INVITED REVIEW

CrossMark

Psychiatric disorders biochemical pathways unraveled by human brain proteomics

Verônica M. Saia‑Cereda1 · Juliana S. Cassoli1 · Daniel Martins‑de‑Souza1,2 · Juliana M. Nascimento1,3

Received: 15 June 2016 / Accepted: 25 June 2016 / Published online: 4 July 2016 © Springer-Verlag Berlin Heidelberg 2016

Abstract Approximately 25 % of the world population is affected by a mental disorder at some point in their life. Yet, only in the mid-twentieth century a biological cause has been proposed for these diseases. Since then, several studies have been conducted toward a better comprehension of those disorders, and although a strong genetic influence was revealed, the role of these genes in disease mechanism is still unclear. This led most recent studies to focus on the molecular basis of mental disorders. One line of investigation that has risen in the post-genomic era is proteomics, due to its power of revealing proteins and biochemical pathways associated with biological systems. Therefore, this review compiled and analyzed data of differentially expressed proteins, which were found in postmortem brain studies of the three most prevalent psychiatric diseases: schizophrenia, bipolar disorder and major depressive disorders. Overviewing both the proteomic methods used in postmortem brain studies, the most consistent metabolic pathways found altered in these diseases. We have unraveled those disorders share about 21 % of proteins affected, and though most are related to energy metabolism pathways deregulation, the main differences found are

Electronic supplementary material The online version of this article (doi[:10.1007/s00406-016-0709-2](http://dx.doi.org/10.1007/s00406-016-0709-2)) contains supplementary material, which is available to authorized users.

- ² UNICAMP's Neurobiology Center, Campinas, Brazil
- ³ D'Or Institute for Research and Education (IDOR), Rio de Janeiro, Brazil

14-3-3-mediated signaling in schizophrenia, mitochondrial dysfunction in bipolar disorder and oxidative phosphorylation in depression.

Keywords Proteome · Mass spectrometry-based proteomics · Schizophrenia · Bipolar disorder · Major depressive disorder · Postmortem brain

Introduction

Since ancient Egypt, mankind attempts to understand mental illness, however, only around the mid-twentieth century, a probable biological cause was confirmed. Since then, modern psychiatry has established a set of systematic criteria for diagnosis, psychological therapies and the development of new drugs. And despite all progress, the prevalence of neuropsychiatric disorders has not diminished. Strong genetic influence of these diseases has been elucidated, yet the role of such genes is still unclear [\[1\]](#page-10-0). Hence, more detailed molecular-based studies are necessary for a better understanding of mental disorders. Approximately 25 % of the world population will, at some point in their lifetime, be affected by a mental disorder [[2](#page-10-1)]. Among the leading causes of disability, especially among woman between 15 and 44 years old, there are several mental disorders, of which five of these disorders are listed as the first cause of burden, schizophrenia as the fifth and bipolar disorder as the seventh [[3\]](#page-10-2). These diseases cause an increased risk of additional health problems, premature death, in addition to suicide attempts [\[4](#page-10-3), [5\]](#page-10-4).

The burden of mental disorders

Currently, depressive disorders are the most common mental illnesses worldwide, estimated to affect about 350

 \boxtimes Daniel Martins-de-Souza dmsouza@unicamp.br

Laboratory of Neuroproteomics, Department of Biochemistry and Tissue Biology, Institute of Biology, University of Campinas (UNICAMP), Rua Monteiro Lobato, 255, Campinas, SP 13083-862, Brazil

million people of all ages [[3\]](#page-10-2). Major depressive disorder (MDD) is associated with high health costs [[6\]](#page-10-5), and according to the National Comorbidity Survey (NCS) it has a prevalence of 14.4 % over lifetime and 7.1 % on a 12-month period [[7\]](#page-10-6). The main symptoms of this disease are depressed mood and/or loss of interest or pleasure [\[8](#page-10-7)], while secondary symptoms are change in sleep, appetite, fatigue/energy loss, feelings of worthlessness or guilt, diminished concentration and suicidal thoughts [\[8](#page-10-7), [9\]](#page-10-8). The basis for MDD treatment still consists in antidepressants, and only 30–40 % of the patients satisfactorily respond to them [\[10](#page-10-9), [11](#page-10-10)]. There are several hypotheses aiming to explain the molecular basis of MDD; however, the pathophysiology of these diseases is only partially understood [\[12](#page-10-11)[–14](#page-10-12)].

Schizophrenia (SCZ) is a chronic mental disorder that usually emerges at the end of adolescence and develops slowly for months or even years [\[15](#page-10-13)]. SCZ may affect up to 1 % of the world population and presents a hereditability of 80–85 % [\[16](#page-10-14)], which may cause a lifespan reduction in almost 20 years [\[17](#page-10-15)]. A burden that displays different symptoms, which are classified as positive, such as hallucinations, deliria and thought disorders, and as negative, such as social interaction disorders, lack of motivation and anhedonia. Furthermore, cognitive deficiencies, such as the reduction in executive functions, selective attention, working memory and mental flexibility, may also be present [\[18](#page-10-16)]. As a multifactorial disease, SCZ involves exogenous and endogenous factors since the beginning of neurodevelopment [\[19](#page-10-17)]. Some of the molecular aspects of SCZ are still to be unraveled, while the connection among the known aspects has still to be improved toward a more integrated understanding of its physiopathology.

Bipolar disorder (BPD) is another psychiatric disorder that may affect up to 4 % of the world adult population [\[20](#page-10-18)]. It is characterized by two well-defined mood shifts, from manic to depressive mood, and it is possible to have periods with symptoms from both states. The diagnosis of BPD is performed clinically. This is a challenge, since the disease has a great heterogeneity, with unclear limits when compared to other psychiatric disorders [\[21](#page-10-19)]. Lithium is by far the most commonly used drug for BPD and works as a mood-stabilizing agent; nevertheless, its specific way of action was not yet entirely unraveled. However, there are several theories trying to explain its action mechanism, such as ionic channel alterations, or gene expression modulation [\[21](#page-10-19)[–23](#page-10-20)]. A cause factor for BPD remains unknown, what is already known is that some biochemical, genetic and environmental disturbed patterns may trigger the disease.

The study of the brain is the most natural way to understand these neuropathologies, aiming to find possible causes for those brain disorders [[24\]](#page-10-21). In addition, cellular and molecular arrangements differ between brain regions; thus, a better comprehension arises from characterizing them at the molecular level and their correlation with the disease pathobiology. Hence, this review aims to evaluate and connect proteomic studies of human brain from patients with SCZ, MDD and BPD, in order to better understand those diseases. These studies employed a myriad of proteomic techniques, which are described in detail below.

Proteomic methods used in neuropsychiatric studies

2DE/2D‑DIGE

The pioneer technique employed not only in psychiatric studies, but also in proteomic investigations in general, was the two-dimensional electrophoresis (2DE) [\[25](#page-10-22)]. Since the 1970s, when developed [\[25](#page-10-22)], this technique is widely used in proteomic studies. Hence, between 2000 and 2010 half of the articles in proteomics in PubMed employed 2DE as its main method of study [[26\]](#page-10-23), which is still used for some particular questions, such as the study of intact proteins [\[26](#page-10-23)]. The 2DE combines two techniques: isoelectric focusing (IEF), followed by a separation by SDS-PAGE. Therefore, as all techniques of electrophoretic separation, molecules with charge migrate under the influence of an electric field, and their migration velocity will depend on specific features of these molecules, such as size, shape and electrical charge.

Thus, by late 1990s this technique was powered by the development of differential two-dimensional electrophoresis (2D-DIGE) [\[27](#page-10-24)]. Herein, proteins are covalently labeled in their lysine residues with fluorescent cyanins (-Cy3, Cy5 and Cy2), and the samples are mixed prior to electrophoretic separation, enabling precise and more sensitive proteome quantification. Consequently, increasing reproducibility and sensitivity, samples can be compared in a single gel [[28\]](#page-10-25). Inherent limitations of 2DE, which also applies to 2D-DIGE, are the difficulty of separating hydrophobic and extremely acidic or basic proteins, which can be partially solved by protein extraction methods using detergents. Moreover, proteins larger than 150 kDa and smaller than 10 kDa can be missed, demanding experiments using several gels with variable acrylamide concentrations. Computational analyses of gels are rather semi-automated, demanding manual corrections for the quantification of protein expression.

Even with all limitations, 2DE and 2D-DIGE are genuinely a top-down analytical approach [[26](#page-10-23)]. Their resolution power is remarkable as they are capable of resolving more than 10,000 protein spots in a single run [[29\]](#page-10-26), besides resolving protein isoforms and posttranslational modifications.

But their application to proteomics heavily relies on mass spectrometry for the identification of proteins.

Mass spectrometry‑based proteomics

A combination of liquid chromatography and mass spectrometry used for large-scale proteome analysis became popular by the end of the 1990s, when the term "Shotgun Proteomics" was coined [[30](#page-10-27)]. At that point, shotgun proteomics could be simply referred as mass spectrometry-based proteomics. But considering all the recent developments, both terms are actually referring to "mass spectrometrybased bottom-up proteomics." This consists primarily of the analysis of a digested proteome, which undergoes a chromatographic separation followed by MS/MS analysis [[31,](#page-10-28) [32\]](#page-10-29). Although this is virtually impossible, the aim here is to unveil the whole proteome of a given sample. For that, depending on the sample analyzed, single liquid chromatography could be insufficient to resolve the complexity of biological protein mixtures, requiring a multidimensional chromatography separation. This concept has been applied since the description of MudPIT (multidimensional protein identification technology) in 2001 [[33\]](#page-10-30), but only more recently multidimensional chromatography separation has been more often used by a significant number of proteomic studies [\[34,](#page-10-31) [35\]](#page-10-32). A given 2D-LC system may employ, for instance, a first separation of peptides by strong cation exchange (SCX) or by reversed-phase column, and this last has recently been more used, followed by separation on a reversed-phase column (RP), which could be employed.

Mass spectrometry‑based quantitative proteomics

In recent years, mass spectrometry-based quantitative proteomics has earned significant space among quantitative techniques for proteins [[36\]](#page-10-33). It is an alternative for antibody-based protein analysis, as virtually any protein can be accurately measured in a large number of samples [[37\]](#page-10-34).

Stable isotope/isobaric labeling approaches

Quantification of proteins is a key aspect in proteomic studies. Efforts in developing effective methods to increase sensitivity and accuracy led to the development of stable chemical (e.g., ICAT, iTRAQ, ICPL and TMT) and metabolic (SILAC, SILAM and ^{15}N) labeling techniques. Isotope-coded affinity tags (ICATs) were the first application of stable isotope labeling to quantitative proteomics [[38\]](#page-10-35). It relies on heavy and light mass tags containing either eight or no deuterium atoms, respectively, allowing the comparison of two samples in one experiment. Isotope-coded protein label (ICPL) follows the same principle, but up to 4 samples could be labeled at once [\[39](#page-10-36)].

Isobaric tags for relative and absolute quantification (iTRAQ) are one the most used in vitro labeling technique in proteomic studies. Quantification consists of different isobaric tags, which label up to eight different samples, and can be used in any biological system [\[40](#page-10-37)]. Proteolytic peptides of each sample are labeled with an iTRAQ specific tag, and then, samples are mixed and further analyzed in LC–MS/MS [[40\]](#page-10-37). iTRAQ tags present three distinct regions: one that reacts with the peptide, a reporter region, and a balance that complements the reporter region mass, making iTRAQ tags isobaric [\[41](#page-11-0)]. Once a given labeled peptide is submitted to MS/MS, the balance and reporter break apart, and the masses of the reporters are measured. The intensity of these reporters is linearly correlated with the quantity of the given peptide.

In addition, among metabolic labeling techniques, stable isotope labeling by amino acids in cell culture (SILAC) is the most employed and relies on the incorporation of non-radioactive, stable isotope containing amino acids in newly synthesized proteins. Culture medium is supplemented with "heavy" amino acids instead of natural amino acids to be incorporated into proteins. Then, both light and heavy-treated cells are mixed and processed together, until analysis by LC–MS/MS, when labeled peptides can be distinguished, and therefore, abundance was determined by relative signal intensities [\[42](#page-11-1)].

Label‑free

On the other hand, there are label-free approaches, which are simpler, require no additional wet-lab experiments and are reproducible and cheaper compared to stable isotope labeling techniques [\[43](#page-11-2)]. However, label-free quantification requires hard and specialized in silico analysis, thus turning relative quantification possible [[44–](#page-11-3)[46\]](#page-11-4). Label-free quantitative analysis is based on two main approaches, the first is to count and compare fragment ion in spectra acquired from peptide derived from a precursor protein [[47,](#page-11-5) [48](#page-11-6)]. The second parameter is the measurement of the chromatographic peaks' areas of peptide precursor ions, which is possible since these peaks are supposed to have a linear correlation with the amount of protein present in the sample [\[49](#page-11-7)]. In addition, label-free quantification does not limit the number of samples and conditions to be compared, which is suitable to longitudinal and clinical proteomics [[50\]](#page-11-8).

Targeted proteomics

SRM

Selected/multiple reaction monitoring (SRM/MRM) is able to detect and perform accurate quantitation of a target protein, or set of proteins, present in complex biological

samples [\[51](#page-11-9)]. This technique is performed with high efficiency in triple quadrupole mass spectrometers (TQ or QqQ), wherein the first analyzer (Q1) achieves the isolation of a given intact peptide (parent ion); the second analyzer (Q2, which is not a proper quadrupole in current mass spectrometers) works as a collision chamber, generating fragments (daughter ions) that will be measured separately and accurately in the third quadrupole (Q3). The various transitions between the precursor and fragment ion pairs are monitored over time, and when combined with standard chromatogram, peak retention time and intensity produce a high selectivity for quantification [[52–](#page-11-10)[55\]](#page-11-11).

More recently, targeted-MS has been also employed in Q-TOF and Orbitrap mass spectrometers. The latest performs the so-called parallel reaction monitoring (PRM), which measures daughter ions on a HR/AM mass analyzer instead of a quadrupole. This allows the parallel detection of all daughter ions from a given parent ion at once [\[56](#page-11-12), [57](#page-11-13)]. PRM offers alternative ways to conduct targeted proteomics studies with comparable performance as SRM [\[58](#page-11-14)]. In a recent study, this type of acquisition was also implemented in an instrument of the type quadrupole time of flight (QqTOF). Using complex biological samples, selectivity and reproducibility of PRM compared to SRM were evaluated, showing a satisfactory performance of this instrument using this technique [\[59](#page-11-15)].

Antibody‑based techniques

Immunoassays have been the basis for protein measurement for over half a century, with a limited range of tests available mainly for diagnosis [[60\]](#page-11-16). Historically, Western blotting is the most common technique for immunodetection of proteins in complex samples. It consists basically in the transference of proteins from a gel to a membrane where the specific protein labeling with the respective antibodies will be performed [[61\]](#page-11-17). Alternatively, commercially available enzyme-linked immunosorbent assay (ELISA) can be more sensitive if compared to Western blot, in addition to better relative quantification using recombinant proteins [[62\]](#page-11-18).

Western blot and ELISA are both commonly used in proteomics as validation tools for differences in protein expression. The major drawback is that these techniques depend on specific and well-characterized antibodies, which can be challenging, especially for the study of posttranslational modifications. More recently, an analysis of large-scale antibody-based proteomic technique has emerged. Though using a lower throughput compared to mass spectrometry, this analysis employs multiplexed dye-coded microspheres, coated with antibodies. Those microspheres are used for identification and quantification of hundreds of proteins simultaneously, depending on the antibody composition of the assay, in dozens of individual samples [\[63](#page-11-19)]. The amount of sample required is also an advantage compared to other antibody-based techniques, though it still depends on the quality of those for greater reproducibility [[64–](#page-11-20)[68\]](#page-11-21).

Biochemical pathways associated with neuropsychiatric disorders unraveled by proteomics

This review analyzes every proteomics study published thus far in several postmortem brain regions of patients with schizophrenia (SCZ), bipolar disorder (BPD) and major depression disorder (MDD). All differentially expressed proteins found in these studies were computed. The survey was conducted in PubMed with the following keywords "proteomic/proteome brain and schizophrenia/ bipolar disorder/major depressive disorder," We found 14 articles on SCZ studies [[69–](#page-11-22)[83\]](#page-12-0), 4 on BPD [[82,](#page-12-1) [84,](#page-12-2) [85\]](#page-12-3) and 7 on MDD [[69,](#page-11-22) [79](#page-12-4), [86–](#page-12-5)[88\]](#page-12-6), which found up- and downregulated proteins that were compiled and are presented in Supplementary table 1. BPD studies unraveled 731 differentially expressed proteins, while 412 proteins were discovered in SCZ studies and 187 proteins in MDD. All these proteins were further analyzed only by Ingenuity Pathway Analysis software (IPA, Ingenuity Systems, QIAGEN, Redwood, CA, USA; [www.ingenuity.com\)](http://www.ingenuity.com), using curated connectivity information from the literature to determine interactions network among differentially expressed proteins and determine canonical pathways in which they are involved [[89\]](#page-12-7). Parameters used in the IPA software were: "genes only," "include direct and indirect relationship" and "do not include endogenous chemicals." Only molecules and/or relationships in humans were considered, and all cell types/tissues were taken into account, using prediction mode assigned to experimentally observed OR high.

Similarities among disorders

We compared the similarities of differentially expressed proteins associated with SCZ, MDD and BPD and found a small overlap among them (Fig. [1\)](#page-4-0). About 26 proteins are common among SCZ, MDD and BPD; additional 28 are common between MDD and BPD, and 24 between MDD and SCZ. On the other hand, comparing SCZ and BPD, a greater similarity is observed, with about 146 proteins in common, which supports genomic studies [[90–](#page-12-8)[92\]](#page-12-9). The low overlap among the main psychiatric disorders might support disease specificity at the proteome level.

Additionally, analysis on the STRING—search tool for the retrieval of interacting genes/proteins [\(http://string-db.](http://string-db.org/) [org/](http://string-db.org/))—was performed. This platform consists of a database devoted to protein–protein interaction, which provides a

Fig. 1 Venn diagram depicting differences between expressed proteins in schizophrenia (SCZ), bipolar disorder (BPD) and major depressive disorder (MDD)

comprehensive view of the interactions between proteins in the dataset (Jensen, 2008). Therefore, 24 proteins found differentially expressed in all three diseases were analyzed, as observed in Fig. [2.](#page-4-1) These proteins have a high degree of connectivity between them and are directly related to axonal region of neuronal cells (axon, 0.000334 FDR; neuron projection terminus, 0.000919 FDR; axon terminus, 0.000725 FDR; dendrite, 0.00726 FDR). However, the most correlated cellular component to those proteins was myelin sheath (1.45E−12 FDR), which is formed by oligodendrocytes. These cells are known to be altered in SCZ, MDD and BPD, as shown by both proteomic studies and neuroimaging [[94–](#page-12-10)[96\]](#page-12-11), including its maturation process [\[97](#page-12-12), [98](#page-12-13)].

Myelin is a multilaminar structure surrounding the axons of neurons made by oligodendrocytes in the central nervous system and by Schwann cells in the peripheral nervous system, being an essential structure for the proper functioning of nerve impulse transmission, providing strength and speed [\[99](#page-12-14), [100\]](#page-12-15). Studies of postmortem tissue in SCZ, MDD and BPD patients showed that myelination-related genes have a reduction in mRNA transcripts in patients [[101](#page-12-16)[–103](#page-12-17)]. Damage to myelin can cause sensory-motor dysfunction, cognitive impairment, mental retardation and even death [[100\]](#page-12-15).

The top network found dysregulated, according to IPA (score 28), is related to neurological, psychological and skeletal/muscular disorders, as shown in Fig. [3.](#page-5-0) This network is composed of 31 molecules, among them 12 proteins are altered in diseases, together with 19 partners of these molecules. This network has a central protein, the TP53, which directly or indirectly connects differentially expressed proteins from all three diseases. This protein has

Fig. 2 Differentially expressed proteins commonly found in SCZ, MDD and BPD and their functional correlations using STRING database

Fig. 3 Differentially expressed proteins and predicted proteins as affected by deregulation them in SCZ, MDD and BPD

anti-proliferation function, plays a role in the maintenance of somatic stem cells [[104\]](#page-12-18) and regulates the proliferation and differentiation of neural stem cells (NSC) [\[105](#page-12-19)]. Therefore, NSCs maintenance/renewal, migration, differentiation and death can thus be disturbed and have a link with various nervous system disorders, including neurodegeneration and psychiatric disorders [[106\]](#page-12-20).

Another molecule, REST (repressor element 1-silencing transcription factor), is also connected to the differentially expressed proteins. This transcription factor is required during differentiation, as induces the expression of neural-specific phenotypes. REST-dependent genes encode transcription factors, transmitter release proteins, voltagedependent receptor channels and signaling proteins [\[107](#page-12-21)]. REST is connected to the differentially expressed synaptosomal-associated protein of 25 kDa (SNAP25), which also plays a critical role in modulating voltage-gated calcium channels and neurotransmitter release [[108\]](#page-12-22). In addition, REST is connected to Alpha-internexin (INA) and GAP43, proteins related to cytoskeleton organization and important function in synapsis and plasticity [\[109](#page-12-23), [110\]](#page-13-0). These similar connections observed in all three disorders suggest a common synaptic and neurodevelopmental deregulation among them.

Main biochemical pathways of each disorder

In addition to those common features presented, several differentially expressed proteins were only observed in patients with SCZ, which have shown a link with neurological disease (*p* value 4.18E−03 to 1.09E−40) with 180 proteins involved and psychological disorders (*p* value 1.29E−03 to 1.09E−40) with 132 proteins. Those observed only in BPD resulted in greater overlap with neurological disease (*p* value 2.88E−03 to 3.15E−31) with 255 proteins involved and psychological disorders (2.48E−03 to 3.15E−31) with 197 proteins. Similarly, differentially expressed proteins in MDD had 75 proteins involved in neurological disease (*p* value 2.39E−02 to 3.66E−13) and 55 proteins in psychological disorders (*p* value 1.92E−02 to 3. 66E−13).

According to IPA, the canonical pathways to which these disorders are associated are energy metabolism pathways deregulation, mainly related to oxidative phosphorylation

mitochondrial dysfunction and gluconeogenesis, followed by cell signaling pathways, including signaling by Rho GTPases family, semaphorin signaling in neurons and 14-3-3 mediated signaling.

Among the leading networks to which SCZ differentially expressed proteins are involved were molecular transport, survival, cell death and neurological disease, with 47 connected proteins. For BPD, the main networks were related to neurological disease, psychological disorders, cellular assembly and organization, with 41 proteins included in those networks. While differentially expressed proteins on MDD were mainly related to cellular assembly and organization, cellular function and maintenance, cardiovascular system development and function, they have shown 37 proteins connected to those networks. As presented in Fig. [4,](#page-7-0) those proteins have a high connectivity among them, indicating a stronger correlation between pathways, which was recently reinforced by large genome studies with patients of these three disorders [\[111](#page-13-1)]. Similarities between those diseases indicate common deregulation basis, yet it is proposed [\[112\]](#page-13-2) that at some point in development they follow different paths to become distinct disorders.

Schizophrenia

Getting an insight into differentially expressed proteins on schizophrenia brains (21.6 %—25 out of 116 differentially expressed proteins), we observe significant association with 14-3-3-mediated signaling (*p* = 1.35E−18) (Fig. [5\)](#page-8-0). The 14-3-3 proteins are abundantly expressed in the brain and interact with a wide variety of cellular proteins, including kinases, phosphatases and transmembrane receptors [[113,](#page-13-3) [114](#page-13-4)]. These proteins regulate intracellular signaling, cell division and differentiation, ion channel function, apoptosis, neurodegeneration and dopamine synthesis [[114,](#page-13-4) [115](#page-13-5)]. Moreover, 14-3-3 proteins have already been implicated in neurological disorders such as Parkinson's, Alzheimer's and Huntington diseases [[116\]](#page-13-6), additionally to psychiatric diseases, such as SCZ [[117–](#page-13-7)[120\]](#page-13-8).

Indeed, the 14-3-3ζ-deficient mice have significant defects in functions such as working memory, sensory gating and associative learning [\[121](#page-13-9), [122\]](#page-13-10), which are related to long-term synaptic plasticity [[122\]](#page-13-10), and defects in neuronal migration [\[121](#page-13-9)], integrating symptoms associated with SCZ-like behavior. Antipsychotic medications, such as haloperidol and olanzapine, affect the expression of 14-3-3 proteins [[119\]](#page-13-11), endorsing association of this protein family with the disease.

Furthermore, proteins 14-3-3 zeta/delta (YWHAZ) have a broader incidence in the studied brain regions, which was found in five regions (corpus callosum (CC), anterior temporal lobe (ATL), anterior cingulate cortex (ACC), dorsolateral prefrontal cortex (DLPC) and mediodorsal thalamus (MDT)). This protein is involved in cell cycle, recognition of DNA alterations, apoptosis, dynamic changes of cytoskeleton and control of gene expression transcription [[123\]](#page-13-12). Furthermore, new evidence indicates an important role in neurogenesis and cell migration [\[124](#page-13-13)].

Another protein commonly found altered was glial fibrillary acidic protein (GFAP). The protein was differentially expressed in seven distinct regions [MDT, DLPC, CC, ACC, insular cortex (IC), frontal cortex (FC) and Wernicke's area (WA)]. GFAP is found in glial cells of the central nervous system, being a classical marker for astrocytes [\[125](#page-13-14)]. Previous studies showed both GFAP mRNA and protein are decreased in patients with SCZ and BPD [[79,](#page-12-4) [126](#page-13-15)]. Astrocytes play important roles in brain immune response, synaptic function, debug ions and cellular transmitters, neuronal metabolism and migration [[127–](#page-13-16)[129](#page-13-17)]. Thus, GFAP, as the main protein of intermediate filament in astrocytes, is a widely studied protein in diseases related to brain and is also very important during development [[130\]](#page-13-18).

Bipolar disorder

Several diseases, such as SCZ, BPD, Alzheimer's and Parkinson's, have some pathophysiological mechanisms in common, including the production of reactive species of oxygen (ROS) and the accumulation of mitochondrial DNA damage (mtDNA), which together result in mitochondrial dysfunction [\[131](#page-13-19)]. Recent studies with BPD patients have revealed differentially expressed proteins and mRNA related to mitochondrial dysfunction [[132,](#page-13-20) [133\]](#page-13-21), particu-larly oxidative phosphorylation [\[134](#page-13-22)]. Mitochondrial dysfunction had higher correlation with BPD, with *p* value of 9.86E−23 and 24.2 % (40/165), as observed in Fig. [6.](#page-9-0) In addition, BPD is also associated with mitochondrial DNA (mtDNA) mutations and polymorphisms [\[135](#page-13-23), [136\]](#page-13-24). These mutations cause an imbalance of mitochondrial enzymes, which can affect energy metabolism. This imbalance can lead to prejudices in major mitochondrial functions, as to synaptogenesis and neuronal plasticity, shown altered in BPD [\[137](#page-13-25), [138](#page-13-26)].

Furthermore, the proteins superoxide dismutase [Cu– Zn] (SOD1), GFAP and stathmin (STMN1) have been identified differentially expressed in several brain regions of patients with BPD. SOD1 is the major intracellular form of the SOD enzyme family, which catalyze the removal of superoxide free radicals within the cells and are increasingly recognized for their key role in response to oxidative stress [[139\]](#page-13-27). This protein is also associated with replication of stress response genes, DNA damage response, stress response and general Cu/Fe homeostasis [\[140](#page-13-28)]. SOD1 is widely associated with psychiatric illnesses on proteomic

Fig. 4 Interaction network depicting similar proteins among diseases

studies, such as BPD and SCZ, on both blood and brain samples [\[141](#page-13-29)[–146](#page-13-30)].

Stathmin, on the other hand, has an important function in mitosis. Moreover, it plays several roles in cellular processes such as the regulation of cell cycle progression, microtubule dynamics, intracellular transport, cell motility, cell polarity and maintenance of cell shape [\[147](#page-14-0)]. Stathmin expression has been reported increased during neuronal differentiation, plasticity and regeneration, key functions for proper brain functioning, thus explaining its possible alteration in many neurodegenerative diseases [[148\]](#page-14-1).

Major depressive disorder

The leading canonical pathway correlated with MDD was the oxidative phosphorylation (OXPHOS), with *p* value of 3.48E−15, and overlap of 15.4 % of molecules from the pathway (16/104). This pathway, as shown in Fig. [7](#page-9-1), is directly related to the mitochondrial dysfunction shown in BPD results. Disorder of the mitochondrial OXPHOS causes biochemical imbalance of the primary route of ATP production, as this pathway is responsible for coordinating the transport of protons and electrons, which leads to energy production [[149\]](#page-14-2). As OXPHOS is a complex pathway, with about 85 proteins, there are a variety of phenotypes related to this route [\[150](#page-14-3)]. Disturbances in this pathway frequently occur in psychiatric diseases like SCZ, BPD and MDD [[151–](#page-14-4)[154\]](#page-14-5). Recent studies, using a mutant MDD mouse model, have observed dysregulated OXPHOS pathway gene expression on the hippocampus, which may play an important role in the disease [[155\]](#page-14-6).

The main differentially expressed protein in BPD was dihydropyrimidinase-related protein 2 (DPYSL2). This protein participates in regulation of hippocampal neuronal axon formation and establishes neuronal polarization [[156,](#page-14-7) [157](#page-14-8)]. Recently, the protein interactome of DPYSL2 was

Fig. 5 14-3-3-mediated signaling as the main canonical pathway related to differentially expressed proteins in SCZ

described. Among DPLYSL2 interacting proteins are those involved in axon guidance, along with semaphorin interactions and WNT5A signaling [[158\]](#page-14-9). The protein gene is located on chromosome 8 and is widely associated with neuropsychiatric diseases, such as SCZ, BPD and MDD, and neurodegenerative diseases, such as Parkinson's and Alzheimer [[159\]](#page-14-10). This protein was found at significantly lower levels in the frontal cortex of patients with MDD [[79,](#page-12-4) [87](#page-12-24)], in addition to the anterior cingulate cortex [\[69](#page-11-22)], which may cause abnormalities in neurodevelopment.

Concluding remarks

Mental disorders are common worldwide, affecting 1 out of 5 people [\[160](#page-14-11)], and psychiatric disorders contribute significantly to this group. Since they are in general diseases of early ages of onset, these disorders often cause severe damage on patients' lives, such as low level of education, marital instability, occupational status and financial downgrade, as well as high social costs $[161–163]$ $[161–163]$ $[161–163]$. Normally, SCZ, MDD and BPD are only diagnosed when symptoms appear; hence, at this point the disease is already established. As a consequence, disease severity is much higher, proportionally to less effective treatments. Therefore, greater efforts are needed to understand these diseases, aiming for an efficient treatment, thus preventing such damage. Studies in neuroscience have reached enormous progress in understanding the cellular and molecular processes involved in psychiatric diseases [[164\]](#page-14-14), but the pathophysiology of these disorders remains undefined [[165\]](#page-14-15).

Proteomics holds great promise in the understanding of psychiatric disorders [\[165](#page-14-15)], mainly through identification of protein changes in postmortem brains of patients [\[164](#page-14-14)], and toward the large-scale analysis of posttranslational modifications [[166,](#page-14-16) [167](#page-14-17)]. By compiling data from SCZ, MDD and BPD patient research, we have uncovered some similarities referring to signaling pathways altered,

Fig. 6 Mitochondrial dysfunction as the main canonical pathway related to differentially expressed proteins in BPD

Fig. 7 Oxidative phosphorylation as the main canonical pathways related to differentially expressed proteins in MDD

which are mainly related to energy metabolism and signaling. Nevertheless, each of these diseases has molecular particularities, including most frequent differentially expressed proteins, and deregulation of several canonical pathways, which are unveiled by proteomics. However, currently available data are still inconclusive, and several efforts are in course for better comprehension of biological mechanisms of these diseases. This study also highlights the importance of postmortem brains banks for the better comprehension of psychiatric diseases, particularly regarding proteomic studies, which are used for multifactorial diseases such as psychiatric. In addition, we need more engagement among psychiatrists–researchers: It is possible to improve the access of researchers to these samples, thus increasing the number of researches in this area.

Acknowledgments Authors thank Dr. Sabine Bahn (University of Cambridge, UK) for providing access to IPA. JMN, DMS and JCS were supported by the São Paulo Research Foundation (FAPESP) Grants 13/08711-3, 14/21035-0 and 14/14881-1 and by the Brazilian National Council for Scientific and Technological Development (CNPq) Grant 460289/2014-4. VMSC was supported by a CNPq fellowship 151787/2F2014-0.

Compliance with ethical standards

Conflict of interest The authors have no conflict of interest regarding this subject.

References

- 1. Chou I, Chouard T (2008) Neuropsychiatric disease. Nature 455:889. doi:[10.1038/455889a](http://dx.doi.org/10.1038/455889a)
- 2. Gillies D, Buykx P, Ag P, Se H (2015) Consultation liaison in primary care for people with mental disorders. Cochrane Libr. doi:[10.1002/14651858.CD007193.pub2](http://dx.doi.org/10.1002/14651858.CD007193.pub2)
- 3. World Health Organization N (2008) The global burden of disease: 2004 update. Update 2010:146. doi:[10.1038/npp.2011.85](http://dx.doi.org/10.1038/npp.2011.85)
- 4. Van Heeringen C, Marušič A (2003) Understanding the suicidal brain. Br J Psychiatry 183:282–284. doi:[10.1192/bjp.183.4.282](http://dx.doi.org/10.1192/bjp.183.4.282)
- 5. Nock MK, Hwang I, Sampson NA, Kessler RC (2010) Mental disorders, comorbidity and suicidal behavior: results from the National Comorbidity Survey Replication. Mol Psychiatry 15:868–876. doi:[10.1038/mp.2009.29](http://dx.doi.org/10.1038/mp.2009.29)
- 6. Kessler R, Chiu W (2005) Prevalence, severity, and comorbidity of twelve-month DSM-IV disorders in the National Comorbidity Survey Replication (NCS-R). Arch Gen Psychiatry 62:617– 627. doi:[10.1001/archpsyc.62.6.617.Prevalence](http://dx.doi.org/10.1001/archpsyc.62.6.617.Prevalence)
- 7. Kessler RC, Petukhova M, Sampson NA et al (2012) Twelvemonth and lifetime prevalence and lifetime morbid risk of anxiety and mood disorders in the United States. Int J Methods Psychiatr Res 21:169–184. doi:[10.1002/mpr.1359](http://dx.doi.org/10.1002/mpr.1359)
- 8. To S, Zepf R, Woods AG (2005) The symptoms, neurobiology, and current pharmacological treatment of depression. J Neurosci Nurs 32:102–107
- 9. American Psychiatric Association (2013) Diagnostic and statistical manual of mental disorders (DSM-5®). doi[:10.1176/appi.](http://dx.doi.org/10.1176/appi.books.9780890425596.744053) [books.9780890425596.744053](http://dx.doi.org/10.1176/appi.books.9780890425596.744053)
- 10. Rush A (2006) Acute and longer-term outcomes in depressed outpatients requiring one or several treatment steps: a STAR*D report. Am J Psychiatry 163:1905. doi[:10.1176/appi.](http://dx.doi.org/10.1176/appi.ajp.163.11.1905) [ajp.163.11.1905](http://dx.doi.org/10.1176/appi.ajp.163.11.1905)
- 11. Trivedi MH, Rush AJ, Wisniewski SR et al (2006) Factors associated with health-related quality of life among outpatients with major depressive disorder: a STAR* D report. J Clin Psychiatry 67:185–195
- 12. Kupfer DJ, Frank E, Phillips ML (2012) Major depressive disorder: new clinical, neurobiological, and treatment perspectives. Lancet 379:1045–1055. doi:[10.1016/S0140-6736\(11\)60602-8.](http://dx.doi.org/10.1016/S0140-6736(11)60602-8.Major) [Major](http://dx.doi.org/10.1016/S0140-6736(11)60602-8.Major)
- 13. Belmaker RH, Agam G (2008) Major depressive disorder. N Engl J Med 358:55–68. doi:[10.1056/NEJMra073096](http://dx.doi.org/10.1056/NEJMra073096)
- 14. Krishnan V, Nestler EJ (2009) The molecular neurobiology of depression. Nature 455:894–902. doi:[10.1038/nature07455.The](http://dx.doi.org/10.1038/nature07455.The)
- 15. Freedman R (2003) Schizophrenia. N Engl J Med 349:1738– 1749. doi:[10.3760/cma.j.issn.0366-6999.20122551](http://dx.doi.org/10.3760/cma.j.issn.0366-6999.20122551)
- 16. Sullivan PF, Kendler KS, Neale MC (2003) Schizophrenia as a complex trait. Arch Gen Psychiatry 60:1187–1192. doi[:10.1001/](http://dx.doi.org/10.1001/archpsyc.60.12.1187) [archpsyc.60.12.1187](http://dx.doi.org/10.1001/archpsyc.60.12.1187)
- 17. Hegarty JD, Baldessarini RJ, Oepen G (1994) One hundred years of schizophrenia. Am J Psychiatry 151:1409–1416
- 18. Weickert TW, Goldberg TE, Gold JM et al (2000) Cognitive impairments in patients with schizophrenia displaying preserved and compromised intellect. Arch Gen Psychiatry 57:907–913. doi:[10.1001/archpsyc.57.9.907](http://dx.doi.org/10.1001/archpsyc.57.9.907)
- 19. Rapoport J (2012) Neurodevelopmental model of schizophrenia: update 2012. Mol Psychiatry 17:1228–1238. doi[:10.1038/](http://dx.doi.org/10.1038/mp.2012.23.Neurodevelopmental) [mp.2012.23.Neurodevelopmental](http://dx.doi.org/10.1038/mp.2012.23.Neurodevelopmental)
- 20. Keller T (2010) Diagnostic features, prevalence, and impact of bipolar disorder. J Clin Psychiatry. doi[:10.4088/](http://dx.doi.org/10.4088/JCP.8125tx11c) [JCP.8125tx11c](http://dx.doi.org/10.4088/JCP.8125tx11c)
- 21. Sussulini A (2014) Proteomics and metabolomics of bipolar disorder. Proteom Metabol Psychiatry 29:116. doi:[10.1159/000358037](http://dx.doi.org/10.1159/000358037)
- 22. Marmol F (2008) Lithium: bipolar disorder and neurodegenerative diseases Possible cellular mechanisms of the therapeutic effects of lithium. Prog Neuropsychopharmacol Biol Psychiatry 32:1761–1771. doi[:10.1016/j.pnpbp.2008.08.012](http://dx.doi.org/10.1016/j.pnpbp.2008.08.012)
- 23. Maj M (2003) The effect of lithium in bipolar disorder: a review of recent research evidence. Bipolar Disord 5:180–188
- 24. Fornito A, Harrison BJ (2012) Brain connectivity and mental illness. Front Psychiatry 3:1–2. doi[:10.3389/fpsyt.2012.00072](http://dx.doi.org/10.3389/fpsyt.2012.00072)
- 25. O'Farrell PH (1975) High resolution two-dimensional electrophoresis of proteins. J Biol Chem 250:4007–4021. doi:[10.1016/j.bbi.2008.05.010](http://dx.doi.org/10.1016/j.bbi.2008.05.010)
- 26. Oliveira BM, Coorssen JR, Martins-de-Souza D (2014) 2DE: the phoenix of proteomics. J Proteom 104:140–150. doi:[10.1016/j.jprot.2014.03.035](http://dx.doi.org/10.1016/j.jprot.2014.03.035)
- 27. Unlü M, Morgan ME, Minden JS (1997) Difference gel electrophoresis: a single gel method for detecting changes in protein extracts. Electrophoresis 18:2071–2077. doi[:10.1002/](http://dx.doi.org/10.1002/elps.1150181133) [elps.1150181133](http://dx.doi.org/10.1002/elps.1150181133)
- 28. Cramer R (2009) Difference gel electrophoresis (DIGE). Methods Mol Biol. doi:[10.1007/978-1-62703-239-1_1](http://dx.doi.org/10.1007/978-1-62703-239-1_1)
- 29. Klose J, Kobalz U (1995) Two-dimensional electrophoresis of proteins: an updated protocol and implications for a functional analysis of the genome. Electrophoresis 16:1034–1059
- 30. Link AJ, Eng J, Schieltz DM et al (1999) Direct analysis of protein complexes using mass spectrometry. Nat Biotechnol 17:676–682. doi:[10.1038/10890](http://dx.doi.org/10.1038/10890)
- 31. Aebersold R, Mann M (2003) Mass spectrometry-based proteomics. Nature 422:198–207. doi:[10.1038/nature01511](http://dx.doi.org/10.1038/nature01511)
- 32. Kislinger T, Emili A (2003) Going global: protein expression profiling using shotgun mass spectrometry. Curr Opin Mol Ther 5:285–293
- 33. Wolters DA, Washburn MP, Yates JR (2001) An automated multidimensional protein identification technology for shotgun proteomics. Anal Chem 73:5683–5690. doi:[10.1021/ac010617e](http://dx.doi.org/10.1021/ac010617e)
- 34. Taylor P, Nielsen PA, Trelle MB et al (2009) Automated 2D peptide separation on a 1D nano-LC-MS system. J Proteome Res 8:1610–1616. doi:[10.1021/pr800986c](http://dx.doi.org/10.1021/pr800986c)
- 35. Rinas A, Jones LM (2014) Fast photochemical oxidation of proteins coupled to multidimensional protein identification technology (MudPIT): expanding footprinting strategies to complex systems. J Am Soc Mass Spectrom 26:540–546. doi[:10.1007/](http://dx.doi.org/10.1007/s13361-014-1017-6) [s13361-014-1017-6](http://dx.doi.org/10.1007/s13361-014-1017-6)
- 36. MacLean B, Tomazela DM, Shulman N et al (2010) Skyline: an open source document editor for creating and analyzing targeted proteomics experiments. Bioinformatics 26:966–968. doi:[10.1093/bioinformatics/btq054](http://dx.doi.org/10.1093/bioinformatics/btq054)
- 37. Pernikářová V, Bouchal P (2015) Targeted proteomics of solid cancers: from quantification of known biomarkers towards reading the digital proteome maps. Expert Rev Proteomics 9450:1– 17. doi:[10.1586/14789450.2015.1094381](http://dx.doi.org/10.1586/14789450.2015.1094381)
- 38. Gygi SP, Rist B, Gerber SA et al (1999) Quantitative analysis of complex protein mixtures using isotope-coded affinity tags. Nat Biotechnol 17:994–999. doi[:10.1038/13690](http://dx.doi.org/10.1038/13690)
- 39. Schmidt A, Kellermann J, Lottspeich F (2005) A novel strategy for quantitative proteomics using isotope-coded protein labels. Proteomics 5:4–15. doi[:10.1002/pmic.200400873](http://dx.doi.org/10.1002/pmic.200400873)
- 40. Ross PL (2004) Multiplexed protein quantitation in saccharomyces cerevisiae using amine-reactive isobaric tagging reagents. Mol Cell Proteomics 3:1154–1169. doi:[10.1074/mcp.](http://dx.doi.org/10.1074/mcp.M400129-MCP200) [M400129-MCP200](http://dx.doi.org/10.1074/mcp.M400129-MCP200)
- 41. Gafken PR, Lampe PD (2006) Methodologies for characterizing phosphoproteins by mass spectrometry. Cell Commun Adhes 13:249–262. doi[:10.1080/15419060601077917](http://dx.doi.org/10.1080/15419060601077917)
- 42. Ong S, Mann M (2007) PROTOCOL A practical recipe for stable isotope labeling by amino acids in cell culture (SILAC). Nat Protoc. doi[:10.1038/nprot.2006.427](http://dx.doi.org/10.1038/nprot.2006.427)
- 43. Latosinska A, Vougas K, Makridakis M et al (2015) Comparative analysis of label-free and 8-plex iTRAQ approach for quantitative tissue proteomic analysis. PLoS ONE 10:e0137048. doi:[10.1371/journal.pone.0137048](http://dx.doi.org/10.1371/journal.pone.0137048)
- 44. Wong JWH, Cagney G (2010) An overview of label-free quantitation methods in proteomics by mass spectrometry. Proteome Bioinforma 604:273–283. doi[:10.1007/978-1-60761-444-9](http://dx.doi.org/10.1007/978-1-60761-444-9)
- 45. Lu P, Vogel C, Wang R et al (2007) Absolute protein expression profiling estimates the relative contributions of transcriptional and translational regulation. Nat Biotechnol 25:117–124. doi:[10.1038/nbt1270](http://dx.doi.org/10.1038/nbt1270)
- 46. Mallick P, Schirle M, Chen SS et al (2007) Computational prediction of proteotypic peptides for quantitative proteomics. Nat Biotechnol 25:125–131. doi[:10.1038/nbt1275](http://dx.doi.org/10.1038/nbt1275)
- 47. Ishihama Y (2005) Exponentially modified protein abundance index (emPAI) for estimation of absolute protein amount in proteomics by the number of sequenced peptides per protein. Mol Cell Proteomics 4:1265–1272. doi[:10.1074/mcp.](http://dx.doi.org/10.1074/mcp.M500061-MCP200) [M500061-MCP200](http://dx.doi.org/10.1074/mcp.M500061-MCP200)
- 48. Silva JC (2005) Absolute quantification of proteins by LCMSE: a virtue of parallel MS acquisition. Mol Cell Proteomics 5:144– 156. doi:[10.1074/mcp.M500230-MCP200](http://dx.doi.org/10.1074/mcp.M500230-MCP200)
- 49. Chelius D, Bondarenko P (2002) Quantitative profiling of proteins in complex mixtures using liquid chromatography and mass spectrometry. J Proteome Res 1:317–323
- 50. Martins-de-Souza D, Guest PC, Vanattou-Saifoudine N et al (2011) Proteomic technologies for biomarker studies in psychiatry: advances and needs. Int Rev Neurobiol. doi[:10.1016/](http://dx.doi.org/10.1016/B978-0-12-387718-5.00004-3) [B978-0-12-387718-5.00004-3](http://dx.doi.org/10.1016/B978-0-12-387718-5.00004-3)
- 51. Picotti P, Aebersold R (2012) Selected reaction monitoringbased proteomics: workflows, potential, pitfalls and future directions. Nat Methods 9:555–566. doi[:10.1038/nmeth.2015](http://dx.doi.org/10.1038/nmeth.2015)
- 52. Lange V, Picotti P, Domon B, Aebersold R (2008) Selected reaction monitoring for quantitative proteomics: a tutorial. Mol Syst Biol 4:222. doi:[10.1038/msb.2008.61](http://dx.doi.org/10.1038/msb.2008.61)
- 53. Burgess MW, Keshishian H, Mani DR et al (2014) Simplified and efficient quantification of low-abundance proteins at very high multiplex via targeted mass spectrometry. Mol Cell Proteomics 13:1137–1149. doi[:10.1074/mcp.M113.034660](http://dx.doi.org/10.1074/mcp.M113.034660)
- 54. Keshishian H, Addona T, Burgess M et al (2007) Quantitative, multiplexed assays for low abundance proteins in plasma by targeted mass spectrometry and stable isotope dilution. Mol Cell Proteomics 6:2212–2229. doi[:10.1074/mcp.M700354-MCP200](http://dx.doi.org/10.1074/mcp.M700354-MCP200)
- 55. Anderson L, Hunter CL (2006) Quantitative mass spectrometric multiple reaction monitoring assays for major plasma proteins. Mol Cell Proteomics 5:573–588. doi[:10.1074/mcp.](http://dx.doi.org/10.1074/mcp.M500331-MCP200) [M500331-MCP200](http://dx.doi.org/10.1074/mcp.M500331-MCP200)
- 56. Peterson AC (2012) Parallel reaction monitoring for high resolution and high mass accuracy quantitative, targeted proteomics. Mol Cell Proteomics 11(11):1475–1488. doi[:10.1074/mcp.](http://dx.doi.org/10.1074/mcp.O112.020131) [O112.020131](http://dx.doi.org/10.1074/mcp.O112.020131)
- 57. Tong L, Zhou XY, Jylha A et al (2015) Quantitation of 47 human tear proteins using high resolution multiple reaction monitoring (HR-MRM) based-mass spectrometry. J Proteom 115:36–48. doi[:10.1016/j.jprot.2014.12.002](http://dx.doi.org/10.1016/j.jprot.2014.12.002)
- 58. Domon B, Gallien S (2015) Recent advances in targeted proteomics for clinical applications. PROTEOMICS Clin Appl. doi:[10.1002/prca.201400136](http://dx.doi.org/10.1002/prca.201400136)
- 59. Schilling B, MacLean B, Held JM et al (2015) Multiplexed, scheduled, high-resolution parallel reaction monitoring on a

full scan QqTOF instrument with integrated data-dependent and targeted mass spectrometric workflows. Anal Chem 87:10222– 10229. doi[:10.1021/acs.analchem.5b02983](http://dx.doi.org/10.1021/acs.analchem.5b02983)

- 60. Ellington AA, Kullo IJ, Bailey KR, Klee GG (2010) Antibody-based protein multiplex platforms: technical and operational challenges. Clin Chem 56:186–193. doi[:10.1373/](http://dx.doi.org/10.1373/clinchem.2009.127514.Antibody-Based) [clinchem.2009.127514.Antibody-Based](http://dx.doi.org/10.1373/clinchem.2009.127514.Antibody-Based)
- 61. Kurien BT, Scofield RH (2015) Western blotting methods and protocols. doi:[10.1007/978-1-4939-2694-7](http://dx.doi.org/10.1007/978-1-4939-2694-7)
- 62. Lisitsyn NA, Chernyi AA, Nikitina IG et al (2014) Methods of protein immunoanalysis. Mol Biol 48:624–633. doi[:10.1134/](http://dx.doi.org/10.1134/S0026893314050094) [S0026893314050094](http://dx.doi.org/10.1134/S0026893314050094)
- 63. Lynch HE, Sancheza AM, D'Souza MP (2015) Development and implementation of a proficiency testing program for Luminex bead-based cytokine assays. J Immunol Methods. doi:[10.1016/j.jim.2014.04.011.Development](http://dx.doi.org/10.1016/j.jim.2014.04.011.Development)
- 64. Khan SS, Smith MS, Reda D et al (2004) Multiplex bead array assays for detection of soluble cytokines : comparisons of sensitivity and quantitative values among kits from multiple manufacturers. Cytometry B Clin Cytom 39:35–39. doi[:10.1002/](http://dx.doi.org/10.1002/cyto.b.20021) [cyto.b.20021](http://dx.doi.org/10.1002/cyto.b.20021)
- 65. Djoba Siawaya JF, Siawaya D, Roberts T et al (2008) An evaluation of commercial fluorescent bead-based luminex cytokine assays. PLoS ONE 3:1–12. doi[:10.1371/journal.pone.0002535](http://dx.doi.org/10.1371/journal.pone.0002535)
- 66. Nechansky A, Grunt S, Roitt IM, Kircheis R (2008) Comparison of the calibration standards of three commercially available multiplex kits for human cytokine measurement to WHO standards reveals striking differences. Biomark Insights 43:227–235
- 67. Butterfield LH, Potter DM, Kirkwood JM (2011) Multiplex serum biomarker assessments : technical and biostatistical issues. J Transl Med 9:173. doi:[10.1186/1479-5876-9-173](http://dx.doi.org/10.1186/1479-5876-9-173)
- 68. Scott ME, Wilson SS, Cosentino LA (2012) Interlaboratory reproducibility of female genital tract cytokine measurements by luminex: implications for microbicide safety studies. Cytokine 56:430–434. doi[:10.1016/j.cyto.2011.06.011.Interlaboratory](http://dx.doi.org/10.1016/j.cyto.2011.06.011.Interlaboratory)
- 69. Beasley CL, Pennington K, Behan A et al (2006) Proteomic analysis of the anterior cingulate cortex in the major psychiatric disorders: evidence for disease-associated changes. Proteomics 6:3414–3425. doi:[10.1002/pmic.200500069](http://dx.doi.org/10.1002/pmic.200500069)
- 70. English JA, Dicker P, Föcking M et al (2009) 2-D DIGE analysis implicates cytoskeletal abnormalities in psychiatric disease. Proteomics 9:3368–3382. doi[:10.1002/pmic.200900015](http://dx.doi.org/10.1002/pmic.200900015)
- 71. Sivagnanasundaram S, Crossett B, Dedova I et al (2007) Abnormal pathways in the genu of the corpus callosum in schizophrenia pathogenesis: a proteome study. Proteomics Clin Appl 1:1291–1305. doi:[10.1002/prca.200700230](http://dx.doi.org/10.1002/prca.200700230)
- 72. Martins-De-Souza D, Gattaz WF, Schmitt A et al (2009) Prefrontal cortex shotgun proteome analysis reveals altered calcium homeostasis and immune system imbalance in schizophrenia. Eur Arch Psychiatry Clin Neurosci 259:151–163. doi[:10.1007/](http://dx.doi.org/10.1007/s00406-008-0847-2) [s00406-008-0847-2](http://dx.doi.org/10.1007/s00406-008-0847-2)
- 73. Saia-Cereda VM, Cassoli JS, Schmitt A et al (2015) Proteomics of the corpus callosum unravel pivotal players in the dysfunction of cell signaling, structure, and myelination in schizophrenia brains. Eur Arch Psychiatry Clin Neurosci 265:601–612. doi:[10.1007/s00406-015-0621-1](http://dx.doi.org/10.1007/s00406-015-0621-1)
- 74. Martins-De-Souza D, Gattaz WF, Schmitt A et al (2009) Alterations in oligodendrocyte proteins, calcium homeostasis and new potential markers in schizophrenia anterior temporal lobe are revealed by shotgun proteome analysis. J Neural Transm 116:275–289. doi:[10.1007/s00702-008-0156-y](http://dx.doi.org/10.1007/s00702-008-0156-y)
- 75. Martins-de-Souza D, Gattaz WF, Schmitt A et al (2009) Proteomic analysis of dorsolateral prefrontal cortex indicates the involvement of cytoskeleton, oligodendrocyte, energy metabolism and new potential markers in schizophrenia. J Psychiatr Res 43:978–986. doi:[10.1016/j.jpsychires.2008.11.006](http://dx.doi.org/10.1016/j.jpsychires.2008.11.006)
- 76. Martins-de-Souza D, Schmitt A, Röder R et al (2010) Sexspecific proteome differences in the anterior cingulate cortex of schizophrenia. J Psychiatr Res 44:989–991. doi:[10.1016/j.](http://dx.doi.org/10.1016/j.jpsychires.2010.03.003) [jpsychires.2010.03.003](http://dx.doi.org/10.1016/j.jpsychires.2010.03.003)
- 77. Martins-de-Souza D, Maccarrone G, Wobrock T et al (2010) Proteome analysis of the thalamus and cerebrospinal fluid reveals glycolysis dysfunction and potential biomarkers candidates for schizophrenia. J Psychiatr Res 44:1176–1189. doi:[10.1016/j.jpsychires.2010.04.014](http://dx.doi.org/10.1016/j.jpsychires.2010.04.014)
- 78. Föcking M, Lopez LM, English JA et al (2014) Proteomic and genomic evidence implicates the postsynaptic density in schizophrenia. Mol Psychiatry. doi[:10.1038/mp.2014.63](http://dx.doi.org/10.1038/mp.2014.63)
- 79. Johnston-Wilson NL, Sims CD, Hofmann JP et al (2000) Disease-specific alterations in frontal cortex brain proteins in schizophrenia, bipolar disorder, and major depressive disorder. The Stanley Neuropathology Consortium. Mol Psychiatry 5:142– 149. doi:[10.1038/sj.mp.4000696](http://dx.doi.org/10.1038/sj.mp.4000696)
- 80. Prabakaran S, Swatton JE, Ryan MM et al (2004) Mitochondrial dysfunction in schizophrenia: evidence for compromised brain metabolism and oxidative stress. Mol Psychiatry 9(684– 697):643. doi:[10.1038/sj.mp.4001532](http://dx.doi.org/10.1038/sj.mp.4001532)
- 81. Clark D, Dedova I, Cordwell S, Matsumoto I (2006) A proteome analysis of the anterior cingulate cortex gray matter in schizophrenia. Mol Psychiatry 11(459–470):423. doi[:10.1038/](http://dx.doi.org/10.1038/sj.mp.4001806) [sj.mp.4001806](http://dx.doi.org/10.1038/sj.mp.4001806)
- 82. Pennington K, Beasley CL, Dicker P et al (2008) Prominent synaptic and metabolic abnormalities revealed by proteomic analysis of the dorsolateral prefrontal cortex in schizophrenia and bipolar disorder. Mol Psychiatry 13:1102–1117. doi:[10.1038/sj.mp.4002098](http://dx.doi.org/10.1038/sj.mp.4002098)
- 83. Martins-de-Souza D, Gattaz WF, Schmitt A et al (2009) Proteome analysis of schizophrenia patients Wernicke's area reveals an energy metabolism dysregulation. BMC Psychiatry 9:17. doi[:10.1186/1471-244X-9-17](http://dx.doi.org/10.1186/1471-244X-9-17)
- 84. Wesseling H, Gottschalk MG, Bahn S (2014) Targeted multiplexed selected reaction monitoring analysis evaluates protein expression changes of molecular risk factors for major psychiatric disorders. Int J Neuropsychopharmacol 18:pyu015. doi:[10.1093/ijnp/pyu015](http://dx.doi.org/10.1093/ijnp/pyu015)
- 85. Föcking M, Dicker P, English JA et al (2011) Common proteomic changes in the hippocampus in schizophrenia and bipolar disorder and particular evidence for involvement of cornu ammonis regions 2 and 3. Arch Gen Psychiatry 68:477–488. doi:[10.1001/archgenpsychiatry.2011.43](http://dx.doi.org/10.1001/archgenpsychiatry.2011.43)
- 86. Gottschalk MG, Wesseling H, Guest PC, Bahn S (2015) Proteomic enrichment analysis of psychotic and affective disorders reveals common signatures in presynaptic glutamatergic signaling and energy. Metabolism. doi:[10.1093/ijnp/pyu019](http://dx.doi.org/10.1093/ijnp/pyu019)
- 87. Martins-de-Souza D, Guest PC, Harris LW et al (2012) Identification of proteomic signatures associated with depression and psychotic depression in post-mortem brains from major depression patients. Transl Psychiatry 2:e87. doi:[10.1038/](http://dx.doi.org/10.1038/tp.2012.13) [tp.2012.13](http://dx.doi.org/10.1038/tp.2012.13)
- 88. Stelzhammer V, Alsaif M, Chan MK et al (2015) Distinct proteomic profiles in post-mortem pituitary glands from bipolar disorder and major depressive disorder patients. J Psychiatr Res 60:40–48. doi[:10.1016/j.jpsychires.2014.09.022](http://dx.doi.org/10.1016/j.jpsychires.2014.09.022)
- 89. Calvano SE, Xiao W, Richards DR et al (2005) A networkbased analysis of systemic inflammation in humans. Nature 437:1032–1037. doi[:10.1038/nature04362](http://dx.doi.org/10.1038/nature04362)
- 90. Möller HJ (2003) Bipolar disorder and schizophrenia: distinct illnesses or a continuum? J Clin Psychiatry 64:23–27
- 91. Craddock N (2005) Genes for schizophrenia and bipolar disorder? Implications for psychiatric nosology. Schizophr Bull 32:9–16. doi:[10.1093/schbul/sbj033](http://dx.doi.org/10.1093/schbul/sbj033)
- 92. Berrettini WH (2000) Are schizophrenic and bipolar disorders related ? A review of family and molecular studies. Biolo Psychiatry 48(6):531–538. doi[:10.1016/S0006-3223\(00\)00883-0](http://dx.doi.org/10.1016/S0006-3223(00)00883-0)
- 93. Davis KL, Stewart DG, Friedman JI, Buchsbaum M, Harvey PD, Hof PR, Haroutunian V (2003) White matter changes in schizophrenia: evidence for myelin-related dysfunction. Arch Gen Psychiatry 60(5):443–456
- 94. Du F, Cooper AJ, Thida T et al (2013) Myelin and axon abnormalities in schizophrenia. Biol Psychiatry 74:451–457. doi:[10.1016/j.biopsych.2013.03.003](http://dx.doi.org/10.1016/j.biopsych.2013.03.003)
- 95. Flynn SW, Lang DJ, MacKay AL et al (2003) Abnormalities of myelination in schizophrenia detected in vivo with MRI, and post-mortem with analysis of oligodendrocyte proteins. Mol Psychiatry 8:811–820. doi[:10.1038/sj.mp.4001337](http://dx.doi.org/10.1038/sj.mp.4001337)
- 96. Cassoli JS, Guest PC, Malchow B et al (2015) Disturbed macro-connectivity in schizophrenia linked to oligodendrocyte dysfunction: from structural findings to molecules. NPJ Schizophr 1:15034. doi[:10.1038/npjschz.2015.34](http://dx.doi.org/10.1038/npjschz.2015.34)
- 97. Deloulme JC, Raponi E, Gentil BJ et al (2004) Nuclear expression of S100B in oligodendrocyte progenitor cells correlates with differentiation toward the oligodendroglial lineage and modulates oligodendrocytes maturation. Mol Cell Neurosci 27:453–465. doi:[10.1016/j.mcn.2004.07.008](http://dx.doi.org/10.1016/j.mcn.2004.07.008)
- 98. Steiner J, Schmitt A, Schroeter ML et al (2014) S100B is downregulated in the nuclear proteome of schizophrenia corpus callosum. Eur Arch Psychiatry Clin Neurosci 264:311–316. doi:[10.1007/s00406-014-0490-z](http://dx.doi.org/10.1007/s00406-014-0490-z)
- 99. Taveggia C (2016) Schwann cells–axon interaction in myelination. Curr Opin Neurobiol 39:24–29. doi:[10.1016/j.](http://dx.doi.org/10.1016/j.conb.2016.03.006) [conb.2016.03.006](http://dx.doi.org/10.1016/j.conb.2016.03.006)
- 100. Fields RD (2008) White matter in learning, cognition and psychiatric disorders. Trends Neurosci 31:361–370. doi[:10.1038/](http://dx.doi.org/10.1038/nature13314.A) [nature13314.A](http://dx.doi.org/10.1038/nature13314.A)
- 101. Tkachev D, Mimmack ML, Ryan MM et al (2003) Oligodendrocyte dysfunction in schizophrenia and bipolar disorder. Lancet 362:798–805
- 102. Georgieva L, Moskvina V, Peirce T et al (2006) Convergent evidence that oligodendrocyte lineage transcription factor 2 (OLIG2) and interacting genes influence susceptibility to schizophrenia. Proc Natl Acad Sci USA 103:12469–12474. doi:[10.1073/pnas.0603029103](http://dx.doi.org/10.1073/pnas.0603029103)
- 103. Aston C, Jiang L, Sokolov BP (2005) Transcriptional profiling reveals evidence for signaling and oligodendroglial abnormalities in the temporal cortex from patients with major depressive disorder. Mol Psychiatry 10:309–322. doi[:10.1038/](http://dx.doi.org/10.1038/sj.mp.4001565) [sj.mp.4001565](http://dx.doi.org/10.1038/sj.mp.4001565)
- 104. Aparicio S, Eaves C (2009) p53: a new kingpin in the stem cell arena. Cell 138:1060–1062
- 105. Liu H, Jia D, Li A et al (2013) p53 regulates neural stem cell proliferation and differentiation via BMP-Smad1 signaling and Id1. Stem Cells Dev 22:913–927. doi:[10.1089/scd.2012.0370](http://dx.doi.org/10.1089/scd.2012.0370)
- 106. Eisch AJ, Cameron HA, Encinas JM et al (2009) Adult neurogenesis, mental health, and mental illness: hope or hype? J Neurosci 28:11785–11791. doi[:10.1523/JNEURO-](http://dx.doi.org/10.1523/JNEUROSCI.3798-08.2008.Adult)[SCI.3798-08.2008.Adult](http://dx.doi.org/10.1523/JNEUROSCI.3798-08.2008.Adult)
- 107. Baldelli P, Meldolesi J (2015) The transcription repressor REST in adult neurons: physiology, pathology, and diseases (1, 2, 3). eNeuro. doi[:10.1523/ENEURO.0010-15.2015](http://dx.doi.org/10.1523/ENEURO.0010-15.2015)
- 108. Wang Q, Wang Y, Ji W et al (2015) SNAP25 is associated with schizophrenia and major depressive disorder in the Han Chinese population. J Clin Psychiatry 76:76–82. doi[:10.4088/](http://dx.doi.org/10.4088/JCP.13m08962) [JCP.13m08962](http://dx.doi.org/10.4088/JCP.13m08962)
- 109. Biewenga JE, Schrama LH, Gispen WH (1996) Presynaptic phosphoprotein B-50/GAP-43 in neuronal and synaptic plasticity. Acta Biochim Pol 43:327–338
- 110. Suzuki T, Mitake S, Okumura-Noji K et al (1997) Excitable membranes and synaptic transmission: postsynaptic mechanisms. Localization of alpha-internexin in the postsynaptic density of the rat brain. Brain Res 765:74–80
- 111. O'Dushlaine C, Rossin L, Lee PH et al (2015) Psychiatric genome-wide association study analyses implicate neuronal, immune and histone pathways. Nat Neurosci 18:199–209. doi:[10.1038/nn.3922](http://dx.doi.org/10.1038/nn.3922)
- 112. Lee SH (2013) Genetic relationship between five psychiatric disorders estimated from genome-wide SNPs. Nat Genet. doi:[10.1038/ng.2711](http://dx.doi.org/10.1038/ng.2711)
- 113. Foote M, Zhou Y (2012) 14-3-3 proteins in neurological disorders. Int J Biochem Mol Biol 3:152–164
- 114. Tzivion G, Avruch J (2002) 14-3-3 proteins: active cofactors in cellular regulation by serine/threonine phosphorylation. J Biol Chem 277:3061–3064. doi:[10.1074/jbc.R100059200](http://dx.doi.org/10.1074/jbc.R100059200)
- 115. Berg D, Holzmann C, Riess O (2003) 14-3-3 proteins in the nervous system. Nat Rev Neurosci 4:752–762. doi[:10.1038/](http://dx.doi.org/10.1038/nrn1197) [nrn1197](http://dx.doi.org/10.1038/nrn1197)
- 116. Steinacker P, Aitken A, Otto M (2011) 14-3-3 proteins in neurodegeneration. Semin Cell Dev Biol 22:696–704. doi:[10.1016/j.](http://dx.doi.org/10.1016/j.semcdb.2011.08.005) [semcdb.2011.08.005](http://dx.doi.org/10.1016/j.semcdb.2011.08.005)
- 117. Bell R, Munro J, Russ C et al (2000) Systematic screening of the 14-3-3 eta (eta) chain gene for polymorphic variants and case-control analysis in schizophrenia. Am J Med Genet 96:736–743
- 118. Foote M, Qiao H, Graham K et al (2015) Inhibition of 14-3-3 proteins leads to schizophrenia-related behavioral phenotypes and synaptic defects in mice. Biol Psychiatry 78:386–395. doi:[10.1016/j.biopsych.2015.02.015](http://dx.doi.org/10.1016/j.biopsych.2015.02.015)
- 119. Schubert KO, Föcking M, Cotter DR (2015) Proteomic pathway analysis of the hippocampus in schizophrenia and bipolar affective disorder implicates 14-3-3 signaling, aryl hydrocarbon receptor signaling, and glucose metabolism: potential roles in GABAergic interneuron pathology. Schizophr Res. doi:[10.1016/j.schres.2015.02.002](http://dx.doi.org/10.1016/j.schres.2015.02.002)
- 120. Wong AHC, Likhodi O, Trakalo J et al (2005) Genetic and post-mortem mRNA analysis of the 14-3-3 genes that encode phosphoserine/threonine-binding regulatory proteins in schizophrenia and bipolar disorder. Schizophr Res 78:137–146. doi:[10.1016/j.schres.2005.06.009](http://dx.doi.org/10.1016/j.schres.2005.06.009)
- 121. Cheah P-S, Ramshaw HS, Thomas PQ et al (2012) Neurodevelopmental and neuropsychiatric behaviour defects arise from 14-3-3ζ deficiency. Mol Psychiatry 17:451–466. doi[:10.1038/](http://dx.doi.org/10.1038/mp.2011.158) [mp.2011.158](http://dx.doi.org/10.1038/mp.2011.158)
- 122. Qiao H, Foote M, Graham K et al (2014) 14-3-3 proteins are required for hippocampal long-term potentiation and associative learning and memory. J Neurosci 34:4801–4808. doi[:10.1523/](http://dx.doi.org/10.1523/JNEUROSCI.4393-13.2014) [JNEUROSCI.4393-13.2014](http://dx.doi.org/10.1523/JNEUROSCI.4393-13.2014)
- 123. Ruce MIC, Asile IONV, Asile MAV, Îlcea ALMAV (2015) c-abl and YWHAZ gene expression in gastric cancer. Rom J Morphol Embryol 56:717–723
- 124. Toyo-oka K, Wachi T, Hunt RF et al (2014) 14-3-3Ε and Ζ regulate neurogenesis and differentiation of neuronal progenitor cells in the developing brain. J Neurosci 34:12168–12181. doi:[10.1523/JNEUROSCI.2513-13.2014](http://dx.doi.org/10.1523/JNEUROSCI.2513-13.2014)
- 125. Deka H, Sarmah R, Sharma A, Biswas S (2015) Modelling and characterization of glial fibrillary acidic protein. Bioinformation 11:393
- 126. Webster MJ, O'Grady J, Kleinman JE, Weickert CS (2005) Glial fibrillary acidic protein mRNA levels in the cingulate cortex of individuals with depression, bipolar disorder and schizophrenia. Neuroscience 133:453–461. doi:[10.1016/j.](http://dx.doi.org/10.1016/j.neuroscience.2005.02.037) [neuroscience.2005.02.037](http://dx.doi.org/10.1016/j.neuroscience.2005.02.037)
- 127. Coyle JT, Schwarcz R (2013) Mind glue. Arch Gen Psychiatry 57:90–93
- 128. Magistretti PJ, Pellerin L (1999) Cellular mechanisms of brain energy metabolism and their relevance to functional brain imaging. Philos Trans R Soc Lond B Biol Sci 354:1155–1163. doi:[10.1098/rstb.1999.0471](http://dx.doi.org/10.1098/rstb.1999.0471)
- 129. Araque A, Sanzgiri RP, Parpura V, Haydon PG (1999) Astrocyte-induced modulation of synaptic transmission. Can J Physiol Pharmacol 77:7. doi:[10.1139/cjpp-77-9-699](http://dx.doi.org/10.1139/cjpp-77-9-699)
- 130. Middeldorp J, Hol EM (2011) GFAP in health and disease. Prog Neurobiol 93:421–443. doi[:10.1016/j.pneurobio.2011.01.005](http://dx.doi.org/10.1016/j.pneurobio.2011.01.005)
- 131. Pieczenik SR, Neustadt J (2007) Mitochondrial dysfunction and molecular pathways of disease. Exp Mol Pathol 83:84–92. doi:[10.1016/j.yexmp.2006.09.008](http://dx.doi.org/10.1016/j.yexmp.2006.09.008)
- 132. Akarsu S, Torun D, Erdem M et al (2015) Mitochondrial complex I and III mRNA levels in bipolar disorder. J Affect Disord 184:160–163. doi:[10.1016/j.jad.2015.05.060](http://dx.doi.org/10.1016/j.jad.2015.05.060)
- 133. Kim D (2015) Methods of integrating data to uncover genotype–phenotype interactions. Nat Publ Gr. doi[:10.1038/nrg3868](http://dx.doi.org/10.1038/nrg3868)
- 134. Konradi C, Eaton M, MacDonald ML et al (2004) Molecular evidence for mitochondrial dysfunction in bipolar disorder. Arch Gen Psychiatry 61:300–308. doi[:10.1001/archpsyc.61.3.300](http://dx.doi.org/10.1001/archpsyc.61.3.300)
- 135. Quiroz JA, Gray NA, Kato T, Manji HK (2008) Mitochondrially mediated plasticity in the pathophysiology and treatment of bipolar disorder. Neuropsychopharmacology 33:2551–2565. doi:[10.1038/sj.npp.1301671](http://dx.doi.org/10.1038/sj.npp.1301671)
- 136. Rollins B, Martin MV, Sequeira PA et al (2009) Mitochondrial variants in schizophrenia, bipolar disorder, and major depressive disorder. PLoS ONE 4:e4913. doi[:10.1371/journal.](http://dx.doi.org/10.1371/journal.pone.0004913) [pone.0004913](http://dx.doi.org/10.1371/journal.pone.0004913)
- 137. Ohgi Y, Futamura T, Hashimoto K (2015) Glutamate signaling in synaptogenesis and NMDA receptors as potential therapeutic targets for psychiatric disorders. Curr Mol Med 15:206–221. doi[:10.2174/1566524015666150330143008#sthash.K0d80Sgj.](http://dx.doi.org/10.2174/1566524015666150330143008#sthash.K0d80Sgj.dpuf) [dpuf](http://dx.doi.org/10.2174/1566524015666150330143008#sthash.K0d80Sgj.dpuf)
- 138. Du J, Machado-Vieira R, Khairova R (2011) Synaptic plasticity in the pathophysiology and treatment of bipolar disorder. Curr Top Behav Neurosci. doi:[10.1007/7854](http://dx.doi.org/10.1007/7854)
- 139. Che M, Wang R, Wang H, Zheng XFS (2015) Expanding roles of superoxide dismutases in cell regulation and cancer. Drug Discov Today 21:1–7
- 140. Tsang CK, Liu Y, Thomas J et al (2014) Superoxide dismutase 1 acts as a nuclear transcription factor to regulate oxidative stress resistance. Nat Commun 5:1–11. doi:[10.1038/ncomms4446](http://dx.doi.org/10.1038/ncomms4446)
- 141. Coughlin JM, Ishizuka K, Kano SI et al (2013) Marked reduction of soluble superoxide dismutase-1 (SOD1) in cerebrospinal fluid of patients with recent-onset schizophrenia. Mol Psychiatry 18:10–11. doi:[10.1038/mp.2012.6](http://dx.doi.org/10.1038/mp.2012.6)
- 142. Gigante AD, Andreazza AC, Lafer B et al (2011) Decreased mRNA expression of uncoupling protein 2, a mitochondrial proton transporter, in post-mortem prefrontal cortex from patients with bipolar disorder and schizophrenia. Neurosci Lett 505:47–51. doi[:10.1016/j.neulet.2011.09.064](http://dx.doi.org/10.1016/j.neulet.2011.09.064)
- 143. Reddy R, Sahebarao MP, Mukherjee S, Murthy JN (1991) Enzymes of the antioxidant defense system in chronic schizophrenic patients. Biol Psychiatry 30:409–412. doi:[10.1016/0006-3223\(91\)90298-Z](http://dx.doi.org/10.1016/0006-3223(91)90298-Z)
- 144. Tsai M-C, Huang T-L (2015) Thiobarbituric acid reactive substances (TBARS) is a state biomarker of oxidative stress in bipolar patients in a manic phase. J Affect Disord 173:22–26. doi:[10.1016/j.jad.2014.10.045](http://dx.doi.org/10.1016/j.jad.2014.10.045)
- 145. Tunçel ÖK, Sarısoy G, Bilgici B et al (2015) Oxidative stress in bipolar and schizophrenia patients. Psychiatry Res 228:688– 694. doi:[10.1016/j.psychres.2015.04.046](http://dx.doi.org/10.1016/j.psychres.2015.04.046)
- 146. Yao JK, Reddy RD, van Kammen DP (2001) Oxidative damage and schizophrenia: an overview of the evidence and its therapeutic implications. CNS Drugs 15:287–310. doi:[10.2165/00023210-200115040-00004](http://dx.doi.org/10.2165/00023210-200115040-00004)
- 147. Rubin CI, Atweh GF (2004) The role of stathmin in the regulation of the cell cycle. J Cell Biochem 93:242–250. doi[:10.1002/](http://dx.doi.org/10.1002/jcb.20187) [jcb.20187](http://dx.doi.org/10.1002/jcb.20187)
- 148. Chauvin S, Sobel A (2014) Neuronal stathmins: A family of phosphoproteins cooperating for neuronal development, plasticity and regeneration. Prog Neurobiol 126:1–18. doi:[10.1016/j.](http://dx.doi.org/10.1016/j.pneurobio.2014.09.002) [pneurobio.2014.09.002](http://dx.doi.org/10.1016/j.pneurobio.2014.09.002)
- 149. Smeitink J, van den Heuvel L, DiMauro S (2001) The genetics and pathology of oxidative phosphorylation. Nat Rev Genet 2:342–352. doi[:10.1038/35072063](http://dx.doi.org/10.1038/35072063)
- 150. Moslemi A-R, Darin N (2007) Molecular genetic and clinical aspects of mitochondrial disorders in childhood. Mitochondrion 7:241–252. doi[:10.1016/j.mito.2007.02.002](http://dx.doi.org/10.1016/j.mito.2007.02.002)
- 151. Martins-de-Souza D (2014) Proteomics, metabolomics, and protein interactomics in the characterization of the molecular features of major depressive disorder. Dialogues Clin Neurosci 16:63–73
- 152. Rice MW, Smith KL, Roberts RC et al (2014) Assessment of cytochrome C oxidase dysfunction in the substantia nigra/ventral tegmental area in schizophrenia. PLoS ONE 9:e100054. doi:[10.1371/journal.pone.0100054](http://dx.doi.org/10.1371/journal.pone.0100054)
- 153. Sawai H, Takai-igarashi T, Tanaka H (2015) Identification of collaborative activities with oxidative phosphorylation in bipolar disorder. Bioinformation 11:207
- 154. Wesseling H, Chan MK, Tsang TM et al (2013) A combined metabonomic and proteomic approach identifies frontal cortex changes in a chronic phencyclidine rat model in relation to human schizophrenia brain pathology. Neuropsychopharmacology 38:2532–2544. doi:[10.1038/npp.2013.160](http://dx.doi.org/10.1038/npp.2013.160)
- 155. Zubenko GS, Hughes HB, Jordan RM et al (2014) Differential hippocampal gene expression and pathway analysis in an etiology-based mouse model of major depressive disorder. Am J Med Genet B Neuropsychiatr Genet 165B:457–466. doi:[10.1002/ajmg.b.32257](http://dx.doi.org/10.1002/ajmg.b.32257)
- 156. Conde C, Cáceres A (2009) Microtubule assembly, organization and dynamics in axons and dendrites. Nat Rev Neurosci 10:319–332. doi:[10.1038/nrn2631](http://dx.doi.org/10.1038/nrn2631)
- 157. Inagaki N, Chihara K, Arimura N et al (2001) CRMP-2 induces axons in cultured hippocampal neurons. Nat Neurosci 4:781– 782. doi:[10.1038/90476](http://dx.doi.org/10.1038/90476)
- 158. Martins-de-souza D, Cassoli JS, Nascimento JM (2015) The protein interactome of collapsing response mediator protein-2 (CRMP2/DPYSL2) reveals novel partner proteins in brain tissue. Clin Relevance 2:1–25. doi[:10.1002/prca.201500004.This](http://dx.doi.org/10.1002/prca.201500004.This)
- 159. Koide T, Aleksic B, Ito Y et al (2010) SHORT COMMUNI-CATION A two-stage case–control association study of with schizophrenia in Japanese subjects. J Hum Genet 55:469–472. doi:[10.1038/jhg.2010.38](http://dx.doi.org/10.1038/jhg.2010.38)
- 160. Kessler RC, Aguilar-Gaxiola S, Alonso J et al (2011) The global burden of mental disorders: an update from the WHO World Mental Health (WMH) surveys. Epidemiol Psichiatr Soc 18:23–33. doi[:10.1017/S1121189X00001421](http://dx.doi.org/10.1017/S1121189X00001421)
- 161. Kessler RC, Berglund PA, Foster CL et al (1997) Social consequences of psychiatric disorders. 2. Teenage parenthood. Am J Psychiatry 154:1405–1411. doi[:10.1176/ajp.154.10.1405](http://dx.doi.org/10.1176/ajp.154.10.1405)
- 162. Kessler RC, Walters EE, Forthofer MS (1998) The social consequences of psychiatric disorders, III: probability of marital stability. Am J Psychiatry 155:1092–1096. doi:[10.1176/](http://dx.doi.org/10.1176/ajp.155.8.1092) [ajp.155.8.1092](http://dx.doi.org/10.1176/ajp.155.8.1092)
- 163. Kessler R, Foster C, Saunders W, Stang P (1995) Social consequences of psychiatric disorders, I: educational attainment. Am J Psychiatry 152:1026–1032
- 164. Pennington K, Cotter D, Dunn MJ (2005) The role of proteomics in investigating psychiatric disorders. Br J Psychiatry 187:4– 6. doi[:10.1192/bjp.187.1.4](http://dx.doi.org/10.1192/bjp.187.1.4)
- 165. Filiou MD, Turck CW, Martins-De-Souza D (2011) Quantitative proteomics for investigating psychiatric disorders. Proteomics Clin Appl 5:38–49. doi:[10.1002/prca.201000060](http://dx.doi.org/10.1002/prca.201000060)
- 166. Martins-de-Souza D, Guest PC, Vanattou-Saifoudine N et al (2011) The need for phosphoproteomic approaches in psychiatric research. J Psychiatr Res 45:1404–1406. doi:[10.1016/j.](http://dx.doi.org/10.1016/j.jpsychires.2011.04.007) [jpsychires.2011.04.007](http://dx.doi.org/10.1016/j.jpsychires.2011.04.007)
- 167. Martins-de-Souza D, Guest PC, Vanattou-Saifoudine N et al (2012) Phosphoproteomic differences in major depressive disorder postmortem brains indicate effects on synaptic function. Eur Arch Psychiatry Clin Neurosci 262:657–666. doi[:10.1007/](http://dx.doi.org/10.1007/s00406-012-0301-3) [s00406-012-0301-3](http://dx.doi.org/10.1007/s00406-012-0301-3)