

# Organic anion transporters OAT1 and OAT4 mediate the high affinity transport of glutarate derivatives accumulating in patients with glutaric acidurias

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**Abstract** Glutaric acidurias are rare inherited neurodegenerative disorders accompanied by accumulation of the metabolites glutarate (GA) and 3-hydroxyglutarate (3OHGA), glutaconate, L-, or D-2-hydroxyglutarate (L-2OHGA, D-2OHGA) in all body fluids. Oocytes expressing the human (h) sodium-dicarboxylate cotransporter (NaDC3) showed sodium-dependent inward currents mediated by GA, 3OHGA, L-, and D-2OHGA. The organic anion transporters (OATs) were examined as additional transporters for GA derivatives. The uptake of [<sup>3</sup>H]*p*-aminohippurate in hOAT1-transfected human embryonic kidney (HEK293) cells was inhibited by GA, 3OHGA, D-, or L-2OHGA in a concentration-dependent manner. None of these compounds affected the hOAT3-mediated uptake of [<sup>3</sup>H]estrone sulfate (ES). In hOAT4-expressing cells and oocytes, ES uptake was strongly increased by intracellular GA derivatives. The data provide a model for the concerted action of OAT1 and NaDC3 mediating the basolateral uptake, and OAT4 mediating apical secretion of GA derivatives from proximal tubule cells and therefore contribute to the renal clearance of these compounds.

**Keywords** NaDC3 · Organic anion transporters · OAT1 · OAT4 · Renal excretion of glutarate derivatives · Sodium-dependent dicarboxylate transporters

## Introduction

Several inborn metabolic diseases lead to the accumulation of dicarboxylic acids such as glutaric acid. Glutaric aciduria type 1 (GA1) is caused by deficiency of glutaryl-CoA dehydrogenase (EC 1.3.99.7; GCDH), a mitochondrial enzyme involved in the degradation of tryptophan and lysine. Defective GCDH causes insufficient glutaryl-CoA breakdown and results in an accumulation of glutaric acid (GA), 3-hydroxyglutaric acid (3OHGA), glutaconate (GC), and glutarylcarnitine in tissues, plasma, cerebrospinal fluid, and urine [1–4]. During catabolic situations, GA1 patients are subject to the development of encephalopathic crises, leading to striatal degeneration with a subsequent irreversible dystonic–dyskinetic movement disorder. Histopathological findings include neuronal loss and astrogliosis in the striatum, subdural hemorrhages, and spongiform vacuolation of the white matter. These symptoms and the associated histopathological changes in putamen and caudatum mainly occur between the 3rd and 36th month of age triggered by a nonspecific disease like fever, respiratory infection, or diarrhea [1–5].

Glutaric aciduria type 2 (GA2) is an autosomal recessive disorder associated with hypoketotic hypoglycemia, metabolic acidosis, liver-, kidney-, and cardiomegaly, often leading to an intractable neonatal illness with early death. GA2 is caused by a defect of electron transfer flavoprotein (ETF) or ETF cytochrome Q oxidoreductase and subsequent deficiencies of multiple acyl-CoA dehydrogenases.

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Defective fatty acid oxidation as well as amino acid degradation led to accumulation of GA, lactate, and dicarboxylic acids such as ethylmalonate and adipate [6]. Two other clinical entities, both affecting the nervous system, are characterized by accumulation of D- and L-2-hydroxyglutaric acids (D-2OHGA, L-2OHGA), respectively, in tissues and body fluids [7–9]. Patients with D-2-hydroxyglutaric aciduria suffer from developmental delay, macrocephalus, epilepsy, and cerebral atrophy. Defects of either hydroxyacid-oxoacid transhydrogenase or D-2OHGA dehydrogenase have been observed in these patients. L-2-hydroxyglutaric aciduria is caused by deficiency of the FAD-dependent L-2-hydroxyglutarate dehydrogenase and accompanied by psychomotor retardation, ataxia, hypotonia, tremor, and seizures [10].

For diagnosis, newborn screening programs have been developed based mainly on the detection of glutaryl-carnitine in dried blood spots [11]. GA1 is considered treatable if diagnosed before onset of encephalopathic crises. Patients excrete with the urine large amounts of GA and 3OHGA which exceeds glomerular filtration rate, indicating that these substances have to be secreted [1].

At physiological pH, GA derivatives exist as highly hydrophilic dicarboxylate anions requiring specific transport systems to translocate these compounds across biological membranes. Recently, we showed that GA,  $\alpha$ -ketoglutarate ( $\alpha$ KG) [12], and 3OHGA [13] are substrates of the sodium dicarboxylate cotransporter 3 (NaDC3, aliases NaC3, SDCT2 [14]). Further renal transporters interacting with  $\alpha$ KG and GA are the organic anion transporters (OAT)1 and OAT3 located in the basolateral membrane and OAT4 in the apical membrane of proximal tubular cells [15–17]. OATs are in the focus of medical interest due to their roles in the excretion of many clinically drugs such as antibiotics, antivirals, H<sub>2</sub> antagonists, antihypertensives, cytostatics, diuretics, nonsteroidal anti-inflammatory drugs, statins, and uricosurics [15–17]. In the present study, we report on the capability of OATs to transport GA derivatives across membranes. Since these transporters are present also in the brain [18], OATs may not only influence plasma levels of GA derivatives by securing their renal excretion but may also have an important impact on their levels in brain tissue.

## Materials and methods

**Solutions** A standard oocyte Ringer solution (ORi) was used for oocyte preparation, storage, and for the uptake as well as for the electrophysiological measurements. ORi contained (in mM): 110 NaCl, three KCl, two CaCl<sub>2</sub>, five 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid (HEPES)/Tris adjusted to pH 7.5. All chemicals, including those for ORi, for oocyte preparation and storage as well as

for the electrophysiological experiments on oocytes and the uptake experiments on oocytes and HEK293 cells were purchased from Sigma (Taufkirchen, Germany). 3OHGA and [<sup>3</sup>H]3OHGA were kindly provided by J. Thiem (Institute of Organic Chemistry, University of Hamburg, Hamburg, Germany). Their synthesis was described in the additional material provided electronically in [13]. Radioactively labeled [<sup>3</sup>H]estrone sulfate (ES, 43.5 Ci/mmol) and [<sup>3</sup>H]para-aminohippurate (PAH, 4.18 Ci/mmol) were from Perkin Elmer Life Sciences (Shelton, CT, USA).

**In vitro transcription of cRNA** hNaDC3 and hOAT4 cDNAs were used as templates for cRNA synthesis. Plasmids were linearized with Not I and in vitro cRNA transcription was performed using T7 mMessage mMachine kit (Ambion, Austin, TX, USA) according to the manufacturer's instructions. The resulting cRNA was suspended in purified, RNase-free water to a final concentration of 1  $\mu$ g/ $\mu$ l.

**Oocyte preparation and storage** Female *Xenopus laevis* (Nasco, Fort Atkinson, WI, USA) were subjected to partial ovariectomy according to protocols approved by the Universitätsmedizin Göttingen, Medical Center Animal Care and Use Committee. Stage V and VI oocytes were prepared by an overnight treatment in ORi supplemented with collagenase (Typ CLS II, Biochrom, Berlin, Germany). Subsequently, after being washed with Ca<sup>2+</sup>-free ORi to remove adhering follicle cells, the oocytes were injected with 23 nl of 1  $\mu$ g/ $\mu$ l cRNA coding for hNaDC3, hOAT4, or water. Afterward, the injected oocytes were incubated at 16°–18°C in ORi supplemented with gentamycin (12 mg/ml) and 2.5 mM sodium pyruvate. After 3 to 4 days of incubation with daily medium changes, oocytes were used for tracer uptake and electrophysiological studies.

**hOAT1-, hOAT3-, and hOAT4-transfected HEK293 cells** Human OAT1 and OAT3 (GeneBank accession number: AF097490 and BI760120) were obtained from Resource Center for Genome Research (RZPD, Berlin, Germany). Human OAT4 (GeneBank accession number AL514126) was from Invitrogen (Groningen, The Netherlands). The stably transfected human epithelial kidney cells T-REX™-HEK293-hOAT1, HEK293-hOAT3, and HEK293-hOAT4 were established by using the Flp-In™ expression system (Invitrogen) as previously described [19]. Stably transfected HEK293 cells were selected by hygromycin (10  $\mu$ g/ml) and grown in flasks in high glucose DMEM medium (Invitrogen) supplemented with 10% fetal calf serum, 1% penicillin/streptomycin, and blasticidine (5 mg/l; Sigma). For control, the cells were transfected with the vector alone. Cultures were maintained in humidified atmosphere that contained 5% CO<sub>2</sub> at 37°C.

**Electrophysiological experiments** These studies were carried out 3 days after cRNA injection at room temperature. Oocytes were placed in a 0.5 ml bath chamber on the stage of a dissecting microscope and impaled under direct view with borosilicate glass (BioMedical Instruments, Zöllnitz, Germany) microelectrodes filled with 3 M KCl. Current recordings were made using the two electrode voltage clamp technique in the voltage clamp mode with a commercial amplifier (OC725, Warner, Hamden, CT, USA). Borosilicate glass microelectrodes had resistances of  $\sim 1\text{ M}\Omega$ .

**Tracer uptake experiments** HEK293-hOAT1, HEK293-hOAT3, HEK293-hOAT4 and vector-transfected control cells were harvested and plated into 24-well plastic dishes (Sarstedt, Nümbrecht, Germany) at a density of  $2 \times 10^5$  cells/well. Transport assays were performed 48 h after seeding in mammalian Ringer solution that contained (in mM): 130 NaCl, four KCl, one  $\text{CaCl}_2$ , one  $\text{MgSO}_4$ , one  $\text{NaH}_2\text{PO}_4$ , 20 HEPES, and 18 glucose at pH 7.4. The cells were washed twice with 0.5 ml Ringer and incubated for 5 min in Ringer that contained 10 or 20 nM [ $^3\text{H}$ ]estrone sulfate for OAT3- and OAT4-expressing HEK293 cells, or 1.2  $\mu\text{M}$  [ $^3\text{H}$ ]PAH or 20 nM [ $^3\text{H}$ ]3OHGA for OAT1-expressing HEK293 cells. Uptake was terminated by removal of the radioactive medium and immediate  $3 \times 3$  ml washes with ice-cold Ringer solution. The cells were dissolved in 0.5 ml one N NaOH by gently shaking for 120 min followed by neutralization with 0.5 ml one N HCl. The ( $^3\text{H}$ ) content was determined by liquid scintillation counting (Tricarb 2900TR, Perkin Elmer, Rodgau, Germany).

Some experiments required high intracellular concentrations of the GA derivatives within the cells or the oocytes. For that purpose, OAT4-transfected HEK293 cells were incubated for 2 h in Ringer containing in addition 1 mM 3OHGA, D-, or L-2OHGA. In a second set, oocytes were injected with 23 nl of either 60 mM 3OHGA, D-, or L-2OHGA or an equivalent amount of ORi (control). Afterward, uptake experiments in cells were performed as described above. The uptake of [ $^3\text{H}$ ]ES in oocytes was measured as described previously [20].

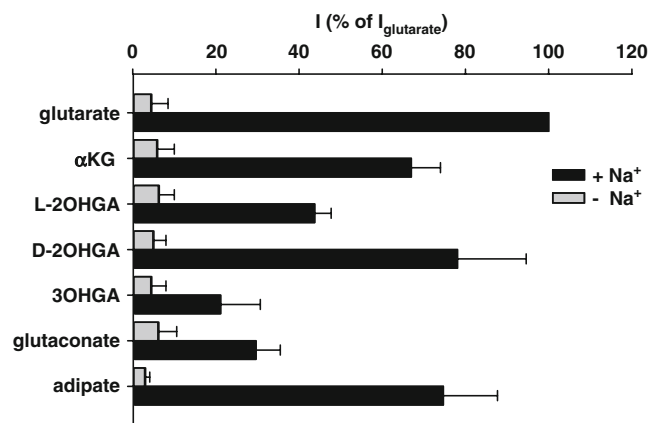
**Statistics** Paired Student's *t*-tests were used to show statistically significant differences of GA derivative-associated inward currents to the currents evoked by GA in the same oocyte. The same test was also used for demonstrating statistical significance of PAH or ES uptake in *cis*-inhibition or *trans*-stimulation experiments. Statistical significance was set at  $p < 0.01$ .  $K_i$  values were determined by Dixon plot analysis. To estimate the kinetic parameters for the transport by hNaDC3, the equation  $I = I_{\text{max}} [S]/(K_m + [S])$  (SigmaPlot 10 software, Systat, Point Richmond, CA, USA) was used, where *I* is the current actually measured,  $I_{\text{max}}$  the maximum

current,  $K_m$  the Michaelis–Menten constant, and *S* the substrate concentration.

## Results

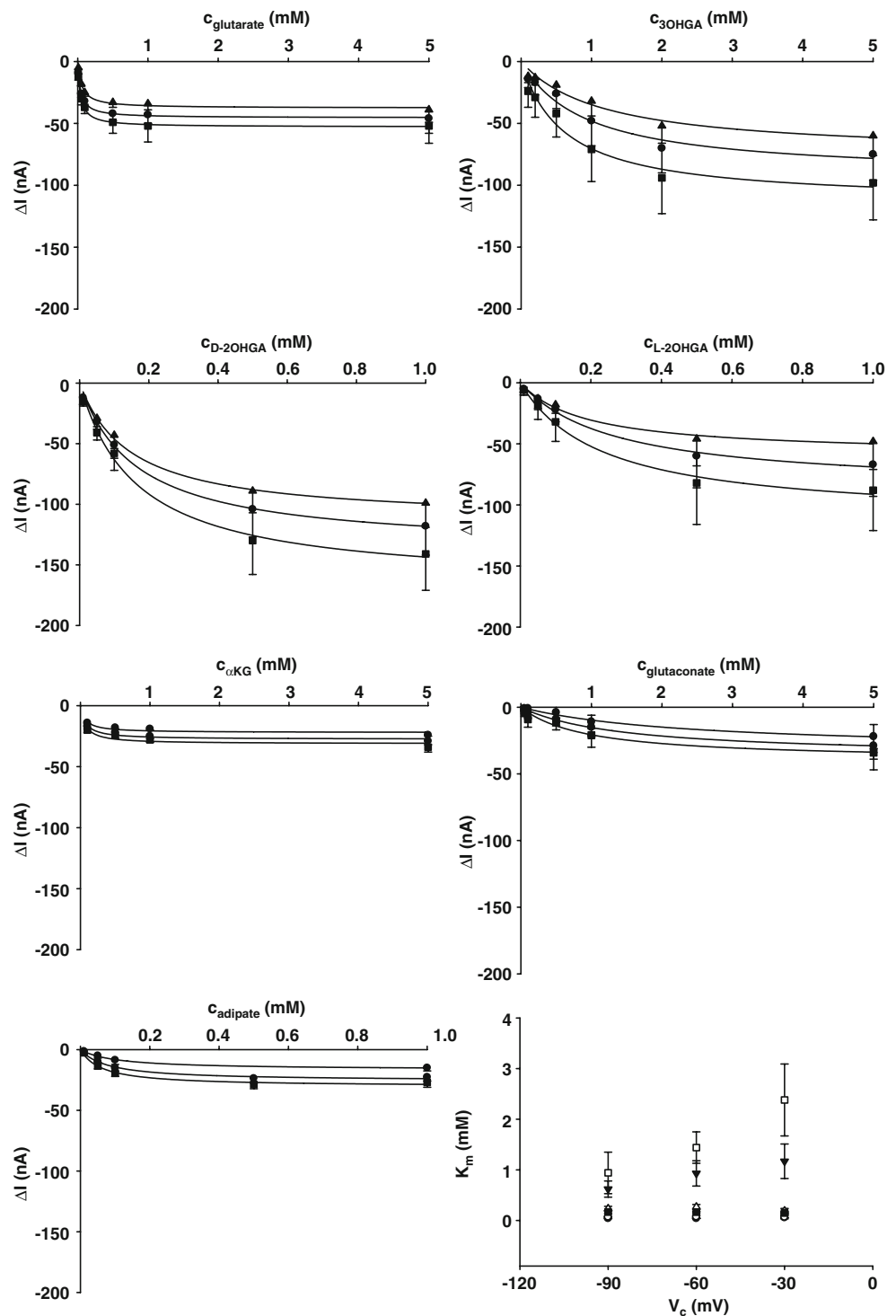
**Substrate specificity for glutarate derivatives and other dicarboxylates of human NaDC3 expressed in *X. laevis* oocytes** Perfusion of hNaDC3-expressing oocytes with L-, D-2OHGA, GC, and adipate (each 1 mM) elicited inward currents (Fig. 1). GA,  $\alpha\text{KG}$ , and 3OHGA were used as controls [13]. In the absence of sodium, these currents were reduced by 81–99%, indicating their dependence on sodium (Fig. 1). The magnitude of the steady state currents was a saturable function of the concentration of the GA derivatives (Fig. 2). Although the  $K_m$  was the lowest for GA and adipate (0.04 mM), the transport capacity ( $I_{\text{max}}$ ) for D-, L-2OHGA, and 3OHGA was greater than that for GA and adipate (Table 1). For three distinct voltages  $-90$ ,  $-60$ , and  $-30$  mV, the data were fitted to the Michaelis–Menten equation and the  $K_m$  values obtained were plotted as a function of membrane potential (Fig. 2, bottom panel, right side). For substitutions at position C2 ( $\alpha\text{KG}$ , D-, and L-2OHGA),  $K_m$  was unaffected by the potential applied. Substitutions at C3 (3OHGA) or introduction of a double bond (GC) yielded comparably higher  $K_m$  values at any membrane potential. Furthermore,  $K_m$  values for 3OHGA and GC increased at depolarizing potentials.

**Effect of various GA derivatives on PAH or ES uptake by the human organic anion transporters OAT1, OAT3, and OAT4** In proximal tubule cells, the transport of dicarboxylates is



**Fig. 1** Inward currents induced by GA derivatives in NaDC3-expressing oocytes. At a holding potential of  $-60$  mV, oocytes were superfused with ORi or the indicated substrates (1 mM) in the presence (black columns) or absence (gray columns) of sodium. Substrate-associated currents, *I*, are presented as percent of the current *I* produced by glutarate in the same batch of oocytes and are shown as mean  $\pm$  SEM from *n* oocytes/*m* frogs: GA 17/11,  $\alpha\text{KG}$  4/3, L-2OHGA 4/3, D-2OHGA 4/3, 3OHGA 8/4, glutaconate 4/3, and adipate 4/3

**Fig. 2** Effect of membrane potential on substrate-associated inward currents and affinities of NaDC3. Steady state substrate-associated currents were measured as a function of three different membrane potentials ( $V_c$ :  $-90$  (filled square),  $-60$  (filled circle),  $-30$  (filled triangle) mV) using each of the indicated substrate and five individual concentrations between 0.01 and 5 mM.  $K_m$  was obtained by fitting  $I = I_{max} [S] / (K_m + [S])$  to the data. The results represent the mean  $\pm$  SEM from  $n$  oocytes/m frogs. The last graph shows the calculated  $K_m$  values. For the GA derivatives, the following symbols are used: (4/3, unfilled square) glutacolate; (7/5, filled inverted triangle) 3OHGA; (5/3, unfilled circle) glutarate and (3/3) adipate; (4/4, filled circle)  $\alpha$ KG; (5/5, unfilled triangle) L-2OHGA; (3/3, filled circle) D-2OHGA



tightly coupled to the transport of organic anions via the organic anion transporters OAT1, OAT3, and OAT4 [15, 16]. Therefore, the impact of GA derivatives on OAT-mediated substrate transfer was investigated. HEK293 cells transfected with the human OAT1, OAT3, or OAT4 cDNA exhibited different inhibition patterns upon simultaneous

application of their prototypical substrate and GA derivatives, adipate, or glutamate. Whereas [ $^3$ H]PAH uptake in hOAT1-transfected HEK293 cells was inhibited by more than 80% of 0.1 mM GA,  $\alpha$ KG, or adipate (Fig. 3a), these compounds were less effective on hOAT3-mediated [ $^3$ H]ES uptake (Fig. 3b). 3OHGA, D-, and L-2OHGA inhibited [ $^3$ H]

**Table 1** Kinetic parameters for NaDC3 substrates

Compound	$K_m$ (mM)	$I_{max}$ (nA)	$I_{max}/K_m$ (nA/mM)	$n/m$
Glutarate	0.040±0.02	-45±1	1125±37	5/3
αKG	0.072±0.027	-28±2	452±135	4/4
L-2OHGA	0.267±0.048	-87±5	336±48	5/5
D-2OHGA	0.164±0.014	-138±4	847±58	3/3
3OHGA	0.93±0.25	-93±9	107±23	7/5
Glutaconate	1.44±0.31	-37±3	26±4	4/3
Adipate	0.040±0.003	-45±1	1129±82	3/3

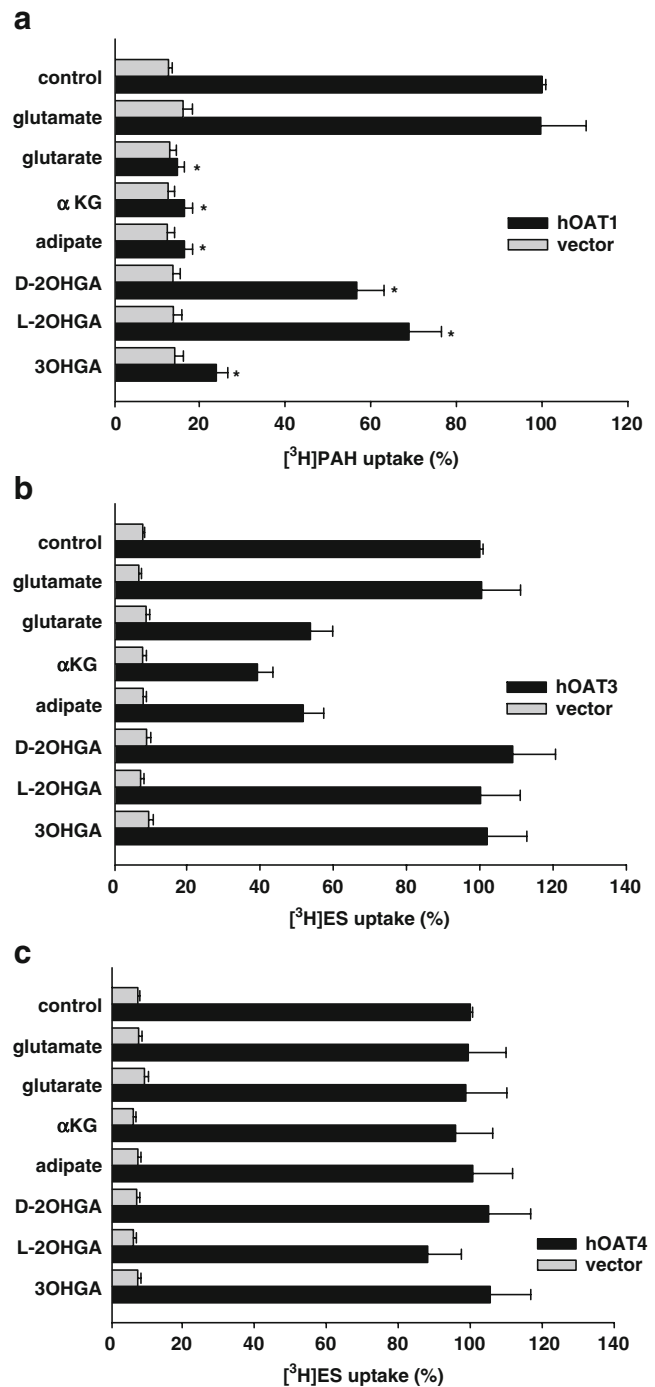
Values are means±SEM from  $n$  oocytes of  $m$  donors.  $K_m$  (Michaelis constant) is the substrate concentration at half-maximal current;  $I_{max}$  is the maximum current observed at saturating substrate concentrations at a holding potential of -60 mV.

PAH uptake in OAT1-expressing cells significantly (Fig. 3a), but showed no effects on OAT3-mediated [<sup>3</sup>H]ES uptake (Fig. 3b). All compounds failed to impair the [<sup>3</sup>H]ES uptake in OAT4-expressing HEK293 cells (Fig. 3c). The dicarboxylic amino acid glutamate (0.1 mM) did neither inhibit [<sup>3</sup>H]PAH nor [<sup>3</sup>H]ES uptake in cells transfected with OAT1, OAT3, or OAT4 cDNA, respectively.

**Inhibition of OAT1-mediated PAH uptake by 3OHGA, D-, and L-2OHGA** To analyze the kinetics of the inhibitory effect of GA derivatives on hOAT1, Dixon plots were used. In three independent experiments, a competitive inhibition of PAH uptake by 3OHGA, D-2OHGA, and L-2OHGA was observed (Fig. 4a,b,c) showing a  $K_i$  of 0.098±0.030 mM, 0.369±0.081 mM, and 0.748±0.032 mM, respectively.

**Direct evidence of transport of 3OHGA, D-, and L-2OHGA by hOAT1-transfected HEK293 cells** Since 3OHGA (0.1 mM) inhibited PAH uptake competitively, *trans*-stimulation experiments with the GA derivatives on OAT1-transfected HEK293 cells were performed (Fig. 5a). All three GA derivatives increased [<sup>3</sup>H]PAH uptake indicating exchange of PAH with either 3OHGA, D-, or L-2OHGA. 3OHGA which showed the lowest  $K_i$  in the Dixon plots (Fig. 4a), evoked the largest *trans*-stimulation. In addition, transport of [<sup>3</sup>H]3OHGA (20 nM) in OAT1-transfected HEK293 cells was assayed. The uptake of [<sup>3</sup>H]3OHGA measured within 5 min was 3.8±1.6 pmol/5 min×2×10<sup>5</sup> cells in hOAT1-transfected and 1.2±0.1 pmol/5 min×2×10<sup>5</sup> cells in vector-transfected cells (Fig. 5b). Despite the low concentration of labeled 3OHGA used, uptake of 3OHGA was enhanced 3.1±1.4 times in OAT1-transfected over vector-transfected cells.

**Trans-stimulation of ES uptake by 3OHGA** Because OAT4 accepted GA only from the intracellular compartment [20], OAT4-transfected HEK293 cells were preincubated for 2 h



**Fig. 3** *Cis*-inhibitory effects of dicarboxylates on uptake of [<sup>3</sup>H]PAH and [<sup>3</sup>H]ES in HEK293 cells expressing OAT1 (a), OAT3 (b), or OAT4 (c). The study was performed in Ringer solution without or with 0.1 mM of the indicated dicarboxylates. The concentrations of labeled substrates were 1.2 μM [<sup>3</sup>H]PAH (a) and 10 nM [<sup>3</sup>H]ES (b, c). The gray columns represent the uptake in vector-transfected cells, and the black columns represent OAT1-, OAT3-, or OAT4-expressing HEK293 cells. All experiments were standardized by setting the control (without inhibitor) of each experiment to 100%. The values represent the mean±SEM of three independent experiments with four repeats each. Significance calculated by Student's *t*-test relative to Ringer solution was accepted at  $p < 0.01$  (\*)

**Fig. 4** Dixon plot analysis for 3OHGA (a), D-2OHGA (b), and L-2OHGA (c) on OAT1-mediated PAH uptake. OAT1-transfected HEK293 cells were incubated in 10  $\mu\text{M}$  (filled circle; 1.2  $\mu\text{M}$  [ $^3\text{H}$ ] PAH plus 8.8  $\mu\text{M}$  unlabeled PAH) or 100  $\mu\text{M}$  (unfilled circle; 1.2  $\mu\text{M}$  [ $^3\text{H}$ ] PAH plus 98.8  $\mu\text{M}$  unlabeled PAH) PAH in the absence or presence of dicarboxylates at the indicated concentrations. Each plot shows a representative experiment out of a series of four independent experiments for each experimental condition

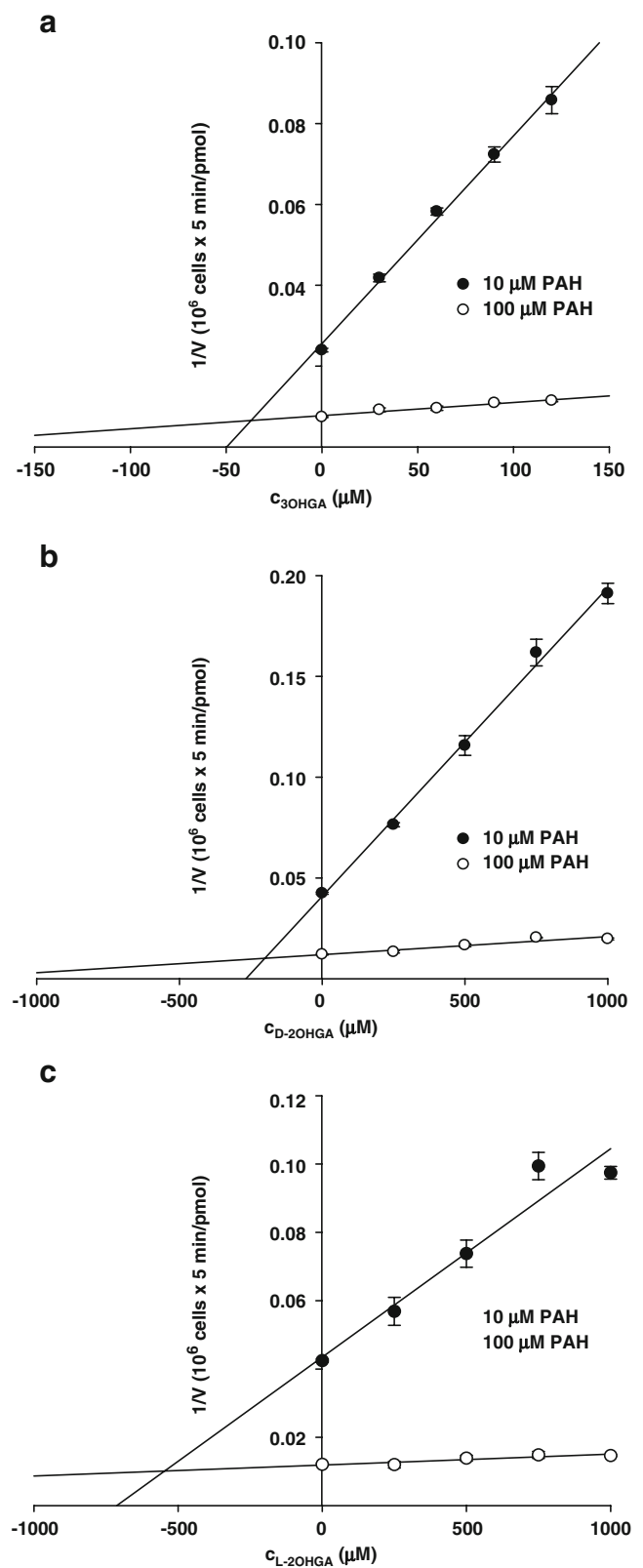
in solutions containing either 1 mM 3OHGA, D-, or L-2OHGA to increase the intracellular concentration of these compounds. Subsequent measurement of ES uptake for 5 min resulted in a *trans*-stimulation of [ $^3\text{H}$ ]ES uptake (Fig. 6a) by a factor of  $1.44 \pm 0.03$ ,  $1.54 \pm 0.02$ ,  $1.73 \pm 0.02$ , indicating an exchange of 3OHGA, D-, and L-2OHGA present within the cell against ES from the bath.

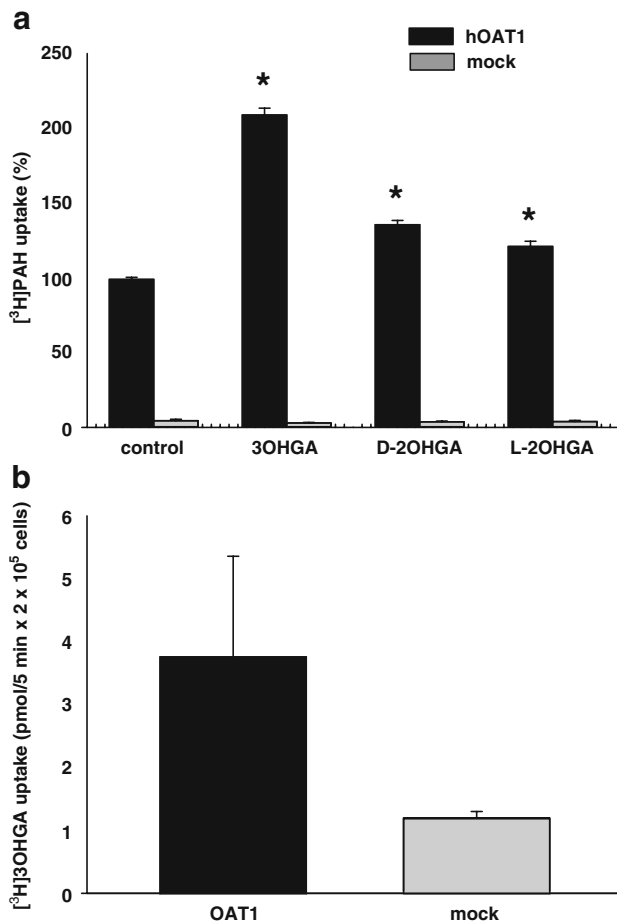
OAT4-expressing oocytes were injected with 3OHGA, L-, and D-2OHGA to efficiently increase the intracellular concentration of these compounds. When ES uptake in buffer-injected oocytes was set to 100%, ES uptake was increased by 3OHGA, L-, and D-2OHGA by a factor of  $1.82 \pm 0.07$ ,  $2.38 \pm 0.26$ , and  $2.11 \pm 0.21$ , respectively (Fig. 6b).

## Discussion

GA1 is caused by a defect in the mitochondrial glutaryl-CoA dehydrogenase. Individuals suffering from GA1 exhibit high concentrations of GA and 3OHGA in their body fluids and tissues. An additional increase in plasma and brain concentrations of these metabolites during catabolic crises is thought to be responsible for acute striatal degeneration. Prerequisite for the cytotoxic effects of GA and 3OHGA on brain and vascular cells [4, 21] is the transport of metabolites across various biological membranes, including mitochondrial and plasma membranes and *trans*-endothelial transport.

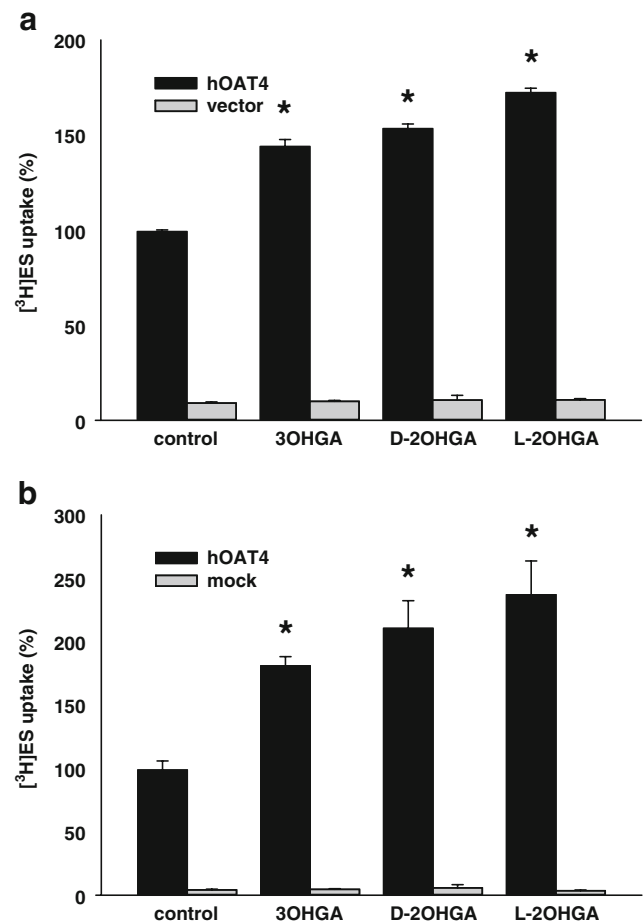
Functional investigations *in vitro* recently demonstrated the capability of the NaDC3 to translocate the GA1-specific metabolite 3OHGA with low affinity [13]. Here, we show that also L-, D-2OHGA, and GC evoked sodium-dependent inward currents in *X. laevis* oocytes expressing hNaDC3. Additionally, adipate, a metabolite accumulating in GA2, showed a sodium-dependent inward current. The affinities of hNaDC3 for these dicarboxylates decreased in the order  $\text{GA} = \text{adipate} > \alpha\text{KG} > \text{D-2OHGA} > \text{L-2OHGA} > 3\text{OHGA} > \text{GC}$ . This sequence indicates that hNaDC3 binds preferentially unsubstituted dicarboxylates with five (GA) and six carbon (adipate) backbones. A double bond within the backbone as in GC largely decreased the affinity, suggesting that the carbon backbone must be highly flexible for interaction with hNaDC3. Substitutions at C2 ( $\alpha\text{KG}$ ,





**Fig. 5** *Trans*-stimulation of OAT1-mediated uptake by [<sup>3</sup>H]PAH and direct evidence for [<sup>3</sup>H]3OHGA uptake by OAT1. *Trans*-stimulation studies were performed by preloading OAT1-transfected HEK293 cells for 2 h with 1 mM of 3OHGA, D-, or L-2OHGA (a). Subsequently, OAT1-mediated PAH-uptake (1.2 μM [<sup>3</sup>H]PAH plus 8.8 μM unlabeled PAH) was measured. *Black columns* represent the uptake measured in OAT1 and *gray columns* such measured in vector-transfected cells. The values represent mean±SEM of four independent repeats. Significance was calculated by Student's *t*-test, \**p*<0.01. In (b), the uptake of 20 nM [<sup>3</sup>H]OHGA in hNaDC3-transfected HEK293 cells and mocks is compared. Data represent mean values of three determinations, where the uptake of [<sup>3</sup>H]OHGA in hOAT1-transfected cells of each individual experiments was set to 100%

D-, and L-2OHGA) appeared to be better tolerated than a substitution at C3 (3OHGA). The two dicarboxylates with the lowest affinities (3OHGA, GC) exhibited a potential-dependent  $K_m$ , whereas unsubstituted and C2-substituted dicarboxylates showed  $K_m$  values independent of membrane potential between  $-90$  and  $-30$  mV. The heavy metal chelator, 2,3-dimercaptosuccinate, also had a  $K_m$  increasing with depolarization [22], suggesting that hNaDC3 has to perform a voltage-dependent step in interacting with compounds carrying a substitution at C3 or a double bond between C2 and C3. The maximal currents,  $I_{max}$ , decreased in the order D-2OHGA>3OHGA=L-2OHGA>GA=adipate>



**Fig. 6** *Trans*-stimulation of OAT4-mediated uptake by [<sup>3</sup>H]estrone sulfate. *Trans*-stimulation studies were performed by preloading (a) OAT4-expressing HEK293 cells for 2 h with 1 mM of the indicated effectors or (b) OAT4-expressing oocytes by injecting 23 nl of either 60 mM 3OHGA, D-, or L-2OHGA or an equivalent amount of ORi (control). Subsequently, OAT4-mediated 20 nM [<sup>3</sup>H]JES uptake was examined. The *gray columns* represent the uptake in vector-transfected HEK293 cells (a) or mock-transfected oocytes (b). *Black columns* represent uptake of radioactively labeled substrates by OAT4-expressing HEK293 cells (a) or oocytes (b). All experiments were standardized by setting the control (Ringer solution) of each experiment to 100%. The values represent the mean±SEM of three independent experiments with four repeats of cells or six to 15 oocytes each. Significance was calculated by Student's *t*-test, \**p*<0.01

GC>αKG. Thereby, the turnover rate of hNaDC3 with a preference for OH substitutions at C2 and C3 appears to follow a different sequence than the affinity. The transport efficiency,  $I_{max}/K_m$ , was the highest for adipate and GA, followed by D-2OHGA>αKG>L-2OHGA>3OHGA>GC. Thus, hNaDC3 may play an important role in the renal secretion particularly of adipate, GA, and 2OHGAs, but not for 3OHGA and GC. A reabsorption of these compounds by hNaDC1 located in the apical [14] is unlikely, because NaDC1 has a low affinity for C5 dicarboxylates ([23] and own unpublished results).

NaDC3 is not only localized in the basolateral membrane of human and rat kidneys [24] but also in rat astrocytes [25, 26] where it mediates  $\alpha$ KG-inhibitable transport of [ $^{14}$ C]*N*-acetyl-L-aspartate, a compound with a dicarboxylic acid like structure. It needs to be established which role hNaDC3 plays in the brain of affected patients.

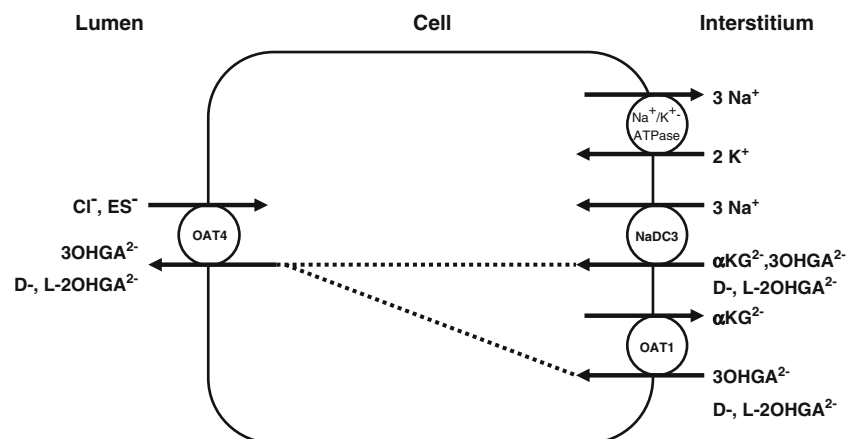
The multispecific OATs may represent additional candidates for high affinity transporters of GA derivatives [15–17]. In our studies, 3OHGA, D-, and L-2OHGA did not interact with hOAT3 expressed in HEK293 cells, while hOAT1 translocated [ $^3$ H]3OHGA. [ $^3$ H]PAH uptake in HEK293 cells transfected with hOAT1 cDNA was markedly reduced by 3OHGA, D-, L-2OHGA, and adipate in *cis*-inhibition (competition) studies. *Vive versa*, [ $^3$ H]PAH uptake was increased by preloading these cells with 3OHGA and D-, or L-2OHGA indicating exchange of PAH with either 3OHGA, D-, or L-2OHGA. These observations provide the first evidence that OAT1 not only interacts with its natural substrate  $\alpha$ KG but also with glutarates possessing hydroxyl groups at position C2 or C3. All GA derivatives exhibited competitive inhibition with PAH. 3OHGA showed a  $K_i$  much lower than D- or L-2OHGA, suggesting that a C3-substituted dicarboxylate may fit better into the binding pocket of hOAT1 than dicarboxylates with substitutions at C2. Glutamate and GC at a concentration of 0.1 mM only weakly interacted with hOAT1 indicating that positively charged substitutions, the amino group in glutamate, or a double bond in the carbon backbone (GC) do not fulfill an ideal configuration for substrates of hOAT1. Acceptance of dicarboxylates by hOAT1 is not restricted to C5 carbon backbone dicarboxylates as demonstrated by inhibition of PAH uptake by adipate (C6). A higher chain length (pimelate; C7 and

suberate; C8) is not suitable for transport (data not shown). Suberate, present in the urine of GA2 patients, has to be excreted by transporters different from OAT1.

3OHGA, D-, and L-2OHGA did not interact with OAT3. These substances also did not inhibit OAT4, but *trans*-stimulated ES uptake into OAT4-transfected cells or OAT4-expressing oocytes. Recently, we have shown that OAT4 which is located at the apical membrane of human proximal tubule cells is working in an asymmetric mode: it accepts dicarboxylates only from the inside (cytosolic side), but not from the outside (urine side) [20]. One of the physiological functions of OAT4 is the reabsorption of estrone sulfate and urate from the filtrate in exchange for intracellular dicarboxylates. Here, we provide experimental evidence of an exchange between extracellular ES and intracellular 3OHGA, L-, and D-2OHGA mediated by OAT4.

Our observations describe for the first time the basic components of the renal secretion of cytotoxic metabolites accumulating in patients suffering from glutaric acidurias. These data are summarized in a cell model for the renal excretion of GA and GA derivatives in Fig. 7. GA and GA derivatives are taken up from the plasma across the basolateral membrane by OAT1 and NaDC3 and are secreted into the urine via OAT4. It is, however, likely that NaDC3 is not only directly involved in the uptake of 3OHGA and 2OHGAs but mediates the transfer of  $\alpha$ KG into proximal tubule cells to supply anionic exchange equivalents for OAT1. The coordinated action of these transporters may guarantee a high renal clearance of GA and GA derivatives under metabolically stable conditions. In states of fever, however, downregulation of the expression of OAT1 [27] may partially impair renal secretion and cause accumulation of GA derivatives with possible side effects.

**Fig. 7** Cell model for the renal secretion of GA derivatives. GA derivatives are taken up from the plasma by OAT1 and NaDC3 across the basolateral membrane and exit the cell by OAT4 in exchange for estrone sulfate or the more physiological anion chloride. The gradient required for sodium-dependent uptake via NaDC3 is maintained by the  $\text{Na}^+/\text{K}^+$ -ATPase





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