



Complete mitogenome data for the Serbian population: the contribution to high-quality forensic databases

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Abstract

Mitochondrial genome (mtDNA) is a valuable resource in resolving various human forensic casework. The usage of variability of complete mtDNA genomes increases their discriminatory power to the maximum and enables ultimate resolution of distinct maternal lineages. However, their wider employment in forensic casework is nowadays limited by the lack of appropriate reference database. In order to fill in the gap in the reference data, which, considering Slavic-speaking populations, currently comprises only mitogenomes of East and West Slavs, we present mitogenome data for 226 Serbians, representatives of South Slavs from the Balkan Peninsula. We found 143 (sub)haplogroups among which West Eurasian ones were dominant. The percentage of unique haplotypes was 85%, and the random match probability was as low as 0.53%. We support previous findings on both high levels of genetic diversity in the Serbian population and patterns of genetic differentiation among this and ten studied European populations. However, our high-resolution data supported more pronounced genetic differentiation among Serbians and two Slavic populations (Russians and Poles) as well as expansion of the Serbian population after the Last Glacial Maximum and during the Migration period (fourth to ninth century A.D.), as inferred from the Bayesian skyline analysis. Phylogenetic analysis of haplotypes found in Serbians contributed towards the improvement of the worldwide mtDNA phylogeny, which is essential for the interpretation of the mtDNA casework.

Keywords Complete mitogenomes · Demographic changes · Molecular phylogeography · Serbian population

Introduction

The mitochondrial genome (mtDNA, mitogenome) is particularly suitable in forensic casework when STR profiling

cannot be performed due to the degraded and/or scarce nuclear DNA [1, 2]. Given its matrilineal inheritance, individuals may be identified along maternal lineage across many generations [3] and that enables, for instance, testing for putative exclusion

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of individuals during the person identification [4]. Traditionally, mtDNA typing is based on ~ 600 bp of the hypervariable segments I and II (HVS-I and HVS-II) of the control region (CR, ~ 1100 bp) (e.g., [2, 5, 6]). However, the information available in these segments sometimes does not have sufficient discriminatory power to resolve distinct maternal lineages, and that has led to extensions of the mtDNA analyses in some cases to the third hypervariable segment, HVS-III, or to the entire CR [7–9]. Further extension to the complete mtDNA genome, generated today by Sanger or next generation sequencing, enables usage of sequence variation in the coding region as well, and thus analysis at the maximum resolution [10–12].

One of the essential aspects in forensic casework is interpretation of obtained mtDNA profiles, their comparison against the reference database for a possible match, and estimation of their frequency in the context of the relevant population sample [13, 14]. EMPOP database (<https://empop.online>) has specifically been developed for forensic applications, and currently, it comprises 48,572 quality-controlled mitotypes [15]. The number of complete mitogenomes in EMPOP, however, is still insufficient [16]. Given the continuous advancement in sequencing technologies, it appears that the wider usage of the complete mtDNA typing in forensic casework is more limited by the lack of appropriate database than by the costs of analyses which are decreasing over time [16].

The Slavs, which constitute more than one third of Europeans and inhabit a rather large part of the continent [17], are an ethnolinguistic group with a complex history which has been studied at the molecular level as well (e.g., [18–25]). They are characterized by a marked geographical stratification to South-, East-, and West Slavic-speaking groups. South Slavs inhabit the Balkan Peninsula, a region characterized by exceptionally turbulent historic, demographic, and other processes during the entire history of humans. It served, in addition, as a starting point for the postglacial recolonization of Europe [24, 26, 27] and for human migrations in later periods. However, the representation of the mtDNA diversity of South Slavs in the EMPOP database is rather limited since high-quality HVS-I and HVS-II haplotypes found in individuals from five populations are only available: Bosnia and Herzegovina [28], Bulgaria [29], Northern Macedonia [30], Serbia [18], and Slovenia [28, 31]. On the other hand, complete mitogenomes of Russians and Poles, representatives of East and West Slavs, respectively, have recently been included into this database [23]. Since the enrichment and refinement of the complete mtDNA reference database contributes towards the improvement of the worldwide mtDNA phylogeny, which is essential for the interpretation of the mtDNA casework [32], our aim was to depict complete mtDNA diversity of Serbians, representatives of South Slavs.

Materials and methods

Population samples

We analyzed complete mitochondrial genomes of 226 maternally unrelated individuals from the general population of Serbia, of which 170 were sequenced de novo and 56 were taken from our previous work (GenBank accession numbers KT697997–KT698032, KM096761–KM096763, and KM096765–KM096781) [18, 33]. For comparative purposes, we used 3145 complete mitogenomes originating from ten Eurasian populations. All mitogenomes belonging to one population were pooled together and used as a representative sample of that population. In that way, we included into our analysis complete mitogenomes of 1998 Danes [34], 53 French [35], 116 Poles [23, 36], 401 Russians [23, 35], 118 Tuscans [35, 37], 80 Hungarians [36], 114 Estonians [38], 101 Spaniards [39], 91 Sardinians [35, 40], and 73 Volga Tatars [41].

Samples (saliva or blood) were collected with written informed consent from all individual participants included in the study, and genealogical information regarding the place of birth of participant's maternal grandparents was obtained. The Serbian ancestry was confirmed for at least two generations, and, according to the geographical origin of donor's maternal grandmothers, they were classified into the following regions of Serbia: Belgrade ($N = 16$), Central Serbia ($N = 36$), Eastern Serbia ($N = 19$), Kosovo and Metohija ($N = 11$), Southern Serbia ($N = 39$), Vojvodina ($N = 17$), and Western Serbia ($N = 49$), or to the neighboring countries: Bosnia and Herzegovina ($N = 13$), Croatia ($N = 18$), Northern Macedonia ($N = 1$), and Montenegro ($N = 7$).

The study was approved by the Research Ethics Committees of the Institute of Molecular Genetics and Genetic Engineering, University of Belgrade, and Institute of Biological Problems of the North, Russian Academy of Sciences.

DNA extraction and sequencing of complete mtDNA genomes

High-quality genomic DNA was obtained from saliva (163 samples) according to Quinque et al. [42] or from blood (63 samples) using QIAamp® DNA Blood Mini Kit (QIAGEN GmbH, Hilden, Germany) in the Laboratory for Human Molecular Genetics at the Institute of Molecular Genetics and Genetic Engineering, University of Belgrade. Sequencing of complete mtDNA genomes was carried out in the Genetics Laboratory at the Institute of Biological Problems of the North, Russian Academy of Sciences, Magadan, Russia (saliva samples), and in the Division of Molecular and Forensic Genetics, Department of Forensic Medicine, Ludwik Rydygier Collegium Medicum, Nicolaus Copernicus University, Torun, Poland (blood samples).

Sequencing was performed using ABI3500xL and ABI3130xL Genetic Analyzers, according to the methods described in Torroni et al. [43] and Fendt et al. [44]. As recommended by Just et al. [16], all cases of point heteroplasmy were confirmed through additional independent PCRs and sequencing reactions.

De novo sequenced mitogenomes from Serbian population were deposited in GenBank under accession numbers MK134267-MK134373 and MK617218-MK617280. The complete dataset of 226 Serbian mitogenomes went through the EMPOP quality check [15] and was made available for forensic searches in the EMPOP database (www.empop.org) under the accession number EMP00739.

Haplogroup estimation

Generated sequences were compared with the revised Cambridge reference sequence (rCRS) [45]; haplotypes were defined with the HaploSearch software [46], and sequence alignment was performed following the phylogenetic concept and the recommendations of the International Society for Forensic Genetics (ISFG) [6, 47, 48]. SAM2 [49], implemented in EMPOP [15], was used for alignment and haplogroup estimation according to PhyloTree build 17 [50]. Along with the assignment of haplotypes to the most recent common ancestor (MRCA) by SAM2, which is a conservative estimate suitable in forensics, we also performed haplotype assignment through reconstruction of most-parsimonious trees using mtPhyl v4.015 software (<http://eltsov.org>). Since this version of program does not use the updated mtDNA phylogeny available in PhyloTree build 17, the trees were modified manually according to the latest PhyloTree build 17 and the following literature [18, 23, 25, 33, 36, 51–55]. New subclades were defined when they comprised at least two different mitogenomes having at least one shared mutation which is not characterized as a hotspot [32]. The length variations at nps 303-315, 522-524, 573-576, and 16180-16193, as well as polymorphism at np 16519 and A-C transversions at nps 16182 and 16183, were excluded from phylogenetic analysis.

Statistical analysis

Basic parameters of genetic diversity in Serbian and other studied populations (number of haplotypes, H ; number of polymorphic sites, P ; haplotype diversity, HD ; nucleotide diversity ND ; and mean pairwise difference, MPD) were assessed with Arlequin 3.5.2.2 [56]. The same tool was used for assessing genetic differentiation among populations via the analysis of molecular variance (AMOVA) and estimating pairwise population and overall F_{ST} values. Point heteroplasmy were treated as differences, indels were excluded, and statistical significance of all tests was assessed

with 10,000 permutations. The matrix of pairwise population F_{ST} values was visualized by two-dimensional scaling (MDS) using STATISTICA10 (StatSoft Inc., Tulsa, OK, USA).

The probability that two randomly selected individuals from a population share the identical mtDNA haplotype, i.e., the random match probability (RMP), was calculated according to Stoneking et al. [57].

Tajima's D test implemented in Arlequin was performed to detect departures from population equilibrium. The significance of the test statistics was assessed with 10,000 bootstrap replicates. DnaSP 5.10.01 software [58] was used to assess the demographic history of populations by a mismatch distribution analysis.

Demographic changes over time were assessed from the Bayesian skyline plot (BSP) analysis carried out with BEAST 1.10.4 [59]. We used the HKY+G+I model of sequence evolution which had the best fit to our dataset as inferred from the Akaike information criterion (AIC) value calculated in MEGA 6.06 [60]. We used the strict molecular clock which, according to the `uclsddev` parameter, cannot be rejected for our dataset, and mutation rate of 1.665×10^{-8} [61]. All parameters were sampled once every 1000 steps from 150 million of Markov chain Monte Carlo (MCMC) steps. Tracer v.1.7.1 [61, 62] was used to assess acceptable mixing, likelihood stationarity of the MCMC chain, and adequate effective sample sizes for each parameter (≥ 200).

Results

mtDNA haplogroup assignment

SAM2 classified 226 complete Serbian mitogenomes into 143 different mtDNA (sub)haplogroups (Table S1) following the nomenclature of PhyloTree build 17, the widely accepted phylogenetic tree of human mtDNA lineages [50]. As expected, Serbian maternal gene pool was composed mainly of West Eurasian (sub)haplogroups: the prevalent haplogroup was H (47.78%), followed by U (16.37%), J (8.85%), K (6.19%), and T (5.31%). Less frequent lineages found in Serbians comprised N1 (including I) (3.54%), W (3.1%), HV (excluding H and V) (3.1%), V (2.2%), X2 (1.32%), and R0a (0.44%). In addition, two mitogenomes belonged to East Asian haplogroup D4j (0.88%) and one to the "European" branch of African haplogroup L2a1k (0.44%).

We subsequently compared the results of haplogroup assignment obtained with SAM2 and mtPhyl. In this respect, it is worth noting that both tools provide haplogroup estimates using the PhyloTree build 17; they employ, however, different approaches: SAM2 does not rely on strict decision trees but rather on variation and fluctuation rates for all sites and regions observed in substantial number of confirmed haplotypes, while mtPhyl uses maximum parsimony for

haplogrouping. The assignment of haplogroups by SAM2 and mtPhyl was generally concordant (Table S1). Nevertheless, in a few cases where multiple haplogroup assignments were feasible by SAM2, haplogrouping by mtPhyl corresponded to one of them but not to the most conservative estimate (MRCA) (Table S1, samples 20_Sb, 69_Sb, 73_Sb, 86_Sb, 92_Sb, 143_Sb, 223_Sb, and 252_Sb). For example, mtPhyl assigned the haplotype 20_Sb to haplogroup H1c9'9 (H1c + 152) whereas SAM2 assigned it to MRCA H1c.

In addition, further refinement of haplogroups found outside of the widely accepted PhyloTree build 17 were estimated with mtPhyl by updating the phylogeny according to new findings from the present study as well as those found in available literature [18, 23, 25, 33, 36, 51–55] (Fig. S1). As a result, 22 haplotypes were assigned to 10 new subclades, namely H1cm, H5a10, H7j, H13d, H15b3, H15b3a, H30a1, H110, V6a, and W3b2, not defined previously in PhyloTree build 17 and available literature (Fig. S1 and Table S1). Subclades which comprise haplotypes from the Serbian population that were defined previously in other publications but not in PhyloTree build 17 encompass, for instance, H1c23a, H24a3 [23], H8b1b [52], X2q1a [25], U7b5 [55], U8a1a1a2, and U4d2b [33].

mtDNA diversity in Serbian population

The parameters describing mtDNA diversity in Serbians and other studied European populations are shown in Table 1. The number of unique haplotypes found in 226 Serbians was 192 (85.0%) while 15 haplotypes (15.0%) were observed more than once (Table S2).

Point heteroplasmies (PHP) were observed in 9 Serbian individuals (3.98%) as single occurrences (Table S3). They were all transitions, of which three were present in the control

region and six were in the coding region of mtDNA (Table S3). Three PHPs recorded in Serbian mitogenomes were found in EMPOP database (198Y, 8843Y, and 15466Y).

Length heteroplasmy (LHP) was observed in more than 50% of the samples, with 27 mitogenomes (11.95%) showing more than one LHP (Table S4). Majority of length variants were found in polycytosine stretches of the HVS-II (48.2%) and HVS-I (12.8%), while LHP of C-stretch at np 573 in HVS-III was observed in six instances (2.65%). In the coding region, LHP was observed at position 965 in three mitogenomes (1.3%). Upon inclusion of indel polymorphisms into the analysis, the number of identical haplotypes in the sample was reduced to 12, improving slightly Serbian mtDNA haplotype resolution (87.6% with indels vs. 85.0% without indels).

The RMP for mitogenomic data of Serbians was 0.53%. The lowest RMP value was observed in Danes (0.06%), represented by 1998 mitogenomes, and the highest in French (1.89%), represented only by 53 complete mtDNA genomes (Table 1).

mtDNA pairwise nucleotide differences in Serbian population

The number of mean pairwise differences (MPDs) in Serbian population of 27.82 was in the range of that observed in other European populations (Table 1). The lowest MPD value (27.00) was detected in Sardinian population and the highest (35.18) in Volga Tatars (Table 1), as already shown by Malyarchuk et al. [23].

Negative and statistically significant Tajima's *D* values observed in Serbian and other studied European populations indicated their recent expansions. However, all populations were characterized by a bimodal distribution of pairwise

Table 1 Parameters of genetic diversity in studied populations based on complete mitogenome sequences

Population	<i>N</i>	<i>H</i>	<i>P</i>	HD	ND	MPD	RMP	Tajima's <i>D</i> (<i>p</i> value)
Serbian	226	207	790	0.999 ± 0.000	0.0017 ± 0.0008	27.82	0.0053	− 2.51 (< 0.001)
Polish	116	111	615	0.999 ± 0.001	0.0019 ± 0.0009	31.83	0.0095	− 2.43 (< 0.001)
Russian	401	387	1106	0.999 ± 0.000	0.0017 ± 0.0008	29.05	0.0027	− 2.54 (< 0.001)
Tuscan	118	117	730	0.999 ± 0.001	0.0019 ± 0.0009	30.83	0.0086	− 2.59 (< 0.001)
Hungarian	80	78	475	0.999 ± 0.002	0.0018 ± 0.0009	30.33	0.0131	− 2.35 (< 0.001)
Estonian	114	108	493	0.999 ± 0.001	0.0017 ± 0.0008	28.39	0.0097	− 2.32 (< 0.001)
Spanish	101	97	541	0.999 ± 0.001	0.0018 ± 0.0009	29.96	0.0107	− 2.42 (< 0.001)
Sardinian	91	77	296	0.996 ± 0.003	0.0016 ± 0.0008	27.00	0.0153	− 1.78 (< 0.05)
Volga Tatars	73	68	509	0.998 ± 0.003	0.0021 ± 0.0010	35.18	0.0156	− 2.32 (< 0.001)
Danish	1998	1878	2108	0.999 ± 0.000	0.0017 ± 0.0008	28.05	0.0006	− 2.55 (< 0.001)
French	53	53	305	1.000 ± 0.004	0.0016 ± 0.0008	27.39	0.0189	− 2.11 (< 0.01)

N number of individuals, *H* number of haplotypes, *P* number of polymorphic sites, *HD* haplotype diversity, *ND* nucleotide diversity, *MPD* mean pairwise difference, *RMP* random match probability

nucleotide differences (data not shown). In Serbian population, the number of pairwise differences ranged from 0 to 75, and two peaks had modes at 12 and 32 differences (Fig. S2).

Differentiation of European populations based on complete mtDNAs

AMOVA revealed low but statistically significant genetic differentiation among populations ($F_{ST} = 0.74\%$, $p = 0$, Table S5). The pairwise population F_{ST} values between Serbian and other studied populations were generally low but significant in the case of five populations (Danes, Poles, Russians, Sardinians, and Volga Tatars) (Table S6). In MDS plot constructed using pairwise population F_{ST} values, Sardinian and Volga Tatar populations were outliers, while among other European populations, two genetically close groups of populations were observed (Fig. 1). One group comprised Romance-speaking populations from Western and Southwestern Europe (Tuscan, Spanish, and French) and the second comprised populations from Central and Eastern Europe (Hungarian, Polish, and Estonian). Serbian and Russian populations were positioned between these two groups.

Bayesian skyline plot in Serbian population

BSP for 226 Serbians was performed to detect changes in population size over time (Fig. 2). An expansion of a population until 45.8 kya (95% CI 41.9–47.4 kya) was followed by a more stable population size with a slow and continuous decrease. The lowest population size was observed during the Last Glacial Maximum (LGM), around 23.7 kya (95% CI 22.9–24.5 kya) and was followed by a steady growth until 7.1 kya (95% CI 7.1–8.7 kya) when the highest population size was recorded. This time frame corresponds to the post-LGM expansion which

was followed by a decline in population size until 1.6 kya (95% CI 1.6–2.4 kya) when the lowest population size was reached. The subsequent growth of the population size coincides with the Migration period that occurred during the Early Middle Ages (fourth to ninth century A.D.).

Discussion

Mitochondrial genome variability, especially that present in hypervariable regions of the control region, has been successfully used in forensics for more than 30 years [1, 2]. Nowadays, a trend towards the usage of variability of complete mitogenomes is evident in the forensic community because that enables a maximum increase in discriminatory power of this molecular tool [12, 16] thus providing ultimate resolution of distinct maternal lineages. However, the wider employment of complete mitogenomes in forensic casework is currently limited by the lack of appropriate reference database [16]. For instance, only mitogenomes found in East Slavs (Russians) and West Slavs (Poles) are currently available in the EMPOP database [23] despite the fact that Slavs are an important component of the extant European population. In order to overcome this issue and to contribute to a better representation of the Slavic-speaking populations in the EMPOP database, we present complete mitogenome data for 226 Serbians, representatives of South Slavs, which is now available in this centrally curated database essential for the forensic casework.

Complete Serbian mitogenomes were classified into 143 different mtDNA (sub)haplogroups of predominantly West Eurasian origin, with East Asian (D4j) and African (L2a1k) haplogroups observed at a low frequency. It is worth mentioning that although haplogroup assignment obtained with mtPhyl and SAM2 was generally concordant, we detected

Fig. 1 Multidimensional scaling plot of F_{ST} distances between Serbian population and selected European populations based on complete mtDNA sequences (stress value 0.00031). Population pairwise F_{ST} values are presented in Table S6

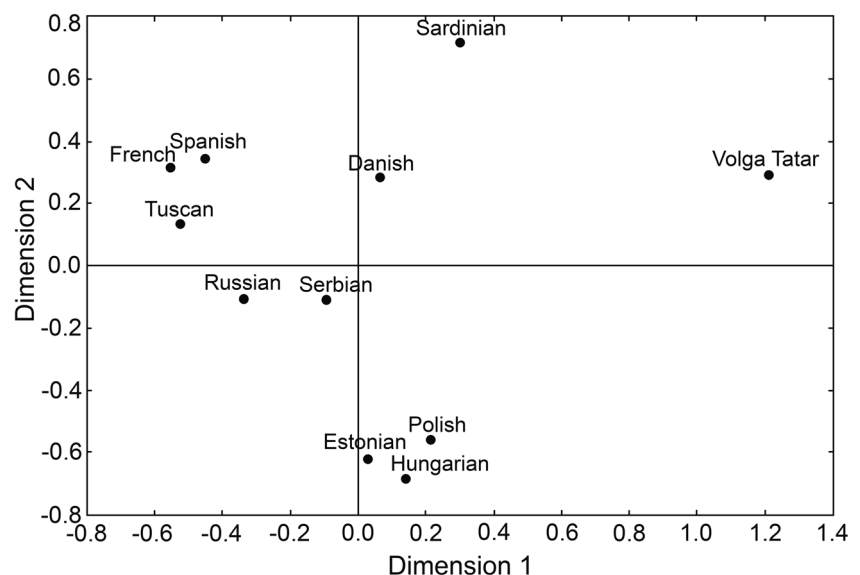
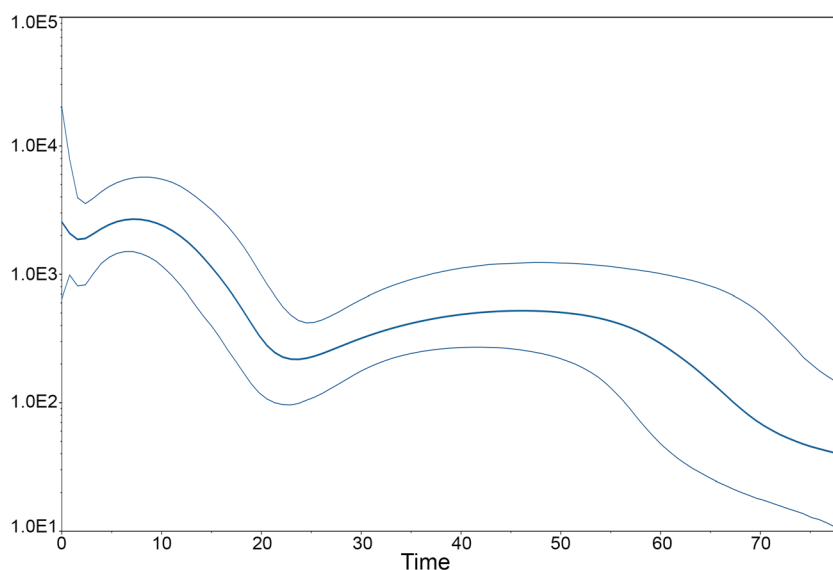


Fig. 2 BSP indicating the median of the hypothetical effective population size through time based on 226 Serbian complete mitogenome sequences. The *x*-axis is the time from the present in units of thousands of years, and the *y*-axis is equal to $Ne\mu$ (the product of the effective population size and mutation rate). The thick solid line represents the median posterior effective population size through time, and the thin lines show the 95% highest posterior density limits



some differences in haplogrouping. In a few cases, mtPhyl haplogroup assignment matched one of the multiple possible haplogroups provided by SAM2 instead of MRCA which represents conservative estimate that should preferably be used in order to avoid bias in forensic casework. On the other hand, we took advantage of the mtDNA data available in the literature [18, 23, 25, 33, 36, 51–55] which is not (yet) included by the PhyloTree, and assigned haplotypes found outside the PhyloTree build 17 using mtPhyl to 10 new subhaplogroups or those previously proposed by us and other research groups.

The number of unique mitogenomes in Serbian population of 85% was lower than that found in Polish and Russian populations [23], and the random match probability was as low as 0.53%. This value, along with that observed in Danes (0.06%) and in Russians (0.27%), was among the lowest values in our study.

Both PHP and LHP, which are much more common in humans than previously thought and have been addressed in several recent forensic studies [16, 63, 64], were observed among Serbian mitogenomes. Contrary to the frequency of control region LHPs observed in Serbians, which was comparable with that reported in other surveys [16, 63], the frequency of PHPs was lower than that found in other human populations [16, 39]. The variation in incidence of PHPs in various human populations, has, however, been observed previously [16, 63]. Moreover, the PHP detection is certainly dependent on the sequencing chemistries and electrophoretic separation systems used by different laboratories [48]. It is also worth mentioning that several studies showed that PHP occurrence is age-related, dependent on the tissue type, and position-specific [64–66]. Given the tissue-specific occurrence of heteroplasmies, it is justified from a forensic perspective to compare the distribution of heteroplasmic sites between

tissues [67] in order to differentiate between exclusion, mixtures, and occurrence of tissue-specific differences within an individual [64]. In addition, while PHPs may increase the strength of the evidence in forensics [68], LHPs are usually disregarded in forensic databasing [48]. Nonetheless, we observed that the inclusion of indel polymorphisms may slightly improve the resolution of Serbian mtDNA haplotypes (87.6% with indels vs. 85.0% without indels).

The level of mtDNA genetic diversity in Serbian population obtained in our study increased in comparison with that observed in a previous survey employing variability of the HVS-I region [18]. Furthermore, somewhat higher values of among population variation were detected in AMOVA based on complete mitogenomes compared with those based on HVS-I/HVS-II analysis (e.g., [7, 69, 70]). In comparison with the previous analyses based on HVS-I variability [18], our investigation at the maximum resolution has not led to the significant increase in F_{ST} values among Serbian and Hungarian population (F_{ST} of 0.00184 vs. 0.00178), while these values almost doubled in cases of the Serbian and Polish (F_{ST} of 0.00418 vs. 0.00279) and Serbian and Russian population (F_{ST} of 0.00357 vs. 0.00185). This may be due to the difference in population sizes used for the analyses, or alternatively, because of increased resolution achieved by the analysis of complete mitogenomes. Higher genetic affinity among Serbians and Russians than among Serbians and Poles observed in our MDS graph is in accordance with previous findings based on HVS-I variability [18].

The number of mean pairwise differences in Serbian population of 27.82 is in the range of that observed in other European populations (this study, [23]). A recent expansion of the Serbian and other studied populations, inferred from the negative and statistically significant Tajima's D values, was not indicated by the mismatch distribution analysis which

revealed bimodal distribution of nucleotide pairwise differences in all analyzed populations. However, bimodal distribution may be obtained in cases of subdivided ancestral populations even in the context of exponential population growth [71]. On the other hand, it may also indicate the presence of two groups of pairwise distributions, with nucleotide differences between recently diverged lineages accounting for the first, smaller peak, and differences between more distantly related haplotypes accounting for the second, larger peak [16].

A recent expansion of the Serbian population was indicated also by the BSP analysis which is very accurate in detecting relatively recent demographic changes [35, 72, 73]. Furthermore, based on this analysis, we were able to correlate time frames of main changes in population sizes over time to relevant prehistoric/historic events. For instance, it is well known that the climate change after the LGM triggered expansions of human populations throughout Eastern and Central Europe from southern refugia [23, 35, 72]. Post-LGM expansions have been demonstrated recently based on the analysis of complete mitogenomes of Russians and Poles [23], and the same trend was observed in our study based on complete mitogenomes of Serbians, with the maximum population size recorded around 7.1 kya, i.e., during the Neolithic agricultural transition [73]. Afterwards, during the Neolithic transition, we found a decline in the population size, which is in accordance with the documented decline of Neolithic cultures throughout the Europe [74, 75]. Different mechanisms have been put forward to explain this Neolithic decline, such as environmental overexploitation (i.e., decrease or disappearance of forests associated with the expansion of steppe environment), and/or a confrontation with foraging Steppe populations, [76, 77], or alternatively, the outbreak of the infectious diseases that spread rapidly due to the increased population density and close contact with domesticated animals. New findings suggest that an outbreak of a plague, *Yersinia pestis*, may account for the collapse of different Neolithic societies throughout the Europe [78]. Further analysis of the Neolithic archaeological remains from the territory of Serbia may shed more light on the mechanisms underlying the observed Neolithic decline.

BSP for Serbian mitogenomic data shows also an onset of a rapid population growth ~ 1.6 kya, i.e., during the Migration period (fourth to ninth century A.D.). During this turbulent period, many different populations, among which were different Germanic and Slavic tribes, went through or settled in the Balkans [79–81]. While the analysis of segments identical by descent revealed that these migrations have had a significant impact on the formation of the contemporary autosomal gene pool of populations that inhabit nowadays Southeast, Central, and East Europe [82], our findings suggest that they could have caused the population growth in the parts of the Balkan Peninsula as well. Furthermore, the importance of the migrations of different Germanic and Slavic tribes for the formation of the contemporary mtDNA gene pool of Serbians has been

suggested previously, based on the presence of subclades shared among Serbian and different populations of a Slavic and/or Germanic origin [18, 33].

In conclusion, we present complete mitogenome data for 226 Serbians of which 170 were sequenced de novo and 56 were taken from our previous work [18, 33]. Since recent studies that employed low-resolution mtDNA data and larger sample of European populations revealed that Serbian population occupies an intermediate position between eastward (Bulgarians and Macedonians) and westward South Slavic populations (Slovenians, Croatians, Bosnians, and Herzegovinians) inhabiting the Balkans [18, 21], we argue that the Serbian population, which is found in the central part of the Balkan Peninsula, may be used as a representative population for South Slavs. Although additional population datasets are needed to represent adequately high genetic diversity in this part of Europe, our data, as the first report on complete mitogenomes in South Slavs, may constitute a reference database of their complete mitogenomes which are of interest not only for forensics but also for studies focusing on evolutionary history of human populations. We also demonstrate that the usage of complete mitogenomes increased the power of discrimination among individuals, which is essential in the forensic casework, and that the enrichment of the complete mtDNA reference database with Serbian mitogenomes contributed to a certain extent towards the improvement of the worldwide mtDNA phylogeny, which is important for the interpretation of the mtDNA casework [32].

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Compliance with ethical standards

This study was approved by the Research Ethics Committees of the Institute of Molecular Genetics and Genetic Engineering, University of Belgrade, and Institute of Biological Problems of the North, Russian Academy of Sciences.

Conflict of interest The authors declare that they have no conflict of interest.

References

1. Butler JM (2015) Advanced topics in forensic DNA typing: interpretation. Elsevier
2. Holland M, Parsons J (1999) Mitochondrial DNA Sequence analysis-validation and use for forensic casework. *Forensic Sci Rev* 11:21–50

3. King TE, Fortes GG, Balaesque P, Thomas MG, Balding D, Delsler PM, Neumann R, Parson W, Knapp M, Walsh S, Tonasso L, Holt J, Kayser M, Appleby J, Forster P, Ekserdjian D, Hofreiter M, Schurer K (2014) Identification of the remains of King Richard III. *Nat Commun* 5:5631
4. Parson W, Bandelt HJ (2007) Extended guidelines for mtDNA typing of population data in forensic science. *Forensic Sci Int Genet* 1:13–19
5. Butler JM, Levin BC (1998) Forensic applications of mitochondrial DNA. *Trends Biotechnol* 16:158–162
6. Carracedo A, Bar W, Lincoln P, Mayr W, Morling N, Olaisen B, Schneider P, Budowle B, Brinkmann B, Gill P, Holland M, Tully G, Wilson M (2000) DNA commission of the international society for forensic genetics: guidelines for mitochondrial DNA typing. *Forensic Sci Int* 110:79–85
7. Brandstatter A, Niederstatter H, Pavlic M, Grubwieser P, Parson W (2007) Generating population data for the EMPOP database—an overview of the mtDNA sequencing and data evaluation processes considering 273 Austrian control region sequences as example. *Forensic Sci Int* 166:164–175
8. Lutz S, Weisser HJ, Heizmann J, Pollak S (1998) Location and frequency of polymorphic positions in the mtDNA control region of individuals from Germany. *Int J Legal Med* 111:67–77
9. Lutz S, Wittig H, Weisser HJ, Heizmann J, Junge A, Dimo-Simonin N, Parson W, Edelmann J, Anslinger K, Jung S, Augustin C (2000) Is it possible to differentiate mtDNA by means of HVIII in samples that cannot be distinguished by sequencing the HVI and HVII regions? *Forensic Sci Int* 113:97–101
10. Bodner M, Iuvaro A, Strobl C, Nagl S, Huber G, Pelotti S, Pettener D, Luiselli D, Parson W (2015) Helena, the hidden beauty: resolving the most common West Eurasian mtDNA control region haplotype by massively parallel sequencing an Italian population sample. *Forensic Sci Int Genet* 15:21–26
11. Coble MD, Just RS, O’Callaghan JE, Letmanyi IH, Peterson CT, Irwin JA, Parsons TJ (2004) Single nucleotide polymorphisms over the entire mtDNA genome that increase the power of forensic testing in Caucasians. *Int J Legal Med* 118:137–146
12. King JL, LaRue BL, Novroski NM, Stoljarova M, Seo SB, Zeng X, Warshauer DH, Davis CP, Parson W, Sajantila A, Budowle B (2014) High-quality and high-throughput massively parallel sequencing of the human mitochondrial genome using the Illumina MiSeq. *Forensic Sci Int Genet* 12:128–135
13. Smith DR (2016) The past, present and future of mitochondrial genomics: have we sequenced enough mtDNAs? *Brief Funct Genomics* 15:47–54
14. Tully G, Bar W, Brinkmann B, Carracedo A, Gill P, Morling N, Parson W, Schneider P (2001) Considerations by the European DNA profiling (EDNAP) group on the working practices, nomenclature and interpretation of mitochondrial DNA profiles. *Forensic Sci Int* 124:83–91
15. Parson W, Dur A (2007) EMPOP—a forensic mtDNA database. *Forensic Sci Int Genet* 1:88–92
16. Just RS, Scheible MK, Fast SA, Sturk-Andreaggi K, Rock AW, Bush JM, Higginbotham JL, Peck MA, Ring JD, Huber GE, Xavier C, Strobl C, Lyons EA, Diegoli TM, Bodner M, Fendt L, Kralj P, Nagl S, Niederwieser D, Zimmermann B, Parson W, Irwin JA (2015) Full mtGenome reference data: development and characterization of 588 forensic-quality haplotypes representing three U.S. populations. *Forensic Sci Int Genet* 14:141–155
17. UN (2015) World population prospects: The 2015 revision, key findings and advance tables. UN Department of Economic and Social Affairs
18. Davidovic S, Malyarchuk B, Aleksic JM, Derenko M, Topalovic V, Litvinov A, Stevanovic M, Kovacevic-Grujicic N (2015) Mitochondrial DNA perspective of Serbian genetic diversity. *Am J Phys Anthropol* 156:449–465
19. Grzybowski T, Malyarchuk BA, Derenko MV, Perkova MA, Bednarek J, Wozniak M (2007) Complex interactions of the Eastern and Western Slavic populations with other European groups as revealed by mitochondrial DNA analysis. *Forensic Sci Int Genet* 1:141–147
20. Juras A, Dabert M, Kushniarevich A, Malmstrom H, Raghavan M, Kosicki JZ, Metspalu E, Willerslev E, Piontek J (2014) Ancient DNA reveals matrilineal continuity in present-day Poland over the last two millennia. *PLoS One* 9:e110839
21. Kushniarevich A, Utevska O, Chuhryaeva M, Agdzhoyan A, Dibirova K, Uktveryte I, Mols M, Mulahasanovic L, Pshenichnov A, Frolova S, Shanko A, Metspalu E, Reidla M, Tambets K, Tamm E, Koshel S, Zaporozhchenko V, Atramentova L, Kucinskas V, Davydenko O, Goncharova O, Evseeva I, Churnosov M, Pocheshchova E, Yunusbayev B, Khusnutdinova E, Marjanovic D, Rudan P, Rootsi S, Yankovsky N, Endicott P, Kassian A, Dybo A, Genographic C, Tyler-Smith C, Balanovska E, Metspalu M, Kivisild T, Villems R, Balanovsky O (2015) Genetic heritage of the Balto-Slavic speaking populations: a synthesis of autosomal, mitochondrial and Y-chromosomal data. *PLoS One* 10:e0135820
22. Malyarchuk B, Grzybowski T, Derenko M, Perkova M, Vanecek T, Lazur J, Gomolcak P, Tsybovsky I (2008) Mitochondrial DNA phylogeny in Eastern and Western Slavs. *Mol Biol Evol* 25:1651–1658
23. Malyarchuk B, Litvinov A, Derenko M, Skonieczna K, Grzybowski T, Grosheva A, Shneider Y, Rychkov S, Zhukova O (2017) Mitogenomic diversity in Russians and Poles. *Forensic Sci Int Genet* 30:51–56
24. Mielnik-Sikorska M, Daca P, Malyarchuk B, Derenko M, Skonieczna K, Perkova M, Dobosz T, Grzybowski T (2013) The history of Slavs inferred from complete mitochondrial genome sequences. *PLoS One* 8:e54360
25. Sarac J, Saric T, Augustin DH, Jeran N, Kovacevic L, Cvjetan S, Lewis AP, Metspalu E, Reidla M, Novokmet N, Vidovic M, Nevajda B, Glasnovic A, Marjanovic D, Missoni S, Villems R, Rudan P (2014) Maternal genetic heritage of Southeastern Europe reveals a new Croatian isolate and a novel, local sub-branching in the X2 haplogroup. *Ann Hum Genet* 78:178–194
26. Rootsi S, Magri C, Kivisild T, Benuzzi G, Help H, Bermisheva M, Kutuev I, Barac L, Peric M, Balanovsky O, Pshenichnov A, Dion D, Grobei M, Zhivotovsky LA, Battaglia V, Achilli A, Al-Zahery N, Parik J, King R, Cinnioglu C, Khusnutdinova E, Rudan P, Balanovska E, Scheffrahn W, Simonescu M, Brehm A, Goncalves R, Rosa A, Moisan JP, Chaventre A, Ferak V, Furedi S, Oefner PJ, Shen P, Beckman L, Mikerezi I, Terzić R, Primorac D, Cambon-Thomsen A, Krumina A, Torroni A, Underhill PA, Santachiara-Benerecetti AS, Villems R, Semino O (2004) Phylogeography of Y-chromosome haplogroup I reveals distinct domains of prehistoric gene flow in Europe. *Am J Hum Genet* 75:128–137
27. Semino O, Passarino G, Oefner PJ, Lin AA, Arbuzova S, Beckman LE, De Benedictis G, Francalacci P, Kouvatsi A, Limborska S, Marcikiae M, Mika A, Mika B, Primorac D, Santachiara-Benerecetti AS, Cavalli-Sforza LL, Underhill PA (2000) The genetic legacy of Paleolithic Homo sapiens sapiens in extant Europeans: a Y chromosome perspective. *Science* 290:1155–1159
28. Malyarchuk BA, Grzybowski T, Derenko MV, Czamy J, Drobnic K, Miscicka-Sliwka D (2003) Mitochondrial DNA variability in Bosnians and Slovenians. *Ann Hum Genet* 67:412–425
29. Karachanak S, Carossa V, Nesheva D, Olivieri A, Pala M, Hooshiar Kashani B, Grugni V, Battaglia V, Achilli A, Yordanov Y, Galabov AS, Semino O, Toncheva D, Torroni A (2012) Bulgarians vs the other European populations: a mitochondrial DNA perspective. *Int J Legal Med* 126:497–503
30. Zimmermann B, Brandstatter A, Duftner N, Niederwieser D, Spiroski M, Arsov T, Parson W (2007) Mitochondrial DNA control

- region population data from Macedonia. *Forensic Sci Int Genet* 1: e4–e9
31. Zupanic Pajnic I, Balazic J, Komel R (2004) Sequence polymorphism of the mitochondrial DNA control region in the Slovenian population. *Int J Legal Med* 118:1–4
 32. Heinz T, Pala M, Gomez-Carballa A, Richards MB, Salas A (2017) Updating the African human mitochondrial DNA tree: relevance to forensic and population genetics. *Forensic Sci Int Genet* 27:156–159
 33. Davidovic S, Malyarchuk B, Aleksic J, Derenko M, Topalovic V, Litvinov A, Skonieczna K, Rogalla U, Grzybowski T, Stevanovic M, Kovacevic-Grujicic N (2017) Mitochondrial super-haplogroup U diversity in Serbians. *Ann Hum Biol* 44:408–418
 34. Li ST, Besenbacher S, Li YR, Kristiansen K, Grarup N, Albrechtsen A, Sparso T, Korneliusen T, Hansen T, Wang J, Nielsen R, Pedersen O, Bolund L, Schierup MH (2014) Variation and association to diabetes in 2000 full mtDNA sequences mined from an exome study in a Danish population. *Eur J Hum Genet* 22: 1040–1045
 35. Lippold S, Xu H, Ko A, Li M, Renaud G, Butthof A, Schroder R, Stoneking M (2014) Human paternal and maternal demographic histories: insights from high-resolution Y chromosome and mtDNA sequences. *Investig Genet* 5:13
 36. Malyarchuk B, Derenko M, Denisova G, Litvinov A, Rogalla U, Skonieczna K, Grzybowski T, Pentelenyi K, Guba Z, Zeke T, Molnar MJ (2018) Whole mitochondrial genome diversity in two Hungarian populations. *Mol Gen Genomics* 293:1255–1263
 37. Gomez-Carballa A, Pardo-Seco J, Amigo J, Martinon-Torres F, Salas A (2015) Mitogenomes from The 1000 Genome Project reveal new Near Eastern features in present-day Tuscans. *PLoS One* 10:e0119242
 38. Stoljarova M, King JL, Takahashi M, Aaspollu A, Budowle B (2015) Whole mitochondrial genome genetic diversity in an Estonian population sample. *Int J Legal Med* 130:67–71
 39. Ramos A, Santos C, Mateiu L, Gonzalez Mdel M, Alvarez L, Azevedo L, Amorim A, Aluja MP (2013) Frequency and pattern of heteroplasmy in the complete human mitochondrial genome. *PLoS One* 8:e74636
 40. Fraumene C, Belle EMS, Castri L, Sanna S, Mancosu G, Cosso M, Marras F, Barbuji G, Pirastu M, Angius A (2006) High resolution analysis and phylogenetic network construction using complete mtDNA sequences in Sardinian genetic isolates. *Mol Biol Evol* 23:2101–2111
 41. Malyarchuk B, Derenko M, Denisova G, Kravtsova O (2010) Mitogenomic diversity in Tatars from the Volga-Ural region of Russia. *Mol Biol Evol* 27:2220–2226
 42. Quinque D, Kittler R, Kayser M, Stoneking M, Nasidze I (2006) Evaluation of saliva as a source of human DNA for population and association studies. *Anal Biochem* 353:272–277
 43. Torroni A, Rengo C, Guida V, Cruciani F, Sellitto D, Coppa A, Calderon FL, Simionati B, Valle G, Richards M, Macaulay V, Scozzari R (2001) Do the four clades of the mtDNA haplogroup L2 evolve at different rates? *Am J Hum Genet* 69:1348–1356
 44. Fendt L, Zimmermann B, Daniaux M, Parson W (2009) Sequencing strategy for the whole mitochondrial genome resulting in high quality sequences. *BMC Genomics* 10:139
 45. Andrews RM, Kubacka I, Chinnery PF, Lightowlers RN, Turnbull DM, Howell N (1999) Reanalysis and revision of the Cambridge reference sequence for human mitochondrial DNA. *Nat Genet* 23: 147
 46. Fregel R, Delgado S (2011) HaploSearch: a tool for haplotype-sequence two-way transformation. *Mitochondrion* 11:366–367
 47. Bandelt HJ, Parson W (2008) Consistent treatment of length variants in the human mtDNA control region: a reappraisal. *Int J Legal Med* 122:11–21
 48. Parson W, Gusmao L, Hares DR, Irwin JA, Mayr WR, Morling N, Pokorak E, Prinz M, Salas A, Schneider PM, Parsons TJ (2014) DNA Commission of the International Society for Forensic Genetics: revised and extended guidelines for mitochondrial DNA typing. *Forensic Sci Int Genet* 13:134–142
 49. Huber N, Parson W, Dur A (2018) Next generation database search algorithm for forensic mitogenome analyses. *Forensic Sci Int Genet* 37:204–214
 50. van Oven M (2015) PhyloTree Build 17: growing the human mitochondrial DNA tree. *Forensic Sci Int Genet: Genet Suppl Ser* 5: e392–e394
 51. Costa MD, Pereira JB, Pala M, Fernandes V, Olivieri A, Achilli A, Perego UA, Rychkov S, Naumova O, Hatina J, Woodward SR, Eng KK, Macaulay V, Carr M, Soares P, Pereira L, Richards MB (2013) A substantial prehistoric European ancestry amongst Ashkenazi maternal lineages. *Nat Commun* 4:2543
 52. Derenko M, Malyarchuk B, Denisova G, Perkova M, Litvinov A, Grzybowski T, Dambueva I, Skonieczna K, Rogalla U, Tsybovsky I, Zakharov I (2014) Western Eurasian ancestry in modern Siberians based on mitogenomic data. *BMC Evol Biol* 14:217
 53. Hernandez CL, Soares P, Dugoujon JM, Novelletto A, Rodriguez JN, Rito T, Oliveira M, Melhaoui M, Baali A, Pereira L, Calderon R (2015) Early holocene and historic mtDNA African signatures in the Iberian Peninsula: the Andalusian Region as a paradigm. *PLoS One* 10:e0139784
 54. Kovacevic L, Tambets K, Ilumae AM, Kushniarevich A, Yunusbayev B, Solnik A, Bego T, Primorac D, Skaro V, Leskovic A, Jakovski Z, Drobnic K, Tolk HV, Kovacevic S, Rudan P, Metspalu E, Marjanovic D (2014) Standing at the gateway to Europe—the genetic structure of Western Balkan populations based on autosomal and haploid markers. *PLoS One* 9:e105090
 55. Sahakyan H, Hooshiar Kashani B, Tamang R, Kushniarevich A, Francis A, Costa MD, Pathak AK, Khachtryan Z, Sharma I, van Oven M, Parik J, Hovhannisyan H, Metspalu E, Pennarun E, Karmin M, Tamm E, Tambets K, Bahmanimehr A, Reisberg T, Reidla M, Achilli A, Olivieri A, Gandini F, Perego UA, Al-Zahery N, Houshmand M, Sanati MH, Soares P, Rai E, Sarac J, Saric T, Sharma V, Pereira L, Fernandes V, Cerny V, Farjadian S, Singh DP, Azakli H, Ustek D, Ekomasova Trofimova N, Kutuev I, Litvinov S, Bermisheva M, Khusnutdinova EK, Rai N, Singh M, Singh VK, Reddy AG, Tolk HV, Cvjetan S, Lauc LB, Rudan P, Michalodimitrakis EN, Anagnou NP, Pappa KI, Golubenkov MV, Orekhov V, Borinskaya SA, Kaldma K, Schauer MA, Simionescu M, Gusar V, Grechanina E, Govindaraj P, Voevoda M, Damba L, Sharma S, Singh L, Semino O, Behar DM, Yepiskoposyan L, Richards MB, Metspalu M, Kivisild T, Thangaraj K, Endicott P, Chaubey G, Torroni A, VILLEMS R (2017) Origin and spread of human mitochondrial DNA haplogroup U7. *Sci Rep* 7:46044
 56. Excoffier L, Lischer HE (2010) Arlequin suite ver 3.5: a new series of programs to perform population genetics analyses under Linux and Windows. *Mol Ecol Resour* 10:564–567
 57. Stoneking M, Hedgecock D, Higuchi RG, Vigilant L, Erlich HA (1991) Population variation of human mtDNA control region sequences detected by enzymatic amplification and sequence-specific oligonucleotide probes. *Am J Hum Genet* 48:370–382
 58. Librado P, Rozas J (2009) DnaSP v5: a software for comprehensive analysis of DNA polymorphism data. *Bioinformatics* 25:1451–1452
 59. Suchard MA, Lemey P, Baele G, Ayres DL, Drummond AJ, Rambaut A (2018) Bayesian phylogenetic and phylodynamic data integration using BEAST 1.10. *Virus Evolution* 4:vey016
 60. Tamura K, Stecher G, Peterson D, Filipinski A, Kumar S (2013) MEGA6: molecular evolutionary genetics analysis version 6.0. *Mol Biol Evol* 30:2725–2729
 61. Soares P, Ermini L, Thomson N, Mormina M, Rito T, Rohlf A, Salas A, Oppenheimer S, Macaulay V, Richards MB (2009) Correcting

- for purifying selection: an improved human mitochondrial molecular clock. *Am J Hum Genet* 84:740–759
62. Rambaut A, Drummond AJ, Xie D, Baele G, Suchard MA (2018) Posterior summarization in Bayesian phylogenetics using Tracer 1.7. *Syst Biol* 67:901–904
 63. Irwin JA, Saunier JL, Niederstatter H, Strouss KM, Sturk KA, Diegoli TM, Brandstatter A, Parson W, Parsons TJ (2009) Investigation of heteroplasmy in the human mitochondrial DNA control region: a synthesis of observations from more than 5000 global population samples. *J Mol Evol* 68:516–527
 64. Naue J, Horer S, Sanger T, Strobl C, Hatzler-Grubwieser P, Parson W, Lutz-Bonengel S (2015) Evidence for frequent and tissue-specific sequence heteroplasmy in human mitochondrial DNA. *Mitochondrion* 20:82–94
 65. Li M, Schroder R, Ni S, Madea B, Stoneking M (2015) Extensive tissue-related and allele-related mtDNA heteroplasmy suggests positive selection for somatic mutations. *Proc Natl Acad Sci U S A* 112:2491–2496
 66. Samuels DC, Li C, Li B, Song Z, Torstenson E, Boyd Clay H, Rokas A, Thornton-Wells TA, Moore JH, Hughes TM, Hoffman RD, Haines JL, Murdock DG, Mortlock DP, Williams SM (2013) Recurrent tissue-specific mtDNA mutations are common in humans. *PLoS Genet* 9:e1003929
 67. Salas A, Lareu MV, Carracedo A (2001) Heteroplasmy in mtDNA and the weight of evidence in forensic mtDNA analysis: a case report. *Int J Legal Med* 114:186–190
 68. Ivanov PL, Wadhams MJ, Roby RK, Holland MM, Weedn VW, Parsons TJ (1996) Mitochondrial DNA sequence heteroplasmy in the Grand Duke of Russia Georgij Romanov establishes the authenticity of the remains of Tsar Nicholas II. *Nat Genet* 12:417–420
 69. Garcia O, Fregel R, Larruga JM, Alvarez V, Yurrebaso I, Cabrera VM, Gonzalez AM (2011) Using mitochondrial DNA to test the hypothesis of a European post-glacial human recolonization from the Franco-Cantabrian refuge. *Heredity* 106:37–45
 70. Prieto L, Zimmermann B, Goios A, Rodriguez-Monge A, Paneto GG, Alves C, Alonso A, Fridman C, Cardoso S, Lima G, Anjos MJ, Whittle MR, Montesino M, Cicarelli RMB, Rocha AM, Albarran C, de Pancorbo MM, Pinheiro MF, Carvalho M, Sumita DR, Parson W (2011) The GHEP-EMPOP collaboration on mtDNA population data—a new resource for forensic casework. *Forensic Sci Int Genet* 5:146–151
 71. Marjoram P, Donnelly P (1994) Pairwise comparisons of mitochondrial DNA sequences in subdivided populations and implications for early human evolution. *Genetics* 136:673–683
 72. Zheng HX, Yan S, Qin ZD, Jin L (2012) MtDNA analysis of global populations support that major population expansions began before Neolithic Time. *Sci Rep* 2:745
 73. Gignoux CR, Henn BM, Mountain JL (2011) Rapid, global demographic expansions after the origins of agriculture. *Proc Natl Acad Sci USA* 108:6044–6049
 74. Hinz M, Feesser I, Sjogren KG, Muller J (2012) Demography and the intensity of cultural activities: an evaluation of Funnel Beaker Societies (4200–2800 cal BC). *J Archaeol Sci* 39:3331–3340
 75. Shennan S, Downey SS, Timpson A, Edinborough K, Colledge S, Kerig T, Manning K, Thomas MG (2013) Regional population collapse followed initial agriculture booms in mid-Holocene Europe. *Nat Commun* 4:2486
 76. Anthony D (2007) *The Horse, the wheel, and language: how Bronze-Age riders from the Eurasian steppes shaped the modern world*. Princeton University Press, Princeton
 77. Kirleis W, Dreibrodt S (2016) *The Natural Background: Forest, Forest Steppe or Steppe Environment*. In: Muller J, Rassmann K, Videiko M (eds) *Trypillia mega-sites and European prehistory 4100–3400 BCE*. Routledge Taylor and Francis, New York, pp 171–180
 78. Rascovan N, Sjogren KG, Kristiansen K, Nielsen R, Willerslev E, Desnues C, Rasmussen S (2019) Emergence and spread of basal lineages of *Yersinia pestis* during the Neolithic decline. *Cell* 176:295–305
 79. Ostrogorski G (1998) *Istorija Vizantije*. Narodna Knjiga Alfa, Belgrade
 80. Wilkes J (1996) *The Illyrians*. Wiley-Blackwell, Oxford
 81. Živković T (2002) *South Slavs under the Byzantine rule (600–1025)*. Institute of History, Belgrade
 82. Ralph P, Coop G (2013) The geography of recent genetic ancestry across Europe. *PLoS Biol* 11:e1001555

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