrites of different neurons as demonstrated by drawings (Cole 1930).

In another view, the nerve cells within the gut were regarded as simple postganglionic neurons of sympathetic and parasympathetic preganglionic nerves. Important arguments for this reduction were deduced from the works of E. Müller (1921), Lawrentjew (1929, 1931), Sokolowa (1931) as well as Iljina and Lawrentjew (1932), who reported gradients of the distributions of type I and II neurons along the gastrointestinal tract and related these gradients to the segmental levels of parasympathetic and sympathetic preganglionic neurons, respectively. This was exaggerated by Botár et al. (1942), who moreover denied that structural differences between enteric neurons imply functional differences and that neurons in the gut establish local reflex arches as was suggested by Dogiel. They obviously gave up the concept of the “enteric nervous system” as defined by Langley.

Furthermore, methodological factors influenced the progress in this field. In contrast to Dogiel, most later authors did not use whole-mount specimens, but sections. Some researchers recognized that the classical histological methods used (methylene blue technique, metal impregnations) that stain only subsets of neurons, may not demonstrate equivalent neuron populations in different species (Ottaviani 1940). The suggestion was made that unstained neurons may be functionally different from stained neurons (Schofield 1962).

Nevertheless, some new aspects were introduced during the decades between Dogiel and the 1970s. L.R.

Müller (1912) distinguished two myenteric neuron types based on the topographical vicinity of their processes to the musculature. Besides Langley (1900) and in contrast to Dogiel, he was one of the first to suggest that ganglion cells in the gut are quite different from other autonomic neurons. He already recognized the absence of pericellular sheaths around enteric neurons, a feature that has been confirmed more recently by fine structural studies. Kuntz (1913, 1923) described terminations of nerve fibres (“synapses”) on enteric neurons, thus favouring the neuron theory. Iwanow (1930) found axons of type I neurons to be oriented in one direction and Temesrékási (1955) estimated lengths of axonal projections within the different ganglionated plexuses. Ito (1936) as well as Ito and Nagahiro (1937) differentiated three classes of neurons based on their size, and found that the excentricity of nuclei is a frequent non-pathological feature of enteric nerve cells. Small neurons were observed that did not belong to the types of Dogiel (Stöhr 1949, 1952; Gunn 1959, 1968), whereas Fehér and Vajda (1972) distinguished three types purely on the basis of cell-soma size. Based on their staining intensity, enteric neurons were classified by Honjin et al. (1959), Michail and Karamanlidis (1967) and Sutherland (1967) into argentophobe and argentophile neurons. Schofield (1962) considered the number of processes to be relevant for functional properties, not their lengths and shapes.

In the late 1970s, the development of new techniques, first of all immunohistochemistry, led to a dramatic increase of our knowledge of the diversity of enteric neurons (Furness et al. 1991). However, important reviews of that period neither mentioned any light-microscopical classification of enteric neurons (Furness and Costa 1980), nor did they refer to Dogiel’s types (Gershon and Erde 1981).

In the 1980s, Stach (1980, 1981, 1982a, b, 1985, 1989) introduced a morphological classification in the pig small intestinal ENS. He took advantage of the eclectic nature of silver impregnation. This allows the undisturbed observation of some of the neurons with their completely stained processes since a number of other neurons – glia cells and surrounding tissues – remain scarcely or not at all represented. The directions of axonal projections, which have direct functional relevance, are thus traceable over considerable distances whereas axonal terminals remain unstained in most cases. For this purpose, intracellular dye injection techniques are more suitable (Bornstein et al. 1991). This classification is far from being finished: the definition of further criteria to distinguish, e.g., different subpopulations of type III neurons, is the focus of ongoing work. The six types hitherto described make up at least 20% of the whole myenteric population (Brehmer and Stach 1998). It uses all structural features that can be observed in silver-impregnated whole mounts: dendritic architecture, direction of axonal projection, location of cell bodies within ganglia, within the different ganglionated plexuses and along the length of the small intestine, and position of the nucleus. Some of these features had been considered already by earlier

investigators but had almost never been incorporated into a consistent concept. Furthermore, the old and inappropriate term “multipolar” was replaced by “multiaxonal” (for type II neurons) and “multidendritic, uniaxonal” (for the other classes of neurons possessing one axon). In this classification, a strong correlation between dendritic architecture and axonal projection in the three neuron types of Dogiel is emphasized (I, oral; II, circumferential and vertical; III, aboral). Stach (1982b) was compelled to introduce a fourth type when he observed neurons with a characteristic dendritic tree pattern combined with an exclusively vertical axonal projection. Combinations of silver impregnation with a vascular injection technique revealed a specific blood supply of some morphologically characterized neuron populations – those division still based on the reduced classification into two neuron types (Stach 1977, 1979). The preferentially vascularized “type I/2” neurons (Stach 1979) were later recognized as being type III neurons (Stach 1982a).

At the same time, numerous attempts were made to demonstrate the shapes of enteric neurons simultaneously with their chemical and/or electrophysiological features (Hodgkiss and Lees 1980, 1983; Costa et al. 1982; Furness et al. 1983, 1985; Bornstein et al. 1984; Kobayashi et al. 1984; 1985; Erde et al. 1985; Katayama et al. 1986). The realization that in the ENS there exists a great variety of chemically coded neuron types (reviewed in Furness and Costa 1980; Llewellyn-Smith et al. 1983; Furness and Costa 1987) triggered the search for morphological correlates of the distinct neurochemical types, particularly in the guinea-pig. Furness et al. (1988a) used an intracellular dye injection technique for the demonstration of cellular morphology. A correlation between dendritic architecture and axonal projection could not be established due to incomplete filling of most of the longer processes. Thus, their criteria for describing four types of neurons remained restricted to shapes and lengths of (short) processes and to soma sizes. All these studies yielded excellent and important findings extending our knowledge about functional properties of single enteric neurons and the ENS as a whole but did not result in a more appropriate morphological classification.

Conventional ultrastructural studies revealed different types of synaptic vesicles in enteric neurons (Baumgarten et al. 1970; Gabella 1972; Wilson et al. 1981a, b) and up to nine different neuron types according to fine structural criteria (Cook and Burnstock 1976), but it was hardly possible to reconcile them with any of the cell types recognized by light microscopy.

Neuron types

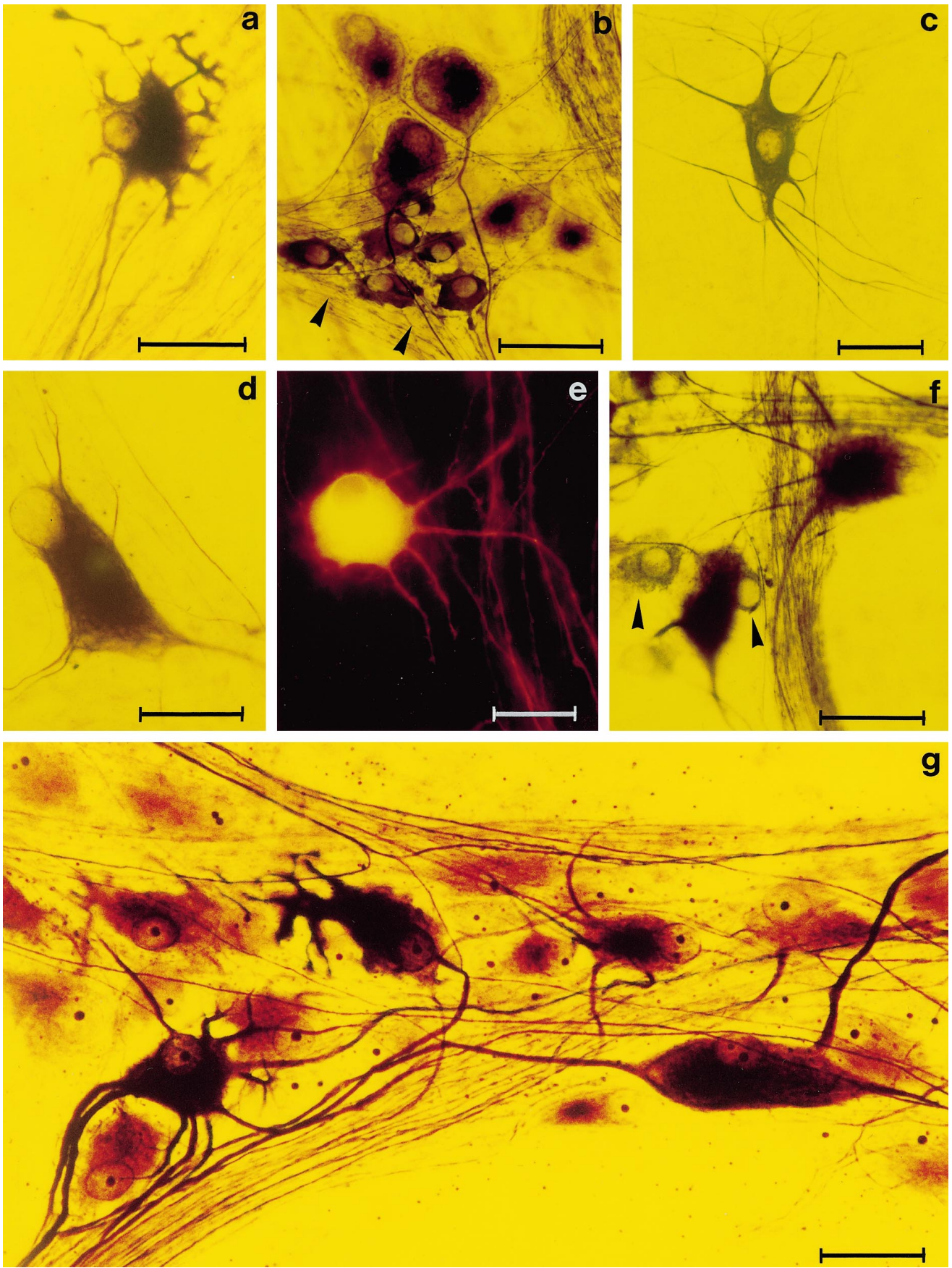
In the following section enteric neuron types will be described from a morphological point of view as they are discussed in the recent literature. Since most advance was made in type II neurons, we will start with this type. Figure 2 (a-d, g) displays the neuron types I to IV as seen in silver impregnated material. A further character-

ization using a tracing technique (Fig. 2e) and a combination of silver impregnation with a histochemical reaction (Fig. 2f) led to the unequivocal distinction of type IV neurons of Stach in contrast to types I to III of Dogiel. Thus, the need for an extended classification beyond Dogiels concept is illustrated.

Dogiel type II (Fig. 2b, g)

The most striking advance to correlate structural and functional features of enteric neurons was achieved in type II neurons. Regardless of interspecies differences and different methodological approaches, the disagreement between researchers may be the lowest in this type. These neurons have smoothly contoured outlines and several long processes that can be regarded structurally (Stach 1981: “multiaxonal”) and functionally (Hendricks et al. 1990; Bornstein et al. 1994) as axons. Type II cells occur in two forms, an adendritic and a dendritic form (Stach 1981, 1989; Furness et al. 1988a, 1990; Bornstein et al. 1991). The adendritic form has similar projection patterns in pig, guinea-pig and, probably, rat small intestinal myenteric plexus: one axon extends typically into the mucosa whereas others run circumferentially within secondary strands of the myenteric plexus (Stach 1981; Furness et al. 1990; Mann et al. 1997; Brehmer et al. 1999). The myenteric dendritic type II neurons have, in the guinea-pig small intestine, long aboral projections (Brookes et al. 1995). In this species and in contrast to uniaxonal neurons, a majority of type II neurons shows the electrophysiological after-hyperpolarization (AH) phenomenon following their action potentials (Hirst et al. 1974; Hodgkiss and Lees 1983; Bornstein et al. 1984; Erde et al. 1985; Katayama et al. 1986; Iyer et al. 1988; Tamura 1992, 1997; Clerc et al. 1998). However, in human and pig, this phenomenon seems to be not as widely present as in the guinea-pig (Brookes et al. 1987; Cornelissen et al. 1996). Using immunohistochemistry, a number of peptides have been shown within these neurons (Scheuermann et al. 1987; Furness et al. 1988b; Bornstein et al. 1989; Timmermans et al. 1989, 1992a; Song et al. 1991; 1994; Neunlist and Schemann 1997). Calbindin (in guinea-pig) and calcitonin gene-related peptide (in pig) were used as markers in ultrastructural studies of myenteric type II neurons (Pompolo and Fur-

Fig. 2a–c Silver-impregnated, myenteric neuron types I, II, III from pig small intestine, whose shapes correspond most closely to the original descriptions given by Dogiel. In **b** 6 small NADPHdiaphorase-positive neurons are visible that are not silverimpregnated (*blue-stained, arrowheads*). **a, c** Silverimpregnation. **b** NADPH diaphorase reaction and subsequent silver impregnation. **d–f** Myenteric Stach type IV neurons. **d** Silverimpregnation, **e** post mortem DiI-tracing from a single mucosal villus, **f** NADPHdiaphorase reaction and subsequent silver impregnation (*arrowheads* NADPHdiaphorase-stained neurons). **g** Myenteric ganglion from pig jejunum with four strongly impregnated (*black-brown stained*) neurons; from left to right: type III, I, IV, II. (Silver impregnation). Bars 50 µm



ness 1988; Scheuermann et al. 1991). These neurons are cholinergic in the guinea-pig (Li and Furness 1998) and, throughout all species investigated, non-nitroergic (Costa et al. 1992b, 1996; Ward et al. 1992; Aimi et al. 1993; Furness et al. 1994; Timmermans et al. 1994 a, b; Brehmer and Stach 1997). Calbindin-immunoreactive terminals, arising exclusively from type II cells, can be detected forming synapses on calbindin-positive nerve-cell bodies in guinea-pig myenteric plexus (Pompolo and Furness 1988; Furness et al. 1990). Based on work in this species, type II neurons are regarded as intrinsic primary afferent neurons (Kunze et al. 1995; Furness et al. 1998) that establish self-reinforcing networks. In an alternative view, they represent enteric interneurons (Wood 1994).

Dogiel type I (Fig. 2a, g)

Besides classical histological methods (see above), these multidendritic uniaxonal neurons were frequently diagnosed using more modern techniques, including histochemistry, immunohistochemistry, dye injection and tracing methods. Recently, their original structural feature, the shapes of the endings of the short processes ("lamellar", "flat", "stubby", "club-shaped") became a more obligatory criterium. Attempts to reconstruct three-dimensionally the typical lamellar dendrites of myenteric type I neurons of the guinea-pig were undertaken by Pompolo and Furness (1990) based on thin sections, as well as by Hanani et al. (1998) and Meedeniya et al. (1998) using confocal microscopy. In the pig small intestine, the occurrence of silver-impregnated type I neurons is restricted to the myenteric plexus. Here, they are located mainly in the oral parts of ganglia and the great majority (about 85%) projects orally in the plane of the plexus (Stach 1980). These neurons are, in the pig small intestine, non-nitroergic (Brehmer and Stach 1997) whereas in other species (guinea-pig, rat, dog, human) nitroergic type I neurons were identified (Costa et al. 1992b; Ward et al. 1992; Young et al. 1992, 1995; Bogers et al. 1994; Cracco and Filogamo 1994; Furness et al. 1994). At present, and in contrast to the type II neurons, it is impossible to assign a common functional role to them in the guinea-pig. A number of functional classes of neurons are regarded to be of Dogiel type I-morphology in this species: orally and aborally projecting interneurons, inhibitory and excitatory motoneurons for circular and longitudinal musculature (Brookes and Costa 1990; Pompolo and Furness 1990; Brookes et al. 1991, 1992, 1997; Steele et al. 1991; Costa et al. 1992a; Watchow et al. 1995; Young and Furness 1995; Young et al. 1995; Portbury et al. 1996; Song et al. 1996; Clerc et al. 1998). This may be due to the existence of subtypes bearing the common morphological feature of a characteristic dendritic shape. On the other hand, the demonstration of dendritic architecture depends extremely on methodology. The nicotinamide-adenine-dinucleotide-phosphate-diaphorase (NADPHd) reaction used as a marker for ni-

trergic neurons may serve as an example. NADPHd-positive neurons were classified as Dogiel type I (Aimi et al. 1993; Cracco and Filogamo 1994). However, caution is recommended when classifying neuron types based on this histochemical reaction which may represent dendritic shapes incompletely (Brehmer and Stach 1997).

Dogiel type III (Fig. 2c, g)

These most common "multipolar" enteric neurons have, after Dogiel's (1899) description, dendrites that branch and end within the ganglion of origin (in contrast to the processes of type II neurons he classified as dendrites). They become thinner towards their endings ("tapering"). Morphologically, in the pig small intestine, Stach (1982a) separated them clearly from type I neurons because of striking differences in dendritic tree pattern (long-dendritic, tapering) and main axonal projections (about 85% aborally). However, there seem to exist subpopulations in this species that can be distinguished by their axonal projections and/or their chemistry, e.g. nitroergic and non-nitroergic ones (Timmermans et al. 1992b, 1993; Brehmer and Stach 1997), but not yet by their dendritic tree patterns. At least some type III neurons in the pig small intestine may be regarded as intrinsic or intestinofugal interneurons (Timmermans et al. 1992b, 1993). In the guinea-pig small intestine, besides submucosal nerve cells (Bornstein et al. 1986), myenteric neurons projecting to the mucosa were classified as type III (Furness et al. 1985). Based on the drawings provided and on later descriptions (Portbury et al. 1995), neither the submucosal nor the myenteric cells correspond to Dogiel's original delineation of type III neurons. Their dendrites are scarcely longer than those of type I neurons, and Dogiel did not describe short dendritic myenteric neurons projecting vertically. The guinea-pig myenteric type III neurons (Furness et al. 1985) more closely resemble pig myenteric type IV neurons (Stach 1982b), which also project to the mucosa (Brehmer et al. 1999). In the guinea-pig small intestine, a novel class of neurons was termed filamentous neurons.

Filamentous neurons

These have "numerous relatively short (usually less than 50 μm), tapering, and sometimes branching processes, and their single long process" (Furness et al. 1988a). They can be found also in guinea-pig large intestine (Messenger et al. 1994) and are regarded as descending interneurons (Portbury et al. 1995; Clerc et al. 1998).

Type IV neurons (Fig. 2 d-g)

These are found in the pig small intestine and have short, scarcely branched dendrites emanating asymmetrically from the soma. Typically, their nucleus is eccentrically

placed within the cell body (Stach 1982b). Using morphometry, it could be shown that the peculiar dendritic tree pattern of type IV neurons is clearly separable from that of, at first glance structurally similar, single type V neurons (Brehmer and Beleites 1996). Type IV neurons are non-nitrergic (Brehmer and Stach 1997), they project from the myenteric plexus to the mucosa (Brehmer et al. 1999) and may represent secretomotor neurons. External submucosal type IV neurons project similarly vertically but mainly to the myenteric plexus (Stach 1983; Brehmer et al. 1998).

Type V neurons

These make up a characteristic non-nitrergic population within myenteric and external submucosal ganglia mainly in the pig lower small intestine (Stach 1985, 1989; Brehmer and Stach 1997; 1998; Brehmer et al. 1998). They appear as single cells or as very conspicuous aggregates. Within the latter, the compact clustering of dendrites suggests the possibility of a simultaneous activation of different type V neurons. In the myenteric plexus, these neurons project mainly aborally whereas the course of external submucosal type V axons is not known.

Type VI neurons

These are found in the pig small intestine (Stach 1989) and are present in myenteric as well as external submucosal ganglia and display a nitrergic phenotype (Timmermans et al. 1994b; Brehmer and Stach 1997). Their frequency along the small intestine is highest in the ileum (Brehmer and Stach 1998). The morphological hallmark of type VI neurons are dendrites arising from the axon hillock and the proximal axon ("axonal dendrites"). The myenteric type VI neurons project mainly aborally within the plane of the plexus whereas the submucosal type VI neurons project frequently vertically, crossing circular muscle bundles and running into the myenteric plexus (Stach 1983; Brehmer et al. 1998).

Small neurons (Fig. 2 b, f)

Besides larger neuron types that can be distinguished from each other more or less clearly by shape criteria, there seem to exist many smaller neurons that may differ in function from the larger ones in guinea-pig and pig. They were named "small neurons with few processes" (Furness et al. 1988a), "minineurons" (Stach 1989) or "simple neurons" (Messenger et al. 1994) and are probably functionally heterogeneous. As compared with the filamentous neurons of guinea-pig myenteric plexus (see above), significantly smaller neurons with filamentous dendrites send their axons to the tertiary component of the myenteric plexus and to the circular musculature. They are regarded as longitudinal and circular muscle

motor neurons (Clerc et al. 1998). Others, e.g. in the pig internal submucosal plexus, may represent mucosal effector neurons (Timmermans et al. 1997; Brehmer et al. 1999).

From morphé to morphology

From the above examples, it is obvious in guinea-pig and pig that significant species differences are an important factor impeding a thorough understanding of the ENS. To remedy this, consistent criteria for a morphological classification may be helpful. Besides guinea-pig and pig, there exists a considerable amount of data about neurochemical diversity and axonal projections of enteric neurons in the rat (Sundler et al. 1993) although information on a morphological classification in this species is scarcely available (Browning and Lees 1994). Some authors observed neurons even in other species (opossum, Christensen 1988; guinea-pig, Lees et al. 1992, Nichols et al. 1992; horse, Pearson 1994) resembling Stach type IV or VI neurons in the pig. Since these authors did not use all the criteria for identification that were applied by Stach, it is not clear whether these neurons are equivalent to the respective neuron types in the pig small intestine. Otherwise, the discrepancy in naming structurally similar neuron populations as type IV in the pig (Stach 1982b) and type III in the guinea-pig (Furness et al. 1985) indicates the possibility of a further divergence of the currently used morphological nomenclature of enteric neurons.

The diversity of chemically defined and functionally differing subtypes of enteric neurons established in guinea-pig, rat and pig (Furness and Costa 1987; Sundler et al. 1993; Costa et al. 1996; Timmermans et al. 1997) and the attempts to transfer concepts from these species to the human gut require a sound basis of categorizing enteric neurons. Recently, human enteric neurons were morphologically described and classified using different methods (Timmermans et al. 1994a; Wattachow et al. 1995, 1997; Porter et al. 1996; Krammer et al. 1998). Besides the variety of modern methods, silver impregnation should hold its place as an efficient technique for an accurate representation of the shapes of large numbers of neurons simultaneously within a single specimen. Even in brain research, there is a "definite renaissance" (Jones 1984) of silver-impregnation techniques for, e.g. the classification of cortical neurons. That the shape of a neuron "provides the key to its role in the nervous system" (Peters et al. 1991) may be true also for the ENS. The classification scheme proposed by Stach (1989) offers criteria surpassing pure description of shapes (from morphé ...), thus achieving a more valuable framework for categorization of typical and significant rather than individual and variable structural features (... to morphology). It is our belief that this classification represents an important step to bridging the gap between the basic morphological types of Dogiel and the multiplicity of neurochemical and functional types recognized today.

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References

- Aimi Y, Kimura H, Kinoshita T, Minami Y, Fujimura M, Vincent SR (1993) Histochemical localization of nitric oxide synthase in rat enteric nervous system. *Neuroscience* 53:553–560
- Baumgarten HG, Holstein A-F, Owman C (1970) Auerbach's plexus of mammals and man: Electron microscopic identification of three different types of neuronal processes in myenteric ganglia of the large intestine from rhesus monkeys, guinea-pigs and man. *Z Zellforsch* 106:376–397
- Bayliss WM, Starling EH (1899) The movements and innervation of the small intestine. *J Physiol (Lond)* 24:99–143
- de Biscop G (1949) Über pathologische Veränderungen am Auerbachschen Plexus bei Megacolon. *Z Zellforsch* 34: 141–159
- Bogers JJ, Timmermans J-P, Scheuermann DW, Pelckmans PA, Mayer B, Marck EA van (1994) Localization of nitric oxide synthase in enteric neurons of the porcine and human ileocaecal junction. *Ann Anat* 176:131–135
- Bornstein JC, Costa M, Furness JB, Lees GM (1984) Electrophysiology and enkephalin immunoreactivity of identified myenteric plexus neurones of guinea-pig small intestine. *J Physiol (Lond)* 351:313–325
- Bornstein JC, Costa M, Furness JB (1986) Synaptic inputs to immunohistochemically identified neurones in the submucous plexus of the guinea-pig small intestine. *J Physiol (Lond)* 381:465–482
- Bornstein JC, Furness JB, Costa M (1989) An electrophysiological comparison of substance P-immunoreactive neurons with other neurons in the guinea-pig submucous plexus. *J Auton Nerv Syst* 26:113–120
- Bornstein JC, Hendriks R, Furness JB, Trussell DC (1991) Ramifications of the axons of AH-neurons injected with the intracellular marker biocytin in the myenteric plexus of the guinea pig small intestine. *J Comp Neurol* 314:437–451
- Bornstein JC, Furness JB, Kunze WAA (1994) Electrophysiological characterization of myenteric neurons: how do classification schemes relate? *J Auton Nerv Syst* 48:1–15
- Botár J, Battancs L, Becker A (1942) Die Nervenzellen des Dünndarms. *Anat Anz* 93: 138–149
- Brehmer A, Beleites B (1996) Myenteric neurons with different projections have different dendritic tree patterns: a morphometric study in the pig ileum. *J Auton Nerv Syst* 61:43–50
- Brehmer A, Stach W (1997) Morphological classification of NADPHd-positive and -negative myenteric neurons in the porcine small intestine. *Cell Tissue Res* 287:127–134
- Brehmer A, Stach W (1998) Regional structural differences in the neuronal composition of myenteric ganglia along the pig small intestine. *Anat Rec* 250:109–116
- Brehmer A, Stach W, Krammer HJ, Neuhuber W (1998) Distribution, morphology and projections of nitrergic and non-nitrergic submucosal neurons in the pig small intestine. *Histochem Cell Biol* 109:87–94
- Brehmer A, Schrödl F, Neuhuber W, Hens J, Timmermans J-P (1999) Comparison of enteric neuronal morphology as demonstrated by DiI-tracing under different tissue handling conditions. *Anat Embryol* 199:57–62
- Brookes SJH, Costa M (1990) Identification of enteric motor neurones which innervate the circular muscle of the guinea pig small intestine. *Neurosci Lett* 118:227–230
- Brookes SJH, Ewart WR, Wingate DL (1987) Intracellular recordings from myenteric neurones in the human colon. *J Physiol (Lond)* 390:305–318
- Brookes SJH, Steele PA, Costa M (1991) Identification and immunohistochemistry of cholinergic and non-cholinergic circular muscle motor neurons in the guinea-pig small intestine. *Neuroscience* 42:863–878
- Brookes SJH, Song Z-M, Steele PA, Costa M (1992) Identification of motor neurons to the longitudinal muscle of the guinea pig ileum. *Gastroenterology* 103:961–973
- Brookes SJH, Song Z-M, Ramsay GA, Costa M (1995) Long aboral projections of Dogiel type II, AH neurons within the myenteric plexus of the guinea pig small intestine. *J Neurosci* 15:4013–4022
- Brookes SJH, Meedeniya ACB, Jobling P, Costa M (1997) Orally projecting interneurons in the guinea-pig small intestine. *J Physiol* 505:473–491
- Browning KN, Lees GM (1994) Reappraisal of the innervation of rat intestine by vasoactive intestinal polypeptide and neuropeptide Y-immunoreactive neurons. *Neuroscience* 62:1257–1266
- Cavazzana P, Borsetto PL (1948) Recherches sur l'aspect microscopique des plexus nerveux intramuraux et sur les modifications morphologiques de leurs neurones dans les divers traits de l'intestin humain pendant la vie. *Acta Anat* 5:17–41
- Chen TS, Chen PS (1995) The history of gastroenterology. Parthenon, New York
- Christensen J (1988) The forms of argyrophilic ganglion cells in the myenteric plexus throughout the gastrointestinal tract of the opossum. *J Auton Nerv Syst* 24:251–260
- Clarke E, O'Malley CD (1996) The human brain and spinal cord. A historical study illustrated by writings from antiquity to the twentieth century. Norman Publishing, San Francisco
- Clerc N, Furness JB, Bornstein JC, Kunze WAA (1998) Correlation of electrophysiological and morphological characteristics of myenteric neurons of the duodenum in the guinea-pig. *Neuroscience* 82:899–914
- Cole EC (1930) Anastomosing neurons in the myenteric plexus of the human sigmoid flexure. *J Comp Neurol* 50:209–215
- Cook RD, Burnstock G (1976) The ultrastructure of Auerbach's plexus in the guinea-pig. I. Neuronal elements. *J Neurocytol* 5:171–194
- Cornelissen W, Timmermans J-P, Bogaert P-P van, Scheuermann DW (1996) Electrophysiology of porcine myenteric neurons revealed after vital staining of their cell bodies. A preliminary report. *Neurogastroenterol Motil* 8:101–109
- Costa M, Furness JB, Cuello AC, Verhofstad AAJ, Steinbusch HWJ, Elde RP (1982) Neurons with 5-hydroxytryptamine-like immunoreactivity in the enteric nervous system: their visualization and reactions to drug treatment. *Neuroscience* 7:351–362
- Costa M, Brookes S, Waterman S, Mayo R (1992a) Enteric neuronal circuitry and transmitters controlling intestinal motor function. In: Holle GE, Wood JD (eds) *Advances in the innervation of the gastrointestinal tract*. Excerpta Medica, Amsterdam, pp 115–121
- Costa M, Furness JB, Pompolo S, Brookes SJH, Bornstein JC, Bredt DS, Snyder SH (1992b) Projections and chemical coding of neurons with immunoreactivity for nitric oxide synthase in the guinea-pig small intestine. *Neurosci Lett* 148:121–125
- Costa M, Brookes SJH, Steele PA, Gibbins I, Burcher E, Kandiah CJ (1996) Neurochemical classification of myenteric neurons in the guinea-pig ileum. *Neuroscience* 75:949–967
- Cracco C, Filogamo G (1994) Quantitative study of the NADPH-diaphorase-positive myenteric neurons of the rat ileum. *Neuroscience* 61:351–359
- Dogiel AS (1895a) Zur Frage über die Ganglien der Darmgeflechte bei den Säugetieren. *Anat Anz* 10:517–528
- Dogiel AS (1895b) Zur Frage über den feineren Bau des sympathischen Nervensystems bei den Säugetieren. *Arch Mikrosk Anat* 46: 305–344
- Dogiel AS (1896) Zwei Arten sympathischer Nervenzellen. *Anat Anz* 11:679–687
- Dogiel AS (1899) Ueber den Bau der Ganglien in den Geflechten des Darms und der Gallenblase des Menschen und der Säugetiere. *Arch Anat Physiol Leip Anat Abt*: 130–158

- Erde SM, Sherman D, Gershon MD (1985) Morphology and serotonergic innervation of physiologically identified cells of the guinea pig's myenteric plexus. *J Neurosci* 5: 617–633
- Esveld LW van (1928) Über die nervösen Elemente in der Darmwand. *Z Mikrosk Anat Forsch* 15:1–42
- Fehér E, Vajda J (1972) Cell types in the nerve plexus of the small intestine. *Acta Morphol Acad Sci Hung* 20:13–25
- Furness JB, Costa M (1980) Types of nerves in the enteric nervous system. *Neuroscience* 5:1–20
- Furness JB, Costa M (1987) The enteric nervous system. Churchill Livingstone, Edinburgh
- Furness JB, Costa M, Emson PC, Hokanson R, Moghizadeh E, Sundler F, Taylor IL, Chance RE (1983) Distribution, pathways and reactions to drug treatment of nerves with neuropeptide Y- and pancreatic polypeptide-like immunoreactivity in the guinea-pig digestive tract. *Cell Tissue Res* 234:71–92
- Furness JB, Costa M, Gibbins IL, Llewellyn-Smith IJ, Oliver JR (1985) Neurochemically similar myenteric and submucous neurons directly traced to the mucosa of the small intestine. *Cell Tissue Res* 241:155–163
- Furness JB, Bornstein JC, Trussell DC (1988a) Shapes of nerve cells in the myenteric plexus of the guinea-pig small intestine revealed by the intracellular injection of dye. *Cell Tissue Res* 254:561–571
- Furness JB, Keast JR, Pompolo S, Bornstein JC, Costa M, Emson PC, Lawson DEM (1988b) Immunohistochemical evidence for the presence of calcium-binding proteins in enteric neurons. *Cell Tissue Res* 252:79–87
- Furness JB, Trussell DC, Pompolo S, Bornstein JC, Smith TK (1990) Calbindin neurons of the guinea-pig small intestine: quantitative analysis of their numbers and projections. *Cell Tissue Res* 260:261–272
- Furness JB, Bornstein JC, Smith TK (1991) Historical aspects of the investigation of the organisation of the enteric nervous system. *Verh Anat Ges* 85:67–73
- Furness JB, Li ZS, Young HM, Förstermann U (1994) Nitric oxide synthase in the enteric nervous system of the guinea-pig: a quantitative description. *Cell Tissue Res* 277:139–149
- Furness JB, Kunze WAA, Bertrand PP, Clerc N, Bornstein JC (1998) Intrinsic primary afferent neurons of the intestine. *Prog Neurobiol* 54:1–18
- Gabella G (1972) Fine structure of the myenteric plexus in the guinea-pig ileum. *J Anat* 111:69–97
- Gershon MD, Erde SM (1981) The nervous system of the gut. *Gastroenterology* 80: 1571–1594
- Greving R (1951a) Histologische Studien am Plexus myentericus des Magens. 1. Der Plexus myentericus und seine Zelltypen. *Dtsch Z Nervenheilk* 165:622–643
- Greving R (1951b) Histologische Studien am Plexus myentericus des Magens. 2. Das Problem der Ganglienzellfortsätze. *Z Anat Entwicklungsgesch* 115:541–554
- Gunn M (1959) Cell types in the myenteric plexus of the cat. *J Comp Neurol* 111:83–99
- Gunn M (1968) Histological and histochemical observations on the myenteric and submucous plexuses of mammals. *J Anat* 102:223–239
- Hanani M, Ermilov LG, Schmalz PF, Louzon V, Miller SM, Szurszewski JH (1998) The three-dimensional structure of myenteric neurons in the guinea-pig ileum. *J Auton Nerv Syst* 71:1–9
- Harting K (1931) Über die feinere Innervation der extrahepatischen Gallenwege. I. Über die mikroskopische Innervation der Gallenblase. *Z Zellforsch* 12:518–543
- Hendriks R, Bornstein JC, Furness JB (1990) An electrophysiological study of the projections of putative sensory neurons within the myenteric plexus of the guinea pig ileum. *Neurosci Lett* 110:286–290
- Hill CJ (1927) A contribution to our knowledge of the enteric plexuses. *Philos Trans R Soc Lond B Biol Sci* 215:355–387
- Hirst GDS, Holman ME, Spence I (1974) Two types of neurones in the myenteric plexus of duodenum in the guinea-pig. *J Physiol (Lond)* 236: 303–326
- Hodgkiss JP, Lees GM (1980) Morphological features of guinea pig myenteric plexus neurones. In: Christensen J (ed) *Gastrointestinal motility*. Raven Press, New York, pp 111–117
- Hodgkiss JP, Lees GM (1983) Morphological studies of electrophysiologically identified myenteric plexus neurones of the guinea-pig ileum. *Neuroscience* 8:593–608
- Honjin R, Izumi S, Ōsugi H (1959) The distribution and morphology of argentophile and argentophobe nerve cells in the myenteric plexus of the digestive tube of the mouse: a quantitative study. *J Comp Neurol* 111:291–319
- Ilijina WJ, Lawrentjew BJ (1932) Zur Lehre von der Cytoarchitektonik des peripherischen autonomen Nervensystems. III. Ganglien des Rektums und ihre Beziehungen zu dem sakralen Parasympathikus. *Z Mikrosk Anat Forsch* 30:530–542
- Ito T (1936) Zytologische Untersuchungen über die intramuralen Ganglienzellen des Verdauungstraktes. Über die Ganglienzellen der menschlichen Wurmfortsätze, mit besonderer Berücksichtigung auf Golgiapparat, Mitochondrien, Nisslsubstanz und Pigmentgranula. *Okajimas Folia Anat Jpn* 14: 621–663
- Ito T, Nagahiro K (1937) Zytologische Untersuchungen über die intramuralen Ganglienzellen des Verdauungstraktes. Über die Ganglienzellen der Darmwand der Ratte, mit besonderer Berücksichtigung auf die Sekretkörnchen ähnlichen Granula in den intramuralen Ganglienzellen. *Okajimas Folia Anat Jpn* 15:609–634
- Iwanow IF (1930) Die sympathische Innervation des Verdauungstraktes einiger Vogelarten. *Z Mikrosk Anat Forsch* 22:469–492
- Iwanowa TS (1958) Über den Bau der Nervenzellen vom Typus II nach Dogiel. *Z Mikrosk Anat Forsch* 63: 523–528
- Iyer V, Bornstein JC, Costa M, Furness JB, Takahashi Y, Iwanaga T (1988) Electrophysiology of guinea-pig myenteric neurons correlated with immunoreactivity for calcium binding proteins. *J Auton Nerv Syst* 22:141–150
- Jabonero V (1951) Études sur le système neurovégétatif périphérique. III. Innervation de l'estomac humain. *Acta Anat* 11:490–532
- Jabonero V (1958) Mikroskopische Studien über die Innervation des Verdauungstraktes. I. Ösophagus. *Acta Neuroveg* 17:308–353
- Jabonero V (1960) Über die feinere Innervation der menschlichen Gallenblase. *Acta Neuroveg* 20:109–154
- Johnson SE (1925) Experimental degeneration of the extrinsic nerves of the small intestine in relation to the structure of the myenteric plexus. *J Comp Neurol* 38:299–314
- Jones EG (1984) History of cortical cytology. In: Peters A, Jones EG (eds) *Cerebral cortex*, vol 1. Cellular components of the cerebral cortex. Plenum Press, New York London, pp 1–32
- Katayama Y, Lees GM, Pearson GT (1986) Electrophysiology and morphology of vasoactive-intestinal-peptide-immunoreactive neurones of the guinea-pig ileum. *J Physiol (Lond)* 378:1–11
- Kirsche W (1984) Ramon y Cajals Lebenswerk im Wandel der Wertung. Ein Beitrag aus Anlaß der 50. Wiederkehr seines Todestages. *Z Mikrosk Anat Forsch* 98:641–658
- Kobayashi S, Suzuki M, Uchida T, Yanaihara N (1984) Enkephalin neurons in the guinea pig duodenum: a light and electron microscopic immunocytochemical study using an antiserum to methionine-enkephalin-Arg⁶-Gly⁷-Leu⁸. *Biomed Res* 5:489–506
- Kobayashi S, Suzuki M, Yanaihara N (1985) Enkephalin neurons in the guinea pig proximal colon: an immunocytochemical study using an antiserum to methionine-enkephalin-Arg⁶-Gly⁷-Leu⁸. *Arch Histol Jpn* 48:27–44
- Koelliker A (1896) *Handbuch der Gewebelehre des Menschen*. Engelmann, Leipzig
- Kolossow NG, Milochin AA (1963) Die afferente Innervation der Ganglien des vegetativen Nervensystems. *Z Mikrosk Anat Forsch* 70:426–464
- Kolossow NG, Sabussow GH (1928) Die sympathische Innervation des Verdauungstraktes der Sumpfschildkröte (*Emys europaea* L.). *Z Mikrosk Anat Forsch* 15:157–190

- Kolossow NG, Sabussow GH (1932) Zur Frage der Innervation des menschlichen Magen-Darmkanals. *Z Mikrosk Anat Forsch* 29:541–560
- Krammer H-J, Brehmer A, Wedel T, Stach W (1998) Das Typ I-Neuron im Dünndarm des Menschen – eine nach oral projizierende Neuronenpopulation. *Verh Anat Ges* 93: 302–303
- Kuntz A (1913) On the innervation of the digestive tube. *J Comp Neurol* 23:173–192
- Kuntz A (1923) On the occurrence of reflex arcs in the myenteric and submucous plexuses. *Anat Rec* 24:193–210
- Kunze WAA, Bornstein JC, Furness JB (1995) Identification of sensory nerve cells in a peripheral organ (the intestine) of a mammal. *Neuroscience* 66:1–4
- Langley JN (1900) The sympathetic and other related systems of nerves. In: Schäfer EA (ed) *Text-book of physiology*. Pentland, Edinburgh, pp 616–696
- Langley JN (1921) *The autonomic nervous system, part 1*. Heffer, Cambridge
- La Villa I (1898) Estructura de los ganglios intestinales. *Rev Trimestral Micrografica* III:1–13
- Lawrentjew BJ (1929) Experimentell-morphologische Studien über den feineren Bau des autonomen Nervensystems. II. Über den Aufbau der Ganglien der Speiseröhre nebst einigen Bemerkungen über das Vorkommen und die Verteilung zweier Arten von Nervenzellen in dem autonomen Nervensystem. *Z Mikrosk Anat Forsch* 18:233–262
- Lawrentjew BJ (1931) Zur Lehre von der Cytoarchitektonik des peripherischen Nervensystems. I. Die Cytoarchitektonik der Ganglien des Verdauungskanal beim Hunde. *Z Mikrosk Anat Forsch* 23:527–551
- Lees GM, Mackenzie GM, Pearson GT (1992) Complex correlations between the morphology, electrophysiology and peptide immunohistochemistry of guinea-pig enteric neurons. *Eur J Morphol* 30:123–136
- Li ZS, Furness JB (1998) Immunohistochemical localisation of cholinergic markers in putative intrinsic primary afferent neurons of the guinea-pig small intestine. *Cell Tissue Res* 294:35–43
- Llewellyn-Smith IJ, Furness JB, Wilson AJ, Costa M (1983) Organization and fine structure of enteric ganglia. In: Elfvin L-G (ed) *Autonomic ganglia*. Wiley, Chichester, pp 145–181
- Mann PT, Southwell BR, Ding YQ, Shigemoto R, Mizuno N, Furness JB (1997) Localisation of neurokinin 3 (NK3) receptor immunoreactivity in the rat gastrointestinal tract. *Cell Tissue Res* 289:1–9
- Martini GA (1996) Geschichte der Gastroenterologie. In: Hahn EG, Reimann JF (eds) *Klinische Gastroenterologie*, 3rd ed. Thieme, Stuttgart, pp 2–13
- Meedeniya ACB, Brookes SJH, Hennig GW, Costa M (1998) The projections of 5-hydroxytryptamine-accumulating neurones in the myenteric plexus of the small intestine of the guinea-pig. *Cell Tissue Res* 291:375–384
- Messenger JB, Bornstein JC, Furness JB (1994) Electrophysiological and morphological classification of myenteric neurons in the proximal colon of the guinea-pig. *Neuroscience* 60:227–244
- Michail S, Karamanlidis A (1967) Morphologie du plexus myentérique d'Auerbach de l'intestin grêle du chien. *Acta Anat* 67:424–436
- Müller E (1921) Über das Darmnervensystem. *Ups Läkaref Förh* 26:XXII. 1–22
- Müller LR (1912) Die Darminnervation. *Dtsch Arch Klin Med* 105:1–43
- Murat VN (1933) Sur la question de la cytoarchitectonique des ganglions nerveux de l'intestin de l'homme. *Trab Lab Invest Biol Univ Madrid* 28:387–401
- Neunlist M, Schemann M (1997) Projections and neurochemical coding of myenteric neurons innervating the mucosa of the guinea pig proximal colon. *Cell Tissue Res* 287:119–125
- Nichols K, Krantis A, Staines W (1992) Histochemical localization of nitric oxide-synthesizing neurons and vascular sites in the guinea-pig intestine. *Neuroscience* 51:791–799
- Okamura C (1934) Über die Darstellung des Nervenapparates in der Magen-Darmwand mittels der Vergoldungsmethode. *Z Mikrosk Anat Forsch* 35:218–253
- Oshima D (1929) Über die Innervation des Darmes. *Z Anat Entwicklungsgesch* 90:725–767
- Ottaviani G (1940) Histologisch-anatomische Untersuchungen über die Innervation des Mastdarmes (Intestinum terminale-Rectum). *Z Mikrosk Anat Forsch* 47:151–182
- Pearson GT (1994) Structural organization and neuropeptide distributions in the equine enteric nervous system: an immunohistochemical study using whole-mount preparations from the small intestine. *Cell Tissue Res* 276:523–534
- Peters A, Palay SL, Webster H deF (1991) *The fine structure of the nervous system*. Oxford University Press, New York
- Pompolo S, Furness JB (1988) Ultrastructure and synaptic relationships of calbindin-reactive, Dogiel type II neurons, in myenteric ganglia of guinea-pig small intestine. *J Neurocytol* 17:771–782
- Pompolo S, Furness JB (1990) Ultrastructure and synaptology of neurons immunoreactive for gamma-aminobutyric acid in the myenteric plexus of the guinea pig small intestine. *J Neurocytol* 19:539–549
- Portbury AL, Pompolo S, Furness JB, Stebbing MJ, Kunze WAA, Bornstein JC, Hughes S (1995) Cholinergic, somatostatin-immunoreactive interneurons in the guinea pig intestine: morphology, ultrastructure, connections and projections. *J Anat* 187:303–321
- Portbury AL, Furness JB, Young HM, Southwell BR, Vigna SR (1996) Localisation of NK1 receptor immunoreactivity to neurons and interstitial cells of the guinea-pig gastrointestinal tract. *J Comp Neurol* 367:342–351
- Porter AJ, Wattchow DA, Brookes SJH, Schemann M, Costa M (1996) Choline acetyltransferase immunoreactivity in the human small and large intestine. *Gastroenterology* 111:401–408
- Ramón y Cajal S (1893) Sur les ganglions et plexus nerveux de l'intestin. *C R Soc Biol* 5:217–223
- Ramón y Cajal S (1911) *Histologie du système nerveux de l'homme et des vertébrés*. Maloine, Paris
- Reiser KA (1932) Der Nervenapparat im Processus vermiformis nebst einigen Bemerkungen über seine Veränderungen bei chronischer Appendicitis. *Z Zellforsch* 15:761–799
- Rossi O (1929) Contributo alla conoscenza degli apparati nervosi intramurali dell' intestino tenue. *Arch Ital Anat Embriol* 26: 632–644
- Scheuermann DW, Stach W, De Groodt-Lasseel MHA, Timmermans J-P (1987) Calcitonin gene-related peptide in morphologically well-defined type II neurons of the enteric nervous system in the porcine small intestine. *Acta Anat* 129:325–328
- Scheuermann DW, Krammer H-J, Timmermans J-P, Stach W, Adriaensen D, De Groodt-Lasseel MHA (1991) Fine structure of morphologically well-defined type II neurons in the enteric nervous system of the porcine small intestine revealed by immunoreactivity for calcitonin gene-related peptide. *Acta Anat* 142:236–241
- Schierhorn H (1981) Die Hirnanatomie im Spiegel der ersten 50 Bände des Anatomischen Anzeigers. *Anat Anz* 150:77–118
- Schofield GC (1962) Experimental studies on the myenteric plexus in mammals. *J Comp Neurol* 119:159–185
- Sokolowa ML (1931) Zur Lehre von der Cytoarchitektonik des peripherischen autonomen Nervensystems. II. Die Architektur der intramuralen Ganglien des Verdauungstrakts des Rindes. *Z Mikrosk Anat Forsch* 23:552–570
- Song Z-M, Brookes SJH, Costa M (1991) Identification of myenteric neurons which project to the mucosa of the guinea-pig small intestine. *Neurosci Lett* 129:294–298
- Song Z-M, Brookes SJH, Costa M (1994) All calbindin-immunoreactive neurons project to the mucosa of the guinea-pig small intestine. *Neurosci Lett* 180:219–222
- Song Z-M, Brookes SJH, Costa M (1996) Projections of specific morphological types of neurons within the myenteric plexus of the small intestine of the guinea-pig. *Cell Tissue Res* 285:149–156

- Stach W (1977) Die differenzierte Gefäßversorgung der Dogiel-schen Zelltypen und die bevorzugte Vaskularisation der Typ I-Zellen in den Ganglien des Plexus submucosus externus (Schabadasch) des Schweins. *Z Mikrosk Anat Forsch* 91:421–429
- Stach W (1979) Die differenzierte Gefäßversorgung der Dogiel-schen Zelltypen und die bevorzugte Vaskularisation der Typ I/2-Zellen in den Ganglien des Plexus myentericus (Auerbach) des Schweins. *Anat Anz* 145:464–473
- Stach W (1980) Zur neuronalen Organisation des Plexus myentericus (Auerbach) im Schweinedünndarm. I. Typ I-Neurone. *Z Mikrosk Anat Forsch* 94:833–849
- Stach W (1981) Zur neuronalen Organisation des Plexus myentericus (Auerbach) im Schweinedünndarm. II. Typ II-Neurone. *Z Mikrosk Anat Forsch* 95:161–182
- Stach W (1982a) Zur neuronalen Organisation des Plexus myentericus (Auerbach) im Schweinedünndarm. III. Typ III-Neurone. *Z Mikrosk Anat Forsch* 96:497–516
- Stach W (1982b) Zur neuronalen Organisation des Plexus myentericus (Auerbach) im Schweinedünndarm. IV. Typ IV-Neurone. *Z Mikrosk Anat Forsch* 96:972–994
- Stach W (1983) Über morphologisch definierte vertikale Verbindungen innerhalb des Darmwandnervensystems im Schweinedünndarm. *Verh Anat Ges* 77:577–578
- Stach W (1985) Zur neuronalen Organisation des Plexus myentericus (Auerbach) im Schweinedünndarm. V. Typ V-Neurone. *Z Mikrosk Anat Forsch* 99:562–582
- Stach W (1989) A revised morphological classification of neurons in the enteric nervous system. In: Singer MV, Goebell H (eds) *Nerves and the gastrointestinal tract*. Kluwer, Lancaster, pp 29–45
- Steele PA, Brookes SJH, Costa M (1991) Immunohistochemical identification of cholinergic neurons in the myenteric plexus of guinea-pig small intestine. *Neuroscience* 45:227–239
- Stöhr P (1931) Mikroskopische Studien zur Innervation des Magen-Darmkanales. *Z Zellforsch* 12:66–154
- Stöhr P (1949) Mikroskopische Studien zur Innervation des Magen-Darmkanales V. *Z Zellforsch* 34:1–54
- Stöhr P (1952) V. Zusammenfassende Ergebnisse über die mikroskopische Innervation des Magen-Darmkanals. *Ergeb Anat Entwicklungsgesch* 34:250–401
- Sundler F, Ekblad E, Hokanson R (1993) Localization and colocalization of gastrointestinal peptides. In: Brown DR (ed) *Gastrointestinal regulatory peptides*. Springer, Berlin, pp 1–28
- Sutherland SD (1967) The neurons of the gall bladder and gut. *J Anat* 101:701–709
- Tamura K (1992) Morphology of electrophysiologically identified myenteric neurons in the guinea pig rectum. *Am J Physiol* 262:G545–G552
- Tamura K (1997) Synaptic inputs to morphologically identified myenteric neurons in guinea pig rectum from pelvic nerves. *Am J Physiol* 273:G49–G55
- Temesrékási D (1955) Die Synaptologie der Dünndarmgeflechte. *Acta Morph Acad Sci Hung* 5:53–69
- Tiegs OW (1927) The structure of the neurone junctions in sympathetic ganglia, and in the ganglia of Auerbach's plexus. *Aust J Exp Biol Med Sci* 4:79–98
- Timmermans J-P, Scheuermann DW, Stach W, Adriaensen D, De Groodt-Lasseel MHA, Polak JM (1989) Neuromedin U-immunoreactivity in the nervous system of the small intestine of the pig and its coexistence with substance P and CGRP. *Cell Tissue Res* 258:331–337
- Timmermans J-P, Scheuermann DW, Barbiers M, Adriaensen D, Stach W, Hee R van, De Groodt-Lasseel MHA (1992a) Calcitonin gene-related peptide-like immunoreactivity in the human small intestine. *Acta Anat* 143:48–53
- Timmermans J-P, Scheuermann DW, Stach W, Adriaensen D, De Groodt-Lasseel MHA (1992b) Functional morphology of the enteric nervous system with special reference to large mammals. *Eur J Morphol* 30:113–122
- Timmermans J-P, Barbiers M, Scheuermann DW, Stach W, Adriaensen D, De Groodt-Lasseel MHA (1993) Occurrence, distribution and neurochemical features of small intestinal neurons projecting to the cranial mesenteric ganglion in the pig. *Cell Tissue Res* 272:49–58
- Timmermans J-P, Barbiers M, Scheuermann DW, Bogers JJ, Adriaensen D, Fekete É, Mayer B, Van Marck EA, De Groodt-Lasseel MHA (1994a) Nitric oxide synthase immunoreactivity in the enteric nervous system of the developing human digestive tract. *Cell Tissue Res* 275:235–245
- Timmermans J-P, Barbiers M, Scheuermann DW, Stach W, Adriaensen D, Mayer B, De Groodt-Lasseel MHA (1994b) Distribution pattern, neurochemical features and projections of nitrergic neurons in the pig small intestine. *Ann Anat* 176:515–525
- Timmermans J-P, Adriaensen D, Cornelissen W, Scheuermann DW (1997) Structural organization and neuropeptide distribution in the mammalian enteric nervous system, with special attention to those components involved in mucosal reflexes. *Comp Biochem Physiol* 118A:331–340
- Trendelenburg P (1917) Physiologische und pharmakologische Versuche über die Dünndarmperistaltik. *Arch Exp Pathol Pharmacol* 81:55–129
- Waldeyer W (1891) Über einige neuere Forschungen im Gebiete der Anatomie des Centralnervensystems. *Dtsch Med Wochenschr* 17:1213–1218; 1244–1246; 1267–1269; 1287–1289; 1331–1332; 1352–1356
- Ward SM, Xue C, Shuttleworth CW, Bredt DS, Snyder SH, Sanders KM (1992) NADPH diaphorase and nitric oxide synthase colocalization in enteric neurons of canine proximal colon. *Am J Physiol* 263:G277–G284
- Wattchow DA, Brookes SJH, Costa M (1995) The morphology and projections of retrogradely labeled myenteric neurons in the human intestine. *Gastroenterology* 109:866–875
- Wattchow DA, Porter A, Brookes SJH, Costa M (1997) The polarity of neurochemically defined myenteric neurons in the human colon. *Gastroenterology* 113:497–506
- Wilson AJ, Furness JB, Costa M (1981a) The fine structure of the submucous plexus of the guinea-pig ileum. I. The ganglia, neurons, Schwann cells and neuropil. *J Neurocytol* 10:759–784
- Wilson AJ, Furness JB, Costa M (1981b) The fine structure of the submucous plexus of the guinea-pig ileum. II. Description and analysis of vesiculated nerve profiles. *J Neurocytol* 10:785–804
- Wood JD (1994) Physiology of the enteric nervous system. In: Johnson LR (ed) *Physiology of the gastrointestinal tract*. Raven Press, New York, pp 423–482
- Young HM, Furness JB (1995) Ultrastructural examination of the targets of serotonin-immunoreactive descending interneurons in the guinea pig small intestine. *J Comp Neurol* 356:101–114
- Young HM, Furness JB, Shuttleworth CWR, Bredt DS, Snyder SH (1992) Co-localization of nitric oxide synthase immunoreactivity and NADPH diaphorase staining in neurons of the guinea-pig intestine. *Histochemistry* 97:375–378
- Young HM, Furness JB, Povey JM (1995) Analysis of connections between nitric oxide synthase neurons in the myenteric plexus of the guinea-pig small intestine. *J Neurocytol* 24:257–263