

Connections between the zona incerta and superior colliculus in the monkey and squirrel

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Abstract The zona incerta contains GABAergic neurons that project to the superior colliculus in the cat and rat, suggesting that it plays a role in gaze changes. However, whether this incertal connection represents a general mammalian pattern remains to be determined. We used neuronal tracers to examine the zona incerta connections with the midbrain tectum in the gray squirrel and macaque monkey. Collicular injections in both species revealed that most incertotectal neurons lay in the ventral layer, but anterogradely labeled tectoincertain terminals were found in both the dorsal and ventral layers. In the monkey, injections of the pretectum also produced retrograde labeling, but mainly in the dorsal layer. The dendritic fields of incertotectal and incertopretectal cells were generally contained within the layer inhabited by their somata. The macaque, but not the squirrel, zona incerta extended dorsolaterally, within the external medullary lamina. Zona

incerta injections produced retrogradely labeled neurons in the superior colliculus of both species. In the squirrel, most cells inhabited the lower sublamina of the intermediate gray layer, but in the monkey, they were scattered throughout the deeper layers. Labeled cells were present among the pretectal nuclei in both species. Labeled terminals were concentrated in the lower sublamina of the intermediate gray layer of both species, but were dispersed among the pretectal nuclei. In summary, an incertal projection that is concentrated on the collicular motor output layers and that originates in the ventral layer of the ipsilateral zona incerta is a common mammalian feature, suggesting an important role in collicular function.

Keywords Gaze · Eye movements · GABA · Primate · Pretectum · Inhibition

Abbreviations

III	Oculomotor nucleus
APt	Anterior pretectal nucleus
BDA	Biotinylated dextran amine
BP	Basilar pons
DLG	Dorsal lateral geniculate nucleus
EML	External medullary lamina
GP	Globus pallidus
HRP	Horseshoe peroxidase
IC	Inferior colliculus
ICa	Internal capsule
Lim	Nucleus limitans
MG	Medial geniculate nucleus
MpT	Medial pretectal nucleus
nPC	Nucleus of the posterior commissure
nOT	Nucleus of the optic tract
Opt	Stratum opticum
OT	Optic tract

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PAG	Periaqueductal gray
PB	Parabrachial nucleus
PG	Pregenulate
Pul	Pulvinar
Pt	Pretectum
PPt	Posterior pretectal nucleus
RT	Reticular thalamic nucleus
SAI	Intermediate white layer
SC	Superior colliculus
SGS	Stratum griseum superficiale
SGI	Stratum griseum intermediale
ISGI	SGI, lower sublamina
uSGI	SGI, upper sublamina
SGP	Stratum griseum profundum
SNc	Substantia nigra pars compacta
SNl	Substantia nigra pars lateralis
SNr	Substantia nigra pars reticulata
SO	Stratum opticum
STh	Subthalamic nucleus
WGA-HRP	Wheat germ agglutinin conjugated HRP
VLG	Ventral lateral geniculate nucleus
VP	Ventral posterior nucleus
ZI	Zona incerta
dZI	Dorsal layer of ZI
vZI	Ventral layer of ZI

Introduction

Inhibitory processes help control the bursts of activity in the superior colliculus (SC) that precede each saccade (Hikosaka and Wurtz 1985). Unlike the widespread activity present in the superficial, visuosensory layer, in the deeper layers, activity needs to be concentrated at one location in the motor map once a target has been selected, because only one saccade can be made at a time. There is evidence that the direct inputs from superficial to deep collicular layers are modulated by inhibition to arrange this pattern of activation and that selective release of this inhibition may be important in producing express saccades (Ozen et al. 2000; Isa and Saito 2001). Inhibition also plays a role in the relationship between the two colliculi, which have activity that codes for saccades to the left and the right, and between the hemifields within each colliculus, which code for upward and downward eye movements (Munoz and Istvan 1998; Takahashi et al. 2005, 2007). Inhibition may even play a role in defining the duration of burst activity and the location of cells activated within the collicular motor map (Kaneda et al. 2008). Since this activity pattern results in both an orienting movement and a shift in attention by the animal (Basso and May 2017), one may argue that inhibition is critical to the primary roles of the colliculus. Furthermore, the most salient target must be

chosen from a host of possibilities, and the property of salience depends on the memory of the animal, species specific behavioral presets (e.g., what is food and what is a threat), previous reinforcements, and the internal state of the animal (Ingle 1971; Dean et al. 1989; Westby et al. 1990; Sato and Hikosaka 2002; Yasuda et al. 2012). All of these decisions may involve inhibiting alternative choices.

Clearly, the roles of inhibition in the SC are various and numerous. In line with this, GABAergic terminals are very numerous on the surface of collicular motor output neurons (Lu et al. 1985). Indeed, the deep layers contain a prominent collection of GABAergic local circuit interneurons (Mize 1988; Mize et al. 1991). In addition, they are targeted by a variety of extrinsic GABAergic inputs (Appell and Behan 1990). The best studied among these is the substantia nigra pars reticulata. The nigrotectal projection provides an extensive input to the motor output layers of the SC (Jayaraman et al. 1977; May and Hall 1984; Harting et al. 1988; Bickford and Hall 1992; Redgrave et al. 1992). It is likely that this projection plays an important role in releasing the burst of activity in collicular cells before saccades, including those to remembered targets (Hikosaka and Wurtz 1983b). Another major extrinsic source of GABAergic input is the central mesencephalic reticular formation (cMRF). The cMRF receives direct input from collicular gaze output neurons and then provides a GABAergic feedback projection back onto these output cells (Moschovakis et al. 1988; Chen and May 2000; Zhou et al. 2008; Wang et al. 2010). While it is known that the cMRF contains neurons that display long lead, saccade-related bursts, the precise role of this feedback pathway has yet to be ascertained (Waitzman et al. 1996; Cromer and Waitzman 2007).

Yet, another major extrinsic GABAergic projection to the SC comes from the zona incerta (ZI). This projection exists in both the rat and cat (Edwards et al. 1979; Watanabe and Kawana 1982; Shammah-Lagnado et al. 1985; Ficalora and Mize 1989; Appell and Behan 1990). While some ZI neurons in monkeys display saccade-related activity (Hikosaka and Wurtz 1983a; Ma 1996), there is evidence from non-primates that they modify movement with respect to internal state and autonomic nervous system activity levels (Mitrofanis 2005). Nevertheless, the ZI's precise role with respect to tectally generated gaze changes is poorly understood. Towards this end, we have endeavored to better define the pattern of connections between the ZI and the SC. Previously, we demonstrated a number of features of these connections in the cat (May et al. 1997; Perkins et al. 2006). These included the fact that the source of the incertotectal projection was largely confined to the ventral layer of ZI and that this layer extended dorsolaterally between the reticular thalamic nucleus and the external medullary lamina. Within the SC, the incertotectal

terminal field was concentrated in the lower sublamina of the intermediate gray layer (SGI), and this same sublamina is the primary source of the tectoincercal projection. Finally, in the cat, we noted connections between the dorsal sublamina of ZI and the pretectum, particularly the anterior pretectal nucleus. While connections between the ZI and the SC are also present in the rat, a number of the details of this projection differed from those seen in the cat (Sham-mah-Lagnado et al. 1985; Roger and Cadusseau 1985). To determine which characteristics of the connections between the ZI and the tectum represent general mammalian features and to provide an anatomical basis for studying this system in primates, we explored the connections of the ZI in two species: the gray squirrel (*Sciurus carolinensis*) and cynomolgous monkey (*Macaca fascicularis*). While both these species have evolved in a diurnal, arboreal habitat that puts a premium on vision and accurate gaze, they occupy fairly different branches of the mammalian radiation.

Methods

The data presented here were obtained from slide collections produced during previous studies (squirrels: May and Hall 1986; monkeys: May et al. 1990; Chen and May 2000; Perkins et al. 2009). All procedures were undertaken in accordance with the NIH rules and regulations governing animal research, as enumerated in the Guide for Care and Use of Laboratory Animals, using protocols approved by the Institutional Animal Care and Use Committee of the University of Mississippi Medical Center and as required by USDA regulations. Tissue from 7 squirrels (*Sciurus carolinensis*) and 16 monkeys (*Macaca fascicularis*) of both sexes was analyzed.

Squirrel procedures

Squirrels were anesthetized with sodium pentobarbital (50 mg/kg, ip) and placed in a stereotaxic head holder (Trent-Wells). Injections (0.05 μ l) of horseradish peroxidase (HRP) (30% in a 10% saponin solution) were placed in the SC ($n = 3$) by use of a 1.0 μ l Hamilton syringe held in a micromanipulator. Inclusion of saponin was intended to increase homogeneous filling of the dendritic trees of the retrogradely labeled cells (May and Hall 1984). The SC ($n = 3$) and the ZI ($n = 1$) were also injected iontophoretically. In this case, a 10% solution of wheat germ agglutinin conjugated to horseradish peroxidase (WGA-HRP) was placed in a glass micropipette with a tip diameter of 25 μ m that was attached to a micromanipulator. The tracer was ejected by use of 7 s, 50% duty cycle square wave positive pulses for 10 min. All injection placements

utilized stereotaxic coordinates developed from an in-house atlas. These animals were deeply anesthetized with sodium pentobarbital (70 mg/kg, ip) 2–3 days later, and were perfused through the heart with a buffered saline rinse, followed by a fixative containing 0.5% paraformaldehyde and 2.5% glutaraldehyde in 0.1 M, pH 7.2 phosphate buffer (PB). The brains were postfixed in the same solution for 2 h and then blocked in the frontal plane.

The brains with HRP injections were cut into 100 μ m frontal sections using a Vibratome. A one in three series of sections was reacted using a modification of the orthotoluidine method of Somogyi et al. (1979). These sections were mounted out of 0.1 M, pH 6.0 PB, counterstained with cresyl violet, cleared, and coverslipped. The brains with WGA-HRP injections were equilibrated in 30% sucrose in 0.1 M, pH 7.2 PB at 4 °C as a cryoprotectant, before being frozen and sectioned on an AO sliding microtome at 40 μ m. A one in three series of sections was reacted to reveal the presence of HRP using the tetramethylbenzidine method of Mesulam (1978). Further details of the surgical and histological procedures can be found in May and Hall (1984, 1986).

Monkey procedures

The monkeys used in these procedures were sedated with ketamine HCL (10 mg/kg, im) and anesthetized with either sodium pentobarbital (35 mg/kg, iv) or with isoflurane (2–3%). They were then placed in stereotaxic frame (Kopf). For SC injections ($n = 8$), the surface of the midbrain tectum was visualized by aspirating the medial surface of the parieto-occipital junction. In some cases ($n = 5$), a 1–2% WGA-HRP/10% HRP solution was used. In other cases ($n = 3$), a 10% solution of biotinylated dextran amine (BDA) was injected. In both cases, the solution was contained within a 1.0 μ l Hamilton syringe held by a micromanipulator at a 20°–25° angle, tip up in the sagittal plane. The needle was advanced 1.5 mm into the SC and 0.01–0.02 μ l of WGA-HRP or 0.1–0.2 μ l of BDA was injected. One-to-three sites in the SC were injected. For the pretectal injections ($n = 6$), the same approach was used to make WGA-HRP ($n = 4$) or BDA ($n = 2$) injections, but the entry point of the needle was located at the point on the surface where the tectum meets the pulvinar, and the needle was located 1–2 mm off the midline. Finally, for the injection of WGA-HRP into the ZI ($n = 2$), the needle was oriented vertically and the nucleus was located using stereotaxic coordinates, without aspiration (Szabo and Cowan 1984). Butorphanol tartrate (0.3 mg/kg, im) or Buprenex (0.01 mg/kg, im) was given for postoperative analgesia. After a 1–2 day survival for the WGA-HRP injections or a 2–3 week survival for the BDA injections, the animals were sedated with ketamine

HCL (10 mg/kg, im) and deeply anesthetized with sodium pentobarbital (50 mg/kg, ip). They were then perfused with a 0.1 M, pH 7.2 PB saline rinse, followed by a fixative solution containing 1.0% paraformaldehyde and 1.25% glutaraldehyde in 0.1 M, pH 7.2 PB. The brains were blocked postfixed for 1–2 h in this solution.

Frontal sections were cut at either 100 μm using a Vibratome or 50 μm using an AO sliding microtome. In the latter case, the brains were first cryoprotected in 30% sucrose PB before being frozen. To reveal the HRP, sections were reacted using one of two tetramethylbenzidine methods: that of Mesulam (1978) or that of Olucha et al. (1985). To visualize the BDA, sections were treated with avidin–HRP, and then the HRP was visualized by use of the diaminobenzidine with nickel cobalt enhancement technique of Adams (1977). Details of the surgical and histological procedures used in these experiments can be found in our previous reports (May and Porter 1992; Chen and May 2000; Perkins et al. 2009). With respect to nomenclature, we divided the ZI into dorsal and ventral layers, and noted the presence of a dorsolateral extension when present, as per our previous studies (May et al. 1997; Perkins et al. 2006).

Analysis

The labeled elements were charted and drawn on a BH-2 Olympus microscope equipped with a drawing tube. Images were taken using a Nikon Eclipse 600 microscope equipped with a Nikon Ds-Ri1 digital camera using the Nikon Elements software. Contrast and color were manipulated in Photoshop (Adobe) to best match the appearance of the material to the eye.

Results

Squirrel collicular injections

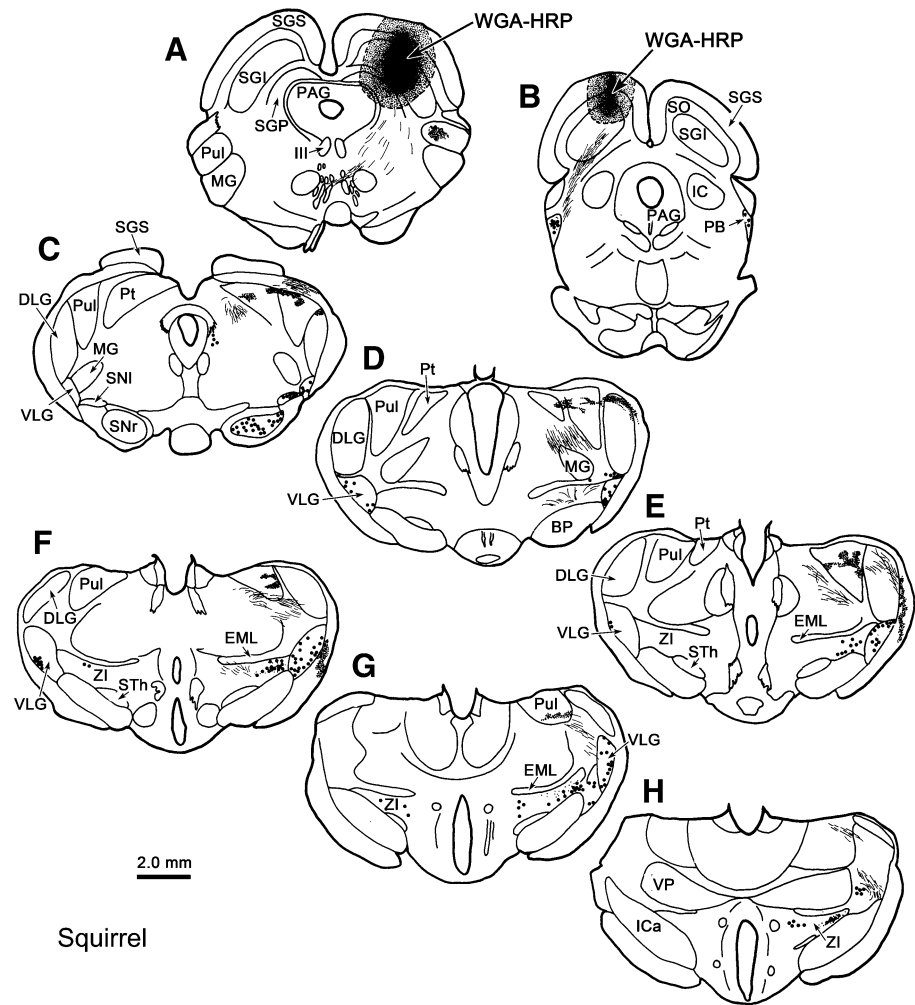
We will cover the connections seen in the gray squirrel first. Figure 1 shows some essential elements of the pattern of connections observed following injections of WGA-HRP into the SC in the context of the squirrel midbrain and thalamus. In this animal, a full thickness injection was placed in the rostral, right SC (Fig. 1a). A second, more caudal, injection that included the superficial gray layer (SGS), stratum opticum (SO), and the upper half of SGI was placed in the left SC (Fig. 1b). Both injections included the SGS, so terminal fields (stipple) were observed in the pretectal, pulvinar, dorsal lateral geniculate, and parabigeminal nuclei on both sides of the brain (Fig. 1b–f). The last also contained retrogradely labeled cells (dots) (Fig. 1b). Within the ventral thalamus, retrogradely labeled cells were present in both the ventral lateral geniculate and ZI from both injections (Fig. 1c–g). However, the more

superficial injection on the left produced considerably fewer cells in the ventral lateral geniculate nucleus and only a couple of scattered cells in ZI, suggesting the latter projects more ventrally within SGI. Similarly, nearly, all the labeled nigrotectal cells were found on the right side (Fig. 1c), in agreement with studies showing the squirrel nigrotectal projection terminates in the lower sublamina of SGI (May and Hall 1984). Labeled terminals were evident in the ZI only on the side of the deeper injection (right).

Figure 2 shows a second squirrel injection case with a unilateral WGA-HRP injection site located rostrally in the SC (Fig. 2a–c). It was centered in the deep gray layer (SGP), but still included all the layers. It spread to include the lateral portion of the periaqueductal gray and portions of the pretectum, specifically the nucleus of the optic tract and posterior pretectal nucleus. The retrograde labeling (dots) within the ventral thalamus was extensive. Most of the retrogradely labeled cells were found within the ventral half of the ipsilateral ZI (Fig. 2f–h), although scattered cells were present in the dorsal half of ZI, particularly rostrally (Fig. 2d–f). A small number of labeled cells were found in contralateral ZI in this and other cases, but they were so few in number that we did not pursue this point. Labeled terminals (stipple) produced a distinct band along the top edge of the dorsal half of ZI. Scattered puncta were also present beneath this band, including some among the cell bodies in the bottom half of ZI (Fig. 2d–f). Additional labeled cells and terminals were present in the ventral lateral geniculate (Fig. 2d–g). The subdivision of ZI into two layers with different patterns of tectal connections can be further appreciated in Fig. 3a showing images from this case. The ventral layer of ZI (vZI) is filled with densely labeled neurons. The dorsal layer of ZI (dZI) lacks these labeled cells, but contains a prominent band of anterogradely labeled terminals in its upper half. Scattered retrogradely labeled cells are present in dZI, but these are much more lightly labeled than those in vZI (compare cells indicated by arrowheads in dZI to those in vZI in Fig. 3b), suggesting that they may be labeled by injection site spread outside the SC. Labeled puncta are present amongst the retrogradely labeled cells in vZI, but it is not clear if these represent terminals or pieces of labeled dendrite (Fig. 3b).

The morphology of the incertotectal population is illustrated in Fig. 4. In this case, HRP was injected primarily in the layers of the colliculus beneath SO (Fig. 4c). The injection site spread into the adjacent periaqueductal gray and the rostral pole of the inferior colliculus. The somata of the labeled multipolar cells tended to be fusiform in shape and oriented parallel to the mediolateral axis of ZI (Fig. 4a, cells C–F and H–J). The primary and secondary dendrites of the neurons were filled with tracer, and these also tended to extend horizontally, parallel with the mediolateral axis of the nucleus. As a group (Fig. 4b), most of the incertotectal

Fig. 1 Laminar differences in squirrel incertotectal neurons. Chartings of the distribution of labeled cells (*dots*), axons (*lines*), and terminals (*stipple*) in the diencephalon following injections of WGA-HRP into the *right* (a) and *left* (b) SC (a, b) in a gray squirrel. Note the presence of many more labeled cells and terminals in the ventral lateral geniculate (VLG) and zona incerta (ZI) following the deeper (*right*) injection. Sections are arranged in rostral to caudal order in this and other figures with chartings



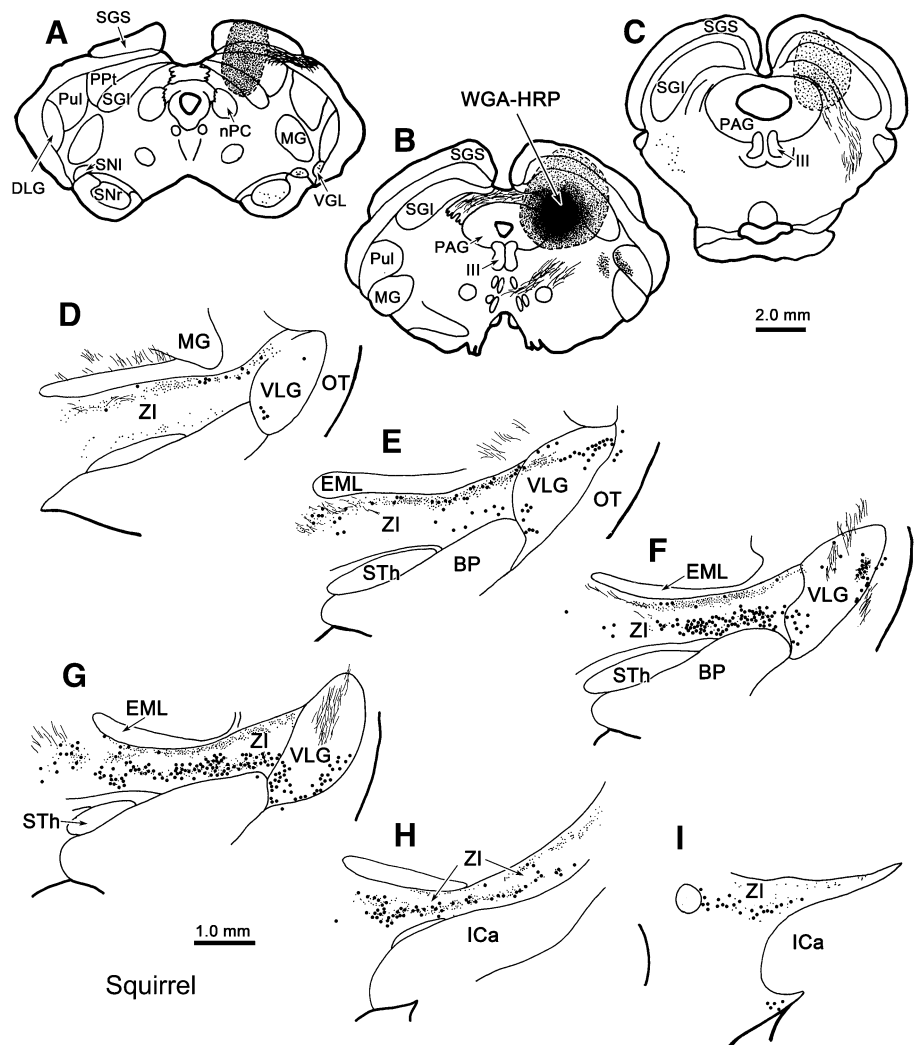
cells labeled in this and the other cases examined lay within vZI, and their labeled dendrites were constrained within this same layer. At the medial edge of ZI, the horizontal orientation became less evident and cells took on more of a multipolar character (Fig. 4a, cells A and B). At the lateral edge of ZI, the layer takes on diagonal orientation, instead of a horizontal one, and the orientation of the cells follows that change (Fig. 4, cells K and L). A second case with retrograde labeling following a more superficial WGA-HRP injection (Fig. 5b) filled even tertiary dendrites. It still demonstrated that the somata and dendrites of incertotectal cells are generally oriented in the same plane as ZI (Figs. 3d, 5a, cells B–E). Even those cells that had a more multipolar organization (Fig. 5a, cell A) displayed dendrites that were largely constrained to the lower half of ZI.

Squirrel zona incerta injection

The pattern of labeling in the squirrel tectum following an injection of ZI is demonstrated in Fig. 6. The WGA-HRP

injection site was quite small, and was located in the medial portion of ZI towards its caudal end (Fig. 6a–c). It spread slightly into the substantia nigra pars compacta. Within the SC, the labeled terminal field (stipple) was largely confined to the lower sublamina of SGI and to SGP (Fig. 6e–j) and extended throughout much of the rostro-caudal and mediolateral extents of these layers. The lack of terminal fields in upper SGI explains the near lack of retrograde ZI labeling from the superficial injection on the left in the case illustrated in Fig. 1. Scattered retrogradely labeled neurons (dots) were present in these same sublamina (Fig. 6e–j). As can be seen in Fig. 3c, the terminal field from this small injection was fairly sparse and the retrogradely labeled neurons (arrowheads) were relatively small. Labeled terminals were also present in the pretectal nuclei, including the anterior, posterior and medial nuclei, as well as the nucleus of the posterior commissure (Fig. 6a–d). Terminals were also observed in thalamic nuclei, in agreement with previous studies (Fig. 6a–d) (Power et al. 1999).

Fig. 2 Chartings of the distribution of labeled cells (dots), axons (lines), and terminals (stipple) in the ventral thalamus (d–i) following a unilateral injection of WGA-HRP into the SC (a–c) in a gray squirrel. Note the large numbers of cells in the ventral half of ZI and the terminal field in the dorsal half of ZI



Monkey midbrain tectum injections

The connections of ZI with the midbrain tectum of the monkey will be illustrated by a series of cases. In the first, the WGA-HRP injection site was centered in the pretectum, including the anterior and posterior pretectal nuclei, portions of the nucleus of the optic tract, and nucleus of the posterior commissure. This injection spreads into the rostral SC and included portions of SGI (Fig. 7a–c). Retrogradely labeled neurons (dots) and anterogradely labeled terminals (stipple) were present throughout ipsilateral ZI, with no regard to lamination (Fig. 7d–i). An occasional labeled cell was observed in contralateral ZI in this and other cases, but we did not pursue this question due to their small number. Note that the band of labeled cells and terminals extends dorsolaterally to sit medial to the thalamic reticular nucleus (Fig. 7d–h). The morphology of the labeled incertotectal neurons labeled in this same case is demonstrated in Fig. 8. As was seen in the squirrel, the

majority of the labeled cells had fusiform somata, whose long axes, as well as primary and secondary dendrites, were oriented parallel to the mediolateral axis of ZI (Fig. 8a, b, cells G, H, and K–M). This orientation was less evident medially (Fig. 8b, cells N–Q). The cells located in dZI were generally smaller (Fig. 8b, cells F and I–K) than those located in vZI (Fig. 8b, cells G, H, L–N, and Q), with a few notable exceptions (Fig. 8b, cell O). The cells located in the dorsolateral extension of ZI (dlZI) were also smaller, and their dendrites were oriented parallel to the axis of this part of the nucleus (Fig. 8b, cells A–E).

A different pattern was observed when the WGA-HRP injection site was confined to the SC. In the illustrated example, the tracer lay primarily in lateral SGI, with some spread into more dorsal layers (Fig. 9a–c). There was no spread to the pretectum. In this case, the retrogradely labeled cells (dots) were largely confined to the ventral half of ZI (Fig. 9e–h), although this lamination was less evident at the very rostral and caudal poles of the nucleus (Fig. 9d,

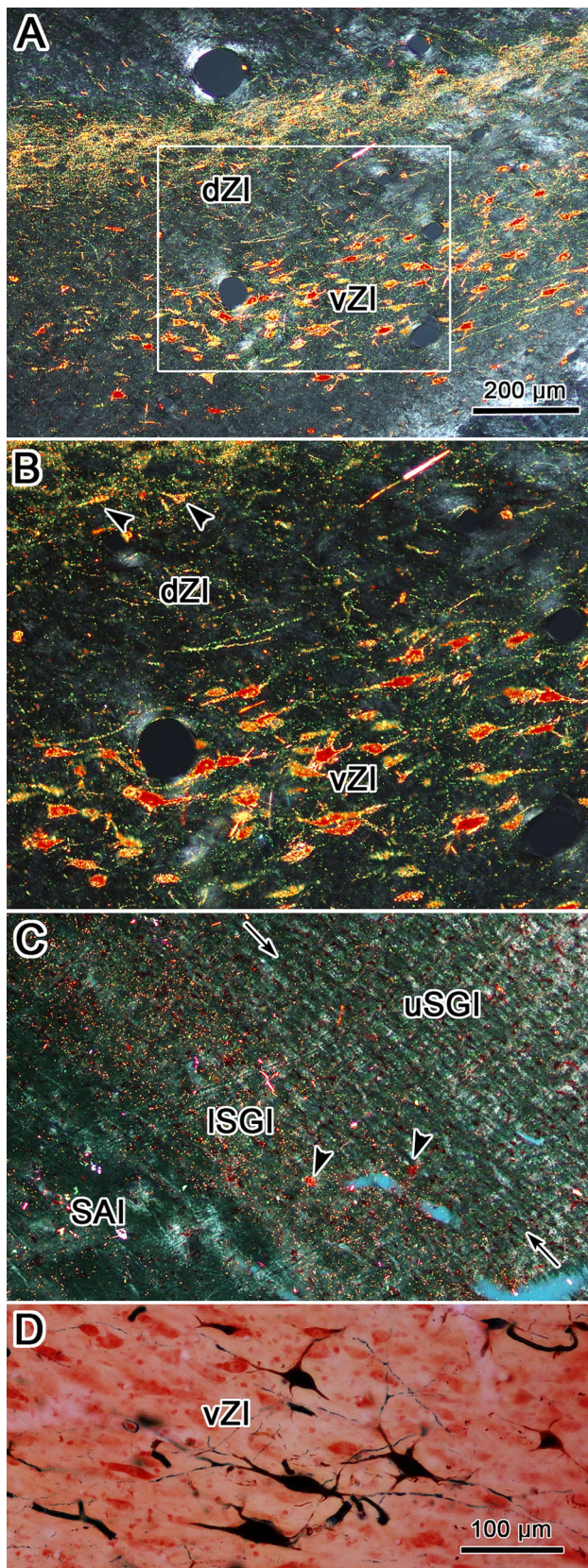


Fig. 3 Images of labeled tectoincercal and incertotectal elements in the gray squirrel. **a** Appearance of ZI following an injection of WGA-HRP into the SC. Note the band of terminals in the upper part of the dorsal layer of ZI (dZI) and the band of labeled cells filling the ventral layer of ZI (vZI). *Box* indicates the region shown at higher magnification in **b**, where the multipolar nature of the incertotectal cells in vZI can be appreciated. In addition, the scattered, more lightly labeled, cells in dZI are indicated by *arrowheads*. **c** Numerous scattered puncta are present in the lower sublamina of the intermediate gray layer (ISGI) of the squirrel following an injection of WGA-HRP into the zona incerta (see Fig. 6). Retrogradely labeled tectoincercal cells (*arrowheads*) are also present. Relatively little label is seen in either the upper sublamina of SGI (uSGI) or the intermediate white layer (SAI). **d** The dendritic morphology of incertotectal cells is evident following injections of HRP in saponin into the SC. Images **a–c** were taken with crossed polarizers. Scale in **c** = **a**, in **d** = **b**

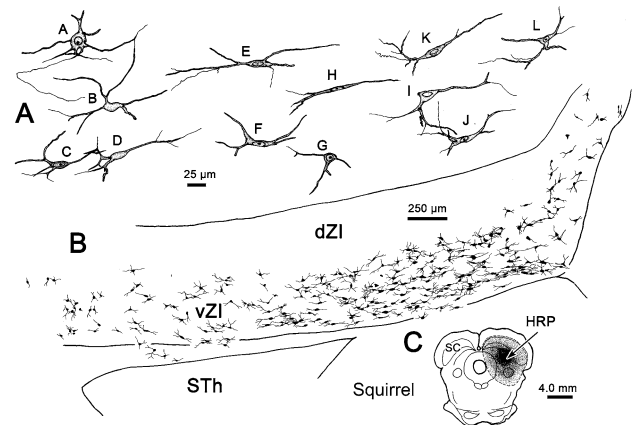


Fig. 4 Dendritic distribution and somatic morphology of incertotectal neurons in the gray squirrel following a large injection of HRP and saponin into the SC. **c** Injection extended into the adjacent periaqueductal gray and midbrain reticular formation. Labeled somata and dendrites were arrayed in vZI, not dZI (**b**). **a** Most somata were fusiform and multipolar, with dendrites that were typically oriented horizontally. This orientation was less evident at the medial end of the layer, where the dendrites showed no orientation, and the lateral end of the layer where it turned in a dorsolateral direction

i, respectively). A distinct band of labeled terminals (stipple) was present in the dorsal half of ZI, along with scattered retrogradely labeled cells (Fig. 9e–g). Scattered labeled puncta were also present in the ventral half of ZI. Labeling also extended dorsolaterally in this case, but it consisted mainly of terminals found in islands of neuropil within the external medullary lamina (Fig. 9e–g). The appearance of ZI labeling following a collicular WGA-HRP injection in another case is shown in Fig. 10a. Numerous labeled cells occupy vZI, but only scattered examples are seen in dZI (blue arrows). Both dZI and vZI contain many small terminal puncta. To determine whether

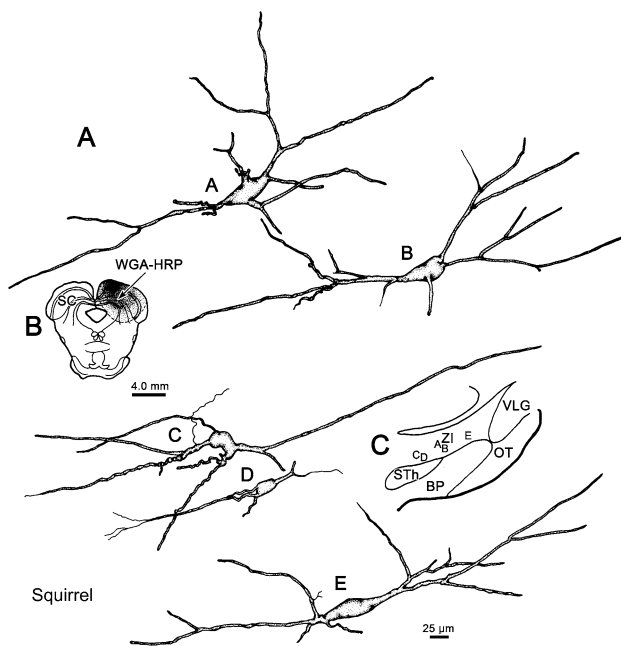


Fig. 5 Detailed morphology of incertotectal neurons in the gray squirrel following a large injection of WGA-HRP into the SC. **b** Injection site in this case only extended slightly into the periaqueductal gray. **a** Somata of labeled incertotectal cells (cells A–E) are typically fusiform in appearance. The dendrites branch sparsely and even the tertiary dendrites extended in the plane of the nucleus. The locations of the labeled cells in **a** are indicated in **c**

these terminal puncta actually represent axon terminals, we examined cases in which BDA was injected into the SC. Figure 10c reveals that BDA labeled terminal fields were located in both dZI and vZI. The latter could be observed to have close associations (arrowheads) with retrogradely labeled incertotectal neurons (Fig. 10d).

The morphology of macaque incertotectal neurons is demonstrated in another case with a collicular injection of WGA-HRP. This was a larger injection that filled much of the SC and extended into the periaqueductal gray and adjacent mesencephalic reticular formation (Fig. 11c). The retrogradely labeled neurons were arrayed in vZI and extended into dlZI (Fig. 11b). As in the squirrel, the cells and their dendrites tended to be oriented parallel to the mediolateral axis of the nucleus, and the primary and secondary dendrites were constrained within the layer. There was some range in the size of the somata, with the smaller cells having long axes of around 20 μm and the largest cells having long axes of around 40 μm (Fig. 11a). The cells in dlZI tended to be smaller (Fig. 11a, cells H–J), although some small cells inhabited vZI (Fig. 11a, cell G).

A different pattern of labeling was present when the WGA-HRP injection was located solely in the pretectum, without involving the SC (Fig. 12). In this case, the injection involved the rostral pole of the posterior pretectal nucleus, the anterior pretectal nucleus, and the nucleus of

the posterior commissure (Fig. 12b, c). It spread rostral and dorsal to the pretectum to involve a small portion of the dorsal thalamus (Fig. 12a, b). This injection site produced considerable terminal labeling (stipple) throughout the dorsal half of ZI (Fig. 12e–h). Numerous retrogradely labeled cells (dots) were found in this same layer. The band of label extended dorsolaterally to wrap around the lateral aspect of the dorsal thalamus. At the rostral and caudal pole of ZI, the layering pattern was not as discrete (Fig. 12d, i, respectively). The appearance of labeling in this case is further demonstrated in Fig. 10e, f. In the main body of ZI, most of the labeled cells (blue arrows) and labeled puncta were found in dZI. Both types of label were less evident in vZI (Fig. 10e). Retrograde and anterograde labeling were also evident in the dlZI (Fig. 10f). To determine whether these labeled puncta were actual axonal boutons, we examined cases with BDA injections of the pretectum. As shown in Fig. 10b, labeled axons with bouton enlargements were observed throughout ZI, but these were much more evident in dZI.

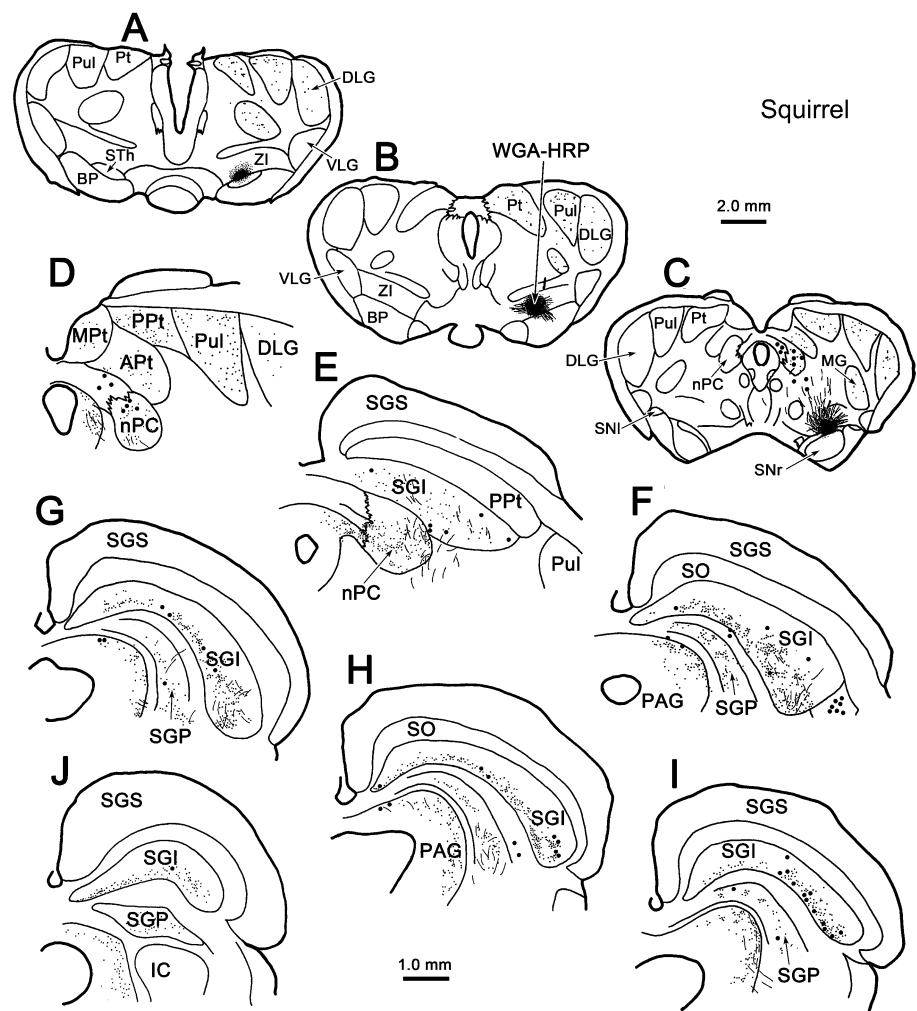
Monkey zona incerta injections

The pattern of incertal connections with the midbrain tectum was investigated by making WGA-HRP injections into the ZI of monkeys. In the case illustrated in Fig. 13, the tracer was centered in ZI, but spread into the adjacent subthalamic nucleus, the external medullary lamina, and internal capsule (Fig. 13a, b). There was also some spread along the needle track through the dorsal thalamus and slight involvement of the dorsolateral corner of the substantia nigra. Within the SC, retrogradely labeled neurons (dots), were mainly located in SGI, although cells were found in SO and SGP (Fig. 13e–j). The labeled cells in this case were relatively small, multipolar neurons with roughly spherical somata having diameters in the 11–15 μm range (Fig. 10g). Labeled terminals were also present (Fig. 13e–j). These were somewhat sparse, but appeared to be more concentrated in the lower sublamina of SGI. Within the pretectum, labeled cells and terminals were observed in the anterior and posterior pretectal nuclei, nucleus of the posterior commissure, and the nucleus of the optic tract (Fig. 13c–f).

Discussion

The data presented here suggest that certain properties of the incertotectal circuitry are common mammalian features, and these are illustrated in Fig. 14. First, the incertotectal projection to the ipsilateral SC originates in vZI. While its laminar terminal distribution differs somewhat between species, its distribution correlates with the location

Fig. 6 Incertotectal termination in the squirrel SC. Chartings show the distribution of labeled cells (*dots*), axons (*lines*), and terminals (*stipple*) in the SC (**d–j**) following an injection of WGA-HRP into ZI (**a–c**). Most of the collicular labeling is found in the lower sublamina of the intermediate gray layer (SGI), but some is also present in the deep gray layer (SGP)



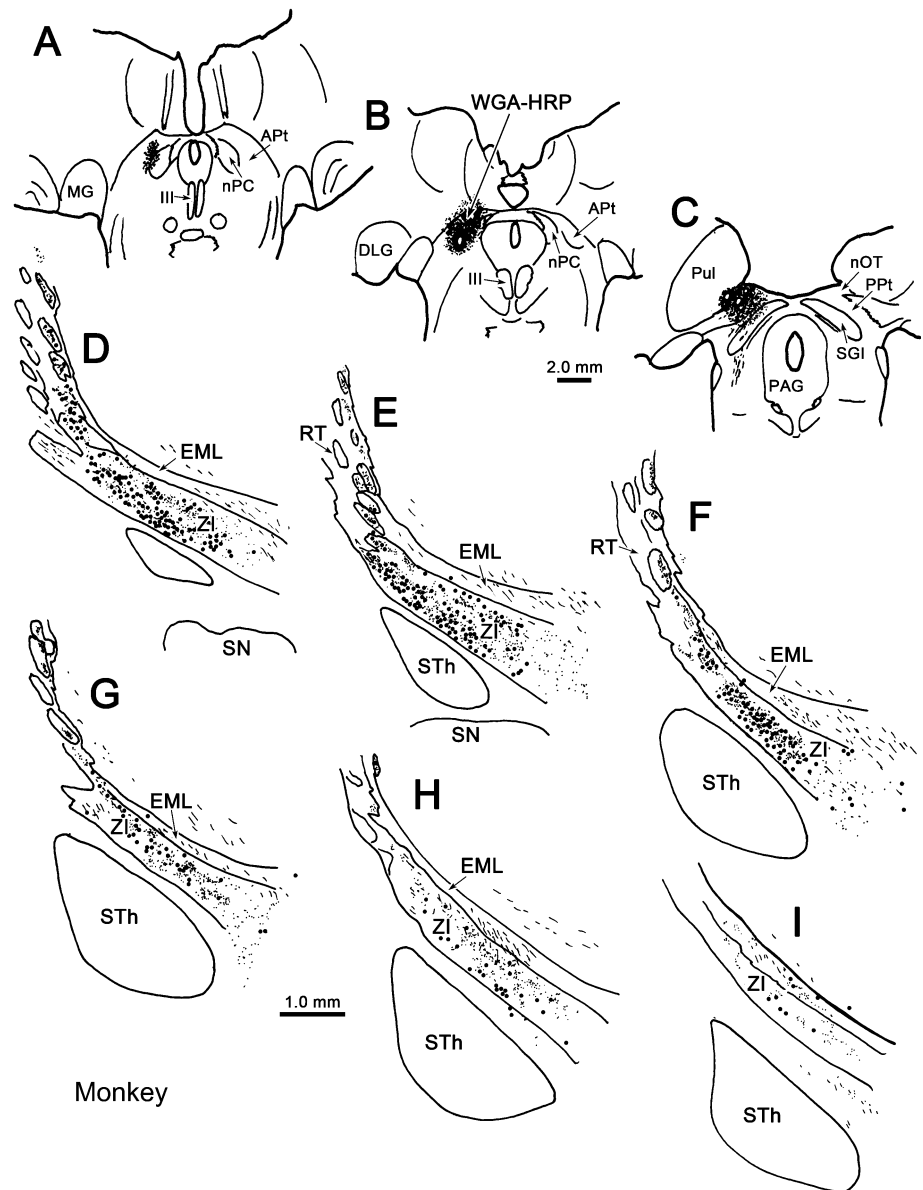
of the cells of origin of the crossed descending output of the colliculus in all species tested. Second, the tectoincertain projection targets both layers of the ipsilateral ZI. It originates from a population of smaller cells, whose distribution within the SC co-localizes with that of collicular saccade-related output cells. Third, the incertopretectal projection derives from cells in ipsilateral dZI. Similarly, the return projection from the pretectum preferentially targets the ipsilateral dZI. These patterns of ZI connectivity support the contention that the main body of this nucleus should be divided into dorsal and ventral layers. The presence of a dorsolateral extension of ZI in the monkey, like that seen in the cat, suggests this characteristic becomes more prominent in animals which have larger, more complex diencephalons.

Technical considerations

Extensive labeling of cells in vZI was a feature of all animals that had retrograde tracer placed in their SC, and the fact that animals with ZI injections displayed terminals

in the SC supports this finding. The sparse nature of the terminal labeling in the colliculus following ZI injections was surprising, considering the degree of retrograde labeling following collicular injections, but this finding is consistent with data from previous studies (Ricardo 1981; May et al. 1997). Several factors may play a role in the limited terminal labeling. First, due to its plate-like shape, the ZI injections only involved a portion of the nucleus. Second, individual incertotectal cells may project broadly, but sparsely to the SC. Third, the incertotectal projection may represent collateral projections of axons supplying other targets. Such collateral projections do not display very effective anterograde labeling with WGA-HRP. While there is evidence that many incertal cells supply a single target (Power and Mitrofanis 1999), it appears that vZI cells project to both the thalamus and the SC (Urbain and Deschênes 2007). In the rodent, there is strong evidence for topography in the ZI projection to SC (Kim et al. 1992). No clear topography was noted in the present study, but the large injection sites employed may have masked its presence.

Fig. 7 Overall pattern of connections between ZI and midbrain tectum in the monkey. Chartings show the distribution of labeled cells (*dots*), axons (*lines*), and terminals (*stipple*) in the ZI (**d–i**) following an injection of WGA-HRP into pretectum (**a–c**) and rostral SC (**c**). Labeled cells and terminals were present throughout ZI and this pattern of labeling extended medial to the reticular thalamic nucleus (RT)



The tectoincertal projection originated in a set of small cells scattered within the layers beneath SGS in both species. It is possible that these cells were labeled by spread into the substantia nigra, as tectal cells with similar morphologies and distributions supply this nucleus (McHaffie et al. 2006; May et al. 2009). However, since tectal injections of both WGA-HRP and BDA produced terminals in ZI, it seems more reasonable to suspect that the same population of cells provides input to both the ZI and the nigra. A pretectoincertal projection was also observed. The anterograde evidence for this projection was only available in the monkey, but it was confirmed by the monkey and squirrel ZI injection cases, which retrogradely labeled cells in the pretectal nuclei.

The incertopretectal projection was only examined in detail in monkeys. A clear pattern of increased cell labeling was observed in dZI with injections that included the pretectum. Pretectal injections are more likely to have involved dorsal thalamic structures, so the possibility exists that some of the cells labeled in dZI might actually be projecting to the thalamus (Watanabe and Kawana 1982; Power et al. 1999). On the other hand, the fact that ZI injections produced terminal labeling among the pretectal nuclei supports the presence of an incertopretectal projection. Incertopretectal terminal labeling was relatively sparse, possibly for the same reasons as noted above for the incertotectal projection.

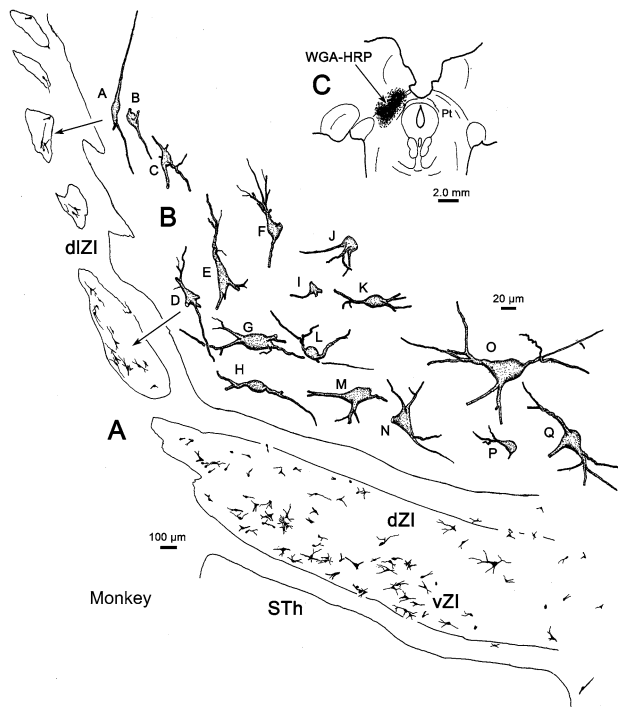


Fig. 8 Morphology of incertotectal neurons in a macaque monkey following a large injection of WGA-HRP into the pretegmentum and rostral SC. This is the same case, as charted in Fig. 7. The injection shown in **c**. **a** Labeled cells were arrayed throughout dZI and vZI. **b** Cells were multipolar with somata and dendrites that were typically oriented horizontally in the main body of the nucleus (cells G–N). This orientation was less evident at the medial end of the layer (cells O–Q). At the lateral end of the layer, where it extended into dlZI, the primary orientation of the dendritic fields is also dorsolateral (cells A–F). The cells in dZI (cells I–K) tended to be smaller, with a few notable exceptions (cell O)

Incertain subdivisions

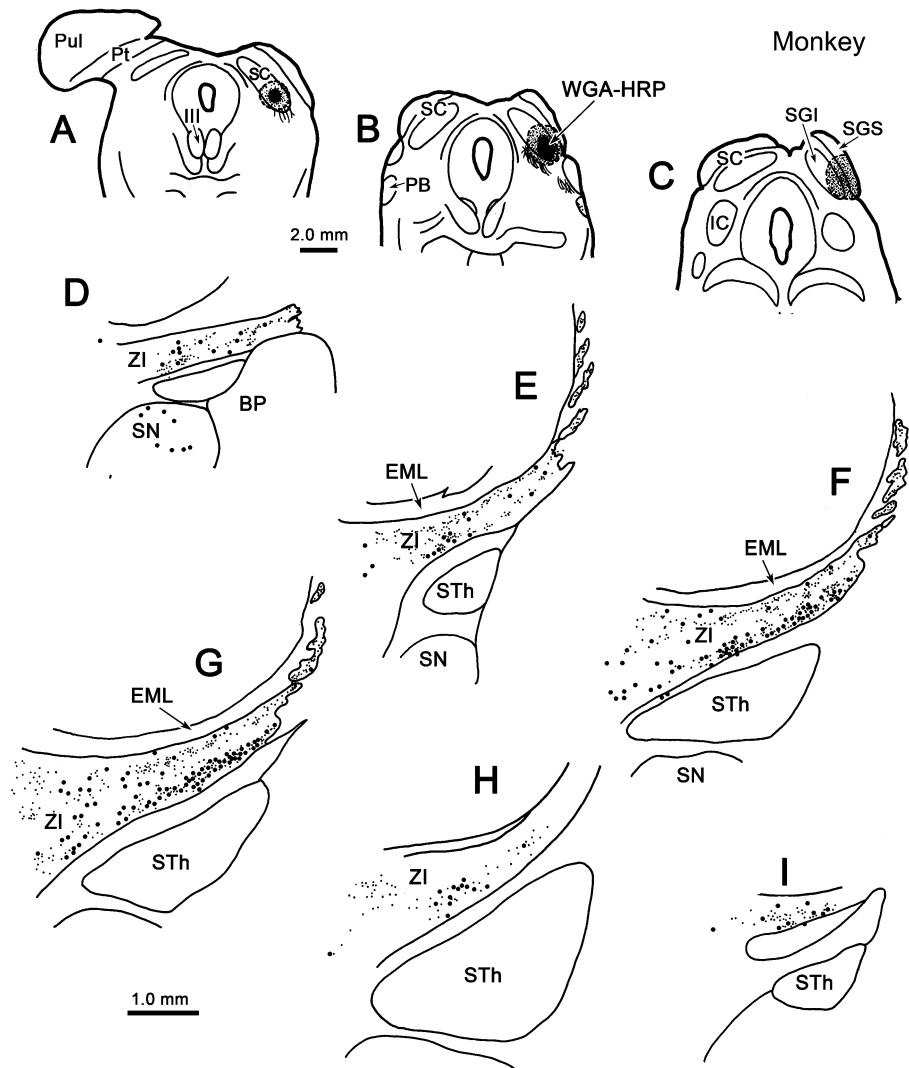
The rat ZI has been subdivided into up to six parts (Kuzemský 1977; Watanabe and Kawana 1982). Here, we have adopted a simple division into two layers: dZI and vZI. This approach works well with respect to the patterns of connectivity of this structure with the midbrain tectum (rat: Kim et al. 1992; cat: May et al. 1997; Perkins et al. 2006). We also noted that the dendritic fields of the labeled cells (Figs. 4, 5, 8, 11), which were elongated in the mediolateral dimension and similar to the cells classified as principal neurons in Golgi preparations (Ma et al. 1992), appeared to largely respect the boundary between these two layers, in agreement with previous rodent studies (Trageser et al. 2006; Urbain and Deschênes 2007). Furthermore, physiological studies indicate that dZI and vZI cells have different physiological characteristics (Trageser et al. 2006; Barthó et al. 2007). Likewise, the connections of these layers differ. In addition to the SC, vZI projects to the lateral dorsal, lateral posterior, and posterior nuclei of the

thalamus (Power and Mitrofanis 1999). This layer receives preferential input from frontal and parietal cortices, as well as the interposed nucleus (rat: Mitrofanis and Mikuletic 1999), and from the frontal eye fields (cat; Perkins et al. 2006). In addition to targeting the pretegmentum, the dorsal ZI projects to the midbrain and pontine reticular formation, periaqueductal gray, pedunculopontine nuclei and parafascicular nucleus (Kolmac et al. 1998; Power and Mitrofanis 1999), as well as to cerebral cortex (Nicolelis et al. 1995; Lin et al. 1997). It receives input from these same brainstem nuclei, as well as from the cingulate cortex (Kolmac et al. 1998; Mitrofanis and Mikuletic 1999). In the present study, this division into layers was not clear at the rostral and caudal poles of ZI. Consequently, others have chosen to define these poles as separate regions of ZI (rat: Watanabe and Kawana 1982; Kolmac et al. 1998; Mitrofanis and Mikuletic 1999; monkey: Ma et al. 1992). We did not do this since the bands of labeled cells continued into these regions.

One of the features that we observed in the ZI of the macaque is a dorsolateral extension (dlZI) that runs from the main nucleus around the lateral edge of the dorsal thalamus. In this region, ZI is arranged as islands of neuropil within the external medullary lamina. This area is commonly included in the thalamic reticular nucleus. However, in view of the fact that bands of label continue from ZI proper into dlZI after both tectal and pretectal injections, we believe that it represents a part of ZI and has the same function. Within dlZI, a division into layers is not evident, but this may be due to its narrow width. The presence of this extension was also observed in the cat (Clemence and Mitrofanis 1992; May et al. 1997; Kolmac and Mitrofanis 1999; Perkins et al. 2006), but not the rat (Watanabe and Kawana 1982; Mitrofanis 2005). We did not observe this extension in the squirrel, but did note a change in the orientation of the incertotectal cell dendrites at the lateral edge of ZI, reminiscent of that seen in the macaque dlZI. In cats and primates, there is considerable expansion and rotation of the elements of the diencephalon that parallels the expansion of the cortex. For example, the rodent ventral lateral geniculate nucleus, sitting beneath the dorsal lateral geniculate nucleus, becomes the macaque pregeniculate nucleus, sitting above the dorsal lateral geniculate. It seems reasonable to propose that this process has pulled a portion of the ZI in a dorsolateral direction in these species.

One characteristic of these layers is that the two projections to ZI shown here terminate in both of them. The presence of separate projections targeting these two layers is a feature of many ZI afferents (Mitrofanis and Mikuletic 1999; Mitrofanis and deFonseka 2001; Power et al. 2001; Shaw and Mitrofanis 2002; Perkins et al. 2006; Simpson et al. 2008). Since the ventral layer provides a

Fig. 9 Pattern of collicular connections with ZI in the monkey. Chartings show the distribution of labeled cells (*dots*), axons (*lines*) and terminals (*stipple*) in the zona incerta (**d–i**) following an injection of WGA-HRP confined to the SC (**a–c**). Note the large numbers of cells in the lower half of ZI and numerous terminals in the upper half of ZI. This labeled band extended dorsolaterally, in islands of neuropil



predominantly inhibitory output and the dorsal layer provides a predominantly excitatory output [except the GABAergic projection to cortex (Lin et al. 1990, 1997; but see Shaw et al. 2013)], the ZI may be in position to act as a multiplex switch—turning the targets of dZI on, while it turns the targets of the vZI off in response to these bilaminar inputs (Fig. 14). Perhaps, it is this capability that makes deep brain stimulation of ZI effective in relief from the symptoms of a variety of motor diseases, including Parkinsons, Tourettes, and multiple sclerosis (Périer et al. 2000; Babel et al. 2001; Nandi et al. 2002; Plaha et al. 2006).

This leads to the question of what information is supplied by these midbrain inputs. The SC and posterior pretectum both contain cells that display saccade-related signals (for review: Sparks and Hartwich-Young 1989; Gamlin 2006). In addition, tonically firing cells that pause for saccades in all directions are found within the primate nucleus of the optic tract (Mustari et al. 1997).

Omnidirectional pause cells have also been described in ZI (Ma 1996), along with cells that stop their tonic discharge for saccades made to visual targets and spontaneous saccades made in the dark (Hikosaka and Wurtz 1983a). Since the connections between the ZI and both the pretectum and colliculus are reciprocal, we cannot specify in which direction the information is passed, but it seems more likely that any saccade-related activity changes seen in the ZI would be due to inputs from the SC and pretectum. Unfortunately, the ZI layer that contains saccade-related cells has not been determined, but the fact that they are tonically active suggests that they are inhibitory neurons in vZI (Trageser et al. 2006). We will further discuss the roles that are suggested by the connections of each layer, below.

Dorsal zona incerta

Our results indicate the pretectum projects to ZI in the monkey, in agreement with previous studies (monkey:

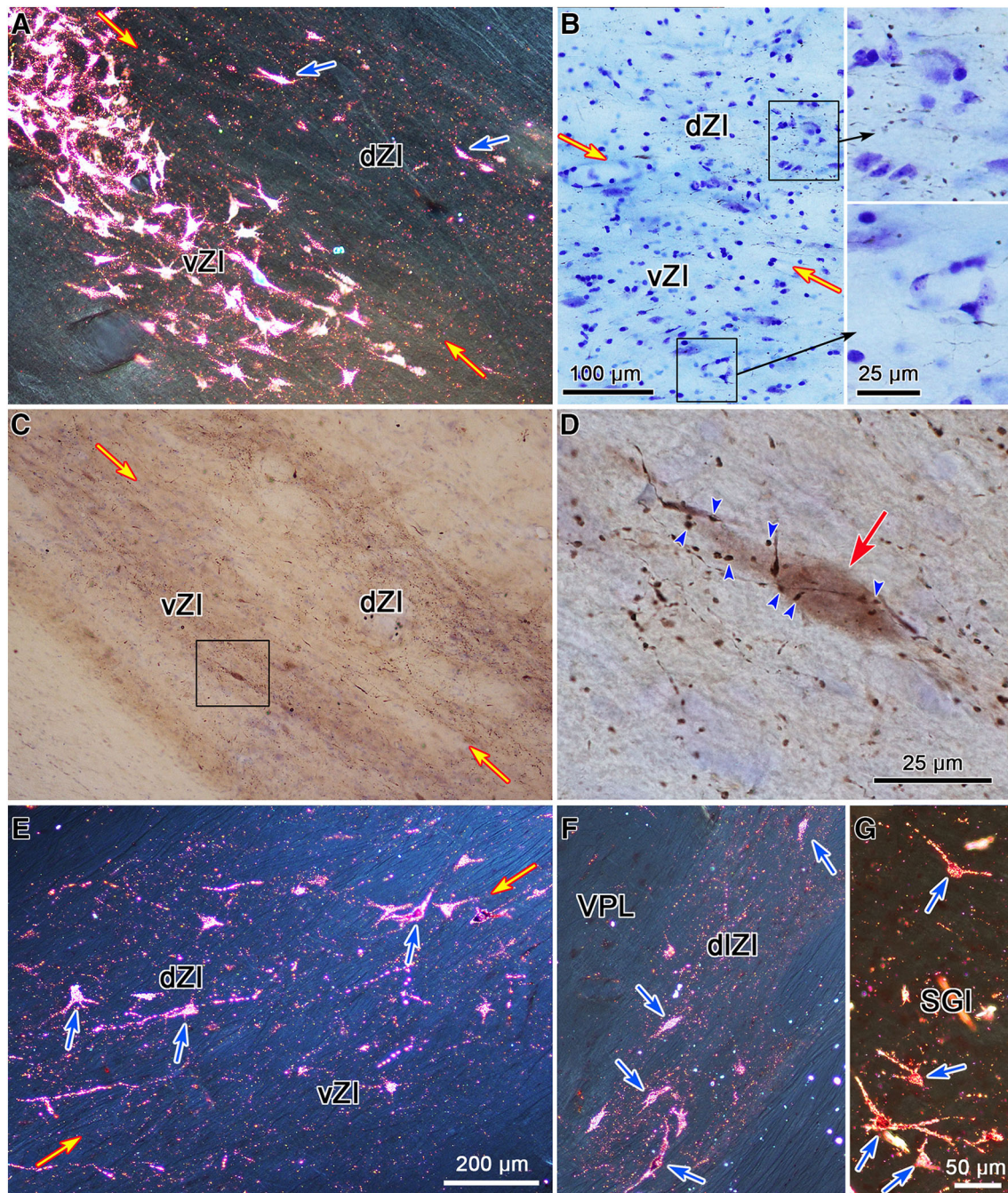


Fig. 10 Images of labeled tectoincercal, incertotectal, and pretectoincercal elements in the macaque monkey. **a** Appearance of ZI following an injection of WGA-HRP into the SC. Note the band of labeled cells filling vZI and the terminal puncta in dZI. Scattered, more lightly labeled cells in dZI are indicated by *blue arrows*. **b** Labeled axonal arbors following a BDA injection of the pretectum. *Boxes* indicate regions of dZI and vZI shown in the high magnification *insets*. Terminations are much denser in dZI. **c** Numerous labeled axonal arbors are present in both dZI and vZI following an injection of BDA into the SC. *Box* indicates area shown at higher

magnification in **d**, where numerous labeled boutons are shown in close association (*arrowheads*) with a retrogradely labeled incertotectal neuron (*red arrow*). Terminal puncta are present in both dZI and vZI (**e**) and in the dorsolateral extension of ZI (dlZI) (**f**) following a WGA-HRP injection of the pretectum (shown in Fig. 12). Most of the labeled cells (*blue arrows*) are found in dZI (**e**) and dlZI (**f**). **g** Labeled neurons (*blue arrows*) are present in the intermediate gray layer (SGI) following an injection of WGA-HRP into ZI (shown in Fig. 13), along with scattered terminal puncta. Images **a** and **e–g** were taken with crossed polarizers. *Scale* in **e** = **a** and **f**, in **g** = **c**

Harting et al. 1980; Benevento et al. 1977; Mustari et al. 1994; Büttner-Ennever et al. 1996; cat: May et al. 1997) and preferentially terminates in dZI (Fig. 14). A similar

pretectoincercal projection has been seen in the cat, although the terminal field was densest along the border of dZI and vZI (Berman 1977; May et al. 1997). The location

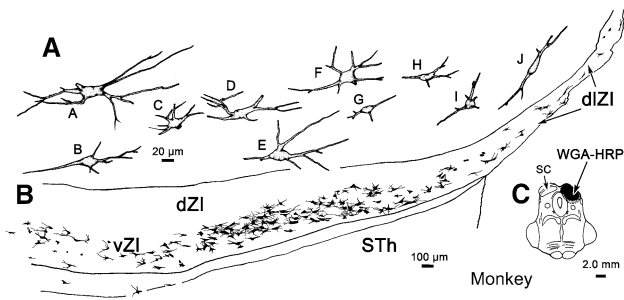
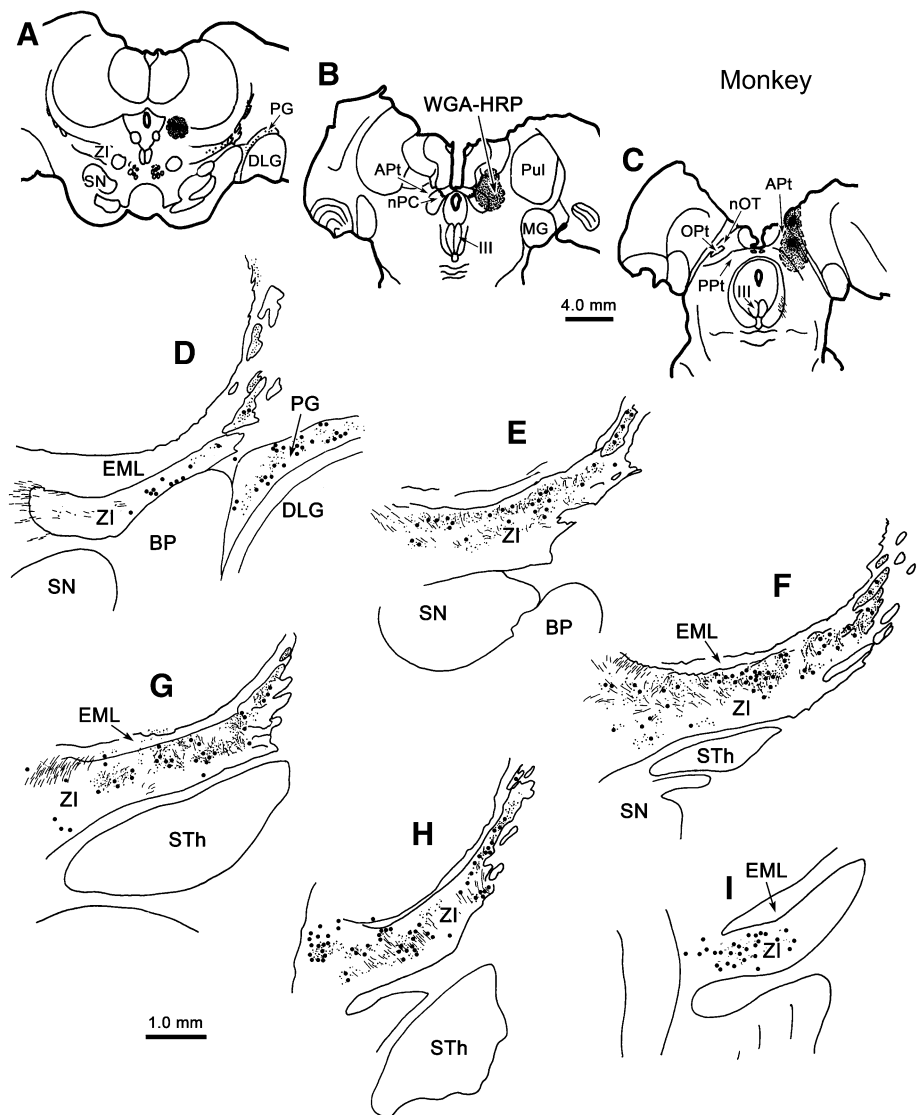


Fig. 11 Morphology of intertectal neurons in a macaque monkey following a large injection of WGA-HRP into the SC. The injection shown in **c** spreads slightly into the periaqueductal gray. **b** Labeled cells were arrayed throughout vZI, but not dZI. **a** Cells were multipolar with somata and dendrites that were typically oriented horizontally in the main body of the nucleus (cells B–H). This orientation was less evident at the medial end of the layer (cell A). Within dZI, the primary orientation of the dendrites was dorsolateral (cells I–J)

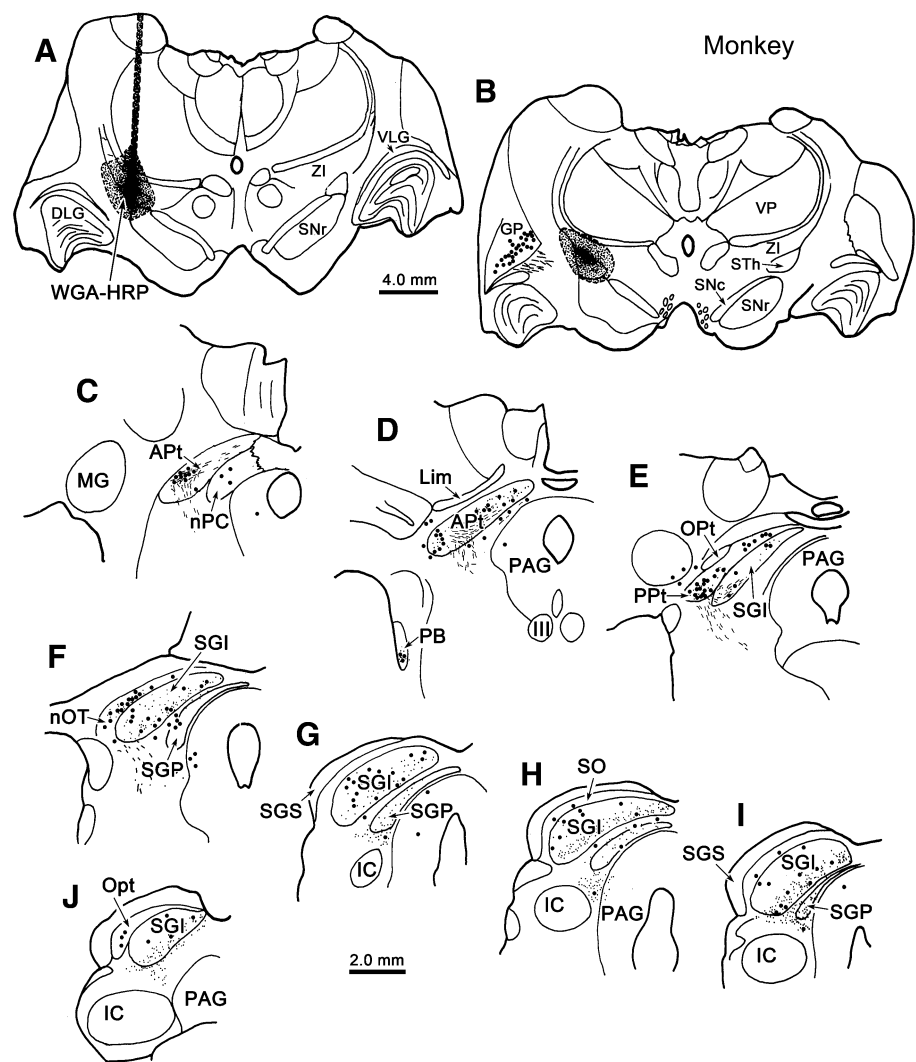
Fig. 12 Pattern of pretectal connections with ZI in the monkey. Chartings show the distribution of labeled cells (dots), axons (lines), and terminals (stipple) in ZI (**d–i**) following an injection of WGA-HRP confined to the pretectum (**a–c**) in a macaque monkey. The injection site involved the anterior and posterior pretectal nuclei, nucleus of the posterior commissure, and nucleus of the optic tract. Note the large numbers of cells and terminals in the dorsal half of ZI. These extended dorsolaterally, in islands of neuropil



of the cells of origin for this projection is fairly diffuse. In the cat and rat, most of the cells labeled retrogradely from ZI injections were located in the anterior and posterior pretectal nuclei and in the nucleus of the optic tract (rat: Roger and Cadusseau 1985; cat: May et al. 1997). Labeled cells were found in these nuclei in the present experiments, as well as in the nucleus of the posterior commissure. In the rat and cat, the largest number of cells was found in the anterior pretectal nucleus (Roger and Cadusseau 1985; May et al. 1997), but this was not true of the monkey and squirrel. In the rat, the anterior pretectal nucleus projects mainly to vZI (Giber et al. 2008), so it is likely that the pretectal terminals observed in the macaque dZI are from the other pretectal nuclei.

The dZI projects throughout much of the neuraxis, including the brainstem, dorsal thalamus, and cerebral cortex (Watanabe and Kawana 1982; Ricardo 1981;

Fig. 13 Incertotectal termination in the monkey midbrain tectum. Chartings show the distribution of labeled cells (*dots*), axons (*lines*), and terminals (*stipple*) in the SC (e–j) and pretectum (c–f) following an injection of WGA-HRP into ZI (a–c) in a macaque monkey. Most of the terminal labeling is found in the intermediate gray layer (SGI) and deep gray layer (SGP). Labeled cells were scattered in these layers, as well as in stratum opticum (SO). Labeled terminals and cells were present in the anterior pretectal nucleus (APt) (c, d), posterior pretectal nucleus (PPt) (e), nucleus of the posterior commissure (nPC), and the nucleus of the optic tract (nOT) (f)



Romanowski et al. 1985; Power et al. 1999). The present results indicate that the dZI also provides a projection back to the pretectum. A similar projection was observed in the rat and cat (Ricardo 1981; May et al. 1997). Thus, dZI is in a position to modulate activity in a wide variety of structures. Mitrofanis (2005) has suggested that the dZI plays a role in arousal, based on its projection to the intralaminar nuclei. In agreement with this, incertal neuron activity is inhibited by ascending cholinergic inputs (Trageser et al. 2006). However, Watanabe and Kawana (1982) asserted that the intralaminar projection arises from the vZI and it is the cells in this layer that are more influenced by cholinergic inputs (Trageser et al. 2006). A related possibility is suggested by the dZI inputs from the SC and pretectum shown here. Due to the fact that attention is commonly directed with the eyes, it is believed that the ascending projections of the SC are also utilized to direct attention (for review: Basso and May 2017). In the case of dZI, such information might be used to change activity levels in its

cortical, thalamic, and brainstem targets when interesting stimuli appear.

Alternatively, a sensory, as opposed to a saccade-related, function may be supported by the tectal and pretectal inputs to dZI. It has been suggested that the tectonigral projection may carry multimodal sensory information (rat: Coizet et al. 2003; cat: McHaffie et al. 2006; monkey: May et al. 2009), and, as noted above, the tectoincertal cells resemble these tectonigral cells. Similarly, the pretectum receives both retinal and somatosensory input (cat: Schweigart and Hoffmann 1992; Rees and Roberts 1993; monkey: Hutchins and Weber 1985; Mustari and Fuchs 1990; Hutchins 1991), which it may pass on to dZI. The dZI in rats projects to the substantia nigra, as well as to the pedunculopontine tegmental nucleus and entopeduncular nucleus (Heise and Mitrofanis 2004). Another major target of dZI is the parafascicular nuclei (Power et al. 1999; Power and Mitrofanis 2002), which in turn supplies the striatum. In view of these connections with components of

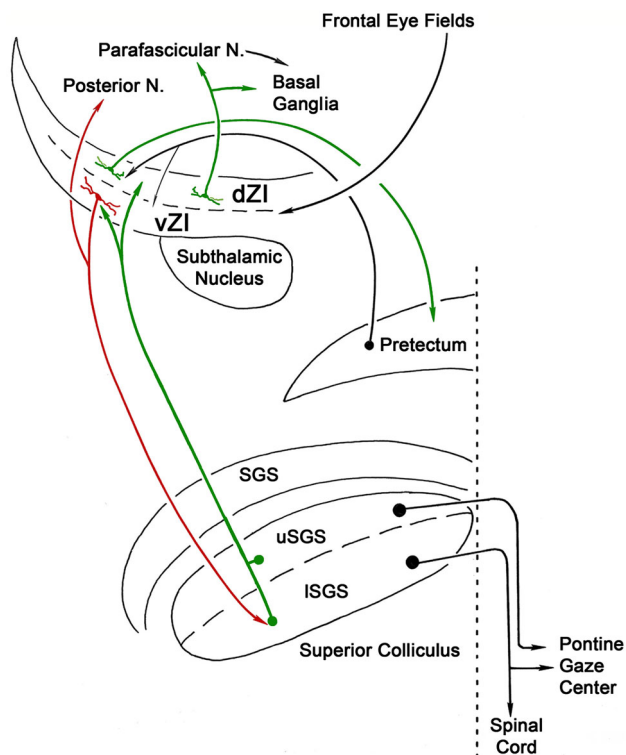


Fig. 14 Circuit diagram of connections between zona incerta (ZI) and the midbrain tectum. The collicular layers that provide gaze-related signals to the brainstem and spinal cord are targeted by a largely inhibitory (*red*) incertotectal projection from the ventral ZI (vZI). These same SC layers provide input to ZI that is denser in the dorsal layer (dZI) than vZI. The projection to vZI terminates on incertotectal neurons. The dZI supplies a largely excitatory (*green*) input to the nuclei of the pretectum. This same layer is the predominant target of inputs from the pretectal nuclei. The two layers have different targets in the dorsal thalamus and receive different inputs, a few of which are indicated

the basal ganglia (Fig. 14), this multimodal excitatory pathway through ZI to the basal ganglia may be important for directing learned and habitual behaviors (McHaffie et al. 2006; May et al. 2009; Watson et al. 2015). The fact that dZI is particularly targeted by cingulate cortex supports this idea (Mitrofanis and Mikuletic 1999).

Ventral zona incerta

We have observed a tectoincertain projection that terminates in vZI in squirrels, monkeys and cats (present results, Fig. 14; May et al. 1997). A similar projection was observed in the rat (Kim et al. 1992; Kolmac et al. 1998). Therefore, it appears that SC information is made available to the incertotectal cells in vZI. Indeed, this input appears to monosynaptically contact these cells (May et al. 1997; Fig. 10d), as well as cells that project to the posterior thalamic nucleus (Watson et al. 2015). In all animals studied to date, the source of this tectoincertain projection

appears to be the deeper, motor output layers of the SC (rat: Roger and Cadusseau 1985; Kim et al. 1992; cat: May et al. 1997). The overlap between this population and the location of the predorsal bundle output cells in the SC is particularly reinforced by the results in the squirrel, where both populations are concentrated in inner SGI (May and Hall 1984; Fig. 6). However, the cells of origin of the tectoincertain pathway appeared to be relatively smaller and more homogeneous (Fig. 10) than the cells projecting in the predorsal bundle (squirrel: May and Hall 1984; monkey: May and Porter 1992), so it seems unlikely that the ZI receives an efference copy of the saccade-related long lead burst output of the tectum. Instead, as discussed above, tectoincertain cells may carry a sensory signal.

A projection from the vZI to the SC has been observed in all species investigated, suggesting that this is a common mammalian feature (rat: Watanabe and Kawana 1982; Romanowski et al. 1985; Kolmac et al. 1998; cat: Edwards et al. 1979; Ficalora and Mize 1989; May et al. 1997; squirrel and monkey: present results). The distribution of incertotectal terminals shows some species variation. The terminal field in the monkey was denser in the lower sublamina of SGI, but sparse terminations were observed in the upper sublamina and in SGP, a pattern also seen in cats (May et al. 1997). In the squirrel, the terminal field was largely restricted to the lower sublamina of SGI and SGP, and appears similar to that described for rats (Ricardo 1981, Kim et al. 1992). In all cases, incertotectal terminal fields are well correlated with the location of the cells of origin of the predorsal bundle (cat: Kawamura and Hashikawa 1978; squirrel: May and Hall 1984; monkey: May and Porter 1992; Robinson et al. 1994; rat: Redgrave et al. 1990). Indeed, Kim et al. (1992) demonstrated synaptic contact by incertotectal terminals onto predorsal bundle cells. Thus, it appears that the crossed descending output pathway from the SC that initiates orienting movements of the eyes and head is targeted by the incertotectal projection (Grantyn and Grantyn 1982; Moschovakis et al. 1988) and that vZI is specifically involved in this function. Since this layer is primarily made up of GABAergic cells (Kolmac and Mitrofanis 1999; Mitrofanis et al. 2004), it is highly likely that the vZI exerts an inhibitory influence over collicular saccade-related activity (Ficalora and Mize 1989; Appell and Behan 1990) (Fig. 14).

As noted in the introduction, inhibition plays many roles in the SC. The role played by the ZI's inhibitory input to SC saccade initiation cells is unknown. In addition to the inputs from the SC and pretectum described here, vZI is targeted preferentially by the trigeminal sensory nuclei and somatosensory cortex (Nicoletis et al. 1992; Mitrofanis and Mikuletic 1999; Shaw and Mitrofanis 2002; Perkins et al. 2006; Simpson et al. 2008). These inputs produce a

topographic body representation within vZI (Nicollelis et al. 1992), although the size of receptive fields is relatively large, and many cells have high tonic activity that is suppressed by stimulation. The inferior colliculus also preferentially projects to a portion of vZI (Mitrofanis 2002b). Taken together, these projections suggest that the incertotectal projection could form a pathway, whereby multi-sensory information modulates the saccade-related activity in the SC. However, vZI also receives additional inputs from the interposed nucleus of the cerebellum and the red nucleus (Mitrofanis and deFonseka 2001; Mitrofanis 2002a). While these inputs do not preferentially involve vZI, they do indicate that its role is unlikely to be purely sensory in nature.

Studies of the vZI projection to the posterior thalamic nucleus in rats may provide insight into the workings of the incertotectal pathway. This thalamic projection, like the one to the SC, is GABAergic (Barthó et al. 2002). Tonically firing inhibitory, incertothalamic projections block the capacity of ascending trigeminal projections conveying vibrissal stimulation to directly activate thalamocortical cells within the posterior nucleus (Trageser and Keller 2004; Lavallée et al. 2005). Furthermore, the activity of these inhibitory cells is actually increased by trigeminal inputs to vZI, so this incertothalamic projection represents a feed forward inhibitory pathway gating the access of somatosensory inputs to the posterior nucleus. This inhibition can be suppressed, allowing vibrissal activation of the posterior nucleus, by stimulating motor cortex. This is due to motor cortex projections that cause tonically active vZI cells to pause (Urbain and Deschênes 2007). The proposed mechanism for this pause involves local connections between GABAergic incertal neurons (Urbain and Deschênes 2007). Indeed, there is considerable evidence for interconnections between incertal neurons (rat: Power and Mitrofanis 1999; cat: May et al. 1997), due to local axon collaterals emitted by ZI projection cells (Urbain and Deschênes 2007). Based on these findings, Urbain and Deschênes (2007) have proposed that when the rat actively investigates an object with its whiskers, motor cortex projections cause their ZI target cells to inhibit incertothalamic neurons. The loss of inhibition in the posterior nucleus then permits ascending sensory information to activate thalamocortical neurons. This model may be applicable to the incertotectal projections. Connections between its GABAergic neurons may explain the pauses observed in ZI neuron activity with respect to saccadic eye movements (Hikosaka and Wurtz 1983a; Ma 1996). Perhaps, based on the pattern of activity coming to vZI, the incertotectal projection helps determine which of the sensory inputs competing to direct the next saccade has access to saccade-related output cells in the intermediate gray layers. In light of the role of motor cortex described above,

it is worth noting that inputs to vZI come from the frontal eye fields (Fig. 14) (cat: Perkins et al. 2006; monkey: Huerta et al. 1986).

Many have suggested that the ZI is closely allied with the hypothalamus, and the borders between these structures are unclear, with some transmitter-specific populations extending between them (see Mitrofanis 2005 for review). Of particular note are studies linking the ZI to ingestion of food, water, and salt (Walsh and Grossman 1973; Huang and Mogenson 1974; Evered and Mogenson 1977; Kendrick and Baldwin 1989). Studies in awake behaving sheep indicate that neurons in the ZI are activated by the sight of approaching food at levels defined by individual preference (Kendrick and Baldwin 1986, 1989). This response extinguishes if the food is not available for consumption and it can be paired to non-food stimuli by classical conditioning. Furthermore, these stimuli produce a release of GABA in the ZI (Kendrick et al. 1991), indicating that it is highly likely that they involve vZI tectoincertain cells. Interestingly, this response is specific to the internal needs of the animal. The ZI in salt-deprived animals only responds to the presence of salt, not food, while the ZI in food-deprived animals responds just to food. Thus, these studies suggest that the incertotectal projection may be crucial to matching the selection of targets for orientation with the internal needs of the animal.

Conventional behavioral testing of collicular function in non-human primates has traditionally involved the presentation of individual neutral targets that have been paired with a sweetened fluid reinforcer to induce the animal to saccade to them. Given the findings described above, it seems possible that vZI could play an important role in the target selection process under such conditions. Furthermore, given its inhibitory nature, the incertotectal projection may be particularly important for suppressing activity related to less attractive targets with respect to the current needs of the individual. The high level of trigeminal input to this circuit in the rat may be due to the important role that the vibrissa play in food identification in this species (Comoli et al. 2003). When we open the refrigerator door, whether we select a food item or a drink is dependent on whether we are thirsty or hungry. Perhaps, the incertotectal pathway is providing crucial information to direct our gaze towards the targets that match our internal needs and away from targets that do not match our needs within a highly complex visual environment.

Acknowledgements This manuscript is dedicated to the memory of Dr. Rick Chia-Sheng Lin, whose infectious enthusiasm for the zona incerta and all his other interests will be sorely missed. We would also like to thank Drs. Susan Warren, Kimberly Simpson and William C. Hall for their thoughtful readings of earlier versions of this work. This work was supported by the National Eye Institute of Health Grant #EY019663 to MAB and PJM.

Compliance with ethical standards

Conflict of interest The authors have no actual or apparent conflicts of interest to report.

Statement on the welfare of animals All applicable international, national, and/or institutional guidelines for the care and use of animals were followed.

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