

Hyperhomocysteinemia impairs regional blood flow: involvements of endothelial and neuronal nitric oxide

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Abstract Increasing evidence support the idea that hyperhomocysteinemia (HHcy) is responsible for pathogenesis underlying cerebral, coronary, renal, and other vascular circulatory disorders and for hypertension. Impaired synthesis of nitric oxide (NO) in the endothelium or increased production of asymmetric dimethylarginine and activated oxygen species are involved in the impairment of vasodilator effects of NO. Impaired circulation in the brain derived from reduced synthesis and actions of NO would be an important triggering factor to dementia and Alzheimer's disease. Reduced actions of NO and brain hypoperfusion trigger increased production of amyloid- β that inhibits endothelial function, thus establishing a vicious cycle for impairing brain circulation. HHcy is involved in the genesis of anginal attack and coronary myocardial infarction. HHcy is also involved in renal circulatory diseases. The homocysteine (Hcy)-induced circulatory failure is promoted by methionine and is prevented by increased folic acid and vitamin B₆/B₁₂. Eliminating poor life styles, such as smoking and being sedentary; keeping favorable dietary habits; and early treatment maintaining constitutive NOS functions healthy, reducing oxidative stresses would be beneficial in protecting HHcy-induced circulatory failures.

Keywords Hyperhomocysteinemia · Nitric oxide · Cerebral blood flow · Coronary blood flow · Oxidative stress

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Abbreviations

ACh	Acetylcholine
A β	Amyloid- β
AD	Alzheimer's disease
ADMA	Asymmetric dimethylarginine
A β PP	A β -protein precursor (A β PP)
DDAH	Dimethylaminohydrolase
EPC	Endothelial progenitor cell
GABA	γ -Amino butylic acid
Hcy	Homocysteine
HHcy	Hyperhomocysteinemia
NO	Nitric oxide
PPAR γ	Peroxisome proliferator-activated receptor- γ
SOD	Superoxide dismutase

Introduction

Homocysteine (Hcy) is a thiol-containing amino acid that is formed from methionine, an essential amino acid contained in large quantities in meat. Recent evidence indicates that elevated plasma level of Hcy is a risk factor for occlusive artery disease, especially in the brain [9, 17], the heart [16, 55, 69], and the kidney [48].

Cerebral blood flow and peripheral circulation are controlled by vasodilator factors, including endothelium-derived nitric oxide (NO), prostacyclin, endothelium-derived hyperpolarizing factors, and vasodilator peptides as well as vasoconstrictors, such as vasoconstrictor prostaglandins, oxidative stress, and asymmetric dimethylarginine (ADMA), an endogenous inhibitor of nitric oxide synthase (NOS). Neural control of cerebral blood flow is regulated mainly by parasympathetic nitrgic nerves, whereas adrenergic vasoconstrictor nerves are mainly involved in peripheral blood flow control; coronary blood flow is controlled by norepinephrine liberated from

adrenergic nerves that induces vasodilatation via activation of adrenergic β -adrenoceptors and vasoconstriction by α -adrenoceptor stimulation [77]. Hyperhomocysteinemia (HHcy) interferes with cerebral and peripheral blood flow mainly by impairment of actions and syntheses of vasodilator factors, such as NO [15, 26, 45, 48]. HHcy also increases oxidative stress and accumulate ADMA, which acts as an endogenous inhibitor of NOS [72, 78], and prevents expression of endothelial dimethylaminohydrolase (DDAH), the main catabolic enzyme of ADMA [23, 49, 70]. Dietary folic acid decreases the plasma HHcy levels [44]. Folate supplementation to adults was found to protect against ischemic stroke or myocardial infarction [37].

Here, we will review and discuss the importance of current research on the modulation of regional blood flow or vascular tone, mainly in the cerebral, coronary, and renal vasculatures, maintained by basal and stimulated release of NO from the endothelium and nitrenergic nerves or by oxygen radicals following HHcy in humans and experimental animals.

General features of vascular function in hyperhomocysteinemia

Hcy is an independent risk factor for atherothrombosis [50]. Hcy is a nonprotein amino acid generated during nucleic acid methylation and demethylation of methionine (Fig. 1). Folic acid decreases Hcy levels by remethylating the Hcy to methionine. HHcy is a defined state connected to the increased risk of atherothrombotic, atherosclerotic, and vascular constrictive effects, largely as a result of facilitating the generation of hydrogen peroxide from oxygen. Endothelium-derived relaxing factor (EDRF) [31] or NO is suggested to S-nitrosate Hcy, rendering it nontoxic to the endothelium [67]. Prolonged exposure of endothelial cells to Hcy impaired vasodilator responses to EDRF, whereas S-NO-Hcy neither supported H_2O_2 generation nor impaired the response to EDRF [68]. These authors suggested that the normal endothelium modulates the adverse effects of Hcy by releasing EDRF. EDRF/NO acts as a detoxifier of Hcy, both by limiting the generation of hydrogen peroxide and by serving as a reducing equivalent for the nitrosation reaction [83]. Responses of resistance vessels to endothelium-dependent vasodilators, acetylcholine (ACh) and ADP, were impaired in cynomolgus monkeys with diet-induced HHcy [45]. These authors suggested that altered vascular function may contribute to vasospasm and thrombosis. Increase in vascular oxidant stress through imbalanced thiol redox status and inhibition of antioxidant enzymes by Hcy results in decreased bioavailability of endothelial NO via oxidative inactivation [85, 87]. Vasodilator effects of endothelium-derived NO and exogenously applied NO donors are compromised by HHcy in anesthetized rats [30]. Elevated plasma levels of ADMA and

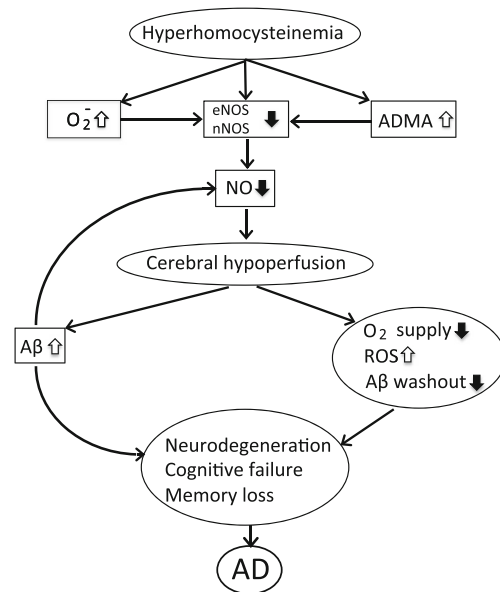


Fig. 1 Possible roles hyperhomocysteinemia in inhibiting eNOS and nNOS functions, impairing cerebral blood flow, and increasing A β deposition in the pathogenesis of Alzheimer's (AD) disease. O_2^- superoxide anion, ADMA asymmetric dimethylarginine

symmetric dimethylarginine are associated with an increased risk of cardiovascular events [8]; the concentrations of these endogenous NOS inhibitors in rat plasma are actually decreased upon methionine administration.

Endothelium-dependent flow-mediated dilation in brachial arteries was significantly lower in HHcy adult subjects than in those with low Hcy levels, suggesting that HHcy is an independent risk factor for arterial endothelial dysfunction [89]. In the forearm circulation of healthy subjects, increased Hcy plasma levels reduce NO availability by producing oxidative stress; in essential hypertensive patients, the presence of hypercholesterolemia causes a further reduction in endothelium-dependent vasodilatation by exacerbating oxidative stress [73]. Atherogenic effects of Hcy appear to depress endothelial function through NO-dependent mechanisms in humans [34].

Oral methionine loading increased Hcy levels and resulted in a decrease in flow-mediated vasodilatation of the brachial artery in healthy subjects, suggesting that acute HHcy impairs endothelial function [1]. Oral methionine raises plasma Hcy and impairs flow-mediated endothelium-dependent brachial artery vasodilatation in healthy adults [6]. In healthy volunteers, methionine increased plasma Hcy and reduced flow-mediated, NO-mediated brachial artery dilatation [14]. Folic acid and vitamin B₁₂ supplementation improved endothelial function in patients with coronary heart disease [16]. In hyperhomocysteinemic patients, folic acid treatment for 12 months lowered elevated plasma Hcy and reversed Hcy-induced impairment of NO vasoreactivity. Acute HHcy decreases flow-mediated vasodilatation of the brachial artery in

healthy subjects; pretreatment with aged garlic extracts diminished the adverse effects of acute HHcy [86].

Cerebral circulation

In anesthetized rats, cerebrocortical blood flow was reduced by superfusion with Hcy-Cu²⁺ but not with Cu²⁺ alone, and the effect of Hcy-Cu²⁺ was prevented by co-administration of superoxide dismutase (SOD); Hcy-Cu²⁺ attenuated the vasodilator response to NO-dependent vasodilators, such as ACh and hypercapnia, suggesting that O₂⁻ generated by the reaction of Hcy with Cu²⁺ inhibits NO-related cerebrovascular responses by scavenging NO [90]. Lee et al. [43] provided evidence suggesting that endothelial dysfunction induced by HHcy is mediated through impairments of endothelial NOS (eNOS) activity in cerebrovascular endothelial cells and that these effects are ameliorated by dietary folic acid supplementation in rats. Dilatation to ACh of cerebral arterioles in mice fed a high-methionine diet was impaired, as compared with those fed a control diet; the vasodilator responses were restored toward normal by the superoxide scavenger tiron, suggesting that superoxide is a key mediator of endothelial dysfunction in the cerebral circulation during HHcy [22]. Betzen et al. [7] suggested that oxidative stress upregulates N-methyl-D-aspartate receptor on murine cerebrovascular endothelium and heightens susceptibility to glutamate-induced blood–brain barrier disruption. Hcy-mediated brain “microvascular endothelial cell” collagen gel constriction was ameliorated by muscimol (γ -amino butylic acid (GABA)-A receptor agonist), baclofen (GABA-B receptor agonist), and eNOS gene ablations [64]. Amelioration of Hcy-induced endothelial cell collagen gel constriction may be induced by NO through GABA-A and GABA-B receptors.

Methionine-induced HHcy caused age-related impairment of cerebrovascular reactivity in healthy adults; this effect was ameliorated by treatment with quinapril, possibly due to improvement of endothelial NO-mediated vasodilatation [17]. Plasma Hcy levels both under basal conditions and post-methionine load were higher in patients with Alzheimer’s disease (AD) and patients with vascular dementia than in normal controls; vitamin B₁₂ basal levels negatively correlated with basal Hcy levels only in AD patients, suggesting the possible role of chronically elevated Hcy in neuronal degeneration in demented patients [84]. Clinical findings show that elevated Hcy levels in plasma are associated with decreasing levels of circulating endothelial progenitor cells (EPC) and that the Hcy-induced EPC reduction may be due to apoptosis through caspase-8-mediated release of cytochrome c; B vitamin (B₆ and B₉) intervention impairs Hcy-induced EPC apoptosis leading to increments in EPC populations [2].

Homocysteine, cerebral blood flow, and Alzheimer’s disease

Supplementation of betaine, an Hcy metabolizer, ameliorated the Hcy-induced memory deficit and enhanced long-term potentiation and also decreased A β production in HHcy rats, possibly due to metabolism of Hcy [13]. Diet-induced high Hcy resulted in an exacerbation of memory deficits and A β and tau neuropathology in mice [47]. Diet-induced HHcy develops cerebral amyloid angiopathy via a reduction of A β clearance and transport within the brain in mice [46].

There is a significant increase in plasma concentration of Hcy and ADMA and a decrease of NO in plasma in AD patients, as compared with control subjects; the inhibition of endothelial NO synthesis by ADMA impairs cerebral blood flow, and the inhibition of nNOS by ADMA may cause cognitive dysfunction in AD [62]. It is suggested that there is a negative, significant relationship between blood and urine Hcy levels and between blood Hcy levels and Mini-Mental State Examination scores, suggesting that the reduced urinary excretion induces elevated Hcy in blood, resulting in cognitive dysfunctions [32]. As already stated, HHcy decreases cerebral blood flow possibly by impairment of eNOS actions and enhancement of NO degradation via oxidative stress. From studies on patients with AD and with normal cognitive function, Cankurtaran et al. [11] suggested that a decrease in antioxidants and an increase in oxidative damage induced by HHcy are linked to AD. Huang et al. [36] reported that cognitive performance in mild AD can be reflected by hypoperfusion of the temporo-parietal region and total Hcy level is an independent risk factor for rapid cognitive decline. Elevated Hcy in adults aged 80 years or older was suggested to contribute to increased AD-type pathology [35]. Behavioral and psychological symptoms of dementia seem to be associated with plasma Hcy concentration in AD-type dementia [91]. It was suggested that high-Hcy and low-folate levels correlate with increased risk of AD occurrence [65].

Vicious cycle in endothelial dysfunction, hypoperfusion, and amyloid- β peptide deposition

As already been mentioned, HHcy is a risk factor for impairing cerebrovascular endothelial function and interfering with NO-mediated cerebral blood flow. Brain and cerebral microvessels from eNOS-deficient mice had higher protein levels of amyloid- β -protein precursor (A β PP) and β -site A β PP cleaving enzyme 1 (BACE1), as well as BACE1 activity and A β , suggesting that endothelial NO plays an important role in modulating A β -protein precursor expression [3]. A β PP and A β _{1–40} were increased in hippocampal tissue of eNOS-deficient mice as compared to wild-type mice; eNOS-deficient mice performed worse in a radial arm maze test of spatial learning and memory, as compared to wild-type mice

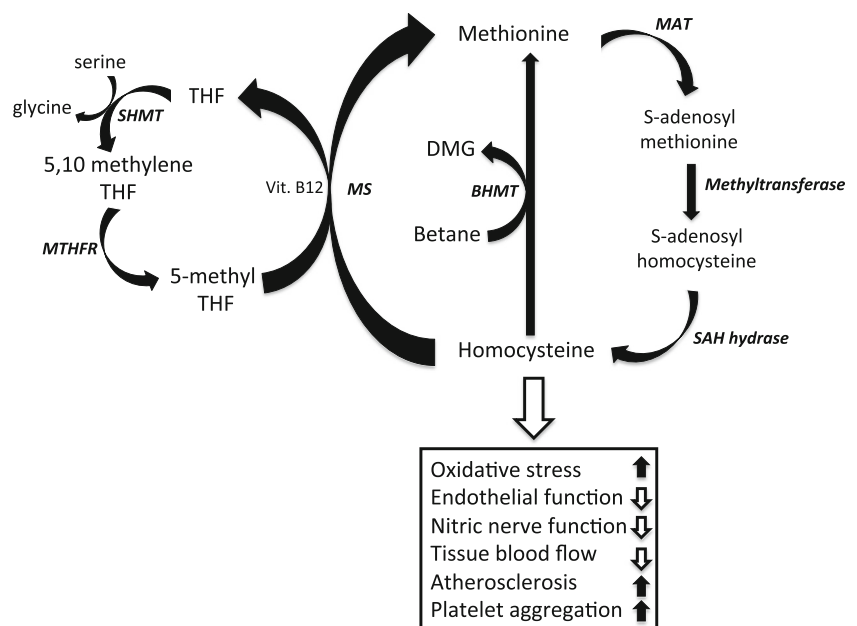
[4]. These authors suggest that NO/cyclic GMP pathway may be an important therapeutic target in preventing and treating mild cognitive impairment, as well as AD. Cerebral hypoperfusion triggers vascular deposition of peripherally applied human A β -42, which is shifted from vasculature to the parenchyma [27]. Reduced cerebral autoregulation is associated with increased amyloid deposition and increased white matter hypersensitivity volume in humans [10]. Aging- and lifestyle-associated damages of the brain microcirculation may affect A β clearance and perivascular drainage, promoting cerebrovascular A β deposition [42]. Increased amounts of A β impair endothelial function in cerebral blood vessels [20, 57]. Endothelial dysfunction causes a decreased cerebral blood flow and as a result increases the A β accumulation that in turn impairs NO bioavailability and promotes cerebral hypoperfusion (Fig. 2) [76]. A decreased cerebral glucose metabolism may precede the A β deposition and A β accumulation in turn leading to further decreases in the cerebral blood flow, closing a vicious cycle [56, 75]. A β -degrading enzymes, including neprilysin- and angiotensin-converting enzymes, may help to maintain cerebral perfusion by reducing the accumulation of A β in cerebral blood vessels [51].

Coronary circulation

In rats with methionine diet-induced HHcy, coronary arterial dilatation in response to increased intraluminal flow was abolished, possibly because of enhanced production of superoxide anions [82]. In HHcy rats and mice fed methionine in drinking water, superoxide produced by the p22phox subunit of NADPH oxidase reduces the ability of NO to regulate

mitochondrial function in the myocardium [5]. In rat cardiac microvascular endothelial cells, Hcy increased inducible NO synthase and decreased endothelial NOS without altering neuronal NOS levels; there was accumulation of ADMA, possibly because of reduced DDAH expression [81]. In conscious dogs fed with methionine to increase plasma Hcy, veratrine-induced, NO-dependent coronary vasodilatation was reduced but was restored by infusion of ascorbic acid or apocinin; HHcy decreased bradykinin- or carbachol-induced reduction of myocardial oxygen consumption, and this effect was restored by co-incubation with ascorbic acid, tempol, or apocinin, suggesting that HHcy impairs NO bioavailability through oxidative stress [71]. In rats, pretreatment with folic acid blunted myocardial dysfunction during ischemia and ameliorated post-reperfusion injury. It was suggested that folic acid preserves high-energy phosphate and reduces subsequent reactive oxygen species generation, eNOS uncoupling, and post-reperfusion injuries [52]. BH₄ supplementation of murine endothelial cells revealed that calcium ionophore-evoked NO bioactivity correlates with intracellular BH₄/BH₂ and not with absolute intracellular BH₄; superoxide production negatively correlated with intracellular BH₄/BH₂, suggesting that diminished BH₄/BH₂ rather than BH₄ depletion per se is the trigger for NO insufficiency [21]. Folic acid administration to post-myocardial infarction in mice improved cardiac ejection fraction and induced tissue inhibitor of metalloproteinase, Hcy-metabolizing enzymes, and 5-methylene tetrahydrofolate reductase, suggesting that folic acid improves myocardial function in myocardial infarction [58]. Plasma HHcy induced by high-methionine diet in rats was prevented by treatment with palm tocotrienol-rich fraction; this compound also reversed the decreased glutathione peroxidase

Fig. 2 Schematic overview of homocysteine metabolism and its relationship with folic acid and vitamin B₁₂. *THF* tetrahydrofolic acid, *SHMT* serine hydroxymethyl transferase, *MTHFR* methylene THF reductase, *MS* methionine synthase, *DMG* 5*N*,10*N*-dimethylglycine, *BHMT* betane homocysteine methyltransferase, *MAT* methionine adenosyltransferase



activity, suggesting that the palm extract is comparable to folate-reducing methionine diet-induced HHcy and oxidative stress [54].

Elevated serum Hcy is an established risk factor for myocardial infarction not only in men but also in middle-aged women [92]. Adenosine-stimulated myocardial blood flow increase was reduced in patients with HHcy, and flow-mediated brachial artery dilatation was also impaired, suggesting that acute HHcy impairs coronary microvascular dilatation as a result of reduced NO bioavailability [74]. In hyperhomocysteinemic patients with coronary artery disease who were treated with folic acid/cobalamin, coronary blood flow increased after ACh infusion, compared with a decrease in blood flow in the placebo-treated group, suggesting that coronary endothelial function improves after treatment with folic acid and cobalamin [88]. Hcy levels were positively correlated with endothelin-1 level and negatively correlated with NO/endothelin-1 in patients with coronary lesions [18]. Chronic HHcy impairs coronary endothelial function, and plasma levels of NO and BH₄ are positively correlated and significantly decreased in patients with HHcy compared with controls [41]. In patients with chronic HHcy, plasma levels of NO and BH₄ were lower and coronary flow velocity reserve was impaired as compared with the control group [33]. Tsuda and Nishio [79] suggested that decreased bioavailability of NO due to ADMA may partially explain the increased risk for coronary heart disease in women with HHcy.

Renal circulation

Incubation of isolated rat small renal arteries with Hcy attenuates the increase in NO in the renal arterial endothelium; decreased NO is not the only mechanism resulting in endothelial dysfunction but also increased superoxide levels in the arterial endothelium [48]. In HHcy rats, superoxide anion concentrations and nitrite + nitrate levels were higher, glomerular filtration rate was lower, renal plasma flow was lower, and renal vascular resistance was higher, as compared with control rats; after treatment with L-arginine, the responses of glomerular filtration rate, renal plasma flow, and renal vascular resistance were attenuated [29]. HHcy seems to induce oxidative stress, NO inactivation, and renal dysfunction involving disturbances in the NO pathway. The renal blood flow increase mediated via endothelium-dependent hyperpolarizing factor was also inhibited in HHcy rats [24]. Chien et al. [19] reported that in children with early chronic kidney disease, blood pressure abnormalities assessed by ambulatory blood pressure monitoring were associated with L-arginine-to-ADMA ratio, Hcy, and L-cysteine. These authors emphasized that the effect of NO and the Hcy pathway is important in the genesis of chronic kidney disease-related hypertension.

The glomerular filtration rate increased by diabetic induction in mice was normalized following treatment with the peroxisome proliferator-activated receptor- γ (PPAR γ) agonist ciglitazone that ameliorated the decreased bioavailability of NO in diabetic mice [63]. Ciglitazone appears to protect against diabetic nephropathy by activating PPAR γ and clearing glomerular tissue Hcy. Several lines of evidence suggest that PPAR γ ligands protect endothelial function and improve vascular functions [40, 60, 66]. Kidneys isolated from HHcy rats showed increased levels of superoxide anions and lipid peroxides; folic acid supplementation antagonized HHcy-induced oxidative stress via its HHcy-lowering effect, attenuated xanthine oxidase activity, and restored SOD activity in the kidney [38]. Folic acid appears to be effective in protecting kidneys against oxidative stress. Lowering plasma Hcy by folic acid inhibits transforming growth factor- β 1 expression and attenuates HHcy-induced glomerular damage [12].

Betaine is the substrate of the kidney- and liver-specific betaine-Hcy methyltransferase, an alternative pathway for Hcy remethylation. In the diet-induced HHcy rat model, betaine was decreased in all tissues analyzed (kidney, liver, and heart); in the mouse cystathionine β -synthase deficiency model, betaine was decreased in plasma, liver, heart, and brain but was conserved in kidney [39].

Others

When endothelial cells were exposed to Hcy or to its precursor methionine, ADMA concentration in the cell culture medium increased. Hcy-induced accumulation of ADMA was associated with reduced NO synthesis by endothelial cells and segments of pig aorta; Hcy reduced the activity of dimethylarginine dimethylaminohydrolase (DDAH), the enzyme that degrades ADMA [70]. Hcy appears to inhibit DDAH enzyme activity, causing ADMA to accumulate and inhibit NO synthesis.

In rats with diet-induced HHcy, the vasodilator response to ACh was attenuated, suggesting that Hcy might contribute to defective NO availability; Hcy-induced endothelial dysfunction is compensated by H₂S in a NO-independent manner [25]. These authors suggested that impaired vasodilatation, possibly via the eNOS inhibition and hepatic stellate cell contraction caused by Hcy, contributes to the dynamic component of portal hypertension. Hcy treatment increased blood pressure in wild-type mice, whereas in HHcy-cystathionine β synthase heterozygote knockout (CBS^{-/+}) mice, Hcy did not increase blood pressure; DDAH-2 and eNOS expressions in mesenteric arteries were decreased in mesenteric arteries in CBS^{-/+} mice compared with wild-type mice, suggesting that HHcy causes mesenteric remodeling and narrowing by decreasing eNOS expression [53]. The tibial bone blood flow and eNOS protein levels in HHcy-CBS^{+/-} mice decreased, as

compared with those in wild-type mice; these effects in CBS mice were ameliorated by folic acid supplementation, suggesting the efficacy of folic acid on HHcy-induced bone loss [80]. There were increases in flow-mediated blood flow in control rat gracilis muscle venules, but constrictions were induced in HHcy rat; flow-induced constrictions seen in HHcy rats were converted to dilatations in the presence of thromboxane A₂-receptor antagonists, which were abolished by the NOS inhibitor L-NAME and the cyclooxygenase inhibitor indomethacin. A superoxide dismutase diminished flow-induced venular constrictions in HHcy rats, suggesting that TxA₂ and reactive oxygen species overcome NO and vasodilator prostaglandins in the venule circulation of HHcy rats [59].

It was suggested that elevated serum Hcy is responsible for the endothelial damage in Bahçet's disease and may be an additional risk factor for the development of retinal vascular occlusive disease [28]. Microparticles (MPs) are small membrane vesicles released by stimulated or apoptotic cells. Human umbilical vein endothelial cells stimulated by Hcy produced more microparticulates than umbilical endothelial cells under the control conditions; endothelial function impairment due to HHcy is related to microparticulate shedding, which may involve platelets and vascular cells [61].

Summary

In this review, we have discussed impaired regional blood flow in experimental animals and humans by HHcy in relation to suppressed function of NO produced in the endothelium and parasympathetic vasodilator neurons. Endothelial NO plays important roles in decreased vascular resistance and increased regional blood flow in the heart, kidney, and other peripheral organs and tissues, whereas nitroergic neuronal vasodilatation plays important roles in increasing cerebral blood flow. In cerebral arteries and arterioles, both endothelial NO and neuronal NO are involved in vasodilatation and blood flow increase; in renal vasculatures, endothelial NO and neuronal NO participate in vasodilatation, but adrenergic nerves play a role as vasoconstrictors via α -adrenoceptor stimulation. In coronary vasculatures, endothelial NO and norepinephrine as a β -agonist contribute to vasodilatation but neuronal NO does not appear to play a role as a vasodilator [77]. Cerebral hypoperfusion functionally induced via NO-related mechanisms elicits not only reduced blood supply but also increased production of β -amyloid, this substance interfering with brain circulation via reduced endothelial function [76]. This is in contrast to brain embolism chronically and histologically induced. These effects induced by increased plasma concentrations of Hcy are expected to be involved in the genesis and development of cognitive failure and AD. Impaired coronary blood flow by HHcy is one of the risk factors in eliciting anginal attack and myocardial infarction. Impaired circulation

in the kidney and other organs and tissues by increased concentrations of Hcy not only participate in local blood flow impairment but also in increasing systemic blood pressure. Folic acid, vitamin B₆/B₁₂, and related substances decrease plasma concentrations of Hcy and expected to play roles in the prevention and treatment of circulatory diseases and AD. Increased HHcy is one of the risk factors for defective functional control of cerebral, coronary, renal, and other vasculatures. It would be important to recognize that HHcy-induced cerebral hypoperfusion due to impaired endothelial function triggers a vicious cycle of cerebral hypoperfusion-increased, β -amyloid deposition-induced endothelial dysfunction [76], leading to an incidence of cognitive failure and AD. Further efforts devoted to advance our understanding of physiological and pathophysiological actions of endothelial and neuronal NOs, and its counteracting molecules such as oxidative stress and ADMA on regional and systemic blood flow regulation will contribute to development of novel ways for prevention and therapy against cardiovascular disorders.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

References

1. Abahji TN, Nill L, Ide N, Keller C, Hoffmann U, Weiss N (2007) Acute hyperhomocysteinemia induces microvascular and macrovascular endothelial dysfunction. *Acta Med Res* 38:411–416
2. Alam MM, Mohammad AA, Shuaib U, Wang C, Ghani U, Schwandt B, Todd KG, Shyaoib A (2009) Homocysteine reduces endothelial progenitor cells in stroke patients through apoptosis. *J Cereb Blood Flow Metab* 29:157–165
3. Austin SA, Santhanam AV, Katusic ZS (2010) Endothelial nitric oxide modulates expression and processing of amyloid precursor protein. *Circ Res* 107:1498–1502
4. Austin SA, Santhanam AV, Hinton DJ, Choi DS, Katusic ZS (2013) Endothelial nitric oxide deficiency promotes Alzheimer's disease pathology. *J Neurochem* 127:691–700
5. Becker JS, Adler A, Schneeberger A, Huang H, Wang Z, Walsh E, Koller A, Hintze TH (2005) Hyperhomocysteinemia, a cardiac metabolic disease: role of nitric oxide and the p22phox subunit of NADPH oxidase. *Circulation* 111:2112–2118
6. Bellamy MF, McDowell IF, Ramsey MW, Brownlee M, Bonses C, Newcombe RG, Lewis (1998) Hyperhomocysteinemia after an oral methionine load acutely impairs endothelial function in human adults. *Circulation* 98:1848–1852
7. Betzen C, White R, Zehender CM, Pietrowski E, Bender B, Luhmann HJ, Kuhlmann CR (2009) Oxidative stress upregulates the NMDA receptor on cerebrovascular endothelium. *Free Radic Biol Med* 47:1212–1220
8. Boger RH (2006) Asymmetric dimethylarginine (ADMA): a novel risk marker in cardiovascular medicine and beyond. *Ann Med* 38:126–136

9. Brattsström L, Lindgren A, Israelsson B, Malinow MR, Norrving B, Upson B, Hamfelt A (1992) Hyperhomocysteinemia in stroke: prevalence, cause, and relationships to type of stroke and stroke risk factors. *Eur J Clin Invest* 22:214–221
10. Brickman AM, Guzman VA, Gonzalez-Castellon M, Razlighi Q, Gu Y, Narkhede A, Janicki S, Ichise M, Stern Y, Manly JJ, Schupf N, Marshall RS (2015) Cerebral autoregulation, beta amyloid, and white matter hyperintensities are interrelated. *Neurosci Lett* 592: 54–58
11. Cankurtaran M, Yesil Y, Kuyumcu ME, Oztet ZA, Yavuz BB, Halil M, Ulger Z, Cankurtaran ES, Arilogul S (2013) Altered levels of homocysteine and serum natural antioxidants links oxidative damage to Alzheimer's disease. *J Alzheimers Dis* 33:1051–105
12. Cao L, Lou X, Zou Z, Mou N, Wu W, Huang X, Tan H (2013) Folic acid attenuates hyperhomocysteinemia-induced glomerular damage in rats. *Microvasc Res* 89:146–152
13. Chai GS, Jiang X, Ni ZF, Ma ZW, Xie AJ, Cheng XS, Wang JZ, Liu GP (2013) Betaine attenuates Alzheimer-like pathological changes and memory deficits induced by homocysteine. *J Neurochem* 124: 388–396
14. Chambers JC, Obeid OA, Kooner JS (1999) Physiological increments in plasma homocysteine induce vascular endothelial dysfunction in normal human subjects. *Arterioscler Thromb Vasc Biol* 19:22–292
15. Chambers JC, Obeid OA, Refsum H, Ueland P, Hackett D, Hopper J, Turner RM, Thompson SG, Kooner JS (2000) Plasma homocysteine concentrations and risk of coronary heart disease in UK Indian Asian and European men. *Lancet* 355:523–527
16. Chambers JC, Ueland PM, Obeid OA, Wrigley J, Refsum H, Kooner JS (2000) Improved vascular endothelial function after oral B vitamins: an effect mediated through reduced concentrations of free plasma homocysteine. *Circulation* 102:2479–2483
17. Chao C-L, Lee Y-T (2000) Impairment of cerebrovascular reactivity by methionine-induced hyperhomocysteinemia and amelioration by quinapril treatment. *Stroke* 31:907–2911
18. Chen Z, Li CS, Zhang J, Pang BS, Xia CQ, Liu XF (2005) Relationship between endothelial dysfunction and serum homocysteine in patients with coronary lesions. *Clin Med Sci J* 20:63–66
19. Chien S-J, Lin I-C, Hsu CN, Lo MH, Tain YL (2015) Homocysteine and arginine-to-asymmetric dimethylarginine ratio associated with blood pressure abnormalities in children with early chronic kidney disease. *Circ J* 79:2031–2037
20. Chisari M, Merlo S, Sortino MA, Solomone S (2010) Long-term incubation with b-amyloid peptides impairs endothelium-dependent vasodilatation in isolated rat basilar artery. *Pharmacol Res* 61: 157–161
21. Crabtree MJ, Smith CL, Lam G, Goligorsky MS, Gross SS (2008) Ratio of 5,6,7,8-tetrahydrobiopterin to 7,8-dihydrobiopterin in endothelial cells determines glucose-elicited changes in NO vs. superoxide production by eNOS. *Am J Physiol* 294:H1530–H1540
22. Dayal S, Arming E, Bottiglieri T, Böger RH, Sigmund CD, Faraci FM, Lentz SR (2004) Cerebral vascular dysfunction mediated by superoxide in hyperhomocysteinemic mice. *Stroke* 35:1957–196
23. Dayal S, Rodinov RN, Arming E, Bottiglieri T, Kimoto M, Murray DJ, Cooke JP, Faraci FM, Lentz SR (2008) Tissue-specific down-regulation of dimethylarginine dimethylaminohydrolase in hyperhomocysteinemia. *Am J Physiol* 295:H816–H825
24. De Vriese AS, Blom HJ, Heil SG, Mortier S, Kluijtmans LA, Van de Voorde J, Lameite NH, Mortier S, Kluijtmans LA, Van de Voorde J, Lameite NH (2004) Endothelium-derived hyperpolarizing factor-mediated renal vasodilatory response is impaired during acute and chronic hyperhomocysteinemia. *Circulation* 109:2331–2336
25. Distrutti E, Mencarelli A, Santucci L, Renga B, Orlandi S, Donini A, Shah V, Fiorucci S (2008) The methionine connection: homocysteine and hydrogen sulfide exert opposite effects on hepatic microcirculation in rats. *Hepatology* 47:659–667
26. Eberhardt RT, Forgione MA, Cap A, Leopold JA, Rudd MA, Troillet M, Heydrick S, Stark R, Klings ES, Moldovan NI, YM, Goldschmidt-Clermont PJ, Farber HW, Cohen R, Loscalzo J, Moldovan NI, Yaghoubi M, Goldschmidt-Clermont PJ, Farber HW, Cohen R, Loscalzo J (2000) Endothelial dysfunction in a murine model of mild hyperhomocyst(e)inemia. *J Clin Invest* 106:483–49
27. Elali A, Thériault P, Préfontaine P, Rivest S (2013) Mild chronic cerebral hypoperfusion induces neurovascular dysfunction, triggering peripheral β -amyloid brain entry and aggregation. *Acta Neuropath Commun* 1:75
28. Er H, Evereklioglu C, Cumurcu T, Türköz Y, Ozerol E, Sahin K, Doganay S (2002) Serum homocysteine levels is increased and correlated with endothelin-1 and nitric oxide in Bahçet's disease. *Br J Ophthalmol* 86:653–657
29. Fischer PA, Dominguez GN, Cuniberti LA, Matinez V, Werba JP, Ramirez AJ, Masnatta LD (2003) Hyperhomocysteinemia induces renal emodynamic dysfunction: is nitric oxide involved? *J Am Soc Nephrol* 14:653–660
30. Fu WY, Dudman NP, Perry MA, Wang XL (2002) Homocysteine attenuates hemodynamic responses to nitric oxide in vivo. *Atherosclerosis* 161:169–176
31. Furchgott RF, Zawadzki JV (1980) The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. *Nature (Lond)* 288:373–376
32. Hasegawa T, Ichiba M, Matsumoto SE, Kasanuki K, Hatano T, Fujishiro H, Iseki E, Hattori N, Yamada T, Tabira T (2012) Urinary homocysteic acid levels correlate with mini-mental state examination scores in Alzheimer's disease patients. *J Alzheimers Dis* 31:59–64
33. He L, Zeng H, Li F, Feng J, Liu S, Yu J, Mao J, Hong T, Chen AF, Wang X, Wang G (2010) Homocysteine impairs coronary artery endothelial function by inhibiting tetrahydrobiopterin in patients with hyperhomocysteinemia. *Am J Physiol* 299:E1061–1065
34. Holven KB, Holm T, Aukrust P, Christensen B, Kjekshus J, Andreassen AK, Gullestad L, Hagve TA, Svilaas A, Ose L, Nenseter MS (2001) Effect of folic acid treatment on endothelium-dependent vasodilation and nitric oxide-derived end products in hyperhomocysteinemic subjects. *Am J Med* 11:536–542
35. Hooshmand B, Polvikoski T, Kivipelto M, Tanskanen M, Mlyllkangas L, Erkinjuntti T, Mäkelä M, Oinas M, Paetau A, Scheltens P, van Straaten EC, Sulkava R (2013) Plasma homocysteine, Alzheimer and cerebrovascular pathology: a population-based autopsy study. *Brain* 136:2707–2716
36. Huang CW, Chang WN, Huang SH, Lui CC, Chen NC, Chang YT, Lee CC, Chang CC, Chang AY (2013) Impact of homocysteine on cortical perfusion and cognitive decline in mild Alzheimer's disease. *Eur J Neurol* 20:1191–1197
37. Huo Y, Li J, Qin X, Huang Y, Wang X, Gottesman RF, 27 collaborators (2015) Efficacy of folic acid therapy in primary prevention of stroke among adults with hypertension in China: the CSPPT randomized clinical trial. *JAMA* 313:1325–1335
38. Hwang SY, Siow YL, Au-Yeung KK, House J (2011) Folic acid supplementation inhibits NADPH oxidase-mediated superoxide anion production in the kidney. *Am J Physiol* 300:F189–F198
39. Imbard A, Benoist JF, Esse R, Gupta S, Lebon S, de Vriese AS, de Baulny HO, Kruger W, Sciff M, Blom HJ (2015) High homocysteine induces betaine depletion. *Biosci Rep* 35, e0222. doi:10.1042/BSR20150094
40. Ketsawatsonkron P, Pelham CJ, Groh S, Keen L, Faraci FM, Sigmund CD (2010) Does peroxisome proliferator-activated receptor- γ (PPAR- γ) protect from hypertension directly through effects in the vasculature? *J Biol Chem* 285:9311–9316

41. Kietadisorn R, Kietselaer BL, Schmidt HHHW, Moens An L (2011) Role of tetrahydrobiopterin (BH4) in hyperhomocysteinemia-induced endothelial dysfunction: new indication for this orphan drug? *Am J Physiol* 300, E1176
42. Kuznetsova E, Schliebs R (2013) β -Amyloid, cholinergic transmission and cerebrovascular system—a developmental study in a mouse model of Alzheimer's disease. *Curr Pharm Des* 19:6749–6765
43. Lee BJ, Huang MC, Chung LJ, Cheng CH, Lin KL, Su KH, Juang YC (2004) Folic acid and vitamin B12 are more effective than B6 in lowering fasting plasma homocystein concentration in patients with coronary artery disease. *Eur J Clin Nutr* 58:481–487
44. Lee H, Kim HJ, Kim J, Chang N (2004) Effects of dietary folic acid supplementation on cerebrovascular endothelial dysfunction in rats with induced hyperhomocysteinemia. *Brain Res* 996:139–147
45. Lentz SR, Sobey CG, Piegors DJ, Bhopatkar MY, Faraci FM, René Malinow M, Heistad DD (1996) Vascular dysfunction in monkey with diet-induced hyperhomocysteinemia. *J Clin Invest* 98:24–29
46. Li JG, Praticó D (2015) High levels of homocysteine results in cerebral amyloid angiopathy in mice. *J Alzheimers Dis* 43:29–35
47. Li JG, Chu J, Barrero C, Merali S, Praticó D (2014) Homocysteine exacerbates β -amyloid pathology, tau pathology, and cognitive deficit in a mouse model of Alzheimer disease with plaques and tangles. *Ann Neurol* 75:851–863
48. Li N, Yi F-X, Rute E, Zang DX, Slocum GR, Zou A-P (2002) Effects of homocysteine on intracellular nitric oxide and superoxide levels in the renal arterial endothelium. *Am J Physiol* 283:H1237–1243
49. Magne J, Huneau J-F, Borderie D et al (2015) Plasma asymmetric and symmetric dimethylarginine in a rat model of endothelial dysfunction induced by acute hyperhomocysteinemia. *Amino Acids* 47:1975–1982
50. Malinow MR (1990) Hyperhomocyst(e)inemia. A common and easy reversible risk factor for occlusive atherosclerosis. *Circulation* 81:2004–2006
51. Miners JS, Palmer JC, Taylor H, Palmer LE, Ashby E, Kehoe PG, Love S (2014) A β degradation or cerebral perfusion? Divergent effects of multifunctional enzymes *Front Aging Neurosci* 6:238. doi:10.3389/fnagi.2014.00238, **eCollection 2014**
52. Moens AL, Champion HC, Claeys MJ, Tavazzi B, Kaminski PM, Wolin MS, Borgonjon DJ, Van Nassauw L, Haile A, Zviman M, Bedja D, Wuyts FL, Elsaesser RS, Cos P, Gabrielson KL, Lazzarino G, Paolucci N, Timmermans JP, Vrints CJ, Kass DA (2008) High-dose folic acid pretreatment blunts cardiac dysfunction during ischemia coupled to maintenance of high-energy phosphates and reduces postreperfusion injury. *Circulation* 117:1810–1819
53. Munjal C, Givvimani S, Qipshidze N, Tyagi N, Falcone JC, Tyagi SC (2011) Mesenteric vascular remodeling in hyperhomocysteinemia. *Mol Cell Biochem* 348:99–108
54. Norsidah KZ, Asmadi AY, Azizi A, Faizah O, Kamisah Y (2013) Palm tocotrienol-rich fraction reduced plasma homocysteine and heart oxidative stress in rats with a high-methionine diet. *J Physiol Biochem* 69:441–449
55. Nygård O, Nordrehaug EJ, Refsum H, Ueland PM, Farstad M, Vollset SE (1997) Plasma homocysteine levels and mortality in patients with coronary artery disease. *NEng J Med* 337:230–236
56. Popa-Wagner A, Buga AM, Popescu B, Muresanu D (2013) Vascular cognitive impairment, dementia, aging and energy demand. A vicious cycle. *J Neural Traum* (Viena) 122(1):S47–S54
57. Price JM, Hellermann A, Hellermann G, Sutton ET (2004) Aging enhances vascular dysfunction induced by the Alzheimer's peptide β -amyloid. *Neurol Res* 26:305–311
58. Qipshidze N, Tyagi N, Sen U, Givvimani S, Metreveli N, Lominadze D, Tyagi SC (2010) Folic acid mitigates cardiac dysfunction by normalizing the levels of tissue inhibitor of metalloproteinase and homocysteine- metabolizing enzymes postmyocardial infarction in mice. *Am J Physiol* 299:H1484–1493
59. Racz A, Veresh Z, Lotz G, Bagi Z, Koller A (2010) Ctclooxygenase-2 derived thromboxane A2 and reactive oxygen species mediates flow-induced constriction of venules in hyperhomocysteinemia. *Atherosclerosis* 208:43–49
60. Ryan MJ, Didion SP, Mathur S, Faraci FM, Sigmund CD (2004) PPAR γ agonist rosiglitazone improves vascular function and lowers blood pressure in hypertensive transgenic mice. *Hypertension* 43:661–666
61. Sekula M, Janawa G, Stankiewicz E, Stepień E (2011) Endothelial microparticle formation in moderate concentrations of homocysteine and methionine in vitro. *Cell Mol Biol Lett* 16:69–78
62. Selley ML (2003) Increased concentrations of homocysteine and asymmetric dimethylarginine and decreased concentrations of nitric oxide in the plasma of patients with Alzheimer's disease. *Neurobiol Aging* 24:903–907
63. Sen U, Rodriguez WE, Tyagi N, Kumar M, Kundu S, Tyagi SC (2008) Ciglitazone, a PPAR γ agonist, ameliorates diabetic nephropathy in part through homocysteine clearance. *Am J Physiol* 295:E1205–E1212
64. Shastri S, Moning L, Tyagi N, Steed M, Tyagi SC (2005) GABA receptors and nitric oxide ameliorate constrictive collagen remodeling in hyperhomocysteinemia. *J Cell Physiol* 205:422–427
65. Shen L, Ji HF (2015) Associations between homocysteine, folic acid, Vitamin B₁₂ and Alzheimer's disease: insights from meta-analysis. *J Alzheimers Dis* 46:777–790
66. Sigmund CD (2010) Endothelial vascular muscle PPAR γ in arterial pressure regulation: lessons from genetic interference and deficiency. *Hypertension* 55:437–444
67. Stamler JS, Loscalzo J (1992) Endothelium-derived relaxing factor modulates atherothrombotic effects of homocysteine. *J Cardiovasc Pharmacol* 20(Suppl 12):S202–S204
68. Stamler JS, Osborne JA, Jaraki O, Robbani LE, Mullins M, Singel D, Loscalzo J (1993) Adverse vascular effects of homocysteine are modulated by endothelium-derived relaxing factor and related oxides of nitrogen. *J Clin Invest* 91:308–318
69. Stampfer MJ, Malinow MR, Willett WC, Newcomer LM, Upson B, Ullmann D, Tishler PV, Hennekens CH (1992) A prospective study of plasma homocyst(e)ine and risk of myocardial infarction in US physicians. *JAMA* 268:877–881
70. Stühlinger MC, Tsao PS, Her JH, Kimoto M, Balint RF, Cooke JP (2001) Homocysteine impairs the nitric oxide synthase pathway: role of asymmetric dimethylarginine. *Circulation* 104:2569–2575
71. Suematsu N, Ojaimi C, Kinugawa S, Wang Z, Xu X, Koller A, Recchia FA, Hintze TH (2007) Hyperhomocysteinemia alters cardiac substrate metabolism by impairing nitric oxide bioavailability through oxidative stress. *Circulation* 115:255–262
72. Sydow K, Schwedhelm E, Arakawa N, Bode-Bögar SM, Tsikas D, Hornig B, Frölich JC BRH (2003) ADMA and oxidative stress are responsible for endothelial dysfunction in hyperhomocyst(e)inemia: effects of L-arginine and B vitamins. *Cardiovasc Res* 57:244–252
73. Taddei S, Virdis A, Ghiadoni L, Sudano I, Salvetti A (2001) Endothelial dysfunction in hypertension. *J Cardiovasc Pharmacol* 38(Suppl 2):S11–S14
74. Tawakol A, Forgiione MA, Stuehlinger M, Alpert NM, Cooke JP, Loscalzo J, Fishman AJ, Creager MA, Gewirtz H (2002) Homocysteine impairs coronary microvascular dilator function in humans. *J Am Coll Cardiol* 40:1051–1058
75. Toda N, Ayajiki K (2014) Okamura T (2014) Obesity-induced cerebral hypoperfusion derived from endothelial dysfunction: one of the risk factors for Alzheimer's disease. *Curr Alzheimer Res* 11: 733–744
76. Toda N, Okamura T (2012) Cerebral blood flow regulation by nitric oxide in Alzheimer's disease. *J Alzheimer Dis* 32:569–578

77. Toda N, Okamura T (2015) Recent advances in research on nitrenergic-nerve mediated vasodilatation. *Pflugers Arch Eur J Physiol* 467:1165–1178
78. Topal G, Brunet A, Millanvoye E, Boucher JL, Rendu F, Devynck MA, Devid-Dyfillo M (2004) Homocysteine induces oxidative stress by uncoupling of NO synthase activity through reduction of tetrahydrobiopterin. *Free Radic Biol Med* 36:1532–1541
79. Tsuda K, Nishio I (2004) Serum homocysteine and endothelial dysfunction in circulation disorders in women. *Circulation* 120, e37
80. Tyagi N, Kandel M, Munjal C, Qipshidze N, Vacek JC, Pushpakumar SB, Metreveli N, Tyagi SC (2011) Homocysteine mediated decrease in bone blood flow and remodeling: role of folic acid. *J Orthop Res* 29:1511–1516
81. Tyagi N, Sedoris KC, Steed M, Ovechkin V, Moshal KS, Tyagi SC (2005) Mechanisms of homocysteine-induced oxidative stress. *Am J Physiol* 289:H2649–H2656
82. Ungvari Z, Zsolt A, Bagi Z, Koller A (2002) Impaired nitric oxide-mediated flow-induced coronary dilation in hyperhomocysteinemia. *Am J Pathol* 161:145–153
83. Upchurch GR Jr, Welch GN, Loscalzo J (1996) Homocysteine, EDRF, and endothelial function. *J Nutr* 126(4 Suppl):1290S–1294S
84. Villa P, Bosco P, Ferri R, Perric C, Suriano R, Costantini B, Macri F, Proto C, Cento RM, Lanzone A (2009) Fasting and post-methionine homocysteine levels in Alzheimers disease. *Int J Vitam Nutr Res* 79:166–172
85. Weiss N, Heydrick SJ, Postea O, Keller C, Keaney JF, Loscalzo J (2003) Influence of hyperhomocysteinemia on the cellular redox state—impact on homocysteine-induced endothelial dysfunction. *Clin Chem Lab Med* 41:1455–1461
86. Weiss N, Ide N, Abahji T, Nill L, Keller C, Hoffmann U (2006) Aged garlic extract improves homocysteine-induced endothelial dysfunction in macro- and microcirculation. *J Nutr* 136(3 Suppl): 750S–754S
87. Weiss N, Keller C, Hoffmann U, Loscalzo J (2002) Endothelial dysfunction and atherothrombosis in mild hyperhomocysteinemia. *Vasc Med* 7:227–239
88. Willems FF, Aengevaeren WR, Boers GH, Blom HJ, Verheugt FW (2002) Coronary endothelial function in hyperhomocysteinemia: improvement after treatment with folic acid and cobalamin in patients with coronary artery disease. *J Am Coll Cardiol* 40:766–772
89. Woo KS, Chook P, Lolin YI, Cheung AS, Chan LT, Sun YY, Sanderson JE, Metreveli C, Celermajer DS (1997) Hyperhomocyst(e)inemia is a risk factor for arterial endothelial dysfunction in humans. *Circulation* 96:2542–2544
90. Zhang F, Slungaard A, Vercellotti GM, Iadecola C (1998) Superoxide-dependen cerebrovascular effects of homocysteine. *Am J Physiol* 274:R1704–R1711
91. Zheng Z, Wang J, Yi L, Yu H, Kong L, Cui W, Chen H, Wang C (2014) Correlation between behavioural and psychological symptoms of Alzheimer type dementia and plasma homocysteine concentration. *Biomed Res Int* 2014, 383494. doi:10.1155/2014]383494
92. Zylberstein DE, Bengtsson C, Björkelund C, Landaas S, Sundh V, Thelle D, Lissner L (2004) Serum homocysteine in relation to mortality and morbidity from coronary heart disease: a 24-year follow-up of the population study of women in Gothenburg. *Circulation* 109:601–606