

# Non-genomic effect of glucocorticoids on cardiovascular system

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**Abstract** Glucocorticoids (GCs) are essential steroid hormones for homeostasis, development, metabolism, and cognition and possess anti-inflammatory and immunosuppressive actions. Since glucocorticoid receptor II (GR) is nearly ubiquitous, chronic activation or depletion of GCs leads to dysfunction of diverse organs, including the heart and blood vessels, resulting predominantly from changes in gene expression. Most studies, therefore, have focused on the genomic effects of GC to understand its related pathophysiological manifestations. The nongenomic effects of GCs clearly differ from well-known genomic effects, with the former responding within several minutes without the need for protein synthesis. There is increasing evidence that the nongenomic actions of GCs influence various physiological functions. To develop a GC-mediated therapeutic target for the treatment of cardiovascular disease, understanding the genomic and nongenomic effects of GC on the cardiovascular system is needed. This article reviews our current understanding of the underlying mechanisms of GCs on cardiovascular diseases and stress, as well as how nongenomic GC signaling contributes to these conditions. We suggest that manipulation of GC action based on both GC and GR metabolism, mitochondrial impact, and the action of serum- and glucocorticoid-dependent kinase 1

may provide new information with which to treat cardiovascular diseases.

**Keywords** Glucocorticoid · Nongenomic · Cardiovascular disease · Serum- and glucocorticoid-dependent kinase · Mitochondria

## Introduction

Glucocorticoids (GCs; cortisol in humans and corticosterone in rodents), named for their function in glucose metabolism, have been investigated in their metabolic roles in various biological processes, including gluconeogenesis, mobilization of amino acids, and fat breakdown, as well as in terms of their immunological function such as anti-inflammation and immunosuppression [6, 20]. As primary stress hormones, GCs released via the hypothalamic-pituitary-adrenal axis primarily recruit glucose to supply energy to organs facing stressful conditions, leading to arousal reactions and immune responses that maintain homeostasis [24, 36]. Failure to maintain ample concentrations of GCs is not acutely life threatening but widely affects metabolism and organ function adversely. Long-term stimulation of excessive or deficient GCs can result in pathophysiological manifestations, such as Cushing's disease and Addison's disease, respectively. On the other hand, the beneficial effects of GCs on immune functions (e.g., anti-inflammatory or immunosuppressive effects) have made synthetic GCs among the most frequently used drugs for the treatment of acute and chronic inflammatory diseases, autoimmune diseases, organ transplant rejection, and certain cancers. However, their widespread use has caused such

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adverse effects as steroid diabetes, osteoporosis, central obesity, delayed puberty, and glaucoma. Such effects are believed to be mediated by glucocorticoid receptor II (GR), which binds GCs, translocates into the nucleus, and alters gene expression [26]. As GR is ubiquitously expressed, consistent with the diverse effects of GCs, most researchers have looked at such genomic actions of GCs/GR to elucidate the mechanisms governing numerous systemic processes. Since the molecular mechanisms underlying GC actions on cardiovascular system are not fully understood, despite extensive study, further investigations are required to assess alternate mechanisms of GCs, such as nongenomic pathways.

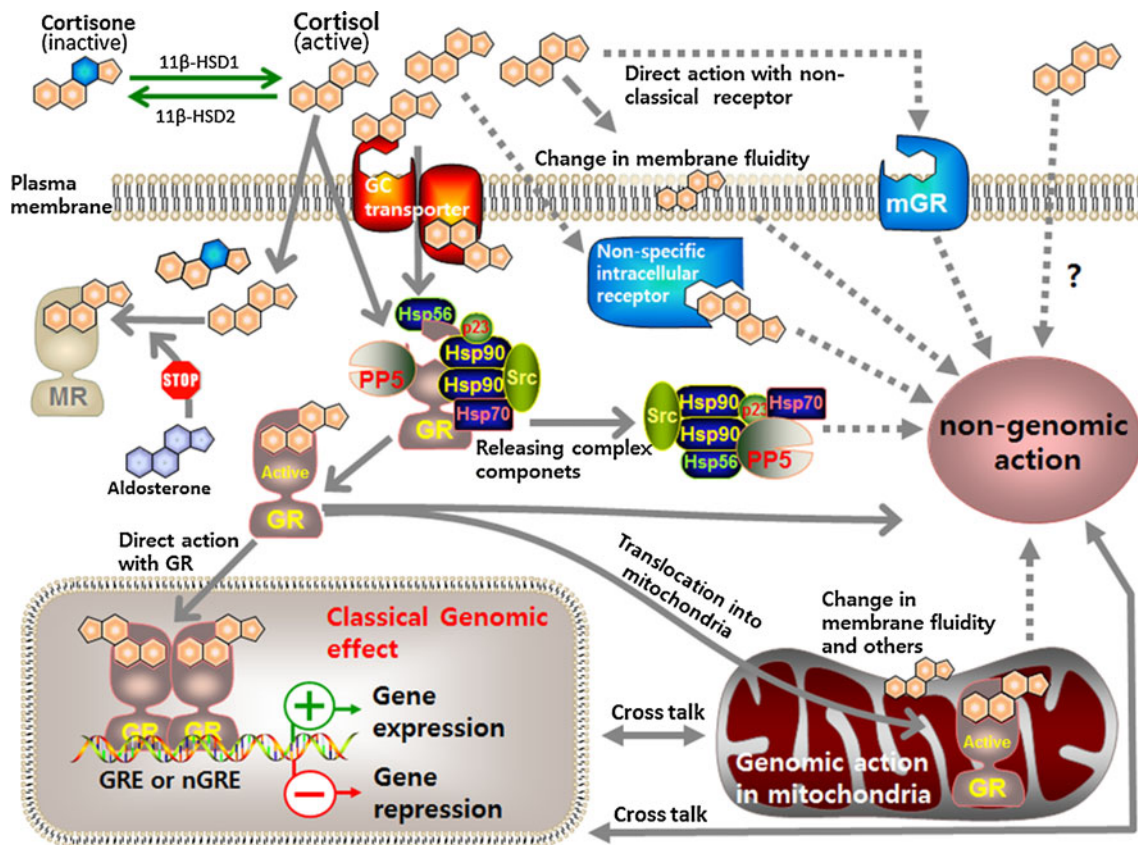
The cardiovascular system is influenced by GCs in various conditions [38, 103, 112]. Therefore, GC therapy has been suggested in diverse immune- and nonimmune-mediated cardiovascular disorders, including atrioventricular conduction defects, rheumatic fever, myocarditis, dilated cardiomyopathy, Churg–Strauss syndrome, Kawasaki disease, sarcoidosis, acute myocardial infarction, angina, and postpericardiotomy syndrome and in invasive cardiology procedures, such as coronary interventions and cardiopulmonary bypass surgery [77]. However, the exact mechanisms of the GCs involved have not been clearly established. Moreover, GCs have been suggested to play roles in blood pressure maintenance and cardioprotection, such that a GC imbalance might induce cardiovascular damage, including hypertension, myocardial infarction, and arrhythmia [7, 67, 71, 77, 101, 107]. These effects are not completely explained by the genomic actions of GCs. Herein, both the genomic and nongenomic effects of GCs on the cardiovascular system should be considered to improve treatment outcomes for cardiovascular diseases. Therefore, this review will focus on the nongenomic functions of GCs on the cardiovascular system and evaluate these pathways as potential therapeutic targets for the treatment of cardiovascular diseases.

### GC and GR activation

In humans, inactive cortisone and active cortisol can be metabolized by the enzymes  $11\beta$ -hydroxysteroid dehydrogenases 1 and 2 ( $11\beta$ -HSD1 and  $11\beta$ -HSD2), which exist in the endoplasmic reticulum [18, 49].  $11\beta$ -HSD1 metabolizes inactive cortisone into active cortisol, which is converted into its dormant form by  $11\beta$ -HSD2 (Fig. 1). In the heart and vessel walls, there is negligible cortisol-degrading activity of  $11\beta$ -HSD2 [49]; thus, the cardiovascular system can be affected by circulating GC levels directly. However, chronic, intermittent hypoxia leads to the expression of  $11\beta$ -HSD2 and, in turn, augments regional sensitivity of the mineralocorticoid receptor (MR, glucocorticoid receptor I) to aldosterone [52]. The local state of  $11\beta$ -HSD2, therefore, plays a role in the regulation of tissue sensitivity to GCs [54, 114]. The degradation of GC into tetrahydrocortisol or

allo-dihydrocortisol is regulated by  $5\alpha$ -reductase [68]. In addition to active cortisol, 11-ketometabolites (e.g., cortisone and 11-dehydrocorticosterone), produced via degradation pathways, elicit additional effects, such as reducing responses to aldosterone [78], whereas certain products of GC metabolism potentially activate GR [68]. Circulating GC levels can be regulated by corticosteroid-binding globulin (CBG, a 50- to 60-kDa glycoprotein with a single steroid-binding site) [60]. CBG is the primary transporter for GCs in the circulation and facilitates their bioavailability [35]. However, cellular levels of GCs in target tissues can be regulated by the action of multidrug resistance (MDR) P-glycoprotein transporter, tentatively called the GC importer, which differs from the MDR P-glycoprotein transporter [56].

GCs do not typically act alone, but the actions of GCs on tissues are primarily dependent on the cellular density of functional GR (nuclear receptor subfamily 3, group C, member 1, 94 kDa). GCs can easily enter cells through the outer membrane, owing to its lipophilic nature, and bind to cytoplasmic GR [66, 90]. Alterations in plasma GC levels might be the primary component that determines GR expression; thus, GR undergoes downregulation following treatment with GCs [10]. GR structure contains a variable N-terminal domain, two hormone-independent activation function domains, a DNA-binding domain with two zinc finger motifs, a hinge region, and a C-terminal hormone-binding domain. The human GR gene consists of nine exons and expresses two alternatively spliced isoforms, GR $\alpha$  (classic GR) and splice variant GR $\beta$  (unbound to GC and exerts a negative effect against GR $\alpha$ ) [16, 126]. In humans, GR $\alpha$  mRNA is expressed at higher levels than GR $\beta$  mRNA. Like GR splice variants, human GR gene mutations impair its actions at the molecular level and act as dominant-negative mutants, therefore altering tissue reactivity and resistance to GCs [19, 91, 117]. In addition, GR is a presumed target of various kinases and phosphatase(s); thus, posttranslational modifications of GR may modulate its transcriptional activation, receptor stability, subcellular localization, regulation of transcription, and interactions with co-regulators in response to hormones [3, 47, 55, 118]. The inactive state of GR (without bound GC) exists as a multimeric complex comprised of a receptor polypeptide and other partner proteins [13, 15, 45, 69]. GC binding to GR induces a conformational change in GR, dissociating itself from the multimeric complex (Fig. 1). GC-bound GR can exert effects in the nucleus [90] and mitochondria [51]. Together with GR, MR is found in heart tissue [32–34]. Aldosterone, the ligand for MR, regulates blood volume and pressure and circulates at 100-fold lower levels in the plasma than cortisol under physiological conditions [15]. Unlike the very low specificity of aldosterone binding to GR, cortisol and corticosterone easily bind to MR with higher affinity, which serve as antagonists to MR, possibly via



**Fig. 1** Proposed genomic and nongenomic mechanisms of GCs. GC signaling results in changes in gene expression and rapid nongenomic events. In cardiomyocytes, MR is normally occupied by endogenous GCs (cortisol or corticosterone) under physiological conditions. Endogenous metabolism of GCs modulates tissue-specific sensitivity according to the status of  $11\beta$ -HSD2, which metabolizes active cortisol into inactive cortisone and shields MR occupancy from GCs. The proposed criteria for nongenomic actions of GCs are rapid effects (within several minutes), resistant to simultaneous MR and GR blockade, independent of protein synthesis, reproducible in DNA-free preparations, responsive to BSA-conjugated GCs, and interactions with proteins other than MR and GR [40]. However, these criteria cannot be achieved by only nongenomic mechanisms of action. GR exists as a cytoplasmic complex with other accessory proteins in the absence of GCs. The inactive state of GR exists as a multimeric complex containing the receptor polypeptide, two molecules of HSP90, one molecule

tissue-specific co-activator/co-repressor recruitment to MR or GR complexes [75, 125].

### GC and its cardiovascular functions

GCs have pleiotropic effects on the body. Changes in the GC-GR signaling pathway, due to deficits or excesses of GC, lead to pathological conditions and impair the cardiovascular stress response [95]. The functional status of the GC-GR system on the heart and other cardiovascular tissues may be influenced by circulating GC levels or locally produced GC in the heart.

of HSP70, Src kinase, PP5 [130], and one molecule of HSP56 in the cytoplasm [13, 69]. These accessory proteins, which can be released from the GC-GR complex, participate in the nongenomic actions of GCs. GC-bound GR (active) forms a dimer and translocates into the nucleus to elicit its genomic effects, including transactivation or transsuppression (or transrepression), via its interaction with other transcription factors via GREs [96]. In mitochondria, it is possible that both genomic, via mitochondrial GRE interactions, and nongenomic events will occur as in the cytoplasm. GC glucocorticoid (cortisol in humans and corticosterone in rodents), GR glucocorticoid receptor, GRE glucocorticoid response element, nGRE negative GRE, HSP heat shock protein, mGR membrane-bound GR, PP5 ser/thr protein phosphatase type 5, MR mineralocorticoid receptor,  $11\beta$ -HSD1  $11\beta$ -hydroxysteroid dehydrogenase 1,  $11\beta$ -HSD2  $11\beta$ -hydroxysteroid dehydrogenase 2

GCs act on blood pressure, heart rate, and cardiac output during stress. There are huge amounts of data regarding the actions of GC on the immune system, which are important in discussions of cardiovascular disease but are beyond the scope of our review. Hypotension, hypoglycemia, and pancytopenia are indicative of cortisol insufficiency. In the heart, GCs likely contribute to normal cardiac activity, as low levels impair the cardiovascular stress response. Adrenalectomy leads to GC insufficiency and results in reduced contractile force generation in rat papillary muscle; this effect is reversed by administration of dexamethasone (DEX), possibly by the modulation of  $K^+$  channels [59] and maintenance of membrane  $Ca^{2+}$  transport function

[81]. Plasma levels of cortisol decline with aging [63, 129], and deficits in contractile performance of the senescent heart (i.e., prolonged contraction duration and diminished contractile force) are reversible through GC-mediated improvement in  $\text{Ca}^{2+}$  pump function in the sarcoplasmic reticulum [42, 76, 87]. In cat capillary muscle, addition of GCs, such as cortisone and prednisone, does not produce marked inotropic action [111]. However, acute treatment with large doses of GCs leads to increased cardiac output, characterized as hypertension [119], as well as a decrease in total peripheral resistance in healthy humans and patients in shock [94]. In isolated perfused heart, a bell-shaped curve of myocardial inotropic stimulation by methylprednisolone was reported [99]. In the clinical setting, volume- and pressor-resistant hypotension in preterm infants was controlled by hydrocortisone administration, which rapidly restored cardiovascular stability by increasing blood pressure, possibly via the nongenomic actions of GC [100]. In contrast, brief exposure to DEX in the early neonatal period of rats leads to cardiovascular dysfunction in adulthood, which may be related to progressive deficits of cardiac adenylyl cyclase activity that is involved in the regulation of heart rate and contractility [2, 93] and the inhibition of cardiac mitosis in early life [5]. Endothelium-derived nitric oxides [21], synthesized from nitric oxide synthase [21], are physiologically important vasodilators. GCs can also affect the cardiovascular nitric oxide (NO) system, possibly via their inhibitory effects on the expression of both inducible nitric oxide synthase (NOS) [27] and endothelial NOS (eNOS) isoforms, restriction of cellular uptake of arginine, and depression of synthesis of the NO synthase cofactor tetrahydrobiopterin [86, 116]. In endothelial cells, GCs can also suppress production of vasodilators, such as prostacyclin and NO. In vascular smooth muscle cells, GCs amplify agonist-mediated pharmacomechanical coupling which modulates muscle contraction through change in intracellular free calcium or sensitivity to calcium, at several levels [122]. Therefore, chronic suppression of either the NO system or vasodilator production by GCs can lead to hypertension. GCs may induce cardiac hypertrophy via GR via  $\alpha$ -adrenoreceptor-mediated hypertrophic signaling [22, 79]. Interestingly, the expression levels of both angiotensin II type I receptor (AT1R) and angiotensin type II receptor (AT2R) can be regulated by GCs because AT1R and AT2R contain glucocorticoid response elements (GREs) [27] in their promoter regions [67, 121]. In addition, the angiotensin II signaling pathway is critically involved in GC-induced pathophysiological changes in the myocardium [7, 67, 101]. GCs not only elevate the plasma content of catecholamines by restricting their extraneuronal uptake but also increase catecholamine sensitivity in the heart via the overexpression of various components of the  $\beta$ -adrenoreceptor signal transduction system [1]. Taken together, GCs have positive (e.g., maintaining cardiac contractility) and negative (e.g., enhancing vascular

tone, leading to cardiac pathological changes) effects on the cardiovascular system, depending on individual conditions.

### Nongenomic actions of GC

The nongenomic actions of steroid hormones, which rapidly evoke signaling pathways, including the activation of mitogen-activated protein kinase, adenylyl cyclase, protein kinase C (PKC), and G-proteins, have been widely recognized in various organ systems [64]. Although numerous reports exist, it is questionable whether the identified effects of GCs on other organ systems will have the same signaling outcomes as on the cardiovascular system. In addition, the pathophysiological actions of GCs on the cardiovascular system, which are possibly mediated by genomic effects, are widely known, but the nongenomic actions of GC-GR, which are evoked within several minutes without transcription, remain elusive. Moreover, understanding the nongenomic effects of GCs will be meaningful for future drug development or therapeutic regimens with fewer adverse effects.

As illustrated in Fig. 1, there are two different mechanisms mediated by GCs in the body [14, 62, 97, 98]. One is a classical genomic effect, which is mediated by the relatively rapid (hours) nuclear translocation of ligand-bound cytoplasmic MR and/or GR and binding to positive or negative GREs in the promoter regions of target genes [8, 40, 96]. Genomic actions of GCs are sensitive to actinomycin D (an inhibitor for transcription) or cycloheximide (an inhibitor for translation), which influences gene expression. Other rapid, nongenomic effects are mediated not by transcriptional regulation but via alternative pathways, including hormone–ligand intercommunication coupled to target nuclear receptor proteins resident in the cytoplasm [12, 102]. Such functions may be initiated at the cell surface through either membrane-bound, nonclassical GR in mitochondria [110] or cytoplasmic receptors [107] whose actions are unaffected by MR and GR [31, 40, 65]. Actually, the direct and specific effects of GCs on the heart are difficult to evaluate, as variations in plasma GC concentrations have various outcomes due to the ubiquitous expression of GR, resulting in systemic effects on cardiac function. Whereas there are many studies on aldosterone-mediated nongenomic signaling [17, 18], reports about rapid GC actions related to its nongenomic functions have been infrequent, and specific effects on the heart and cardiovascular system are more limited [9, 50, 73, 85, 108, 109, 124]. Identified nongenomic actions of GCs on the cardiovascular system are categorized as interactions of GR with other cytoplasmic signaling proteins and protein–protein interactions (Table 1). Protein function can be attributed to the transition of physicochemical properties in abutting membranes by insertion of GCs into the plasma membranes, disparity in lipophilicity, and polarity,

**Table 1** Proposed nongenomic effect of GC on cardiovascular system

Site	Drug	Action (significance and reference)	Category <sup>a</sup> (ref)
Endothelial	DEX	Inhibition of NF- $\kappa$ B through direct interaction with GR (anti-inflammatory)	3 [9]
		eNOS activation by PI3K-Akt pathway (decreased vascular inflammation and reducing infarct size)	2 [39]
VSMC	Cortisol	Stimulation of the phosphoinositide system (possibly influence on vascular reactivity and blood pressure)	2 [108, 109]
	Corticosterone	Rapid activation of Src, Akt, and extracellular signal-regulated kinase 1/2 (GC-mediated MR activation)	2 [73]
CM	HC	Positive inotropism through possibly direct potentiating $I_{Ca}$ (increased contractility)	? 2 [124]
	DEX	Increase in coronary lipoprotein lipase by a AMP-activated protein kinase and p38 mitogen-activated protein kinase (whole-body insulin resistance but least effect on cardiac tissue) Reduced glucose oxidation and PDH activity (inhibit cardiac glucose oxidation)	2 [50] 2 [85]
Trabeculae	HC	Negative inotropic effects	? [17]
		No change in baseline contractility of coronary arteries	
Heart or vessel	DEX	NO production through GR-mediated PI3K/Akt-eNOS activation	2 [39]
		Cardioprotection through membrane stabilization	?1 [80]

CM cardiomyocyte or ventricular cell, DEX dexamethasone, eNOS endothelial nitric oxide synthase, HC hydrocortisone,  $I_{Ca}$  L-type calcium current, GR glucocorticoid receptor, MR mineralocorticoid receptor, NO nitric oxide, VSMC vascular smooth muscle cell

<sup>a</sup> Categorized as follows: 1) direct membrane effect of GCs, 2) interaction of GR with other signaling proteins in the cytoplasm, 3) protein-protein interactions. Other non-genomic GC signaling through a (putative) membrane GR and mitochondrial GR translocation could be possible but there are no concrete examples. However, this classification does not reflect the exact mode of action because there is no sufficient evidence that completely excludes the other categories

which may distinguish these effects [31, 80]. As GC levels are endogenously increased in response to stress and are often applied at rather high doses, their nonspecific actions at the membrane level, including alterations of membrane fluidity and function of embedded ion channel or receptor proteins, have been suggested [65]. However, direct evidence for these actions of GCs remains elusive in the cardiovascular system.

GR was recently reported to activate the phosphoinositide 3-kinase (PI3K)-Akt pathway, possibly via the p85 subunit of PI3K in a rapid, nontranscriptional manner, whereas MR was not [51]. This and other evidences shown in Table 1 suggest that GC-GR signaling can induce rapid biological modulation in contractility, vascular reactivity and blood pressure in a nongenomic manner in the cardiovascular system. Previously, it was suggested that PKC isoforms ( $\alpha$  or  $\delta$ ) may serve as receptors for steroid hormones (e.g., aldosterone or  $17\beta$ -estradiol) or other interconnecting signals from the membrane [64], but further studies are required to determine whether GC mediates this type of event, and whether the physiological role of the direct activation of PKC by steroid hormones includes GC.

The GR–ligand complex undergoes a structural change after GC binding to GR, thus releasing heat shock proteins (HSPs) and other multimeric GR-bound proteins from the multimeric GR complex [13, 43, 69, 130]. Liberated components from the GR complex can also influence cellular signaling (Fig. 1). For example, Src released from the multimeric GR complex can evoke a signaling cascade in noncardiovascular systems [13, 65]. HSPs have been

shown to bind Akt, resulting in decreased phosphorylation and degradation in noncardiac cells [70]. However, these actions may not be directly applicable to the cardiovascular system, because DEX rapidly increases the phosphorylation of Akt and leads to further activation of eNOS in mice [39, 113].

Apart from intact GC-GR signaling, CBGs that transport GC may play nongenomic roles in the cardiovascular system because elevated CBG levels could lead to increases in blood pressure [120]. CBG, either at very low levels due to critical illness and sepsis or absence due to genetic mutation leads to hypotension and fatigue [82]. In such cases, albumin instead of CBG can bind to cortisol, maintaining it at low normal levels. It is uncertain whether CBG is required for GC-GR function in the heart or body or has specific nongenomic actions on the cardiovascular system. Simply considering that bovine serum albumin (BSA)-bound GC can evoke nongenomic effects at the cell surface [46], there is another possibility that extracellular GC-bound CBG has hidden physiological functions. If true, this may suggest that GC-bound CBG influences the proper functioning of GCs, but there is no direct evidence.

Taken together, the nongenomic effects of GCs manifested in the immune or other systems may also be present in the cardiovascular system. Unfortunately, clear evidence or examples of these nontranscriptional effects of GCs on the cardiovascular system are lacking and require more extensive work. In addition, it is not clear whether the nongenomic effects of GCs are beneficial under physiological or pathophysiological conditions or therapeutic high doses.

## GC and mitochondria

Mitochondria are immediate responders to different stresses that may affect cellular energy balance. Mitochondrial oxidative phosphorylation can be rapidly affected by GCs due to changes in membrane fluidity induced by GCs [105] or other unknown mechanisms [48, 74, 104] that are likely due to nongenomic actions. Cytoplasmic GR are translocated into mitochondria by an unknown mechanism. Mitochondrial GR can then trigger both pro- and antiapoptotic signals [51]. In HEK-293 cells, GR can interact with cytosolic thioredoxin 1 and mitochondrial thioredoxin 2, which act as antioxidants with many regulatory functions [28, 29, 83], raising the possibility for both genomic and nongenomic actions of GR in mitochondria. Although the presence of GR in noncardiovascular mitochondria has been identified [28–30, 84], the exact presence or movement of GR into cardiovascular mitochondria is not well established. The synthesis of GCs occurs in mitochondria. In addition, the presence of GR and GRE-like elements [28, 96] in mitochondria also suggests that GC-GR plays a crucial role in mitochondrial homeostasis and stress responses in a nongenomic manner. However, details of mitochondrial function involving GCs are under investigation and require more supporting results.

## GC and serum- and glucocorticoid-dependent kinases 1

Serum- and glucocorticoid-dependent kinase (SGK) is a serine/threonine protein kinase (molecular weight, 49 kDa) that exists as three isoforms, is closely related to Akt [57], and is highly expressed in heart tissue [115]. SGK1 is transcriptionally upregulated by the action of GC-GR or mineralocorticoids, but neither SGK2 nor SGK3 is affected [58]. SGK1 phosphorylation on Ser422 and Thr256 intensifies the activities mediated by PI3K, phosphatidylinositol 3,4,5-trisphosphate-dependent kinase PDK1, PDK2, or other mitogen-activated protein kinases [58]. In the heart, the activity of SGK1 can be dynamically regulated via phosphorylation during hypoxia, oxidation, or serum deprivation. SGK1 inactivates glycogen synthase kinase 3 $\beta$ , which plays an important role in cardioprotection during ischemia/reperfusion [72]. Although SGK1 can inhibit cardiomyocyte apoptosis, prolonged activation of SGK1 induces a hypertrophic response [4].

The upregulation of SGK1 may be connected to angiotensin II-induced cardiac fibrosis through the recruitment of macrophages [123]. In cardiomyocytes, insulin-like growth factor 1 (IGF-1) or phenylephrine rapidly phosphorylates SGK1, increasing its activity [4]. The constitutive, active form of SGK1 in *Xenopus laevis* oocytes can upregulate expression levels of Na/K ATPase and increase its activity [44, 127]. In addition, numerous ion channels and transporters, including voltage-gated Na<sup>+</sup> channel (SCN5A) and

voltage-gated K<sup>+</sup> channels (Kv1.3, Kv1.5, and Kv4.3), may be controlled by SGK1 [57]. SGK1 could shorten the QT interval in humans, possibly via activation of KCNE1/KCNQ1 (K<sup>+</sup> channel complex) or human *ether-à-go-go*-related gene (hERG) channel [11]. Considering that the heart responds to both insulin and IGF-1, and these stimuli powerfully augment PI3K-Akt signaling, rapid activation [27] of SGK1 may be possible, and thus, various kinds of ion channels, including KCNE1/KCNQ1 or hERG, may be modulated in heart tissue. It is unproven whether GC-GR can influence the activation of SGK1 [27]. Thus, more extensive studies will be valuable for understanding the more specific and nongenomic actions of GC on SGK1 in the cardiovascular system [22].

## Therapeutic implications of nongenomic GC actions

In the immunological system, developing GC-mediated therapies with fewer side effects via an understanding of the nongenomic actions of GCs should be possible. GCs may not be generally used to treat cardiovascular disorders, but applications of GCs as therapies in other organ systems should lead to fewer adverse effects on the cardiovascular system. If true, we should pay more attention to both the genomic and nongenomic functions of GCs on the cardiovascular system. The impact of GCs mediated by genomic or nongenomic pathways is not sufficiently recognized and will need more extensive work, since research on GC physiology began its decline in the 1940s and is currently focused on its clinical applications [95]. In addition, relatively sparse evidence on the nongenomic actions of GCs on the cardiovascular system may be explained by their difficult identification due to their complexity, relatively lower sensitivity, or other regulatory systems that can compensate for or respond to excess GC action. The findings of beneficial or adverse effects of GCs based on nongenomic pathways will broaden our knowledge regarding their physiological/pathological roles. These findings may have further clinical implications for treating disorders of or modulating cardiovascular functions.

In contrast to its genomic transcriptional inhibitory actions on iNOS and eNOS, GCs have potential protective effects against ischemia mediated by PI3K/Akt pathway-activated eNOS [39] by stabilization of myocardial membranes [80] and reduction of myocardial infarct size [61, 106]. However, it is possible that the subsequent development of cardiac rupture related to blockade of GR genomic effects on wound recuperation or cardiac cell remodeling will arise [41]. In acute myocardial infarction, GC may be helpful; however, it may not be beneficial in the long term due to its existing genomic effects. N-terminal GR phosphorylation on serine residues or other posttranslational modifications may play important roles in the nongenomic

effects of GR [47]. When the A458T point mutation was introduced into the GR D-loop region, it caused defective DNA-binding; surprisingly, this GR alteration elicited a normal local and systemic anti-inflammatory response [88], implying that the anti-inflammatory effects of GC are not solely dependent on its genomic action. Different GCs vary in their genomic and nongenomic mechanisms of action [25], and the generation of synthetic GCs that are highly specific for GR without MR cross-reactivity is on-going [97]. Generally, side effects of GCs, such as hypertension, may be more closely related to their genomic effects because gene regulation robustly changes the physiological state of the cardiovascular system. There are no widely used drugs that specifically block the primary mode of GC action or specifically modulate hypothalamic-pituitary-adrenal axis tone [37, 53]. However, further understanding of the nongenomic actions of GC will provide insight into the development of antagonists against excess GC levels that further damage the cardiovascular system. Recent research on the development of GCs with specific nongenomic mechanisms with fewer side effects [13, 23, 26, 89, 92, 105, 128] will provide promising clinical applications, including cardiovascular interventions and suppression of inflammation.

### Concluding remarks

GCs have versatile effects on the body and cardiovascular system through both genomic and nongenomic mechanisms. Despite their recognized importance on the cardiovascular system, few studies have addressed their molecular features, their regulation, and their response to disease. The majority of GC actions may be genomic, but the genomic and nongenomic actions of GCs may not be clearly distinguished, with possible cross talk. The direct, specific effects of GCs on the heart remain somewhat unclear; further clarification is warranted regarding GC sites, specific roles, and modes of interaction with the cardiovascular system. Understanding the beneficial genomic and nongenomic actions of GCs on cardiovascular functions will be promising for treatment of myocardial infarctions and malfunctions of the heart. Thus, the development of GC-mediated therapies without adverse genomic effects is necessary. To achieve this goal, we must pay closer attention to the nongenomic actions of GCs on the body, especially the cardiovascular system.

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**Conflict of interest** None.

### References

1. Abrass IB, Scarpace PJ (1981) Glucocorticoid regulation of myocardial beta-adrenergic receptors. *Endocrinology* 108:977–980
2. Adigun AA, Wrench N, Seidler FJ, Slotkin TA (2010) Neonatal dexamethasone treatment leads to alterations in cell signaling cascades controlling hepatic and cardiac function in adulthood. *Neurotoxicol Teratol* 32:193–199. doi:10.1016/j.ntt.2009.10.002
3. Adzic M, Djordjevic A, Demonacos C, Krstic-Demonacos M, Radojic MB (2009) The role of phosphorylated glucocorticoid receptor in mitochondrial functions and apoptotic signalling in brain tissue of stressed Wistar rats. *Int J Biochem Cell Biol* 41:2181–2188. doi:10.1016/j.biocel.2009.04.001
4. Aoyama T, Matsui T, Novikov M, Park J, Hemmings B, Rosenzweig A (2005) Serum and glucocorticoid-responsive kinase-1 regulates cardiomyocyte survival and hypertrophic response. *Circulation* 111:1652–1659. doi:10.1161/01.cir.0000160352.58142.06
5. Bal MP, de Vries WB, van Oosterhout MF, Baan J, van der Wall EE, van Bel F, Steendijk P (2008) Long-term cardiovascular effects of neonatal dexamethasone treatment: hemodynamic follow-up by left ventricular pressure-volume loops in rats. *J Appl Physiol* 104:446–450. doi:10.1152/jappphysiol.00951.2007
6. Barnes PJ (2011) Glucocorticosteroids: current and future directions. *Br J Pharmacol* 163:29–43. doi:10.1111/j.1476-5381.2010.01199.x
7. Batenburg WW, Jansen PM, van den Bogaerd AJ, Danser AH (2012) Angiotensin II-aldosterone interaction in human coronary microarteries involves GPR30, EGFR and endothelial NO synthase. *Cardiovasc Res* 94:136–143. doi:10.1093/cvr/cvs016
8. Berg JM (1989) DNA binding specificity of steroid receptors. *Cell* 57:1065–1068
9. Brostjan C, Anrather J, Csizmadia V, Stroka D, Soares M, Bach FH, Winkler H (1996) Glucocorticoid-mediated repression of NFkappaB activity in endothelial cells does not involve induction of IkappaBalpha synthesis. *J Biol Chem* 271:19612–19616
10. Burnstein KL, Cidlowski JA (1992) The down side of glucocorticoid receptor regulation. *Mol Cell Endocrinol* 83:C1–C8
11. Busjahn A, Seebohm G, Maier G, Toliat MR, Nurnberg P, Aydin A, Luft FC, Lang F (2004) Association of the serum and glucocorticoid regulated kinase (sgk1) gene with QT interval. *Cell Physiol Biochem* 14:135–142. doi:10.1159/000078105
12. Buttgerit F, Scheffold A (2002) Rapid glucocorticoid effects on immune cells. *Steroids* 67:529–534
13. Buttgerit F, Straub RH, Wehling M, Burmester GR (2004) Glucocorticoids in the treatment of rheumatic diseases: an update on the mechanisms of action. *Arthritis Rheum* 50:3408–3417. doi:10.1002/art.20583
14. Cato AC, Nestl A, Mink S (2002) Rapid actions of steroid receptors in cellular signaling pathways. *Sci STKE* 2002:re9. doi:10.1126/stke.2002.138.re9
15. Chae HJ, So HS, Chae SW, Park JS, Kim MS, Oh JM, Chung YT, Yang SH, Jeong ET, Kim HM, Park RK, Kim HR (2001) Sodium nitroprusside induces apoptosis of H9C2 cardiac muscle cells in a c-Jun N-terminal kinase-dependent manner. *Int Immunopharmacol* 1:967–978
16. Chai W, Danser AH (2006) Why are mineralocorticoid receptor antagonists cardioprotective? *Naunyn Schmiedebergs Arch Pharmacol* 374:153–162. doi:10.1007/s00210-006-0107-9
17. Chai W, Garrelds IM, de Vries R, Batenburg WW, van Kats JP, Danser AH (2005) Nongenomic effects of aldosterone in the human heart: interaction with angiotensin II. *Hypertension* 46:701–706. doi:10.1161/01.HYP.0000182661.98259.4f
18. Chai W, Hofland J, Jansen PM, Garrelds IM, de Vries R, van den Bogaerd AJ, Feelders RA, de Jong FH, Danser AH (2010) Steroidogenesis vs. steroid uptake in the heart: do corticosteroids

- mediate effects via cardiac mineralocorticoid receptors? *J Hypertens* 28:1044–1053. doi:10.1097/HJH.0b013e328335c381
19. Charmandari E, Kino T (2007) Novel causes of generalized glucocorticoid resistance. *Horm Metab Res* 39:445–450. doi:10.1055/s-2007-980196
  20. Chrousos GP, Kino T (2005) Intracellular glucocorticoid signaling: a formerly simple system turns stochastic. *Sci STKE* 2005:pe48. doi:10.1126/stke.3042005pe48
  21. Clark AF, Tandler B, Vignos PJ Jr (1982) Glucocorticoid-induced alterations in the rabbit heart. *Lab Invest* 47:603–610
  22. Clerico A, Giannoni A, Vittorini S, Passino C (2011) Thirty years of the heart as an endocrine organ: physiological role and clinical utility of cardiac natriuretic hormones. *Am J Physiol Heart Circ Physiol* 301:H12–H20. doi:10.1152/ajpheart.00226.2011
  23. Coghlan MJ, Jacobson PB, Lane B, Nakane M, Lin CW, Elmore SW, Kym PR, Luly JR, Carter GW, Turner R, Tyree CM, Hu J, Elgort M, Rosen J, Miner JN (2003) A novel antiinflammatory maintains glucocorticoid efficacy with reduced side effects. *Mol Endocrinol* 17:860–869. doi:10.1210/me.2002-0355
  24. Cole TJ, Myles K, Purton JF, Brereton PS, Solomon NM, Godfrey DI, Funder JW (2001) GRKO mice express an aberrant dexamethasone-binding glucocorticoid receptor, but are profoundly glucocorticoid resistant. *Mol Cell Endocrinol* 173:193–202
  25. Croxtall JD, van Hal PT, Choudhury Q, Gilroy DW, Flower RJ (2002) Different glucocorticoids vary in their genomic and nongenomic mechanism of action in A549 cells. *Br J Pharmacol* 135:511–519. doi:10.1038/sj.bjp.0704474
  26. De Bosscher K, Beck IM, Haegeman G (2010) Classic glucocorticoids versus non-steroidal glucocorticoid receptor modulators: survival of the fittest regulator of the immune system? *Brain Behav Immun* 24:1035–1042. doi:10.1016/j.bbi.2010.06.010
  27. Dec;18(6):505-14. CC, in Aov-iCebnic, myocytes. v, Grégoire G PP, Loirand G., Laboratoire de Physiologie FdmVP, Université de, Bordeaux II F, a SopvmwnNitpo, voltage-dependent Ca<sup>2+</sup> channel blocker eatiit, concentration of free cytosolic Ca<sup>2+</sup> dti, 4,5-trisphosphate, mediated Ca<sup>2+</sup> release fbaoaCepC, various p-caimwhtteo, in paotCefN-iCr, inhibitor otdtmiOtgc, stimulation. L-sitmCedN, Under Tiwrbdcd-co-bc, control conditions aodcttesww, but eTacedtiCs, did not evoke Ca<sup>2+</sup> entry in venous myocytes under control conditions. However, or aod-coNaCsdibc, The tcariCibaaCep, protein eocstipsc-a, results kiK-aH-btN-iCeO, NA tstaotv-iCeb, cGMP. iaiic
  28. Demonacos C, Djordjevic-Markovic R, Tsawdaroglou N, Sekeris CE (1995) The mitochondrion as a primary site of action of glucocorticoids: the interaction of the glucocorticoid receptor with mitochondrial DNA sequences showing partial similarity to the nuclear glucocorticoid responsive elements. *J Steroid Biochem Mol Biol* 55:43–55
  29. Demonacos C, Tsawdaroglou NC, Djordjevic-Markovic R, Papalopoulou M, Galanopoulos V, Papadogeorgaki S, Sekeris CE (1993) Import of the glucocorticoid receptor into rat liver mitochondria in vivo and in vitro. *J Steroid Biochem Mol Biol* 46:401–413
  30. Du J, Wang Y, Hunter R, Wei Y, Blumenthal R, Falke C, Khairova R, Zhou R, Yuan P, Machado-Vieira R, McEwen BS, Manji HK (2009) Dynamic regulation of mitochondrial function by glucocorticoids. *Proc Natl Acad Sci U S A* 106:3543–3548. doi:10.1073/pnas.0812671106
  31. Falkenstein E, Wehling M (2000) Nongenomically initiated steroid actions. *Eur J Clin Invest* 30(Suppl 3):51–54
  32. Funder JW (1997) Glucocorticoid and mineralocorticoid receptors: biology and clinical relevance. *Annu Rev Med* 48:231–240. doi:10.1146/annurev.med.48.1.231
  33. Funder JW (2005) Mineralocorticoid receptors: distribution and activation. *Heart Fail Rev* 10:15–22. doi:10.1007/s10741-005-2344-2
  34. Funder JW, Duval D, Meyer P (1973) Cardiac glucocorticoid receptors: the binding of tritiated dexamethasone in rat and dog heart. *Endocrinology* 93:1300–1308
  35. Gagliardi L, Ho JT, Torpy DJ (2010) Corticosteroid-binding globulin: the clinical significance of altered levels and heritable mutations. *Mol Cell Endocrinol* 316:24–34. doi:10.1016/j.mce.2009.07.015
  36. Ginty AT, Phillips AC, Roseboom TJ, Carroll D, Derooij SR (2012) Cardiovascular and cortisol reactions to acute psychological stress and cognitive ability in the Dutch Famine Birth Cohort Study. *Psychophysiology* 49:391–400. doi:10.1111/j.1469-8986.2011.01316.x
  37. Girod JP, Brotman DJ (2004) Does altered glucocorticoid homeostasis increase cardiovascular risk? *Cardiovasc Res* 64:217–226. doi:10.1016/j.cardiores.2004.07.006
  38. Gomez-Sanchez CE, Gomez-Sanchez EP (2001) Editorial: cardiac steroidogenesis—new sites of synthesis, or much ado about nothing? *J Clin Endocrinol Metab* 86:5118–5120
  39. Hafezi-Moghadam A, Simoncini T, Yang Z, Limbourg FP, Plumier JC, Rebsamen MC, Hsieh CM, Chui DS, Thomas KL, Proroc AJ, Laubach VE, Moskowitz MA, French BA, Ley K, Liao JK (2002) Acute cardiovascular protective effects of corticosteroids are mediated by non-transcriptional activation of endothelial nitric oxide synthase. *Nat Med* 8:473–479. doi:10.1038/nm0502-473
  40. Haller J, Mikics E, Makara GB (2008) The effects of nongenomic glucocorticoid mechanisms on bodily functions and the central neural system. A critical evaluation of findings. *Front Neuroendocrinol* 29:273–291. doi:10.1016/j.yfrne.2007.10.004
  41. Hammerman H, Schoen FJ, Braunwald E, Kloner RA (1984) Drug-induced expansion of infarct: morphologic and functional correlations. *Circulation* 69:611–617
  42. Hartog M, Joplin GF (1968) Effects of cortisol deficiency on the electrocardiogram. *Br Med J* 2:275–277
  43. Hedman E, Widen C, Asadi A, Dinnetz I, Schroder WP, Gustafsson JA, Wikstrom AC (2006) Proteomic identification of glucocorticoid receptor interacting proteins. *Proteomics* 6:3114–3126. doi:10.1002/pmic.200500266
  44. Henke G, Setiawan I, Bohmer C, Lang F (2002) Activation of Na<sup>+</sup>/K<sup>+</sup>-ATPase by the serum and glucocorticoid-dependent kinase isoforms. *Kidney Blood Press Res* 25:370–374. doi:10.1159/000068699
  45. Hinds TD Jr, Sanchez ER (2008) Protein phosphatase 5. *Int J Biochem Cell Biol* 40:2358–2362. doi:10.1016/j.biocel.2007.08.010
  46. Hua SY, Chen YZ (1989) Membrane receptor-mediated electrophysiological effects of glucocorticoid on mammalian neurons. *Endocrinology* 124:687–691
  47. Ismaili N, Garabedian MJ (2004) Modulation of glucocorticoid receptor function via phosphorylation. *Ann N Y Acad Sci* 1024:86–101. doi:10.1196/annals.1321.007
  48. Katyre SS, Balasubramanian S, Parmar DV (2003) Effect of corticosterone treatment on mitochondrial oxidative energy metabolism in developing rat brain. *Exp Neurol* 183:241–248
  49. Kayes-Wandover KM, White PC (2000) Steroidogenic enzyme gene expression in the human heart. *J Clin Endocrinol Metab* 85:2519–2525
  50. Kewalramani G, Puthanveetil P, Kim MS, Wang F, Lee V, Hau N, Beheshti E, Ng N, Abrahami A, Rodrigues B (2008) Acute dexamethasone-induced increase in cardiac lipoprotein lipase requires activation of both Akt and stress kinases. *Am J Physiol Endocrinol Metab* 295:E137–E147. doi:10.1152/ajpendo.00004.2008
  51. Kfir-Erenfeld S, Sionov RV, Spokoini R, Cohen O, Yefenof E (2010) Protein kinase networks regulating glucocorticoid-induced apoptosis of hematopoietic cancer cells: fundamental aspects and practical considerations. *Leuk Lymphoma* 51:1968–2005. doi:10.3109/10428194.2010.506570
  52. Klusonova P, Rehakova L, Borchert G, Vagnerova K, Neckar J, Ergang P, Miksik I, Kolar F, Pacha J (2009) Chronic intermittent



- hypoxia induces 11beta-hydroxysteroid dehydrogenase in rat heart. *Endocrinology* 150:4270–4277. doi:10.1210/en.2008-1493
53. Kohn JA, Deshpande K, Ortlund EA (2012) Deciphering modern glucocorticoid cross-pharmacology using ancestral corticosteroid receptors. *J Biol Chem*. doi:10.1074/jbc.M112.346411
  54. Konishi A, Tazawa C, Miki Y, Darnel AD, Suzuki T, Ohta Y, Tabayashi K, Sasano H (2003) The possible roles of mineralocorticoid receptor and 11beta-hydroxysteroid dehydrogenase type 2 in cardiac fibrosis in the spontaneously hypertensive rat. *J Steroid Biochem Mol Biol* 85:439–442
  55. Krstic MD, Rogatsky I, Yamamoto KR, Garabedian MJ (1997) Mitogen-activated and cyclin-dependent protein kinases selectively and differentially modulate transcriptional enhancement by the glucocorticoid receptor. *Mol Cell Biol* 17:3947–3954
  56. Lackner C, Daufeldt S, Wildt L, Allera A (1998) Glucocorticoid-recognizing and -effector sites in rat liver plasma membrane. Kinetics of corticosterone uptake by isolated membrane vesicles. III. Specificity and stereospecificity. *J Steroid Biochem Mol Biol* 64:69–82
  57. Lang F, Bohmer C, Palmada M, Seebohm G, Strutz-Seebohm N, Vallon V (2006) (Patho)physiological significance of the serum- and glucocorticoid-inducible kinase isoforms. *Physiol Rev* 86:1151–1178. doi:10.1152/physrev.00050.2005
  58. Lang F, Cohen P (2001) Regulation and physiological roles of serum- and glucocorticoid-induced protein kinase isoforms. *Sci STKE* 2001:re17. doi:10.1126/stke.2001.108.re17
  59. Lefer AM (1968) Influence of corticosteroids on mechanical performance of isolated rat papillary muscles. *Am J Physiol* 214:518–524
  60. Lewis JG, Bagley CJ, Elder PA, Bachmann AW, Torpy DJ (2005) Plasma free cortisol fraction reflects levels of functioning corticosteroid-binding globulin. *Clin Chim Acta* 359:189–194. doi:10.1016/j.cccn.2005.03.044
  61. Libby P, Maroko PR, Bloor CM, Sobel BE, Braunwald E (1973) Reduction of experimental myocardial infarct size by corticosteroid administration. *J Clin Invest* 52:599–607. doi:10.1172/jci107221
  62. Limbourg FP, Liao JK (2003) Nontranscriptional actions of the glucocorticoid receptor. *J Mol Med (Berl)* 81:168–174. doi:10.1007/s00109-003-0418-y
  63. Liu SJ, Wyeth RP, Melchert RB, Kennedy RH (2000) Aging-associated changes in whole cell K<sup>+</sup> and L-type Ca<sup>2+</sup> currents in rat ventricular myocytes. *Am J Physiol Heart Circ Physiol* 279: H889–H900
  64. Losel RM, Falkenstein E, Feuring M, Schultz A, Tillmann HC, Rossol-Haseroth K, Wehling M (2003) Nongenomic steroid action: controversies, questions, and answers. *Physiol Rev* 83:965–1016. doi:10.1152/physrev.00003.2003
  65. Losel R, Wehling M (2003) Nongenomic actions of steroid hormones. *Nat Rev Mol Cell Biol* 4:46–56. doi:10.1038/nrm1009
  66. Lu NZ, Cidlowski JA (2004) The origin and functions of multiple human glucocorticoid receptor isoforms. *Ann N Y Acad Sci* 1024:102–123. doi:10.1196/annals.1321.008
  67. Matsubara H (1998) Pathophysiological role of angiotensin II type 2 receptor in cardiovascular and renal diseases. *Circ Res* 83:1182–1191
  68. McInnes KJ, Kenyon CJ, Chapman KE, Livingstone DE, Macdonald LJ, Walker BR, Andrew R (2004) 5alpha-reduced glucocorticoids, novel endogenous activators of the glucocorticoid receptor. *J Biol Chem* 279:22908–22912. doi:10.1074/jbc.M402822200
  69. McMaster A, Ray DW (2007) Modelling the glucocorticoid receptor and producing therapeutic agents with anti-inflammatory effects but reduced side-effects. *Exp Physiol* 92:299–309. doi:10.1113/expphysiol.2006.036194
  70. Meares GP, Zmijewska AA, Jope RS (2004) Heat shock protein-90 dampens and directs signaling stimulated by insulin-like growth factor-1 and insulin. *FEBS Lett* 574:181–186. doi:10.1016/j.febslet.2004.08.026
  71. Miner JN, Hong MH, Negro-Vilar A (2005) New and improved glucocorticoid receptor ligands. *Expert Opin Investig Drugs* 14:1527–1545. doi:10.1517/13543784.14.12.1527
  72. Miyamoto S, Murphy AN, Brown JH (2009) Akt mediated mitochondrial protection in the heart: metabolic and survival pathways to the rescue. *J Bioenerg Biomembr* 41:169–180. doi:10.1007/s10863-009-9205-y
  73. Molnar GA, Lindschau C, Dubrovskaja G, Mertens PR, Kirsch T, Quinkler M, Gollasch M, Wresche S, Luft FC, Muller DN, Fiebeler A (2008) Glucocorticoid-related signaling effects in vascular smooth muscle cells. *Hypertension* 51:1372–1378. doi:10.1161/hypertensionaha.107.105718
  74. Morin C, Zini R, Simon N, Charbonnier P, Tillement JP, Le Louet H (2000) Low glucocorticoid concentrations decrease oxidative phosphorylation of isolated rat brain mitochondria: an additional effect of dexamethasone. *Fundam Clin Pharmacol* 14:493–500
  75. Mune T, Rogerson FM, Nikkila H, Agarwal AK, White PC (1995) Human hypertension caused by mutations in the kidney isozyme of 11 beta-hydroxysteroid dehydrogenase. *Nat Genet* 10:394–399. doi:10.1038/ng0895-394
  76. Narayanan N, Yang C, Xu A (2004) Dexamethasone treatment improves sarcoplasmic reticulum function and contractile performance in aged myocardium. *Mol Cell Biochem* 266:31–36
  77. Nussinovitch U, de Carvalho JF, Pereira RM, Shoenfeld Y (2010) Glucocorticoids and the cardiovascular system: state of the art. *Curr Pharm Des* 16:3574–3585
  78. Odermatt A, Arnold P, Frey FJ (2001) The intracellular localization of the mineralocorticoid receptor is regulated by 11beta-hydroxysteroid dehydrogenase type 2. *J Biol Chem* 276:28484–28492. doi:10.1074/jbc.M100374200
  79. Ohtani T, Mano T, Hikoso S, Sakata Y, Nishio M, Takeda Y, Otsu K, Miwa T, Masuyama T, Hori M, Yamamoto K (2009) Cardiac steroidogenesis and glucocorticoid in the development of cardiac hypertrophy during the progression to heart failure. *J Hypertens* 27:1074–1083. doi:10.1097/HJH.0b013e328326cb04
  80. Okuda M, Young KR Jr, Lefer AM (1976) Localization of glucocorticoid uptake in normal and ischemic myocardial tissue of isolated perfused cat hearts. *Circ Res* 39:640–646
  81. Penefsky ZJ, Kahn M (1971) Inotropic effects of dexamethasone in mammalian heart muscle. *Eur J Pharmacol* 15:259–266
  82. Perogramvros I, Underhill C, Henley DE, Hadfield KD, Newman WG, Ray DW, Lightman SL, Hammond GL, Trainer PJ (2010) Novel corticosteroid-binding globulin variant that lacks steroid binding activity. *J Clin Endocrinol Metab* 95:E142–E150. doi:10.1210/jc.2010-0746
  83. Psarra AM, Hermann S, Panayotou G, Spyrou G (2009) Interaction of mitochondrial thioredoxin with glucocorticoid receptor and NF-kappaB modulates glucocorticoid receptor and NF-kappaB signaling in HEK-293 cells. *Biochem J* 422:521–531. doi:10.1042/bj20090107
  84. Psarra AM, Sekeris CE (2008) Steroid and thyroid hormone receptors in mitochondria. *IUBMB Life* 60:210–223. doi:10.1002/iub.37
  85. Puthanveetil P, Wang Y, Wang F, Kim MS, Abrahami A, Rodrigues B (2010) The increase in cardiac pyruvate dehydrogenase kinase-4 after short-term dexamethasone is controlled by an Akt-p38-forkhead box other factor-1 signaling axis. *Endocrinology* 151:2306–2318. doi:10.1210/en.2009-1072
  86. Radomski MW, Palmer RM, Moncada S (1990) Glucocorticoids inhibit the expression of an inducible, but not the constitutive, nitric oxide synthase in vascular endothelial cells. *Proc Natl Acad Sci U S A* 87:10043–10047
  87. Rao MK, Xu A, Narayanan N (2001) Glucocorticoid modulation of protein phosphorylation and sarcoplasmic reticulum function

- in rat myocardium. *Am J Physiol Heart Circ Physiol* 281:H325–H333
88. Reichardt HM, Tuckermann JP, Gottlicher M, Vujic M, Weih F, Angel P, Herrlich P, Schutz G (2001) Repression of inflammatory responses in the absence of DNA binding by the glucocorticoid receptor. *EMBO J* 20:7168–7173. doi:10.1093/emboj/20.24.7168
  89. Reuter KC, Loitsch SM, Dignass AU, Steinhilber D, Stein J (2012) Selective non-steroidal glucocorticoid receptor agonists attenuate inflammation but do not impair intestinal epithelial cell restitution in vitro. *PLoS One* 7:e29756. doi:10.1371/journal.pone.0029756
  90. Revollo JR, Cidlowski JA (2009) Mechanisms generating diversity in glucocorticoid receptor signaling. *Ann N Y Acad Sci* 1179:167–178. doi:10.1111/j.1749-6632.2009.04986.x
  91. Rivers C, Levy A, Hancock J, Lightman S, Norman M (1999) Insertion of an amino acid in the DNA-binding domain of the glucocorticoid receptor as a result of alternative splicing. *J Clin Endocrinol Metab* 84:4283–4286
  92. Rosen J, Miner JN (2005) The search for safer glucocorticoid receptor ligands. *Endocr Rev* 26:452–464. doi:10.1210/er.2005-0002
  93. Rubin JM, Hidalgo A, Bordallo C, Cantabrana B, Sanchez M (1999) Positive inotropism induced by androgens in isolated left atrium of rat: evidence for a cAMP-dependent transcriptional mechanism. *Life Sci* 65:1035–1045
  94. Sambhi MP, Weil MH, Udhoji VN (1965) Acute pharmacodynamic effects of glucocorticoids; cardiac output and related hemodynamic changes in normal subjects and patients in shock. *Circulation* 31:523–530
  95. Sapolsky RM, Romero LM, Munck AU (2000) How do glucocorticoids influence stress responses? Integrating permissive, suppressive, stimulatory, and preparative actions. *Endocr Rev* 21:55–89
  96. Schacke H, Docke WD, Asadullah K (2002) Mechanisms involved in the side effects of glucocorticoids. *Pharmacol Ther* 96:23–43
  97. Schmidt BM, Gerdes D, Feuring M, Falkenstein E, Christ M, Wehling M (2000) Rapid, nongenomic steroid actions: a new age? *Front Neuroendocrinol* 21:57–94. doi:10.1006/frne.1999.0189
  98. Schoneveld JL, Fritsch-Stork RD, Bijlsma JW (2011) Nongenomic glucocorticoid signaling: new targets for immunosuppressive therapy? *Arthritis Rheum* 63:3665–3667. doi:10.1002/art.30635
  99. Sellevold OF, Jynge P (1989) Bell-shaped concentration-response curve for myocardial stimulation by glucocorticoids. An experimental study in the rat. *Acta Anaesthesiol Scand* 33:61–65
  100. Seri I, Tan R, Evans J (2001) Cardiovascular effects of hydrocortisone in preterm infants with pressor-resistant hypotension. *Pediatrics* 107:1070–1074
  101. Shaltout HA, Rose JC, Figueroa JP, Chappell MC, Diz DI, Averill DB (2010) Acute AT(1)-receptor blockade reverses the hemodynamic and baroreflex impairment in adult sheep exposed to antenatal betamethasone. *Am J Physiol Heart Circ Physiol* 299:H541–H547. doi:10.1152/ajpheart.00100.2010
  102. Shivaji S, Jagannadham MV (1992) Steroid-induced perturbations of membranes and its relevance to sperm acrosome reaction. *Biochim Biophys Acta* 1108:99–109
  103. Silvestre JS, Robert V, Heymes C, Aupetit-Faisant B, Mouas C, Moalic JM, Swynghedauw B, Delcayre C (1998) Myocardial production of aldosterone and corticosterone in the rat. Physiological regulation. *J Biol Chem* 273:4883–4891
  104. Simon N, Jolliet P, Morin C, Zini R, Urien S, Tillement JP (1998) Glucocorticoids decrease cytochrome c oxidase activity of isolated rat kidney mitochondria. *FEBS Lett* 435:25–28
  105. Song IH, Buttgerit F (2006) Non-genomic glucocorticoid effects to provide the basis for new drug developments. *Mol Cell Endocrinol* 246:142–146. doi:10.1016/j.mce.2005.11.012
  106. Spath JA Jr, Lane DL, Lefer AM (1974) Protective action of methylprednisolone on the myocardium during experimental myocardial ischemia in the cat. *Circ Res* 35:44–51
  107. Stahn C, Buttgerit F (2008) Genomic and nongenomic effects of glucocorticoids. *Nat Clin Pract Rheumatol* 4:525–533. doi:10.1038/ncprheum0898
  108. Steiner A, Locher R, Sachinidis A, Vetter W (1989) Cortisol-stimulated phosphoinositide metabolism in vascular smooth muscle cells: a role for glucocorticoids in blood pressure control? *J Hypertens Suppl* 7:S140–S141
  109. Steiner A, Vogt E, Locher R, Vetter W (1988) Stimulation of the phosphoinositide signalling system as a possible mechanism for glucocorticoid action in blood pressure control. *J Hypertens Suppl* 6:S366–S368
  110. Talaber G, Boldizsar F, Bartis D, Palinkas L, Szabo M, Berta G, Setalo G Jr, Nemeth P, Berki T (2009) Mitochondrial translocation of the glucocorticoid receptor in double-positive thymocytes correlates with their sensitivity to glucocorticoid-induced apoptosis. *Int Immunol* 21:1269–1276. doi:10.1093/intimm/dxp093
  111. Tanz RD, Kerby CF (1961) The inotropic action of certain steroids upon isolated cardiac tissue; with comments on steroidal cardiotoxic structure-activity relationships. *J Pharmacol Exp Ther* 131:56–64
  112. Taves MD, Gomez-Sanchez CE, Soma KK (2011) Extra-adrenal glucocorticoids and mineralocorticoids: evidence for local synthesis, regulation, and function. *Am J Physiol Endocrinol Metab* 301:E11–E24. doi:10.1152/ajpendo.00100.2011
  113. Thiemermann C (2002) Corticosteroids and cardioprotection. *Nat Med* 8:453–455. doi:10.1038/nm0502-453
  114. Tomlinson JW, Walker EA, Bujalska IJ, Draper N, Lavery GG, Cooper MS, Hewison M, Stewart PM (2004) 11beta-hydroxysteroid dehydrogenase type 1: a tissue-specific regulator of glucocorticoid response. *Endocr Rev* 25:831–866. doi:10.1210/er.2003-0031
  115. Waldegger S, Barth P, Raber G, Lang F (1997) Cloning and characterization of a putative human serine/threonine protein kinase transcriptionally modified during anisotonic and isotonic alterations of cell volume. *Proc Natl Acad Sci U S A* 94:4440–4445
  116. Wallerath T, Witte K, Schafer SC, Schwarz PM, Prellwitz W, Wohlfart P, Kleinert H, Lehr HA, Lemmer B, Forstermann U (1999) Down-regulation of the expression of endothelial NO synthase is likely to contribute to glucocorticoid-mediated hypertension. *Proc Natl Acad Sci U S A* 96:13357–13362
  117. Wang Z, Chen W, Kono E, Dang T, Garabedian MJ (2007) Modulation of glucocorticoid receptor phosphorylation and transcriptional activity by a C-terminal-associated protein phosphatase. *Mol Endocrinol* 21:625–634. doi:10.1210/me.2005-0338
  118. Weigel NL, Moore NL (2007) Steroid receptor phosphorylation: a key modulator of multiple receptor functions. *Mol Endocrinol* 21:2311–2319. doi:10.1210/me.2007-0101
  119. Whitworth JA (1994) Studies on the mechanisms of glucocorticoid hypertension in humans. *Blood Press* 3:24–32
  120. Whitworth JA, Kelly JJ, Brown MA, Williamson PM, Lawson JA (1997) Glucocorticoids and hypertension in man. *Clin Exp Hypertens* 19:871–884
  121. Xue Q, Dasgupta C, Chen M, Zhang L (2011) Foetal hypoxia increases cardiac AT(2)R expression and subsequent vulnerability to adult ischaemic injury. *Cardiovasc Res* 89:300–308. doi:10.1093/cvr/cvq303
  122. Yang S, Zhang L (2004) Glucocorticoids and vascular reactivity. *Curr Vasc Pharmacol* 2:1–12
  123. Yang M, Zheng J, Miao Y, Wang Y, Cui W, Guo J, Qiu S, Han Y, Jia L, Li H, Cheng J, Du J (2012) Serum-glucocorticoid regulated kinase 1 regulates alternatively activated macrophage polarization contributing to angiotensin II-induced inflammation and cardiac

- fibrosis. *Arterioscler Thromb Vasc Biol* 32:1675–1686. doi:10.1161/atvbaha.112.248732
124. Yano K, Tsuda Y, Kaji Y, Kanaya S, Fujino T, Niho Y (1994) Effects of hydrocortisone on transmembrane currents in guinea pig ventricular myocytes—possible evidence for positive inotropism. *Jpn Circ J* 58:836–843
125. Young MJ, Funder JW (1996) The renin-angiotensin-aldosterone system in experimental mineralocorticoid-salt-induced cardiac fibrosis. *Am J Physiol* 271:E883–E888
126. Yudit MR, Jewell CM, Bienstock RJ, Cidlowski JA (2003) Molecular origins for the dominant negative function of human glucocorticoid receptor beta. *Mol Cell Biol* 23:4319–4330
127. Zecevic M, Heitzmann D, Camargo SM, Verrey F (2004) SGK1 increases Na, K-ATP cell-surface expression and function in *Xenopus laevis* oocytes. *Pflugers Arch* 448:29–35. doi:10.1007/s00424-003-1222-9
128. Zhou J, Li M, Sheng CQ, Liu L, Li Z, Wang Y, Zhou JR, Jing ZP, Chen YZ, Jiang CL (2011) A novel strategy for development of glucocorticoids through non-genomic mechanism. *Cell Mol Life Sci* 68:1405–1414. doi:10.1007/s00018-010-0526-0
129. Zietz B, Hrach S, Scholmerich J, Straub RH (2001) Differential age-related changes of hypothalamus-pituitary-adrenal axis hormones in healthy women and men—role of interleukin 6. *Exp Clin Endocrinol Diabetes* 109:93–101. doi:10.1055/s-2001-14833
130. Zuo Z, Urban G, Scammell JG, Dean NM, McLean TK, Aragon I, Honkanen RE (1999) Ser/Thr protein phosphatase type 5 (PP5) is a negative regulator of glucocorticoid receptor-mediated growth arrest. *Biochemistry* 38:8849–8857. doi:10.1021/bi990842e