#### REVIEW



# Breast carcinomas of low malignant potential

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#### Abstract

Some breast carcinomas have a very low likelihood of metastasis to regional lymph nodes and distant sites and may be considered carcinomas of low malignant potential. In this article, we review the clinical, pathologic, immunophenotypic, and molecular features of selected breast carcinomas of low malignant potential including low-grade adenosquamous carcinoma, fibromatosis-like metaplastic carcinoma, encapsulated papillary carcinoma, solid papillary carcinoma, and tall cell carcinoma with reversed polarity.

**Keywords** Breast cancer  $\cdot$  Low malignant potential  $\cdot$  Low-grade adenosquamous carcinoma  $\cdot$  Fibromatosis-like metaplastic carcinoma  $\cdot$  Encapsulated papillary carcinoma  $\cdot$  Solid papillary carcinoma  $\cdot$  Tall cell carcinoma with reversed polarity

Most lesions in the breast are fairly common, readily categorized by pathologists as benign or malignant, and associated with relatively predictable clinical outcomes. However, there are some breast lesions currently categorized as carcinomas that have been repeatedly shown to have a very low risk of metastasizing to regional lymph nodes and distant sites. Included among this group are low-grade adenosquamous carcinoma, fibromatosis-like metaplastic carcinoma, encapsulated papillary carcinoma, solid papillary carcinoma, and tall cell carcinoma with reversed polarity. In this article, we will review the clinical and pathologic features and, where relevant, the molecular features of these breast carcinomas of low malignant potential. Not included in this review are more common breast carcinomas with a low risk of metastatic spread (e.g., tubular carcinomas) or salivary glandtype carcinomas arising in the breast, many of which also show indolent behavior.

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# Low-grade adenosquamous carcinoma

Low-grade adenosquamous carcinoma (LGASC) is a rare subtype of metaplastic breast carcinoma with 155 cases published to date [1–4]. These tumors most often occur in women older than 50 years of age (Supplementary Table 1). LGASC may arise de novo, but frequently occur in association with other breast lesions including sclerosing papillomas, complex sclerosing lesions, and adenomyoepitheliomas [3, 5].

Macroscopically, LGASCs are firm, poorly defined lesions with a white or pale yellow cut surface. The average size of reported cases is 20 mm with a wide range (0.5–86 mm) (Supplementary Table 1).

Microscopically, LGASCs show a variable admixture of well-formed tubular glands and solid clusters and cords of cells with squamous differentiation, arranged in a haphazard pattern and typically surrounded by a pale, cuff-like, mildly edematous desmoplastic stroma containing bland spindle cells (Fig. 1). The remainder of the stroma is often cellular but may sometimes be collagenous or even hyalinized. The spindle cell component often focally expresses cytokeratins (Fig. 2) and myoepithelial markers. Furthermore, a variable layer of myoepithelial cells is seen around the tubules, a feature usually associated with benign lesions. Even within a given case, the myoepithelial layer around the tubules can vary from complete to partial to absent [3, 6] (Fig. 3). Both the glandular and the solid components, as well as the spindle cell component, show only mild cytologic atypia,

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**Fig. 2** Cytokeratin AE1/AE3 immunostain demonstrating focal expression of cytokeratins by the spindle cells surrounding a tubule in low-grade adenosquamous carcinoma

few mitoses, and a low Ki67 proliferation rate. In addition, small aggregates of stromal lymphocytes are typically present. Rarely focal chondroid and/or osseous metaplasia has been reported [2, 7].

The relationship between benign LGASC and benign sclerosing lesions of the breast is of particular interest. The benign lesions with which LGASC are associated are characterized by the presence of the common element of adenosquamous proliferation (ASP). ASP shares a nearly identical histological and immunohistochemical profile with LGASC [6] ASP in sclerosing lesions has been characterized as a clonal proliferation and has been postulated to be a nonobligate precursor of LGASC [6, 8]. LGASCs are typically ER, PR, and HER2 negative (triple negative). The absence of staining for hormone receptors [[1, 3, 8, 9] in combination with expression of CK5/6, CK7 and CK14, E-cadherin, p63, and SMA [21] is considered a typical immunophenotype for LGASC.

Gene expression profiling studies showed a basal-like phenotype for LGASC, confirming the immunohistochemical impression [9]. The same genetic aberrations have been found in the lesional epithelial cells as well as in part of the spindle cells confirming the metaplastic nature of LGASC with typical epithelial-mesenchymal transition.

LGASCs show a high rate of PIK3CA mutations and absence of TP53 mutations [9, 10] [11, 21]. Baitallion et al.

Fig. 3 At least some of the tubules of the glandular component of this low-grade adenosquamous carcinoma are more or less continuously surrounded by myoepithelial cells in this CK5/6 immunostain. However, other histologically similar tubules show no surrounding myoepithelial cell layer



state that these findings, combined with the triple-negative phenotype and androgen receptor negativity, confirm LGASC as a distinct genetic entity among metaplastic carcinomas [10].

LGASC must be distinguished from a variety of benign and malignant breast lesions.

Due to the infiltrative pattern composed of a variable mixture of well-formed glands and solid cell strands with squamous differentiation and with typically minimal cytologic atypia, radial scar/complex sclerosing lesion may be in the differential diagnosis in some circumstances (especially in core needle biopsies). The more cell-rich spindle cell desmoplastic background, but especially the triple-negative phenotype, supports the diagnosis of LGASC [10]. Adenomyoepithelioma (AME) may also enter the differential diagnosis. Whereas classic AME is composed of glandular structures surrounded by a double cell layer, AME of spindle cell subtype is characterized by a prominent growth of spindle myoepithelial cells surrounding and compressing glandular structures lined by epithelial cells, which can sometimes be difficult to visualize. There are no strict criterion-based boundaries here. Accordingly, cases in which an association between AME and LGASC is observed seem to be more than coincidental [3, 5]. A major differential diagnostic consideration is a syringomatous tumor of the nipple (ST). LGASC and ST have a similar histological appearance and immunophenotype, differing only in their location [11]. However, it was proposed early on that ST arises from the pluripotent adnexal epidermal keratinocytes with dual follicular and sweat gland differentiation, whereas LGASC arises primarily from the myoepithelium [12]. Accordingly, LGASCs are also found within the mammary parenchyma without reference to the overlying skin or nipple epidermis,

while STs are restricted to the dermis of the nipple [4, 13]. Both lesions appear to have similar variable myoepithelial staining patterns, although the luminal and peripheral myoepithelial cytokeratin staining patterns mentioned above are usually absent in ST [6]. Both lesions are locally aggressive with a propensity for local recurrence. LGASC should be distinguished from other triple-negative/basal-like carcinomas that are associated with aggressive behavior. In highgrade adenosquamous carcinoma, there is always a mixture of an atypical squamous component and an NST type adenocarcinoma component whose architecture and cytology are not compatible with the LGASC-type glandular components described above [14]. Any sarcomatous spindle cell or heterologous (chondroid, osseous, rhabdomyoid) component in a metaplastic carcinoma excludes LGASC by definition and a high-grade spindle cell or high-grade metaplastic carcinoma with heterologous mesenchymal differentiation is present [14]

The prognosis of LGASC is excellent. Among the 155 published cases, follow-up data from a total of 102 patients (with 103 lesions) are available to confirm this: LGASCs show a low rate of local recurrence (10/103, 10%) and an extremely low rate of regional lymph node metastases (1/103 cases, 1%) and distant metastases (1/103 cases, 1%). Only one patient died from metastatic tumor [2, 3, 7, 15] (Supplemental Table 1).

## Fibromatosis-like metaplastic carcinoma

Fibromatosis-like metaplastic carcinoma (FLMC) is a very rare subtype of metaplastic breast carcinoma with a total of 70 published cases to date [16–28] (Supplementary Table 2).

FLMC usually affects women over the age of 50; only one case under the age of 40 has been reported [26]. The published cases were almost exclusively detected as solitary palpable masses; however, it must be considered that FLMCs are today increasingly detected by imaging studies [16–28].

On macroscopic examination, FLMCs are compact whitish tumors and typically 2–3 cm in size (reported range, 1.9–7.0 cm) (Supplementary Table 2). They are sometimes well circumscribed but may have irregular borders. They are very rarely cystic and never encapsulated [16, 17] (Fig. 4).

On histologic examination, FLMC is characterized by a proliferation of spindle-shaped, fibroblast-like, and myofibroblast-like cells, which account for more than 95% of the tumor [14, 16]. These cells are arranged in interlacing fascicles, often infiltrating the adjacent breast tissue, and histologically resembling desmoid-type fibromatosis (Fig. 5). Focally, bands of hyalinized collagen may dominate the interspersed spindle cells. Normal pre-existing ducts and lobules may be entrapped within the tumor. Patchy round cell inflammatory infiltrates may be seen inside the tumor, but are more often present at the periphery [16, 17] (Fig. 6).

The neoplastic cells are cytologically bland with no or minimal nuclear atypia and pale eosinophilic cytoplasm. Within the same tumor, the neoplastic nuclei may vary from thin, slender, and spindled with tapered ends to more plump, round to oval nuclei with discrete nucleoli within finely distributed chromatin. There may be a gradual transition from the slender spindle cells to the plump cell component (Fig. 7). The more plump spindle cells may assume an epithelioid appearance when cut in cross-section and tend to be arranged in a perivascular pattern [16, 17, 29]. High nuclear grade is not seen [14, 16, 17]. Mitotic figures are either completely absent or rare [16, 17]. Necrosis is not identified.

Neoplastic squamous or glandular epithelial elements may be admixed among the spindle cells. By definition, however, these should represent less than 5% of the total tumor area and FLMC should not contain any otherwise differentiated component [14].

Expression of epithelial markers by tumor spindle cells is the characteristic immunophenotypic feature of FLMC. However, FLMC may occasionally show only focal cytokeratin expression (Fig. 8). Therefore, the epithelial origin should preferably be verified by immunohistochemistry with a broad panel of cytokeratin antibodies that includes antibodies to both low and high molecular weight cytokeratins such as AE1/AE3, pan-keratin MNF116, CK5/6, CK14, and  $34\beta$ E12 [16–18, 22, 29, 30]. Additionally, nuclear expression of p63 in the spindle cell component has proven to be a sensitive and specific diagnostic feature [30]; thus, p63 should also be included in the panel of immunostains. Myoepithelial markers, such as CD10, calponin, and  $\alpha$ -SMA, may be also expressed [17, 30]. These tumors are typically ER, PR, and HER2 negative (triple negative) [30].

FLMCs show a claudin-low phenotype, with low expression of claudin 1–4 and E-cadherin, evidence of epithelialmesenchymal transition and tumor-infiltrating immune cells. Activating EGFR mutations have not been found [31].

The diagnosis of FLMC requires, on the one hand, the exclusion of benign and other low-grade spindle cell lesions of the breast and, on the other hand, the exclusion of higher-grade spindle cell metaplastic carcinoma. Benign and low-grade neoplastic spindle cell lesions of the breast may cause differential diagnostic problems, particularly in core needle biopsies. FLMC may occasionally show



**Fig. 4** Scanning magnification of a large format section of a 32-mm fibromatosis-like metaplastic carcinoma with irregular, radiating infiltration of surrounding tissue **Fig. 5** Low-power view of a fibromatosis-like metaplastic carcinoma showing interlacing fascicles of cytologically bland spindle cells with absent to minimal nuclear atypia and pale eosinophilic cytoplasm



a storiform pattern, fibromyxoid or pseudoangiomatoid areas, or collagen-rich foci with hyalinized collagen bands and a scattered inflammatory infiltrate of lymphocytes and plasma cells. The resulting spectrum of differential diagnoses, therefore, includes exaggerated scar tissue, benign fibrous histiocytoma, pseudoangiomatous stromal hyperplasia, nodular fasciitis, myofibroblastoma, inflammatory myofibroblastic tumor (IMT), solitary fibrous tumor (SFT), phyllodes tumor, and dermatofibrosarcoma protuberans. In addition to morphology, clinical history, and imaging, immunohistochemistry can help in making the diagnosis: the expression of epithelial markers by tumor spindle cells of FLMC and negative results for ALK, ER, CD34, and STAT6 helps to rule out these other entities. Desmoid fibromatosis also typically shows nuclear  $\beta$ -catenin expression, which is detectable due to the mutation in the CTNNB1 gene and is present in over 80% of cases [19, 32]. The high-grade or malignant spindle cell lesions that enter into the differential diagnoses of FLMCs include in particular other types of metaplastic carcinoma as well as rarer spindle cell malignancies such as leiomyosarcomas, fibrosarcomas, and spindle cell melanomas.

Because of the favorable prognosis of FLMCs, these tumors should be distinguished from the more common high-grade spindle cell metaplastic carcinomas. In these cases, conspicuous mitoses and a predominance of plump cells with unequivocal nuclear atypia strongly argue against an FLMC (Fig. 6). A proportion of the spindle cell component of less than 95% or a metaplastic carcinoma with a heterologous component also speaks against an FLMC (see above). An NST component in the tumor also rules out a FLMC by definition [14, 30]. Spindle cell sarcomas such as leiomyosarcomas and high-grade pleomorphic sarcomas, which may express not only SMA but also cytokeratin,

**Fig. 6** Cloudy lymphocytic infiltrate at the periphery of the lesion in a fibromatosis-like metaplastic carcinoma



**Fig. 7** Gradual transition from bland spindle cells to more plump cells with round to oval nuclei with discrete nucleoli within finely distributed chromatin



should be clearly distinguished from FLMC. Leiomyosarcomas usually show strong and diffuse staining for musclespecific actin, desmin, and caldesmon, compared to the weak and focal staining observed in FLMC. High-grade pleomorphic sarcoma can be discriminated from FLMC on the basis of a more pronounced nuclear atypia and an elevated number of mitoses [16, 17]. The rare fibrosarcoma is more cellular and has a characteristic "herringbone" growth pattern of impressively atypical spindle cells. Spindle cell melanoma may be negative for HMB-45 and melan A; thus, the strong, diffuse expression of S100 protein distinguishes it from FLMC, in which the expression of S100 protein is a more focal feature [33, 34].

Among the 70 cases of FLMC reported to date in the English-language literature, follow-up data are available for 41 patients. While these tumors have a high rate of local recurrence (14/41 cases, 34%), the low frequency of regional

lymph node metastases (3/41 cases, 2%) and distant metastases (5/41 cases, 12%) indicates clinically indolent behavior [16–28] (Supplementary Table 2).

An association between tumor size and the risk of distant metastasis suggested by some authors is not confirmed by a literature review [16–28] (Supplementary Table 2). Although FLMCs often are of considerable size when detected, they are usually detected at an early stage, unlike other subtypes of metaplastic carcinoma [19, 33]. The literature review suggests that the risk of local recurrence is at least partly related to inadequate local resection [16, 17, 21]. Histologic risk factors associated with local recurrence or distant metastasis have not been identified [14, 16–30, 32].

There are no evidence-based treatment guidelines for FLMC. In terms of local therapy, wide excision appears to be adequate to prevent local recurrence [16, 17, 23]. From the available data, there is no evidence for a beneficial effect of

**Fig. 8** Fibromatosis-like metaplastic carcinoma. AE1/AE3 stain showing that cytokeratin staining may be only focal and weak



adjuvant radiotherapy or chemotherapy in reducing the risk of local recurrence or distant metastasis [16–28]] (Table 1). However, some authors advocate adjuvant radiotherapy for larger lesions [2]. Due to the very low risk of regional lymph node metastases, several authors have argued against axillary lymph node evaluation[16, 30].

#### **Encapsulated papillary carcinoma**

Encapsulated papillary carcinoma (EPC), previously known as intracystic papillary carcinoma and encysted papillary carcinoma, is an expansile, well-circumscribed neoplasm characterized by delicate fibrovascular papillae covered by neoplastic epithelium of low to intermediate nuclear grade and typically lacking myoepithelial cells around the periphery of the lesion and within the fibrovascular cores [35–40]. It is an uncommon tumor comprising 1% of breast cancer cases and is primarily seen in women, mainly in the seventh and eighth decades [41]. Rare cases have been described in men [42].

The majority of EPCs are centrally located. Historically, these lesions came to attention as a palpable mass with or without nipple discharge but EPC now commonly present as mammographically detected masses. Macroscopically, the tumors are circumscribed, soft, friable masses, often within an apparent cystic space, sometimes with associated hemorrhage, especially after fine-needle aspiration or biopsy [39, 43].

Histologically, these tumors are well-circumscribed nodules that may appear to be within a cystic space and surrounded by a fibrous capsule. Some tumors consist of an aggregate of nodules with variable intervening fibrous stroma, especially following core needle biopsy. EPCs are characterized by branched, mostly slender, but occasionally broad fibrovascular stalks covered by one to several layers of monomorphic epithelial cells with low- to intermediategrade nuclear atypia (Fig. 9). In rare cases, the epithelium has apocrine features [44].

In most cases, no myoepithelial cells are present within the fibrovascular stalks or around the periphery of the lesion. The absence of myoepithelial cells both within fibrovascular stalks and at the periphery is a feature that distinguishes EPC from benign papillary lesions and is a useful feature in core needle biopsies. Areas of cribriform or solid growth may be present. The surrounding fibrous capsule may contain entrapped epithelial structures. This needs to be distinguished from frank invasion transgressing the capsule, which usually takes the form of invasive carcinoma of no special type [45]. Therefore, a diagnosis of invasion in a core needle biopsy with a low- to intermediate-grade papillary carcinoma should only be made with clear evidence of infiltrating glands. EPC usually exhibits strong positivity for estrogen receptors.

EPC is characterized by low genomic complexity with a common loss of 16q and gains of 1q and 16p and a high frequency of *P1K3CA* mutations, similar to the genomic profile of low grade, hormone receptor–positive invasive carcinomas [46, 47]. EPCs usually belong to the luminal A group by gene expression profiling.

The differential diagnosis of EPC encompasses a broad range of lesions with papillary features and diverse biological behavior [39]. Differentiation from benign papillary lesions is easy in most cases by virtue of the monotonous appearance of lesional epithelial cells and lack of myoepithelial cells; the latter can be confirmed, if necessary, by staining for myoepithelial markers. Papillary ductal carcinoma in situ (DCIS), which may be present in breast issues adjacent to EPC, shows the presence of myoepithelial cells at the periphery of the involved ducts. Rare cases of EPC show high-grade cytology and these may be ER negative [48]. While it has been recommended to stage high-grade EPC as conventional invasive carcinoma, there are no strong data to support this since only one of ten patients reported developed a recurrence and died of disease [48]. An invasive component accompanying EPC should only be diagnosed, when clearly neoplastic aggregates are found beyond the fibrous capsule. Despite the characteristic absence of myoepithelial cells, EPC has a prognosis similar to that of ductal carcinoma in situ. In order to prevent overtreatment, in the absence of frankly invasive carcinoma that extends beyond the fibrous capsule, EPC should be assigned a nuclear grade and staged as pTis (DCIS) [40]. When there is an accompanying frankly invasive carcinoma, it is the invasive component that should be used for grading (using the Nottingham system) and determining the T stage.

EPC is adequately treated with local therapy. The risk of local recurrence appears to be related to the presence of ductal carcinoma in situ in the adjacent breast tissue [40] Rare cases of lymph node metastases have been reported [49] but the possibility that this was related to undetected invasive carcinoma in these cases cannot be excluded.

## Solid papillary carcinoma

Solid papillary carcinomas (SPCs) are defined by the WHO as tumors characterized by a solid growth pattern with delicate fibrovascular cores that frequently show neuroendocrine differentiation and are biologically indolent. These lesions are most often seen in post-menopausal women, typically during the 7<sup>th</sup> decade or later and may be in situ or invasive [50]. These lesions may present as a palpable or mammographic mass, mammographic microcalcifications, or bloody nipple discharge.



**Fig. 9** Encapsulated papillary carcinoma. **A** Scanning magnification shows an encapsulated papillary lesion. **B** Branched papillae of variable thickness are covered by a monotonous epithelial proliferation. **C** At high power, one to several layers of columnar epithelial cells with increased nuclear to cytoplasmic ratio, loss of nuclear polarity, and mild atypia are evident. **D** Immunostaining for smooth muscle myo-

Histologically, SPC consists of one or more solid, wellcircumscribed expansile nodules of monotonous neoplastic epithelial cells of low to intermediate nuclear grade with a round to spindled nuclei reminiscent of DCIS with solid growth pattern [51, 52]. However, the nodules of SPC contain slender fibrovascular cores which separate SPC from solid pattern ductal carcinoma in situ. Palisading of tumor cells around the fibrovascular cores is a characteristic feature. Signet ring cell morphology, mucin accumulation, a microcystic pattern, and calcifications can be observed. Occasionally, a streaming appearance similar to usual ductal hyperplasia can occur. Myoepithelial cells are usually absent or sparse both within, as well as at the

sin heavy chain demonstrates complete absence of myoepithelial cells both on the papillae, as well as in the inner lining of the cyst. **E** Early invasion in encapsulated papillary carcinoma. Isolated glandular and papillary structures start to infiltrate beyond the fibrous capsule. Entrapped epithelia in the fibrous capsule alone would not qualify for invasion

periphery of individual nodules. [53] As long as the expansile and rounded character of the nodules is preserved, SPC is regarded as in situ lesion irrespective of the presence or absence of surrounding myoepithelial cells (Fig. 10). The assessment of invasion in SPC can be difficult, especially in cases which show a jigsaw puzzle-like appearance with irregularly shaped solid tumor nodules (Fig. 11). Invasive SPC is frequently associated with extracellular mucin pools and must clearly be distinguished from frank mucinous carcinoma or invasive carcinoma NST [54, 55]. SPCs are usually ER and PR positive and HER2 negative.

Neuroendocrine differentiation with expression of neuroendocrine markers such as synaptophysin or chromogranin Fig. 10 Solid papillary carcinoma in situ. A At scanning magnification, the lesion consists of multiple circumscribed nodules. B At higher power, the tumor cells are uniform in appearance and grow in a solid pattern; delicate fibrovascular cores are evident. C An immunostain for p63 demonstrates variable numbers of myoepithelial cells around the periphery of the nodules. The nodule on the upper right has no surrounding myoepithelial cells



A has been described in at least half of the cases of SPC and is commonly present also in the invasive part [54-56].

SPCs, like EPC, share low genomic complexity with a common loss of 16q and gains of 1q and 16p and a high frequency of *PIK3CA* mutations with hormone receptorpositive carcinomas of low grade [46, 47]. SPCs more frequently than EPC are of luminal B molecular type. HER2 amplification and *TP53* mutations are distinctly infrequent.

The differential diagnosis of SPC includes florid, usual ductal hyperplasia (UDH) with solid growth pattern, solid pattern DCIS, and neuroendocrine tumors of the breast. The distinction from UDH should be easily made with immunostains for high molecular weight cytokeratins (variably positive in UDH and negative in SPC), ER (variably positive in UDH and strongly and diffusely positive in SPC), and neuroendocrine markers (negative in UDH and positive in many cases of SPC) [57, 58]. The presence of rare fibrovascular cores should prompt the inclusion of SPC in the differential diagnosis of solid pattern DCIS and staining for neuroendocrine and myoepithelial markers should be performed. SPC shares many features with the

hypercellular or type B mucinous carcinoma, supporting their close relationship. The distinction from tall cell carcinoma with reversed polarity is discussed below [59–65].

The prognosis of SPC is excellent, with a rare occurrence of lymph node (2-3%) or distant metastasis [38]. Among 265 published cases, Guo et al. reported 11 local recurrences in their meta-analysis (2 for in situ; 9 for invasive SPC). In SPC in situ, neither lymph node nor distant metastases were seen in 265 published cases (even in cases without peripheral myoepithelial confinement). Lymph node or distant metastases occurred exclusively in 5 and 7 cases, respectively, of the 135 invasive SPCs, while all 129 in situ SPCs were metastasis-free [66]. The WHO currently recommends that, in the absence of an obvious invasive component, SPC should be staged as pTis (DCIS) regardless of whether or not a myoepithelial cell layer is identified at the periphery of the nodules whereas SPC with invasion is staged according to the size of the invasive component only [50].

The key features of EPC and SPC are summarized in Supplementary Table 3.



Fig. 11 Solid papillary carcinoma with focal invasion. A Scanning magnification shows solid nests of tumor with mostly smooth, rounded contours. B This area with round nodules does not fulfill the criteria for invasion. C The high-power view shows a solid proliferation of epithelial cells with round to oval low-grade nuclei with palisading around delicate fibrovascular cores. D Synaptophysin is

strongly and homogenously expressed and highlights an area of invasion with irregular small tumor islands (arrowheads). **E** The immunostain for cytokeratin CK5/14 of the same area shows tumor nodules with complete, incomplete, and absent basal cell layers. However, only the same area as above between the arrowheads would be considered true invasion. **F** Variable staining for chromogranin A

## Tall cell carcinoma with reversed polarity

Tall cell carcinoma with reversed polarity (TCCRP) was recently recognized by the WHO as a rare subtype of invasive breast carcinoma [67]. This tumor was first reported by Eusebi et al. as "breast tumor resembling tall cell variant of papillary thyroid carcinoma" [68] and subsequently reported as "solid papillary carcinoma with reverse polarity" [59] and "solid papillary carcinoma resembling tall cell variant of papillary thyroid carcinoma" [69]. To date, approximately 80 cases of this tumor have been reported.

TCCRP primarily occurs in post-menopausal women in their 6<sup>th</sup> to 7<sup>th</sup> decades and present most often as relatively

small mammographic abnormalities (typically T1 lesions). Histologically, these tumors are composed of solid, circumscribed nodules consisting of tall columnar epithelial cells. The nodules often have relatively inconspicuous fibrovascular cores imparting a solid papillary appearance, and these cores may contain foamy histiocytes (Fig. 12). Infrequently, true papillae are formed. The nodules are haphazardly distributed in the breast stroma, are present around and between normal ducts and lobule, and may extend into adipose tissue. The stroma is typically collagenous with little or no desmoplasia. In some cases, cystic or follicular structures containing colloid-like secretions are present. The columnar cells comprising the nodules have eosinophilic **Fig. 12** Low-power view of a tall cell carcinoma with reversed polarity showing haphazard distribution of relatively rounded tumor cells nests, many of which contain fibrovascular cores imparting a solid papillary appearance



cytoplasm and usually intermediate-grade nuclear atypia. In some nodules, the cells are present in a double layer with a back-to-back appearance. Nuclear grooves and cytoplasmic pseudo-inclusions may be seen. However, the most distinctive feature of these cells is the presence of the nuclei at the apical rather than the basal pole, creating the impression of reversed polarity (Fig. 13)[59].

The tumor nodules lack a surrounding myoepithelial cell layer supporting the invasive nature of these lesions (Fig. 14). The nodules are surrounded by a rich vascular

**Fig. 13** In this high-power view of a tall cell carcinoma with reversed polarity the nuclei are clearly evident at the apical rather than the basal pole of the columnar epithelial cells. The nest in the center of the field contains foamy histiocytes within the fibrovascular core



**Fig. 14** p63 immunostain of a tall cell carcinoma with reversed polarity demonstrates absence of myoepithelial cells around the tumor cell nests (note the presence of myoepithelial cells around the normal ducts)



network on immunostains for endothelial markers such as CD31 and CD34. The epithelial cells comprising these lesions typically show strong staining for both low molecular weight (luminal) and high molecular weight (basal) cytokeratins and may show staining for GATA3, gross cystic disease fluid protein, and mammaglobin [59–63, 65, 68, 69]. These cells typically show staining for calretinin [62, 65] as well as a basal pattern of staining for mitochondrial antigen [69]. The cells comprising TCCRP are typically negative for estrogen receptor (ER), progesterone receptor (PR), and HER2 (triple negative), but some tumors show low levels of ER and or PR expression [59, 63]. MUC1 staining, which highlights the apical poles of columnar epithelial cells, is present at the end of the cells closest to the nuclei confirming the abnormal nuclear location [59]. The tumor cells have a low Ki67 proliferation rate (<5%) [69]. Despite the superficial resemblance to the tall cell variant of papillary thyroid carcinoma, these lesions are uniformly negative for thyroglobulin and TTF-1 [59, 65, 68, 69].

Chiang et al. were the first to describe the genomic alterations in TCCRP. Among 13 cases studied, 10 (77%) showed *IDH2* R172 hot spot mutations and 8 concurrently displayed mutations in the PI3 kinase pathway (in either *PIK3CA* or *PIK3R1*) [59]. In addition, in that series one *IDH2* wild-type tumor displayed a *TET2* Q548 truncating mutation coupled with a *PIK3CA* mutation. Of note, *IDH2* mutations had not been previously reported in any breast carcinomas. Functional studies demonstrated that *IDH2* and *PIK3CA* hot spot mutations are likely drivers resulting in the reverse polarity phenotype. Subsequent studies by other groups have confirmed the presence of *IDH2* R172 hot spot mutations in these tumors and to date this mutation has been found in 34 of 40 TCCRP studied (85%) [59–65] Pareja et al. have reported that demonstration of the *IDH2* R172 protein by immunohistochemistry is a sensitive and specific marker for TCCRP and provides a rapid and inexpensive alternative to sequencing [70]. Of note, no *BRAF* mutations or *RET/ PTC* rearrangements have been reported in TCCRP providing further evidence of a lack of a relationship to thyroid tumors [59, 61–65]

The differential diagnosis of TCCRP includes several entities. The circumscribed nature of the nodules often gives the impression of DCIS, but the lack of surrounding myoepithelial cells, the haphazard distribution of the nodules in the stroma, and the tall columnar cells with nuclei at the apical poles of the cells distinguish TCCRP from DCIS. The presence of fibrovascular cores in the nodules may raise concern for a SPC (in situ or invasive). However, solid papillary carcinomas do not have tall columnar epithelial cells with reversed polarity. The similarity of some features of TCCRP to some thyroid neoplasms may raise the question of a metastatic lesion of thyroid origin. However, the presence of reversed polarity, lack of expression of thyroid markers, and the characteristic immunophenotype and genotype distinguish TCCRP from tumors of thyroid origin.

TCCRP generally pursue an indolent clinical course. There are four reported cases with axillary lymph node metastases and a single reported case of distant metastases to bone [67]. However, it is unclear if the tumor that metastasized to bone is a *bona fide* example of TCCRP since this tumor at presentation was large (4.1 cm), had associated DCIS with comedo necrosis, demonstrated lymphovascular invasion and lymph node metastases, was ER positive, and had a high Ki67 proliferation rate, all unusual features for TCCRP [71].

In summary, TCCRP is a histologically low-grade invasive breast carcinoma with distinctive morphologic, immunophenotypic, and genotypic features. These tumors typically pursue an indolent clinical course despite a triplenegative phenotype and expression of high molecular weight (basal) cytokeratins and add to the list of low-grade triplenegative neoplasms that includes adenoid cystic carcinomas, secretory carcinomas, and acinic cell carcinomas.

# Conclusion

Some breast carcinomas, including low-grade adenosquamous carcinoma, fibromatosis-like metaplastic carcinoma, encapsulated papillary carcinoma, solid papillary carcinoma, and tall cell carcinoma with reversed polarity have a very low likelihood of aggressive behavior, particularly distant metastases, and are best categorized as carcinomas of low malignant potential. Recognition and appropriate classification are important so that patients with these tumors can avoid overtreatment.

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This manuscript complies with the ethical standards of the authors' institutions.

#### Declarations

Conflict of interest The authors declare no competing interests.

#### References

- Drudis T, Arroyo C, Van Hoeven K, Cordon-Cardo C, Rosen PP (1994) The pathology of low-grade adenosquamous carcinoma of the breast. An immunohistochemical study. Pathol Annu 29(Pt):181–97
- Rosen PP, Ernsberger D (1987) Low-grade adenosquamous carcinoma. A variant of metaplastic mammary carcinoma. Am J Surg Pathol 11(5):351–8
- Van Hoeven KH, Drudis T, Cranor ML, Erlandson RA, Rosen PP (1993) Low-grade adenosquamous carcinoma of the breast. A clinocopathologic study of 32 cases with ultrastructural analysis. Am J Surg Pathol 17(3):248–58
- Soo K, Tan PH (2013) Low-grade adenosquamous carcinoma of the breast. J Clin Pathol 66(6):506–511

- Foschini MP, Pizzicannella G, Peterse JL, Eusebi V (1995) Adenomyoepithelioma of the breast associated with low-grade adenosquamous and sarcomatoid carcinomas. Virchows Arch 427(3):243–250
- Kawaguchi K, Shin SJ (2012) Immunohistochemical staining characteristics of low-grade adenosquamous carcinoma of the breast. Am J Surg Pathol 36(7):1009–1020
- Chuthapisith S, Warnnissorn M, Amornpinyokiat N, Pradniwat K, Angsusinha T (2013) Metaplastic carcinoma of the breast with transformation from adenosquamous carcinoma to osteosarcomatoid and spindle cell morphology. Oncol Lett 6(3):728–732
- Wilsher MJ (2014) Adenosquamous proliferation of the breast and low grade adenosquamous carcinoma: a common precursor of an uncommon cancer? Pathology 46(5):402–410
- Geyer FC, Lambros MB, Natrajan R, Mehta R, Mackay A, Savage K et al (2010) Genomic and immunohistochemical analysis of adenosquamous carcinoma of the breast. Mod Pathol 23(7):951–960
- Bataillon G, Collet JF, Voillemot N, Menet E, Vincent-Salomon A, Klijanienko J (2014) Fine-needle aspiration of low-grade adenosquamous carcinomas of the breast: a report of three new cases. Acta Cytol 58(5):427–431
- Boecker W, Stenman G, Loening T, Andersson MK, Berg T, Lange A et al (2015) Squamous/epidermoid differentiation in normal breast and salivary gland tissues and their corresponding tumors originate from p63/K5/14-positive progenitor cells. Virchows Arch 466(1):21–36
- 12. Ferrara G, Nappi O, Wick MR (1999) Fine-needle aspiration cytology and immunohistology of low-grade adenosquamous carcinoma of the breast. Diagn Cytopathol 20(1):13–18
- Ho BC, Tan HW, Lee VK, Tan PH (2006) Preoperative and intraoperative diagnosis of low-grade adenosquamous carcinoma of the breast: potential diagnostic pitfalls. Histopathology 49(6):603–611
- Reis-Filho JS, Gobbi H, McCart Reed AE, Rakha E, Shin SJ, Sotiriou C, Vincent-Salomon A. Metaplastic carcinoma. In: WHO classification of tumors editorial board, editor. WHO Classification of Tumours: Breast Tumours. Lyon: IARC Press; 2019. p. 134–8.
- Scali EP, Ali RH, Hayes M, Tyldesley S, Hassell P (2013) Lowgrade adenosquamous carcinoma of the breast: imaging and histopathologic characteristics of this rare disease. Can Assoc Radiol J 64(4):339–344
- Gobbi H, Simpson JF, Borowsky A, Jensen RA, Page DL (1999) Metaplastic breast tumors with a dominant fibromatosis-like phenotype have a high risk of local recurrence. Cancer 85(10):2170–2182
- Sneige N, Yaziji H, Mandavilli SR, Perez ER, Ordonez NG, Gown AM et al (2001) Low-grade (fibromatosis-like) spindle cell carcinoma of the breast. Am J Surg Pathol 25(8):1009–1016
- Schafernak KT, Policarpio-Nicolas ML, Wiley EL, Laskin WB, Diaz LK (2006) A 59-year-old woman with a spindle cell lesion of the breast. Low-grade (fibromatosis-like) spindle cell carcinoma of the breast. Arch Pathol Lab Med 130(5):e81-3
- Rekhi B, Shet TM, Badwe RA, Chinoy RF (2007) Fibromatosislike carcinoma-an unusual phenotype of a metaplastic breast tumor associated with a micropapilloma. World J Surg Oncol 5:24
- Podetta M, D'Ambrosio G, Ferrari A, Sgarella A, Dal Bello B, Fossati GS et al (2009) Low-grade fibromatosis-like spindle cell metaplastic carcinoma: a basal-like tumor with a favorable clinical outcome. Report of two cases Tumori 95(2):264–267
- Lamovec J, Gasljevic G (2010) Keloid type of fibromatosis-like metaplastic carcinoma of the breast with transformation into biphasic tumour in recurrences and lymph node metastases. Histopathology 57(2):318–320

- Nonnis R, Paliogiannis P, Giangrande D, Marras V, Trignano M (2012) Low-grade fibromatosis-like spindle cell metaplastic carcinoma of the breast: a case report and literature review. Clin Breast Cancer 12(2):147–150
- 23. Rito M, Schmitt F, Pinto AE, Andre S (2014) Fibromatosis-like metaplastic carcinoma of the breast has a claudin-low immunohistochemical phenotype. Virchows Arch 465(2):185–191
- Takano EA, Hunter SM, Campbell IG, Fox SB (2015) Low-grade fibromatosis-like spindle cell carcinomas of the breast are molecularly exiguous. J Clin Pathol 68(5):362–367
- 25. Pinilla Pagnon I, Perez Mies B, Tulio Martinez M, Pena JL, Roldan Cabanillas AM, Romio de las Hera E, Blazquez Ortiz JM, Sanchez Monforte J, Delgado Moya MA, Rubio Marin D. Metaplastic breast carcinoma "fibromatosis like", associated with intraductal papilloma. Case report and literature review. Hum Pathol Case Reports. 2017;9:15–8.
- 26. Zhu H, Li K, Dong DD, Fu J, Liu DD, Wang L et al (2017) Spindle cell metaplastic carcinoma of breast: a clinicopathological and immunohistochemical analysis. Asia Pac J Clin Oncol 13(2):e72–e78
- Zhao Y, Gong X, Li N, Zhu B, Yu D, Jin X (2018) Fibromatosislike metaplastic carcinoma of breast: a challenge for clinicopathologic diagnosis. Int J Clin Exp Pathol 11(7):3691–3696
- Takatsuka D, Ogura H, Asano Y, Nakamura A, Koizumi K, Shiiya N et al (2021) A difficult-to-diagnose fibromatosis-like metaplastic carcinoma of the breast: a case report. Surg Case Rep 7(1):16
- Gobbi H, Simpson JF, Jensen RA, Olson SJ, Page DL (2003) Metaplastic spindle cell breast tumors arising within papillomas, complex sclerosing lesions, and nipple adenomas. Mod Pathol 16(9):893–901
- Dwyer JB, Clark BZ (2015) Low-grade fibromatosis-like spindle cell carcinoma of the breast. Arch Pathol Lab Med 139(4):552–557
- Victoor J, Bourgain C, Vander Borght S, Vanden Bempt I, De Rop C, Floris G (2020) Fibromatosis-like metaplastic carcinoma: a case report and review of the literature. Diagn Pathol 15(1):20
- 32. Rakha EA, Aleskandarany MA, Lee AH, Ellis IO (2016) An approach to the diagnosis of spindle cell lesions of the breast. Histopathology 68(1):33–44
- Al-Nafussi A (1999) Spindle cell tumours of the breast: practical approach to diagnosis. Histopathology 35(1):1–13
- Behranwala KA, Nasiri N, A'Hern R, Gui GP (2004) Clinical presentation and long-term outcome of pure myoepithelial carcinoma of the breast. Eur J Surg Oncol 30(4):357–361
- Lefkowitz M, Lefkowitz W, Wargotz ES (1994) Intraductal (intracystic) papillary carcinoma of the breast and its variants: a clinicopathological study of 77 cases. Hum Pathol 25(8):802–809
- Collins LC, Carlo VP, Hwang H, Barry TS, Gown AM, Schnitt SJ (2006) Intracystic papillary carcinomas of the breast: a reevaluation using a panel of myoepithelial cell markers. Am J Surg Pathol 30(8):1002–1007
- 37. Mulligan AM, O'Malley FP (2007) Papillary lesions of the breast: a review. Adv Anat Pathol 14(2):108–119
- Rakha EA, Gandhi N, Climent F, van Deurzen CH, Haider SA, Dunk L et al (2011) Encapsulated papillary carcinoma of the breast: an invasive tumor with excellent prognosis. Am J Surg Pathol 35(8):1093–1103
- Ni YB, Tse GM (2016) Pathological criteria and practical issues in papillary lesions of the breast - a review. Histopathology 68(1):22–32
- MacGrogan G, Collins L, Lerwill M, Rakha EA, Tan BY. Encapsulated papillary carcinoma. In: WHO classification of tumors editorial board, editor. WHO Classification of Tumours: Breast Tumours. 5th ed. Lyon: IARC Press; 2019. p. 60–2.
- 41. Tay TKY, Tan PH. Papillary neoplasms of the breast-reviewing the spectrum. Mod Pathol. 2021.

- 42. Zhong E, Cheng E, Goldfischer M, Hoda SA (2020) Papillary lesions of the male breast: a study of 117 cases and brief review of the literature demonstrate a broad clinicopathologic spectrum. Am J Surg Pathol 44(1):68–76
- Collins LC, Schnitt SJ (2008) Papillary lesions of the breast: selected diagnostic and management issues. Histopathology 52(1):20–29
- 44. Seal M, Wilson C, Naus GJ, Chia S, Bainbridge TC, Hayes MM (2009) Encapsulated apocrine papillary carcinoma of the breast–a tumour of uncertain malignant potential: report of five cases. Virchows Arch 455(6):477–483
- 45. Wynveen CA, Nehhozina T, Akram M, Hassan M, Norton L, Van Zee KJ, et al. Intracystic papillary carcinoma of the breast: an in situ or invasive tumor? Results of immunohistochemical analysis and clinical follow-up. Am J Surg Pathol. 2011.
- 46. Duprez R, Wilkerson PM, Lacroix-Triki M, Lambros MB, Mackay A, Hern RA et al (2012) Immunophenotypic and genomic characterization of papillary carcinomas of the breast. J Pathol 226(3):427–441
- 47. Piscuoglio S, Ng CK, Martelotto LG, Eberle CA, Cowell CF, Natrajan R et al (2014) Integrative genomic and transcriptomic characterization of papillary carcinomas of the breast. Mol Oncol 8(8):1588–1602
- Rakha EA, Varga Z, Elsheik S, Ellis IO (2015) High-grade encapsulated papillary carcinoma of the breast: an under-recognized entity. Histopathology 66(5):740–746
- 49. Mulligan AM, O'Malley FP (2007) Metastatic potential of encapsulated (intracystic) papillary carcinoma of the breast: a report of 2 cases with axillary lymph node micrometastases. Int J Surg Pathol 15(2):143–147
- MacGrogan G, Collins LC, Lerwill M, Rakha EA, Tan BY. Solid papillary carcinoma (in situ and invasive). In: WHO classification of tumors editorial board, editor. WHO Classification of Tumours: Breast Tumours. 5th ed. Lyon: IARC Press; 2019. p. 63–5.
- Maluf HM, Koerner FC. Solid papillary carcinoma of the breast. A form of intraductal carcinoma with endocrine differentiation frequently associated with mucinous carcinoma. Am J Surg Pathol. 1995;19(11):1237–44.
- Tan BY, Thike AA, Ellis IO, Tan PH (2016) Clinicopathologic Characteristics of Solid Papillary Carcinoma of the Breast. Am J Surg Pathol 40(10):1334–1342
- Nicolas MM, Wu Y, Middleton LP, Gilcrease MZ (2007) Loss of myoepithelium is variable in solid papillary carcinoma of the breast. Histopathology 51(5):657–665
- Nassar H, Qureshi H, Volkanadsay N, Visscher D (2006) Clinicopathologic analysis of solid papillary carcinoma of the breast and associated invasive carcinomas. Am J Surg Pathol 30(4):501–507
- 55. Otsuki Y, Yamada M, Shimizu S, Suwa K, Yoshida M, Tanioka F et al (2007) Solid-papillary carcinoma of the breast: clinicopathological study of 20 cases. Pathol Int 57(7):421–429
- 56. Moritani S, Ichihara S, Kushima R, Okabe H, Bamba M, Kobayashi TK et al (2007) Myoepithelial cells in solid variant of intraductal papillary carcinoma of the breast: a potential diagnostic pitfall and a proposal of an immunohistochemical panel in the differential diagnosis with intraductal papilloma with usual ductal hyperplasia. Virchows Arch 450(5):539–547
- 57. Tan PH, Schnitt SJ, van de Vijver MJ, Ellis IO, Lakhani SR (2015) Papillary and neuroendocrine breast lesions: the WHO stance. Histopathology 66(6):761–770
- Pareja F, D'Alfonso TM. Neuroendocrine neoplasms of the breast: A review focused on the updated World Health Organization (WHO) 5th Edition morphologic classification. Breast J. 2020;26(6):1160–7.
- 59. Chiang S, Weigelt B, Wen HC, Pareja F, Raghavendra A, Martelotto LG et al (2016) IDH2 mutations define a unique

subtype of breast cancer with altered nuclear polarity. Cancer Res 76(24):7118–7129

- 60. Bhargava R, Florea AV, Pelmus M, Jones MW, Bonaventura M, Wald A et al (2017) Breast tumor resembling tall cell variant of papillary thyroid carcinoma: a solid papillary neoplasm with characteristic immunohistochemical profile and few recurrent mutations. Am J Clin Pathol 147(4):399–410
- 61. Lozada JR, Basili T, Pareja F, Alemar B, Paula ADC, Gularte-Merida R et al (2018) Solid papillary breast carcinomas resembling the tall cell variant of papillary thyroid neoplasms (solid papillary carcinomas with reverse polarity) harbour recurrent mutations affecting IDH2 and PIK3CA: a validation cohort. Histopathology 73(2):339–344
- 62. Alsadoun N, MacGrogan G, Truntzer C, Lacroix-Triki M, Bedgedjian I, Koeb MH et al (2018) Solid papillary carcinoma with reverse polarity of the breast harbors specific morphologic, immunohistochemical and molecular profile in comparison with other benign or malignant papillary lesions of the breast: a comparative study of 9 additional cases. Mod Pathol 31(9):1367–1380
- 63. Zhong E, Scognamiglio T, D'Alfonso T, Song W, Tran H, Baek I et al (2019) Breast tumor resembling the tall cell variant of papillary thyroid carcinoma: molecular characterization by next-generation sequencing and histopathological comparison with tall cell papillary carcinoma of thyroid. Int J Surg Pathol 27(2):134–141
- 64. Haefliger S, Muenst S, Went P, Bihl M, Dellas S, Weber WP et al (2020) Tall cell carcinoma of the breast with reversed polarity (TCCRP) with mutations in the IDH2 and PIK3CA genes: a case report. Mol Biol Rep 47(6):4917–4921
- 65. Jassim M, Premalata CS, Okaly G, Srinivas C. Tall cell carcinoma with reverse polarity of breast: report of a case with unique morphologic and molecular features. Turk Patoloji Derg. 2020.

- 66. Guo S, Wang Y, Rohr J, Fan C, Li Q, Li X et al (2016) Solid papillary carcinoma of the breast: a special entity needs to be distinguished from conventional invasive carcinoma avoiding over-treatment. Breast 26:67–72
- Yang WT, Bu H, Foschini MP, Schnitt SJ. Tall cell carcinoma with reversed polarity. In: Board WCoTE, editor. WHO Classification of Tumours: Breast Tumours. 5th ed. Lyon: IARC Press; 2019. p. 153–4.
- Eusebi V, Damiani S, Ellis IO, Azzopardi JG, Rosai J (2003) Breast tumor resembling the tall cell variant of papillary thyroid carcinoma: report of 5 cases. Am J Surg Pathol 27(8):1114–1118
- 69. Foschini MP, Asioli S, Foreid S, Cserni G, Ellis IO, Eusebi V et al (2017) Solid papillary breast carcinomas resembling the tall cell variant of papillary thyroid neoplasms: a unique invasive tumor with indolent behavior. Am J Surg Pathol 41(7):887–895
- 70. Pareja F, da Silva EM, Frosina D, Geyer FC, Lozada JR, Basili T et al (2020) Immunohistochemical analysis of IDH2 R172 hotspot mutations in breast papillary neoplasms: applications in the diagnosis of tall cell carcinoma with reverse polarity. Mod Pathol 33(6):1056–1064
- Cameselle-Teijeiro J, Abdulkader I, Barreiro-Morandeira F, Ruiz-Ponte C, Reyes-Santias R, Chavez E et al (2006) Breast tumor resembling the tall cell variant of papillary thyroid carcinoma: a case report. Int J Surg Pathol 14(1):79–84

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