

Cardioprotection by remote ischemic conditioning and its signal transduction

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Abstract Cardioprotective strategies aim to salvage myocardium from ischemia/reperfusion injury and to reduce infarct size and its consequences. Different stimuli, acting at sites remote from the heart (remote conditioning), activate molecular self-defense mechanisms at the target organ heart as well as in other parenchymal organs. Remote conditioning of the heart has been established in many experimental studies and successfully translated to patients. Remote ischemic conditioning by short repetitive cycles of ischemia/reperfusion on an extremity reduces infarct size and improves the prognosis of patients with reperfused myocardial infarction. The present review focuses on three levels of remote conditioning and its resulting cardioprotection: I) at the stimulus level, electrical stimulation, chemical/pharmacological substances, mechanical trauma and cycles of ischemia/reperfusion act at sites remote from the heart, II) at the transfer level, neuronal and humoral mediators transfer the protective signal from the periphery to the heart, and III) at the target level, receptor activation and intracellular signal transduction ultimately affect protection of the myocardium and other organs, as established in different animal models and humans/patients. Remote conditioning is obviously a systemic response. Further mechanistic understanding is mandatory to translate the protection by remote conditioning more successfully to patients with cardiovascular disease.

Keywords Cardioprotection · Infarct size · Myocardial ischaemia · Remote conditioning · Signal transduction

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Historical and conceptual background

In acute myocardial infarction [131], sustained coronary occlusion results most often from the rupture of an atherosclerotic plaque in an epicardial coronary artery and superimposed thrombosis (type I myocardial infarction) [157]. The only way to rescue the ischemic myocardium from impending infarction is timely reperfusion. However, reperfusion is a double-edged sword and does not only terminate ischemic injury, but also inflicts additional reperfusion injury [60, 75, 117, 168]. The final infarct size is therefore the aggregate of both ischemic and reperfusion injury. The final infarct size determines left ventricular function, remodeling [66] and the long-term outcome of patients surviving an acute myocardial infarction [102, 152].

Somewhat paradoxically, brief episodes of myocardial ischemia and reperfusion, which do not cause irreversible injury per se, can even protect from the consequences of prolonged myocardial ischemia and reperfusion by activation of molecular self-defense mechanisms [56]. Such conditioning can either precede (ischemic preconditioning) or follow (ischemic postconditioning) the infarct-causing index ischemia.

Interestingly, protection of the myocardium by conditioning can not only be induced locally by brief episodes of occlusion/ reperfusion in the target coronary artery but also from a distance (remote ischemic conditioning) [67]. In the original study in anesthetized dogs, infarct size in the left anterior descending coronary artery-perfused myocardium was reduced by preceding brief episodes of occlusion/ reperfusion in the left circumflex coronary artery [123]. Initially, such protection by conditioning between two adjacent coronary vascular territories was considered as a laboratory curiosity. However, in subsequent studies such remote ischemic conditioning was not only confirmed, but also elicited from a larger distance and from different tissues and organs. Over the years, it became evident that remote

conditioning is a systemic response which can be elicited from different tissues and organs, by a variety of stimuli (apart from ischemia/reperfusion) and protect different parenchymal organs from ischemia/reperfusion injury [67].

With respect to the time frame, remote conditioning is classified as pre- (before the target organ ischemia), per- (during the target organ ischemia) or post- (after the target organ ischemia) conditioning. When the time lag between the remote conditioning stimulus and the target ischemia is of longer duration than minutes, there is delayed remote ischemic conditioning [6]. Some studies suggest a dose-response relationship between stimulus strength and protection. In mice, number and duration of remote ischemia/reperfusion cycles in the hind limbs rather than ischemic tissue mass determined the magnitude of protection, as reflected by the reduction of infarct size: 4 to 6 cycles of 2–5 min ischemia / 2–5 min reperfusion on one or two hind limbs reduced infarct size, whereas with 6 cycles of 10 min ischemia and 5 min reperfusion, respectively, cardioprotection was not observed [78]. This finding is in line with the notion of “hyperconditioning”, i.e. an excessive conditioning protocol may cause additional damage to the target organ [164].

The signal transduction of remote conditioning is largely unclear in its details. Whereas the intramyocardial signal transduction of remote conditioning appears to be very similar to that of local conditioning, in particular the transfer signal from the remote tissue or organ to the heart or other target organs is still enigmatic [57]. Both neuronal and humoral mechanisms appear to be causally involved [56].

Remote ischemic conditioning is also operative in humans [61]. In the original proof-of-concept studies, transient ischemia/reperfusion by inflation and deflation of a blood pressure cuff on the arm protected from ischemia/reperfusion-induced endothelial dysfunction on the contralateral arm [86, 103]. In a further translational study, the myocardial injury by ischemic cardioplegic arrest during elective coronary artery bypass graft (CABG) surgery was taken as a surrogate of myocardial infarction, and the troponin release following CABG was reduced by remote ischemic preconditioning with 3 cycles of 5 min forearm ischemia and 5 min reperfusion [47]. Subsequently, infarct size (single photon emission computed tomography) was indeed reduced by remote ischemic preconditioning with brief cycles of forearm ischemia/reperfusion during the transport of patients with acute myocardial infarction in the ambulance to the hospital and before interventional reperfusion [9].

Signal transduction of remote conditioning

The **remote stimulus** acts, per definition, at a distance from the target organ. A variety of remote stimuli elicit target organ protection: a) electrical stimuli such as direct or percutaneous electrical

nerve stimulation [110, 129] or electroacupuncture [37, 130, 159], b) chemical/pharmacological stimuli such as capsaicin [46, 79, 129] or autacoids (adenosine [99, 151], bradykinin [46, 136]), c) mechanical stimuli such as trauma i.e. by surgical skin incision [45, 46, 79], or d) cycles of ischemia/reperfusion of an extremity or organ [25, 68, 71, 126, 128, 142, 145, 151] which again comprise chemical and mechanical stimuli. Common to all these stimuli is an activation of peripheral sensory nerves. For the ischemia/reperfusion stimulus, it is not entirely clear to which extent the peripheral sensory nerve activation or additional mechanisms such hypoxia/acidosis or vascular shear stress contribute to remote conditioning [67, 88]. Currently it is impossible to compare the different stimuli and to estimate the most effective remote stimulus, since for none of the stimuli a stimulus strength-response relationship has been established. All such comparisons done so far [79, 130, 151] are therefore of limited value.

Neuronal and humoral components are involved in the **signal transfer** from the peripheral stimulus site to the target organ; they act in concert and interact on three different levels. *Stimulus level:* peripheral sensory nerves are directly activated and/or humoral factors are released which subsequently activate the peripheral sensory nerves [43]. *Systemic level:* the peripheral sensory afferent nerves travel to the spinal cord, project into autonomic centers of the central nervous system to result in activation of efferent vagal nerves [43]. Vagal nerve activation a) releases acetylcholine to activate muscarinic receptors in the target organs, and/or b) activates muscarinic receptors in non-target organs which release humoral factors which subsequently act on the target organs [7]. *Target level:* apart from and in addition to a) neuronal activation of receptors in the target organ, b) released humoral factors can activate the intrinsic nervous system of the target organ (e.g. intracardiac ganglia) [7, 121] or c) mediate a receptor-dependent or -independent response of the specific parenchymal cells of the target organ. Neither the quantitative amount nor the exact nature of neuronal and humoral mediators in remote ischemic conditioning are currently clear [43, 56, 57, 67].

The **response** to remote ischemic conditioning is obviously systemic because it occurs in all organs tested so far. Cell injury i.e. the consequence of ischemia/reperfusion is reduced by remote ischemic conditioning in the brain, the heart, the intestine, the kidneys, the liver, the lungs, the ovaries, the pancreas, the skeletal muscle, and the skin [15]. In different cell types, common intracellular signal transduction pathways are involved. However, a direct comparison of intracellular signaling between different cell types has not been performed so far, and signaling may to a certain extent be organ-specific. The intracellular signaling in cardiomyocytes is probably species-specific [56, 67]. Notwithstanding the specific signal transduction pathways, mitochondria are an important end-effector of protection [56, 64]. Ischemia/reperfusion injury causes, independently of the cell type and organ-specific susceptibility to this injury, cell death by different processes (necrosis, necroptosis, apoptosis and/or autophagy) [75, 85], and

preserved mitochondrial function protects from such cell death.

In the following, we discuss the current knowledge about underlying mechanisms of remote conditioning, its stimuli, the signal transfer from the periphery to the target organ and the respective cellular response within the target organ. (Fig. 1) Most studies on cellular response/intracellular signal transduction by remote conditioning were performed in the myocardium. Therefore, we specifically focus on the target organ heart.

Remote stimulus

The different remote stimuli - electrical, chemical/pharmacological, mechanical and also cycles of ischemia/reperfusion - all act as noxious triggers which activate peripheral sensory nerves. A causal involvement of neuronal activation is evidenced by the response to stimuli which exclusively activate peripheral sensory nerves, e.g. electrical nerve stimulation or chemical/pharmacological nociceptor stimuli such as capsaicin [46, 79, 129]. The resulting cardioprotection is abolished by peripheral nerve transection or by local anesthesia [79, 129]. However, even in response to peripheral nerve stimulation humoral factor(s) are causally involved in signal transfer. Whether or not the humoral factor(s) are locally released as a direct response to the

stimulus or are released from other neuronally activated organs is currently unclear [7, 57, 107, 120, 121]. So far, all studies investigating humoral signals in response to different stimuli used blood collected from the systemic circulation and its derivatives, such as plasma or plasma dialysate, respectively. Local release of humoral factor(s) was suggested in a study in mice where occlusion of the femoral vein draining the leg undergoing remote ischemic conditioning by 3 cycles of femoral artery occlusion and reperfusion abrogated infarct size reduction [100].

Electrical nerve stimulation (4 cycles of 5 min femoral nerve stimulation and 5 min rest) in rabbits induced the release of humoral factor(s) into the systemic circulation which reduced infarct size after transfer to an isolated acceptor rabbit heart [129]. Electroacupuncture at the forelimb reduced myocardial infarct size in rodents [37, 159] and induced the release of humoral factor(s) into the systemic circulation of rabbits, which then reduced infarct size after transfer to an isolated acceptor rabbit heart [130]. Cardioprotective humoral factor(s) were also released into the systemic circulation after cycles of transcutaneous electrical nerve stimulation of rabbit and human limbs [110]. Femoral nerve transection prevented the release of cardioprotective humoral factors after transcutaneous electrical nerve stimulation [129], but did not

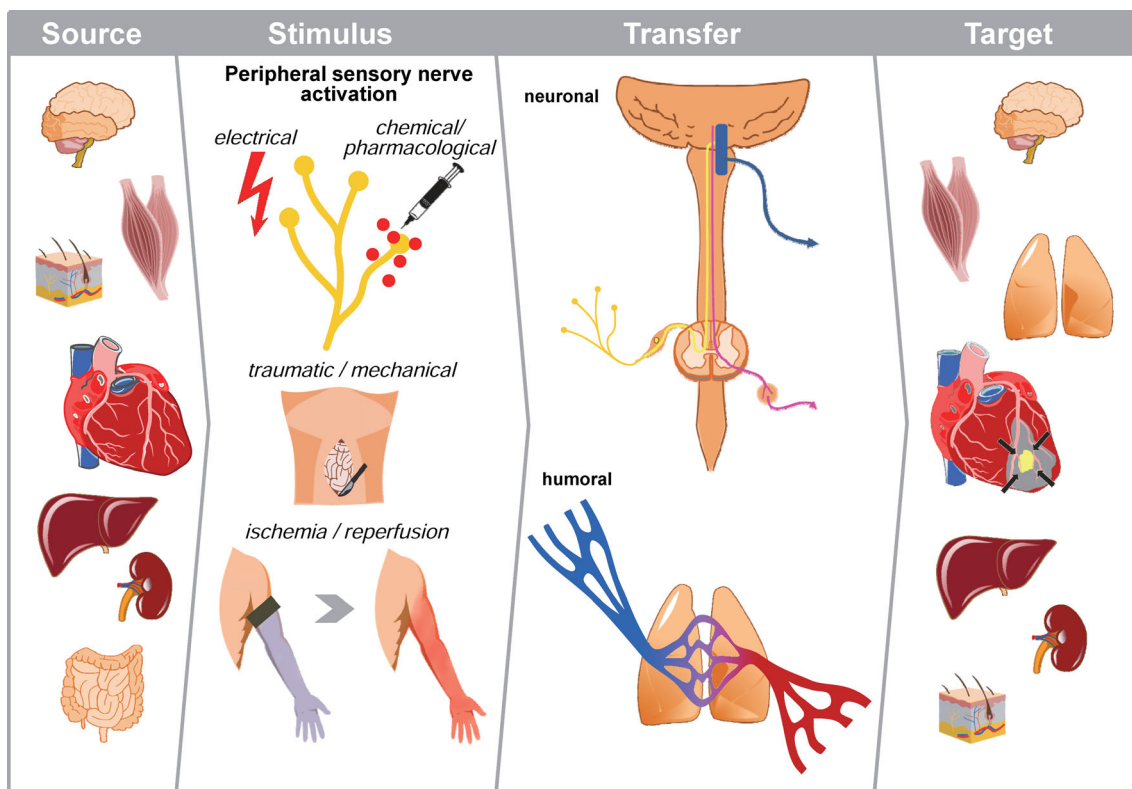


Fig. 1 Remote conditioning - neuronal and humoral signal transfer from the source organ where the stimulus acts to the target organ heart

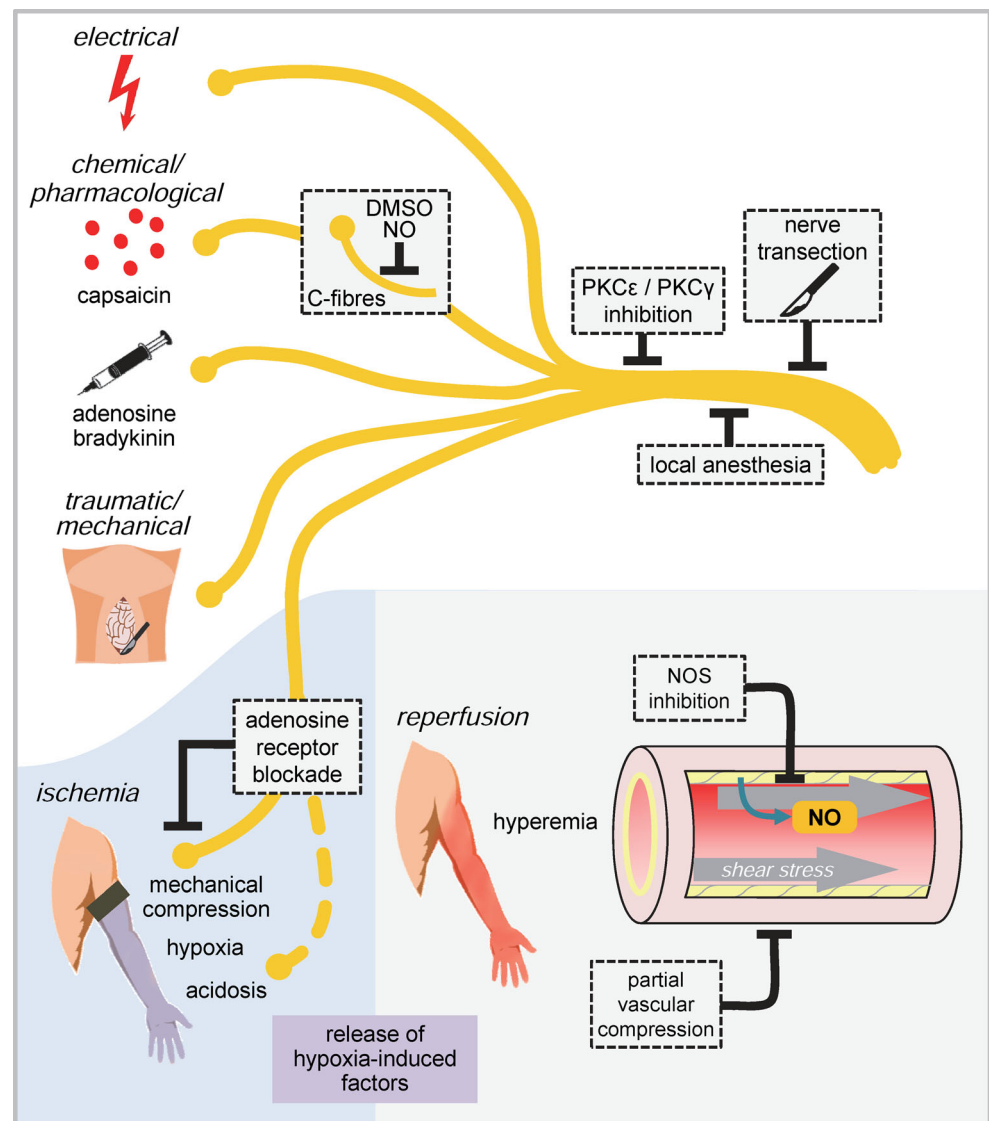
prevent the release of nitric oxide and subsequent infarct size reduction by remote ischemic conditioning [128], suggesting an additive/independent action of shear stress as stimulus. (Fig. 2)

Local chemical/pharmacological stimuli Topical capsaicin selectively activates non-myelinated afferent C-fibers, and capsaicin when applied to the abdominal skin elicited infarct size reduction in rodents [46, 79]. Local anesthesia with lidocaine [79] or blockade of non-myelinated C-fiber activation by dimethyl sulfoxide (DMSO) abrogated the protection by application of capsaicin to the abdominal skin in rabbits [129]. C-fiber activation is nitric oxide-sensitive, and local application of a nitric oxide donor prior to nerve stimulation prevented the release of transferable cardioprotective humoral factor(s) [110]. Noxious mechanical stimuli also activate C-fiber nociceptors [33, 80]. Intra-arterial infusion of the autacoids adenosine [99] or bradykinin [136], in doses activating local sensory neurons

without a systemic effect or direct activation of myocardial receptors, reduced myocardial infarct size in rodents. The infarct size reduction by adenosine or bradykinin can be elicited from different organs. Mesenteric artery infusion of adenosine [99] or bradykinin [136] in rats, femoral artery infusion of adenosine in rabbits [151] and in patients undergoing elective coronary angiography [24], but also application of bradykinin to the abdominal skin of rats [46] reduced infarct size [126]. In rats, in response to bradykinin application to the abdominal skin, two protein kinase C (PKC) isozymes (PKC ϵ and PKC γ) were involved in the local signal, and their inhibition abrogated the cardioprotection by bradykinin [46] (Fig. 2).

Trauma by transverse abdominal skin incision, i.e. partly a *mechanical stimulus*, reduced myocardial infarct size in rodents [46, 79] and dogs [45]. Such abdominal skin incision certainly activates sensory nerves. However, it is unclear to which extent other stimuli (interruption of blood vessels, edema and tissue compression) are also involved. This

Fig. 2 Stimuli of remote conditioning: peripheral sensory nerve activation / vascular shear stress. DMSO dimethyl sulfoxide; NO nitric oxide, NOS nitric oxide synthase, PKC protein kinase C



nociceptive, neuronal activation by skin incision involved also - as did topical bradykinin – a local activation of PKC ϵ and PKC [46]. Thus, there may be an interplay between bradykinin signaling and nerve activation. Indeed, protection in response to skin incision was prevented by blockade of bradykinin 2 receptors in mice and dogs [45, 79]. Since in these studies bradykinin 2 receptors were blocked systemically, it is unclear whether bradykinin is released and acts locally in response to the nociceptive stimulus or more downstream and systemically (Fig. 2).

Ischemia/reperfusion of an extremity or parenchymal organ reduces myocardial infarct size and induces the release of humoral factor(s) into the systemic circulation in rodents, pigs and humans, which elicit cardioprotection after transfer to an isolated acceptor heart [71, 142, 145]. Ischemia/reperfusion involves mechanical stimuli through vascular occlusion and tissue compression as well as chemical factors (adenosine and bradykinin) [88, 126]. Femoral nerve transection abrogated the infarct size reduction by 3 cycles of 5 min ischemia and 5 min reperfusion of one hind limb in rabbits [30]. Sensory nerve activation by remote ischemic conditioning with cycles of blood pressure cuff inflation/deflation of the arm was suggested by the lack of release of transferable humoral cardioprotective factors in humans with diabetic neuropathy as compared to diabetics without neuropathy and healthy controls [77]. However, it is not clear whether the stimulus or the transfer signal is absent in diabetic neuropathy.

Local adenosine receptor blockade by intra-arterial caffeine infusion abrogated the release of humoral factor(s) in response to remote ischemic conditioning in healthy volunteers by 3 cycles of 5 min forearm ischemia and 5 min reperfusion [24], indicating that adenosine mediates a local neuronal activation in response to ischemia/reperfusion.

The non-selective, acid-sensitive, cation channel transient receptor potential vanilloid 1 (TRPV1) is widely expressed on primary sensory nerves including sensory C-fibers [124]. Physical and/or chemical stimuli activate TRPV1 and subsequently trigger the release of neuropeptides such as calcitonin gene-related peptide (CGRP) and substance P from peripheral nerve terminals [112]. In rats, the infarct size reduction by remote ischemic conditioning with 3 cycles of 5 min ischemia and 5 min reperfusion on one hind limb was associated with a TRPV1-mediated CGRP and substance P release [38]. The TRPV1 antagonist capsaizine abolished the release of CGRP and substance P and the observed infarct size reduction [38]. However, capsaizine was applied systemically [38], and thus the participation of TRPV1 activation at other parts of the organism cannot be excluded. In rats, an intrathecal and intravenous administration of CGRP receptor antagonists abolished the infarct size reduction by intrathecal morphine administration [101], indicating that there is an interaction of CGRP- with

opioid-mediated cardioprotection. Ischemia/reperfusion also induces tissue hypoxia, acidosis (during ischemia) and reactive hyperemia (during reperfusion), which trigger the release of local and humoral factor(s). Hypoxia of the perivascular parenchymal tissue may stimulate the expression of hypoxia-sensitive factors, which are subsequently released into the circulating blood [68]. The acidosis resulting from ischemia activates acid-sensitive sensory nociceptors of the vasculature [80]. The reactive hyperemia during reperfusion increases vascular shear stress and induces the release of humoral factor(s) from the endothelium such as nitric oxide [128]. A causal role for such nitric oxide release in response to vascular shear stress was evidenced by its prevention when reactive hyperemia was suppressed by controlled partial vascular compression with an ultrasonic probe in healthy volunteers undergoing remote ischemic conditioning by one cycle of 5 min blood pressure cuff inflation / 5 min deflation on the arm [128] (Fig. 2).

Signal transfer

Neuronal signal transfer

Neuronal signal transfer is certainly involved in remote conditioning of the heart [43], but details are not entirely clear.

Hexamethonium inhibits impulse transmission from pre-ganglionic to postganglionic neurons of both the sympathetic and parasympathetic nerve systems. Hexamethonium abrogates cardioprotection by remote conditioning in some but not in all studies. Remote conditioning by surgical trauma in mice [79] and by one cycle of 15 min ischemia and 10 min reperfusion of the mesentery in rats [41] was abrogated by hexamethonium. Ganglionic blockade with trimetaphan abrogated also the protection of endothelium-dependent vasodilation by remote ischemic conditioning with 3 cycles of 5 min forearm ischemia and 5 min reperfusion in healthy volunteers [103]. However, the infarct size reduction by occlusion/reopening of the infrarenal aorta was not attenuated by hexamethonium in rats [163]. (Fig. 3)

In healthy humans with experimental ischemia/reperfusion in the forearm, remote ischemic conditioning of one leg by 2 cycles of 5 min ischemia and 5 min reperfusion attenuated the ischemia-induced sympathetic nerve activation in the muscle of the contralateral leg [94]. Optogenetic cell-specific silencing of neurons in the brainstem dorsal motor nucleus of the vagus nerve [107], but also bilateral cervical or subdiaphragmatic vagotomy abolished the infarct size reduction by remote ischemic conditioning with one cycle of 15 min ischemia and 10 min reperfusion on both hind limbs in rats [7]. Conversely, electrical vagal nerve stimulation mimicked the effect of remote ischemic conditioning in rabbits [30] and pigs [160] and reduced infarct size. The infarct size reduction

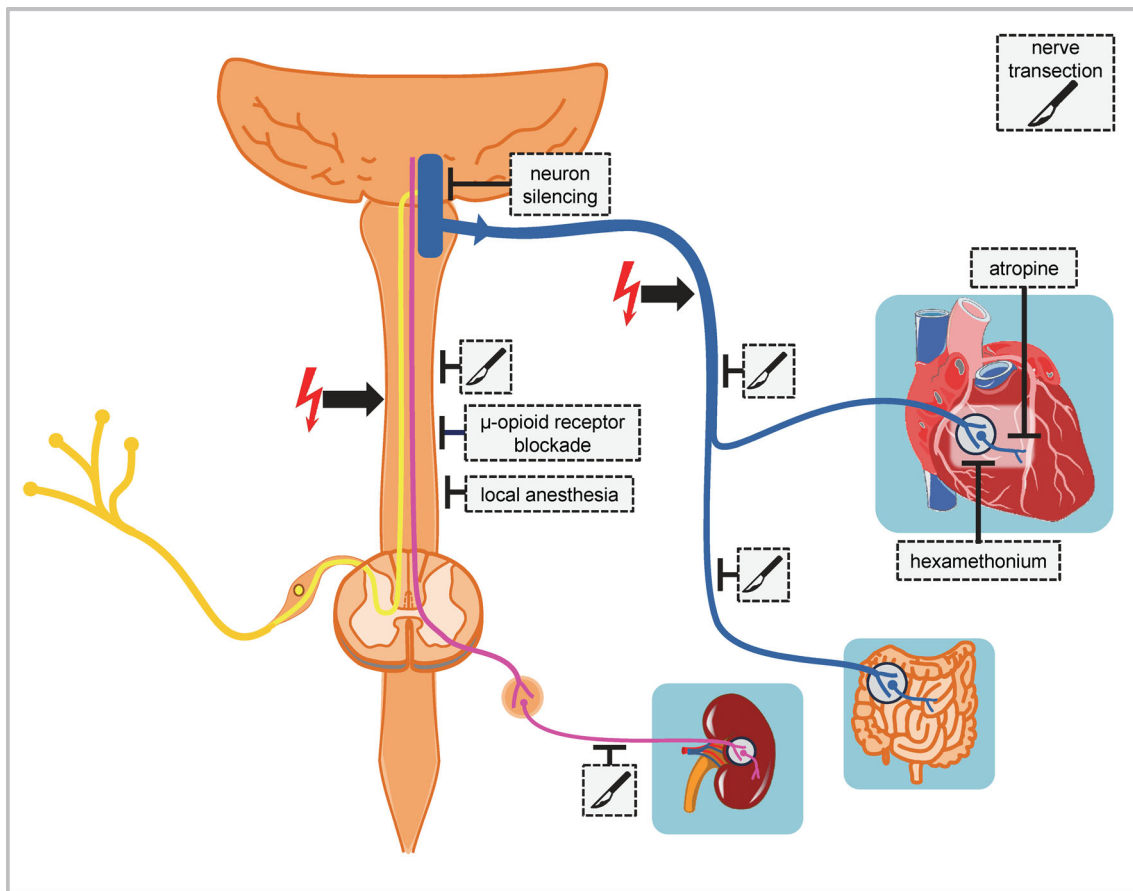


Fig. 3 Neuronal transfer of remote conditioning

by electrical vagal nerve stimulation during myocardial ischemia and reperfusion was sensitive to nitric oxide and abolished by systemic inhibition of nitric oxide synthase [160] (Fig. 3).

The spinal cord also participates in neuronal signal transfer. In rats, blockade of spinal cord nerve transmission by intrathecal lidocaine abrogated the infarct size reduction by intrathecal application of morphine in 3 cycles of 5 min [105]. Also in rabbits, spinal cord section at T9-T10 level abolished the infarct size reduction by remote ischemic conditioning with 3 cycles of 5 min ischemia and 5 min reperfusion on one hind limb [30]. An intrathecal μ -opioid receptor antagonist abrogated the infarct size reduction by skin incision in rodents whereas antagonists of δ - or κ -opioid receptor subtypes did not [109, 165]. Such cardioprotection was associated with an adenosine release into spinal cord tissue of the T1-T5 region, and the μ -opioid receptor antagonist inhibited such adenosine release [109] (Fig. 3).

The involvement of supraspinal central nervous centers is contentious. Spinal cord integrity above C7 was not required for cardioprotection by skin incision in mice, but an intact spinal cord at T7 was required [79]. In rabbits, direct electrical stimulation of the spinal cord at C8-T2 before myocardial ischemia reduced infarct size [149]. However, the above

studies and the involvement of brainstem vagal nuclei and vagal nerve activation support a mandatory role for supraspinal neuronal centers [7, 43, 57, 107].

Thus, given the current data it is unclear a) whether or not the brainstem must be involved, and b) which level(s) of the spinal cord are mandatory for signal transfer. Possibly, the involved neuronal level(s) depend on the remote stimulus and/or on the species under study.

Humoral signal transfer

In response to all remote stimuli, there is humoral signal transfer to the target organ heart. However, it is unclear whether the different stimuli operate via the same or different humoral factor(s) and how these factor(s) possibly interact. The core paradigm for the identification of humoral cardioprotective signals is the transfer of protection with blood, plasma or plasma dialysate from the donor organism which underwent a remote ischemic conditioning protocol to an isolated perfused acceptor heart that serves as a bioassay [28, 56, 71, 142]. In a more reductionist approach, coronary effluent after local ischemic conditioning of a donor heart (rat, rabbit) was used for transfer experiments to an isolated perfused acceptor (rat, rabbit) heart [29, 138]. Such transfer experiments have

then been combined with analytical procedures and pharmacological inhibitor approaches. The analytical procedures comprised non-biased approaches such as gene microarrays and proteomic/metabolomic mass spectroscopy techniques. Other more hypothesis-driven studies have just focused on single humoral factors in isolation.

General properties of the humoral factor(s) Cardioprotection in response to remote ischemic conditioning is transferable with whole blood from a donor to an acceptor rabbit [28], or with diluted plasma or plasma dialysate to an isolated perfused heart of the same species [142] or even across species [56, 145]. Neither the efficiency of cardioprotection by transfer of whole blood versus that of pure plasma nor that of plasma or plasma dialysate at different dilutions have been compared so far. Most studies used a plasma dialysate (< 12–14 kDa) with a final dilution of 1:10–1:50 for transfer experiments. Technical reasons (e.g. extensive foaming of undiluted plasma or adverse side effects by immune responses of the acceptor heart to plasma proteins of another species) are the background for the use of diluted plasma or plasma dialysate for such transfer experiments [71, 142]. However, as a side product of the successful humoral transfer with diluted dialysate, two properties of one or more humoral factor(s) are indirectly identified: a) its/their size is below <12–14 kDa (cut-off), and b) the original factor(s)' plasma concentration is effective in an up to 1:10–1:50 dilution. In plasma of pigs taken after vs. before remote ischemic conditioning by renal artery occlusion for 10 min and 20 min reperfusion no different expression of proteins and peptides >8–10 kDa was identified [95], supporting the notion that proteins and peptides <8–10 kDa are most relevant for the transfer of protection.

The protective, humoral factor(s) were further characterized as hydrophobic, thermolabile and lyophilizable. With a purified hydrophobic and with a lyophilized fraction of coronary effluent of rat and rabbit hearts undergoing local ischemic preconditioning the cardioprotection was transferable to an acceptor rat [138] or rabbit heart [29], respectively. Also, when rabbit plasma dialysate taken after remote ischemic conditioning by 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb was perfused through a hydrophobic column, the infarct size reduction in acceptor rabbit hearts was abrogated [142]. Infarct size reduction was abrogated when coronary effluent was heated at 70 °C [138]. These characteristics (hydrophobic, lyophilizable and thermolabile) suggest that some of the released humoral factor(s) are a peptide or protein.

After a single protocol of 3 cycles of 5 min forearm ischemia and 5 min reperfusion in healthy volunteers, plasma dialysate (<12–14 kDa) of blood taken at 5 min after and in the following 6 days following the protocol still reduced infarct size in isolated mouse hearts, and the cardioprotection disappeared no earlier than after 7 days [71]. Whether there is a

circulating cardioprotective signal over one week following a single remote ischemic conditioning protocol or whether there is a rapid and continuous release of one or more factor(s) with a short half-life remains unclear from this study. A proteomic approach on plasma taken from healthy volunteers 15 min and 24 h after 3 cycles of 5 min forearm ischemia and 5 min reperfusion identified a time-dependent difference in up/down-regulated plasma proteins [54], indicating that different factors are appearing at a different time-scale, but this study neither identified these factors nor did it attribute a causal role to them in mediating cardioprotection.

Non-biased “omic” approaches In response to remote ischemic conditioning in mice and humans by 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb or forearm, respectively, a microRNA (miRNA) array identified an association of increased systemically circulating miRNA144 with remote ischemic conditioning. The causal involvement of miRNA144 in cardioprotection was proven by induction of cardioprotection with exogenous miRNA144 and abrogation of infarct size reduction by remote ischemic conditioning with a specific antisense oligonucleotide [98]. An increase of plasma miRNA144, induced by remote ischemic conditioning with 4 cycles of 5 min forearm ischemia and 5 min reperfusion, was confirmed in patients with suspected ischemic coronary artery disease undergoing diagnostic cardiac positron emission tomography [122]. (Fig. 4)

Several proteomic/metabolomic approaches did not identify a specific humoral protective factor or a “protective cocktail” of factor(s) after remote ischemia/reperfusion cycles in the limbs [70] or kidneys of rats [95], in adult healthy volunteers [52] and in children undergoing heart surgery for repair of Fallot's tetralogy [55]. However, two different proteomic approaches on rat and human plasma after remote ischemic conditioning identified an increase of apolipoprotein A1 [54, 69]. In rats, this apolipoprotein A1 increase in response to one cycle of 10 min ischemia and 5 min reperfusion on one hind limb was associated with infarct size reduction [69]. Apolipoprotein A1 infusion before myocardial ischemia/reperfusion reduced infarct size [69] and shared intracellular signaling pathways with that of remote ischemic conditioning in rats [84]. Using a metabolomic approach in rat and patient plasma after remote ischemic conditioning by 4/3 cycles of 5 min hind limb/forearm ischemia and 5 min reperfusion, respectively, there was a decrease of ornithine and an increase of kynurenine and glycine in both species. In rats, the intraperitoneal infusion of exogenous kynurenine and glycine alone or in a combination 10 min before myocardial ischemia, respectively, reduced infarct size [20]. However, the doses of such exogenous amino acids were up to 10-fold higher than those determined in response to the remote ischemic conditioning stimulus, raising doubts about the causal involvement

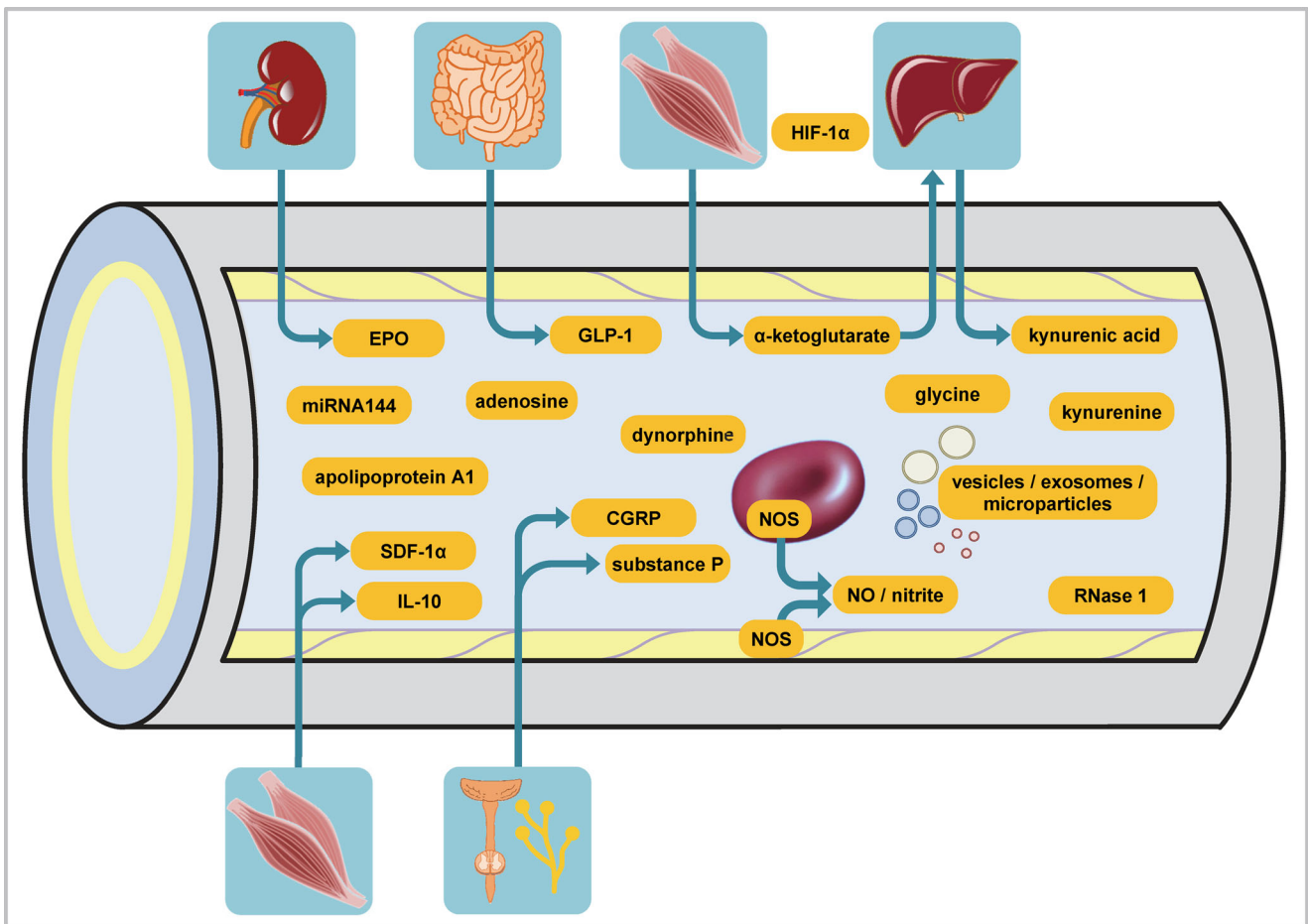


Fig. 4 Humoral transfer of remote conditioning. CGRP calcitonin gene-related peptide; EPO erythropoietin; GLP-1 glucagon-like peptide-1; HIF-1 α hypoxia-inducible factor-1 α ; IL-10 interleukin-10; miRNA

microRNA; NO nitric oxide, NOS nitric oxide synthase, SDF-1 α stromal cell-derived factor-1 α

of kynurenine and glycine in the transfer of cardioprotection by remote ischemic conditioning (Fig. 4).

Hypothesis-driven approaches on individual humoral factors Autacoids: Autacoids are endogenously released during local ischemic pre- and postconditioning, and exogenous application of autacoids induces cardioprotection. Adenosine as well as bradykinin receptor blockade abrogates the infarct size reduction by local conditioning [88]. Whether there is an endogenous release of autacoids into the systemic circulation in response to remote conditioning is not really clear.

Indeed, adenosine was increased in the coronary effluent of rabbits after local ischemic preconditioning, and this effluent reduced infarct size after transfer to another rabbit heart. Infarct size reduction was abrogated with an adenosine receptor blocker [97], suggesting the potential for adenosine to also contribute to the humoral transfer of remote conditioning. In rats in response to remote conditioning by skin incision, the infarct size reduction was associated with an adenosine release into spinal cord tissue [109], but circulating blood levels were not determined. In mice, in response to persistent brain

ischemia through bilateral ligation of the internal carotid arteries there was an increase of arterial plasma, but not cerebral adenosine, and this increase in arterial adenosine was subsequently associated with infarct size reduction of isolated perfused mice hearts [137]. Persistent brain ischemia, however, is not really the same as remote ischemic conditioning which requires brief intermittent episodes of reversible ischemia and reperfusion.

In non-diabetic patients undergoing elective coronary angiography, intra-arterial infusion of exogenous adenosine was associated with the release of transferable cardioprotective factor(s). Transfer of plasma dialysate, taken after the arterial adenosine infusion, reduced infarct size in isolated perfused mouse hearts to an extent which could not be reproduced by the adenosine concentration in the dialysate [24]. Thus, adenosine in fact stimulated the release of cardioprotective factor(s), but adenosine itself was not the cardioprotective factor (Fig. 4).

Currently, there is no evidence that bradykinin is released into the systemic circulation in response to remote conditioning. Furthermore, the intravenous infusion of a bradykinin

B(2) receptor antagonist did not abrogate the attenuation of ischemia/reperfusion-induced endothelial dysfunction in humans by remote ischemic conditioning by 3 cycles of 5 min forearm ischemia and 5 min reperfusion [119], suggesting that bradykinin is not a humoral transfer factor of cardioprotection.

Hormones: The glycoprotein hormone erythropoietin (EPO) (phylogenetically, EPO is related to the family of cytokines) is produced by the kidneys, the liver, the vasculature and the heart [36]. In mice, remote ischemic conditioning by 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb increased serum EPO and reduced infarct size. This increase in serum EPO was associated with increased EPO mRNA expression in the kidneys, and exogenous EPO antibodies before remote ischemic conditioning abolished the infarct size reduction [113]. Interestingly, there was an interaction between neuronal signal transduction and EPO release, since renal denervation abolished both the increase in serum EPO and the infarct size reduction by remote ischemic conditioning [113] (Fig. 4).

Recently, the incretin hormone glucagon-like peptide-1 (GLP-1) was reported as a potential humoral transfer factor. In rats, in response to remote ischemic conditioning by one cycle of ischemia/reperfusion of both hind limbs for 15 min, activation of the vagal nerves appeared to mediate a release of GLP-1 release from the gut. Subdiaphragmatic vagotomy abolished the infarct size reduction, and infarct size reduction was also achieved by a GLP-1 receptor agonist. Systemic muscarinic receptor M3 blockade abolished the infarct size reduction by the GLP-1 agonist [7]. Thus, GLP-1 appeared to be released from visceral organs which are innervated by the posterior gastric branch of the vagus nerve, and it involves the activation of muscarinic M3 receptors in the heart. However, the reported increase of circulating GLP-1 in response to remote ischemic conditioning was minimal and could not explain the observed protection [57]. Also, there was no distinction between the inactive full length GLP-1 and the active peptide of GLP-1 [72] (Fig. 4).

Cytokines/Chemokines: Local ischemic conditioning is associated with the inhibition of inflammatory responses [56]. In mice, 24 h after remote ischemic conditioning by 3 cycles of 5 min ischemia and 5 min reperfusion on one hind limb, the plasma concentration of the anti-inflammatory interleukin-10 (IL-10) was increased. The infarct size reduction by remote ischemic conditioning was abrogated, when the activation of IL-10 receptors was prevented by anti-IL-10 receptor antibodies or by a genetic knock out of IL-10 [13] (Fig. 4).

The chemokine stromal cell-derived factor-1 α (SDF-1 α) is abundant in ischemic tissue with a vascular and perivascular distribution after dermal ischemia in athymic nude mice [19]. In rats, SDF-1 α was released into the plasma in response to remote ischemic conditioning by 3 cycles of 5 min ischemia and 5 min reperfusion on one hind limb. Inhibition of the

SDF-1 α chemokine 4 receptor attenuated the infarct size reduction by remote ischemic conditioning, but did not completely abrogate it [25] (Fig. 4).

Neuro-peptides: Opioid peptides are widely distributed in the central nervous system but they are also - eventually as a spillover from neuronal release- released into the blood in response to different remote stimuli [125]. A quantitative release of dynorphin, but not of met-enkephalin, into the plasma was reported in rats in response to remote ischemic conditioning by 3 cycles of 5 min ischemia and 5 min of reperfusion of one hind limb [172]. In healthy volunteers, however, remote ischemic conditioning by 4 cycles of 5 min forearm ischemia and 5 min reperfusion did not induce an increase of pro-opiomelanocortin derivatives [8]. Non-specific opioid receptor blockade with naloxone abrogated the infarct size reduction by remote ischemic conditioning in mice and rats [51]. Also in patients undergoing primary percutaneous coronary intervention, the opioid receptor blocker naloxone abrogated the reduction of troponin release by remote ischemic conditioning with 3 cycles of 4 min forearm ischemia and 4 min reperfusion [132], emphasizing the relevance of opioids for cardioprotection by remote ischemic conditioning. (Fig. 4) However, whether the cardioprotection is mediated/transferred by circulating opioids or whether they are just a spillover of local neuronal release and the cardioprotection is actually mediated by local neuronal release in the myocardium is currently unclear.

In rats, the release of the neuropeptides CGRP and substance P into the plasma was triggered by the activation of TRPV1 in response to 3 cycles of 5 min ischemia and 5 min reperfusion of one hind limb. Both peptides were causally involved in the infarct size reduction by remote ischemic conditioning, and CGRP and substance P receptor antagonists abolished the infarct size reduction [38] (Fig. 4).

Amino acids: Several studies suggested that one or more amino acids may act as humoral transfer factors of protection. The enzyme hypoxia-inducible factor prolyl hydroxylase 2 or prolyl hydroxylase domain-containing protein 2 is encoded by the EglN1 gene and therefore termed EglN1. EglN1 is ubiquitously present in nearly all cell types and requires oxygen for its enzymatic activity [81]. In skeletal muscle, EglN1 converts α -ketoglutarate to succinate, and its activity is reduced with tissue hypoxia. Accordingly, the α -ketoglutarate concentration in the systemic circulation is increased with lack of EglN1 activity. In the liver, α -ketoglutarate is metabolized to kynurenic acid. In mice, the genetic deletion or pharmacological inhibition of EglN1, either systemically or in skeletal muscles, reduced infarct size. The humoral transfer of protection was determined in parabiosis experiments ($n = 2$) between donor mice with a skeletal muscle-specific EglN1 deletion and wild type recipient mice undergoing coronary occlusion/reperfusion. Infarct size was reduced in the recipient mice. Exogenous kynurenic acid also reduced infarct size [116]. In

response to EglN1 deletion, the hypoxia-inducible factor-1 α (HIF-1 α) protein expression was increased [116], further relating these data to hypoxia-sensitive pathways. However, this particular study did not actually perform a single remote ischemic conditioning protocol and the conclusions of this study on mechanisms of remote ischemic conditioning are therefore largely speculative. As mentioned above, increased kynurenine was also determined in a metabolomic approach in rat and patient plasma after remote ischemic conditioning by 4/3 cycles of 5 min hind limb/forearm ischemia and 5 min reperfusion, respectively [20], and exogenous kynurenine and glycine also reduced infarct size in rats [20]. However, a causal relation between EglN1, kynurenic acid and remote ischemic conditioning remains to be established (Fig. 4).

Nitric oxide/nitrite: The role of nitric oxide as a humoral transfer signal of remote ischemic conditioning remains controversial. The oxidative metabolite of nitric oxide -nitrite- was increased in plasma in response to remote ischemic conditioning in rodents and humans. The nitric oxide release was prevented by genetic ablation of endothelial nitric oxide synthase but not by femoral nerve transection in mice [128]. Also in rats, systemic nitric oxide synthase inhibition abolished the infarct size reduction by 3 cycles of 5 min ischemia and 5 min reperfusion on one hind limb or one cycle of 15 min ischemia and 10 min reperfusion on both hind limbs, respectively [31, 139], whereas in rabbits, such systemic pre-treatment with a nitric oxide synthase inhibitor did not attenuate the protection by remote ischemic conditioning with 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb [151]. Two studies in humans used not nitrite but the sum of plasma nitrite and nitrate (NO_x) to quantify the nitric oxide release in response to cycles of 5 min ischemia and 5 min reperfusion on one upper leg. Whereas in patients undergoing elective percutaneous coronary intervention NO_x was released into coronary arterial blood after 3 cycles of 5 min forearm ischemia and 5 min reperfusion [3], there was no such release of NO_x into the venous blood of healthy volunteers after 2 cycles of 5 min forearm ischemia and 5 min reperfusion [94]. Nitrite is more stable than the radical nitric oxide itself and enables therefore the transport of eNOS-derived nitric oxide with the circulating blood from the periphery to the heart [127] (Fig. 4).

Ribonucleic acid (RNA) degrading enzyme: With tissue injury, RNA is released into the extracellular space and it amplifies and propagates tissue injury. Circulating RNase1 degrades extracellular RNA and limits tissue damage [171]. Remote ischemic conditioning by 4 cycles of 5 min forearm ischemia and 5 min reperfusion increased endothelial RNase1 in the circulating blood in a smaller cohort of patients undergoing heart surgery [12]. However, troponin release was not

reduced by the remote ischemic conditioning protocol in this particular patient cohort, and therefore a causal involvement of RNase1 in cardioprotection is still elusive (Fig. 4).

HIF-1 α is potentially one *common regulator* of different humoral factors [68]. HIF-1 α is a major regulator of hormone and cytokine signaling in response to hypoxia [106], and in HIF-1 α heterozygous knockout mice the infarct size reduction by remote ischemic conditioning with 3 cycles of 5 min ischemia and 5 min reperfusion on one hind limb was abrogated [14]. EPO synthesis is initiated through HIF-1 α [36]. However, currently it is unclear whether the EPO release in response to remote ischemic conditioning is also HIF-1 α -dependent. The IL-10 release into the plasma in response to remote ischemic conditioning by 3 cycles of 5 min ischemia and 5 min reperfusion on one hind limb was abolished in mice with heterozygous deletion of HIF-1 α [13]. Upregulation of HIF-1 α expression in the hind limb after the remote ischemic conditioning protocol by 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb was confirmed in another study in mice, but this HIF-1 α expression was not a prerequisite for infarct size reduction [83]. This study therefore does not establish a causal role for HIF-1 α in the peripheral release of humoral factor(s) or its participation in humoral transfer. As mentioned above, hypoxia-induced EglN1 inhibition [116] may be one upstream regulator of HIF-1 α .

Particulate carriers of humoral signals Blood cells: Apart from and in addition to endothelial nitric oxide synthase, the nitric oxide synthase of red blood cells was also activated by remote ischemic conditioning with 4 cycles of 5 min forearm ischemia and 5 min reperfusion in healthy volunteers [44]. Thus, the nitric oxide synthase of red blood cells may therefore act as an additional, circulating source of circulating nitrite (Fig. 4).

In patients undergoing elective CABG surgery, remote ischemic conditioning by 3 cycles of 5 min forearm ischemia and 5 min reperfusion down-regulated the expression of bradykinin receptors 1 and 2 expression on leukocytes [135]. The expression of adhesion molecules on neutrophils was bradykinin receptor-dependent [140]. However, whether such differences in bradykinin receptor-dependent adhesion molecule expression result in decreased/increased leukocyte recruitment after myocardial infarction has not been examined so far.

Vesicles/particles: Extracellular vesicles/microparticles and exosomes are circulating extracellular lipid bilayer vesicles with a particle size of 50–1000 / 30–100 nm in diameter, respectively, and considered to be a carrier of information for intercellular communication. Extracellular vesicles/exosomes originate from different cell types and contain proteins, mRNAs, miRNAs, and DNAs [5, 74, 167]. Whether or not extracellular vesicles/exosomes and which particular type of them may contribute to myocardial protection in response to remote ischemic conditioning is currently not clear [26].

The total amount of extracellular vesicles was increased after local ischemic conditioning in the coronary effluent of isolated rat hearts, and administration of the coronary effluent reduced infarct size of isolated recipient hearts, whereas extracellular vesicles-depleted perfusate failed to reduce infarct size [42]. In contrast, after remote ischemic conditioning in mice and humans by 4 cycles of 5 min hind limb / forearm ischemia and 5 min reperfusion, respectively, the total amount of extracellular vesicles was not increased [98]. Extracellular vesicles originating from endothelial cells were increased in circulating blood after remote ischemic conditioning by one cycle of 10 min ischemia and 10 min reperfusion on one hind limb in rats and by 3 cycles of 5 min forearm ischemia and 5 min reperfusion in humans, respectively. However, these vesicles did not reduce infarct size when intravenously transferred to a donor rat [76] (Fig. 4).

Exosomes were increased in response to remote ischemic conditioning by 3 cycles of 5 min ischemia and 5 min reperfusion on one hind limb in rats or the forearm in humans, respectively. When such exosomes were transferred intravenously to an acceptor rat or ex-vivo to an isolated rat heart, the exosomes reduced infarct size, no matter whether they were taken before or after remote ischemic conditioning. In isolated cardiomyocytes, these exosomes activated toll-like receptor 4 and various kinases and induced an activation of cardioprotective heat shock protein27 [161].

Which specific vesicular content is responsible for or involved in the observed cardioprotection is not clear. The total DNA content [155] was unchanged, whereas the total mRNA content [154] was increased in extracellular vesicles in the coronary venous blood of pigs after local myocardial ischemic conditioning. In a more specific analysis, extracellular vesicles carried the cardioprotective miRNA144, but the miRNA144 content was unchanged after remote ischemic conditioning by 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb/forearm in mice and humans, respectively. The content of a miRNA144 precursor, however, was almost fourfold increased in these extracellular vesicles [98].

Differences of the particle type (extracellular vesicles/microparticles versus exosomes) and/or their cellular origin complicate the interpretation of the available data [5, 167].

Neuro-humoral interaction

There is a complex neuro-humoral interaction in remote ischemic conditioning's signal transduction in the periphery, systemically and in the target organ. Activated afferent but also efferent nerves induce the release of humoral factor(s) and, vice versa, humoral factors activate afferent and efferent nerves. Within the myocardium, the intrinsic nervous system is activated by humoral factor(s) and probably also by locally released factors.

At the stimulus level, peripheral adenosine receptor activation after intra-arterial adenosine infusion in human volunteers was mandatory to induce the release of transferable cardioprotective factor(s) [24], suggesting that adenosine may activate peripheral sensory nerve fibers to ultimately cause the release of a humoral factor(s) and reflecting neuro-humoral interaction. (Fig. 5)

At the transfer level, in mice, the infarct size reduction by remote ischemic conditioning with 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb was markedly attenuated by femoral vein occlusion. Also, femoral nerve or sciatic nerve resection only partially abolished the infarct size reduction. Only combined femoral and sciatic nerve resection with femoral vein occlusion completely abolished the cardioprotective effect [100], indicating that there is a neuro-humoral interaction in the afferent signal transfer to the heart.

Efferent renal sympathetic nerve activation in response to remote ischemic conditioning by 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb was mandatory for the release of EPO into the systemic circulation in mice [113]. (Fig. 5) In rats, subdiaphragmatic vagotomy abolished the infarct size reduction by remote ischemic conditioning with one cycle of 15 min ischemia and 10 min reperfusion on both hind limbs and, as mentioned above, GLP-1 was released from visceral organs innervated by the posterior gastric branch of the vagus nerve and was possibly the humoral factor to transfer protection to the heart [7]. Also, supporting the role of vagal gut innervation, in rats, infarct size reduction by remote ischemic conditioning with one cycle of 15 min ischemia and 10 min reperfusion on both hind limbs was not attenuated by section of hepatic, celiac or anterior gastric vagal branches, whereas bilateral cervical vagotomy, subdiaphragmatic vagotomy, gastric vagotomy or sectioning of the posterior gastric branch abolished the infarct size reduction [108]. In other studies in rats only cervical [31, 121] but not subdiaphragmatic vagotomy [31] abrogated the infarct size reduction by remote ischemic conditioning with 3/4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb. However, these latter studies did not investigate a potential impact of vagal innervation on humoral factor release (Fig. 5).

Reflecting the participation of the central nervous system, remote ischemic conditioning by 3 cycles of 5 min ischemia and 5 min reperfusion of one hind limb in rats increased the CGRP and substance P mRNA and protein expression in the dorsal root ganglion, and both peptides were also increased in plasma and myocardial tissue and causally involved in infarct size reduction [38] (Fig. 5).

At the target organ level, humoral factors appear to reduce infarct size through recruitment of intracardiac ganglia. When isolated perfused rat hearts were treated with hexamethonium, the dialysate-mediated infarct size reduction was abrogated [121], and infarct size reduction was also abolished after transfer into an acceptor mouse heart which received an opioid

antagonist [110], suggesting that humoral factors activate intracardiac ganglia. Also, systemic blockade of muscarinic receptor M3 abrogated the infarct size reduction by the GLP-1 analog exendin-4 which mimicked the cardioprotection by remote ischemic conditioning in rats [7], thus reflecting the action of GLP-1 on postganglionic cardiac cholinergic nerves. Indeed, intracardiac ganglia are activated directly by factors released from the myocardium which then, in turn, activate efferent post-ganglionic nerves [144]. Thus, even in isolated, centrally denervated hearts neuronal activation may be involved in cardioprotection [121] (Fig. 5).

Myocardial response

Incoming neuronal and humoral signals activate protective signaling cascades in the myocardium through sarcolemmal receptors or act receptor-independently [88]. The downstream

intracellular signaling cascades then activate their intracellular target structure(s), notably the mitochondria.

Non-biased “omic” approaches on signal transduction Up to now, no genomic/proteomic approach has identified an intracellular signal of particular relevance and with causal involvement in cardioprotection. Genomic profiling of myocardium taken 15 min after remote ischemic conditioning in mice identified an upregulation of genes against oxidative stress (Hadhsc, Prdx4, Fabp4) and of genes involved in cytoprotection (Hsp73). Pro-inflammatory genes (Egr-1, Dusp1, Dusp6) were downregulated [90]. Proteomic profiling for phosphorylated proteins found an increase of several phosphoproteins related to the sarcomeric Z-disk in myocardium of mice in response to remote ischemic conditioning by 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb [1]. Proteomic profiling of right atrial tissue of cardio-surgical

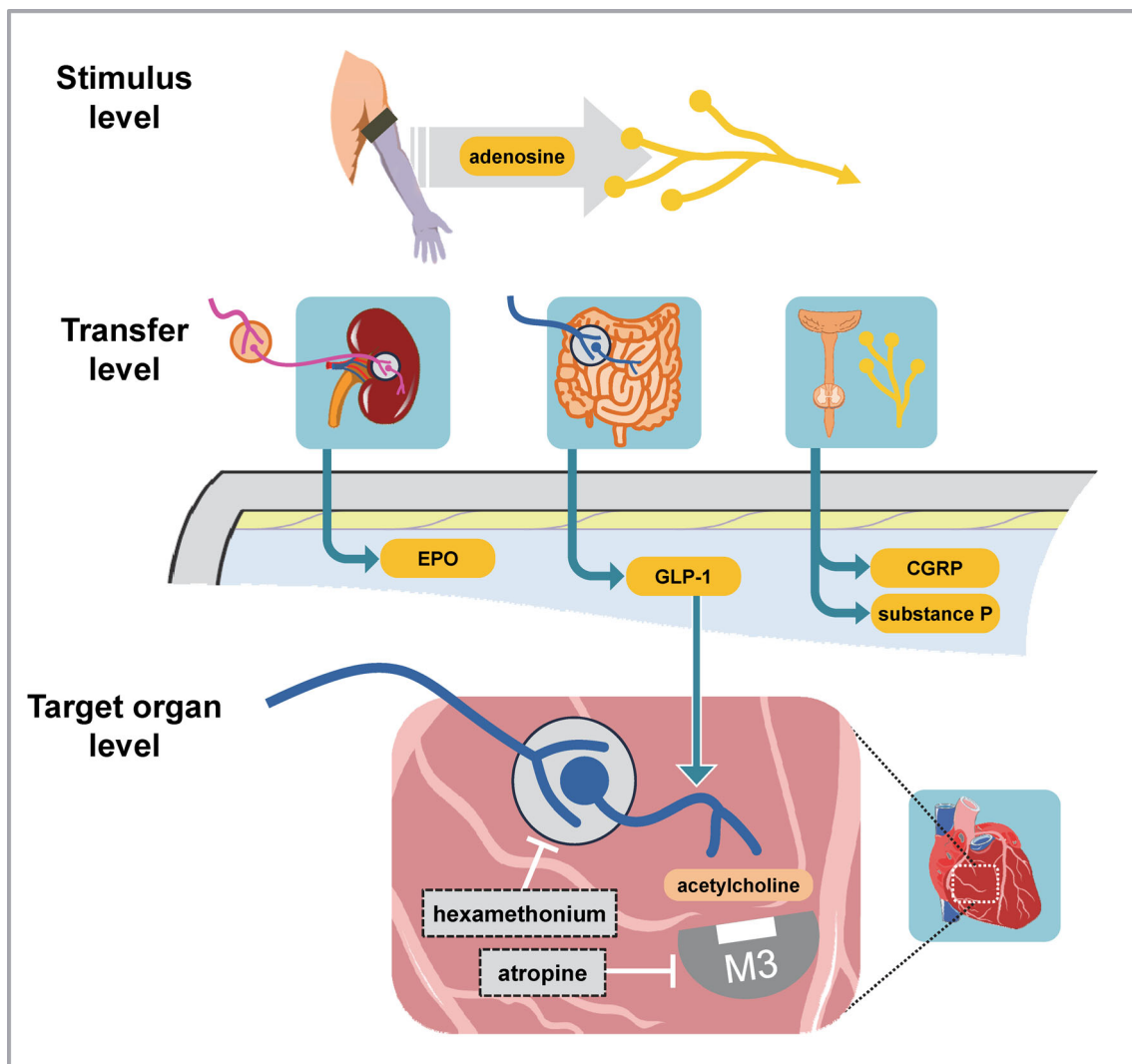


Fig. 5 Neuro-humoral interaction in response to remote conditioning. CGRP calcitonin gene-related peptide; EPO erythropoietin; GLP-1 glucagon-like peptide-1; M3 muscarinic acetylcholine receptor M3

patients, taken after remote ischemic conditioning by 4 cycles of 5 min forearm ischemia and 5 min reperfusion, identified an upregulation of thioredoxin and several cell stress-associated proteins [174], but this particular study did not report a cardioprotective effect. Neither a metabolomic nor a lipidomic approach nor a miRNA array have yet been performed in myocardial tissue after remote conditioning.

Hypothesis-driven approaches on signal transduction The signal transduction of remote ischemic conditioning in the myocardium is remarkably similar to that of local ischemic pre- and postconditioning. Autacoids and neurohormones (acetylcholine, opioids) activate G-protein coupled receptors. Reactive oxygen species such as nitric oxide initiate receptor-independent cardioprotective signaling. The activation of several proteins, notably protein kinases but also others,

converges on the mitochondria and the cytoskeleton. The intracellular signal transduction is conceptually summarized in three major pathways: the endothelial nitric oxide synthase/protein kinase G (eNOS/PKG) pathway, the reperfusion injury salvage kinase (RISK) pathway and the survivor activating factor enhancement (SAFE) pathway [56, 88]. (Fig. 6)

Neurohormone receptors: In response to remote conditioning the following receptors are activated: In a cardiomyocyte cell line, the adenosine receptor A1 was activated with rabbit plasma dialysate after remote ischemic conditioning by 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb [153]. In mice, the bradykinin receptor B2 was activated by traumatic remote conditioning (skin incision) and downstream of bradykinin receptor B2, myocardial PKC ϵ was activated whereas PKC δ was repressed [79]. The infarct size reduction by remote ischemic conditioning with one cycle of 15 min ischemia and 10 min reperfusion on both hind limbs

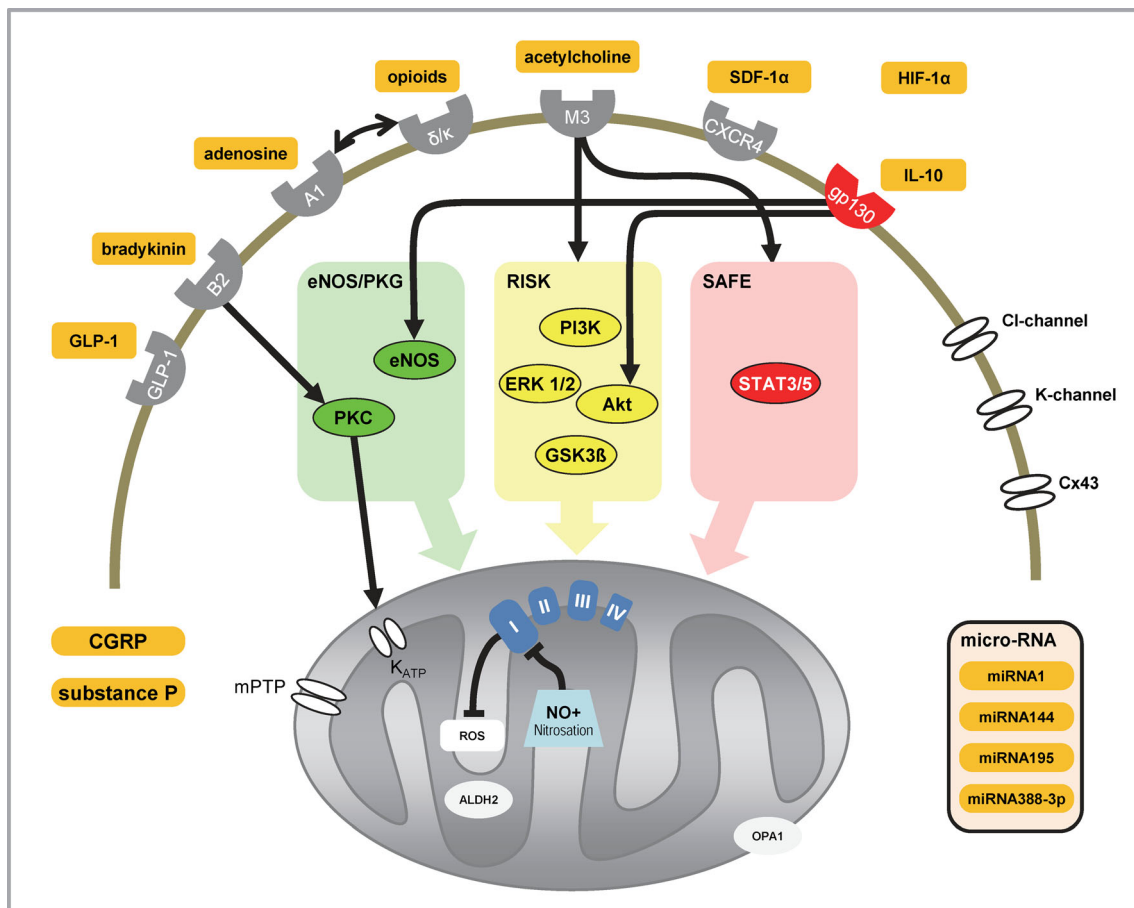


Fig. 6 Myocardial signal transduction - eNOS/PKG pathway in green, RISK pathway in yellow, and SAFE pathway in red. I,II,III, IV mitochondrial respiratory chain complexes I,II,III, IV; δ/κ δ - and κ -opioid receptors; A1 adenosine receptor A1; ALDH2 aldehyde dehydrogenase 2; Akt protein kinase B; B2 bradykinin receptor B2; CGRP calcitonin gene-related peptide; CX43 connexin43; CXCR4 chemokine 4 receptor; eNOS endothelial nitric oxide synthase; ERK 1/2 extracellular-regulated kinases 1/2; GLP-1 glucagon-like peptide-1 and its receptor; gp130 glycoprotein130; GSK3 β glycogen synthase

kinase 3 β ; HIF-1 α hypoxia-inducible factor-1 α ; IL-10 interleukin-10; K_{ATP} ATP-dependent potassium channel; M3 muscarinic receptor M3; miRNA microRNA; mPTP mitochondrial permeability transition pore; NO nitric oxide; OPA1 dynamin-like 120 kDa protein; PI3K phosphatidylinositol-4,5-bisphosphate 3-kinase; PKC protein kinase C; PKG protein kinase G; RISK reperfusion injury salvage kinases; ROS reactive oxygen species; SAFE survivor activating factor enhancement; SDF-1 α stromal cell-derived factor-1 α ; STAT signal transducer and activator of transcription

was GLP-1 receptor- and muscarinic receptor M3 activation-dependent in rats [7]. The precise localization of the involved GLP-1 receptors, however, is currently unknown. The causal involvement of GLP-1 was proven by systemic GLP-1 receptor blockade, but the presence of GLP-1 receptors on cardiomyocytes is still contentious [115]. Muscarinic receptor activation may also trigger an increase in acetylcholine synthesis; a non-selective muscarinic receptor agonist increased the acetylcholine synthesis in cardiomyocytes in vitro [82]. Cardiomyocytes express choline acetyltransferase, and intramyocardial acetylcholine was increased by remote ischemic conditioning with 3 cycles of 3 min ischemia and 3 min reperfusion on one hind limb 24 h after myocardial infarction in mice [114]. However, in this study cardioprotection in response to remote ischemic conditioning was not examined.

Opioid receptors were activated in isolated rabbit cardiomyocytes with rabbit plasma dialysate taken after remote ischemic conditioning by 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb; whereas pretreatment of cardiomyocytes with rabbit plasma dialysate after remote ischemic conditioning reduced the percentage of dead cardiomyocytes after simulated ischemia/reperfusion, unspecific blockade of opioid receptors with naloxone completely abolished this beneficial effect [142]. There is also an interaction of adenosine and opioid receptors. In a cardiomyocyte cell line, pharmacologically induced activation of δ - and κ -opioid receptors mimicked the protective effect on cellular viability induced by plasma dialysate after remote ischemic conditioning, and blockade of adenosine A1 receptors attenuated this cardioprotection [153].

EPO release is increased with remote ischemic conditioning [113]. EPO receptors are expressed in different tissues and cell types, including neurons and cardiomyocytes [133], and selective EPO receptor agonists, which did not stimulate erythropoiesis, reduced infarct size in rats [87]. However, the activation of EPO receptors in the cardioprotection by remote ischemic conditioning has not been directly proven so far.

Receptor(s) for CGRP and substance P as well as their downstream signaling which mediates infarct size reduction [124] have not been identified in the myocardium so far. Apart from the increase of plasmatic levels of both peptides, there was also an increase of both peptides in myocardial tissue in response to remote ischemic conditioning by 3 cycles of 5 min ischemia and 5 min reperfusion on both hind limbs in rats [38].

Cytokine/chemokine receptors: In rats, the chemokine receptor type 4 was activated by SDF-1 α in response to remote ischemic conditioning by 3 cycles of 5 min ischemia and 5 min reperfusion on one hind limb [25]. In mice, anti-IL-10 receptor antibodies attenuated the late infarct size reduction by remote ischemic conditioning with 3 cycles of 5 min ischemia and 5 min reperfusion on one hind limb [13]; however, IL-10 receptors are ubiquitously expressed and their specific participation in the myocardium remains to be identified.

Cytosolic proteins: Different intracellular proteins are activated in response to remote conditioning. In mice, the eNOS/PKG was activated in response to remote ischemic conditioning by 3 cycles of 5 min ischemia and 5 min reperfusion on one hind limb; the key enzyme eNOS was phosphorylated 24 h after myocardial ischemia and reperfusion [13]. PKC was activated in rodents in response to abdominal surgical incision [46, 79]. The RISK pathway was activated regardless of what the specific remote stimulus was, in rodents [7, 11, 13, 46, 79, 145, 158, 166], and pigs [48] with the following key enzymes: protein kinase B/Akt [11, 13, 145, 158], phosphatidylinositol-4,5-bisphosphate 3-kinase [48], extracellular-regulated kinases 1/2 [145, 166], and glycogen synthase kinase 3 β [158]. One key enzyme of the SAFE pathway, the transducer and activator of transcription 3 was activated by remote ischemic conditioning in rats with one cycle of 15 min ischemia and 10 min reperfusion on both hind limbs [7] and in pigs with 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb [145]. When the protection was transferred via plasma from pigs to isolated rat hearts, both signaling pathways, RISK and SAFE were activated and causally involved in infarct size reduction [145]. Key enzymes of the RISK and the SAFE pathway were activated downstream of the muscarinic receptor M3 in the myocardium of rats in response to remote ischemic conditioning by one cycle of 15 min ischemia and 10 min reperfusion on both hind limbs [7]. In left ventricular myocardial biopsies of patients undergoing remote ischemic conditioning by 3 cycles of 5 min forearm ischemia and 5 min reperfusion before elective CABG surgery, none of the established cardioprotective proteins was activated, but only signal transducer and activator of transcription 5 was activated at early reperfusion [39, 65]. The intracellular signaling is obviously species-specific.

Apart from and in addition to the above mentioned cardioprotective signaling pathways, heat shock protein heme oxygenase-1 expression was increased in rats after repetitive remote ischemic conditioning by 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb on 1 to 3 consecutive days [173]. Associated with cardioprotection (reduced troponin T release) by remote ischemic conditioning by 4 cycles of 5 min forearm ischemia and 5 min reperfusion in patients undergoing heart surgery under cardiopulmonary bypass, HIF-1 α protein expression was increased in right atrial tissue [2]. A causal involvement of HIF-1 α in cardioprotection by remote conditioning, however, remains to be identified. As mentioned above, HIF-1 α may be a regulator of different humoral mediators of protection.

In rats, remote ischemic conditioning by 4 cycles of 5 min ischemia and 5 min reperfusion on both hind limbs preserved connexin-43 phosphorylation and immunofluorescence staining, indicating a localization of phosphorylated connexin-43 at intercalated disks [10]. Exposure of cultured rabbit ventricular cardiomyocytes to plasma dialysate from rabbits with

remote ischemic conditioning by 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb reduced hypoxia-induced cell-swelling. Experiments using inhibitors identified the activation of potassium- and chloride-channels which control volume and edema formation [27]. However, the role of chloride-channels and cell-volume control in cardioprotection remains somewhat contentious [62, 63].

In rats, miRNA1 was downregulated in response to remote ischemic conditioning by 3 cycles of 5 min ischemia and 5 min reperfusion on both hind limbs [32]. In mice, along with infarct size reduction by remote ischemic conditioning with 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb, miRNA144 was increased in myocardial tissue. A reduction of myocardial miRNA144 levels with a specific antisense oligonucleotide abrogated the observed infarct size reduction [98]. In association with remote ischemic conditioning by 3 cycles of 4 min forearm ischemia and 4 min reperfusion [73] or by 3 cycles of 5 min forearm ischemia and 5 min reperfusion [147], respectively, miRNA1 and miRNA195 were downregulated and miRNA388-3p was upregulated in right atrial myocardium of patients undergoing double valve replacement [73] or elective CABG surgery [147]. Up to now, all potential downstream pathways or interacting pathways of the miRNAs in conditioning rely on literature search and not on experimental data [4]. Further experiments are necessary to identify a potentially causal role and a relevance of miRNAs in remote conditioning.

Mitochondria as targets and potential effectors: Mitochondria have been identified as a major common intracellular target during local and remote ischemic conditioning [56, 64]. The preservation of mitochondrial integrity and function after ischemia/reperfusion is decisive for survival of cardiomyocytes and thus salvage of myocardium. Mitochondria are the major cellular source of ATP under aerobic conditions and therefore relevant for all cellular functions and cell survival [35, 56]. In ischemia/reperfusion, in contrast, mitochondrial permeability transition pore (mPTP) opening may initiate cell death by cytochrome C release and initiation of intracellular proteolysis.

In rats, myocardial infarct size reduction by remote ischemic conditioning with 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb was associated with preserved mitochondrial morphology [18], maintained mitochondrial membrane potential and increased mitochondrial manganese superoxide dismutase content [17]. Mitochondrial respiration was improved in rat hearts [35] and in right atrial appendages of patients undergoing elective CABG surgery [146, 147] in response to remote ischemic conditioning by 3 cycles of 5 min ischemia and 5 min reperfusion on one hind limb and by 4 cycles of 5 min forearm ischemia and 5 min reperfusion, respectively. Humoral transfer (pig to rat [40] or rabbit to rabbit [162]) of cardioprotection by remote ischemic conditioning with 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb improved mitochondrial function.

Mitochondrial adenosine diphosphate-stimulated respiration was improved [40, 162], calcium retention capacity as an estimate of mPTP opening was increased [40], adenosine triphosphate (ATP) concentration was increased [40] and reactive oxygen species formation decreased [40]. The improved mitochondrial function in rat myocardium was associated with infarct size reduction [40]. Nitrosation and nitrosylation of mitochondrial membrane proteins are causally involved in cardioprotection [88]. In response to remote ischemic conditioning by 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb in mice, myocardial nitrite is increased [128]. Myocardial myoglobin reduces nitrite to bioactive nitric oxide, and nitric oxide subsequently inhibits mitochondrial complex I activity [53]. This nitrosation triggered a reduction of complex I activity and finally reduced myocardial reactive oxygen species formation in response to remote ischemic conditioning by 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb in mice [128]. In rats, remote ischemic conditioning by 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb increased mitochondrial dynamics and the expression of the mitochondrial fusion protein dynamin-like 120 kDa protein (OPA1); mitochondrial morphology was preserved [18]. Remote ischemic conditioning by 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb induced mitochondrial ATP-dependent K(+) channel opening in rat myocardium. The mitochondrial ATP-dependent K(+) channel blocker 5-hydroxydecanoic acid abolished the infarct size reduction by remote ischemic conditioning [93]. Aldehyde dehydrogenase 2 is located in the mitochondrial matrix and detoxifies aldehydes [21, 22]. Aldehyde dehydrogenase 2 was involved in the infarct size reduction by remote ischemic conditioning with 4 cycles of 5 min ischemia and 5 min reperfusion on one hind limb in rabbits and one forearm in humans, respectively. Inhibition of aldehyde dehydrogenase 2 by cyanamide abrogated the myocardial infarct size reduction in rabbits, and a single nucleotide polymorphism which inactivates aldehyde dehydrogenase 2 abrogated the attenuation of ischemia/reperfusion-induced endothelial dysfunction in humans [23] (Fig. 6).

Coronary vascular response

Ischemia/reperfusion-injury of the coronary microcirculation is characterized by increased vascular permeability and edema, endothelial dysfunction and impaired vasomotion, microembolization of atherothrombotic debris, and stasis with intravascular cell aggregates. Finally the vascular ischemia/reperfusion may result in its most severe form: destruction of the capillaries with hemorrhage [58]. Conversely, the coronary microcirculation is a target of cardioprotection, including that by remote conditioning. After ischemia/reperfusion, coronary flow and coronary flow reserve were improved in mice [71], pigs [141] and humans [89]. Red blood cell nitric

oxide synthase activity was stimulated by remote ischemic conditioning with 4 cycles of 5 min forearm ischemia and 5 min reperfusion in healthy volunteers along with improved red blood cell deformability *in vitro* [44]. Leukocyte adhesion was reduced in healthy volunteers by remote ischemic conditioning with 3 cycles of 5 min forearm ischemia and 5 min reperfusion [143]. In healthy volunteers after ischemia/reperfusion injury (20 min ischemia with reperfusion) on one forearm [118], in patients undergoing coronary angiography [96] and in patients undergoing ablation for atrial fibrillation, remote ischemic conditioning with 3 cycles of 5 min forearm ischemia and 5 min reperfusion [150] attenuated the number of monocyte-platelet aggregates.

Translation of remote ischemic conditioning into clinical practice

Remote ischemic conditioning is also operative in humans. In the initial studies in healthy volunteers, endothelium-dependent vasodilation was taken as endpoint, and 3 preceding cycles of 5 min forearm ischemia and 5 min reperfusion attenuated the impairment of vasodilation following 20 min ischemia and reperfusion in the contralateral arm [86, 103, 104]. This protection of endothelium-dependent vasodilation was immediate, but a second window of protection was apparent 24 and 48 h later [103]. Protection was even seen when the remote ischemic conditioning procedure was performed during the sustained arm ischemia (preconditioning) [104], and activation of the autonomous nervous system and of ATP-dependent K(+) channels was causally involved [103, 104].

Subsequently, remote ischemic conditioning by 3 cycles of 5 min blood pressure cuff inflation and 5 min deflation was demonstrated to reduce myocardial injury, as reflected by troponin release, in patients undergoing CABG surgery under cardiopulmonary bypass and ischemic cardioplegic arrest [47]. Such protection by remote ischemic conditioning in patients undergoing cardiovascular surgery was confirmed in a number of, but not all studies [61, 134], and even an improved short-term [16] and long-term [156] clinical outcome became apparent from retrospective analyses. However, two recent large phase III trials on remote ischemic preconditioning in patients undergoing cardiovascular surgery in which clinical outcome was the primary endpoint were neutral, with respect to biomarker release and to clinical outcome [49, 111]. Unfortunately, all [111] or almost all [49] patients in these trials were operated under propofol anesthesia, although it was previously known that propofol specifically abrogates the protection by remote ischemic conditioning whereas volatile anesthesia does not [91, 92, 169]. Therefore, the actual potential of remote ischemic conditioning to confer protection in patients undergoing cardiovascular surgery remains unclear [59]. Interestingly, remote ischemic preconditioning protects

against acute kidney injury in cardiosurgical patients when these are operated without the use of propofol [170].

Importantly, in patients who suffer from an acute myocardial infarction and undergo interventional or thrombolytic reperfusion, remote ischemic preconditioning reduces myocardial injury, as reflected by reduced biomarker release or reduced infarct size on imaging, in all studies published so far [61], and it even improves clinical outcome on retrospective analysis [148]. In acute myocardial infarction, the myocardial injury is obviously greater and protection more needed than in cardiovascular surgery, and patients with acute myocardial infarction have no propofol on board. A large phase III trial with clinical outcome as primary endpoint in patients with reperfused acute myocardial infarction is on its way [50].

While the available translational studies are encouraging, notably those in acute myocardial infarction, we need to develop a better understanding of the mechanisms and signal transduction of remote ischemic conditioning. Such better understanding will help us to avoid pitfalls in translational attempts such as the use of propofol and to deal with other confounding variables such as advanced age, and the multiple co-morbidities and co-medications in those patients who need the additional protection by remote ischemic conditioning the most [34].

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