

Spatial link between nucleoli and expression of the *Zac1* gene

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Abstract Eukaryotic genomes are highly organized within the cell nucleus. Genome organization not only implies the preferential positioning of genetic elements in the interphase nucleus but also the topographic distribution of biological processes. We have investigated the relationship between spatial organization and genome function in single cells. *Myc*, *c-Met*, *Igf2r*, *Asb4*, and *Zac1* genes have the same radial distribution, but they are not positioned in close proximity with respect to each other. Three-dimensional mapping of their transcription sites uncovered a gene-specific pattern of relative positioning with respect to the nucleolus. We found that the *Zac1* gene transcription preferentially occurs juxtaposed to the nucleolus, and that its mRNA accumulates at this site of transcription. Nucleoli

isolation followed by qRT-PCR provided evidence for a physical interaction between *Zac1* mRNA and the nucleolus. Actinomycin-D treatment induced disassembly of the nucleolus, loss of the RNA-FISH signal, and dramatic increase of the ZAC protein level. However, inhibition of RNA polymerase II had no effect over the *Zac1* FISH signal and the protein expression. Induction of cell cycle arrest, which involves participation of the ZAC protein, provoked mRNA release from its retention site and protein synthesis. Our data demonstrate that *Zac1* mRNA preferentially accumulates in close proximity to nucleoli within the cell nucleus. In addition, our results suggest a functional link between such spatial distribution and protein expression.

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Background

Eukaryotic genomes are highly organized within the cell nucleus. Chromosomes exhibit patterns of spatial distribution determined by their size or gene density (Misteli 2007; Parada et al. 2004b). In mouse, the patterns of radial and relative chromosome positioning are tissue type specific indicating that genome reorganization occurs during cell differentiation (Parada et al. 2004a). This conclusion is further supported by the observations that the stem cell-specific genes *Nanog* and *Oct4* acquire differential positioning as their expression levels change during differentiation of human embryonic stem cells (Wiblin et al. 2005). Cellular organization of the genome not only implies non-random distribution of genes and chromosomes in the nuclear space but also higher order of chromatin organization and spatial organization within the interphase nucleus of biological processes, including DNA replication, repair, and transcription (Misteli 2007). Gene transcription

by RNA polymerase II involves many molecular players and occurs throughout the whole nucleus; however, the process occurs highly organized in “factories” that contain more than one polymerase molecule (Cook 1999). Furthermore, transcription of genes that are functionally related yet distant from one another at the genomic level can occur by sharing RNA polymerase II transcription sites, providing support for the notion of spatial organization of biological processes (Dostie et al. 2007; Osborne et al. 2007).

The nucleolus is a clear example of the nuclear functional organization paradigm. In this nuclear domain, ribosomal RNA genes are transcribed by RNA polymerase I in multiple and large transcription centers (Dundr et al. 2000). rRNA genes (18S, 5.8S, and 28S) are arranged in clusters of repeated units associated with nucleolar organizing regions (NORs). In humans, rRNA genes map to the short arm of chromosomes 13, 14, 15, 21, and 22. However, in mouse, the chromosomal localization of NORs varies by strain, for example, NORs map to chromosomes 12, 15, 18, and 19 in C57BL/6J mice and to chromosomes 12, 15, and 16 in the BALB/cJ mice (Henderson et al. 1974; Romanova et al. 2006). Taking into consideration the proposed models for the cellular organization of genome function (Misteli 2007), an obvious prediction would be that chromosomes carrying NORs have to be spatially arranged in the cell nucleus. In support of this model, mouse chromosomes 12 and 15, which contain NORs, are positioned in close proximity to each other as well as to chromosome 14, forming a cluster (Parada et al. 2002). Furthermore, the tendency of NOR-bearing chromosomes to associate with nucleoli correlates with the number of transcriptionally competent NORs in the chromosome (Kalmarova et al. 2007).

Spatial genome organization has been usually assessed by determining the preferential position of chromosomes/genes relative to the nuclear center (radial distribution) or with respect to other genetic elements (relative positioning). In addition, evidence has emerged pointing towards the importance of the third dimension on gene expression (Dekker 2008). Transient gene repositioning in the interphase nucleus has been postulated to optimize coordinated expression of similarly regulated genes (Kumaran et al. 2008). Here, we test the link between proximal positioning with respect to the nucleolus and function of murine *Asb4*, *c-Met*, *Zac1*, *Myc*, and *Igf2r*. These genes were selected due to their similar radial distribution in the cell nucleus. By simultaneously mapping the transcription sites of these genes with respect to the nucleolus and measuring the expression level of this series of loci, we uncovered a gene-specific pattern of relative positioning. Furthermore, our results support the notion that this specialized nuclear compartment may play a role in ZAC protein expression by interacting with the *Zac1* mRNA.

Materials and methods

Cell culture and drug treatments

The mouse embryonic fibroblast (MEF) cell line G7 was cultured in DMEM/F-12 media containing 10% FBS, 1% penicillin/streptomycin, and 1% glutamine (Gibco) under 5% CO₂ atmosphere at 37°C. When appropriate, 18–24 h after cell plating, the old medium was replaced for fresh medium containing 0.5 µg/ml of Actinomycin D (Sigma-Aldrich) for 1 h or 50 µg/ml α-amanitin (Sigma-Aldrich) for 6 h, for selective RNA Pol I or II inhibition, respectively (Lorenzi et al. 2000). Control experiments were done by adding equal amounts of water/DMSO. For induction of double-strand breaks, Etoposide (Sigma) was added at a final concentration of 5 µM during 1 h.

RNA and DNA FISH

Cells were plated on Petri dishes containing glass coverslips 18–24 h prior to the procedure. The cells were rinsed briefly in PBS and in PBS containing paraformaldehyde (2%) and Triton X-100 (0.3%). The cells were then fixed in paraformaldehyde 2% for 10 min, permeabilized with Triton X-100 (0.5%) in PBS for 10 min, rinsed twice with PBS, and stabilized in 2× SSC for 10 min. Immediately after this treatment, coverslips were placed over the probe mixture, sealed with rubber cement, and incubated in a humidified chamber at 37°C overnight. The probes were generated by nick translation of plasmid or BAC clones. In addition, RNA FISH was performed with directly labeled PCR fragments specific to coding exons VIII, IX, and X and intron X–XI of the *Zac1* gene, or to the coding region and a fragment of the 3' UTR of the *β3b5* gene. Apart from the specific DNA (0.1–0.2 µg), the hybridization mixture contained salmon sperm DNA (1.5 µg/µl) and yeast tRNA (1 µg/µl) dissolved in 2× SSC, 50% formamide, and 10% dextran sulfate.

For chromosome painting and locus-specific DNA FISH, the yeast tRNA was substituted by cot1DNA (0.4 µg/µl) in the probe preparation and the target DNA was denatured by heating at 73°C in 70% formamide for 5 min. Post-hybridization washes included 15 min in 2× SSC and 10 min in 1× SSC. The detection was performed according to standard procedures with FITC conjugated avidin (Pierce) or anti-digoxigenin antibodies (Roche) (Parada et al. 2004a). Spectral karyotyping was performed according to the protocol provided by the probe manufacturer (Applied Spectral Imaging Ltd). Nucleolus immunostaining was done during the FISH detection step with a mouse monoclonal antibody against the C-terminus of nucleophosmin (NPM) (B23) and using goat anti mouse IgG conjugated with Texas red or FITC (Southern Biotech).

Biotin- or digoxigenin-labeled probes were generated by nick translation using the following clones: A95504 and RP23-157D7 (*Zac1*), RP24-465F5 (*Myc*), RP23-208N9 (*c-Met*), RP23-81B3 (*Igf2R*), and RP24-169I21 (*Asb4*). For *Zac1* and *Sf3b5* specific probes, we performed PCR on G7 genomic DNA using a mix of dNTP and Biotin-16-dUTP or Digoxigenin-12-dUTP (Roche) and primers designed with Primer3 software (<http://primer3.sourceforge.net/>) for different regions of *Zac1* and *Sf3b5* sequences (see primer sequences in Table 1 of Supplementary File 1). Whole chromosome painting probes were prepared by DOP-PCR from flow sorted mouse chromosomes (Parada et al. 2004a). For detecting NORs, we used a mix of pA and pB rDNA probes, prepared from pA and pB plasmid constructs (Erickson et al. 1981), kindly donated by Miroslav Dundr (Rosalind Franklin University of Medicine and Science, North Chicago, USA).

Imaging and image analysis

Stacks of images scanning the whole nucleus were acquired with an axial separation of 300 nm using a Leica DM IRE2 laser-scanning microscope (Leica Microsystems Heidelberg GmbH). To determine the 3D radial position of any given signal, the shortest distance from the center of the nucleus to the periphery that included the center of the fluorescent signal was directly measured on the z-stack using Volocity.4.3[®] software (Improvision, Image, Processing and Vision Company Limited), using the same threshold value for object segmentation in all images. The absolute distances from the center of the nucleus to the transcription sites were normalized as a fraction of the nuclear radius to account for natural variations in nuclear size, which may influence positioning. Cell nuclei were subdivided in five concentric shells each corresponding to 20% of the nucleus radius and the radial positioning data was binned into these five sub-domains. For quantitative measurements, more than 100 nuclei from multiple experiments were analyzed.

To study relative positioning respect to the nucleolus, we determined the frequency at which the nucleolus and the transcription site fluorescent signals were overlapping in at least two focal planes of the 3D z-stack images using the Leica Software. Since we observed that there were many cells with signals very close, but not juxtaposed, we defined an alternative criterion. The absolute spatial separations of the transcription sites were directly measured from center of the transcription sites to the nearest voxel of the nucleolus on the 3D image, and signals were classified as “close” if the nucleolus–transcript maximum separation was less than or equal to the diameter of the RNA-FISH signal. However, to better characterize trends in relative positioning, we used the first and more stringent criterion for statistical analysis.

Quantitative RT-PCR

RNA was isolated from the same cell plate used for FISH analysis using the RNAeasy Mini kit (Qiagen) and treated with RNAase-free DNAase set (Qiagen). The cDNA was synthesized from 1 µg of RNA using the Reverse Transcriptase kit (Promega) following the manufacturer’s recommendations.

qRT-PCR was performed using the SYBR Green PCR Master Mix (ABgene) in an iCycler thermocycler (Bio-Rad Laboratories Inc.) following the instructions of the manufacturer. Primers for *Zac1*, *Sf3b5*, rRNA, β -Actin, and *Gapdh* were designed with Primer 3 Software (primer sequences and annealing temperatures are shown in Table 1 of Supplementary File 1). The efficiency of each set of primers was determined by serial dilution of the template cDNA. The theoretical value obtained with the iCycler software was $100 \pm 2\%$ (Efficiency(%) = $100 \times [10^{(-1/\text{slope})} - 1]$; Bio – Rad Laboratories Inc.) Gene expression levels were estimated as the differences in the threshold cycle values (C_T) of treated and untreated cells.

Nucleoli from MEF cells were isolated according to the protocols publicly available at <http://www.lamondlab.com/f7nucleolarprotocol.htm>. The nucleolar preparations were screened by phase contrast optical microscopy. The total RNA was extracted and 1 µg of this RNA was used to synthesize the cDNA. Thereafter, the presence of *Zac1* transcripts in the nucleolar preparations was assessed by qRT-PCR with a set of primers for exon VIII of the *Zac1* gene (Table 1 of Supplementary File 1). As positive and negative controls, we used primers for pre-rRNA and β -actin, respectively.

Western blot analysis

Proteins were extracted from the same cell plate used for FISH analysis with cell lysis buffer (Cell Signaling Technology) and quantified with the BCA protein assay kit (Pierce). Aliquots of 30 µg of the total protein lysate were separated on 10% SDS-PAGE gels, transferred to PVDF membranes, and probed with primary antibodies against ZAC (M-300) (Santa Cruz Biotechnology Inc). β -Actin C-11 antibody (Santa Cruz Biotechnology) was used as a loading control. Detection was performed with a bovine anti-goat or anti-rabbit secondary antibody conjugated to horseradish peroxidase (Santa Cruz Biotechnology).

Statistics

The Pearson's chi-squared test and the *T* test were performed using R software <http://www.r-project.org/>.

Results

Spatial position of gene transcription

To study the relationship between spatial distribution and genome function, we determined the 3D distribution of transcription sites of *Asb4*, *c-Met*, *Zac1*, *Myc*, and *Igf2r* mapping to chromosomes 6A1, 6A2, 10A1, 15D3, and 17A2, respectively. The position of each gene was assayed independently and also combined in triplets in a cytogenetically normal MEF cell line by RNA FISH using probes generated with bacterial artificial chromosomes (BAC) clones. This type of material was deliberately chosen to obtain robust RNA signals as a first approach, although has the caveat that the fluorescent signals might include other transcripts in addition to those of interest. Transcripts for all clones were easily detected in roughly 90% of the cells, but with clear differences in their expression pattern. While single signals for *Asb4* and *Igf2R* were detected in the vast majority of cells, *Zac1*, *Myc*, and *c-Met* expression was mostly biallelic (Fig. 1a–c). *Asb4* and *Igf2R* are imprinted genes; therefore, monoallelic expression is in agreement with their form of regulation in a parent-of-origin manner. *Zac1* is also an imprinted gene and methyl-specific PCR (MSP) assay showed that both methylated and unmethylated CpGs are present in its promoter region (Supplementary File 1, Supplementary Fig. 1a,b), suggesting that *Zac1* expression should be monoallelic in this cell type. However, we found two signals per nucleus in the vast majority of cells. Therefore, we favor the idea that the biallelic expression of *Zac1* must be due to the presence of a second promoter in these cells, as has been demonstrated for human *ZAC* (Piras et al. 2000; Valleley et al. 2007).

To assess the spatial position of gene transcription, we first determined the 3D radial positions of all genes by direct measurement of the distances from the signals to the center of the nucleus and binning the data into five concentric shells, each corresponding to 20% of the nucleus radius. Primary transcripts corresponding to these genomic regions exhibited a homogeneous distribution localizing preferentially to an intermediate zone between the periphery and the nuclear center. The average radial position was between 60% and 65% of the relative radius (Fig. 1d). Despite their similar radial distribution, we did not find a clear pattern of proximal positioning among these genes, except for *Asb4* and *Met* which map to the same genomic region and are located close to each other in the interphase nucleus (data not shown).

Positioning of gene loci relative to the nucleolus

The nucleolus is not only involved in ribosome biogenesis, but is also a key player in cell cycle progression and

proliferation (Pederson 1998). To study the role of the nucleolus in the spatial organization of non-ribosomal gene expression, we performed RNA FISH combined with immunofluorescence for NPM, a nucleolus specific marker protein involved in ribosome assembly and transport. We first determined the 3D physical distance between each gene and the nearest nucleolus. The positions of the transcription sites with respect to the nucleolus were different for most of the genes, with variations from 0 to 3 μm away from the nucleolus. To quantitate these observations, we determined the frequency with which the fluorescent signals from the nucleolus and the transcription site were juxtaposed in at least two focal planes of the 3D z-stack. While only 25–28% of the *Asb4* and *Met* transcriptionally active genes were juxtaposed to the nucleolus, *Zac1* gene transcription occurred twice as frequently in juxtaposition to nucleoli. Interestingly, only 46% of the fluorescent signals from the *Myc* gene were found in close proximity to the nucleolus, despite the fact that it maps to chromosome 15 (NOR positive), and is therefore expected to localize close to nucleoli at the highest frequency. This frequency is much lower than the 57% at which the transcription of the *Zac1* gene occurs juxtaposed to the nucleolus (Fig. 1e). Since we found many cells with signals very close to the nucleolus, but not juxtaposed, we also quantitated our observations using an alternative criterion based on distance measurements. Signals were considered “close” when the distance between them and the nucleolus was equal to or less than the diameter of the signal. Based on this criterion, we found that the frequency of signals close to the nucleolar compartment increased to 70% for *Zac1* and 60% for *c-Myc*, suggesting that the nucleolus plays a role in *Zac1* gene spatial/functional organization. Additional support for this idea comes from the observation that more than 35% of the cells with biallelic expression of *Zac1* had both signals juxtaposed to the nucleolus. Furthermore, by comparing radial spatial positioning with relative positioning with respect to the nucleolus, we found that despite their similar radial distribution, proximal positioning with respect to the nucleolus is distinct among these genes (Fig. 1e).

Chromosome territories positioning close to nucleoli

Chromosomes are not randomly distributed in the cell nucleus and the position of many genetic elements is related to their function. In humans, rRNA genes are found on all acrocentric chromosomes and this accounts for the preferential positioning of these chromosomes close to the nucleolus (Kalmarova et al. 2007). Having determined that transcription of the *Zac1* gene occurs in close proximity to the nucleolus, we next sought to determine the relative position of this gene and chromosome 10, where *Zac1*

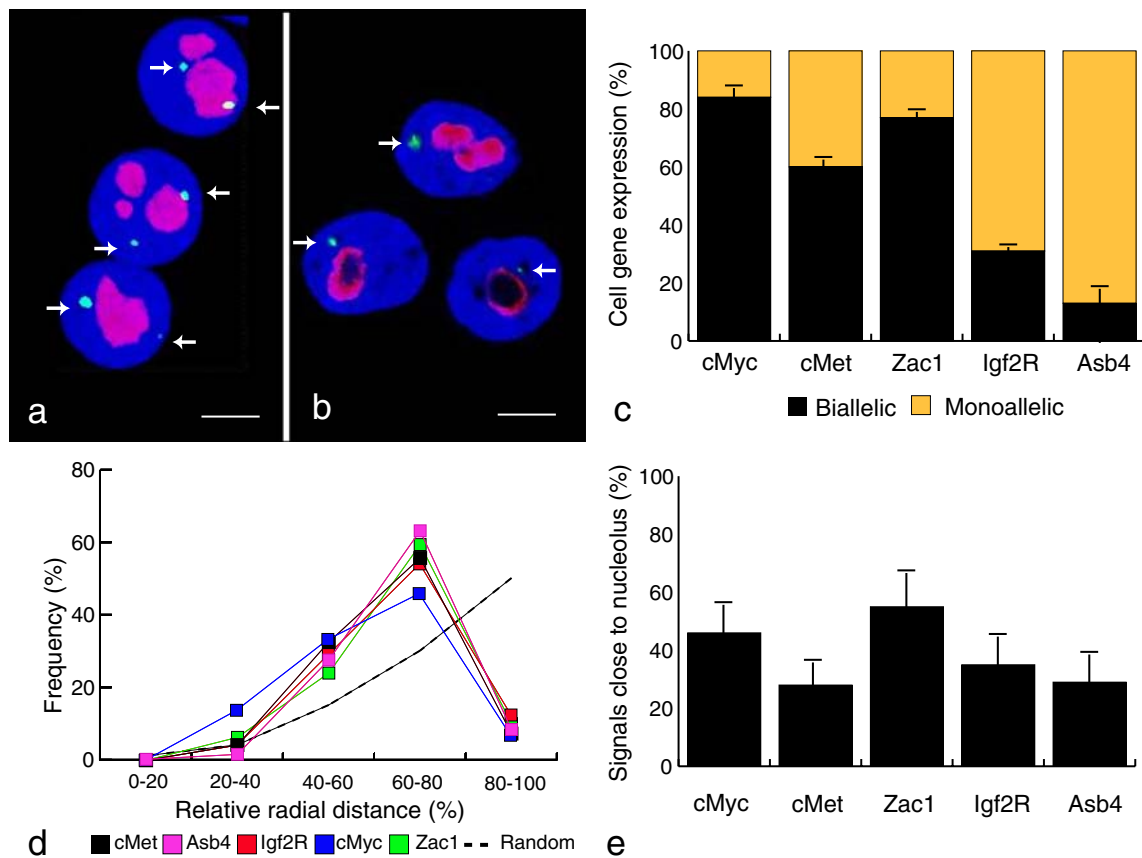


Fig. 1 Nuclear positioning of gene expression. *Zac1* and *Asb4* genes transcription sites detected by RNA FISH with a BAC clone (green, arrows) and nucleolus immunostaining with NPM (red). Counterstaining with DAPI (blue), scale bar=4 μ m. Projected stacks of nuclei are shown (a, b). Gene expression patterns of monoallelic *Asb4*, *Igf2R*, and biallelic *Myc*, *c-Met*, and *Zac1* in MEF cells (c).

The transcription sites have similar radial distribution in the interphase nucleus. 3D actual measurements were normalized as a fraction of nuclear radius (d). Relative position of the genes with respect to the nucleolus. *Zac1* gene transcription occurs frequently juxtaposed to the nucleolus (e). Error bars represent SD from three independent experiments

maps, with respect to the nucleolus. Three-dimensional dual color DNA FISH with probes for chromosome 10 and the *Zac1* gene, combined with immunofluorescence for NPM, revealed that 59% of chromosome 10 and their *Zac1* allele signals were juxtaposed to the nucleolus ($N=190$) (Fig. 2a). Since the chromosomal distribution of ribosomal RNA coding genes varies among different mouse strains, we asked whether chromosome 10 harbor ribosomal RNA coding genes whose function may be responsible for this location. To this end, we performed DNA FISH with specific probes addressed to rDNA encoding the three major classes of rRNA (5.8S, 18S, and 28S) followed by SKY on methanol/acetic acid fixed metaphases. In addition, we performed dual DNA-FISH analysis with rDNA probes combined with whole painting probes for selected chromosomes to confirm the SKY results. Interestingly, we did not find NORs mapping to chromosome 10, but only to chromosomes 12, 15, and 18 in this cell type (Fig. 2b,c; Supplementary Fig. 2). Furthermore, we assessed the spatial positioning of chromosomes 12 and 15 with respect

to the nucleolus by chromosome painting combined with immunofluorescence for NPM. We determined that these chromosomes are closely positioned one to another and juxtaposed to the same nucleolus in 47% of the analyzed cells ($N=200$) (data not shown). Taken together, these results demonstrate that the positioning of *Zac1* transcription close to the nucleolus correlates with the localization of the gene. More importantly, we show that such nuclear distribution is not due the presence of NORs on chromosome 10.

Spatial organization of the *Zac1* gene transcription

Our initial analysis of the topological distribution of *Zac1* gene transcription was performed using a BAC probe which also covers two additional genes: *Stx11* and *Sf3b5*. In order to confirm our observations, we repeated the RNA-FISH experiments using probes addressed specifically to coding exons VIII, IX, and X and intron X–XI generated by PCR. Except for the probe addressed to exon IX, the others

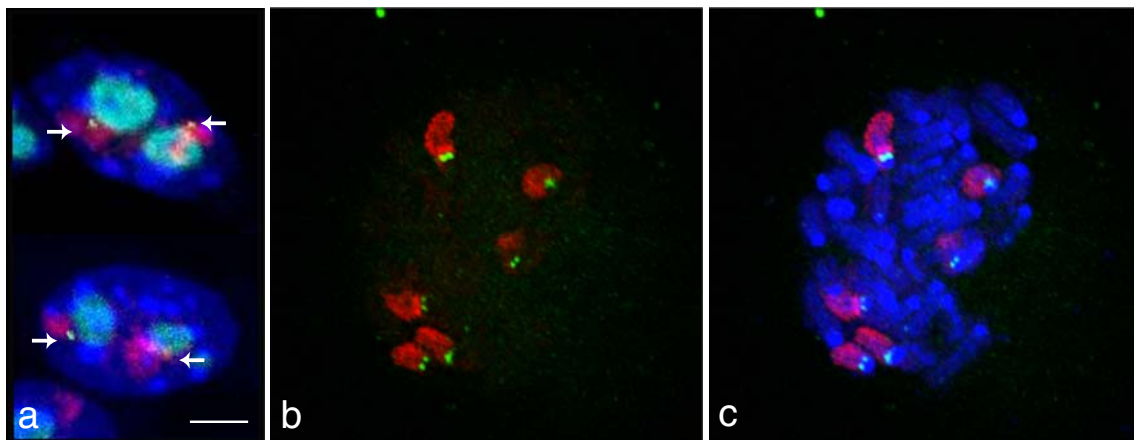


Fig. 2 Chromosome 10 and *Zac1* gene proximal positioning to the nucleolar compartment and NORs mapping in the G7 cell line. Simultaneous chromosome painting and locus-specific DNA FISH on PFA fixed cells showing the *Zac1* gene (yellow, arrows), chromosome 10 (red), and nucleoli (cyan) clustering. Average image of two

consecutive focal planes of the 3D z-stack used for data collection are shown (a). Dual FISH analysis with pA and pB rDNA (green) and whole painting probes for chromosomes 12, 15, and 18 (red) on metaphase chromosomes. Counterstaining with DAPI (blue), scale bar=2 μ m (b, c)

detected their target, however, with heterogeneous results in terms of signal intensity. The hybridizations with these fragments, in particular the hybridization with the intron probe, indicate that we detect both the primary transcript and mature RNA from this gene (Supplementary Figs. 1a and 3a). Control RNA FISH on RNase-treated cells did not give any signal. Furthermore, combined RNA–DNA FISH experiments with the *Zac1*-5 fragment, which reproducibly detects transcripts from exon X of *Zac1* and the BAC RP23-82B19, showed co-localization of both signals (Supplementary Fig. 3b). We then performed the RNA-FISH analysis with the *Zac1*-5 probe combined with immunostaining for NPM and found that 54% of all signals were juxtaposed to the nucleolus (Fig. 3a). In summary, the frequencies of signals proximally positioned (juxtaposed) to nucleoli were similar across the three different types of probes used, i.e., 57% for RNA FISH with the BAC clone, 54% for RNA FISH with the probe for exon X, and 59% for DNA FISH with the BAC clone.

In addition to confirming that the preferential positioning of *Zac1* gene transcription occurred close to the nucleolus, these experiments showed that the size and the fluorescent intensity of the signals located juxtaposed and far from the nucleolus were clearly different (Fig. 3a). Therefore, we determined the 3D volume and the average intensity level of the FISH signals on the images acquired with the *Zac1*-5 probe. Statistical analysis of the measurements showed that there is no significant difference in the average intensity between both alleles ($p=0.80$). However, the analysis proved that the signals juxtaposed to nucleoli are bigger than those further away ($p<0.001$) (Fig. 3b, c). Similar comparative analyses performed for the Splicing Factor 3B Subunit 5 gene (*Sf3b5*) mRNA, the nearest gene to *Zac1*, showed that there are no statistical differences in the

volume and average intensity between both transcription sites (see below). Furthermore, RNA-FISH experiments with a probe generated by nick translation of a cDNA clone (A95504) for *Zac1* on G7 cells and on a breast cancer cell line confirmed that signals located close to nucleoli are larger than those located far from this nuclear compartment (Fig. 3d, e). There may be differences in the transcription level between *Zac1* alleles; however, it cannot be inferred from these experiments. Indeed, qRT-PCR analysis showed that the transcription level of *Sf3b5* is higher than that of *Zac1* gene and yet its signals are smaller and weaker (see below). Our RNA-FISH results suggest, instead, that a retention mechanism may cause an accumulation of mRNA at the transcription site near the nucleolus.

Zac1 gene transcripts associate with nucleoli

To determine whether *Zac1* mRNA is bound to nucleoli, we isolated this nuclear compartment from MEF cells, screened the preparations using phase contrast optical microscopy and isolated their total RNA for cDNA synthesis. Thereafter, the presence of *Zac1* transcripts in the nucleolus preparations was assessed by qRT-PCR with a set of primers for exon VIII of the *Zac1* gene. As positive and negative controls, we used primers for *M. musculus* 45S pre-rRNA (AN: X82564.1) and β -actin, respectively (Supplementary Fig. 1a; Table 1 in Supplementary File 1). We effectively detected an enrichment of *Zac1* mRNA in the nucleolar fraction in comparison to the whole-cell extract. Similar results were obtained for pre-rRNA, although with marked differences in the C_T , reflecting the differences in the amounts of both transcript types in the nucleolus. Conversely, β -Actin transcripts were more abundant in the cell extract than in the isolated nucleoli (Fig. 3f). Additional qualitative

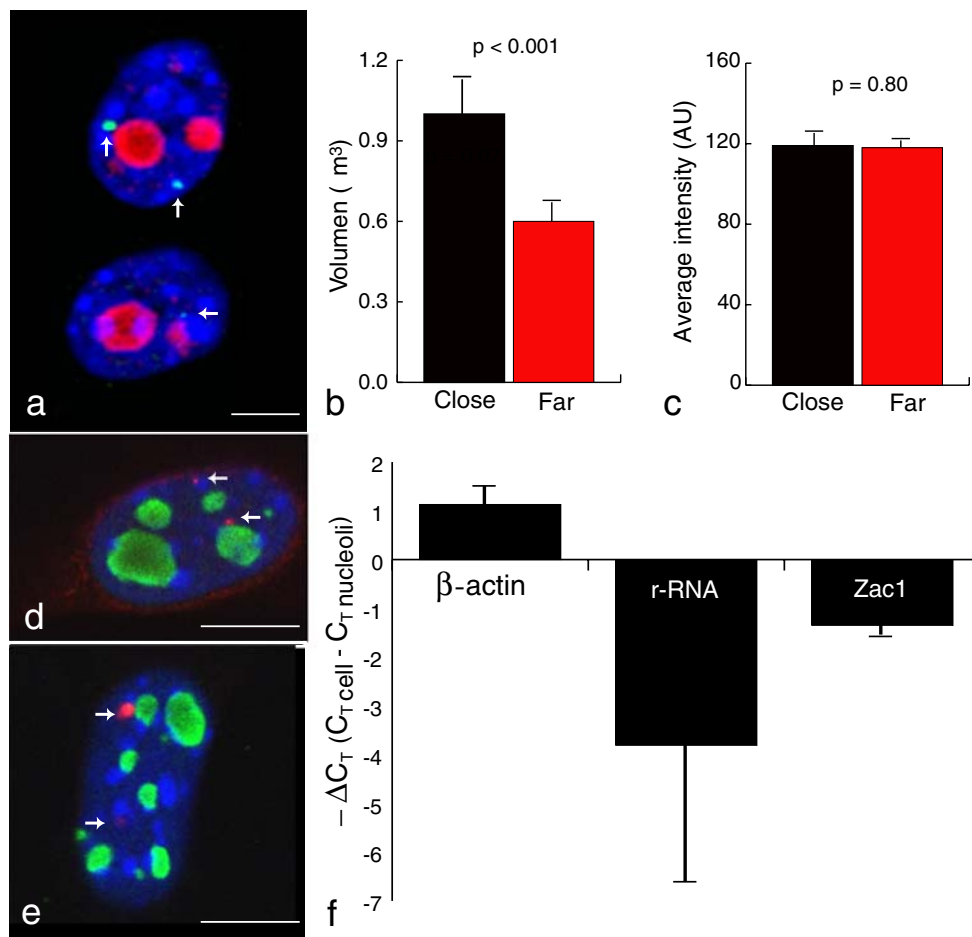


Fig. 3 Spatial organization of *Zac1* gene transcription. RNA FISH with the *Zac1*-5 fragment (green, arrows), nucleoli immunostaining with NPM (red), and DAPI counterstaining (blue) showed differences in the size of the signals localized close to and far from the nucleolus. Average image of two consecutive focal planes of the 3D z-stack used for data collection is shown. Scale bar=4 μm (a). Graphs represent the mean volume and the mean of the average intensity of close and far localized signals. Error bars represent SE ($N=100$). Differences between these two groups were statistically probed by *T* test ($p<0.001$ for volume and $p=0.80$ for average intensity) (b–c). RNA FISH with a plasmid probe (red, arrows) for *Zac1* on G7 cells (d) and on a breast cancer cell line (e), nucleoli immunostaining with NPM (green) and DAPI counterstaining (blue) showing the difference in the size of the

signals localized close to and far from the nucleolus (d–e). qRT-PCR was performed on cDNA synthesized from 1 μg of total RNA from whole-cell extracts and isolated nucleoli. *Zac1* and *β-Actin* genes mRNA and pre-rRNA levels in these two RNA sources were estimated from the information regarding the number of cycles necessary for reaching the lineal amplification threshold (C_T). The graph expresses the differences in the C_T value between the two RNA sources (ΔC_T). Note that the positive value for *β-actin* means that less number of cycles are necessary to reach lineal amplification in the whole-cell extract compared to the nucleoli fraction, and vice versa for *Zac1* and rRNA. Four independent experiments were performed with similar results. Error bars represent SD (f)

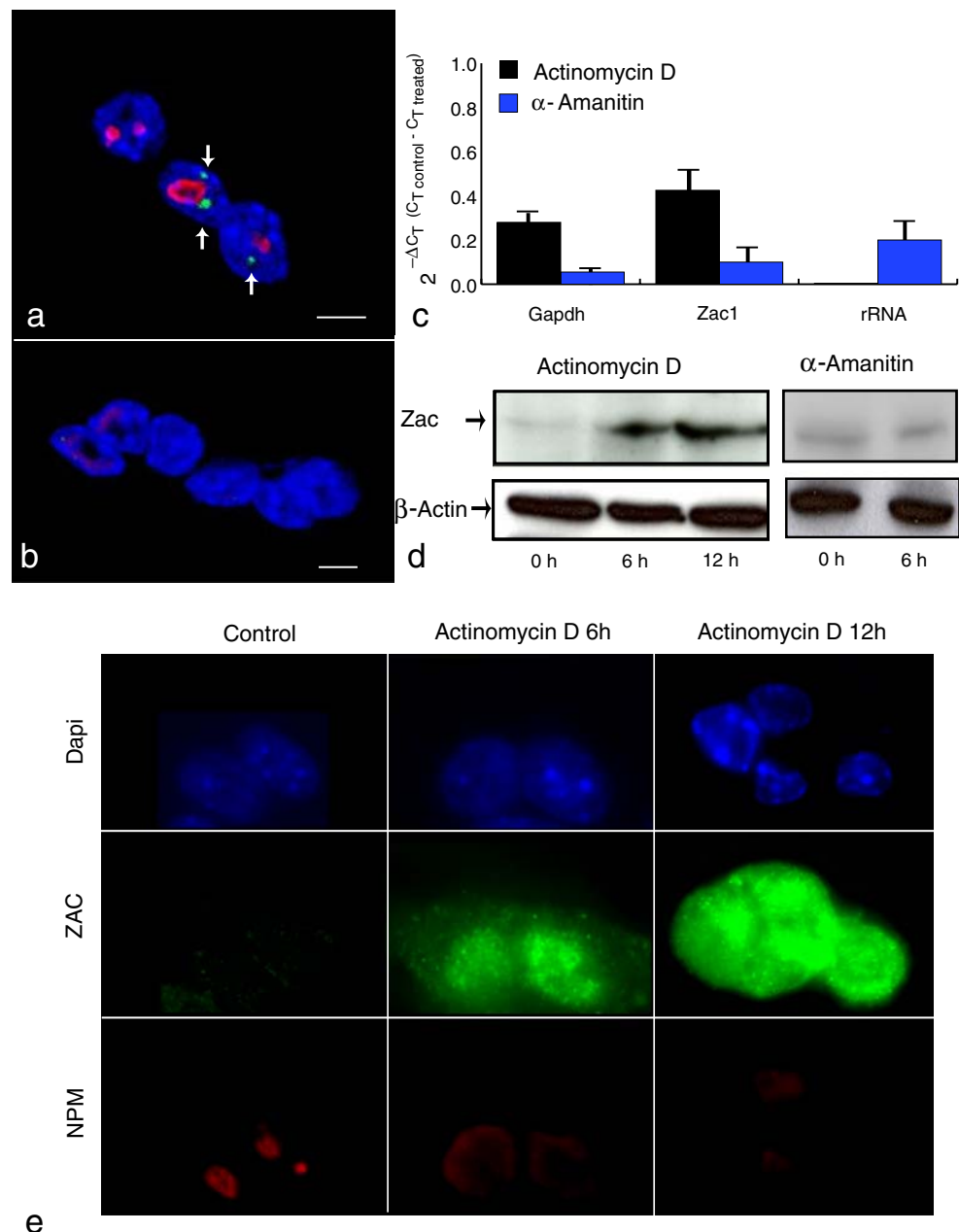
evidence for the existence of such nucleolar retention comes from the observation that MEF cells with high levels of *Zac1* mRNA had multiple FISH signals decorating the nucleoli (Supplementary Fig. 5/movie).

Nucleolus disassembly induces *Zac1* gene expression

To gain insights into the relationship between *Zac1* function and ribosomal gene transcription, we investigated *Zac1* transcription in the context of RNA polymerase inhibition. Treatment of MEFs with Actinomycin D, a known repressor of ribosomal RNA transcription that causes nucleolar components to segregate, resulted in disappear-

ance of *Zac1* mRNA fluorescent signal concomitantly with nucleolar disassembly. In contrast, the *Zac1* mRNA signal remained unchanged after cell treatment with α -amanitin (Fig. 4a, b). Interestingly, nucleoli reassembling occurred after Actinomycin D removal in a dose-dependent manner. Six hours after the drug withdrawal, the *Zac1* gene mRNA signal was again found positioned close to nucleoli at a frequency similar to that observed before treatment (Supplementary Fig. 4). To further explore this behavior, we measured the transcription level of *Zac1*, 45S pre-rRNA, and Glyceraldehyde-3-phosphate dehydrogenase (*Gapdh*) genes by qRT-PCR and ZAC protein expression by immunofluorescence and Western blot analyses after

Fig. 4 *Zac1* gene transcription after RNA polymerase I and II inhibition. *Zac1* gene RNA FISH (green, arrows) with a probe for exon X (*Zac1*-5 fragment) combined with nucleolus immunostaining with an antibody against NPM (red) after treatment with α -amanitin (a) and Actinomycin D (b), respectively. Counterstaining with DAPI (blue), scale bar=4 μ m (a–b). qRT-PCR for *Zac1*, *rRNA*, and *Gapdh* genes using as templates the cDNA synthesized from RNA extracted from the same MEF cells subjected to RNA Pol I and II inhibition treatment and FISH analyses. mRNA levels were quantified and are expressed as fold changes with respect to the control untreated cells ($2^{-\Delta\Delta C_T(C_{T\text{control}} - C_{T\text{treated}})}$). Error bars represent SD of three independent experiments (c). Western blot analyses of MEF cells at different time points after treatment with Actinomycin D and α -amanitin (d). Immunofluorescence analysis of MEF cells at different time points after treatment with Actinomycin D. The ZAC protein level increased concomitantly with the disappearance of *Zac1* and nucleolus fluorescent signals after treatment with Actinomycin D, whereas the level of ZAC protein remained unchanged after treatment with α -amanitin. These experiments were performed four times (e)



both treatments. *Zac1* and *Gapdh* transcription is RNA Pol II dependent and, in agreement with this, their mRNA level diminished after inhibition with α -amanitin. Cell treatment with Actinomycin D blocked rRNA transcription, but also slightly reduced the cellular *Zac1* and *Gapdh* RNA levels (Fig. 4c). Western blot and immunofluorescence analyses showed that the level of ZAC protein changed dramatically, from being practically undetectable at the beginning of the experiment to a several fold increase at 6 h, reaching a maximum at 12 h after treatment with Actinomycin D (Fig. 4d, e). On the contrary, we did not detect changes up to 6 h after treatment with α -amanitin (Fig. 4d). These results demonstrate that *Zac1* RNA synthesis occurs, as

expected, by RNA Pol II. More importantly, the loss of the *Zac1* FISH signal upon treatment with Actinomycin D indicates that the functional integrity of the nucleolus plays a role in retaining *Zac1* mRNA. Whereas the associated increase in the protein level suggests that ZAC protein synthesis may occur from RNA released from its transcription site, rather than from increased transcription of the *Zac1* gene.

We next asked whether the structural and functional integrity of the nucleolus plays a similar role in regulating the behavior of other genes that map to the same genomic region. To this end, we investigated the relative positioning of the *Sf3b5* gene transcripts with respect to the nucleolus

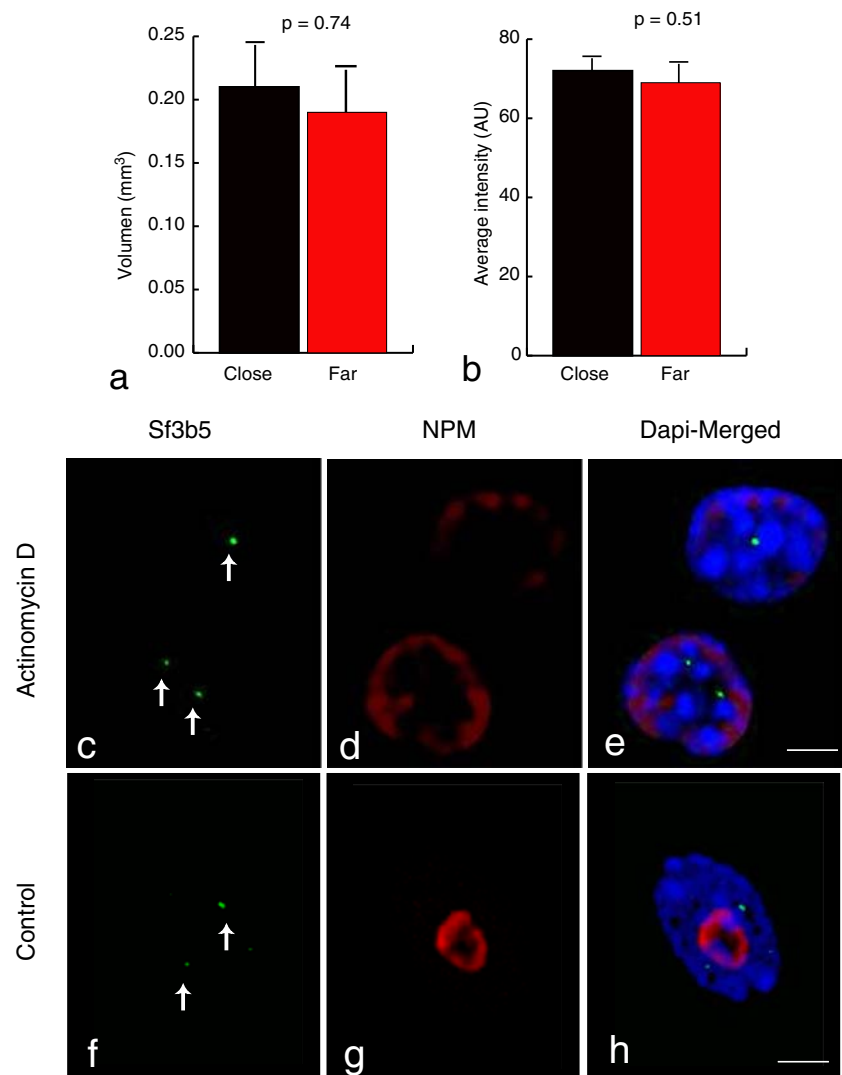
and the effect of nucleoli disassembly on the spatial distribution of *Sf3b5* transcripts. *Sf3b5* is the nearest gene to *Zac1*, mapping approximately 10^5 bp (centromeric) on chromosome band 10A, and encodes the Splicing Factor 3B Subunit 5. We determined the transcription level of this gene by qRT-PCR. Additionally, we performed RNA-FISH analysis with a probe addressed to the single coding region of *Sf3b5* and a fragment of the 3' UTR combined with immunostaining for NPM. The qRT-PCR analysis revealed that the expression of *Sf3b5* is higher than the transcription level of *Zac1* ($C_{T_{Sf3b5}} = 23.4 \pm 0.1$; $C_{T_{Zac1}} = 26.0 \pm 0.4$). Whereas the RNA-FISH experiments showed that the primary transcripts from this gene localize juxtaposed to the nucleolus at a frequency similar to that of *Zac1* (53%). Statistical comparison of the volume and mean intensity of these signals with those that did not localize juxtaposed to nucleoli showed that both signals are similar ($p=0.74$ for volume and $p=0.51$ for average intensity) (Fig. 5a, b). Most importantly, we found that the RNA message remains *in*

situ after RNA Pol I ribosomal RNA transcription blockade with Actinomycin D. The observation that the signal obtained with specific probes does not disappear demonstrates that disassembly of the nucleolus did not affect *Sf3b5* gene spatial organization (Fig. 5c–h). This experiment also provides evidence for the specificity of the link between spatial positioning of *Zac1* transcripts close to the nucleolus and protein expression.

Zac1 mRNA quick release in response to DNA damage

The ZAC protein is a key transcription factor and coactivator for nuclear receptors that induces cell cycle arrest and apoptosis. We hypothesized that inducing DNA damage will trigger the *Zac1* mRNA release and translation. To test this hypothesis, we treated cycling MEF cells in culture with Etoposide to induce cell cycle arrest due to DNA double-strand breaks. We then performed RNA-FISH combined with nucleolus staining and Western blot analy-

Fig. 5 Spatial organization of *Sf3b5* transcripts in the context of RNA polymerase I inhibition. Graphs show that the mean volume and the mean of the average intensity of close and far localized signals are not significantly different by *T* test ($p=0.74$ for volume and $p=0.51$ for average intensity). Error bars represent SE ($N=110$) (a–b). FISH analysis for the *Sf3b5* gene mRNA (green, arrows) with a probe for the single coding region combined with nucleolus immunostaining with an antibody against NPM (red). Counterstaining with DAPI (blue), scale bar=3 μ m. Average image of two consecutive focal planes of the 3D z-stack used for data collection is shown (c–e= treated cells; f–h=untreated control cells)



ses at different time points. We observed that the *Zac1* fluorescent signal was undetectable after 1 h treatment, probably due to the release of the accumulated mRNA, while nucleoli appeared to remain organized (Fig. 6a, b). Interestingly, ZAC protein levels increased dramatically reaching the highest level after 6 h (Fig. 6c). In contrast, in the control experiments (untreated cells), the FISH signals were clearly detected juxtaposed to well-organized nucleoli and the ZAC protein levels were lower than in treated cells. The disappearance of the *Zac1* mRNA signals after DNA damage and consequent protein synthesis indicate that the spatial accumulation of *Zac1* gene transcripts close to nucleoli is not stationary. The findings suggest that such association with nucleoli may serve as a mechanism that makes *Zac1* mRNA readily available when rapid expression of the ZAC protein is needed for cell cycle arrest.

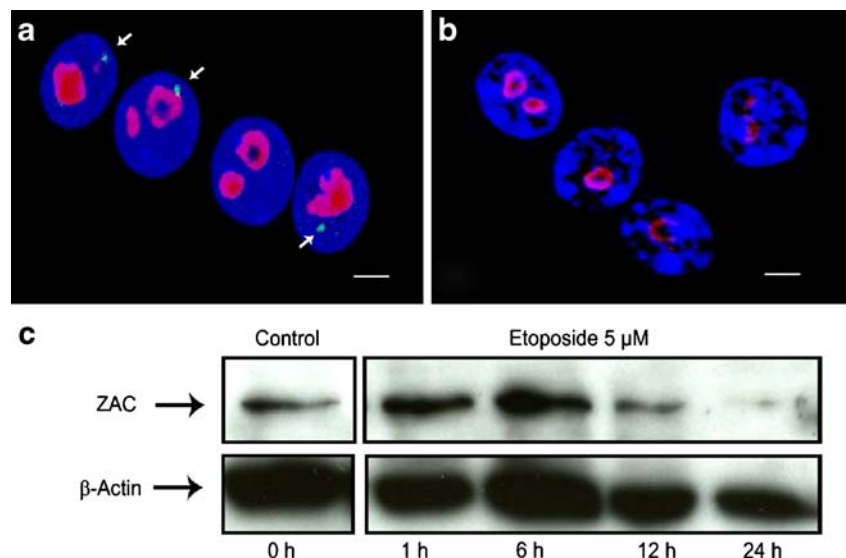
Discussion

The results presented here support the notion that there is a link between the preferential accumulation of *Zac1* mRNA juxtaposed to the nucleolus and ZAC protein expression. We performed RNA FISH for a series of loci including *Asb4*, *c-Met*, *Zac1*, *Myc*, and *Igf2R*, combined with NPM immunostaining to map the position of their transcription sites with respect to the nuclear center (radial distribution) and to each other (relative positioning). We found that the vast majority of active genes were confined to a nuclear sub-domain intermediate between the center and the periphery (60–80% of the nucleus radius). In contrast, no clear pattern of relative positioning with respect to each other was detected. The relative positioning of the fluorescent signals with respect to the nucleolar compart-

ment was different for all these genes. The signals corresponding to *Zac1* mRNA were those that most often localized close to the nucleolus. The gene maps to chromosome 10A1 and 3D DNA FISH revealed that this chromosome is preferentially positioned close to the nucleolus in the interphase nucleus of MEF cells. Ribosomal DNA transcriptional activity accounts for the positioning of transcripts from genes mapping to NOR-bearing chromosomes close to the nucleolus (Kalmarova et al. 2007). This can explain the positioning of *Myc* mRNA, but not the localization of *Zac1* transcripts juxtaposed to the nucleolus, because chromosome 10 does not harbor ribosomal RNA genes. Nucleolar association of NOR-negative chromosomes has been described by Manuelidis and Borden for the human chromosomes 1 and 9 in cells of the central nervous system (Manuelidis and Borden 1988), and more recently Bridger et al. reported the nucleolar association of chromosome 19 in proliferating and quiescent human dermal fibroblasts (Bridger et al. 2000). Of these, only chromosome 19 has homology with mouse chromosome 10, but the syntenic region does not include 10A1 where *Zac1* maps. This indicates that mechanisms other than efficient rDNA transcription determine the localization of this NOR-negative chromosome in close proximity to the nucleolus.

Increasing evidence support the hypothesis that the nucleolus plays a role in nuclear processes other than its primary function in ribosome biogenesis, including mitosis, cell cycle progression, and proliferation and non-rRNAs processing (Boisvert et al. 2007). Here, we show evidence for a link between the nucleolus and *Zac1* gene expression. Murine *Zac1* gene regulation, like its human homolog, is complex and not fully understood. Promoter methylation results in imprinted expression of the paternal allele; however, an additional promoter has been found to control

Fig. 6 Cell cycle arrest induces *Zac1* mRNA release and translation. RNA-FISH analysis with a probe for the *Zac1*-5 fragment on MEF cells untreated (**a**) and after 1-h treatment with 5 μ M of etoposide (**b**). FISH signals in green (arrows), nucleoli were immunoassayed with an anti-NPM antibody (red) and nuclei counterstained with DAPI (blue), scale bar=3 μ m (**a–b**). Western blot analysis to assess the ZAC protein level 1, 6, 12, and 24 h after treatment with Etoposide (5 μ M) (**c**)



the biallelic expression in human and mouse liver, kidney, and skeletal muscle tissue of adult individuals (Piras et al. 2000; Smith et al. 2002; Valleley et al. 2007). In agreement with these results, biallelic expression of *Zac1* occurred in the vast majority of MEFs and 57% of all transcription sites detected with specific probes were juxtaposed to nucleoli. Moreover, we found that the size of the RNA-FISH fluorescent signal correlated positively with the proximity to this compartment. We conclude that such spatial distribution is related to this gene function because disrupting the functional integrity of the nucleolus with Actinomycin D ceased the nuclear retention of the *Zac1* mRNA. Intriguingly, signals localizing far from the nucleolus also disappear upon treatment with Actinomycin D. This suggests that the mechanism responsible of such accumulation must be complex; perhaps it involves main nucleolar components that can be found in small amounts outside the nucleolus, such as NPM (Okuwaki 2008). This notwithstanding, our nucleolus fractionation experiments followed by specific RT-PCR demonstrate that the preferential accumulation of *Zac1* transcripts juxtaposed to nucleoli is due to a physical interaction between this compartment and *Zac1* mRNA. Moreover, we found that the transcripts from the *Sf3b5* gene, which maps only 10⁵bp distant from *Zac1*, also localizes juxtaposed to the nucleolus and that this spatial distribution is not altered by nucleolus disassembly. Thus, it appears that this spatial link with the nucleolus may be specific for *Zac1* transcripts.

Proteomic analysis demonstrated that more than 700 proteins are present in the nucleolus, but only 30% of them participate in ribosome biogenesis and the rest are involved in other molecular processes (Andersen et al. 2005; Boisvert et al. 2007). Interestingly, the nucleolus has a key role in regulating the availability of partner proteins (p19arf and mdm2) involved in p53 stabilization during stress response (Wsierska-Gadek and Horiky 2003). Also, nucleostemin, a protein which interacts with p53, accumulates in the granular area of the nucleolus (Politz et al. 2006). ZAC is a transcriptional coactivator that interacts with p53 and promotes cell cycle arrest and apoptosis by enhancing p53-responsive promoter activity (Huang et al. 2001; Pederson 1998; Rozenfeld-Granot et al. 2002; Visintin and Amon 2000). In contrast to other stress response proteins, ZAC is excluded from the human Nucleolar Proteome Database v 2.0 (<http://www.lamondlab.com/NOPdb/>). Furthermore, Huang et al., using transient transfection assays with a EGFP-tagged fusion ZAC protein, have shown that the full-length protein localize in the nucleus, but not in the nucleolus (Huang et al. 2001). We found, however, that *Zac1* mRNA very often accumulates juxtaposed to nucleoli, suggesting that nuclear retention modulates its expression. Nuclear retention of

CTN-RNA (CAT2 transcribed nuclear RNA), an alternatively spliced primary transcript of the mouse cationic aminoacid transporter 2 gene (*mCAT2*), has been proposed as a mechanism for regulating the level of this gene mRNA, and consequently CAT2 protein expression (Prasanth et al. 2005). Shimizu et al. have recently reported *Myc* mRNA accumulation within the nucleolus, and postulated that such localization may regulate *Myc* gene expression (Shimizu et al. 2007). We found that the *Myc* transcripts were excluded from nucleoli. This difference in the nuclear positioning is likely to be the result of intrinsic differences between the normal embryonic fibroblasts used as a model system in our study and the cancer cells, with over-expression of the *Myc* gene, used by Shimizu et al. More importantly, *Zac1* mRNA signals were not found inside the nucleolus, but positioned very close to this nuclear compartment. This finding poses the question of whether *Zac1* mRNA positioning close to nucleoli is stochastic and if such spatial accumulation has functional relevance. Our data showing that 70% of mRNA signals localize close to nucleoli support the conclusion that *Zac1* mRNA preferentially accumulates in close proximity to nucleoli within the cell nucleus and that such accumulation is relevant for ZAC protein expression. Moreover, the observation of few highly expressing cells in which the primary transcripts were decorating the nucleolus suggests that this particular mRNA positioning may be involved in a post-transcriptional regulation of this gene expression. Alternatively, the nucleolus may contribute to make critical mRNA readily available during the tight cell cycle control or stress response in normal mouse embryonic cells.

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References

- Andersen JS, Lam YW, Leung AK, Ong SE, Lyon CE, Lamond AI, Mann M (2005) Nucleolar proteome dynamics. *Nature* 433:77–83
- Boisvert FM, van Koningsbruggen S, Navascues J, Lamond AI (2007) The multifunctional nucleolus. *Nat Rev Mol Cell Biol* 8:574–585
- Bridger JM, Boyle S, Kill IR, Bickmore WA (2000) Re-modelling of nuclear architecture in quiescent and senescent human fibroblasts. *Curr Biol* 10:149–152
- Cook PR (1999) The organization of replication and transcription. *Science* 284:1790–1795
- Dekker J (2008) Gene regulation in the third dimension. *Science* 319:1793–1794

- Dostie J, Zhan Y, Dekker J (2007) Chromosome conformation capture carbon copy technology. *Curr Protoc Mol Biol* Chapter 21: Unit 21.14.
- Dundr M, Misteli T, Olson MO (2000) The dynamics of postmitotic reassembly of the nucleolus. *J Cell Biol* 150:433–446
- Erickson JM, Rushford CL, Dorney DJ, Wilson GN, Schmickel RD (1981) Structure and variation of human ribosomal DNA: molecular analysis of cloned fragments. *Gene* 16:1–9
- Henderson AS, Eicher EM, Yu MT, Atwood KC (1974) The chromosomal location of ribosomal DNA in the mouse. *Chromosoma* 49:155–160
- Huang SM, Schonthal AH, Stallcup MR (2001) Enhancement of p53-dependent gene activation by the transcriptional coactivator Zac1. *Oncogene* 20:2134–2143
- Kalmarova M, Smirnov E, Masata M, Koberna K, Ligasova A, Popov A, Raska I (2007) Positioning of NORs and NOR-bearing chromosomes in relation to nucleoli. *J Struct Biol* 160:49–56
- Kumaran RI, Thakar R, Spector DL (2008) Chromatin dynamics and gene positioning. *Cell* 132:929–934
- Loreni F, Thomas G, Amaldi F (2000) Transcription inhibitors stimulate translation of 5' TOP mRNAs through activation of S6 kinase and the mTOR/FRAP signalling pathway. *Eur J Biochem* 267:6594–6601
- Manuelidis L, Borden J (1988) Reproducible compartmentalization of individual chromosome domains in human CNS cells revealed by in situ hybridization and three-dimensional reconstruction. *Chromosoma* 96:397–410
- Misteli T (2007) Beyond the sequence: cellular organization of genome function. *Cell* 128:787–800
- Okuwaki M (2008) The structure and functions of NPM1/Nucleophosmin/B23, a multifunctional nucleolar acidic protein. *J Biochem* 143:441–448
- Osborne CS, Chakalova L, Mitchell JA, Horton A, Wood AL, Bolland DJ, Corcoran AE, Fraser P (2007) Myc dynamically and preferentially relocates to a transcription factory occupied by Igh. *PLoS Biol* 5:e192
- Parada LA, McQueen PG, Munson PJ, Misteli T (2002) Conservation of relative chromosome positioning in normal and cancer cells. *Curr Biol* 12:1692–1697
- Parada LA, McQueen PG, Misteli T (2004a) Tissue-specific spatial organization of genomes. *Genome Biol* 5:R44
- Parada LA, Sotiriou S, Misteli T (2004b) Spatial genome organization. *Exp Cell Res* 296:64–70
- Pederson T (1998) The plurifunctional nucleolus. *Nucleic Acids Res* 26:3871–3876
- Piras G, El Kharroubi A, Kozlov S, Escalante-Alcalde D, Hernandez L, Copeland NG, Gilbert DJ, Jenkins NA, Stewart CL (2000) Zac1 (Lot1), a potential tumor suppressor gene, and the gene for epsilon-sarcoglycan are maternally imprinted genes: identification by a subtractive screen of novel uniparental fibroblast lines. *Mol Cell Biol* 20:3308–3315
- Politz JC, Tuft RA, Prasanth KV, Baudendistel N, Fogarty KE, Lifshitz LM, Langowski J, Spector DL, Pederson T (2006) Rapid, diffusional shuttling of poly(A) RNA between nuclear speckles and the nucleoplasm. *Mol Biol Cell* 17:1239–1249
- Prasanth KV, Prasanth SG, Xuan Z, Hearn S, Freier SM, Bennett CF, Zhang MQ, Spector DL (2005) Regulating gene expression through RNA nuclear retention. *Cell* 123:249–263
- Romanova L, Korobova F, Noniashvili E, Dyban A, Zatssepina O (2006) High resolution mapping of ribosomal DNA in early mouse embryos by fluorescence in situ hybridization. *Biol Reprod* 74:807–815
- Rozenfeld-Granot G, Krishnamurthy J, Kannan K, Toren A, Amariglio N, Givol D, Rechavi G (2002) A positive feedback mechanism in the transcriptional activation of Apaf-1 by p53 and the coactivator Zac-1. *Oncogene* 21:1469–1476
- Shimizu N, Kawamoto JK, Utani K (2007) Regulation of c-myc through intranuclear localization of its RNA subspecies. *Biochem Biophys Res Commun* 359:806–810
- Smith RJ, Arnaud P, Konfortova G, Dean WL, Beechey CV, Kelsey G (2002) The mouse Zac1 locus: basis for imprinting and comparison with human ZAC. *Gene* 292:101–112
- Valleley EM, Cordery SF, Bonthron DT (2007) Tissue-specific imprinting of the ZAC/PLAGL1 tumour suppressor gene results from variable utilization of monoallelic and biallelic promoters. *Hum Mol Genet* 16(8):972–981
- Visintin R, Amon A (2000) The nucleolus: the magician's hat for cell cycle tricks. *Curr Opin Cell Biol* 12:752
- Wiblin AE, Cui W, Clark AJ, Bickmore WA (2005) Distinctive nuclear organisation of centromeres and regions involved in pluripotency in human embryonic stem cells. *J Cell Sci* 118:3861–3868
- Wsierska-Gadek J, Horky M (2003) How the nucleolar sequestration of p53 protein or its interplayers contributes to its (re)-activation. *Ann N Y Acad Sci* 1010:266–272