

Lambert–Eaton myasthenic syndrome (LEMS): a rare autoimmune presynaptic disorder often associated with cancer

Benedikt Schoser¹  · Bruno Eymard² · Joe Datt³ · Renato Mantegazza⁴

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Abstract Lambert–Eaton myasthenic syndrome (LEMS) is a rare autoimmune neuromuscular junction disorder that is related to the loss of functional P/Q-type voltage-gated calcium channels (VGCCs) on presynaptic nerve terminals. Up to 60% of cases occur as a paraneoplastic disorder (SCLC-LEMS), most commonly in association with small cell lung cancer. The remaining cases have an idiopathic non-tumor etiology but are associated with underlying autoimmune disease (NT-LEMS). Patients with LEMS invariably experience progressive proximal muscle weakness, often accompanied by general fatigue and autonomic symptoms. Some LEMS clinical symptoms overlap with those of other myasthenic syndromes, most commonly myasthenia gravis, which can contribute to misdiagnosis or delayed diagnosis. Prognosis is related to the presence of cancer or autoimmune disease and the severity/distribution of muscle weakness. Cause of death in patients with SCLC-LEMS is typically tumor progression, whereas NT-LEMS does not reduce life expectancy. LEMS diagnosis is supported by a threefold approach: clinical features, electromyography, and anti-VGCC antibody serology. LEMS is a clinically important early indicator of possible cancer;

therefore, a LEMS diagnosis should immediately prompt rigorous oncological screening and surveillance. Symptomatic treatment of LEMS typically involves medications that improve neurotransmission (e.g., the potassium channel blocker amifampridine [3,4-diaminopyridine]), with addition of immunosuppressants/modulators (e.g., prednisone plus azathioprine) in individuals with persistent symptoms. Where a tumor is identified, oncological treatment should take priority. It should be remembered, however, that LEMS has a significant impact on a patient's quality of life and ability to perform daily activities, and therefore warrants timely diagnosis and appropriate treatment in and of itself.

Keywords Autoimmunity · Lambert–Eaton myasthenic syndrome · Neuromuscular junction · Quality of life · Small cell lung carcinoma · Voltage-gated calcium channels

Introduction

Lambert–Eaton myasthenic syndrome (LEMS) is a rare autoimmune disorder of the neuromuscular junction that was first characterized in 1956 by Drs. Edward Lambert, Lee Eaton, and Douglas Rooke of the Mayo Clinic [1, 2]. This myasthenic disorder was given its eponym 15 years later when Dr. Lambert and Dr. Dan Elmqvist presented a detailed microelectrophysiological analysis of the pathological neuromuscular transmission that distinguished the disease from similar conditions such as myasthenia gravis (MG) [3].

Lambert–Eaton myasthenic syndrome (LEMS) may occur as a paraneoplastic disorder (SCLC-LEMS), most commonly in association with small cell lung cancer

✉ Benedikt Schoser
bschoser@med.uni-muenchen.de

¹ Friedrich-Baur-Institute, Department of Neurology Klinikum München Ludwig-Maximilians-University Munich, Munich, Germany

² Institute de Myologie, 47 Boulevard de l'Hôpital, 75013 Paris, France

³ BioMarin Europe Ltd, 10 Bloomsbury Way, London WC1A 2SL, UK

⁴ Fondazione IRCCS Istituto Neurologico, Via Giovanni Celoria, 11, 20133 Milan, Italy

(SCLC), or as an autoimmune disease in the absence of cancer [non-tumor (NT)-LEMS] [4]. Symptoms include gradual onset of fatigue, skeletal muscle weakness, weight loss, and autonomic symptoms such as dry mouth, male impotence, and constipation [2, 5, 6]. Both CA- and NT-LEMS demonstrate circulating immunoglobulin G antibodies against presynaptic P/Q-type voltage-gated calcium channels (VGCCs); these antibodies modulate expression of functional VGCC and thereby inhibit neurotransmission [2].

Because LEMS is a rare disease with fluctuating symptoms, it can be misdiagnosed as MG or as an oncological sequela, or diagnosis can be significantly delayed [2, 5–8]. It is essential that neurologists are aware of LEMS so that affected patients can be correctly diagnosed in a timely fashion. This will allow proper treatment of the neurological disease and any underlying tumor. This review discusses the clinical picture and pathophysiology of LEMS, together with a recommended diagnostic approach.

Epidemiology

Lambert–Eaton myasthenic syndrome (LEMS) is a rare disease, with a world-wide prevalence of around 3–4 per million population [9]. A decade-long study performed from 1990 to 1999 in South Holland found that the annual incidence of LEMS was 0.48 cases per million, with a prevalence of 2.32 per million [10]; a follow-up study across all of the Netherlands indicated an annual incidence of 0.4 cases per million and a prevalence of 2.5 per million in 2003 [11]. Extrapolating the data from the Netherlands to the overall European population suggests that LEMS currently affects around 1850 individuals in this region, with potentially 300 new cases diagnosed each year. A recent study in the US Veterans Affairs population (2013) found a confirmed crude LEMS annual incidence of 0.6 cases per million (or 0.7 cases per million when confirmed and probable cases were combined) [12]. The crude confirmed and confirmed/probable prevalence of LEMS was 2.8 and 3.8 per million, respectively.

CA-LEMS accounts for between 47 and 62% of cases of the syndrome [4, 11, 13, 14]. SCLC is the most frequently occurring underlying tumor, although rare cases of other lung and non-lung cancers have been reported in patients with LEMS [13, 15–30]. Diagnosis of LEMS typically precedes detection of the tumor [31]. In an unselected population of 63 patients with SCLC, two individuals (3%) had a confirmed diagnosis of LEMS [32]. The median age of onset of CA-LEMS is around 60 years, with 59–70% of cases occurring in males [4, 31]. On the other hand, NT-LEMS has a closer age and sex distribution to that seen

with MG. Median age of onset is 50 years [4], with the condition also affecting children [33]. In general, NT-LEMS has an equal representation of males and females, although a female predominance has been reported in individuals diagnosed at less than 45 years old, with a male predominance in those diagnosed after the age of 60 years [34]. NT-LEMS is often associated with an underlying autoimmune disease, including autoimmune thyroid disease, diabetes mellitus, rheumatoid arthritis, and systemic lupus erythematosus [35–37]. In a 2003 study in the Netherlands, the annual incidence of LEMS associated with SCLC was around 0.2 cases per million, with a prevalence of 0.5 per million [11]. The incidence of NT-LEMS was similar in this population (0.2 per million) but the prevalence was higher (2.0 per million) [11], reflective of the better survival associated with NT-LEMS than with CA-LEMS.

Clinical picture and prognosis

Patients with LEMS almost invariably experience proximal weakness in the legs as a first symptom. This typically spreads to proximal weakness in the arms, distal weakness in the arms and legs, and finally weakness involving hands, feet, and cranial muscles [2, 31]. Other commonly reported symptoms include general fatigue, dry mouth, slurred or slow speech, male impotence, droopy eyelids, double vision, difficulty swallowing or chewing, neck weakness, dry eyes, and constipation [4, 5, 31]. Although usually a hallmark of MG, ptosis can also be reported by patients with LEMS, albeit generally in a mild form and later in the disease course, [6, 38]. Cerebellar ataxia is a relatively uncommon symptom, but it appears almost exclusively in patients with CA-LEMS [31]. Symptoms appear to develop more quickly in patients with CA-LEMS than those with NT-LEMS: in a cohort of 97 patients with LEMS, those with CA-LEMS reported a mean of seven different symptoms in the first 6 months of diagnosis, compared with a mean of two symptoms in patients with NT-LEMS [31]. The insidious nature of LEMS symptoms mean that patients may wait for months or years before presenting to their doctor, with a likely further delay from presentation until diagnosis [5]. Recently, a Firdapse registry based on voluntary submission of data by investigators from 29 EU centers has been established. Data are entered into a centralized database via a validated web-portal application. Interestingly, results from specific clinical assessments using the quantitative myasthenia gravis (QMG) score showed that the raw QMG score was 7.2 ± 6.8 and the % standardized QMG total score 20.4 ± 18.8 , thus indicating that the severity of muscle weakness and fatigability was mild to moderate [39].

Little formal documentation has been undertaken to define the burden of illness associated with LEMS. In one of the only studies published in this area, researchers in Germany performed a series of interviews in 12 patients with LEMS (three of whom had a diagnosis of SCLC) and found that leg weakness and general fatigue were considered the most troublesome symptoms [5]. Tellingly, all respondents had mobility issues, 50% reported severe pain or discomfort, and 75% reported frequent restrictions in their activities of daily living. In a study of 47 consecutive patients with NT-LEMS, 25% required a wheelchair at all times or while mobilizing outside [7]. Health-related quality of life, measured by the generic EuroQol five-dimensions five-level (EQ-5D-5L) instrument, was similar in patients with LEMS to that in patients hospitalized with a severe exacerbation of asthma or those with severe multiple sclerosis [5].

The prognosis of patients with LEMS is related to the presence of cancer or autoimmune disease and the severity/distribution of muscle weakness. Given that SCLC is typically an aggressive cancer, the cause of death in patients with CA-LEMS is likely progression of the underlying tumor. Interestingly, however, patients with SCLC have longer overall survival when they have LEMS than when they do not (median of 17.3 months vs 10 months, respectively) [40]. It has been debated whether this is a lead-time bias in identifying SCLC tumors in patients presenting with LEMS or whether it is related to a biological mechanism such as elevated anti-VGCC antibodies [41, 42]. Prognosis in NT-LEMS is variable but is notably different from that in CA-LEMS as the condition does not appear to reduce life expectancy [7]. In a series of 47 patients with NT-LEMS treated at a UK center between 1987 and 1998, around half achieved sustained clinical remission [7]. The majority of patients were symptomatically treated with amifampridine (Firdapse; BioMarin Europe Ltd, London, UK; 79% at final follow-up or death). In most cases, significant and ongoing immunosuppressant treatment was required for patients to remain clinically stable. The only independent predictor of clinical remission or independent ambulation was initial clinical score, comprising strength measurements in proximal limb muscles. Interestingly, neither electrophysiological findings nor anti-VGCC antibody levels correlated with outcome. In a US-based analysis of the Veterans Affairs population, the majority of patients with LEMS treated pharmacologically had some degree of improvement (37/46; 80%) [12]. Of the more frequently used medications, amifampridine was associated with the highest rate of clinical improvement or resolution (14/18; 78%).

Mechanisms of disease

Etiology and risk factors

Voltage-gated calcium channels (VGCCs) are heteromeric multi-subunit complexes and can be classified according to their characteristic voltage activation threshold (high- or low-voltage activated), their sequence similarities at the pore-forming α -1 subunit (Ca.v1, Ca.v2, or Ca.v3) or their pharmacological properties (P/Q, N, L, T, or R). The P/Q-type VGCC is primarily involved in neurotransmitter release from motor nerve terminals, while the effects of the other subtypes include neurotransmitter release from autonomic nerve terminals (N-type) and specialized terminals such as the retina or auditory hair cells (L-type) [43].

Lambert–Eaton myasthenic syndrome (LEMS) is an autoimmune disease caused by the interaction of autoantibodies with P/Q-type VGCCs on presynaptic nerve terminals [44]. Antibodies against the P/Q-type VGCC have been demonstrated in serum for approximately 90% of non-immunosuppressed patients with LEMS [45]. Recent studies have indicated that autoantibodies from patients with LEMS bind to multiple subunits of the P/Q-type VGCC complex [46]. This autoimmune etiology is additionally supported by the observations that immunomodulation improves muscle weakness in many patients with LEMS [2] and that transfer of immunoglobulins from patients with LEMS to mice results in changes at the neuromuscular junction that are consistent with clinical observations [47].

The etiological basis for the development of CA-LEMS is the presence of high concentrations of functional P/Q-type VGCC on SCLC cells, which presumably induce autoimmune production of pathogenic anti-VGCC antibodies [2, 48, 49]. These autoantibodies then cross-react with components of VGCC on presynaptic nerve endings [2, 50, 51], affecting neuromuscular function. Autoantibody production appears to begin at an early stage in tumor development, typically before detection of the tumor itself. As cigarette smoking is a strong risk factor for SCLC, it is also a risk factor for CA-LEMS.

The specific trigger for NT-LEMS is unknown; however, the syndrome is strongly associated with underlying autoimmune disease [4, 35] with a notable maternal link [35]. Furthermore, there is a correlation between NT-LEMS and haplotypes associated with autoimmunity (e.g., HLA-B8, -A1, -A2, and -DR3), particularly in patients with young-onset disease [2, 34, 52].

Pathophysiology

Under physiological conditions, transmembrane P/Q-type VGCCs are expressed on the presynaptic membrane in regular arrays [53]. Depolarization of the presynaptic membrane causes the VGCCs to open, allowing an influx of calcium ions into the nerve terminal. This influx induces fusion of acetylcholine (ACh)-containing synaptic vesicles with the presynaptic membrane, resulting in quantal release of ACh from areas known as “active zones” into the synaptic cleft. ACh binds to ACh receptors on the adjacent postsynaptic endplate of the muscle fiber; this opens ligand-gated sodium and potassium ion channels, inducing depolarization of the endplate. Once the depolarization threshold is met, an action potential occurs and the muscle contracts [4].

In the situation of LEMS, presynaptic ACh stores and postsynaptic response to ACh quanta at the neuromuscular junction are normal, but there is a reduction in release of ACh from the presynaptic nerve terminal that translates into a reduced postsynaptic endplate action potential [4, 54]. The reduction in ACh quanta is attributed to loss of functional P/Q-type VGCCs at the presynaptic terminal, presumably due to autoantibody binding. Immunoelectron microscopy has shown that P/Q-type VGCCs in the presynaptic membrane decrease in number and are expressed in a clustered pattern in LEMS [47, 53]. This not only reduces calcium ion influx at depolarization, but may also decrease the number of presynaptic active zones. P/Q-type and N-type VGCCs act as scaffolding proteins in active zones and are involved in tethering ACh vesicles and bringing them into proximity to the presynaptic membrane [55, 56]. The autoimmune response seen in LEMS not only affects muscle function, but also has an impact on the autonomic nervous system (presumably through interaction of autoantibodies with N-type VGCCs [57]) and, in some patients, on the central nervous system [31] (through interaction of autoantibodies with P/G-type VGCCs located in the cerebellum [3]).

Around 10% of patients with LEMS (predominantly those with NT-LEMS) are seronegative for anti-P/Q-type VGCC antibodies [45]. Clinical features are similar in seronegative and seropositive patients, although the electrophysiological profile may be less pronounced in seronegative patients [58, 59]. It may be that seronegative patients do actually have anti-P/Q-type VGCC antibodies, but at concentrations below the level of detection with current assays; alternatively, they may have antibodies to a different VGCC epitope or a different molecule that generates a similar phenotype [58, 60, 61]. Interestingly, antibodies to SOX1 (a transcription factor expressed in the developing nervous system) have been reported with higher frequency in patients with CA-LEMS (64–65%) than those with SCLC without LEMS

(22–32%), and in very few patients with NT-LEMS (0–5%) [62, 63]; however, no conclusive pathogenicity of anti-SOX1 antibodies has yet been established.

A diagnostic approach

Diagnosis of LEMS is supported by a threefold approach: history and physical examination, electromyography (showing a presynaptic deficit of neuromuscular transmission), and autoantibody (anti-VGCC antibody) serology (Box 1; Fig. 1). Patients with suspicion of LEMS should be examined and treated by a neurologist and, if appropriate, an oncologist. If an underlying tumor is identified, then the treatment priority should be the cancer.

History and physical examination

Lambert–Eaton myasthenic syndrome (LEMS) should be considered in a patient presenting with progressive proximal muscle weakness (particularly in the legs, but may also include arms), together with reduced or absent tendon reflexes (areflexia) [2, 8]. A waddling gait may be noted. One of the hallmarks of LEMS is that tendon reflexes normalize and muscle strength improves immediately after brief maximal contraction; however, not all

Box 1 Diagnostic assessment of LEMS

Features

Risk factors

- Small cell lung cancer
- Cigarette smoking
- Autoimmune disease
- Family history of autoimmune disease

Clinical features

- Progressive proximal muscle weakness (should be present)
 - Areflexia
 - Autonomic symptoms
 - Ocular/bulbar symptoms
 - General fatigue
 - Postoperative muscle weakness after neuromuscular blockers

Electromyography

- Repetitive nerve stimulation studies (should all be present)
 - Low CAMP at rest (0.1–6 mV)
 - Decrease of >10% in CAMP at low frequency (2–5 Hz)
 - Increase of >60% in CAMP after maximum voluntary contraction or at high frequency (20–50 Hz)

Autoantibody serology

- Anti-P/Q-type VGCC antibodies

CAMP compound muscle action potential, LEMS Lambert–Eaton myasthenic syndrome, VGCC voltage-gated calcium channel

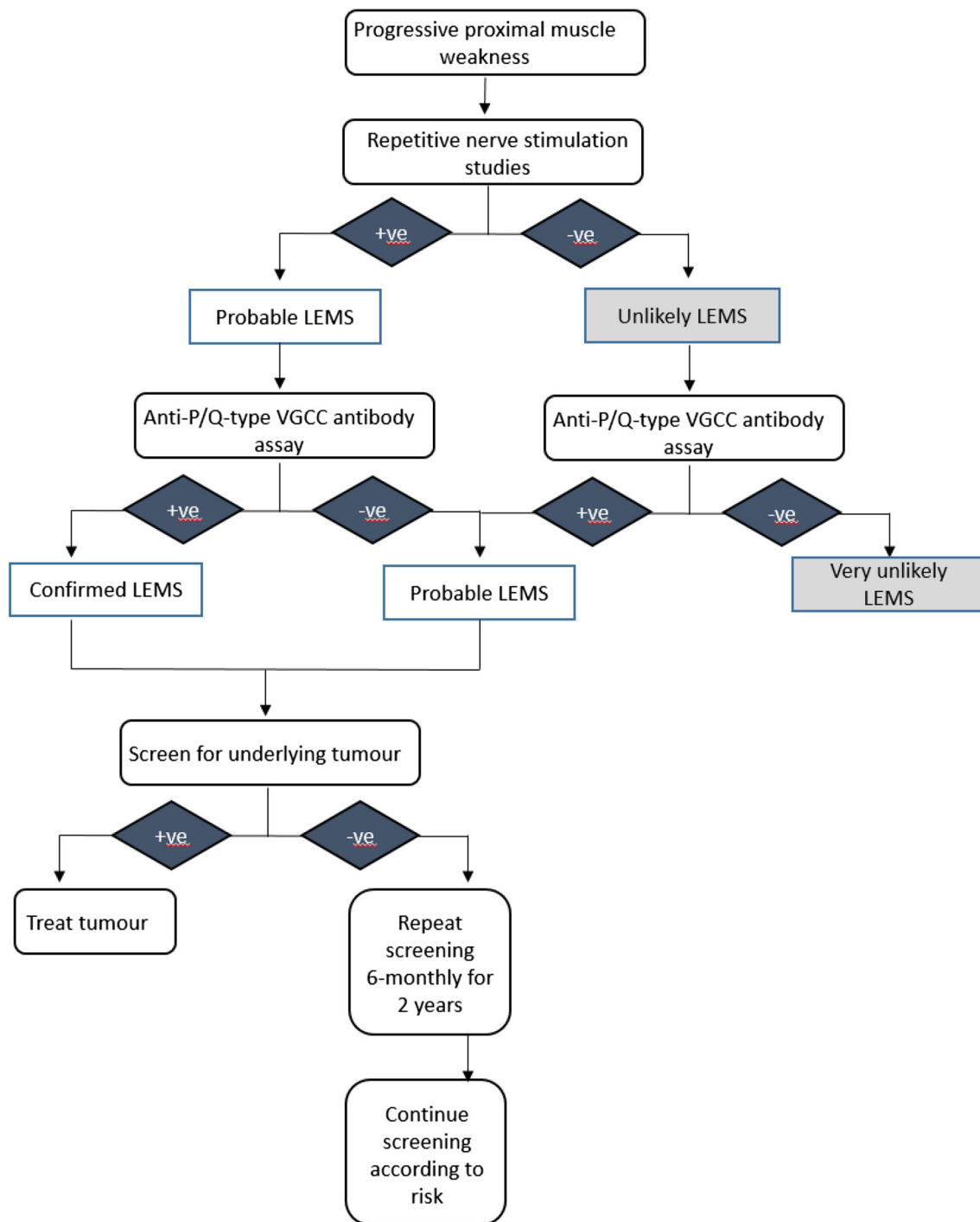


Fig. 1 LEMS diagnosis and oncological screening algorithm. LEMS Lambert–Eaton myasthenic syndrome, VGCC voltage-gated calcium channel

patients demonstrate this post-exercise facilitation [64]. Because this phenomenon can mask lowered tendon reflexes, these should be tested after a rest period [2, 8]. Additional indicative signs of LEMS include general fatigue, autonomic dysfunction (e.g., dry mouth, impotence, or constipation), or weakness of facial or ocular muscles (e.g., ptosis or double vision) [2, 5, 38]. These

symptoms may fluctuate throughout the day. LEMS should also be considered in the case of any unexplained postoperative muscle weakness after neuromuscular-blocking drugs [65]. History should include an assessment of LEMS risk factors such as underlying SCLC or autoimmune disease, a family history of autoimmune disease, or cigarette smoking.

Electromyography

Repetitive nerve stimulation (RNS) tests should be performed on at least two distal muscles, with positive tests showing (1) low amplitude of the compound muscle action potential (CMAP) at rest (0.1–6 mV), and (2) further decrease of the CMAP amplitude (at least 10%) during low-rate (2–5 Hz) RNS [2, 8]. These are the most sensitive diagnostic tests. However, as the decrease in CMAP amplitude during low-rate RNS does not discriminate in total between LEMS and MG, at least two muscles should be tested with either the painful high-frequency (20–50 Hz) RNS or much better and preferably RNS immediately after a brief maximal voluntary contraction (15–20 s) [8]. An increase in the CMAP amplitude (increment) of at least 100% is considered specific for LEMS, although it has been suggested that the threshold can be decreased to 60% to improve sensitivity (97%) while retaining specificity (99%) [66]. RNS after maximal contraction is less painful than high-frequency RNS and involves applying a single supra-maximal stimulus to generate a baseline CMAP [8]. After the brief period of maximal voluntary contraction, the post-exercise CMAP is produced by applying a second stimulus. Consideration should be given to withdrawing symptomatic medication 12 h before electrophysiological testing and to maintaining the temperature of the examined muscles at above 35 °C to further increase sensitivity [67].

Single-fiber electromyography is slightly more sensitive than RNS for diagnosis of LEMS [66] and jitter appears to correlate with clinical and electrophysiological disease severity [68]. However, single-fiber electromyography is less specific than RNS and has limited availability [67]. For these reasons, RNS is the preferred initial test for LEMS [67].

Autoantibody serology

Detection of anti-P/Q-type VGCC antibodies is a highly specific diagnostic confirmation of LEMS; however, the absence of detectable VGCC antibodies does not exclude LEMS [2, 8]. It should be noted that there is no correlation between antibody titres and disease severity [7] and that antibody levels may be affected by the use of immunosuppressants [69]. The presence of antibodies to domain IV of the alpha-1A P/Q-VGCC subunit is highly suggestive of NT-LEMS (occurring in 38% of patients with NT-LEMS and 5% of patients with SCLC-LEMS) [70]. Conversely, the presence of SOX1 antibodies is indicative of SCLC-LEMS [62, 63, 71, 72]. While antibodies against synaptotagmin and M1 muscarinic ACh receptors have been detected in some patients with LEMS, these specific antibodies have no diagnostic value [8].

Misdiagnosis and differential diagnosis

Many clinical symptoms of LEMS overlap with those of other myasthenic syndromes, most commonly MG. In a cohort of 241 Dutch or British individuals diagnosed with LEMS between 1990 and 2009 [14], 58% were initially misdiagnosed [2]. The most common diagnosis was MG (21%), with other diagnoses including myopathy not otherwise specified (11%), polyneuropathy (3%), and depression or psychosomatic causes (4%). Although rare, co-occurrence of LEMS and MG have been previously reported in the literature and might render the diagnosis of LEMS more complicated [73, 74].

SCLC-LEMS is typically diagnosed more quickly after the onset of symptoms than NT-LEMS [11]. Reasons for this are not clear, but it may be that SCLC-LEMS has a more progressive course, which could shorten both patient presentation time and doctor delay in diagnosis [11].

Clinical features that strongly support a diagnosis of LEMS include progression of symptoms over weeks to months, spreading from proximal to distal muscles and in a caudocranial direction. Symptoms are likely to be symmetrical, with fluctuating severity over the course of the day. Patients may exhibit prominent autonomic symptoms, normal (or slightly elevated) creatine kinase, and cerebellar ataxia. Sensory symptoms or prominent pain are unlikely [2, 8].

While LEMS typically starts with leg weakness, which progresses in a caudocranial direction, MG typically begins with oculobulbar weakness, and muscle weakness spreads craniocaudally [75]. Autonomic dysfunction and diminished tendon reflexes are rarely seen with MG [75]. High-frequency RNS or post-exercise RNS below 60% may differentiate LEMS from MG [2].

Polymyositis or immunogenic necrotizing myopathy may also be suspected in patients with proximal symmetrical weakness, although these patients tend to have an absence of autonomic symptoms and instead experience pain, muscle atrophy, and raised creatine kinase, which are uncommonly seen in LEMS [2]. Difficulties getting out of a chair may be suggestive of early-stage Parkinson's disease, and subacute symptoms with abnormal electrophysiology may indicate neuropathy, Guillain-Barré syndrome, myotonic dystrophy type 2, or amyotrophic lateral sclerosis (ALS) [2]. However, these patients will likely have sensory symptoms or pronounced pain or myotonia not seen with LEMS. ALS patients will show fasciculations and progressive muscle atrophy.

Oncological screening

Clinical symptoms of LEMS nearly always precede detection of SCLC, so it is essential that screening for any underlying tumor begins as soon as a diagnosis of LEMS is

made (Fig. 1). With effective screening, 96% of cases of SCLC were diagnosed within a year of LEMS diagnosis [76]. Conventional radiography or bronchoscopy is unlikely to detect early-stage SCLC, so computed tomography (CT) of the thorax is warranted. If negative, this should be followed by ^{18}F -fluorodeoxyglucose-positron emission tomography (PET) or integrated PET/CT [77]. If the first screening is negative, oncological surveillance should be continued at 6-month intervals for at least 2 years after LEMS onset [77]. A simple SCLC prediction score, known as the DELTA-P score, has also been developed to guide assessment of the need for further screening early in the course of disease [14]. This tool scores a series of characteristics (age at onset, smoking status, bulbar involvement, weight loss, erectile dysfunction, and Karnofsky performance status) on a scale from 0 to 6, with a higher score indicative of greater SCLC risk. While this tool can prioritize high-risk patients for oncological cancer screening and reassure those with lower risk, neurologists should be vigilant about signs of possible SCLC in patients with a LEMS diagnosis.

Autoimmune disease screening

In the absence of malignancy, it is likely that the LEMS patient will have an underlying autoimmune disease. Assessment should be made if the patient is symptomatic (e.g., thyroid-stimulating hormone measurement for assessing comorbid thyroid dysfunction, or typical investigations if rheumatoid arthritis, systemic lupus erythematosus, systemic vasculitis, autoimmune inflammatory myopathies, or pernicious anemia is suspected); however, routine screening for autoimmune disease is not necessary.

Treatment

Symptomatic treatment of LEMS involves drugs that increase the release of neurotransmitters from the presynaptic terminal or that prolong the activity of ACh in the synapse.

The compound 4-aminopyridine (4-AP) has been used for the symptomatic treatment for LEMS [78]. Nevertheless, a few reports showed that this mono-aminopyridine has the ability to easily cross the blood–brain barrier causing central nervous system side effects such as seizures. Although limited, these results suggest that 4-AP might not be acceptable for the management of LEMS [6].

Unlike 4-AP, the potassium channel blocker 3,4-diaminopyridine or amifampridine is a viable and effective option for the symptomatic treatment of LEMS [2]. This agent may also directly stimulate the VGCC β -subunit,

potentiating neurotransmission [79]. In four randomized placebo-controlled trials in patients with LEMS, amifampridine showed a significant benefit in terms of muscle strength and CMAP amplitude, and was generally well tolerated [80, 81]. Amifampridine (in any of the possible formulations) was used as a symptomatic drug by the vast majority of the patients included in the EU registry, and steroids or immunosuppressants were used in about one-third of the patients [39]. Patients experiencing persistent symptoms may also require immunosuppressants or immunomodulators (e.g., prednisone plus azathioprine or intravenous immunoglobulin) [2]. Where a tumor is identified, oncological treatment should take priority.

Intravenous immunoglobulin (IVIg) is considered an effective alternative for the management of diseases with immune pathogenesis such as LEMS [82]. In fact, in a placebo-controlled crossover study IVIg was associated with a significant improvement in the amplitude of resting CMAP and a decline in serum VGCV antibodies [83]. Additionally, other case reports have demonstrated the benefit of IVIg for the short and long-term management of LEMS, especially as adjuvant therapy in patients with resistant muscle weakness [82].

Plasma exchange therapy is another alternative for the management of LEMS and other neurological disorders of autoimmune etiology [84]. Several case series reported improvements in clinical and electrophysiological outcome measures. Nevertheless, a case study showing only a transient decrease in the levels of serum VGCV after plasma exchange therapy suggests a limited effect of this therapy in patients with LEMS [84].

Conclusions

While rare, LEMS is a clinically important early indicator of the possible presence of SCLC or another underlying tumor. A LEMS diagnosis initiates a mandatory oncology screening and surveillance process, which may allow identification of early-stage cancer and initiation of cancer treatment, with a positive impact on patient outcomes. Furthermore, it should be remembered that the presence of LEMS has a significant impact on a patient's quality of life and ability to perform daily activities and therefore warrants timely diagnosis and appropriate treatment in and of itself.

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Compliance with ethical standards

Conflicts of interest BS has received speaker honoraria from BioMarin, Sanofi Genzyme, CSL Behring, and Amicus Therapeutics. He is a member of the neuromuscular advisory board of Audentes Therapeutics and has received an unrestricted research grant from Sanofi Genzyme. BE has received honoraria from BioMarin Europe Ltd, Novartis and the LFB Group. JD is a EUMEA Medical Affairs Senior Director for BioMarin Europe Ltd. RM has received honoraria for participation in clinical trials, advisory boards, congresses and travel from Alexion Pharmaceuticals, Biomarin Europe Ltd. and Catalys.

References

- Eaton LM, Lambert EH (1957) Electromyography and electric stimulation of nerves in diseases of motor unit; observations on myasthenic syndrome associated with malignant tumors. *J Am Med Assoc* 163:1117–1124
- Titulaer MJ, Lang B, Verschuuren JJ (2011) Lambert–Eaton myasthenic syndrome: from clinical characteristics to therapeutic strategies. *Lancet Neurol* 10:1098–1107
- Lambert EH, Elmqvist D (1971) Quantal components of end-plate potentials in the myasthenic syndrome. *Ann N Y Acad Sci* 183:183–199
- Wirtz PW, Smallegange TM, Wintzen AR, Verschuuren JJ (2002) Differences in clinical features between the Lambert–Eaton myasthenic syndrome with and without cancer: an analysis of 227 published cases. *Clin Neurol Neurosurg* 104:359–363
- Harms L, Sieb JP, Williams AE et al (2012) Long-term disease history, clinical symptoms, health status, and healthcare utilization in patients suffering from Lambert Eaton myasthenic syndrome: results of a patient interview survey in Germany. *J Med Econ* 15:521–530
- Quartel A, Turbeville S, Lousbury D (2010) Current therapy for Lambert–Eaton myasthenic syndrome: development of 3,4-diaminopyridine phosphate salt as first-line symptomatic treatment. *Curr Med Res Opin* 26:1363–1375
- Maddison P, Lang B, Mills K, Newsom-Davis J (2001) Long term outcome in Lambert–Eaton myasthenic syndrome without lung cancer. *J Neurol Neurosurg Psychiatry* 70:212–217
- Evoli A, Liguori R, Romani A et al (2014) Italian recommendations for Lambert–Eaton myasthenic syndrome (LEMS) management. *Neurol Sci* 35:515–520
- Orphanet (2013) Lambert–Eaton myasthenic syndrome. http://www.orpha.net/consor/cgi-bin/OC_Exp.php?Lng=EN&Expert=43393. Accessed 10 Feb 2017
- Wirtz PW, Nijhuis MG, Sotodeh M et al (2003) The epidemiology of myasthenia gravis, Lambert–Eaton myasthenic syndrome and their associated tumors in the northern part of the province of South Holland. *J Neurol* 250:698–701
- Wirtz PW, van Dijk JG, van Doorn PA et al (2004) The epidemiology of the Lambert–Eaton myasthenic syndrome in the Netherlands. *Neurology* 63:397–398
- Abenroth DC, Smith AG, Greenlee JE, Austin SD, Clardy SL (2016) Lambert–Eaton myasthenic syndrome (LEMS): epidemiology and therapeutic response in the national Veterans Affairs (VA) population. *Muscle Nerve* doi:10.1002/mus.25520
- O’Neill JH, Murray NM, Newsom-Davis J (1988) The Lambert–Eaton myasthenic syndrome. A review of 50 cases. *Brain* 111(Pt 3):577–596
- Titulaer MJ, Maddison P, Sont JK et al (2011) Clinical Dutch–English Lambert–Eaton myasthenic syndrome (LEMS) tumor association prediction score accurately predicts small-cell lung cancer in the LEMS. *J Clin Oncol* 29:902–908
- Sher E, Comola M, Nemni R, Canal N, Clementi F (1990) Calcium channel autoantibody and non-small-cell lung cancer in patients with Lambert–Eaton syndrome. *Lancet* 335:413
- Zivaljevic M, Popovic S, Vujkov T (2005) Lambert–Eaton myasthenic syndrome—a rare manifestation of paraneoplastic syndrome in ovarian cancer—case report. *Med Pregled* 58:495–497
- Arai H, Inui K, Hashimoto K et al (2012) Lung adenocarcinoma with Lambert–Eaton myasthenic syndrome indicated by voltage-gated calcium channel: a case report. *J Med Case Rep* 6:281
- Bombelli F, Lispi L, Calabro F, Corsi FM, Petrucci A (2015) Lambert–Eaton myasthenic syndrome associated to Merkel cell carcinoma: report of a case. *Neurol Sci* 36:1491–1492
- Collins DR, Connolly S, Burns M et al (1999) Lambert–Eaton myasthenic syndrome in association with transitional cell carcinoma: a previously unrecognized association. *Urology* 54:162
- Delahunt B, Abernethy DA, Johnson CA, Nacey JN (2003) Prostate carcinoma and the Lambert–Eaton myasthenic syndrome. *J Urol* 169:278–279
- Galton C, Thomson D, Boyle R (1998) Lambert–Eaton myasthenic syndrome and non-pulmonary small cell carcinoma. *J Neurol Neurosurg Psychiatry* 64:819–820
- Grommes C, Preston DC, Al-Kadhimi Z, Alshehlee A (2008) Lambert–Eaton syndrome with large-cell neuroendocrine carcinoma of the lung. *Muscle Nerve* 37:786–789
- Katirji B (2000) Lambert–Eaton myasthenic syndrome: a harbinger to transitional cell carcinoma of the urinary bladder. *J Clin Neuromuscul Dis* 1:134–136
- Milanez FM, Pereira CA, Trindade PH, Milinavicius R, Coletta EN (2008) Lung adenocarcinoma, dermatomyositis, and Lambert–Eaton myasthenic syndrome: a rare combination. *J Bras Pneumol* 34:333–336
- Monteiro C, Moreira I, Lima JL, Santos E (2015) Lambert–Eaton myasthenic syndrome and prostatic adenocarcinoma. *Neurol Sci* 36:2145–2146
- Nalbantoglu M, Kose L, Uzun N et al (2015) Lambert–Eaton myasthenic syndrome associated with thymic neuroendocrine carcinoma. *Muscle Nerve* 51:936–938
- Romics L Jr, McNamara B, Cronin PA et al (2011) Unusual paraneoplastic syndromes of breast carcinoma: a combination of cerebellar degeneration and Lambert–Eaton myasthenic syndrome. *Ir J Med Sci* 180:569–571
- Shiple E, Krim E, Deminiere C, Laguery A (2008) Lambert–Eaton myasthenic syndrome associated with vocal cord carcinoma. *Rev Neurol (Paris)* 164:72–76
- Simmons DB, Duginski TM, McClean JC, Amato AA, Sladky JH (2016) Lambert–Eaton myasthenic syndrome and merkel cell carcinoma. *Muscle Nerve* 53:325–326
- Argov Z, Shapira Y, Averbuch-Heller L, Wirguin I (1995) Lambert–Eaton myasthenic syndrome (LEMS) in association with lymphoproliferative disorders. *Muscle Nerve* 18:715–719
- Titulaer MJ, Wirtz PW, Kuks JB et al (2008) The Lambert–Eaton myasthenic syndrome 1988–2008: a clinical picture in 97 patients. *J Neuroimmunol* 201–202:153–158
- Payne M, Bradbury P, Lang B et al (2010) Prospective study into the incidence of Lambert Eaton myasthenic syndrome in small cell lung cancer. *J Thorac Oncol* 5:34–38
- Hajjar M, Markowitz J, Darras BT et al (2014) Lambert–Eaton syndrome, an unrecognized treatable pediatric neuromuscular disorder: three patients and literature review. *Pediatr Neurol* 50:11–17
- Titulaer MJ, Verschuuren JJ (2008) Lambert–Eaton myasthenic syndrome: tumor versus nontumor forms. *Ann N Y Acad Sci* 1132:129–134

35. Wirtz PW, Bradshaw J, Wintzen AR, Verschuuren JJ (2004) Associated autoimmune diseases in patients with the Lambert–Eaton myasthenic syndrome and their families. *J Neurol* 251:1255–1259
36. Peris P, Del Olme J, Gratacos J, Munoz J (1990) The Lambert–Eaton myasthenic syndrome in association with rheumatoid arthritis. *Br J Rheumatol* 29:75–76
37. Deodhar A, Norden J, So Y, Bennett R (1996) The association of systemic lupus erythematosus and Lambert–Eaton myasthenic syndrome. *J Rheumatol* 23:1292–1294
38. Young JD, Leavitt JA (2015) Lambert–Eaton myasthenic syndrome: ocular signs and symptoms. *J Neuroophthalmol* 36:20–22
39. Mantegazza R, Meisel A, Sieb JP, Masson GL, Desnuelle C, Essing M (2015) The European LEMS Registry: baseline demographics and treatment approaches. *Neurol Ther* 4:105–124
40. Maddison P, Newsom-Davis J, Mills KR, Souhami RL (1999) Favourable prognosis in Lambert–Eaton myasthenic syndrome and small-cell lung carcinoma. *Lancet* 353:117–118
41. Maddison P, Silcocks P (2010) Prospective study of Lambert–Eaton myasthenic syndrome in small cell lung cancer. *J Thorac Oncol* 5:1309–1310
42. Wirtz PW, Lang B, Graus F et al (2005) P/Q-type calcium channel antibodies, Lambert–Eaton myasthenic syndrome and survival in small cell lung cancer. *J Neuroimmunol* 164:161–165
43. Dolphin AC (2006) A short history of voltage-gated calcium channels. *Br J Pharmacol* 147(Suppl 1):S56–S62
44. Lennon VA, Lambert EH, Whittingham S, Fairbanks V (1982) Autoimmunity in the Lambert–Eaton myasthenic syndrome. *Muscle Nerve* 5:S21–S25
45. Lennon VA, Kryzer TJ, Griesmann GE et al (1995) Calcium-channel antibodies in the Lambert–Eaton syndrome and other paraneoplastic syndromes. *N Engl J Med* 332:1467–1474
46. Hajela RK, Huntoon KM, Atchison WD (2015) Lambert–Eaton syndrome antibodies target multiple subunits of voltage-gated Ca²⁺ channels. *Muscle Nerve* 51:176–184
47. Fukunaga H, Engel AG, Lang B, Newsom-Davis J, Vincent A (1983) Passive transfer of Lambert–Eaton myasthenic syndrome with IgG from man to mouse depletes the presynaptic membrane active zones. *Proc Natl Acad Sci USA* 80:7636–7640
48. Roberts A, Perera S, Lang B, Vincent A, Newsom-Davis J (1985) Paraneoplastic myasthenic syndrome IgG inhibits 45Ca²⁺ flux in a human small cell carcinoma line. *Nature* 317:737–739
49. Benatar M, Blaes F, Johnston I et al (2001) Presynaptic neuronal antigens expressed by a small cell lung carcinoma cell line. *J Neuroimmunol* 113:153–162
50. Sher E, Gotti C, Canal N et al (1989) Specificity of calcium channel autoantibodies in Lambert–Eaton myasthenic syndrome. *Lancet* 2:640–643
51. Wray DW, Lang B, Newsom-Davis J, Peers C (1989) Antibodies against calcium channels in the Lambert–Eaton myasthenic syndrome. *Ann N Y Acad Sci* 560:269–277
52. Wirtz PW, Roep BO, Schreuder GM et al (2001) HLA class I and II in Lambert–Eaton myasthenic syndrome without associated tumor. *Hum Immunol* 62:809–813
53. Fukuoka T, Engel AG, Lang B et al (1987) Lambert–Eaton myasthenic syndrome: I. Early morphological effects of IgG on the presynaptic membrane active zones. *Ann Neurol* 22:193–199
54. Molenaar PC, Newsom-Davis J, Polak RL, Vincent A (1982) Eaton–Lambert syndrome: acetylcholine and choline acetyltransferase in skeletal muscle. *Neurology* 32:1061–1065
55. Nishimune H, Sanes JR, Carlson SS (2004) A synaptic laminin-calcium channel interaction organizes active zones in motor nerve terminals. *Nature* 432:580–587
56. Chen J, Billings SE, Nishimune H (2011) Calcium channels link the muscle-derived synapse organizer laminin beta2 to Bassoon and CAST/Erc2 to organize presynaptic active zones. *J Neurosci* 31:512–525
57. Waterman SA, Lang B, Newsom-Davis J (1997) Effect of Lambert–Eaton myasthenic syndrome antibodies on autonomic neurons in the mouse. *Ann Neurol* 42:147–156
58. Nakao YK, Motomura M, Fukudome T et al (2002) Seronegative Lambert–Eaton myasthenic syndrome: study of 110 Japanese patients. *Neurology* 59:1773–1775
59. Oh SJ, Hatanaka Y, Claussen GC, Sher E (2007) Electrophysiological differences in seropositive and seronegative Lambert–Eaton myasthenic syndrome. *Muscle Nerve* 35:178–183
60. Takamori M (2008) Lambert–Eaton myasthenic syndrome: search for alternative autoimmune targets and possible compensatory mechanisms based on presynaptic calcium homeostasis. *J Neuroimmunol* 201–202:145–152
61. Takamori M, Takahashi M, Yasukawa Y et al (1995) Antibodies to recombinant synaptotagmin and calcium channel subtypes in Lambert–Eaton myasthenic syndrome. *J Neurol Sci* 133:95–101
62. Titulaer MJ, Klooster R, Potman M et al (2009) SOX antibodies in small-cell lung cancer and Lambert–Eaton myasthenic syndrome: frequency and relation with survival. *J Clin Oncol* 27:4260–4267
63. Sabater L, Titulaer M, Saiz A et al (2008) SOX1 antibodies are markers of paraneoplastic Lambert–Eaton myasthenic syndrome. *Neurology* 70:924–928
64. Odabasi Z, Demirci M, Kim DS et al (2002) Postexercise facilitation of reflexes is not common in Lambert–Eaton myasthenic syndrome. *Neurology* 59:1085–1087
65. Weingarten TN, Araka CN, Mogensen ME et al (2014) Lambert–Eaton myasthenic syndrome during anesthesia: a report of 37 patients. *J Clin Anesth* 26:648–653
66. Oh SJ, Kurokawa K, Claussen GC, Ryan HF Jr (2005) Electrophysiological diagnostic criteria of Lambert–Eaton myasthenic syndrome. *Muscle Nerve* 32:515–520
67. Medicine AQCAAoE (2001) Practice parameter for repetitive nerve stimulation and single fiber EMG evaluation of adults with suspected myasthenia gravis or Lambert–Eaton myasthenic syndrome: summary statement. *Muscle Nerve* 24:1236–1238
68. Oh SJ, Ohira M (2013) Single-fiber EMG and clinical correlation in Lambert–Eaton myasthenic syndrome. *Muscle Nerve* 47:664–667
69. Leys K, Lang B, Johnston I, Newsom-Davis J (1991) Calcium channel autoantibodies in the Lambert–Eaton myasthenic syndrome. *Ann Neurol* 29:307–314
70. Pellkofer HL, Armbruster L, Krumbholz M et al (2008) Lambert–Eaton myasthenic syndrome differential reactivity of tumor versus non-tumor patients to subunits of the voltage-gated calcium channel. *J Neuroimmunol* 204:136–139
71. Stich O, Klages E, Bischler P et al (2012) SOX1 antibodies in sera from patients with paraneoplastic neurological syndromes. *Acta Neurol Scand* 125:326–331
72. Tschernatsch M, Singh P, Gross O et al (2010) Anti-SOX1 antibodies in patients with paraneoplastic and non-paraneoplastic neuropathy. *J Neuroimmunol* 226:177–180
73. Oh SJ, Dwyer DS, Bradley RJ (1987) Overlap myasthenic syndrome: combined myasthenia gravis and Eaton–Lamert syndrome. *Neurology* 37:1411–1414
74. Newsom-Davis J, Leys K, Vincent A et al (1991) Immunological evidence for the co-existence of the Lambert–Eaton myasthenic syndrome and myasthenia gravis in two patients. *J Neurol Neurosurg Psychiatry* 54:452–453
75. Hulsbrink R, Hashemolhosseini S (2014) Lambert–Eaton myasthenic syndrome—diagnosis, pathogenesis and therapy. *Clin Neurophysiol* 125:2328–2336
76. Titulaer MJ, Wirtz PW, Willems LN et al (2008) Screening for small-cell lung cancer: a follow-up study of patients with Lambert–Eaton myasthenic syndrome. *J Clin Oncol* 26:4276–4281
77. Titulaer MJ, Soffietti R, Dalmau J et al (2011) Screening for tumors in paraneoplastic syndromes: report of an EFNS task force. *Eur J Neurol* 18:19 (e13)

78. Verschuuren JJG, Wirtz PW, Titulaer MJ et al (2006) Available treatment options for the management of Lambert–Eaton myasthenic syndrome. *Opin Pharmacother* 7:1323–1336
79. Wu ZZ, Li DP, Chen SR, Pan HL (2009) Aminopyridines potentiate synaptic and neuromuscular transmission by targeting the voltage-activated calcium channel beta subunit. *J Biol Chem* 284:36453–36461
80. Wirtz PW, Titulaer MJ, Gerven JM, Verschuuren JJ (2010) 3,4-diaminopyridine for the treatment of Lambert–Eaton myasthenic syndrome. *Expert Rev Clin Immunol* 6:867–874
81. Keogh M, Sedehizadeh S, Maddison P (2011) Treatment for Lambert–Eaton myasthenic syndrome. *Cochrane Database Syst Rev* CD003279 doi:[10.1002/14651858.CD003279.pub3](https://doi.org/10.1002/14651858.CD003279.pub3)
82. Illa I (2005) IVIg in myasthenia gravis, Lambert Eaton myasthenic syndrome and inflammatory myopathies: current status. *J Neurol* 252(Suppl 1):I14–I18
83. Bain PG, Motomura M, Newsom-Davis J et al (1996) Effects of intravenous immunoglobulin on muscle weakness and calcium-channel autoantibodies in the Lambert–Eaton myasthenic syndrome. *Neurology* 47:678–683
84. Lehmann HC, Hartung HP, Hetzel GR et al (2006) Plasma exchange in neuroimmunological disorders: part 2. Treatment of neuromuscular disorders. *Arch Neurol* 63:1066–1071