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## Galectin-3 and HBME-1 expression in oncocytic cell tumors of the thyroid

Received: 9 April 2004 / Accepted: 8 June 2004 / Published online: 14 July 2004  
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**Abstract** Oncocytic cell tumors (OCTs) of the thyroid include oncocytic cell adenomas (OCAs) and oncocytic cell carcinomas (OCCs). Oncocytic variant of papillary carcinoma (OVPC) has also been described. These tumors may present similar diagnostic problems as their non-oncocytic counterparts, in both conventional histology and fine-needle aspiration biopsies. Several markers were shown able to distinguish benign from malignant thyroid follicular tumors, galectin-3 and HBME-1 being the most promising ones. Controversial data have been reported on their discriminatory potential in the small series of OCTs so far analyzed. We aimed to assess the role of galectin-3 and HBME-1 in a large series of 152 OCTs (including 50 OCAs, 70 OCCs and 32 OVPCs). The expression of PPAR $\gamma$  protein was also evaluated. Using a biotin-free detection system, the sensitivity of galectin-3 was 95.1%, while that for HBME-1 was nearly 53%. The combination of galectin-3 and HBME-1 increased the sensitivity up to 99%. However, for both markers, the specificity was 88%, lower than that reported for non-oncocytic follicular tumors. PPAR $\gamma$  protein overexpression was absent in all OCAs tested and present in only 10% of OCCs, con-

firmer previous reports on the low prevalence of PAX8-PPAR $\gamma$  translocations in OCT and ruling out its role as a potential diagnostic marker of malignancy.

**Keywords** Thyroid · Tumors · Oncocytic · HBME-1 · Galectin-3

### Introduction

Oncocytic cell tumors (OCTs) of the thyroid include adenomas and carcinomas of follicular origin characterized by a predominant (usually more than 75% of the tumor area) population of eosinophilic mitochondrion-rich cells [18, 31]. They may be separated into oncocytic cell adenomas (OCAs) and oncocytic cell carcinomas (OCCs). An oncocytic variant of papillary carcinoma (OVPC) has also been described [13, 27]. Their biological behavior has been the matter of long-standing controversies. Originally, these tumors, also referred to as Hürthle cell or oxyphilic tumors, were all considered potentially malignant, based on the reported higher aggressive behavior than their conventional non-oncocytic follicular counterparts [3, 32]. More recently [6, 23], it became evident that the clinical outcome of OCTs was by no means different from that of the corresponding non-oncocytic tumors, irrespective of them being benign (adenomas) or malignant (well or poorly differentiated carcinomas). Nevertheless, their recognition is an important task for pathologists, since oncocytic cells may also be found in non-neoplastic conditions (goiter, thyroiditis). In addition, as in the case of non-oncocytic minimally invasive follicular carcinomas, the differential diagnosis of oncocytic adenomas from carcinomas may not be easily performed in all cases, and it is, in any case, impossible in cytological material from fine-needle aspiration (FNA) biopsies.

Several markers have been proposed, in both surgical and FNA cytological specimens, to distinguish follicular adenomas from carcinomas, including the oncocytic cell types. The most promising appeared to be galectin-3, cytokeratin 19, the mesothelial cell marker HBME-1 and

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thyroperoxidase [1, 4, 5, 8, 10, 11, 12, 22, 24, 25, 26, 28, 29]. In all published series, however, the number of OCTs studied was generally low. For galectin-3 and HBME-1 in particular, controversial data have been reported, indicating that the discriminatory potential of such markers in OCT was of limited value [14, 17]. To this purpose, we aimed to validate the role of galectin-3 and HBME-1 in a large series of 152 OCT, which included 50 OCAs, 70 OCCs and 32 OVPCs. In addition, we searched for PPAR $\gamma$  protein overexpression in all cases of OCA and OCC. PPAR $\gamma$  protein was found to be expressed by follicular carcinoma nuclei but not by adenomas [20], as a consequence of the PAX8-PPAR $\gamma$  translocation t(2;3)(q13;p25). This finding was not confirmed in subsequent reports and the specificity of this marker is rather poor [15]. Moreover, a very low rate of overexpression in OCC has been described in previous reports [20], thus excluding an involvement of PAX8-PPAR $\gamma$  translocation in the pathogenesis of this tumor type.

We here show that using a biotin-free detection system, galectin-3 alone is capable of correctly identifying 95% of malignant OCTs, and the association of galectin-3 and HBME-1 increases the sensitivity up to 99%.

## Materials and methods

### Case selection

We retrospectively analyzed 152 thyroid OCTs (including 50 OCAs, 70 OCCs and 32 OVPCs) and 34 non-neoplastic, oncocyctic, cell-rich control cases (including 19 cases of nodular goiter with oncocyctic changes and 15 cases of Hashimoto's thyroiditis). Tumor samples were collected from the pathology files of the Department of Pathology, University of Turin, from 1974 to 2003. All tumors were classified according to widely accepted histological criteria [13, 27], and a diagnosis of oncocyctic cell neoplasm was made when more than 75% of the cells had oncocyctic features. Diagnostic criteria of malignancy discriminating OCC from OCA were complete capsular penetration and/or invasion of blood vessels within the capsule or external to it. OVPC were diagnosed in the presence of the characteristic nuclear features of papillary carcinoma, irrespective of their growth pattern (either papillary or follicular).

### Immunohistochemical analysis

Immunohistochemistry was performed on paraffin-embedded sections using a standard manual biotin-free immunoperoxidase procedure with monoclonal antibodies against human galectin-3 (clone 9C4, diluted 1:200; Novocastra, Newcastle, UK), HBME-1 (clone

HBME-1, diluted 1:50; DakoCytomation, Glostrup, Denmark), and PPAR $\gamma$  (clone E-8, 1:300; Santa Cruz Biotechnology, Santa Cruz, CA). For all antibodies tested, antigen-retrieval treatment (three 3-min passages in citrate buffer, pH 6.0) was performed, and then immune complexes were detected with the EnVision system (DakoCytomation) to prevent endogenous biotin activity, and visualized by diaminobenzidine precipitation. Slides were then counterstained with Mayer's hematoxylin and mounted.

### Scoring of staining and statistical data

All cases were evaluated by three independent investigators, blind with respect to the histological diagnosis. Cytoplasmic galectin-3 and HBME-1-positive cases were scored as + ( $\leq 10\%$ ), ++ (11–50%) and +++ ( $> 50\%$ ), as previously reported [1, 14]. Nuclear staining for PPAR $\gamma$  was scored as weak (+), moderate (++) or strong (+++), and as focal ( $< 50\%$  of the tumor cells positive) or diffuse ( $> 50\%$  of tumor cells), as proposed by Nikiforova et al. [20]. According to these latter authors, in the final interpretation of the results, only cases with diffuse ( $> 50\%$  of tumor cells) and strong (+++) staining were considered positive (being correlated to the presence of the translocation at a molecular level).

Sensitivity, specificity, and diagnostic accuracy were assessed for each antibody tested, considered separately or in combination. Sensitivity was defined as true positive/(true positive+false negative) and specificity as true negative/(true negative+false positive). Diagnostic accuracy was defined as (true positive+true negative)/(true positive+false positive+true negative+false negative).

## Results

The immunohistochemical expression of galectin-3 and HBME-1 is summarized in Table 1. Among 50 cases of OCA, 44 were negative for galectin-3. In 6 cases (12%), a focal reactivity (score +) was observed. Accurate histological revision failed to detect in these latter cases the presence of capsular penetration or vascular invasion, or nuclear features suggestive of a follicular variant of papillary carcinoma. Cytoplasmic galectin-3 immunostaining was restricted to small clusters of tumor cells located beneath the tumor capsule. Cytoplasmic galectin-3 was expressed in 66 of 70 (94.3%) OCCs (Fig. 1) and in 31 of 32 (96.9%) OVPCs (Fig. 2). The immunoreactivity was typically strong and diffuse in cases of OVPC; while, in cases of OCC, it was mostly located at the periphery of the nodule and particularly in areas of capsular or vascular invasion, although some diffusely positive cases have been observed.

HBME-1 reactivity was detected in 6 of 50 OCAs (12%), 26 of 70 OCCs (37.1%) (Fig. 1) and 28 of 32 (87.5%) OVPCs. HBME-1 pattern of immunostaining

**Table 1** Immunohistochemical results of galectin-3, HBME-1 and PPAR $\gamma$  in a series of 152 oncocyctic cell tumors

Diagnosis	Galectin-3 <sup>a</sup>	HBME-1 <sup>a</sup>	PPAR $\gamma$ <sup>b</sup>
Oncocyctic cell adenoma	6 <sup>c</sup> /50 (12%)	6 <sup>c</sup> /50 (12%)	0/50 (0%)
Oncocyctic cell carcinoma	66/70 (94.3%)	26/70 (37.1%)	7/70 (10%)
Oncocyctic variant of papillary carcinoma	31/32 (96.9%)	28/32 (87.5%)	ND

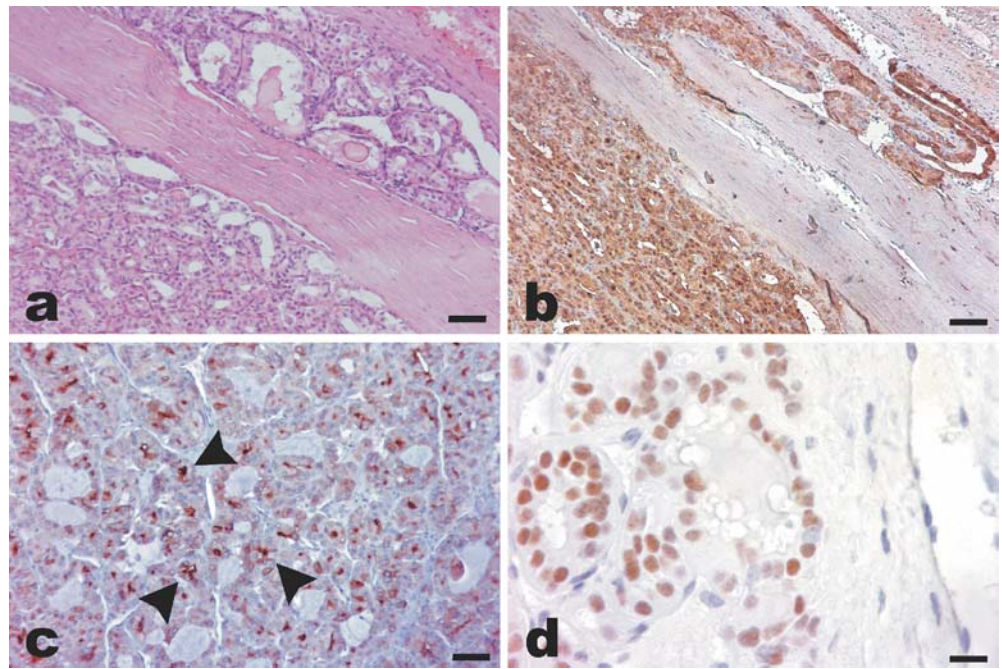
<sup>a</sup> Positive cases included scores + to +++, as detailed in the Materials and methods (see references [1, 14])

<sup>b</sup> Positive cases included only strong (+++) and diffuse ( $> 50\%$  of tumor cells) immunohistochemical features [20]

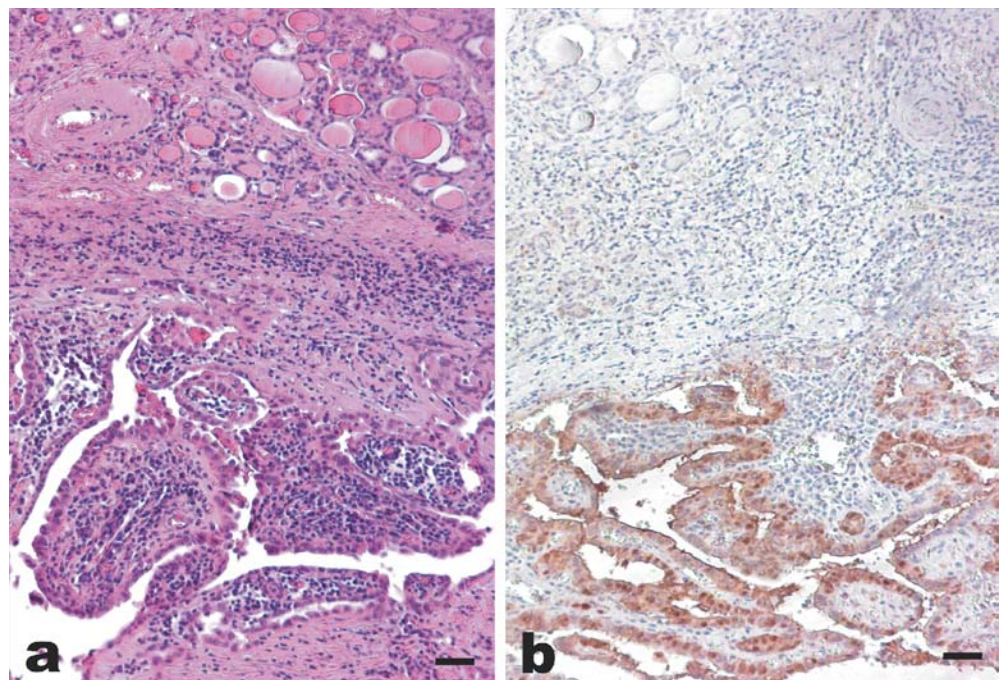
<sup>c</sup> All cases with + score



**Fig. 1** A case of oncocytic cell carcinoma (a), angioinvasive (top right) diffusely expressing cytoplasmic and nuclear galectin-3 in tumor cells and also in a blood vessel (top right; b). Tumor cells also co-express HBME-1 (c), with the typical endoluminal reinforcement (arrowheads) and PPAR $\gamma$  proteins (d). **a** Hematoxylin and Eosin,  $\times 100$ , bar 100  $\mu\text{m}$ . **b** Immunoperoxidase,  $\times 100$ , bar 100  $\mu\text{m}$ . **c** Immunoperoxidase,  $\times 200$ , bar 50  $\mu\text{m}$ . **d** Immunoperoxidase  $\times 400$ , bar 25  $\mu\text{m}$



**Fig. 2** A case of oncocytic variant of papillary carcinoma (a), strongly expressing cytoplasmic and nuclear galectin-3 (b). Normal peritumoral thyroid follicles show no reactivity for galectin-3 (top). **a** Hematoxylin and eosin,  $\times 100$ . **b** Immunoperoxidase,  $\times 100$ . **a, b** Bar 100  $\mu\text{m}$



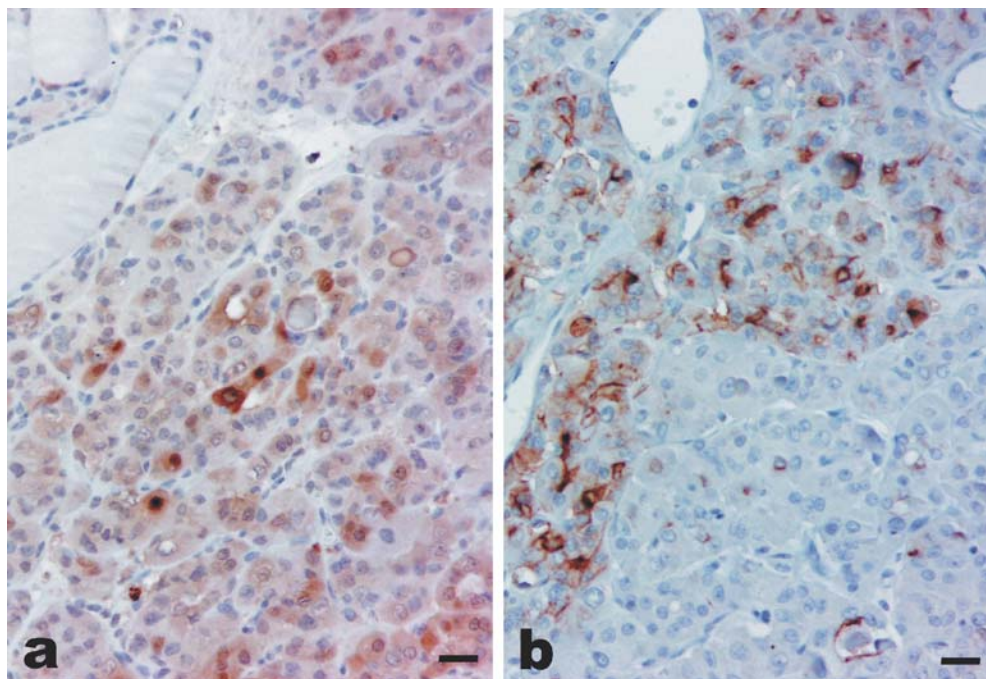
was diffuse in most cases, while in some cases a clustered reactivity was observed either in the center or in the periphery of the nodule (in contrast with that of galectin-3). With respect to “false-positive” OCAs, 4 cases were reactive for both galectin-3 and HBME-1 (Fig. 3). Concerning “false-negative” OCCs, interestingly, 3 of the 4 galectin-3-negative cases were positive for HBME-1. All cases of hyperplastic nodular goiters with oncocytic changes were unreactive for both galectin 3 and HBME-1. Of 15 cases of Hashimoto’s thyroiditis, 14 showed a moderate (score ++ ) to strong (score +++ ) immunoreac-

tivity for both galectin-3 and HBME-1 in clusters of oncocytic epithelial cells within lymphocytic infiltrates.

Immunostaining with PPAR $\gamma$  antibody was performed on 120 oncocytic thyroid tumors, comprising 50 OCA and 70 OCC (Table 1). Seven carcinomas (10%), being all but one minimally invasive, showed a strong and diffuse nuclear staining. In 6 cases (5 OCCs and 1 OCA), a weak to moderate nuclear staining (either focal or diffuse) was observed; since, according to the literature data [9, 20], only the strong diffuse nuclear staining is correlated with the presence of the PAX8-PPAR $\gamma$  rearrangement detected



**Fig. 3** A case of oncocytic cell adenoma expressing both galectin-3 (in the cytoplasm and nuclei of tumor cells; **a**) and HBME-1 proteins (**b**). In both cases, the reactivity is restricted to single cells or small clusters of tumor cells. **a, b** Immunoperoxidase  $\times 400$ , bar 25  $\mu\text{m}$



**Table 2** Sensitivity, specificity and diagnostic accuracy of galectin-3, HBME-1 and PPAR $\gamma$ , used alone or in combination

Immunohistochemical marker	Number of cases	Sensitivity (%)	Specificity (%)	Diagnostic accuracy (%)
GAL-3	152	95.1	88	92.8
HBME-1	152	52.9	88	64.5
PPAR $\gamma$	120	10	100	47.5
GAL-3 + HBME-1	152	99	80	92.8
GAL-3 + HBME-1 + PPAR $\gamma$	120	98.6	80	90.8

by RT-PCR, these latter cases were considered as negative.

Data on sensitivity, specificity and diagnostic accuracy of the three antibodies tested are reported in Table 2. Sensitivity ranged from 10% for PPAR $\gamma$  antibody to 95.1% for galectin-3. Although sensitivity for HBME-1 was very low (52.9%) in the present series of OCTs, the combination of galectin-3 and HBME-1 was shown to increase the general sensitivity up to 99%. Specificity was 88% for both galectin-3 and HBME-1, and slightly decreased using a combination of the two. A strong and diffuse PPAR $\gamma$  reactivity confirmed to be 100% specific of carcinomas. Finally, galectin-3 gained the highest diagnostic accuracy (92.8%) used alone or in combination with HBME-1, while HBME-1 alone showed a diagnostic accuracy of 64.5%, due to its low sensitivity.

## Discussion

In this study, we have shown that galectin-3 is a useful marker of oncocytic carcinomas, having a sensitivity of 95.1%. Its combined use with HBME-1, however, increased sensitivity to 99%, suggesting that their association in a panel of markers is optimal for better characterizing OCCs in histological and, possibly, cytological

specimens. These findings confirm the reported role of galectin-3 and HBME-1 when employed separately in conventional non-oncocytic tumors [1, 22] and also expand their usefulness for OCTs. Actually, the potential role of galectin-3 as a marker of malignancy in OCT as been questioned by some authors. In fact, a few recent papers suggested that galectin-3 immunodetection is not restricted to malignant tumors, but is also observed in adenomas [16, 17, 19].

A possible explanation for such reported discrepancies in OCT comes from the knowledge that oncocytic cells are rich in endogenous biotin and galectin-3 immunocytochemistry (as well as HBME-1) and may provide false-positive results in OCT using biotin-based detection systems, especially when heat-induced antigen retrieval methods (required in the case of galectin-3 immunocytochemistry) are employed. Therefore, as correctly pointed out by some authors [12, 30], galectin-3 immunodetection may be a useful adjunct to distinguish benign from malignant thyroid tumors, only if used in a biotin-free detection system.

Moreover, different technical procedures used to reveal galectin-3 may also explain controversial data from the literature. In some recent reports based on real-time reverse-transcription polymerase chain reaction (RT-PCR) methods [7], high levels of galectin-3 mRNA were de-

tected also in benign thyroid nodules. In our opinion, methods aimed to localize the molecule in the tissue, such as immunohistochemistry, seem to be more reliable in interpreting the final results, the cytoplasmic fraction of galectin-3 being the only specific sign of malignant transformation [2]. PCR-based techniques and in situ hybridization, although highly sensitive, can distinguish neither the cytoplasmic from the nuclear galectin-3 localization nor (with regard to PCR) the cell type expressing galectin-3 mRNA [4]. Since it is well known that macrophages and endothelial cells produce relatively large amounts of galectin-3 in physiological conditions, such cell types are probably present in the tissue submitted to PCR analysis, irrespective of the benign or malignant nature [2]. In addition, although the immunohistochemical expression of galectin-3 was found to correlate with mRNA expression [10], it is not completely understood how mRNA transcription is regulated in benign conditions.

Our data on 152 cases of OCTs, under strictly controlled technical conditions, showed a galectin-3 immunoreactivity in 95.1% malignant OCTs (including OCC and OVPC cases) and in 12% of OCAs, values that are consistent with those observed in non-oncogenic follicular and papillary tumors. Conversely, lower sensitivity, specificity and, therefore, diagnostic accuracy (52.9%, 88% and 64.5%, respectively) were observed for HBME-1, as also pointed out by other authors [14] with special reference to OVPC cases. Our data suggest a limited diagnostic role of HBME-1 used alone in the diagnosis of OCC (since only 37.1% of cases were positive), its sensitivity in OVPC being slightly below that of galectin-3 used alone. Nevertheless, HBME-1 was able to recognize three of the four galectin-3-negative OCCs. Although this result has poor diagnostic utility due to the low HBME-1 specificity (being positive in 12% of OCAs), it suggests that an alternative phenotype is probably present in this small group of tumors. The presence of two alternative and non-overlapping pathogenetic pathways of tumorigenesis in follicular carcinoma have already been postulated by some authors [21], the presence of the PAX8-PPAR $\gamma$  translocation being most frequently associated with a galectin-3-positive/HBME-1-negative immunophenotype and the presence of RAS mutations associated with a galectin-3-negative/HBME-1-positive immunophenotype. These data indirectly indicate that, in the diagnosis of follicular tumors in general, a panel of markers including at least galectin-3 and HBME-1 would probably increase the sensitivity of an immunohistochemical approach up to nearly 100%.

A separate comment is deserved for “false-positive” OCAs, i.e., four cases positive for both galectin-3 and HBME-1 and two cases positive for one of the two markers, each. They may represent, on the one hand, true false-positive cases by immunohistochemistry, but on the other may reflect lesions having a biologically malignant potential in the absence of morphological signs of invasive growth (as already described in conventional follicular adenomas) [1]. As opposed to molecular markers

(such as PAX8-PPAR $\gamma$  gene fusion product), which possess oncogenic implications and therefore are an early event in malignant transformation, galectin-3 and HBME-1 regulation in morphologically benign tumors, in tumors of uncertain malignant potential, as well as in early malignant lesions is poorly understood. In this respect, we were able to confirm the presence of immunoreactivity for both galectin-3 and HBME-1 in clusters of epithelial follicular cells with oncogenic changes in the setting of Hashimoto’s thyroiditis. These markers may be upregulated due to reactive processes rather than neoplastic transformation. The exact interactions between lymphoid and follicular epithelial cells, as well as the processes leading to oncogenic transformation in Hashimoto’s thyroiditis, are far to be understood. However, the positive cells often present a mild to moderate degree of atypia, with nuclear features resembling those of papillary carcinoma, although clear molecular markers of oncogenic transformation, typical of papillary carcinoma, such as RET/PTC translocation, have not been definitely identified. Therefore, further studies are required to better understand the biological significance of such morphological and immunophenotypical features.

With regard to PAX8-PPAR $\gamma$  gene fusion protein, we have investigated a large series of 120 cases by means of immunohistochemistry, to validate previous reports, based on small number of cases, which failed to detect PPAR $\gamma$  abnormalities in OCT (either by RT-PCR or immunohistochemistry). Since a good correlation has been reported between the molecular detection of PAX8-PPAR $\gamma$  translocation and a strong and diffuse PPAR $\gamma$  protein overexpression, only immunohistochemistry was employed (also to check the possible applications of PPAR $\gamma$  as a diagnostic marker of OCC). Our data confirmed that PPAR $\gamma$  protein over-expression occurred in a very small number of OCCs, all being OCA negative. These results support the hypothesis that OCCs may follow distinct molecular pathways from those of non-oncogenic follicular carcinomas [20].

In conclusion, the present data confirm a high sensitivity and diagnostic accuracy of galectin-3 immunohistochemistry in the diagnosis of malignant OCT, at variance with HBME-1, which had a much lower sensitivity and specificity in OCT than that reported for non-oncogenic follicular tumors (an exception being OVPC). However, the combined use of these two markers was able to improve the sensitivity up to 99%, and therefore a panel including galectin-3 and HBME-1 is recommended in the diagnosis of OCT, although the low specificity of HBME-1 has to be considered when dealing with galectin-3-negative/HBME-1-positive OCT cases. Such a panel of markers is probably the best choice in FNA cytological diagnosis of OCT, to obtain the highest sensitivity in selecting presumably malignant cases to be sent to the surgeon.

**Acknowledgements** The work was supported by grants from the Italian Ministry of University and Education (ex 60% to M.P.), from the Regione Piemonte (D.D. no.173 dated October 30, 2003,

to F.O.), and The Compagnia di San Paolo "Progetto Speciale Oncologia".

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