



HER2 status as a potential predictive biomarker for ovarian clear cell carcinoma

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Abstract

Ovarian clear cell carcinoma (OCCC) is a subtype of ovarian carcinoma characterized by unique biological features and highly malignant characteristics including low chemosensitivity. Therefore, new therapeutic targets are needed. These could include the downstream pathways of receptor tyrosine kinases, especially the human epidermal growth factor receptor 2 (HER2). Our main objective was to characterize the HER2 status using immunohistochemistry (IHC) and FISH on 118 OCCCs, also considering the novel paradigm of HER2-zero and HER2-low status. Other aims included determination of the association between HER2 status and survival, HER2 gene DNA and RNA NGS analysis, HER2 gene expression analysis, and correlation between IHC and gene expression in HER2-zero and HER2-low cases. Cases with HER2 overexpression/amplification accounted for 5.1% (6/118), with additional 3% harbouring HER2 gene mutation. The remaining 112 (94.9%) cases were HER2-negative. Of these, 75% were classified as HER2-zero and 25% as HER2-low. This percentage of HER2 aberrations is significant concerning their possible therapeutic influence. Cases from the HER2-zero group showed significantly better survival. Although this relationship lost statistical significance in multivariate analysis, the results have potential therapeutic significance. HER2 gene expression analysis showed a significant correlation with HER2 IHC status in the entire cohort (HER2-positive vs. HER2-negative), while in the cohort of only HER2-negative cases, the results did not reach statistical significance, suggesting that gene expression analysis would not be suitable to confirm the subdivision into HER2-low and HER2-zero. Our results also emphasize the need for standardized HER2 testing in OCCC to determine the best predictor of clinical response.

Keywords HER2 · Ovarian clear cell carcinoma · Immunohistochemistry · Gynecopathology · Ovarian cancer

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Introduction

Primary ovarian carcinoma continues to be the most lethal malignancy of the female genital tract. Ovarian clear cell carcinoma (OCCC) is a histological subtype of ovarian carcinoma characterized by unique biological features and highly malignant characteristics, the development of which has been closely linked to endometriosis [10]. While in Eastern Asian countries the incidence of OCCC is relatively high (reaching up to 30%), among the European and North American population it represents a rarer malignancy [15, 26, 53]. In comparison to high grade serous carcinoma, patients with OCCC have a lower stage-for-stage survival [46, 54]. Contributing to the adverse outcome is the very low sensitivity of OCCC to standard chemotherapeutic regimes based on platinum derivatives and taxanes [46, 47] which, however, even now represent the mainstay of OCCC treatment. Therefore, new therapeutic targets are needed, but until recently, the research into targeted therapy has been limited by the rarity of the tumour.

The main possible molecular targets for OCCC include the downstream pathways of receptor tyrosine kinases, especially the human epidermal growth factor receptor 2 (HER2) [2]. HER2 is encoded by the *ERBB2* oncogene (Erb-B2 Receptor Tyrosine Kinase 2; OMIM#164870) located at the long arm of chromosome 17q12. The evaluation of HER2 immunohistochemical status for predictive purposes has become the standard of care for early and advanced breast cancer, bringing about a significant decrease in recurrence and mortality [8, 19, 40, 41, 44, 45]. While in the past, HER2-positive breast cancer was associated with a worse prognosis; the emergence of targeted anti-HER2 therapy (such as trastuzumab and pertuzumab) has turned this adverse factor into an advantage, leading to a remarkable improvement in the prognosis of these patients. In the past decade, the use of HER2 targeted therapy has also become a major therapeutic tool for advanced stage gastroesophageal cancer, where the addition of trastuzumab to systemic chemotherapy leads to prolonged survival [3, 4, 21, 27, 28]. Recently, it has also been confirmed that trastuzumab provides both longer progression-free and overall survival in patients with advanced and recurrent HER2-positive uterine serous carcinoma [11, 12].

Recently, the introduction of the new antibody–drug conjugate trastuzumab deruxtecan (T-DXd) showed that not only HER2-positive patients but also HER2-low expressing patients were responsive to this therapy [1, 36–38]. This discovery has sparked new interest in exploring the proportion of patients who can be classified as HER2-low (IHC score 1+, or 2+ without amplification on FISH), who could also benefit from targeted therapies [9, 35].

For OCCC, the need for novel therapeutic approaches is undeniable. The evaluation of HER2 status has so far been

reported in a handful of studies, which are mostly performed on limited cohorts (often with less than 20 included patients) [13, 16, 24, 29, 33, 42, 50, 52]. The reported rate of HER2 overexpression covers a wide range from 0 to 45.6% (with the largest study comprising 95 cases), which clearly indicates that the current data is not entirely reliable [33].

The main objectives of this study were to (i) characterize the HER2 status using immunohistochemistry (IHC) and fluorescent in situ hybridization (FISH) on a large, well-defined cohort of 118 OCCCs, taking into consideration the novel paradigm of HER2-zero and HER2-low status; (ii) analyse the relationship between HER2 status and selected clinico-pathological parameters; (iii) determine the association between HER2 status and patient survival; (iv) perform DNA and RNA NGS analysis of the *HER2* gene; (v) perform *HER2* gene expression analysis and examine the correlation between IHC and gene expression in HER2-zero and HER2-low cases.

Materials and methods

Samples

The study was performed on formalin-fixed, paraffin-embedded (FFPE) tissue blocks, which were sourced from the archives of the participating co-authors from departments from the Czech Republic and Hungary. The selection criteria were set to include all patients diagnosed with a clear cell carcinoma of the ovary available in the participating department's archive. All cases were reviewed by two experienced pathologists (PD, MKB) and only those cases fulfilling the diagnostic criteria of clear cell carcinoma were included in the study ($n = 121$). These criteria included immunohistochemical profile compatible with the diagnosis—positivity of PAX8 and at least one marker of “clear cell” differentiation (HNF1B, AMACR, or napsin A) together with the negativity of WT1. During the study, additional 3 cases were excluded due to unsuccessful FISH, which yielded a study cohort of 118 cases in total.

The clinicopathologic characteristics of the cohort are summarized in Table 1. The main parameters evaluated include tumour stage (TNM and FIGO) and age at the time of diagnosis. The most common disease stage was T1c (34%) and T1a (33%). The median follow-up time was 37 months. During the follow-up period, 21 patients had a local recurrence, 13 developed distant metastases, and 24 patients died (14 of those due to diagnosis).

The haematoxylin and eosin-stained slides of all the selected tumour samples were reviewed, and suitable tumour areas were marked for the removal of individual tissue cores, which were used for the construction of tissue

Table 1 Clinico-pathological characteristics of dataset of 118 OCCC

Characteristic	OCCC
Age (years)	
Range (median)	34–87
Mean/median	61/62
Follow-up (months)	
Range	0–198
Mean/median	45/37
Survival status	
NED	73 (62%)
AWD	19 (16%)
DOD/DTC	14 (12%)
DOC/DUC	10 (8%)
NA	2 (2%)
FIGO	
IA	39 (33%)
IB	1 (1%)
IC	38 (32%)
II	9 (7.5%)
III	21 (18%)
IV	1 (1%)
NA	9 (7.5%)
T stage	
T1a	39 (33%)
T1b	1 (1%)
T1c	40 (34%)
T2	11 (9%)
T3	19 (16%)
NA/Tx	8 (7%)
N stage	
N0	50 (42%)
N1	7 (6%)
NA/Nx	61 (52%)
M stage	
M0	6 (5%)
M1	1 (1%)
NA/Mx	111 (94%)
Local recurrence	
No	81 (69%)
Yes	21 (18%)
NA	16 (13%)
Distant recurrence	
No	89 (75%)
Yes	13 (11%)
NA	16 (14%)

NA data not available, NED no evidence of disease, AWD alive with disease, DOD death of disease, DTC death of treatment complication, DUC death of uncertain cause, DOC death of other cause

microarrays (TMAs). From each tumour donor block two tissue cores (each 2.0 mm in diameter) were taken using the tissue microarray instrument TMA Master (3DHISTECH Ltd., Budapest, Hungary) and the evaluation of the studied cases was performed with the use of TMAs.

Immunohistochemical analysis

Using the constructed TMAs, the immunohistochemical (IHC) analysis was performed on 4- μ m thick sections of FFPE tissue. In cases where the TMA approach was not suitable due to technical difficulties with the sample processing ($n = 12$) whole-tissue sections were used.

The IHC evaluation of the HER2 status was performed using the antibody PATHWAY anti-HER-2/neu (clone 4B5, Roche, Basel, Switzerland). The heat induced epitope retrieval using diluted EnVision FLEX Target Retrieval Solution was used for the pre-treatment of HER2. The detection of the primary antibody was performed using the Ventana BenchMark ULTRA instrument (Roche, Basel, Switzerland) with the Ultraview Universal DAB Detection Kit. HER2 scoring was performed in accordance with the 2018 ASCO Guidelines for breast carcinoma [51]. HER2-positive tumours were defined as tumours with a score of 2+ with FISH-confirmed amplification, or tumours with a score of 3+ (complete, strong circumferential immunoreactivity of > 10% of tumour cells). The group of HER2-negative tumours was further divided into two sub-groups: HER2-zero (score 0) and HER2-low (score 1+, score 2+ without amplification), as previously suggested by other works [1, 36]. The IHC expression was double-blindly evaluated by 2 independent pathologists (MKB, KN).

For the assessment of the clinicopathological characteristics and survival outcomes the cases were categorized as HER2-zero, HER2-low, and HER2-positive.

Fluorescent in situ hybridization (FISH) analysis for HER2

All cases with HER2 IHC 2+ were tested for amplification by FISH using 4- μ m thick whole tissue sections of FFPE tumour tissue and ZytoLight® SPEC ERBB2/CEN 17 Dual Color Probe (cat. No. Z-2077, ZytoVision GmbH, Bremerhaven, Germany) according to the manufacturer's protocol. The scoring of ISH results was performed in accordance with the 2018 ASCO Guidelines for breast carcinoma [51]. A HER2/CEP17 ratio ≥ 2.0 was considered as amplification (positive, group 1). The assessment was carried out by a pathologist experienced with FISH analysis (KN).

Next generation sequencing (NGS) analysis of DNA and RNA

The isolation of nucleic acids from FFPE tumour tissue and the following capture DNA and RNA NGS analyses were performed as described previously, with the focus on *ERBB2* (HER2) [7]. The sequence capture NGS analysis of DNA (DNA NGS) and/or RNA (RNA-Seq) was performed for all qualitatively sufficient cases (DNA NGS: 100/118, 84.7%, and RNA-seq: 103/118, 87.3%). Complete DNA and RNA NGS analysis could be carried out for 92 samples had (same sample set was used for comprehensive genomic and transcriptomic analyses, which is a subject of another study from our group, currently under review).

The demultiplexed RNA-Seq data were analysed using the CLC GW by an in-house pipeline which includes targeted RNA-Seq expression analysis (RNA-Seq Analysis module). The bioinformatics pipeline and module settings are available upon request.

Normalization of the mRNA expression was evaluated as RPKM (reads per kilobase of transcript per million reads mapped) and *VCP*, *SF3B1*, and *ATP5F1B* genes were used as reference.

Statistical analyses

All statistical analyses were performed using the program R (version 4.0.2, <https://www.r-project.org/>), Statistica (TIBCO), and/or CLC (Qiagen; CLC GW). The cases were divided according to the IHC expression of HER2 into 3 groups: (1) HER2-zero (score 0), (2) HER2-low (score 1+, or score 2+ without amplification on FISH), (3) HER2 positive (score 3+, or score 2+ with confirmed amplification on FISH). Correlations between the HER2 status and the clinicopathological characteristics were analysed using the Pearson chi-squared test or Fisher exact test according to the expected values.

Survival analyses were performed with four outcomes—overall survival (OS: the period from the date of diagnosis to the date of recorded death), relapse-free survival (RFS: the period from the date of diagnosis to the date of recurrence/death from diagnosis), local recurrence-free survival (LFS: the period from primary diagnosis till the first local recurrence), and distant metastasis-free survival (MFS: the period from primary diagnosis till the first distant metastasis). The date of diagnosis was the date of the surgical procedure. Survival analyses were plotted using the Kaplan–Meier model, and the differences between curves were tested for significance using the log-rank test. If a patient did not have an event, the case was censored in each analysis to the date of the last known follow-up.

To determinate whether HER2 status is an independent prognostic factor, the multivariate Cox's Proportional

Hazard Ratio Model involving age and FIGO stage as covariates was performed. Using the backward elimination of non-significant effects, a minimal adequate model was achieved.

All tests were two-sided and a *p*-value of less than 0.05 was considered as significant.

Differential expression in two-group module which is implemented in CLC GW was used for differential expression of *ERBB2* in group HER2-positive versus HER2-low/HER2-zero and HER2-low versus HER2-zero. This module is a multi-factorial statistics test based on a negative binomial Generalized Linear Model (the statistical model for this module is thoroughly described in the CLC GW manual—<https://digitalinsights.qiagen.com/technical-support/manuals/>).

Results

Immunohistochemical and FISH findings

The representative examples of the immunostaining results are provided in Fig. 1, while the overview of the HER2 immunostaining results is described in Fig. 2. HER2 immunoreactivity was scored as 0 in 84 cases, as 1+ in 7 cases, as 2+ in 23 cases, and as 3+ in 4 cases. The expression of the HER2 protein was categorized into HER2-positive (6/118, 5%) and HER2-negative (comprising HER2-zero and HER2-low cases, total of 112/118, 95%). In the HER2-negative group, 84/112 (75%) cases were HER2-zero and 28/112 (25%) cases HER2-low.

All 23 cases with an IHC score of 2+ were subsequently tested using FISH. Of these, 2 cases (2/23; 9%) were evaluated as positive, belonging to the group of classic HER2 amplified cancer (group 1). The remaining 21 cases were classified as negative (classic non-HER2 amplified cancer, group 5). There were no cases showing monosomy, co-amplification (previously polysomy 17), or borderline/equivocal results.

Prognostic significance of HER2 IHC status

The main evaluated clinicopathological parameters are summarized in Table 2 and included age, FIGO, TNM stage, and local and distant recurrence. None of the observed parameters showed any significant correlation with the HER2 IHC status.

To investigate the prognostic value of the HER2 IHC status, we performed a time to event analysis (OS, RFS, LFS, and MFS) in a 3-tier model (using all three categories HER2-zero, HER2-low, and HER2-positive), and also in a 2-tier model excluding the HER2-positive cases (using only the categories of HER2-zero and HER2-low, given the limitations of the small number of cases in the HER2-positive group).

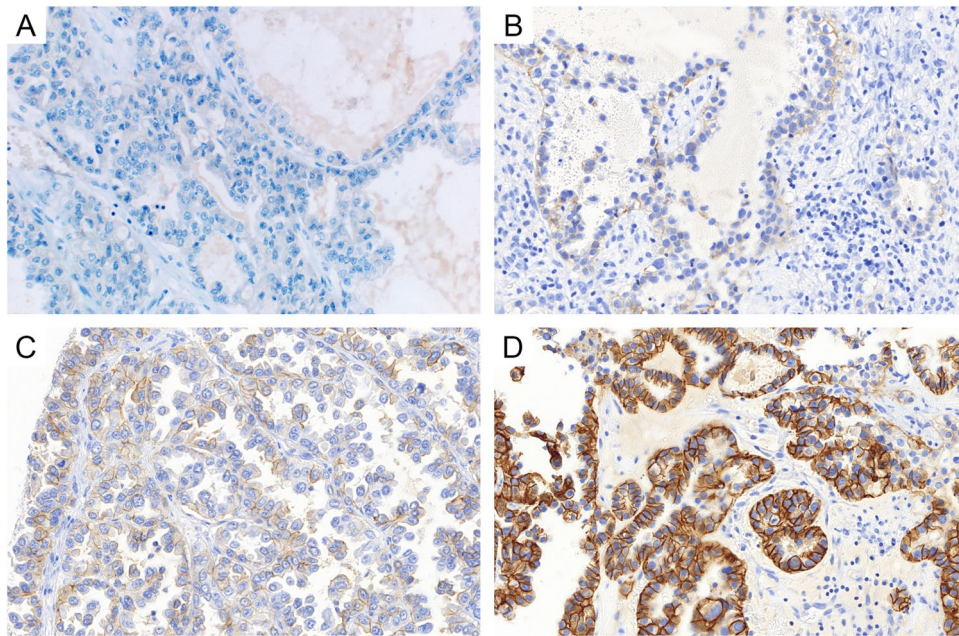
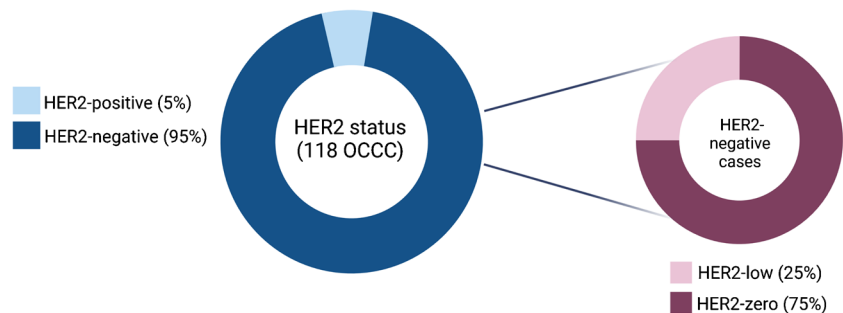


Fig. 1 The representative examples of the immunostaining results of 118 ovarian clear cell carcinomas using the HER2 antibody. All microphotographs are taken at $\times 200$ magnification. **A** Negative result with complete absence of staining or incomplete membrane staining which is faint or barely perceptible and within $\leq 10\%$ of the tumour cells (score 0), **B** Negative result with incomplete membrane staining

which is faint or barely perceptible and within $> 10\%$ of the tumour cells (score 1+). **C** Equivocal result showing weak to moderate complete membrane staining in $> 10\%$ of tumour cells (score 2+). **D** Positive result with circumferential membrane staining which is complete, intense, and in $> 10\%$ of tumour cells

Fig. 2 Graph showing the results of the HER2 status as evaluated by IHC and FISH in 118 ovarian clear cell carcinomas



There was no significant difference in any of the outcomes when applying all three categories (OS: $\chi^2 = 4.48$, $p = 0.106$, RFS: $\chi^2 = 5.93$, $p = 0.051$, LFS: $\chi^2 = 2.75$, $p = 0.252$, MFS: $\chi^2 = 4.99$, $p = 0.082$), although the HER2-zero cases showed a better probability of survival in all outcomes of interest when compared with a group including both the HER2-low and HER2-positive categories (there is a limitation in the number of complete cases, especially in the case of LFS and MFS, Fig. 3).

The 2-tier model including HER2-zero and HER2-low cases revealed better OS and RFS for HER2-zero patients ($Z = 2.01$, $p = 0.044$, and $Z = 2.12$, $p = 0.034$, respectively; Fig. 3). However, the multivariate analysis involving FIGO stage and age as possible covariates determined this relationship as not significant ($\chi^2 = 2.75$, $p = 0.098$), which would

indicate that HER2 status is not an independent prognostic factor, and that the survival parameters are better explained by FIGO stage.

Molecular findings

DNA analysis

The DNA NGS analysis of 100 eligible cases revealed an *ERBB2* pathogenic or likely pathogenic variant (class 4/5) in 3 cases. The *ERBB2* mutation NM_004448.2: c.2524G > A, p.(Val842Ile) was detected in one case (HER2-zero) and the mutation c.2033G > A, p.(Arg678Gln) was detected in two cases, one of which was classified as HER2-zero, while the other was scored as IHC 2+ (the FISH analysis in this case failed).

Table 2 Correlation between HER2 status and clinico-pathological characteristics

Characteristic	HER2-zero	HER2-low	HER2 positive	<i>p</i> -value (HER2-zero × HER2-low)	<i>p</i> -value (HER2-zero × HER2-low × HER2 positive)
Age (years)				0.063	0.163
No. of patients < median	36 (44%)	18 (64%)	2 (40%)		
No. of patients ≥ median	46 (56%)	10 (36%)	3 (60%)		
FIGO				0.238	0.266
(Low) I+II	66 (84%)	19 (73%)	3 (60%)		
(High) III+IV	13 (16%)	7 (27%)	2 (40%)		
T stage				0.514	0.281
(Low) T1 + T2	68 (86%)	21 (81%)	3 (60%)		
(High) T3	11 (14%)	5 (19%)	2 (40%)		
N stage				0.173	0.205
N0	37 (92.5%)	11 (79%)	2 (67%)		
N1	3 (7.5%)	3 (21%)	1 (33%)		
M stage				NULL	NULL
M0	3 (75%)	2 (100%)	1 (100%)		
M1	1 (25%)	0 (0%)	0 (0%)		
Local recurrence				0.434	0.065
Yes	12 (17%)	6 (24%)	3 (60%)		
No	59 (83%)	19 (76%)	2 (40%)		
Distant recurrence				1.000	0.125
Yes	8 (11%)	2 (8%)	2 (40%)		
No	63 (89%)	23 (92%)	3 (60%)		

p-values are based on chi-squared/Fisher exact test

NULL analysed is not possible

RNA analysis and correlation between immunohistochemistry and gene expression

The HER2 gene expression analysis showed a significant correlation with HER2 IHC status when examining the entire cohort (HER2-positive vs. HER2-negative)—the expression of normalized ERBB2 mRNA was 2.97-fold higher in the HER2-positive group than the HER2-low/HER2-zero ($p < 0.001$) (Fig. 4). In the cohort of only HER2-negative cases, the expression of ERBB2 mRNA was 1.06-fold higher in the HER2-low group than the HER2-zero group, but this result did not reach statistical significance ($p = 0.590$).

Discussion

HER2 is one of the receptor kinases which plays an important role in promoting and regulating cell proliferation and differentiation. The overexpression of HER2 has been linked to chemoresistance and poor outcomes, both of which represent the main characteristics of OCCC [22]. The reported overexpression/amplification of HER2 in OCCC shows striking variability. In our study, HER2 amplification/

overexpression was found in 6/118 OCCC (5%), with an additional 28/118 (24%) of cases being classified as HER2-low. This result is comparable to only one other study which reported HER2 overexpression in 6.7% of OCCCs; however, their cohort contained only 15 cases of OCCC and the authors used a different method of HER2 status evaluation (only cases which scored 3+ in IHC were considered positive) [24]. As previously mentioned, other works have reported HER2 overexpression in a very wide range including values of 12.6%; 12.5%; 14%; 20%; 42.9% and 45.6% [13, 29, 31, 33, 49, 50]. The comparison of these results is problematic, because the methodology of HER2 status evaluation lacks standardization (some authors considered HER2 score 2+ and 3+ as overexpression, some required only 3+ score to be considered as overexpression), and the confirmation of amplification by FISH was performed only in some studies [29, 49, 52]. A recent work by Koopman et al. compared three different antibodies (SP3, 4B5, and HercepTest) when evaluating ovarian clear cell carcinoma and pointed out that there is a considerable difference in HER2 overexpression by different antibodies, as well as a marked discordance with ISH [29]. Another important aspect regarding the scoring of HER2 expression by IHC is

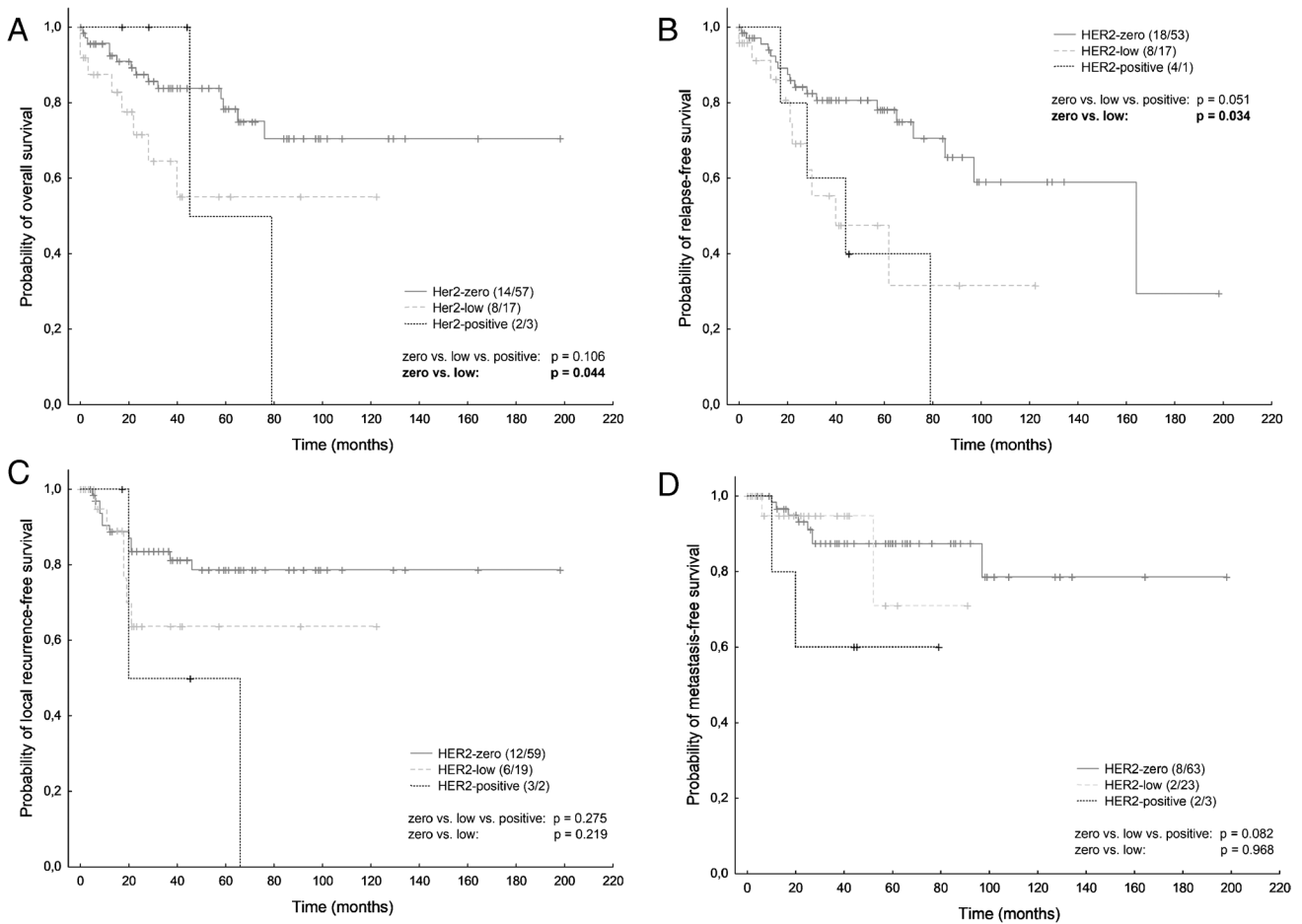


Fig. 3 Correlation of HER2 protein expression on a protein level with prognosis. Representative Kaplan–Meier curves for respective outcomes (A–D) of 101 OCCC patients with known follow-up. Numbers

in parentheses indicate complete/censored cases. Significant p -values are marked in bold

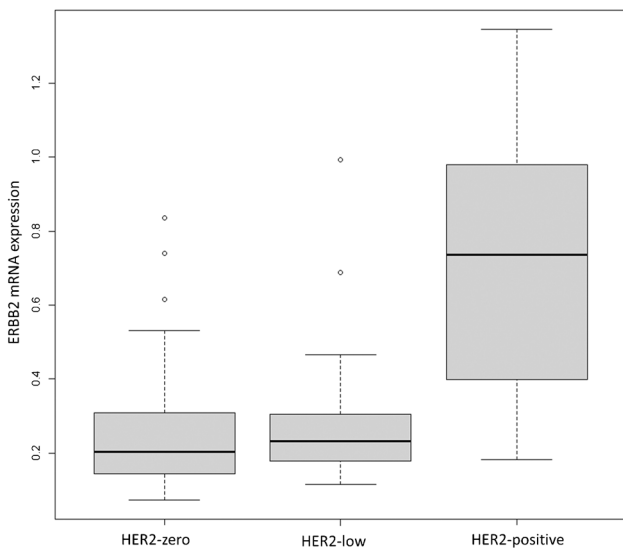


Fig. 4 Comparison of the expression of *ERBB2* (*HER2*) on an mRNA level in the three immunohistochemically defined subgroups. The values are referred to reference gene expression ATP5F1B

tumour heterogeneity, which seems to be more prominent in tumours of other sites apart from the breast and may result in false negative results and contribute to the observed discordance [48]. Given that HER2 may be a useful predictor for patients with OCCC, the question of a suitable method to evaluate HER2 status effectively is an important one.

In our cohort, none of the examined clinico-pathological parameters showed any significant correlation with HER2 IHC status. Considering the prognostic significance of HER2 expression in OCCC, the literary data is sparse. One review and meta-analysis has shown that increased levels of HER2 could predict worse survival, but the included tumours represent a heterogeneous group of ovarian carcinomas without further specification of their histotype [55]. Similarly, the results of the Danish “MALOVA” study (which included 9 OCCCs among the 181 investigated cases) also showed that increased HER2-expression correlated with reduced survival [16]. In contrast, others found that HER2 expression does not correlate with survival and lacks prognostic meaning [17, 50]. Our results showed that in a 3-tier model

using the categories of HER2-zero, HER2-low, and HER2-positive there were no statistically significant differences in any of the outcomes. However, these results are limited by the low number of cases in the HER2-positive group. When comparing only the groups HER2-zero and HER2-low, there was significantly better survival in the HER2-zero patients in case of both overall survival and relapse-free survival. Although this relationship lost its significance when the multivariate analysis was applied (suggesting that HER2 status is not an independent prognostic factor), these results could be of potential therapeutic value, given the recent discovery that patients with HER2-low breast cancer could also benefit from anti-HER2 therapy. However, the number of cases with complete available follow up represents a limitation of our study in this respect.

The frequency of HER2 overexpression/amplification indicates that anti-HER2 targeted therapy could be of value in this setting, especially in the cases of metastatic and/or recurrent tumours. Some authors reported promising results for the effect of trastuzumab on three OCCC cell lines, where trastuzumab significantly and dose-dependently reduced the growth of tumour cells in vitro and prolonged the survival of mice with a xenografted tumour [13]. However, others found that trastuzumab did not inhibit proliferation in any of the four OCCC lines tested [22]. Despite the potentially encouraging results, when trastuzumab was used in a single-agent study of 41 patients with recurrent or refractory ovarian/primary peritoneal carcinoma with an overexpression of HER2 (including 7 OCCCs) the therapeutic response was disappointing [6, 13]. The authors concluded that the value of single-agent trastuzumab therapy is not only limited by the low frequency of HER2 overexpression (in their cohort 11.4% of 837 screened tumour samples), but also by the rather poor overall response rate which reached 7.3% and included one complete and two partial responses, with no further specification provided for the subset of OCCC patients [6]. However, the inclusion criteria were set to include tumours which were HER2 2+ or HER2 3+ by IHC, without further assessment of the amplification status. The discordance between the IHC evaluation and the therapeutic responsiveness could have been caused by the different antibodies used for evaluation [29]. Based on the limited data, it seems that similarly to breast and gastric carcinoma, the efficacy of trastuzumab in OCCC patients could be improved by combining it with chemotherapy [14].

The correlation between the immunohistochemistry and gene expression analysis showed that while there was a strong correlation between HER2 gene expression and HER2 IHC status when the entire cohort was included (HER2-positive vs. HER2-negative); when looking at the HER2-negative cases only, the results were not statistically significant. In our cohort, the gene expression analysis would not be a suitable tool to confirm the subdivision of the

HER2-negative group into HER2-low and HER2-zero. However, it has been suggested that evaluating HER2 expression using more precise methods such as mRNA expression may provide a higher clinical meaning, given that HER2 evaluation by immunohistochemistry can underestimate the HER2 expression in ovarian cancer [30]. Comparable data is currently missing for ovarian carcinomas, although for breast cancer Almstedt et al. reported that the IHC HER2-subdivision in their cohort was significantly confirmed by gene expression analysis (both in the entire cohort and in the HER2-negative subgroup) [1].

Our results also revealed a class 4/5 *HER2* mutation in 3 cases, namely p.(Arg678Gln) in 2 cases and p.(Val842Ile) in one case. Both detected *HER2* variants are known recurrent activating oncogenic mutations found in diverse types of cancers [20, 25, 39]. We did not observe any relationship between mRNA/protein expression and the presence of HER2 mutation. The frequency of HER2-mutated OCCC in our study is comparable with the results of the other authors, which varies between 0 and 4% [5, 23, 34, 43]. The frequency of *HER2* mutations in our cohort (3/100, 3%) suggests that analysis of *HER2* oncogenic mutations may be of clinical interest, as these patients could also benefit from anti-HER2 therapy [18, 32].

We are aware of other limitations of our study. The main limitation is the use of TMA, as the results of the immunohistochemical evaluation could be influenced by tumour heterogeneity. While that is true, the use of TMA has become a widely used routine method in research to enable the evaluation of a large number of samples while preserving the sometimes limited tissue and providing high experimental uniformity. To minimize the potential impact of heterogeneity, two cores were taken from each of the tumour samples and evaluated together, thus increasing the amount of analysed tissue.

Conclusion

We have characterized the HER2 status using immunohistochemistry (IHC) and fluorescent in situ hybridization (FISH) on a large, well-defined cohort of 118 OCCCs fulfilling strict inclusion criteria. The frequency of overexpression/amplification (HER2-positive cases) confirms that anti-HER2 targeted therapy could be of value, especially in cases of metastatic and/or recurrent tumours. This is also the first work to date which has evaluated the HER2 status of OCCC in the context of the novel paradigm of HER2-zero and HER2-low status. Although the relationship between HER2 expression (HER2-zero vs. HER2-low) and survival lost its significance in the multivariate analysis (suggesting that HER2 status is not an independent prognostic factor), these results could be of potential therapeutic value, given the recent discovery that

patients with HER2-low breast cancer could also benefit from anti-HER2 therapy. Our results also emphasize the need for standardized HER2 testing in ovarian clear cell carcinoma to determine the most suitable predictor of clinical response.

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Author contribution All authors contributed to the study conception and design. Michaela Kendall Bártů and Pavel Dunder conceived the study and carried out experiments, Kristýna Němejcová has participated in the assessment of the immunohistochemistry results and the FISH analyses. Ivana Stružinská, Jan Hojný, Nikola Hájková, and Pavel Dunder conceived experiments and analysed data. Romana Michálková performed the statistical analyses. Eva Krkavcová carried out the molecular genetics part of the experiments. Jan Laco, Radoslav Matěj, Jana Drozenová, Gábor Méhes, Pavel Fabian, Jitka Hausnerová, Marián Švajdler, Petr Škapa, David Cibula, and Tomáš Zima participated in the selection and evaluation of cases, provided the clinical data and participated in the data analysis. All authors were involved in writing the paper and had final approval of the submitted and published versions.

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Data availability The bioinformatics pipeline and module settings used during the current study are available from the corresponding author on reasonable request.

Declarations

Ethics approval and consent to participate The study has been approved by the Ethics Committee of the General University Hospital in Prague in compliance with the Helsinki Declaration. The Ethics Committee did not require the procurement of informed consent, given that according to the Czech Law (Act. no. 373/11, and its amendment Act no. 202/17) it is not necessary to provide informed consent in fully anonymized studies.

Conflict of interest The authors declare no competing interests.

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