



Differential *MC5R* loss in whales and manatees reveals convergent evolution to the marine environment

Jian Liu^{1,2} · Mingrong Shu³ · Shaobo Liu¹ · Jingwen Xue¹ · Haidi Chen² · Wen Li¹ · Jingfan Zhou¹ · Amanullah Amanullah¹ · Miao Guan¹ · Ji Bao⁴ · Dan Pu⁵ · Cheng Deng^{1,2}

Received: 31 January 2022 / Accepted: 16 May 2022 / Published online: 1 June 2022
© The Author(s), under exclusive licence to Springer-Verlag GmbH Germany, part of Springer Nature 2022

Abstract

Melanocortin 5 receptor (*MC5R*), which is expressed in the terminally differentiated sebaceous gland, is a G protein-coupled receptor (GPCR). *MC5R* exists mostly in mammals but is completely lost in whales; only the relic of *MC5R* can be detected in manatees, and phenotypically, they have lost sebaceous glands. Interestingly, whales and manatees are both aquatic mammals but have no immediate common ancestors. The loss of *MC5R* and sebaceous glands in whales and manatees is likely to be a result of convergent evolution. Here, we find that *MC5R* in whales and manatees are lost by two different mechanisms. Homologous recombination of *MC5R* in manatees and the insertion of reverse transcriptase in whales lead to the gene loss, respectively. On one hand, in manatees, there are two “TTATC” sequences flanking *MC5R*, and homologous recombination of the segments between the two “TTATC” sequences resulted in the partial loss of the sequence of *MC5R*. On the other hand, in whales, reverse transcriptase inserts between *MC2R* and *RNMT* on the chromosome led to the loss of *MC5R*. Based on these two different mechanisms for gene loss in whales and manatees, we finally concluded that *MC5R* loss might be the result of convergent evolution to the marine environment, and we explored the impact on biological function that is significant to environmental adaptation.

Keywords *MC5R* · Whales · Manatees · Mechanisms of gene lost · Convergent evolution

Responsible Editor: Matthias Hammerschmidt

Jian Liu and Mingrong Shu contributed equally to this work.

✉ Ji Bao
baoji@scu.edu.cn

✉ Cheng Deng
dengcheng2014@126.com

¹ Jiangsu Key Laboratory for Biodiversity and Biotechnology, College of Life Sciences, Nanjing Normal University, 1 Wenyuan Rd, Nanjing 210023, China

² Institutes for Systems Genetics, Frontiers Science Center for Disease-Related Molecular Network, National Clinical Research Center for Geriatrics, West China Hospital, Sichuan University, Chengdu, China

³ Department of Infection Control, West China Hospital, Sichuan University, Chengdu, Sichuan, China

⁴ Institute of Clinical Pathology, Key Laboratory of Transplant Engineering and Immunology, NHC, West China Hospital, Sichuan University, Chengdu 610041, Sichuan Province, China

⁵ West China Medical Simulation Center West China Hospital, Sichuan University, Chengdu, Sichuan Province, China

Introduction

Convergent evolution, which occurs ranging from the molecular level to the behavioral level, is considered a similar organismal trait evolving independently from different ancestors due to similar selective pressures in the environment (Zakon 2002). The dynamic of convergent evolution is to cope with a new stimulus and minimize unnecessary energy consumption when individuals face repetitive innocuous stimuli, which improves the survival of an organism (van Duijn 2017). Echolocation is a case of convergent evolution where similar traits evolved through identical genetic changes in bats and dolphins (Liu et al. 2010). Convergent adaptation shows great potential for the prediction of evolution and the procession of biological diversity. Gene loss is one way to generate convergent evolution (Branco et al. 2018).

G protein-coupled receptors (GPCRs), which are also known as seven-transmembrane (7TM) receptors, represent the largest and most diverse superfamily in vertebrates (Pedersen et al. 2018). Melanocortin 5 receptor (*MC5R*), a classic GPCR and one of five melanocortin receptor genes in

placental mammals, is important in the regulation of energy homeostasis (Cone 2005). *MC5R* is expressed in terminally differentiated sebaceous glands, and *MC5R* knockout mice showed reduced water rejection after swimming (Chen et al. 1997). Interestingly, in both whales and manatees, sebaceous glands are lost or degenerative (Springer and Gatesy 2018), which may be the reason for the reduced requirement for *MC5R* (Springer and Gatesy 2018; Wang et al. 2015). With the exception of the loss of sebaceous glands and the reduction in hair, other similar water-dependent features in whales are retained, such as underwater communication and hearing, underwater birth, loss of scrotal testes, and dense limb bones to overcome buoyancy (Barklow 2004, Boissarie et al. 2011; Coughlin and Fish 2009; Gatesy 1997; Gatesy et al. 2013; Spaulding et al. 2009; Tsagkogeorga et al. 2015).

Homologous recombination and insertion of transposons or retrotransposons leading to gene loss have been reported by several studies (Dahal et al. 2018; Krom and Ramakrishna 2012; Sedivy and Sharp 1989). On one hand, homologous recombination plays an important role in DNA template switching; recombination is a common event leading to allelic loss (Henson et al. 1991). The loss of the agouti signaling protein gene in the lesser apes is mediated by unequal homologous recombination (Nakayama and Ishida 2006). Similarly, micro-homologous recombination of 5–25 base pairs efficiently repairs double-stranded breaks created during murine B lymphocyte development (Nussenzweig and Nussenzweig 2007). On the other hand, the insertion of transposons or retrotransposons leads to gene loss (Kanazawa et al. 2009). Homolog pairing, which plays a critical role in meiosis, poses a potential risk if it occurs in inappropriate tissues or between nonallelic sites, as it can lead to changes in gene expression, chromosome entanglements, and loss-of-heterozygosity due to mitotic recombination (Joyce et al. 2013). In retrotransposons, enzymes and proteins that encode transposons are called autonomous transposons and can independently complete the transposon process (Wisman et al. 1998). The insertion of retrotransposons can induce mutations near or within genes (Niu et al. 2019). The FAIRE signal is lost in promoters and enhancers of active genes and gained in heterochromatic gene-poor regions that make senescent cells smooth. Chromatin of major retrotransposon classes, Alu, SVA, and LINE-1 (Long interspersed nuclear element-1), becomes relatively more open in senescent cells, affecting most strongly the evolutionarily recent elements, and leads to an increase in their transcription and ultimately transposition (Cecco et al. 2013). Furthermore, retrotransposon-induced mutations are stable due to their replication mechanism (Monden et al. 2014). LINE-1 is the largest retrotransposon family in the human genome and is the only family capable of autonomous transposition, accounting for 17% of the genome (Goodier and Kazazian 2008). Active retrotransposons play a great role in biological evolution and species formation (Rajput 2015). For example, a heterozygous

frameshift mutation was detected in Meckel-Gruber syndrome, which is a rare ciliopathy disease. It was detected that the insertion of LINE-1 affected exon 7 in the paternally derived allele (Takenouchi et al. 2017). However, in normal somatic cells, in order to maintain the stability of the genome, host cells have strict control over the transposition of LINE-1 (Ye et al. 2017).

Although *MC5R* was reported to be completely lost in whales and mostly deleted in manatees (Springer and Gatesy 2018), the molecular mechanism underlying *MC5R* loss remained poorly understood. In this study, we found differential molecular mechanism for *MC5R* loss in whales and manatees and revealed evidence of convergent evolution to the marine environment.

Materials and methods

Database querying and BLAST searches

In *Homo sapiens*, the protein-coding sequences for *MC5R* have 978 nucleotides. Protein-coding sequences of representative placental taxa that have annotated genomes (human: *Homo sapiens*; white-tufted-ear marmoset: *Callithrix jacchus*; mouse: *Mus musculus*; Norway rat: *Rattus norvegicus*; bottlenose dolphin: *Tursiops truncatus*; killer whale: *Orcinus orca*; Beluga whale: *Delphinapterus leucas*; sperm whale: *Physeter catodon*; Minke whale: *Balaenoptera acutorostrata scammoni*; Yangtze River dolphin: *Lipotes vexillifer*; wild yak: *Bos mutus*; cattle: *Bos taurus*; dog: *Canis lupus familiaris*; walrus: *Odobenus rosmarus*; Weddell sea: *Leptonychotes weddellii*; African savanna elephant: *Loxodonta Africana*; and Florida manatee: *Trichechus manatus latirostris*) were extracted from NCBI. Sequences of *Homo sapiens* and *Bos mutus* were used as query sequences to BLAST against other placental mammals.

Phylogenetic analysis

The coding sequences downloaded from NCBI were aligned by Clustal W version 1.83 (Thompson and Higgins 1994) with default settings. Phylogenetic trees were constructed using one algorithm neighbor-joining (NJ) with 2000 bootstrap replicates in MEGA version 4 (Tamura et al. 2007).

Results and discussion

Homologous recombination mechanism of *MC5R* loss in manatees

Marine mammals do not represent a distinct taxon or systematic grouping, but have a multil lineage relationship owing to convergent evolution, as they do not have a

direct common ancestor (Jefferson et al. 1995). Based on molecular systematics, among marine mammals, whales belong to Cetartiodactyla and *Trichechus manatus latirostris* belong to Sirenia (Springer and Gatesy 2018) (Fig. 1). Adaptation to the aquatic lifestyle of marine mammals varies considerably between species. Whales and manatees are universally recognized as fully aquatic marine mammals. Additionally, the presence of hair is densely distributed in most mammalian species and is also closely related to the sebaceous glands (Springer and Gatesy 2018; Li et al. 2006). However, some taxa are largely hairless, or hairs are sparse, including whales and manatees, which both lost *MC5R* (Folk and Semken 1991). During the evolution of whales and manatees, the function of sebaceous glands deteriorated slowly after the transition from land to sea. In addition, after sebaceous glands were lost in whales and manatees, *MC5R* was lost or inactivated. Therefore, DNA sequences of the *MC5R* gene among various species were aligned to find molecular evolutionary aspects of *MC5R*. *MC5R* was found to be located between *MC2R* and *RNMT* in most mammals in large-scale studies of 10 classical mammals and 6 whales (Fig. 1). The genomic location of *MC5R* flanking *RNMT* and *MC2R* is highly conserved in mammals (Fig. 1). In most mammals, such as *Homo sapiens* (human), *Callithrix jacchus* (white-tufted-ear marmoset), *Mus musculus* (mouse), *Rattus norvegicus* (Norway rat), *Bos taurus* (cattle), *Canis lupus familiaris* (dog), *Odobenus rosmarus* (walrus), *Leptonychotes*

weddellii (Weddell seal), and *Loxodonta africana* (African savanna elephant), *MC5R* is totally present (Fig. 1). However, *MC5R* is lost in whales and manatees, indicating that the levels of *MC5R* retention vary in mammals (Fig. 1). Only *MC5R* relics are detected in manatees, and no *MC5R* sequences could be found in whales (*Tursiops truncatus*, *Orcinus orca*, *Delphinapterus leucas*, *Lipotes vexillifer*, *Physeter catodon*, and *Balaenoptera acutorostrata scammoni*) (Fig. 2). Based on the above findings, loss of *MC5R* in aquatic animals is speculated to result from evolution to the marine environment. Taken together, our results suggest convergent evolution to the marine environment in whales and manatees.

MC5R is present in *L. africana*, *Desmodus rotundus*, and *Chrysochloris asiatica*, while in *T. m. latirostris*, as described by Springer and Gatesy (Springer and Gatesy 2018), there is a 2823-bp deletion (relative to *L. africana*) which includes 1991 bp of 5'UTR sequences and 832 bp coding sequence of *MC5R* (Fig. 3). By comparing the genomes of *L. africana* and *T. m. latirostris*, we find that in *L. africana*, there are two homologous “TTATC” sequences that are relatively conserved on the chromosomes containing *MC5R* (Fig. 3). The first one is located upstream of coding sequence of *MC5R*, and the second one is 131 bp before the stop codon of *MC5R* (Fig. 3). In *T. m. latirostris*, there is only the second homologous sequence “TTATC” with a relic of *MC5R* retained.

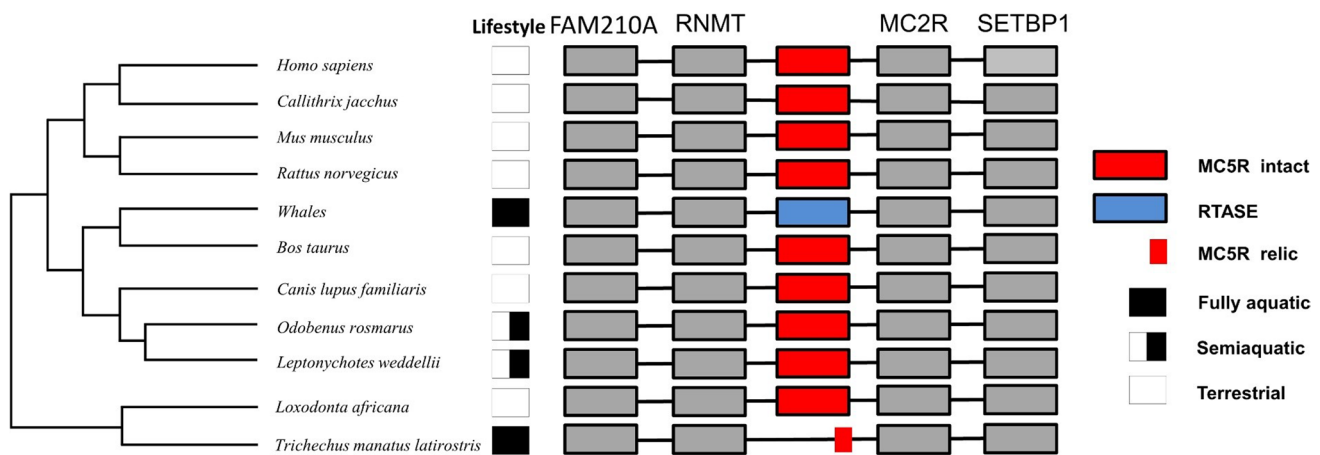


Fig. 1 Genomic location of *MC5R* in mammals. Phylogenetic tree follows previous studies (Tarver, et al. 2016), showing the relationships among 10 mammals and 6 whales used in the study. Outgroup branch lengths are not drawn to scale. Information about the lifestyle and the genomic location of *MC5R* in 16 mammals. The complete boxes are denoted in red (*MC5R* intact), blue (RTASE), and gray (conserved genes around *MC5R*). The incomplete box in red represents an *MC5R* relic. The squares beside the phylogenetic tree indicate the lifestyle. The black square represents fully aquatic, the white one represents terrestrial, and the half black-half white square

indicates semiaquatic. Human: *Homo sapiens*; white-tufted-ear marmoset: *Callithrix jacchus*; mouse: *Mus musculus*; Norway rat: *Rattus norvegicus*; whales (Bottlenose dolphin: *Tursiops truncatus*; killer: *Orcinus orca*; Beluga: *Delphinapterus leucas*; sperm: *Physeter catodon*; Minke: *Balaenoptera acutorostrata scammoni*; Yangtze River dolphin: *Lipotes vexillifer*); cattle: *Bos taurus*; dog: *Canis lupus familiaris*; walrus: *Odobenus rosmarus*; Weddell sea: *Leptonychotes weddellii*; African savanna elephant: *Loxodonta africana*; Florida manatee: *Trichechus manatus latirostris*

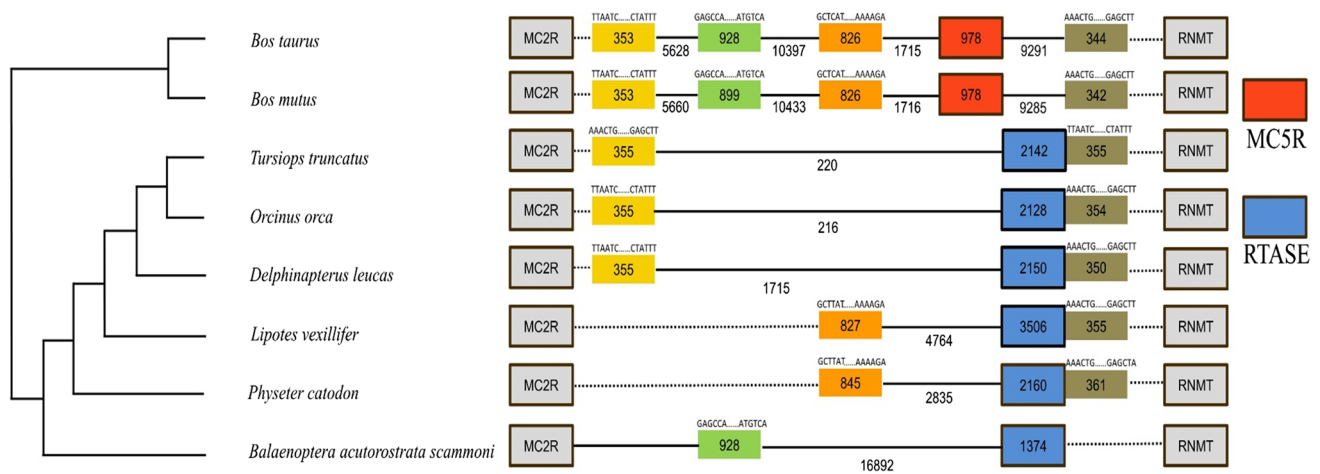


Fig. 2 Genomic structure of whales after converging evolution. The tree topology among different whales follows previous studies (Gatesy et al. 2013). Outgroups branch lengths are not drawn to scale. Schematic representation of the gene organization comparisons in different whales, *Bos taurus* and *Bos mutus*. The sequences located between *MC2R* and *RNMT* are highly conserved. The sequences located in yellow show similarity in *Bos taurus*, *Bos mutus*, *Tursiops truncatus*, *Orcinus orca*, and *Delphinapterus leucas*. The sequences in green are conserved in *Bos taurus*, *Bos mutus*, and *Balaenoptera acutorostrata scammoni*, and the sequences in orange are conserved in *Bos taurus*, *Bos mutus*, *Lipotes vexillifer*, and *Physeter catodon*. The brown

sequences can be detected in all the species except *Balaenoptera acutorostrata scammoni*. The difference between *Bos taurus* and whales is that *MC5R* is located between these conserved sequences in *Bos taurus*, while *RTASE* is inserted in whales, which led to the loss of *MC5R*. The numbers above the solid lines and on the boxes indicate the size of the introns and exons, respectively. The dashed line indicates an uncertain number of the introns and exons. Cattle: *Bos taurus*; wild yak: *Bos mutus*; bottlenose dolphin: *Tursiops truncatus*; killer: *Orcinus orca*; Beluga: *Delphinapterus leucas*; Yangtze River dolphin: *Lipotes vexillifer*; sperm: *Physeter catodon*; Minke: *Balaenoptera acutorostrata scammoni*

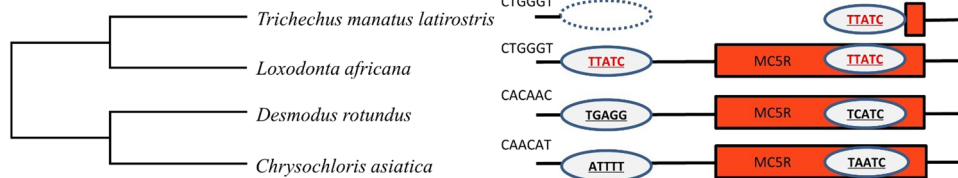


Fig. 3 *MC5R* gene with upstream and downstream sequences of closely related species in manatees. Phylogenetic tree follows previous studies (Tarver, et al. 2016), showing the relationships among *Trichechus manatus latirostris*, *Loxodonta africana*, *Desmodus rotundus*

dusus, and *Chrysochloris asiatica*. *MC5R* is marked in red; “TTATC” and other similar mutation sequences are marked with ellipses. It shows that specific mutations have led to homologous recombination and gene loss in *Trichechus manatus latirostris*

The upstream sequence of the first homologous sequence “TTATC” (“CTGGGT”) is also relatively conserved in *T. m. latirostris* and *L. africana* (Fig. 3). However, the first homologous “TTATC” sequences away from the *MC5R* are not absolutely conserved. The mutations occur in the genome “TGAGG” in *D. rotundus* and “ATTTT” in *C. asiatica* (Figs. 3 and 4; Fig. S2). In terms of *D. rotundus* and *C. asiatica*, according to the results of a genomic BLAST, the second homologous “TTATC” sequences on *MC5R* are still conserved: “TCATC” in *D. rotundus* and “TAATC” in *C. asiatica* (Figs. 3 and 4; Fig. S2A and B). In the genome sequences analysis between *L. africana* and *T. m. latirostris*, we find that the loss of most sequences of *MC5R* in *T. m. latirostris* is caused by the recombination of homologous sequence “TTATC” (Fig. 4A). The high conservatism of the sequence adjacent to the “TAATC” sequence can be regarded as the reliability of the comparison results (Figs. 3 and 4; Fig. S2).

The mechanism of *MC5R* loss in whales

Unlike *T. m. latirostris*, *MC5R* sequences are completely lost in whales. Based on the phylogenetic tree of the species, the relationship among whales, *B. taurus* and *B. mutus*, is relatively close. Genome sequences of *T. truncatus*, *O. orca*, *D. leucas*, *P. catodon*, *L. vexillifer*, and *B. a. scammoni* were obtained to analyze the absence of *MC5R* in whales (Fig. 2). However, the complete *MC5R* coding sequences of six whales are missing. *MC5R* flanking with *MC2R* and *RNMT* and some noncoding sequences were found between *MC2R* and *RNMT* in these species (Fig. 2). Sequences are marked with different colors and highly conserved between sequences marked by the same color (Fig. 3; Figure S1A and B).

MC5R is located between the conserved sequences in *B. taurus* and *B. mutus*, while *MC5R* is replaced by reverse transcriptase (*RTASE*) in whales. Furthermore, the lack of

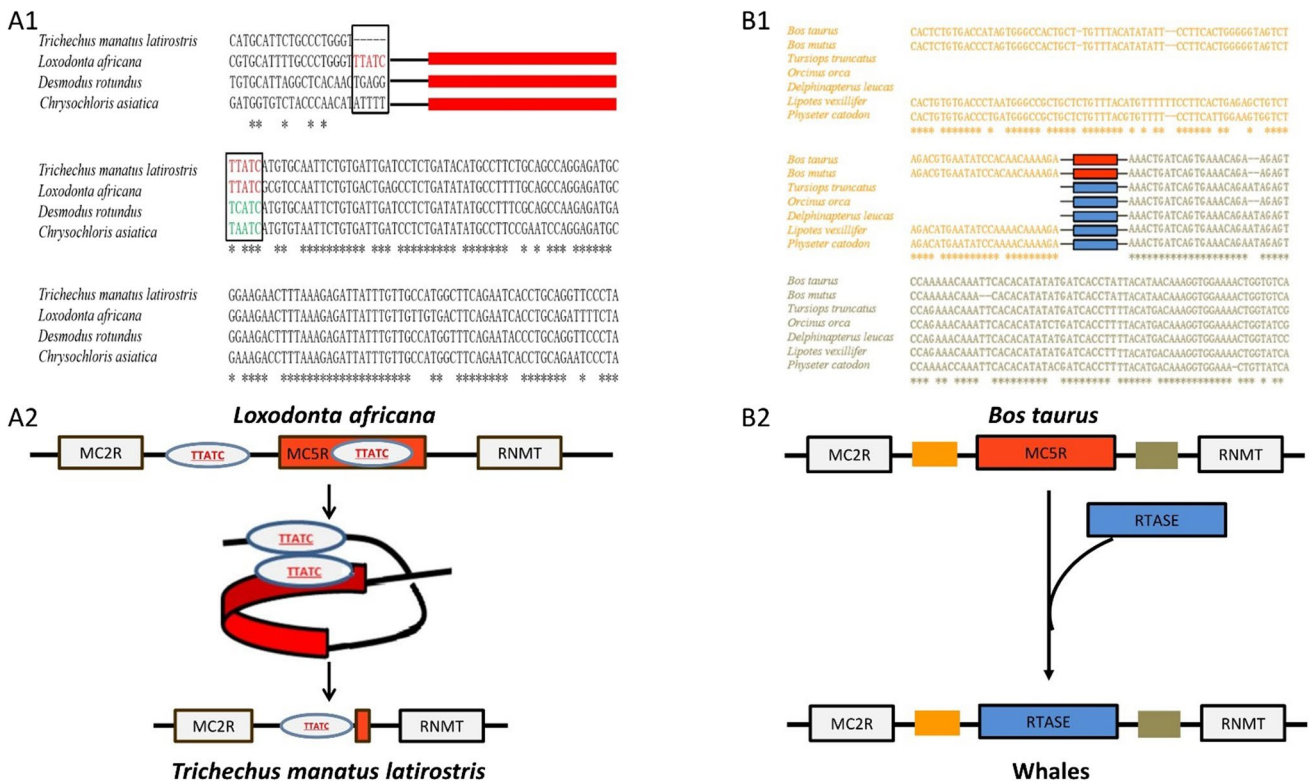


Fig. 4 Two different mechanisms of *MC5R* loss in manatees and whales. **A** Sequence alignment of *Trichechus manatus latirostris*, *Loxodonta africana*, *Desmodius rotundus*, and *Chrysochloris asiatica* is shown in A1. The conserved sequence “TTATC” and mutant sequence are indicated in red and green, respectively. The red box represents *MC5R*. In the genome of manatees, most *MC5R* sequences are lost and only a small number of sequences remain. In the gene of *Loxodonta africana*, there are two homologous sequences “TTATC” on the chromosome containing the *MC5R* gene. One is located between *MC5R* and *MC2R*; the other one is located on *MC5R*. However, in manatees, the sequences between the two “TTATC” are lost,

common *MC5R* ORF in whales indicates that these deficiencies occurred in a common ancestor. L1s are a class of repetitive DNA sequences that can spontaneously “copy-paste” themselves in the human genome (Hancks et al. 2011). After sequence alignment, there is only an RTASE in the LINE-1 insertion, and other elements are lost. Furthermore, the location of the RTASE insertion is like that of *MC5R* on the chromosome in *B. taurus* and *B. mutus* (Fig. 2). Consistent with the *MC5R* flanking sequence of *B. taurus* and *B. mutus*, the RTASE flanking sequences showed high similarity (Fig. 4B). Synthesizing all the discoveries and analyzes above, we suppose that ultimately, the absence of *MC5R* in whales is due to the insertion of a special enzyme, RTASE. Thus, whales tend to adapt to the marine environment during evolution.

The obtained results show that gene loss in whales and manatees exposes an occurrence of convergent evolution to the marine environment, and we try to explore the functional

and together with one “TTATC,” only a relic of *MC5R* is retained. **B** In the whale genome, the whole *MC5R* gene is lost, and RTASE is inserted. Sequence alignment of *Bos taurus*, *Bos mutus*, *Tursiops truncatus*, *Orcinus orca*, *Delphinapterus leucas*, *Lipotes vexillifer*, and *Physeter catodon* is shown in B1. The red box indicates *MC5R*, and the blue indicates RTASE. The conserved sequences before *MC5R* are marked in orange, and the ones after *MC5R* are marked in brown. The intact *MC5R* is located between the conserved sequence of *Bos taurus* and *Bos mutus*. In whales, *MC5R* is completely lost and RTASE is inserted

impact that is significant to environmental adaptation. The reason why this gene is not lost in semiaquatic mammals is currently unidentified and would be an interesting and important topic for further research.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s00427-022-00688-1>.

Author contribution Ji Bao is the cooperation teacher of West China Hospital, Sichuan University. Jian Liu and Mingrong Shu play a guiding role in revise and editing manuscripts. Cheng Deng and Ji Bao designed the study. HaiDi Chen and Miao Guan wrote the first draft. Shaobo Liu, Jingwen Xue, HaiDi Chen, Wen Li, Jingfan Zhou, Amanullah, and Dan Pu designed, performed, and analyzed the data.

Funding This work was supported by National Natural Science Foundation of China (31970388, 32170498); the National Key Research and Development Program of China (2021YFF0702000, 2018YFD0900602); 1.3.5 project for disciplines of excellence, West China Hospital, Sichuan University (ZYJC21050); Science and

Technology Department of Sichuan Province (2022YFH0116); and the Priority Academic Program Development of Jiangsu Higher Education Institutions (PAPD).

Data availability All data generated or analyzed during this study are included in this published article and its supplementary information files.

Declarations

Competing interests The authors declare no competing interests.

References

- Barklow WE (2004) Amphibious communication with sound in hippos *Hippopotamus amphibius*. *Anim Behav* 68(Pt5):1125–1132
- Boisserie JR, Fisher RE, Lihoreau F, Weston EM (2011) Evolving between land and water: key questions on the emergence and history of the Hippopotamidae (Hippopotamoidea, Cetartiodactyla). *Biol Rev* 86(3):601–625
- Branco S, Carpentier F, de la Vega RCR, Badouin H, Snirc A, Le Prieur S, Coelho MA, de Vienne DM, Hartmann FE, Begerow D, Hood ME, Giraud T (2018) Multiple convergent supergene evolution events in mating-type chromosomes. *Nat Commun* 9(1):2000
- Cecco MD, Criscione SW, Peckham EJ, Hillenmeyer S, Hamm EA, Manivannan J, Peterson AL, Kreiling JA, Neretti N, Sedivy JM (2013) Genomes of replicatively senescent cells undergo global epigenetic changes leading to gene silencing and activation of transposable elements. *Aging Cell* 12(2):247–256
- Cone RD (2005) Anatomy and regulation of the central melanocortin system. *Nat Neurosci* 8(5):571–578
- Coughlin BL, Fish FE (2009) Hippopotamus underwater locomotion: reduced-gravity movements for a massive mammal. *J Mammal* 90(3):675–679
- Chen W, Kelly MA, Opitz-Araya X, Thomas RE, Low MJ, Cone RD (1997) Exocrine gland dysfunction in MC5-R-deficient mice: evidence for coordinated regulation of exocrine gland function by melanocortin peptides. *Cell* 91(6):789–798
- Dahal S, Dubey S, Raghavan SC (2018) Homologous recombination-mediated repair of DNA double-strand breaks operates in mammalian mitochondria. *Cell Mol Life Sci* 75(9):1641–1655
- Duijn V, Marc. (2017) Phylogenetic origins of biological cognition: convergent patterns in the early evolution of learning. *Interface Focus* 7(3):20160158
- Folk GE, Semken HA (1991) The evolution of sweat glands. *Int J Biometeorol* 35(3):180–186
- Gatesy J (1997) More DNA support for a Cetacea/Hippopotamidae clade: the blood-clotting protein gene gamma-fibrinogen. *Mol Biol Evol* 14(5):537–543
- Gatesy J, Geisler JH, Chang J, Buell C, Berta A, Meredith RW, Springer MS, McGowen MR (2013) A phylogenetic blueprint for a modern whale. *Mol Phylogenet Evol* 66(2):479–506
- Goodier JL, Kazazian HH Jr (2008) Retrotransposons revisited: the restraint and rehabilitation of parasites. *Cell* 135(1):23–35
- Hancks DC, Goodier JL, Mandal PK, Cheung LE, Kazazian HH (2011) Retrotransposition of marked SVA elements by human L1s in cultured cells. *Hum Mol Genet* 20(17):3386–3400
- Henson V, Palmer L, Banks S, Nadeau JH, Carlson GA (1991) Loss of heterozygosity and mitotic linkage maps in the mouse. *Proc Natl Acad Sci U S A* 88(15):6486–6490
- Jefferson TA, Jefferson TA, Jefferson TA and Jefferson TA. 1995. Marine mammals of the world, G.G. Hararr
- Joyce EF, Apostolopoulos N, Beliveau BJ, Wu CT, Hawley RS (2013) Germline progenitors escape the widespread phenomenon of homolog pairing during *Drosophila* development. *PLoS Genet* 9(12):e1004013
- Kanazawa A, Liu B, Kong F, Arase S, Abe J (2009) Adaptive evolution involving gene duplication and insertion of a novel Ty1/copia-like retrotransposon in soybean. *J Mol Evol* 69(2):164–175
- Krom N, Ramakrishna W (2012) Retrotransposon insertions in rice gene pairs associated with reduced conservation of gene pairs in grass genomes. *Genomics* 99(5):308–314
- Li Z, Li WH, Anthonavage M, Eisinger M (2006) Melanocortin-5 receptor: a marker of human sebocyte differentiation. *Peptides* 27(2):413–420
- Liu Y, Cotton JA, Shen B, Han XQ, Rossiter SJ, Zhang SY (2010) Convergent sequence evolution between echolocating bats and dolphins. *Curr Biol* 20(2):R53–R54
- Monden Y, Fujii N, Yamaguchi K, Ikeo K, Nakazawa Y, Waki T, Hirashima K, Uchimura Y, Tahara M, Bureau TE (2014) Efficient screening of long terminal repeat retrotransposons that show high insertion polymorphism via high-throughput sequencing of the primer binding site. *Genome* 57(5):245–252
- Nakayama K, Ishida T (2006) Alu-mediated 100-kb deletion in the primate genome: the loss of the agouti signaling protein gene in the lesser apes. *Genome Res* 16(4):485–490
- Niu XM, Xu YC, Li ZW, Bian YT, Hou XH, Chen JF, Zou YP, Jiang J, Wu Q, Ge S, Balasubramanian S, Guo LY (2019) Transposable elements drive rapid phenotypic variation in *Capsella rubella*. *Proc Natl Acad Sci U S A* 116(14):6908–6913
- Nussenzweig A, Nussenzweig MC (2007) A backup DNA repair pathway moves to the forefront. *Cell* 131(2):223–225
- Pedersen JE, Bergqvist CA, Larhammar D (2018) Evolution of the muscarinic acetylcholine receptors in vertebrates. *Eneuro* 5(5):0340–18.2018
- Rajput MK (2015) Retrotransposons: the intrinsic genomic evolutionist. *Genes Genom* 37(2):113–123
- Sedivy JM, Sharp PA (1989) Positive genetic selection for gene disruption in mammalian cells by homologous recombination. *Proc Natl Acad Sci U S A* 86(1):227–231
- Spaulding M, O’Leary MA, Gatesy J (2009) Relationships of Cetacea (Artiodactyla) among mammals: increased taxon sampling alters interpretations of key fossils and character evolution. *PLoS One* 4(9):e7062
- Springer MS, Gatesy J (2018) Evolution of the MC5R gene in placental mammals with evidence for its inactivation in multiple lineages that lack sebaceous glands. *Mol Phylogenet Evol* 120:364–374
- Takenouchi T, Kuchikata T, Yoshihashi H, Fujiwara M, Uehara T, Miyama S, Yamada S, Kosaki K (2017) Diagnostic use of computational retrotransposon detection: Successful definition of pathogenic mechanism in a ciliopathy phenotype. *Am J Med Genet A* 173(5):1353–1357
- Tamura K, Dudley J, Nei M, Kumar S (2007) MEGA4: molecular evolutionary genetics analysis (MEGA) software version 4.0. *Mol Biol Evol* 24(8):1596–1599. <https://doi.org/10.1093/molbev/msm092>
- Thompson JD, Higgins D (1994) CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Res* 22(22):4673–4680
- Tsagkogeorga G, McGowen MR, Davies KTJ, Jarman S, Polanowski A, Bertelsen MF, Rossiter SJ (2015) A phylogenomic analysis of the role and timing of molecular adaptation in the aquatic transition of cetartiodactyl mammals. *R Soc Open Sci* 2(9):150156
- Wang Z, Chen Z, Xu S, Ren W, Yang G (2015) ‘Obesity’ is healthy for cetaceans? Evidence from pervasive positive selection in genes related to triacylglycerol metabolism. *Entific Rep* 5:14187

Wisman E, Cardon GH, Fransz P, Saedler H (1998) The behaviour of the autonomous maize transposable element *En/Spm* in *Arabidopsis thaliana* allows efficient mutagenesis. *Plant Mol Biol* 37(6):989–999

Ye ZJ, Liu QP, Shan C, Li XY (2017) The function of LINE-1-encoded reverse transcriptase in tumorigenesis. *Hereditas* 39(5):368–376

Zakon HH (2002) Convergent evolution on the molecular level. *Brain Behav Evol* 59(5–6):250–261

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.