

Association of haptoglobin phenotypes with the development of Kaposi's sarcoma in HIV patients

Reinhart Speeckaert · Bob Colebunders · Johan R. Boelaert · Lieve Brochez · Jos Van Acker · Filip Van Wanzele · Robert Hemmer · Marijn M. Speeckaert · Chris Verhofstede · Marc De Buyzere · Vic Arendt · Jean Plum · Joris R. Delanghe

Received: 10 May 2011 / Revised: 20 June 2011 / Accepted: 28 June 2011 / Published online: 12 July 2011
© Springer-Verlag 2011

Abstract Kaposi's sarcoma (KS) is a rare cutaneous tumor caused by human herpes virus-8 (HHV-8) infection that preferentially develops in case of severe immunosuppression, such as in HIV/AIDS disease. Haptoglobin (Hp), a polymorphic multifunctional plasma protein, exerts several immunomodulatory effects and is characterized by a genetic polymorphism leading to three major phenotypes (Hp 1-1, Hp 2-1 and Hp 2-2). This study investigated the influence of Hp genetic polymorphism on the development of KS in HIV-positive patients. 661 HIV patients were enrolled in the study with a median age of 35 years and a median follow-up time of 57 months. Hp phenotyping was performed using hemoglobin-supplemented starch gel electrophoresis. In case of low Hp concentration high pressure gel permeation chromatography (HPGPC) was used. The Hp 1-1 phenotype was associated with a significant higher risk of KS compared to the combined group

of Hp 2-1 and Hp 2-2 patients ($p < 0.0005$) which remained significant after adjustment for possible confounding variables (age, gender and AIDS status) ($p < 0.001$). In contrast, the Hp 2-1 phenotype carried the lowest risk. These findings point to the involvement of Hp phenotypes in the pathogenesis of KS, which may be due to a difference in skin immunosurveillance between the Hp phenotypes.

Keywords Genetic polymorphism · Haptoglobin · HIV · Kaposi's sarcoma

Introduction

Kaposi's sarcoma (KS) is a cutaneous viral-induced malignant tumor, which preferentially develops in immunocompromised patients. The disease presents with erythematous or violaceous macules, plaques or nodules with frequent mucocutaneous involvement. KS can extend to lymph nodes, lungs, liver, spleen and gastrointestinal tract. It was one of the earliest signs of the AIDS outbreak and has led to major morbidity and mortality. The human herpes virus-8 (HHV-8) has been identified as the causing pathogen. HHV-8, a double-stranded DNA virus, infects primarily B lymphocytes and persists lifelong in a latent phase [16]. In case of immunosuppression, HHV-8 can be reactivated and undergo an exponential proliferation. Due to a deficient immunosurveillance, viral-induced mutations are tolerated which may lead to the development of KS [36]. In HIV patients with HHV-8 infection a 20,000- to 50,000-fold increase of KS has been reported [9]. The seroprevalence of HHV-8 infection is highly dependent on the geographic region with high percentages in areas with endemic KS such as Central Africa ($\pm 50\%$) and in regions

R. Speeckaert · L. Brochez
Department of Dermatology, Ghent University Hospital, Ghent, Belgium

B. Colebunders
Institute for Tropical Medicine, Antwerp University Hospital, Antwerp, Belgium

J. R. Boelaert
Department of Nephrology and Infectious Diseases, AZ Sint-Jan Brugge, Bruges, Belgium

J. Van Acker · F. Van Wanzele · M. M. Speeckaert · C. Verhofstede · M. De Buyzere · J. Plum · J. R. Delanghe (✉)
Department of Clinical Chemistry, Ghent University Hospital, De Pintelaan 185, 9000 Ghent, Belgium
e-mail: joris.delanghe@ugent.be

R. Hemmer · V. Arendt
Centre Hospitalier de Luxembourg, Luxembourg, Luxembourg

with classic KS such as the Mediterranean (± 10 –30%). In Western-Europe, Northern America and Asia, the seroprevalence is less than 10% [23].

Haptoglobin (Hp), an α_2 -sialoglycoprotein, is a member of the family of acute phase proteins. The best known function of Hp is capturing heme-bound iron in the circulation released by hemolysis. The Hp gene is characterized by a genetic polymorphism leading to three phenotypes: Hp 1-1, Hp 2-1 and Hp 2-2. Hp phenotypes have different immunomodulating and iron-scavenging capacities which have been clinically confirmed in different pathologies including cancer and infections [14, 21, 33]. Our group previously demonstrated the association between the Hp 2-2 phenotype and a worse outcome in HIV-infected patients [5].

In this study, we investigated the role of Hp polymorphism in the development of KS in HIV-infected patients. This could provide new insights in the etiopathogenesis of KS and highlight the role of the Hp phenotype in this angioproliferative tumor with skin and visceral presentations.

Materials and methods

Patients with HIV infection were recruited from two AIDS reference centers in Belgium (Ghent University Hospital, Antwerp University Hospital) and one in Luxembourg (Centre Hospitalier de Luxembourg). Six hundred sixty one patients were enrolled [553 males and 108 females; median age of diagnosis 34 years; interquartile range (IQR) 28–42 years] in this prospective study between 1989 and 2000. The median follow-up time was 57 months (IQR 26–87 months). As the recruitment of patients for this study started in 1989 and was completed in 2000, AIDS diagnosis was based on the two main definitions produced by the Centers for Disease Control and Prevention (CDC). In the period of 1989 to 1992, AIDS was diagnosed using the 23 clinical conditions that were associated with it [25]. From 1993, the AIDS surveillance case definition was expanded to include all HIV-infected persons with CD4⁺ T lymphocyte counts of less than 200 cells/ μ L or a CD4⁺ percentage of less than 14 [26]. The diagnosis of KS was based on physician diagnosis supported by laboratory, radiologic and histopathologic results. The study was approved by the local Ethical Committee, following the ethical guidelines of the 1975 Declaration of Helsinki.

Venous blood was drawn and allowed to clot for 30 min at room temperature. After centrifugation at 1,500g for 10 min, the supernatant serum was collected for analysis. Hp phenotyping was performed using hemoglobin (Hb)-supplemented starch gel electrophoresis. Briefly, 11.5% hydrolyzed starch (Connaught Laboratories, Willowdale, Canada) was used to prepare a starch gel in a 0.1 mol/L

Tris–citrate buffer (pH 8.86). Electrophoresis was carried out at 200 V in a 0.3 mol/L borate buffer (pH 8.4) during 1 h. The Hp–Hb complexes were visualized by staining the gel with metal-enhanced peroxidase reagents (Pierce, Rockford, IL, USA), as described previously [5]. High pressure gel permeation chromatography (HPGPG) was used in case of low plasma Hp concentration [6].

The iron status was investigated by measuring the serum ferritin concentration in the absence of an acute phase reaction [C-reactive protein (CRP) <1 mg/dL]. A latex-enhanced immunonephelometric method on a BN II analyzer (Siemens, Marburg, Germany) was used to determine serum ferritin concentration [8].

Statistical analyses were calculated with SPSS 17.0 (SPSS Inc., Chicago, IL, USA). The Hp phenotypes were compared by Mann–Whitney *U* analysis and Kruskal–Wallis test. The Chi-square test was used to investigate agreement with the Hardy–Weinberg equilibrium. Multivariate logistic regression was performed to investigate the independent effect of the Hp phenotypes on the development of KS. All values are expressed as median (IQR) and statistical significance was set at $p < 0.05$.

Results

Hundred fifteen patients were found with the Hp 1-1 phenotype, 335 subjects with the Hp 2-1 phenotype and 211 with Hp 2-2. This corresponded to allele frequencies of 0.427 for the Hp1 allele and 0.573 for the Hp2 allele. These values are similar to the allelic distributions reported from large studies in Western European populations [17]. 296 patients (44.8%) had AIDS and 51 patients (7.7%) developed KS. Seven cases of KS were diagnosed at baseline. With a total follow-up time of 3,334 years, the overall incidence was 0.018 case/year or 1 case in 55 years. Comparing the Hp phenotypes, we found 0.033 cases/year (1 case in 30 years) in the Hp 1-1 group, 0.008 cases/year (1 case in 125 years) in the Hp 2-1 group and 0.017 cases/year (1 case in 60 years) in the Hp 2-2 group.

Median age, gender distribution, frequency of AIDS-positive patients and median follow-up time were similar between the different Hp phenotypes (Table 1). As reported earlier [5], the mortality in the Hp 2-2 group was significantly higher than in the Hp 1-1 and Hp 2-1 group. The occurrence of KS was markedly different between the Hp types. In the Hp 1-1 phenotype group, 19 patients (16.5%) developed KS; in the Hp 2-1 group, 15 patients (4.5%) and in the Hp 2-2 group, 17 patients (8.1%). Multivariate logistic regression analysis with adjustment for age, gender and AIDS status showed an increased development of KS in patients with the Hp 1-1 phenotype ($p < 0.0005$) (Table 2). In patients without AIDS, the influence of the

Table 1 Characteristics of the study population according to the Hp phenotype

	Hp 1-1 phenotype (n = 115)	Hp 2-1 phenotype (n = 335)	Hp 2-2 phenotype (n = 211)	p value
Age (years)	34 (29–41)	34 (27–43)	36 (28–43)	n.s.
Gender (males %)	87.0 (n = 100)	83.0 (n = 278)	82.0 (n = 173)	n.s.
Aids (%)	46.1 (n = 53)	42.7 (n = 143)	46.9 (n = 99)	n.s.
Mortality (%)	21.7 (n = 25)	23.3 (n = 78)	32.2 (n = 68)	0.025
Median follow-up time (months)	50 (25–90)	63 (27–88)	50 (25–82)	n.s.
Kaposi's sarcoma (%)	16.5 (n = 19)	4.5 (n = 15)	8.1% (n = 17)	<0.001

n.s. not significant

Table 2 Logistic regression model of KS development according to Hp phenotypes

	β	Wald χ^2	Odds ratio (95% CI)	p value
<i>Model 1 (multivariate)</i>				
Hp 1-1 vs. Hp 2-1 and Hp 2-2				
Hp 1-1 type	1.199	13.820	3.31 (1.76–6.26)	<0.0005
Age	0.020	2.825	1.02 (1.00–1.04)	n.s.
Gender	0.493	0.816	1.64 (0.56–4.76)	n.s.
AIDS	1.255	13.488	3.51 (1.80–6.85)	<0.0005
<i>Model 1 (univariate)</i>				
Hp 1-1 vs. Hp 2-1 and Hp 2-2				
Hp 1-1 type	1.155	0.310	3.17 (1.73–5.83)	<0.0005
Age	0.025	0.011	1.03 (1.01–1.05)	<0.05
Gender	0.893	0.532	2.44 (0.86–6.93)	n.s.
AIDS	1.383	0.332	3.99 (2.08–7.64)	<0.0001
<i>Model 2 (multivariate)</i>				
Hp 2-1 vs. Hp 1-1 and Hp 2-2				
Hp 2-1 type	0.942	8.491	0.39 (0.21–0.74)	<0.005
Age	0.018	2.293	1.02 (1.00–1.04)	n.s.
Gender	0.545	0.999	1.72 (0.59–5.02)	n.s.
AIDS	1.226	12.986	3.41 (1.75–6.64)	<0.0005
<i>Model 2 (univariate)</i>				
Hp 2-1 vs. Hp 1-1 and Hp 2-2				
Hp 2-1 type	-0.971	0.318	0.38 (0.20–0.71)	<0.005
Age	0.025	0.011	1.03 (1.01–1.05)	<0.05
Gender	0.893	0.532	2.44 (0.86–6.93)	n.s.
AIDS	1.383	0.332	3.99 (2.08–7.64)	<0.0001

Hp 1-1 phenotype was more pronounced ($p < 0.005$) compared to patients with AIDS, where only borderline significance was reached ($p = 0.059$) (Fig. 1a–c). Bias due to an increased frequency of mortality in the Hp 2-2 group is unlikely as the median follow-up time was very similar in the Hp 1-1 group compared to the Hp 2-2 group (Table 1). The Hp 2-1 phenotype was associated with the lowest risk of KS development ($p < 0.005$) (Table 2). Ferritin values, measured in 128 patients, were lowest in

patients with Hp 1-1 phenotype [111 $\mu\text{g/L}$ (49–194 $\mu\text{g/L}$)], followed by Hp 2-1 [131 $\mu\text{g/L}$ (69–211 $\mu\text{g/L}$)] and Hp 2-2 patients [195 $\mu\text{g/L}$ (107–401 $\mu\text{g/L}$)].

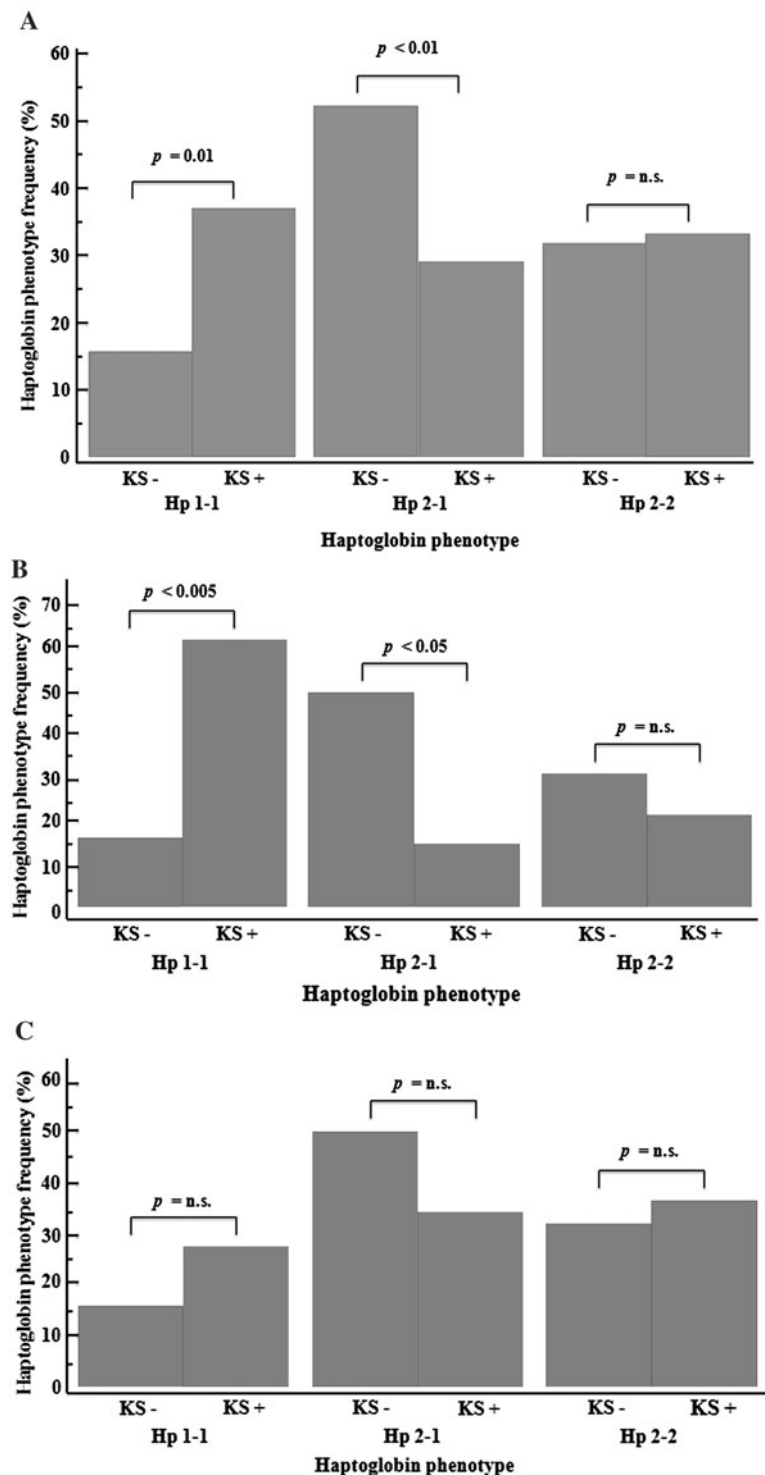
Discussion

As a consequence of the progress in highly active anti-retroviral treatment (HAART) the incidence of KS has decreased dramatically. This was confirmed in our study, since no new cases were diagnosed in the period 1997–2000. Nonetheless, KS may also develop in patients with CD4 counts exceeding $200 \mu\text{L}^{-1}$ [22]. Current treatments for KS (including HAART and chemotherapy) are insufficient to induce complete regression in up to half of the patients [24]. As HIV patients have increasing survival expectations, the future evolution of the incidence of KS on the aging HIV-population is unknown [38]. This implicates that studies of biomarkers identifying a subset of patients at increased risk for KS remain useful.

Our study points to an increased susceptibility of Hp 1-1 patients for the development of KS. A less effective immune response associated with the Hp 1-1 phenotype may partially explain this finding. Hp downregulates T cell proliferation and induces a shift to a Th2 balance which may favor the persistence of viruses [27]. Hp 1-1 complexes phagocytosed by macrophages result in higher Interleukin (IL)-6 and IL-10 levels compared to Hp 2-2 complexes. Interestingly, HHV-8 infection of dendritic cells also induces a Th2 response [13]. IL-6 is overexpressed in patients with KS and in a subset of HHV8+/HIV+ patients, viral-IL-6 can lead to a systemic inflammatory syndrome even in absence of Castleman disease [37]. In transplant patients, a polymorphism in the promoter of the IL-6 gene was associated with an increased risk of KS [11]. IL-6 may play a role in cellular proliferation of HHV-8 infected cells and inhibit apoptosis favoring tumoral development [42, 44].

Hp is mainly produced by the liver, although a small fraction is synthesized by keratinocytes in the skin.

Fig. 1 a Hp phenotype distribution in patients (combined group of subjects with HIV or AIDS) with (*right*) and without (*left*) Kaposi's sarcoma (KS). A significant difference in the haptoglobin (Hp) phenotype frequency was found in the Hp 1-1 ($p = 0.01$) and in the Hp 2-1 group ($p < 0.01$). **b** Hp phenotype distribution in HIV patients with (*right*) and without (*left*) Kaposi's sarcoma (KS). A significant difference in the haptoglobin (Hp) phenotype frequency was found in the Hp 1-1 ($p < 0.005$) and in the Hp 2-1 group ($p < 0.05$). **c** Hp phenotype distribution in AIDS patients with (*right*) and without (*left*) Kaposi's sarcoma (KS). No significant difference in the haptoglobin (Hp) phenotype frequency was found. n.s. = not significant



Moreover, Hp is found in Langerhans cells (LCs) which is probably due to transfer of Hp from surrounding keratinocytes. Hp 1-1 is characterized by the highest concentration of Hp, which may result in a more impaired activity of LCs compared to the other phenotypes as Hp inhibits the functional maturation of LCs [43]. A decreased function of LCs is considered as a primary event in the development of

KS. Human leukocyte antigen (HLA)-DR + LCs are found in decreased numbers in KS lesions [1, 39]. Remarkably, the Hp 2-1 phenotype was a stronger preventive factor to KS development compared to the Hp 2-2 phenotype. This may be due to the higher serum iron concentration, higher transferrin saturation and higher ferritin values reported in Hp 2-2 males, but not in females, than in Hp 1-1 and Hp

2-1 subjects [7]. Multiple studies have described the role of cellular iron in the carcinogenic pathways [15, 20, 29, 34, 40]. However, the pathogenic role of iron in cancer development and/or progression is not fully understood [32]. In comparison with a healthy population, ferritin concentrations are higher in HIV-infected patients [5]. More specifically in cases of KS, the addition of iron salts to KS-derived cells can stimulate their growth [30]. In human dermal microvascular endothelial cells, iron can induce antiapoptosis, which may alter the homeostasis of microvessels and promote neo-angiogenesis [31]. At the early stages of KS development, red blood cell extravasation and the presence of siderophages are encountered, providing a possible continuous source of iron for endothelial and KS cells. Moreover, on histopathological examination, ferritin granules and hemosiderin-loaded macrophages are regarded as the hallmark of KS [32]. Hp 2-2 subjects accumulate more iron and oxidize more vitamin C, suggesting that less efficient protection against hemoglobin/iron-driven oxidative stress may be a direct mechanism for stimulating viral replication [5].

Hp is a multifunctional protein exerting both immunomodulating and iron-scavenging properties. On the one hand, Hp 1-1 has the least inflammatory properties, but on the other hand, the Hp 2-2 phenotype is associated with higher iron levels. Moreover, the three phenotypes have a markedly different molecular structure. As the development of Kaposi sarcoma has also a multifactorial pathophysiology, this may explain why the Hp phenotypes show no gradual change. Previous research has shown that the Hp 2-1 phenotype is not always associated with an intermediate risk, which is nicely discussed in a letter regarding the finding of an increased risk of the Hp 2-1 phenotype for rapid growth of abdominal aortic aneurysms [41]. A similar finding was observed in patients with Legionnaires' disease. Using a multivariate analysis, Hp 2-1 was an independent risk factor for the more severe cases. This association was not influenced by age and could not be explained by differences in serum ferritin, iron and transferrin saturation [19]. The phenomenon of an enhanced or decreased risk for heterozygous individuals has been termed molecular heterosis [3].

Recently, several studies stressed the protective effect of functional promoter single nucleotide polymorphisms (SNPs) of the Hp gene to the susceptibility to infections. The base substitutions -61A-C (rs5471) and -101C-G (rs5470) within the 5' flanking region of the Hp gene are, respectively, associated with ahaptoglobinaemia and hypohaptoglobinaemia [35]. In two independent cohort studies of Gambian children, Hp gene promoter polymorphisms affected susceptibility to malaria (*Plasmodium falciparum*) [4] and trachoma (*Chlamydia trachomatis*) [28]. As the -61A-C and the -101C-G Hp promoter SNPs are highly linked with the Hp2 and Hp1S allele, respectively, those

SNPs may also be involved in the development of Kaposi's sarcoma in HIV patients. Further investigations should focus on the possible associations between Hp promoter SNPs and HHV-8 infection. Besides the Hp phenotypes, polymorphisms in other genes involved in the host immune response have previously been investigated. In HIV patients, the VF genotype of Fc γ R11A was associated with KS risk, while the FF genotype was characterized as a protective factor [18]. HLA-DRB1 alleles with phenylalanine at position 13 instead of glycine had an increased risk for KS [10]. Studies of classic KS identified diplotypes of the interleukin-8 receptor- β (IL8R β , 1235T/1010G) and IL-13 (promoter region variant 98A) as predisposing factors. Furthermore, polymorphisms in HLA have been linked to KS and HHV-8 shedding [2, 12]. Our findings pointing to an association of the Hp 1-1 phenotype with an increased risk for KS add additional evidence to the role of genes involved in the immune response in the pathogenesis of KS.

In conclusion, we found a significant difference in development of KS in HIV patients, according to their Hp phenotypes. If confirmed, HIV patients with the Hp 1-1 phenotype are at increased risk for KS development which may be due to a decreased skin immunosurveillance associated with this phenotype.

References

1. Bedani PL, Risichella IS, Strumia R, Cavazzini L, Gilli P, Calstrini C, Verzola A, Stabellini G (1999) Kaposi's sarcoma in renal transplant recipients: pathogenetic relation between the reduced density of Langerhans cells and cyclosporine-A therapy. *J Nephrol* 12:193–196
2. Brown EE, Fallin D, Ruczinski I, Hutchinson A, Staats B, Vitale F, Lauria C, Serraino D, Rezza G, Mbisa G, Whithby D, Messina A, Goedert JJ, Chanock SJ (2006) Associations of classic Kaposi sarcoma with common variants in genes that modulate host immunity. *Cancer Epidemiol Biomarkers Prev* 15:926–934
3. Comings DE, MacMurray JP (2000) Molecular heterosis: a review. *Mol Genet Metab* 71:19–31
4. Cox SE, Doherty C, Atkinson SH, Nweneka CV, Fulford AJ, Ghattas H, Rockett KA, Kwiatkowski DP, Prentice AM (2007) Haplotype association between haptoglobin (Hp2) and Hp promoter SNP (A-61C) may explain previous controversy of haptoglobin and malaria protection. *PLoS One* 2:e362
5. Delanghe JR, Langlois MR, Boelaert JR, Van Acker J, Van Wanzeele F, van der Groen G, Hemmer R, Verhofstede C, De Buyzere M, De Bacquer D, Arendt V, Plum J (1998) Haptoglobin polymorphism, iron metabolism and mortality in HIV infection. *AIDS* 12:1027–1032
6. Delanghe J, Allcock K, Langlois M, Claeys L, De Buyzere M (2000) Fast determination of haptoglobin phenotype and calculation of hemoglobin binding capacity using high pressure gel permeation chromatography. *Clin Chim Acta* 291:43–51
7. Delanghe JR, Langlois MR (2002) Haptoglobin polymorphism and body iron stores. *Clin Chem Lab Med* 40:212–216
8. Fink P, Roemer M, Haeckel R, Fateh Mogadam A, Delanghe J, Gressner A, Dubs RW (1989) Measurement of proteins with the

- Behring Nephelometer. A multicenter evaluation. *J Clin Chem Clin Biochem* 27:261–276
9. Gallo RC (1998) The enigmas of Kaposi's sarcoma. *Science* 282:1837–1839
 10. Gayá A, Esteve A, Casabona J, McCarthy JJ, Martorell J, Schulz TF, Whitby D, EURO-SHAKS working group (2004) Amino acid residue at position 13 in HLA-DR beta chain plays a critical role in the development of Kaposi's sarcoma in AIDS patients. *AIDS* 18:199–204
 11. Gazouli M, Zavos G, Papaconstantinou I, Lukas JC, Zografidis A, Boletis J, Kostakis A (2004) The interleukin-6-174 promoter polymorphism is associated with a risk of development of Kaposi's sarcoma in transplant recipients. *Anticancer Res* 24:1311–1314
 12. Geuch-Ongey M, Verboom M, Pfeiffer RM, Schulz TF, Ndugwa CM, Owor AM, Bakaki PM, Bhatia K, Figueiredo C, Eiz-Vesper B, Blasczyk R, Mbulaiteye SM (2010) HLA polymorphisms and detection of Kaposi sarcoma-associated herpesvirus DNA in saliva and peripheral blood among children and their mothers in the Uganda sickle cell anemia KSHV Study. *Infect Agent Cancer* 5:21
 13. Hensler HR, Rappocciolo G, Rinaldo CR, Jenins FJ (2009) Cytokine production by human herpesvirus 8-infected dendritic cells. *J Gen Virol* 90:79–83
 14. Kasvosve I, Speeckaert MM, Speeckaert R, Masukume G, Delanghe JR (2010) Haptoglobin polymorphism and infection. *Adv Clin Chem* 50:23–46
 15. Knekt P, Reunanen A, Takkunen H, Aromaa A, Heliovaara M, Hakulinen T (1994) Body iron stores and risk of cancer. *Int J Cancer* 56:379–382
 16. Lagos D, Boshoff C (2007) Immunobiology and host response to KSHV infection. In: Arvin A, Campadelli-Fiume G, Macarski E, Moore PS, Roizman B, Whitley R, Yamanishi K (eds) *Human herpesviruses: biology, therapy, and immunoprophylaxis*, chap 52. Cambridge University Press, Cambridge
 17. Langlois MR, Delanghe JR (1996) Biological and clinical significance of haptoglobin polymorphism in humans. *Clin Chem* 42:1589–1600
 18. Lehrnbecher TL, Foster CB, Zhu S, Venzon D, Steinberg SM, Wyvill K, Metcalf JA, Cohen SS, Kovacs J, Yarchoan R, Blauvelt A, Chanock SJ (2000) Variant genotypes of FcγRIIIA influence the development of Kaposi's sarcoma in HIV-infected men. *Blood* 95:2386–2390
 19. Lettinga KD, Delanghe JR, Prins J, Langlois MR, Speelman P, Reitsma P, Verbon A (2002) Iron status does not influence susceptibility and outcome in patients with Legionnaires' disease (LD). Abstract ICAAC 2002 B-698
 20. Ludin PM (1961) The carcinogenic action of complex iron preparations. *Br J Cancer* 15:838–847
 21. Mahmud SM, Koushik A, Duarte-Franco E, Costa J, Fontes G, Bicho M, Coutlée F, Franco EL, Biomarkers of Cervical Cancer Risk Study Team (2007) Haptoglobin phenotype and risk of cervical neoplasia: a case-control study. *Clin Chim Acta* 385:67–72
 22. Maurer T, Ponte M, Leslie K (2007) HIV-associated Kaposi's Sarcoma with a high CD4 count and a low viral load. *New Engl J Med* 357:1352–1353
 23. Mesri EA, Cesarman E, Boshoff C (2010) Kaposi's sarcoma and its associated herpesvirus. *Nat Rev Cancer* 10:707–719
 24. Nguyen HQ, Magaret AS, Kitahata MM, Van Rompaey SE, Wald A, Casper C (2008) Persistent Kaposi sarcoma in the era of highly active antiretroviral therapy: characterizing the predictors of clinical response. *AIDS* 22:937–945
 25. No authors listed (1987) Revision of the CDC surveillance case definition for acquired immunodeficiency syndrome. Council of State and Territorial Epidemiologists; AIDS Program, Center for Infectious Diseases. *MMWR Morb Mortal Wkly Rep* 36:1S–15S
 26. No authors listed (1992) 1993 revised classification system for HIV infection and expanded surveillance case definition for AIDS among adolescents and adults. *MMWR Recomm Rep* 41:1–19
 27. Palucka K, Banchereau J (2002) How dendritic cells and microbes interact to elicit or subvert protective immune responses. *Curr Opin Immunol* 14:420–431
 28. Savy M, Hennig BJ, Doherty CP, Fulford AJ, Bailey R, Holland MJ, Sirugo G, Rockett KA, Kwiatkowski DP, Prentice AM, Cox SE (2010) Haptoglobin and sickle cell polymorphisms and risk of active trachoma in Gambian children. *PLoS One* 5:e11075
 29. Sherson D, Svane O, Lyng E (1991) Cancer incidence among foundry workers in Denmark. *Arch Environ Health* 46:75–81
 30. Simonart T, Noel JC, Andrei G, Parent D, Van Vooren JP, Hermans P, Lunardi-Yskandar Y, Lambert C, Dieye T, Farber CM, Liesnard C, Snoeck R, Heenen M, Boelaert JR (1998) Iron as a potential cofactor in the pathogenesis of Kaposi's sarcoma. *Int J Cancer* 78:720–726
 31. Simonart T, Degraef C, Stordeur P, Noel JC, Mosselmans R, Van Vooren JP, Parent D, Boelaert JR, Heenen M, Galand P (2001) Iron induces bcl-2 expression in human dermal microvascular endothelial cells. *Free Rad Res* 34:221–235
 32. Simonart T (2004) Iron: a target for the management of Kaposi's sarcoma? *BMC Cancer* 15:1
 33. Speeckaert R, Brochez L, Lambert J, van Geel N, Speeckaert MM, Claeys L, Langlois M, Van Laer C, Peeters P, Delanghe JR (2011) The haptoglobin phenotype influences the risk of cutaneous squamous cell carcinoma in kidney transplant patients. *J Eur Acad Dermatol Venereol*. doi:10.1111/j.1468-3083.2011.04112.x. (Epub ahead of print)
 34. Stevens RG, Graubard BI, Micozzi MS, Neriishi K, Blumberg BS (1994) Moderate elevation of body iron level and increased risk of cancer occurrence and death. *Int J Cancer* 56:364–369
 35. Teye K, Quaye IK, Koda Y, Soejima M, Tsuneoka M, Pang H, Ekem I, Amoah AG, Adjei A, Kimura H (2003) A-61C and C-101G Hp gene promoter polymorphisms are, respectively, associated with ahaptoglobinaemia and hypohaptoglobinaemia in Ghana. *Clin Genet* 64:439–443
 36. Tuttleton AS, Jennings L, Nindl I, Rosl F, Bouwes Bavinck JN, Seçkin D, Trakatelli M, Murphy GM; for the Viral Working Group of the International Transplant Skin Cancer Collaborative (ITSCC) and Skin Care in Organ Transplant Patients, Europe (SCOPE) (2011) Viral oncogenesis and its role in nonmelanoma skin cancer. *Br J Dermatol* (Epub ahead of print)
 37. Uldrick TS, Wang V, O'Mahony D, Aleman K, Wyvill KM, Marshall V, Steinberg SM, Pittaluga S, Maric I, Whitby D, Tosato G, Little RF, Yarchoan R (2010) An interleukin-6-related systemic inflammatory syndrome in patients co-infected with Kaposi sarcoma-associated herpesvirus and HIV but without multicentric Castlemans disease. *Clin Infect Dis* 62:350–358
 38. Uldrick TS, Whitby D (2011) Update on KSHV epidemiology, Kaposi Sarcoma pathogenesis, and treatment of Kaposi Sarcoma. *Cancer Lett* (Epub ahead of print)
 39. Valcuende-Cavero F, Febrer-Bosch MI, Castells-Rodellas A (1994) Langerhans' cells and lymphocytic infiltrate in AIDS-associated Kaposi's sarcoma. An immunohistochemical study. *Acta Derm Venereol* 74:183–187
 40. Weinberg ED (1996) The role of iron in cancer. *Eur J Cancer Prev* 5:19–36
 41. Wiernicki I, Safranow K, Baranowska-Bosiacka I, Piatek J, Gutowski P (2010) Haptoglobin 2–1 phenotype predicts rapid growth of abdominal aortic aneurysms. *J Vasc Surg* 52:691–696
 42. de Wit R, Raasveld MH, ten Berge RJ, van der Wouw PA, Bakker PJ, Veenhof CH (1991) Interleukin-6 concentrations in the serum of patients with AIDS-associated Kaposi's sarcoma

- during treatment with interferon-alpha. *J Intern Med* 229:539–542
43. Xie LX, Li Y, Zhang Q, Stiller MJ, Wang CL, Streilein JW (2000) Haptoglobin is a natural regulator of Langerhans cell function in the skin. *J Dermatol Sci* 24:25–37
44. Xie J, Pan H, Yoo S, Gao SJ (2005) Kaposi's sarcoma-associated herpesvirus induction of AP-1 and interleukin 6 during primary infection mediated by multiple mitogen-activated protein kinase pathways. *J Virol* 79:15027–15037