REVIEW



# **ALS and FTD: an epigenetic perspective**

**Veronique V. Belzil1 · Rebecca B. Katzman1 · Leonard Petrucelli1**

Received: 15 March 2016 / Revised: 17 May 2016 / Accepted: 2 June 2016 / Published online: 9 June 2016 © Springer-Verlag Berlin Heidelberg 2016

**Abstract** Amyotrophic lateral sclerosis (ALS) and frontotemporal dementia (FTD) are two fatal neurodegenerative diseases seen in comorbidity in up to 50 % of cases. Despite tremendous efforts over the last two decades, no biomarkers or effective therapeutics have been identified to prevent, decelerate, or stop neuronal death in patients. While the identification of multiple mutations in more than two dozen genes elucidated the involvement of several mechanisms in the pathogenesis of both diseases, identifying the hexanucleotide repeat expansion in *C9orf72,* the most common genetic abnormality in ALS and FTD, opened the door to the discovery of several novel pathogenic biological routes, including chromatin remodeling and transcriptome alteration. Epigenetic processes regulate DNA replication and repair, RNA transcription, and chromatin conformation, which in turn further dictate transcriptional regulation and protein translation. Transcriptional and post-transcriptional epigenetic regulation is mediated by enzymes and chromatin-modifying complexes that control DNA methylation, histone modifications, and RNA editing. While the alteration of DNA methylation and histone modification has recently been reported in ALS and FTD, the assessment of epigenetic involvement in both diseases is still at an early stage, and the involvement of multiple epigenetic players still needs to be evaluated. As the epigenome serves as a way to alter genetic information not only during aging, but also following environmental

V. V. Belzil and R. B. Katzman contributed equally to this work.

 $\boxtimes$  Leonard Petrucelli petrucelli.leonard@mayo.edu signals, epigenetic mechanisms might play a central role in initiating ALS and FTD, especially for sporadic cases. Here, we provide a review of what is currently known about altered epigenetic processes in both ALS and FTD and discuss potential therapeutic strategies targeting epigenetic mechanisms. As approximately 85 % of ALS and FTD cases are still genetically unexplained, epigenetic therapeutics explored for other diseases might represent a profitable direction for the field.

**Keywords** Amyotrophic lateral sclerosis · Chromatin remodeling · Epigenetic processes · Frontotemporal dementia · RNA-mediated regulation · Transcription regulation

## **Introduction**

Degeneration of motor neurons results in progressive loss of motor skills, a condition first described by Charcot and Joffroy in 1869 [\[29](#page-11-0)] and commonly known as Lou Gherig's disease in honor of the baseball Hall of Famer who gave a memorable farewell speech in 1939. Also referred to as amyotrophic lateral sclerosis (ALS), the disease is characterized by degeneration of upper motor neurons of the motor cortex and corticospinal tract, and lower motor neurons of the brain stem and spinal cord, progressively causing muscle weakness, spasticity, atrophy, and finally lethal respiratory failure within 2–5 years of disease onset [\[20](#page-10-0)]. Only 5–8 % of overall ALS cases report a family history of the disease, where there is much heterogeneity in clinical presentation across affected relatives [[2,](#page-10-1) [24](#page-10-2), [25,](#page-11-1) [63](#page-11-2)]. The most common alternative deficit observed in ALS family members is cognitive impairment, which is also comorbid to ALS in about 50 % of patients. In fact, while deficits in

<sup>1</sup> Department of Research, Neuroscience, Mayo Clinic College of Medicine, 4500 San Pablo Road, Jacksonville, FL 32224, USA

executive functioning, visual and immediate verbal memory, language and fluency, as well as psychomotor speed are commonly present in ALS patients, these symptoms are mostly not acute enough to receive a diagnosis of dementia [\[144](#page-14-0)]. As such, only 15–20  $%$  of ALS patients receive concomitant diagnosis of frontotemporal dementia (FTD) [[23,](#page-10-3) [65](#page-12-0), [67](#page-12-1), [106](#page-13-0), [153](#page-14-1)]. While ALS is the most common motor neuron disease, FTD is the second-most common cause of early onset dementia after Alzheimer's Disease (AD) [\[120](#page-13-1)], and comprises three clinically distinct syndromes: behavioral variant FTD, progressive nonfluent aphasia, and semantic dementia [[136,](#page-13-2) [163](#page-14-2)]. Briefly, FTD is characterized by neuronal degeneration in the frontal and temporal lobes causing progressive deterioration of language, personality, and behavior  $[68]$  $[68]$ . Up to 50 % of FTD cases report a positive family history, with members affected with either FTD or other neurodegenerative diseases [\[68](#page-12-2)].

The link between ALS and FTD was strengthened after the finding that a hexanucleotide repeat expansion (HRE) in the *C9orf72* gene explains disease in multiple family pedigrees counting members diagnosed with either one or both diseases [\[45](#page-11-3), [150\]](#page-14-3). Multiple genetic assessments now predict this HRE to be carried by approximately 34 % of familial and 6 % of sporadic ALS cases, as well as 26 % of familial and 5 % of sporadic FTD patients [\[145](#page-14-4), [177](#page-14-5)]. Although higher and lower frequencies have been reported depending on the population studied [[107\]](#page-13-3), the HRE in *C9orf72* is considered the most common genetic cause of ALS and FTD identified thus far [\[8](#page-10-4), [149](#page-14-6)].

Causative genetic mutations identified in more than two dozen genes currently explain  $~68\%$  of familial and  $~11\%$ of sporadic ALS cases [\[149](#page-14-6)], leaving about 86 % of overall cases, mostly sporadic, unexplained. Similarly, genetic mutations explain about 25 % of familial and 10 % of sporadic FTD, leaving about 83 % of the overall FTD cases genetically unexplained [[77\]](#page-12-3). The fact that genes associated with familial ALS remain typically unaltered in sporadic ALS (sALS) patients, along with the fact that genome-wide association studies have identified variants with only moderate risk, points to the likelihood of other disease culprits [\[34](#page-11-4), [40,](#page-11-5) [99,](#page-12-4) [149](#page-14-6), [179](#page-14-7)]. Specifically, increasing evidence supports altered RNA processing as a central pathological mechanism in ALS [\[71](#page-12-5), [140](#page-13-4), [142](#page-13-5)]. Epigenetic processes are known to regulate RNA transcription, which can in turn dictate protein translation or further regulate downstream transcription [[127\]](#page-13-6). Because of the discovery of an HRE in *C9orf72*, numerous studies using blood, brain tissues, and induced pluripotent stem cells from *C9orf72* repeat expansion carriers demonstrated the involvement of epigenetic and transcriptional dysfunction in ALS and FTD [\[14](#page-10-5), [15,](#page-10-6) [50](#page-11-6), [142,](#page-13-5) [187,](#page-15-0) [190,](#page-15-1) [191\]](#page-15-2). How epigenetic and transcriptomic mechanisms interact with one another, and whether these interactions can be exploited as potential therapeutic targets for ALS and FTD remain unanswered questions. Here, we provide a review of what is currently known about the involvement of altered epigenetic processes in these two devastating diseases and discuss potential strategies for targeting these alterations therapeutically.

#### **Epigenetic regulatory mechanisms**

Since Crick's 1958 central dogma suggested a flow from genetic information to RNA transcription and protein translation [[39\]](#page-11-7), much effort has been devoted to better understand RNA regulation and its role in human diseases. It has been recognized for decades that the genetic material is under epigenetic control through modification of the DNA and chromatin-associated proteins dictating RNA transcription, the template for protein synthesis, as well as regulating DNA replication and repair. The ability of the RNA to also act as an intermediate in gene regulation was only contemplated in 1969 [\[22](#page-10-7)], a proposition refined in 2001 suggesting a direct role for regulatory RNA networks to control epigenetic processes [\[111](#page-13-7)]. It is now recognized that RNA not only functions as a messenger between DNA and protein, but also regulates the organization of the genome as well as gene expression [\[127](#page-13-6)]. Regulatory RNAs play central roles in transcriptional and post-transcriptional epigenetic processes, while their own expression is also under epigenetic control [[113,](#page-13-8) [127](#page-13-6)]. While most epigenetic changes responsible for developmental processes result from anticipated internal processes rooted within the genome, communication between the environment and the genome is reflected through RNA editing, and these changes can further be transmitted from cell to cell to enhance proper physiological adaptation during development [\[112](#page-13-9)]. Specifically, RNA editing serves as a way to alter genetic information following environmental signals, especially in the brain, highlighting a dynamic RNA-mediated interaction between the environment, the epigenome and the transcriptome [[113\]](#page-13-8). Over the last decade, there has been emergent interest to better understand the interaction between the epigenome and the transcriptome, especially in the contexts of cancer and neurodegenerative diseases. A general overview of the major epigenetic processes controlling transcription and chromatin conformation is provided in this section (Fig. [1\)](#page-2-0).

## **Modification of DNA and chromatin‑associated proteins**

During the last few decades, the role of transcriptional and post-transcriptional epigenetic modifications in gene activation and repression has been intensely studied, especially in the cancer field, and neuroscientists are increasingly



<span id="page-2-0"></span>**Fig. 1** Major epigenetic regulatory mechanisms. Schematic representation of the major cellular epigenetic mechanisms: histone modification, DNA methylation, and RNA-mediated regulation. It is suggested that interplay between these mechanisms exists in cells, as depicted by the *arrows* from one pie slice to the next. For each epigenetic mechanism, one example of aberrant regulation demonstrated in ALS and/or FTD is provided (font in *red*)

interested in assessing its role in neurodegeneration. Epigenetic modifications constitute codes that regulate chromatin organization as well as DNA transcription and repair, balancing stability and reversibility of the genetic material to maintain cell identity and/or enable appropriate cellular responses to internal and external stimuli. Epigenetic responses are initiated by chemical modifications of eukaryotic DNA and histone octamers around which the DNA is wrapped. Each histone octamer is composed of four different pairs of histones (H2A, H2B, H3, and H4), and each have N-terminal tails that can be post-transcriptionally modified. DNA cytosine residues, especially at CpG dinucleotides, can also be chemically adjusted, mostly in CpG-rich regions commonly referred to as CpG islands. CpG methylation has been known for a long time to act as an epigenetic repressive modulator in vertebrates [\[167](#page-14-8), [180–](#page-14-9)[182\]](#page-14-10), and only recent evidence demonstrated that DNA methylation is actually highly interconnected to post-transcriptional changes at histone lysine residues, with each system mechanistically relying on the other for normal regulation of chromatin conformation [[155\]](#page-14-11). As such, histone modification can direct DNA methylation patterns, and DNA methylation can serve as a template for histone modifications.

DNA methylation and histone modification pathways can act independently or be mutually dependent on one another through mediation of different biochemical interactions. Such interactions are mediated by a limited number of enzymes and chromatin-modifying complexes that broadly influence the transcription of the genome, with only a few of these having affinity for specific DNA sequences. These enzymes and chromatin-modifying complexes facilitate transcriptional regulation by acting as 'writers' or 'erasers', adding or removing chemicals to DNA and histone proteins [\[105](#page-13-10)]. Specifically, epigenetic writers lay down epigenetic marks on DNA or histones by either covalently modifying the amino-terminal tails of histone proteins or by altering the DNA itself. The covalent modification generated by epigenetic writers can be reversed by epigenetic erasers and recognized by epigenetic readers [[53\]](#page-11-8). DNA methyltransferases (Dnmts), such as Dnmt3a, Dnmt3b, and Dnmt1 are epigenetic writers responsible for establishing and maintaining DNA methylation patterns at different genomic sites [\[31](#page-11-9)]. Other epigenetic writers, such as histone acetyltransferases (HATs), histone methyltransferases (HMTs), protein arginine methyltransferases (PRMTs), and kinases, are responsible for modulating epigenetic histone marks by modifying amino acid residues on histone tails. Epigenetic readers recognize specific epigenetic marks through their protein-containing domains and recruit other chromatin modifiers and remodeling proteins, all together regulating DNA-dependent processes [[53\]](#page-11-8). Proteins containing DNA methyl-binding domains, chromodomains, bromodomains,

and Tudor domains are all considered epigenetic readers. Finally, some enzymes act as erasers after catalyzing the removal of epigenetic marks [[53\]](#page-11-8). Examples of erasers are histone deacetylases (HDACs), lysine demethylases (KDMs), phosphatases, and deubiquitylases [[53\]](#page-11-8).

While histone modification leading to chromatin remodeling is readily reversible, changes in DNA methylation are perceived as stable, long-term changes. Specifically, Dnmt1 is often considered as a 'maintenance' methyltransferase after recognizing hemimethylated CpG sites following semi-conservative DNA replication by reinstating original methylation patterns [[180\]](#page-14-9). On the other hand, Dnmt3a and Dnmt3b are seen as de novo methyltransferases, both catalyzing the methylation of originally unmethylated CpG sites [\[155](#page-14-11), [162](#page-14-12)]. Dnmts thus work in an interconnected fashion, with Dnmt3a and Dnmt3b recognizing and methylating specific genomic regions, and with Dnmt1 faithfully maintaining methylation. Recent reports demonstrated that de novo methylation through Dnmt3a and Dnmt3b action rely partly on pre-existing histone lysine methylation and enzymes catalyzing lysine post-transcriptional modifications [\[155](#page-14-11)]. In contrast, another report suggested that nonmethylated sites in CpG islands influence histone lysine methylation at gene encoding regulatory elements [\[155](#page-14-11)]. There is also an emerging belief that histone lysine methylation protects DNA from active demethylation [\[155](#page-14-11)].

The process by which demethylation can take place was first considered in the early 1970s when Penn et al. suggested that methylated cytosines, or 5-methylcytosines (5mCs), can be oxidized to become 5-hydroxymethylcytosines (5hmC), an observation finally demonstrated in 2009 [\[98](#page-12-6), [134\]](#page-13-11). Growing evidence now shows that 5hmCs, initially thought to represent an intermediate state between methylated DNA and unmethylated DNA, also act as a stable epigenetic marker that might contribute to neurological disease development [[164,](#page-14-13) [166\]](#page-14-14). Of interest, studies using human and mouse tissues have shown that the number of 5hmC sites increases with age, and levels vary across organs [\[30](#page-11-10), [66](#page-12-7), [170](#page-14-15)]. In fact, the highest levels of 5hmC are found in the central nervous system [\[30](#page-11-10), [66](#page-12-7), [170](#page-14-15)]. Enzymes Tet1, Tet2, and Tet3 are Tet family 5mC hydrolases that convert 5mC to 5hmC [\[95](#page-12-8), [98](#page-12-6), [171](#page-14-16)] to potentially restore gene expression. While not essential for 5mC to 5hmC conversion, recent evidence demonstrated that oxidative stress can also trigger this oxidation event [\[94](#page-12-9), [141\]](#page-13-12). Finally, it was recently reported that Tet enzymes can also catalyze the establishment of 5hmC in RNA [[58\]](#page-11-11), which promotes RNA translation [\[46](#page-11-12)].

Enzymes and chromatin-modifying complexes must be purposefully directed to specific genomic positions in different types of cells to elicit DNA and histone tail modification. One important source of guidance for these proteins is RNA species which, as opposed to enzymes and chromatin-modifying complexes, are highly sequence and locus specific. The role of RNAs in mediating epigenetic processes is explained in the next section.

#### **RNA‑mediated epigenetic regulation**

There is now compelling evidence that RNA signaling and editing play a crucial role in chromatin remodeling and nuclear architecture [[5,](#page-10-8) [113](#page-13-8)]. Specifically, coding and noncoding RNAs (ncRNAs) are involved in epigenetic regulation by recruiting chromatin-modifying complexes and Dnmts to particular genomic loci [\[112\]](#page-13-9). ncRNAs, which mostly operate through repressive control while still having the potential to act as gene activators [\[160](#page-14-17), [161\]](#page-14-18), comprise small RNAs (sRNAs) of less than 200 nucleotides and long non-coding RNAs (lncRNAs) of more than 200 nucleotides. sRNAs are further subcategorized as micro-RNAs (miRNAs), short interfering RNAs (siRNAs) and PIWI-associated RNAs (piRNAs), and lncRNAs are classified according to their position and direction of their transcription (e.g. antisense, intergenic, overlapping, intronic, bidirectional, and processed) [\[114](#page-13-13), [137](#page-13-14)]. Several miRNAs originate from introns of protein-coding genes either by canonical Drosha pathway or by splicing (which generates mirtrons) [[17,](#page-10-9) [132](#page-13-15), [156\]](#page-14-19), whereas lncRNAs often overlap with, or are interspersed between, several coding and non-coding transcript variants [[28,](#page-11-13) [90\]](#page-12-10). In this way, lncR-NAs regulate the expression of neighboring protein-coding genes [[6\]](#page-10-10).

Different sRNAs and lncRNAs are classified into groups depending on their genomic origin, but also according to their mechanism of action [[5,](#page-10-8) [137\]](#page-13-14). In fact, sRNAs and lncRNAs modify chromatin structure and silence transcription through distinct but unifying mechanisms. One of these is sRNA-guided gene regulation, which has emerged as a central mechanism that guides Argonaute (AGO) containing complexes to complementary nascent RNA scaffolds [\[118](#page-13-16)]. AGO proteins are highly specialized direct binding partners of miRNAs, siRNAs, and piRNAs, and interact with other proteins to coordinate downstream gene-silencing and RNA splicing [[118\]](#page-13-16). sRNAs also perform as mediators of chromatin structure and transcription repression by facilitating the recruitment of histone and DNA methyltransferases. Besides playing a central role in RNA degradation and translational repression, sRNAs also modulate chromatin and gene expression via RNA interference (RNAi) pathways [[85\]](#page-12-11), which in turn modulate histone or DNA methylation to repress transcription [\[154\]](#page-14-20). sRNAs, such as miRNAs, can also act oppositely: they can stimulate gene expression under stress conditions as a result of new miRNA-ARGO complexes interacting with RNA-binding proteins which relocate during cellular stress [\[102\]](#page-12-12). Transcriptional silencing under sRNA control is also 'memorized' through self-reinforcing epigenetic loops, a process by which sRNAs are fused to histone modification or DNA methylation and form positive feedback systems to maintain epigenetic conditions. These self-reinforcing epigenetic loops are key players in epigenetic inheritance of histone and DNA methylation patterns [[78\]](#page-12-13). Specifically, such association of sRNAs with positive feedback loops in the germ line acts as a fingerprint for internal or environmentally induced alterations, and can be transmitted from parents to offspring [\[60,](#page-11-14) [146\]](#page-14-21).

On the other hand, several lncRNAs and some messenger RNAs (mRNAs) mediate the recruitment of chromatinmodifying complexes independently of sRNAs and RNAi pathways [[154\]](#page-14-20). Specifically, lncRNAs can recruit chromatin-modifying enzymes to specific loci to activate or silence gene-specific transcription [[19,](#page-10-11) [78,](#page-12-13) [154](#page-14-20)]. One example is exonized Alu elements, the most common transposable elements in humans, which are mostly located in lncRNAs and untranslated regions of mRNA [[93\]](#page-12-14). Alu RNAs act as transacting transcriptional repressors after binding RNA polymerase II [[109\]](#page-13-17), and are also involved in the regulation of alternative splicing, modulation of translation, and monitoring of mRNA stability [[74\]](#page-12-15). In addition, several studies attempted to establish whether lncRNAs recruit Polycomb proteins, which are epigenetic regulators of transcription, but the data remain inconclusive [[78\]](#page-12-13). Current evidence does support a role for lncRNAs transcribed from enhancers in transcription regulation. These particular enhancer sequences are different from enhancer sequences that bind transcription factors, as they activate specific target genes [\[101](#page-12-16)]. However, as the exact function of lncRNAs in genome regulation is still largely unknown, much will likely be learned in the near future.

Transcriptional silencing mediated by sRNAs and lncR-NAs is an efficient RNA surveillance system responsible for the detection and silencing of aberrant transcript variants. sRNAs and lncRNAs, while having distinct functions, act together to regulate gene transcription. A good example is the piRNA-induced silencing complex, which protects the integrity of the genome by silencing transposable elements that can in turn act as transcriptional repressors when expressed [\[165](#page-14-22)]. Along with enzymes such as DNA and histone methyltransferases or acetylases/deacetylases, and repressive and permissive chromatin-modifying complexes such as Polycomb and Trithorax groups, RNA-directed processes assist in orchestrating chromatin architecture, gene transcription, and epigenetic memory [\[6](#page-10-10), [18](#page-10-12)]. Moreover, RNA-mediated regulation has the highest affinity for specific DNA sequences, and can thus be an interesting therapeutic target for diseases such as ALS and FTD.

Finally, another way to regulate epigenetic processes is through RNA editing. Environmental information can be transmitted and reflected on the hardwired genetic

information after post-transcriptional editing of the RNA base sequence, which in turn affects the regulation of downstream targeted RNAs. Two classes of enzymes are responsible for editing the RNA: the adenosine deaminases acting on RNA (ADARs), which catalyzes adenosine deamination to inosine [[26\]](#page-11-15), and the apolipoprotein B mRNA editing enzymes (APOBECs), which catalyze cytidine deamination to uracil [\[129](#page-13-18)]. While RNA editing has been shown to take place in most tissues, it is particularly abundant and important in the brain [\[13](#page-10-13)], and has been shown in some cases to alter the amino acid sequence and splicing patterns of neurotransmitter receptors, thereby altering the electrophysiological properties of synapses [[53\]](#page-11-8). Consequently, these RNA alterations can be transmitted across cells and offspring, a phenomenon that was first demonstrated in plants prior to animals [[48,](#page-11-16) [75,](#page-12-17) [185\]](#page-15-3).

RNA editing highlights the dynamic interplay between the environment, the epigenome, and the transcriptome. These post-transcriptional changes in RNA lead to alteration of epigenetic information, which is memorized, and then transmitted between cells, across organ systems, and through different generations. Future studies will have to determine how extensively RNA editing may modulate epigenetic processes and assess how plastic the epigenome may be. This is particularly important for germline and de novo RNA editing occurring in the brain, considering the possibility that edited RNAs may eventually cause disease, including neurodegeneration.

#### **The role of epigenetic regulation in ALS and FTD**

Very few epigenetic studies related to ALS were published before the identification of the *C9orf72* HRE in September 2011 [[45](#page-11-3), [150](#page-14-3)]. As alteration of epigenetic processes has been observed in a number of repeat expansion disorders [[16\]](#page-10-14), the finding of a HRE in *C9orf72* unveiled the possibility that epigenetic modifications and chromatin remodeling might also play a role in ALS and FTD. Since the HRE discovery, more than twenty studies have been reported, half of which are *C9orf72* locusrelated. This sudden increase in ALS/FTD epigenetic reports reflects the enthusiasm of the field to explore new territories in terms of disease mechanism, as the young field of epigenomics might provide novel explanations for these two lethal diseases. The assessment of epigenetic modifications for ALS and FTD is still at an early stage and much remains to be evaluated. For example, it is not known whether unique epigenetic changes contributing to ALS and FTD can be found in all cells or a subgroup of cells more susceptible or vulnerable to specific epigenetic changes. However, coupled with evidence that modifications of the epigenome contribute to ALS and FTD

pathogenesis is the dynamic nature of epigenetic writer, eraser, and reader enzymes. Thus, developing therapeutic strategies that target enzymes regulating epigenetic dynamics to reverse epigenetic changes that lead to neurodegeneration offers an attractive approach to combat these diseases.

The following sections provide an overview of what we currently know about epigenetic modifications in ALS and FTD (Table [1\)](#page-6-0), including therapeutic implications of these findings, environmental factors that may lead to epigenetic changes, and avenues that should be explored in future studies.

#### **DNA methylation in ALS and FTD**

While earlier studies found the promoters of some genes implicated in the pathogenesis of ALS (*SOD1*, *VEGF*, and *GLT1*) largely unmethylated in ALS patients [[131,](#page-13-19) [192](#page-15-4)], several studies indicate that DNA methylation plays a role in neurodegeneration pathophysiology. DNA methylation was reported altered in ALS post-mortem brains after methylation levels were compared between sALS and control subjects using Affymetrix GeneChip Human Tiling 2.0R Arrays [[124\]](#page-13-20). The authors reported 38 differentially methylated regions (DMRs), and their pathway analysis suggested that the genes with DMRs were involved in calcium homeostasis, neurotransmission and oxidative stress. Another study designed to identify epigenetic modifications associated with sporadic ALS reported global changes in both 5mC and 5hmC levels in postmortem spinal cords but not in blood samples [[54\]](#page-11-17). The authors observed hyper- or hypomethylation with corresponding under- or overexpression of 112 genes highly associated with immune and inflammation responses. Furthermore, the whole blood was analyzed to determine whether DNA methylation is a modifier of ALS age of onset [\[174](#page-14-23)]. Here, it was found that DNA methylation may be a marker of epigenetic dysfunction in ALS, as levels of methylation are increased independently from age of onset. Moreover, two studies analyzing the progranulin-encoding gene (*GRN*) reported that *GRN* promoter methylation regulates progranulin expression [\[11](#page-10-15), [59](#page-11-18)]. Both reports found increased *GRN* promoter methylation in FTD subjects negatively correlating with *GRN* mRNA levels [\[11](#page-10-15), [59](#page-11-18)], an interesting finding considering that *GRN* haploinsufficiency is a major cause of FTD [[56\]](#page-11-19). A recent study analyzing genome-wide DNA methylation patterns in the peripheral blood of tau-related progressive supranuclear palsy (PSP) and FTD subjects relative to unaffected controls identified a specific methylation signature associated pathologically with tauopathy, indicating the signature serves as a risk factor for neurodegeneration [[103\]](#page-13-21).

The most extensively studied epigenetic change in ALS and FTD thus far is the methylation of *C9orf72*; different



<span id="page-6-0"></span>Table 1 Epigenetic Modifications in ALS and FTD **Table 1** Epigenetic Modifications in ALS and FTD

studies aimed to determine whether this modification may play a role in mechanisms possibly leading to C9ORF72 loss of function. The loss of function theory has been supported by several reports demonstrating decreased expression of one or multiple *C9orf72* transcript variants in frontal cortex, motor cortex, cerebellum, and cervical spinal cord of FTD or ALS *C9orf72* HRE carriers (c9FTD/ALS) [\[15](#page-10-6), [36,](#page-11-25) [50,](#page-11-6) [57](#page-11-26), [64](#page-12-22), [126,](#page-13-26) [178\]](#page-14-25), as well as in lymphoblastoid cell lines generated from c9FTD/ALS patient blood [[36\]](#page-11-25) and neuronal cell lines differentiated from c9FTD/ALSinduced pluripotent stem cells (iPSCs) [[4,](#page-10-18) [50](#page-11-6)]. Hypermethylation of the 5′ CpG island located in the *C9orf72* promoter region was shown by different groups to be present in about 10–30 % of c9FTD/ALS subjects [\[14](#page-10-5), [104](#page-13-22), [187,](#page-15-0) [191](#page-15-2)], possibly leading to reduced *C9orf72* expression levels. While the cause of reduced *C9orf72* expression in the remaining 70 % of c9FTD/ALS cases not hypermethylated was still obscure, a subsequent study demonstrated that the repeat expansion itself is methylated in all HRE carriers [\[190](#page-15-1)], suggesting that the methylation of the HRE region might be the cause of the reduced expression for most, if not all, c9FTD/ALS patients. The consequence of *C9orf72* downregulation has been evaluated in several model systems, including by knocking down the *C9orf72* orthologue in zebrafish. A knockdown of the zebrafish orthologue led to both altered morphology of motor neuron axons and locomotor deficits, a phenotype rescued by overexpression of human *C9orf72* [[36\]](#page-11-25). Depleting the *C9orf72* orthologue in the nematode also led to motor neuron deficits [\[172](#page-14-26)]. Of note, the fact that the *C9orf72* mouse orthologue is enriched in brain regions susceptible to degenerate in ALS and FTD [[168\]](#page-14-27) suggests that sufficient *C9orf72* expression is critical to neuronal survival. However, since conditional deletion from mouse neuronal and glial cells was not associated with neurodegeneration [[96\]](#page-12-23), reduced *C9orf72* levels alone may not be sufficient to trigger motor neuron degeneration in higher organisms.

One report suggested that the length of the repeat might influence the level of DNA methylation at the *C9orf72* promoter. Specifically, this process was demonstrated in a Canadian family with a father carrying an intermediate length allele (70 repeats) with an unmethylated *C9orf72* promoter, which expanded to approximately 1750 repeats at the time of transmission to four of his children [\[188](#page-15-7)]. The expanded allele carried by the four children, two of whom have developed ALS symptoms thus far, was characterized by *C9orf72* promoter hypermethylation and associated with reduced *C9orf72* expression [[188\]](#page-15-7). The timing associated with hypermethylation of the *C9orf72* promoter and the reason why it happens in only ~30 % of c9FTD/ALS cases is still under investigation. However, in an attempt to explore this phenomenon, one group used iPSCs generated from a hypermethylated c9ALS patient and observed that 5mC levels at the *C9orf72* promoter were reduced during reprogramming but restored upon neuronal differentiation. On the contrary, 5hmC levels in the same region were increased during the reprogramming process, and levels were even higher after neuronal differentiation. The abundance of 5hmC at the *C9orf72* promoter was also confirmed in the brain of hypermethylated c9FTD patients [[52\]](#page-11-27). Several groups attempted to assess whether DNA methylation can be a clinical modifier of disease. So far, very few significant correlations have been identified. Among these, however, the hypermethylation of the CpG island upstream of the HRE has been shown to correlate with shorter disease duration [\[191\]](#page-15-2), which may serve as a prognostic tool for *C9orf72*-associated disease.

The role that methylation at the *C9orf72* locus plays in ALS and FTD pathogenesis is yet to be fully determined, as contradictory results have arisen. For instance, while epigenetic modification of *C9orf72* through hypermethylation has been significantly correlated with shorter disease duration, it has also been shown to be neuroprotective in patients [[15,](#page-10-6) [104,](#page-13-22) [116,](#page-13-27) [157,](#page-14-28) [190](#page-15-1), [191](#page-15-2)]. Specifically, it was shown that methylation of the HRE reduces the pathogenic effects of the HRE, as determined by the quantification of dipeptide poly (GP) levels, one product of non-traditional repeat associated non-ATG (RAN) translation of the HRE, and of RNA foci formation when the HRE was expressed in cells [[12\]](#page-10-19). As such, simultaneously promoting methylation of the mutant allele to reduce its expression and overexpressing the normal allele might avoid both the haploinsufficiency and the toxic gain of function observed in c9FTD/ALS. Consequently, using epigenetic modifiers to independently regulate expression of the normal and mutant alleles might be a novel strategy to explore in the near future. In fact, the mechanism of de novo gene methylation was first demonstrated in plants in 1994, and for the first time embraced the possibility of artificially modulating gene expression through epigenetic modulation [\[186](#page-15-8)]. Strategies to therapeutically reverse pathogenic changes in DNA methylation have been intensely studied since then to treat many diseases, including cancer and neurodegeneration [[76,](#page-12-24) [184](#page-15-9)]. Interestingly, to combat the potential consequences associated with *C9orf72* haploinsufficiency, small molecules targeting bromodomain proteins, proteins that recognize acetylated lysine residues on chromatin, have been shown to specifically enhance *C9orf72* RNA expression without affecting the epigenetic regulation of this gene [\[193](#page-15-10)].

Other therapeutic strategies targeting DNA methylation include developing drugs that alter the enzymatic activity of the hydroxymethylase TET to normalize 5mC or 5hmC levels, as well as altered activity of Dnmts to treat diseases where DNA methylation levels are perturbed [\[44](#page-11-28), [81](#page-12-25)]. While these strategies are being explored in diseases such

as cancer, where aberrantly regulated TET enzymes have both tumor suppressing and promoting capabilities [[86,](#page-12-26) [183](#page-15-11)], their potential to treat neurodegenerative diseases is even less known. DNA methylation studies characterizing changes in 5hmC levels in many neurodegenerative diseases aim to identify whether these changes may serve as potential biomarkers or therapeutic targets for those diseases [[3\]](#page-10-20). Interestingly, the involvement of 5hmC level changes in the regulation of transcription factors have been reported in neurodevelopment, neurodevelopmental diseases, aging, and neurodegenerative diseases [\[164](#page-14-13), [166](#page-14-14)], suggesting that these DNA marks are important for neuronal cell development and maintenance.

Integrative analysis of DNA methylation from ALS subject spinal cords, combined with transcriptome analyses, revealed a potential to use DNA methylation changes to identify suitable biomarker and therapeutic targets [\[54](#page-11-17)]. Experimentally, Dnmts were reported to be pro-apoptotic and increase 5hmC levels in motor neurons [[32\]](#page-11-29). Interestingly, pharmacologically treating a mouse model for motor neuron neurodegeneration and apoptosis with Dnmt inhibitors abrogated both elevated 5mC levels and apoptosis of the motor neurons. Furthermore, in human sporadic ALS, Dnmt1 and Dnmt3a levels were found increased in motor cortex and spinal cord neurons, as were 5mC levels in cortical pyramidal motor neurons. Dnmt3a expression was also found upregulated in FTD patients, and the methyltransferase was shown to regulate *GRN* promoter activity [\[11](#page-10-15)]. Taken together, targeting Dnmts may be an important therapeutic strategy to treat ALS.

#### **Histone modification in ALS and FTD**

Repressive histone marks at the *C9orf72* locus were found to reduce gene expression in both ALS and FTD patients with the HRE, but not in ALS patients without expanded repeats [[15\]](#page-10-6). Interestingly, treating fibroblasts derived from *C9orf72* HRE carriers with the demethylating agent 5-aza-2-deoxycytidine increased *C9orf72* mRNA expression. These data demonstrated that a potentially pathogenic histone modification event regulating *C9orf72* expression can be reversed. Therapeutic strategies targeting histone modifications have been intensely studied and developed as anticancer agents, where multi-faceted HDAC inhibitors have successfully reversed the effects of cancer-induced aberrant epigenetic changes [[100\]](#page-12-27). Among other diseases, HDAC inhibitors have also been explored to treat progressive neurodegeneration such as in Parkinson's disease (PD), where an imbalance between histone acetylation and deacetylation is known [\[73](#page-12-28)]. Because of the pathogenic potential for excessive histone deacetylation in PD, which leads to the altered expression of neuronal genes including those essential for survival, HDAC inhibitors were shown to provide neuroprotection in PD in vitro and in vivo models. However, the detailed molecular mechanisms for neuroprotection, and whether HDAC inhibitors will prove clinically successful in PD remain to be determined. HDAC inhibitors have also been tested for the epigenetic therapy of ALS in in vitro and animal models, with some proceeding to clinical trials. For example, a phase 2 study of sodium phenylbutyrate (NaPB) in ALS, a histone deacetylase inhibitor that was neuroprotective and prolonged the survival of SOD1-Gly93Ala mice [\[158](#page-14-29)], was found safe and well-tolerated by study participants, and importantly, significantly increased histone acetylation in blood buffy-coat specimens [\[42](#page-11-30)]. These data demonstrate the ability for a compound to alter the epigenomic architecture in patients.

#### **miRNAs in ALS and FTD**

Disrupting miRNA biogenesis has the potential to have many downstream consequences that affect multiple cellular pathways. Postmortem human spinal cord tissues from ALS subjects were assessed to identify aberrantly regulated microRNAs [[55\]](#page-11-20). Expression and systems biology analyses revealed that mature miRNAs are globally reduced and miRNA processing is altered. Furthermore, the same study found that cellular redistribution and cytoplasmic aggregation of TDP-43 regulates ALS-associated miRNA expression. Interplay between different cell types is also being explored for pathogenic roles in neurodegeneration, and evidence suggests that alterations in neighboring skeletal muscle may enhance damage to motor neurons in ALS [\[128](#page-13-28)]. In fact, selectively expressing mutant *SOD1* (G39A), a known mutation in ALS, in mouse muscles lead to the altered expression of miRNAs and mRNAs associated with myelin homeostasis in the spinal cords of the mice, therefore revealing interplay between cell types affects epigenetic regulation [\[49](#page-11-21)].

Since altered miRNA regulation could aberrantly regulate many cellular pathways, reversing pathogenic miRNA changes could provide therapeutic benefit. Both miRNA antagonists, to inhibit pathogenic gain-of-function mechanisms, and miRNA mimics, to replace downregulated miR-NAs, have been intensely explored to treat cancer [\[7](#page-10-21), [135](#page-13-29)]. In ALS mouse models, specific oligonucleuotide miRNA inhibitors known as anti-miRs, and miRNA agonists, have been used to demonstrate anti-miR delivery and function in the brain and spinal cord and to support targeting miRNAs to treat ALS. For example, anti-miRs to miR-155 or miR-29a extended the lifespans of *SOD1*-Gly93Ala mice [[97,](#page-12-29) [130](#page-13-30)]. Injecting miR-124a oligonucleotides into *SOD1*-Gly-93Ala mice prevented the pathological loss of the rodent excitatory amino acid transporter 2 (EAAT2/GLT1) [\[125](#page-13-31)], an astroglial glutamate transporter implicated in astroglial dysfunction in ALS. Similarly, in a mouse model expressing FTD-associated mutant *CHMP2B*, miR-124 levels were found decreased; re-expressing miR-124 in those mice decreased the levels of the miR-124 target AMPA receptor (AMPAR) and partially rescued behavioral deficits [\[62](#page-11-23)]. Moreover, the authors found correlative levels of miR-124 and AMPAR in induced pluripotent stem cell-derived neurons from subjects with behavioral variant FTD, supporting the investigation of targeting this miRNA for FTD therapy. Several more studies have identified additional alternatively expressed miRNAs in ALS and FTD that could be developed as drug targets or as circulatory biomarkers for ALS [\[37](#page-11-22), [69](#page-12-18), [87,](#page-12-19) [108,](#page-13-23) [133](#page-13-24), [173](#page-14-24), [194\]](#page-15-5). For example, miR-206 was found consistently altered during the course of disease in *SOD1*-Gly93Ala mice and increased in the circulation of a small cohort of ALS patients [\[173](#page-14-24)]. Furthermore, exploring therapeutic strategies targeting pathogenic miRNAs in ALS is further supported by the recent report on aberrantly regulated miRNAS in sporadic ALS discussed above [\[55](#page-11-20)]. Manipulating miRNAs have the advantage of altering multiple gene targets simultaneously using a single drug, but the disadvantage of non-specific binding to other miR-NAs resulting in deleterious off-target effects. Therefore, designing approaches to target pathogenic miRNA alterations requires great specificity.

## **Environmental signals and epigenetic modifications in ALS and FTD**

Communication between the environment and the genome is reflected through epigenetic alteration of the genetic information, and these changes can further be transmitted from cell to cell [\[139](#page-13-32)]. The dynamic interaction between the environment, the epigenome, and the transcriptome can be initiated by different factors, including age, exercise, diet, and toxic environmental exposure, to name a few. The first association between environmental factors and ALS was observed with the Chamorro indigenous people of Guam, who showed an extremely high incidence of ALS. Disease was linked to the accumulation of cycad neurotoxins, including beta-*N*-methylamino-L-alanine (BMAA) produced across the cyanobacterial order, from consuming flying fox  $[10, 21, 35]$  $[10, 21, 35]$  $[10, 21, 35]$  $[10, 21, 35]$  $[10, 21, 35]$  $[10, 21, 35]$ . It is well know that diet can induce epigenetic changes and cause various diseases [[148\]](#page-14-30), and the high incidence of ALS and other neurodegenerative diseases in Guam might be the first report for the field [\[79](#page-12-20)]. Association between BMAA, reactive oxigene species (ROS), and ALS has also been reported by others [[27,](#page-11-31) [35,](#page-11-24) [43](#page-11-32), [91](#page-12-30), [139](#page-13-32), [143](#page-14-31)].

Precisely, environmental signals can induce the liberation of free radicals leading to oxidative stress and alteration of epigenetic mechanisms, including histone remodeling, RNA editing, as well as DNA methylation and hydroxymethylation, and consequently alter gene expression [[70,](#page-12-31)

[83](#page-12-32), [117,](#page-13-33) [151](#page-14-32)]. For instance, stress after social defeat has been shown to cause both histone methylation and acetylation changes in rodents [\[175](#page-14-33)], and histone modifications after acute or chronic stress was demonstrated to cause gene activation or repression that modulate memory processes [\[152](#page-14-34)]. Acute stress has been shown to have a repressive effect on retrotransposable elements (RTE), as well as on coding and non-coding RNA of the hippocampus after histone 3 trimethylation at lysines 9 and 27. Such repression was shown to be lost after repeated stress, suggesting that induced expression of RTEs after prolonged stress may impair genomic stability and give rise to cognitive impairments [\[51](#page-11-33), [82,](#page-12-33) [84](#page-12-34), [89,](#page-12-35) [92](#page-12-36), [115,](#page-13-34) [147](#page-14-35)]. Interestingly, two recent reports of twins highlight the potential contribution of environmental factors to ALS onset. One study focused on monozygotic twins discordant for ALS, where no SNPs or genome-wide structural differences were found to explain the discordance, and the second study described a pair of *C9orf72* HRE-positive identical twins discordant for ALS [\[119](#page-13-25), [189](#page-15-6)].

Many other studies suggest connections between environmental influences and ALS. For example, accumulating evidence indicates heavy metals induce cellular stress and toxicity by catalyzing ROS formation, thereby mediating protein denaturation and aggregation, together leading to the inability of the ubiquitin/proteasome system to eliminate dysfunctional proteins [[121](#page-13-35)]. One study used a mixed isomer fluorescent indicator (5-(and-6)-carboxy-2′,7′-dichlorofluorescein diacetate or carboxy-DCFDA) to assess ROS generation within human neuronal cells [\[138\]](#page-13-36) after generating stress using different metal sulfates. Of note, aluminum sulfate demonstrated the greatest ability to induce ROS. They found that carboxy-DCFDA is a highly sensitive and long-lasting tracer enabling ROS quantification which can also be useful to evaluate epigenetic changes initiated by metal sulfate to ROS generation in neurodegeneration [[138\]](#page-13-36). Furthermore, regular exposure to electromagnetic fields may trigger DNA methylation and histone modification, as was suggested by a study of a large cohort of workers, especially resistance welders regularly exposed to extremely low frequency magnetic fields who had an increased risk for AD and ALS [\[72](#page-12-21)]. While no robust association has been reported between ALS and other environmental factors, some studies suggested connections with heavy metal exposures such as lead, mercury, and selenium, as well as agricultural and household pesticides- and herbicides-containing organophosphate (OP) [[9,](#page-10-22) [38](#page-11-34), [41,](#page-11-35) [47,](#page-11-36) [88](#page-12-37), [110,](#page-13-37) [123](#page-13-38), [159,](#page-14-36) [169](#page-14-37), [176\]](#page-14-38).

Finally, a common subject of discussion is the increased ALS frequency among soccer and football players [\[1](#page-10-23), [33\]](#page-11-37), as well as in war veterans [\[80](#page-12-38), [122\]](#page-13-39) further supporting that neurological symptoms of ALS arise from environmental exposures. While different factors such as physical activity, drug consumption, and ischemia caused by head injuries have been proposed to increase the production of ROS in ALS, the specific contribution of these exposures to epigenetic changes and neurodegeneration remains to be determined.

## **Concluding remarks**

While much of the epigenome was initially believed to be inherited and mostly stable, that it is in fact dynamic, and that it is altered in ALS and FTD makes the epigenome an attractive target for therapeutic development. Much progress has been made in characterizing the status of different epigenetic mechanisms in neurodegenerative diseases, but whether altered epigenetic regulation is a consequence of neurodegeneration or pathogenic remains a matter of debate [[61](#page-11-38)]. For example, a recent study observed distinct transcriptome alterations in c9ALS and sALS brain [\[142\]](#page-13-5), but whether these unique signatures are the result of aberrant epigenetic regulation has not yet been demonstrated. If epigenetic mechanisms are indeed found to mediate transcriptome alterations, these unique changes would be useful to monitor the effectiveness of epigenetic targeting therapeutics in restoring transcriptome defects. In laboratory and clinical settings, strategies targeting epigenetic writers, readers, and erasers have shown some support to treat neurodegeneration and cancer, suggesting that therapeutic agents that reverse pathogenic epigenetic alterations have the potential to treat ALS and FTD. Because RNAs mediate epigenetic regulation by recruiting important chromatin remodeling factors, RNAs, such as miRNAs, may be interesting therapeutics targets to develop based on their ability to recognize specific sequences. While research identifying epigenetic pathogenic mechanisms is still in its infancy for ALS and FTD, lessons learned from other diseases, especially cancer, have the potential to fast-track the development of epigenetic targeting strategies from bench to bedside to treat ALS and FTD. Drugs already approved to target epigenetic mechanisms may turn out to be efficacious and restore altered cellular pathways in ALS and FTD; already FDA approved to treat cancer, Dnmt and HDAC inhibitors [\[53,](#page-11-8) [76\]](#page-12-24) may be able to reverse aberrant epigenetic changes in the central nervous system [[184](#page-15-9)]. Thus, the emerging field of epigenetics provides new hope for patients with lethal ALS and FTD, and exciting studies exploring new territories should be expected in the near future.

### **References**

- <span id="page-10-23"></span>1. Abel EL (2007) Football increases the risk for Lou Gehrig's disease, amyotrophic lateral sclerosis. Percept Mot Skills 104(3 Pt 2):1251–1254
- <span id="page-10-1"></span>2. Al-Chalabi A, Jones A, Troakes C, King A, Al-Sarraj S, van den Berg LH (2012) The genetics and neuropathology of amyotrophic lateral sclerosis. Acta Neuropathol 124(3):339–352
- <span id="page-10-20"></span>3. Al-Mahdawi S, Virmouni SA, Pook MA (2014) The emerging role of 5-hydroxymethylcytosine in neurodegenerative diseases. Front Neurosci 8:397
- <span id="page-10-18"></span>4. Almeida S, Gascon E, Tran H, Chou HJ, Gendron TF, Degroot S et al (2013) Modeling key pathological features of frontotemporal dementia with C9ORF72 repeat expansion in iPSCderived human neurons. Acta Neuropathol 126(3):385–399
- <span id="page-10-8"></span>5. Amaral PP, Dinger ME, Mercer TR, Mattick JS (2008) The eukaryotic genome as an RNA machine. Science 319(5871):1787–1789
- <span id="page-10-10"></span>6. Amaral PP, Mattick JS (2008) Noncoding RNA in development. Mamm Genome 19(7–8):454–492
- <span id="page-10-21"></span>7. Bader AG, Brown D, Winkler M (2010) The promise of micro-RNA replacement therapy. Cancer Res 70(18):7027–7030
- <span id="page-10-4"></span>8. Baizabal-Carvallo JF, Jankovic J (2016) Parkinsonism, movement disorders and genetics in frontotemporal dementia. Nat Rev Neurol. 12(3):175–185
- <span id="page-10-22"></span>9. Bakir F, Damluji SF, Amin-Zaki L, Murtadha M, Khalidi A, al-Rawi NY et al (1973) Methylmercury poisoning in Iraq. Science 181(4096):230–241
- <span id="page-10-16"></span>10. Banack SA, Cox PA (2003) Biomagnification of cycad neurotoxins in flying foxes: implications for ALS-PDC in Guam. Neurology 61(3):387–389
- <span id="page-10-15"></span>11. Banzhaf-Strathmann J, Claus R, Mucke O, Rentzsch K, van der Zee J, Engelborghs S et al (2013) Promoter DNA methylation regulates progranulin expression and is altered in FTLD. Acta Neuropathol Commun 1:16
- <span id="page-10-19"></span>12. Bauer PO (2016) Methylation of C9orf72 expansion reduces RNA foci formation and dipeptide-repeat proteins expression in cells. Neurosci Lett 612:204–209
- <span id="page-10-13"></span>13. Behm M, Ohman M (2016) RNA editing: a contributor to neuronal dynamics in the mammalian brain. Trends Genet 32(3):165–175
- <span id="page-10-5"></span>14. Belzil VV, Bauer PO, Gendron TF, Murray ME, Dickson D, Petrucelli L (2014) Characterization of DNA hypermethylation in the cerebellum of c9FTD/ALS patients. Brain Res 1584:15–21
- <span id="page-10-6"></span>15. Belzil VV, Bauer PO, Prudencio M, Gendron TF, Stetler CT, Yan IK et al (2013) Reduced C9orf72 gene expression in c9FTD/ALS is caused by histone trimethylation, an epigenetic event detectable in blood. Acta Neuropathol 126(6):895–905
- <span id="page-10-14"></span>16. Belzil VV, Gendron TF, Petrucelli L (2013) RNA-mediated toxicity in neurodegenerative disease. Mol Cell Neurosci 56C:406–419
- <span id="page-10-9"></span>17. Berezikov E, Chung WJ, Willis J, Cuppen E, Lai EC (2007) Mammalian mirtron genes. Mol Cell 28(2):328–336
- <span id="page-10-12"></span>18. Bernstein E, Allis CD (2005) RNA meets chromatin. Genes Dev 19(14):1635–1655
- <span id="page-10-11"></span>19. Bonasio R, Shiekhattar R (2014) Regulation of transcription by long noncoding RNAs. Annu Rev Genet 48:433–455
- <span id="page-10-0"></span>20. Bradley WG (2000) Neurology in clinical practice, 3rd edn. Butterworth-Heinemann, Boston
- <span id="page-10-17"></span>21. Bradley WG, Mash DC (2009) Beyond Guam: the cyanobacteria/BMAA hypothesis of the cause of ALS and other neurodegenerative diseases. Amyotroph Lateral Scler 10(Suppl 2):7–20
- <span id="page-10-7"></span>22. Britten RJ, Davidson EH (1969) Gene regulation for higher cells: a theory. Science 165(3891):349–357
- <span id="page-10-3"></span>23. Burrell JR, Kiernan MC, Vucic S, Hodges JR (2011) Motor neuron dysfunction in frontotemporal dementia. Brain 134(Pt 9):2582–2594
- <span id="page-10-2"></span>24. Byrne S, Heverin M, Elamin M, Bede P, Lynch C, Kenna K et al (2013) Aggregation of neurologic and neuropsychiatric disease in amyotrophic lateral sclerosis kindreds: a population-based case-control cohort study of familial and sporadic amyotrophic lateral sclerosis. Ann Neurol 74(5):699–708
- <span id="page-11-1"></span>25. Byrne S, Walsh C, Lynch C, Bede P, Elamin M, Kenna K et al (2011) Rate of familial amyotrophic lateral sclerosis: a systematic review and meta-analysis. J Neurol Neurosurg Psychiatry 82(6):623–627
- <span id="page-11-15"></span>26. Callahan KP, Butler JS (2008) Lifting the veil on the transcriptome. Genome Biol 9(4):218
- <span id="page-11-31"></span>27. Caller TA, Doolin JW, Haney JF, Murby AJ, West KG, Farrar HE et al (2009) A cluster of amyotrophic lateral sclerosis in New Hampshire: a possible role for toxic cyanobacteria blooms. Amyotroph Lateral Scler 10(Suppl 2):101–108
- <span id="page-11-13"></span>28. Carninci P, Kasukawa T, Katayama S, Gough J, Frith MC, Maeda N et al (2005) The transcriptional landscape of the mammalian genome. Science 309(5740):1559–1563
- <span id="page-11-0"></span>29. Charcot JM, Joffroy A (1869) Deux cas d'atrophie musculaire progressive avec lésions de la substance grise et des faisceaux antéraux de la moelle épinière. Arch Physiol Neurol Pathol 2:744–754
- <span id="page-11-10"></span>30. Chen H, Dzitoyeva S, Manev H (2012) Effect of aging on 5-hydroxymethylcytosine in the mouse hippocampus. Restor Neurol Neurosci 30(3):237–245
- <span id="page-11-9"></span>31. Chen T (2011) Mechanistic and functional links between histone methylation and DNA methylation. Prog Mol Biol Transl Sci 101:335–348
- <span id="page-11-29"></span>32. Chestnut BA, Chang Q, Price A, Lesuisse C, Wong M, Martin LJ (2011) Epigenetic regulation of motor neuron cell death through DNA methylation. J Neurosci 31(46):16619–16636
- <span id="page-11-37"></span>33. Chio A, Benzi G, Dossena M, Mutani R, Mora G (2005) Severely increased risk of amyotrophic lateral sclerosis among Italian professional football players. Brain 128(Pt 3):472–476
- <span id="page-11-4"></span>34. Chio A, Schymick JC, Restagno G, Scholz SW, Lombardo F, Lai SL et al (2009) A two-stage genome-wide association study of sporadic amyotrophic lateral sclerosis. Hum Mol Genet 18(8):1524–1532
- <span id="page-11-24"></span>35. Chiu AS, Gehringer MM, Welch JH, Neilan BA (2011) Does alpha-amino-beta-methylaminopropionic acid (BMAA) play a role in neurodegeneration? Int J Environ Res Public Health 8(9):3728–3746
- <span id="page-11-25"></span>36. Ciura S, Lattante S, Le Ber I, Latouche M, Tostivint H, Brice A et al (2013) Loss of function of C9orf72 causes motor deficits in a zebrafish model of Amyotrophic Lateral Sclerosis. Ann Neurol 74(2):180–187
- <span id="page-11-22"></span>37. Cloutier F, Marrero A, O'Connell C, Morin P Jr (2015) Micro-RNAs as potential circulating biomarkers for amyotrophic lateral sclerosis. J Mol Neurosci 56(1):102–112
- <span id="page-11-34"></span>38. Combs GF Jr (2001) Selenium in global food systems. The British journal of nutrition 85(5):517–547
- <span id="page-11-7"></span>39. Crick FH (1958) On protein synthesis. Symp Soc Exp Biol 12:138–163
- <span id="page-11-5"></span>40. Cronin S, Berger S, Ding J, Schymick JC, Washecka N, Hernandez DG et al (2008) A genome-wide association study of sporadic ALS in a homogenous Irish population. Hum Mol Genet 17(5):768–774
- <span id="page-11-35"></span>41. Cronin S, Greenway MJ, Prehn JH, Hardiman O (2007) Paraoxonase promoter and intronic variants modify risk of sporadic amyotrophic lateral sclerosis. J Neurol Neurosurg Psychiatry 78(9):984–986
- <span id="page-11-30"></span>42. Cudkowicz ME, Andres PL, Macdonald SA, Bedlack RS, Choudry R, Brown RH Jr et al (2009) Phase 2 study of sodium phenylbutyrate in ALS. Amyotroph Lateral Scler 10(2):99–106
- <span id="page-11-32"></span>43. Dastur DK (1964) Cycad toxicity in monkeys: clinical, pathological, and biochemical aspects. Fed Proc 23:1368–1369
- <span id="page-11-28"></span>44. Day JJ, Kennedy AJ, Sweatt JD (2015) DNA methylation and its implications and accessibility for neuropsychiatric therapeutics. Annu Rev Pharmacol Toxicol 55:591–611
- <span id="page-11-3"></span>45. DeJesus-Hernandez M, Mackenzie IR, Boeve BF, Boxer AL, Baker M, Rutherford NJ et al (2011) Expanded GGGGCC

hexanucleotide repeat in noncoding region of C9ORF72 causes chromosome 9p-linked FTD and ALS. Neuron 72(2):245–256

- <span id="page-11-12"></span>46. Delatte B, Wang F, Ngoc LV, Collignon E, Bonvin E, Deplus R et al (2016) RNA biochemistry. Transcriptome-wide distribution and function of RNA hydroxymethylcytosine. Science 351(6270):282–285
- <span id="page-11-36"></span>47. Diekstra FP, Beleza-Meireles A, Leigh NP, Shaw CE, Al-Chalabi A (2009) Interaction between PON1 and population density in amyotrophic lateral sclerosis. NeuroReport 20(2):186–190
- <span id="page-11-16"></span>48. Dinger ME, Mercer TR, Mattick JS (2008) RNAs as extracellular signaling molecules. J Mol Endocrinol 40(4):151–159
- <span id="page-11-21"></span>49. Dobrowolny G, Bernardini C, Martini M, Baranzini M, Barba M, Musaro A (2015) Muscle expression of SOD1(G93A) modulates microRNA and mRNA transcription pattern associated with the myelination process in the spinal cord of transgenic mice. Front Cell Neurosci 9:463
- <span id="page-11-6"></span>50. Donnelly CJ, Zhang PW, Pham JT, Haeusler AR, Mistry NA, Vidensky S et al (2013) RNA toxicity from the ALS/FTD C9ORF72 expansion is mitigated by antisense intervention. Neuron 80(2):415–428
- <span id="page-11-33"></span>51. Erwin JA, Marchetto MC, Gage FH (2014) Mobile DNA elements in the generation of diversity and complexity in the brain. Nat Rev Neurosci 15(8):497–506
- <span id="page-11-27"></span>52. Esanov R, Belle KC, van Blitterswijk M, Belzil VV, Rademakers R, Dickson DW et al (2015) C9orf72 promoter hypermethylation is reduced while hydroxymethylation is acquired during reprogramming of ALS patient cells. Exp Neurol 277:171–177
- <span id="page-11-8"></span>53. Falkenberg KJ, Johnstone RW (2014) Histone deacetylases and their inhibitors in cancer, neurological diseases and immune disorders. Nat Rev Drug Discov 13(9):673–691
- <span id="page-11-17"></span>54. Figueroa-Romero C, Hur J, Bender DE, Delaney CE, Cataldo MD, Smith AL et al (2012) Identification of epigenetically altered genes in sporadic amyotrophic lateral sclerosis. PLoS ONE 7(12):e52672
- <span id="page-11-20"></span>55. Figueroa-Romero C, Hur J, Lunn JS, Paez-Colasante X, Bender DE, Yung R et al (2016) Expression of microRNAs in human post-mortem amyotrophic lateral sclerosis spinal cords provides insight into disease mechanisms. Mol Cell Neurosci 71:34–45
- <span id="page-11-19"></span>56. Finch N, Baker M, Crook R, Swanson K, Kuntz K, Surtees R et al (2009) Plasma progranulin levels predict progranulin mutation status in frontotemporal dementia patients and asymptomatic family members. Brain 132(Pt 3):583–591
- <span id="page-11-26"></span>57. Fratta P, Poulter M, Lashley T, Rohrer JD, Polke JM, Beck J et al (2013) Homozygosity for the C9orf72 GGGGCC repeat expansion in frontotemporal dementia. Acta Neuropathol 126(3):401–409
- <span id="page-11-11"></span>58. Fu L, Guerrero CR, Zhong N, Amato NJ, Liu Y, Liu S et al (2014) Tet-mediated formation of 5-hydroxymethylcytosine in RNA. J Am Chem Soc 136(33):11582–11585
- <span id="page-11-18"></span>59. Galimberti D, D'Addario C, Dell'osso B, Fenoglio C, Marcone A, Cerami C et al (2013) Progranulin gene (GRN) promoter methylation is increased in patients with sporadic frontotemporal lobar degeneration. Neurol Sci 34(6):899–903
- <span id="page-11-14"></span>60. Gapp K, Jawaid A, Sarkies P, Bohacek J, Pelczar P, Prados J et al (2014) Implication of sperm RNAs in transgenerational inheritance of the effects of early trauma in mice. Nat Neurosci 17(5):667–669
- <span id="page-11-38"></span>61. Gascon E, Gao FB (2012) Cause or Effect: misregulation of microRNA Pathways in Neurodegeneration. Front Neurosci 6:48
- <span id="page-11-23"></span>62. Gascon E, Lynch K, Ruan H, Almeida S, Verheyden JM, Seeley WW et al (2014) Alterations in microRNA-124 and AMPA receptors contribute to social behavioral deficits in frontotemporal dementia. Nat Med 20(12):1444–1451
- <span id="page-11-2"></span>63. Gibson SB, Figueroa KP, Bromberg MB, Pulst SM, Cannon-Albright L (2014) Familial clustering of ALS in a populationbased resource. Neurology 82(1):17–22
- <span id="page-12-22"></span>64. Gijselinck I, Van Langenhove T, van der Zee J, Sleegers K, Philtjens S, Kleinberger G et al (2012) A C9orf72 promoter repeat expansion in a Flanders-Belgian cohort with disorders of the frontotemporal lobar degeneration-amyotrophic lateral sclerosis spectrum: a gene identification study. Lancet Neurol 11(1):54–65
- <span id="page-12-0"></span>65. Giordana MT, Ferrero P, Grifoni S, Pellerino A, Naldi A, Montuschi A (2011) Dementia and cognitive impairment in amyotrophic lateral sclerosis: a review. Neurol Sci 32(1):9–16
- <span id="page-12-7"></span>66. Globisch D, Munzel M, Muller M, Michalakis S, Wagner M, Koch S et al (2010) Tissue distribution of 5-hydroxymethylcytosine and search for active demethylation intermediates. PLoS ONE 5(12):e15367
- <span id="page-12-1"></span>67. Gordon PH, Delgadillo D, Piquard A, Bruneteau G, Pradat PF, Salachas F et al (2011) The range and clinical impact of cognitive impairment in French patients with ALS: a cross-sectional study of neuropsychological test performance. Amyotroph Lateral Scler 12(5):372–378
- <span id="page-12-2"></span>68. Graff-Radford NR, Woodruff BK (2007) Frontotemporal dementia. Semin Neurol 27(1):48–57
- <span id="page-12-18"></span>69. Grasso M, Piscopo P, Confaloni A, Denti MA (2014) Circulating miRNAs as biomarkers for neurodegenerative disorders. Molecules 19(5):6891–6910
- <span id="page-12-31"></span>70. Griffiths BB, Hunter RG (2014) Neuroepigenetics of stress. Neuroscience 275:420–435
- <span id="page-12-5"></span>71. Guerreiro R, Bras J, Hardy J (2015) SnapShot: genetics of ALS and FTD. Cell 160(4):798 **(e791)**
- <span id="page-12-21"></span>72. Hakansson N, Gustavsson P, Johansen C, Floderus B (2003) Neurodegenerative diseases in welders and other workers exposed to high levels of magnetic fields. Epidemiology 14(4):420–426 **(discussion 427–428)**
- <span id="page-12-28"></span>73. Harrison IF, Dexter DT (2013) Epigenetic targeting of histone deacetylase: therapeutic potential in Parkinson's disease? Pharmacol Ther 140(1):34–52
- <span id="page-12-15"></span>74. Hasler J, Samuelsson T, Strub K (2007) Useful 'junk': Alu RNAs in the human transcriptome. Cell Mol Life Sci 64(14):1793–1800
- <span id="page-12-17"></span>75. Himber C, Dunoyer P, Moissiard G, Ritzenthaler C, Voinnet O (2003) Transitivity-dependent and -independent cell-to-cell movement of RNA silencing. EMBO J 22(17):4523–4533
- <span id="page-12-24"></span>76. Ho AS, Turcan S, Chan TA (2013) Epigenetic therapy: use of agents targeting deacetylation and methylation in cancer management. Onco Targets Ther 6:223–232
- <span id="page-12-3"></span>77. Hodges J (2012) Familial frontotemporal dementia and amyotrophic lateral sclerosis associated with the C9ORF72 hexanucleotide repeat. Brain 135(Pt 3):652–655
- <span id="page-12-13"></span>78. Holoch D, Moazed D (2015) RNA-mediated epigenetic regulation of gene expression. Nat Rev Genet 16(2):71–84
- <span id="page-12-20"></span>79. Holtcamp W (2012) The emerging science of BMAA: do cyanobacteria contribute to neurodegenerative disease? Environ Health Perspect 120(3):A110–116
- <span id="page-12-38"></span>80. Horner RD, Grambow SC, Coffman CJ, Lindquist JH, Oddone EZ, Allen KD et al (2008) Amyotrophic lateral sclerosis among 1991 Gulf War veterans: evidence for a time-limited outbreak. Neuroepidemiology 31(1):28–32
- <span id="page-12-25"></span>81. Huang Y, Rao A (2014) Connections between TET proteins and aberrant DNA modification in cancer. Trends Genet 30(10):464–474
- <span id="page-12-33"></span>82. Hunter RG, Gagnidze K, McEwen BS, Pfaff DW (2015) Stress and the dynamic genome: steroids, epigenetics, and the transposome. Proc Natl Acad Sci USA 112(22):6828–6833
- <span id="page-12-32"></span>83. Hunter RG, McEwen BS (2013) Stress and anxiety across the lifespan: structural plasticity and epigenetic regulation. Epigenomics 5(2):177–194
- <span id="page-12-34"></span>84. Hunter RG, McEwen BS, Pfaff DW (2013) Environmental stress and transposon transcription in the mammalian brain. Mob Genet Elements 3(2):e24555
- <span id="page-12-11"></span>85. Hutvagner G, Zamore PD (2002) A microRNA in a multipleturnover RNAi enzyme complex. Science 297(5589):2056–2060
- <span id="page-12-26"></span>86. Jeschke J, Collignon E, Fuks F (2016) Portraits of TET-mediated DNA hydroxymethylation in cancer. Curr Opin Genet Dev 36:16–26
- <span id="page-12-19"></span>87. Jiao J, Herl LD, Farese RV, Gao FB (2010) MicroRNA-29b regulates the expression level of human progranulin, a secreted glycoprotein implicated in frontotemporal dementia. PLoS ONE 5(5):e10551
- <span id="page-12-37"></span>88. Johnson FO, Atchison WD (2009) The role of environmental mercury, lead and pesticide exposure in development of amyotrophic lateral sclerosis. Neurotoxicology 30(5):761–765
- <span id="page-12-35"></span>89. Johnson R, Guigo R (2014) The RIDL hypothesis: transposable elements as functional domains of long noncoding RNAs. RNA 20(7):959–976
- <span id="page-12-10"></span>90. Kapranov P, Drenkow J, Cheng J, Long J, Helt G, Dike S et al (2005) Examples of the complex architecture of the human transcriptome revealed by RACE and high-density tiling arrays. Genome Res 15(7):987–997
- <span id="page-12-30"></span>91. Karlsson O, Roman E, Berg AL, Brittebo EB (2011) Early hippocampal cell death, and late learning and memory deficits in rats exposed to the environmental toxin BMAA (beta-*N*-methylamino-l-alanine) during the neonatal period. Behav Brain Res 219(2):310–320
- <span id="page-12-36"></span>92. Khalil AM, Guttman M, Huarte M, Garber M, Raj A, Rivea Morales D et al (2009) Many human large intergenic noncoding RNAs associate with chromatin-modifying complexes and affect gene expression. Proc Natl Acad Sci USA 106(28):11667–11672
- <span id="page-12-14"></span>93. Kim EZ, Wespiser AR, Caffrey DR (2016) The domain structure and distribution of Alu elements in long noncoding RNAs and mRNAs. RNA 22(2):254–264
- <span id="page-12-9"></span>94. Klug M, Heinz S, Gebhard C, Schwarzfischer L, Krause SW, Andreesen R et al (2010) Active DNA demethylation in human postmitotic cells correlates with activating histone modifications, but not transcription levels. Genome Biol 11(6):R63
- <span id="page-12-8"></span>95. Kohli RM, Zhang Y (2013) TET enzymes, TDG and the dynamics of DNA demethylation. Nature 502(7472):472–479
- <span id="page-12-23"></span>96. Koppers M, Blokhuis AM, Westeneng HJ, Terpstra ML, Zundel CA, Vieira de Sa R et al (2015) C9orf72 ablation in mice does not cause motor neuron degeneration or motor deficits. Ann Neurol 78:426–438
- <span id="page-12-29"></span>97. Koval ED, Shaner C, Zhang P, du Maine X, Fischer K, Tay J et al (2013) Method for widespread microRNA-155 inhibition prolongs survival in ALS-model mice. Hum Mol Genet 22(20):4127–4135
- <span id="page-12-6"></span>98. Kriaucionis S, Heintz N (2009) The nuclear DNA base 5-hydroxymethylcytosine is present in Purkinje neurons and the brain. Science 324(5929):929–930
- <span id="page-12-4"></span>99. Laaksovirta H, Peuralinna T, Schymick JC, Scholz SW, Lai SL, Myllykangas L et al (2010) Chromosome 9p21 in amyotrophic lateral sclerosis in Finland: a genome-wide association study. Lancet Neurol 9(10):978–985
- <span id="page-12-27"></span>100. Lakshmaiah KC, Jacob LA, Aparna S, Lokanatha D, Saldanha SC (2014) Epigenetic therapy of cancer with histone deacetylase inhibitors. J Cancer Res Ther 10(3):469–478
- <span id="page-12-16"></span>101. Lam MT, Cho H, Lesch HP, Gosselin D, Heinz S, Tanaka-Oishi Y et al (2013) Rev-Erbs repress macrophage gene expression by inhibiting enhancer-directed transcription. Nature 498(7455):511–515
- <span id="page-12-12"></span>102. Leung AK, Sharp PA (2007) microRNAs: a safeguard against turmoil? Cell 130(4):581–585
- <span id="page-13-21"></span>103. Li Y, Chen JA, Sears RL, Gao F, Klein ED, Karydas A et al (2014) An epigenetic signature in peripheral blood associated with the haplotype on 17q21.31, a risk factor for neurodegenerative tauopathy. PLoS Genet 10(3):e1004211
- <span id="page-13-22"></span>104. Liu EY, Russ J, Wu K, Neal D, Suh E, McNally AG et al (2014) C9orf72 hypermethylation protects against repeat expansion-associated pathology in ALS/FTD. Acta Neuropathol 128(4):525–541
- <span id="page-13-10"></span>105. Lo R, Weksberg R (2014) Biological and biochemical modulation of DNA methylation. Epigenomics 6(6):593–602
- <span id="page-13-0"></span>106. Lomen-Hoerth C, Anderson T, Miller B (2002) The overlap of amyotrophic lateral sclerosis and frontotemporal dementia. Neurology 59(7):1077–1079
- <span id="page-13-3"></span>107. Majounie E, Renton AE, Mok K, Dopper EG, Waite A, Rollinson S et al (2012) Frequency of the C9orf72 hexanucleotide repeat expansion in patients with amyotrophic lateral sclerosis and frontotemporal dementia: a cross-sectional study. Lancet Neurol 11(4):323–330
- <span id="page-13-23"></span>108. Marcuzzo S, Bonanno S, Kapetis D, Barzago C, Cavalcante P, D'Alessandro S et al (2015) Up-regulation of neural and cell cycle-related microRNAs in brain of amyotrophic lateral sclerosis mice at late disease stage. Mol Brain 8:5
- <span id="page-13-17"></span>109. Mariner PD, Walters RD, Espinoza CA, Drullinger LF, Wagner SD, Kugel JF et al (2008) Human Alu RNA is a modular transacting repressor of mRNA transcription during heat shock. Mol Cell 29(4):499–509
- <span id="page-13-37"></span>110. Matin MA, Hussain K (1985) Striatal neurochemical changes and motor dysfunction in mipafox-treated animals. Methods Find Exp Clin Pharmacol 7(2):79–81
- <span id="page-13-7"></span>111. Mattick JS (2001) Non-coding RNAs: the architects of eukaryotic complexity. EMBO Rep 2(11):986–991
- <span id="page-13-9"></span>112. Mattick JS (2007) A new paradigm for developmental biology. J Exp Biol 210(Pt 9):1526–1547
- <span id="page-13-8"></span>113. Mattick JS, Amaral PP, Dinger ME, Mercer TR, Mehler MF (2009) RNA regulation of epigenetic processes. Bioessays 31(1):51–59
- <span id="page-13-13"></span>114. Mattick JS, Rinn JL (2015) Discovery and annotation of long noncoding RNAs. Nat Struct Mol Biol 22(1):5–7
- <span id="page-13-34"></span>115. McEwen BS, Bowles NP, Gray JD, Hill MN, Hunter RG, Karatsoreos IN et al (2015) Mechanisms of stress in the brain. Nat Neurosci 18(10):1353–1363
- <span id="page-13-27"></span>116. McMillan CT, Russ J, Wood EM, Irwin DJ, Grossman M, McCluskey L et al (2015) C9orf72 promoter hypermethylation is neuroprotective: neuroimaging and neuropathologic evidence. Neurology 84(16):1622–1630
- <span id="page-13-33"></span>117. Meaney MJ, Szyf M (2005) Environmental programming of stress responses through DNA methylation: life at the interface between a dynamic environment and a fixed genome. Dialogues Clin Neurosci 7(2):103–123
- <span id="page-13-16"></span>118. Meister G (2013) Argonaute proteins: functional insights and emerging roles. Nat Rev Genet 14(7):447–459
- <span id="page-13-25"></span>119. Meltz Steinberg K, Nicholas TJ, Koboldt DC, Yu B, Mardis E, Pamphlett R (2015) Whole genome analyses reveal no pathogenetic single nucleotide or structural differences between monozygotic twins discordant for amyotrophic lateral sclerosis. Amyotroph Lateral Scler Frontotemporal Degener 16(5–6):385–392
- <span id="page-13-1"></span>120. Mercy L, Hodges JR, Dawson K, Barker RA, Brayne C (2008) Incidence of early-onset dementias in Cambridgeshire, United Kingdom. Neurology 71(19):1496–1499
- <span id="page-13-35"></span>121. Migliore L, Coppede F (2009) Environmental-induced oxidative stress in neurodegenerative disorders and aging. Mutat Res 674(1–2):73–84
- <span id="page-13-39"></span>122. Miranda ML, Alicia Overstreet Galeano M, Tassone E, Allen KD, Horner RD (2008) Spatial analysis of the etiology of amyotrophic lateral sclerosis among 1991 Gulf War veterans. Neurotoxicology 29(6):964–970
- <span id="page-13-38"></span>123. Morahan JM, Yu B, Trent RJ, Pamphlett R (2007) A geneenvironment study of the paraoxonase 1 gene and pesticides in amyotrophic lateral sclerosis. Neurotoxicology 28(3):532–540
- <span id="page-13-20"></span>124. Morahan JM, Yu B, Trent RJ, Pamphlett R (2009) A genomewide analysis of brain DNA methylation identifies new candidate genes for sporadic amyotrophic lateral sclerosis. Amyotroph Lateral Scler 10(5–6):418–429
- <span id="page-13-31"></span>125. Morel L, Regan M, Higashimori H, Ng SK, Esau C, Vidensky S et al (2013) Neuronal exosomal miRNA-dependent translational regulation of astroglial glutamate transporter GLT1. J Biol Chem 288(10):7105–7116
- <span id="page-13-26"></span>126. Mori K, Weng SM, Arzberger T, May S, Rentzsch K, Kremmer E et al (2013) The C9orf72 GGGGCC repeat is translated into aggregating dipeptide-repeat proteins in FTLD/ALS. Science 339(6125):1335–1338
- <span id="page-13-6"></span>127. Morris KV, Mattick JS (2014) The rise of regulatory RNA. Nat Rev Genet 15(6):423–437
- <span id="page-13-28"></span>128. Musaro A (2013) Understanding ALS: new therapeutic approaches. FEBS J 280(17):4315–4322
- <span id="page-13-18"></span>129. Navaratnam N, Sarwar R (2006) An overview of cytidine deaminases. Int J Hematol 83(3):195–200
- <span id="page-13-30"></span>130. Nolan K, Mitchem MR, Jimenez-Mateos EM, Henshall DC, Concannon CG, Prehn JH (2014) Increased expression of microRNA-29a in ALS mice: functional analysis of its inhibition. J Mol Neurosci 53(2):231–241
- <span id="page-13-19"></span>131. Oates N, Pamphlett R (2007) An epigenetic analysis of SOD1 and VEGF in ALS. Amyotroph Lateral Scler 8(2):83–86
- <span id="page-13-15"></span>132. Okamura K, Hagen JW, Duan H, Tyler DM, Lai EC (2007) The mirtron pathway generates microRNA-class regulatory RNAs in Drosophila. Cell 130(1):89–100
- <span id="page-13-24"></span>133. Paez-Colasante X, Figueroa-Romero C, Sakowski SA, Goutman SA, Feldman EL (2015) Amyotrophic lateral sclerosis: mechanisms and therapeutics in the epigenomic era. Nat Rev Neurol 11(5):266–279
- <span id="page-13-11"></span>134. Penn NW, Suwalski R, O'Riley C, Bojanowski K, Yura R (1972) The presence of 5-hydroxymethylcytosine in animal deoxyribonucleic acid. Biochem J 126(4):781–790
- <span id="page-13-29"></span>135. Pereira DM, Rodrigues PM, Borralho PM, Rodrigues CM (2013) Delivering the promise of miRNA cancer therapeutics. Drug Discov Today 18(5–6):282–289
- <span id="page-13-2"></span>136. Perry DC, Miller BL (2013) Frontotemporal dementia. Semin Neurol 33(4):336–341
- <span id="page-13-14"></span>137. Peschansky VJ, Wahlestedt C (2014) Non-coding RNAs as direct and indirect modulators of epigenetic regulation. Epigenetics 9(1):3–12
- <span id="page-13-36"></span>138. Pogue AI, Jones BM, Bhattacharjee S, Percy ME, Zhao Y, Lukiw WJ (2012) Metal-sulfate induced generation of ROS in human brain cells: detection using an isomeric mixture of 5- and 6-carboxy-2′,7′-dichlorofluorescein diacetate (carboxy-DCFDA) as a cell permeant tracer. Int J Mol Sci 13(8):9615–9626
- <span id="page-13-32"></span>139. Polsky FI, Nunn PB, Bell EA (1972) Distribution and toxicity of alpha-amino-beta-methylaminopropionic acid. Fed Proc 31(5):1473–1475
- <span id="page-13-4"></span>140. Polymenidou M, Lagier-Tourenne C, Hutt KR, Bennett CF, Cleveland DW, Yeo GW (2012) Misregulated RNA processing in amyotrophic lateral sclerosis. Brain Res 1462:3–15
- <span id="page-13-12"></span>141. Privat E, Sowers LC (1996) Photochemical deamination and demethylation of 5-methylcytosine. Chem Res Toxicol 9(4):745–750
- <span id="page-13-5"></span>142. Prudencio M, Belzil VV, Batra R, Ross CA, Gendron TF, Pregent LJ et al (2015) Distinct brain transcriptome profiles in C9orf72-associated and sporadic ALS. Nat Neurosci 18(8):1175–1182
- <span id="page-14-31"></span>143. Purdie EL, Samsudin S, Eddy FB, Codd GA (2009) Effects of the cyanobacterial neurotoxin beta-*N*-methylamino-L-alanine on the early-life stage development of zebrafish (Danio rerio). Aquat Toxicol 95(4):279–284
- <span id="page-14-0"></span>144. Raaphorst J, de Visser M, Linssen WH, de Haan RJ, Schmand B (2010) The cognitive profile of amyotrophic lateral sclerosis: a meta-analysis. Amyotroph Lateral Scler 11(1–2):27–37
- <span id="page-14-4"></span>145. Rademakers R, van Blitterswijk M (2013) Motor neuron disease in 2012: novel causal genes and disease modifiers. Nat Rev Neurol 9(2):63–64
- <span id="page-14-21"></span>146. Rechavi O, Houri-Ze'evi L, Anava S, Goh WS, Kerk SY, Hannon GJ et al (2014) Starvation-induced transgenerational inheritance of small RNAs in *C. elegans*. Cell 158(2):277–287
- <span id="page-14-35"></span>147. Reilly MT, Faulkner GJ, Dubnau J, Ponomarev I, Gage FH (2013) The role of transposable elements in health and diseases of the central nervous system. J Neurosci 33(45):17577–17586
- <span id="page-14-30"></span>148. Remely M, Stefanska B, Lovrecic L, Magnet U, Haslberger AG (2015) Nutriepigenomics: the role of nutrition in epigenetic control of human diseases. Curr Opin Clin Nutr Metab Care 18(4):328–333
- <span id="page-14-6"></span>149. Renton AE, Chio A, Traynor BJ (2014) State of play in amyotrophic lateral sclerosis genetics. Nat Neurosci 17(1):17–23
- <span id="page-14-3"></span>150. Renton AE, Majounie E, Waite A, Simon-Sanchez J, Rollinson S, Gibbs JR et al (2011) A hexanucleotide repeat expansion in C9ORF72 is the cause of chromosome 9p21-linked ALS-FTD. Neuron 72(2):257–268
- <span id="page-14-32"></span>151. Reul JM (2014) Making memories of stressful events: a journey along epigenetic, gene transcription, and signaling pathways. Front Psychiatry 5:5
- <span id="page-14-34"></span>152. Reul JM, Chandramohan Y (2007) Epigenetic mechanisms in stress-related memory formation. Psychoneuroendocrinology 32(Suppl 1):S21–25
- <span id="page-14-1"></span>153. Ringholz GM, Appel SH, Bradshaw M, Cooke NA, Mosnik DM, Schulz PE (2005) Prevalence and patterns of cognitive impairment in sporadic ALS. Neurology 65(4):586–590
- <span id="page-14-20"></span>154. Rinn JL, Chang HY (2012) Genome regulation by long noncoding RNAs. Annu Rev Biochem 81:145–166
- <span id="page-14-11"></span>155. Rose NR, Klose RJ (2014) Understanding the relationship between DNA methylation and histone lysine methylation. Biochim Biophys Acta 1839 12:1362–1372
- <span id="page-14-19"></span>156. Ruby JG, Jan CH, Bartel DP (2007) Intronic microRNA precursors that bypass Drosha processing. Nature 448(7149):83–86
- <span id="page-14-28"></span>157. Russ J, Liu EY, Wu K, Neal D, Suh E, Irwin DJ et al (2015) Hypermethylation of repeat expanded C9orf72 is a clinical and molecular disease modifier. Acta Neuropathol 129(1):39–52
- <span id="page-14-29"></span>158. Ryu H, Smith K, Camelo SI, Carreras I, Lee J, Iglesias AH et al (2005) Sodium phenylbutyrate prolongs survival and regulates expression of anti-apoptotic genes in transgenic amyotrophic lateral sclerosis mice. J Neurochem 93(5):1087–1098
- <span id="page-14-36"></span>159. Saeed M, Siddique N, Hung WY, Usacheva E, Liu E, Sufit RL et al (2006) Paraoxonase cluster polymorphisms are associated with sporadic ALS. Neurology 67(5):771–776
- <span id="page-14-17"></span>160. Sanchez-Elsner T, Gou D, Kremmer E, Sauer F (2006) Noncoding RNAs of trithorax response elements recruit *Drosophila* Ash1 to Ultrabithorax. Science 311(5764):1118–1123
- <span id="page-14-18"></span>161. Schwartz JC, Younger ST, Nguyen NB, Hardy DB, Monia BP, Corey DR et al (2008) Antisense transcripts are targets for activating small RNAs. Nat Struct Mol Biol 15(8):842–848
- <span id="page-14-12"></span>162. Seisenberger S, Peat JR, Hore TA, Santos F, Dean W, Reik W (2013) Reprogramming DNA methylation in the mammalian life cycle: building and breaking epigenetic barriers. Philos Trans R Soc Lond B Biol Sci 368(1609):20110330
- <span id="page-14-2"></span>163. Seltman RE, Matthews BR (2012) Frontotemporal lobar degeneration: epidemiology, pathology, diagnosis and management. CNS Drugs 26(10):841–870
- <span id="page-14-13"></span>164. Sherwani SI, Khan HA (2015) Role of 5-hydroxymethylcytosine in neurodegeneration. Gene 570(1):17–24
- <span id="page-14-22"></span>165. Siomi MC, Sato K, Pezic D, Aravin AA (2011) PIWI-interacting small RNAs: the vanguard of genome defence. Nat Rev Mol Cell Biol 12(4):246–258
- <span id="page-14-14"></span>166. Sun W, Zang L, Shu Q, Li X (2014) From development to diseases: the role of 5hmC in brain. Genomics 104(5):347–351
- <span id="page-14-8"></span>167. Suzuki MM, Bird A (2008) DNA methylation landscapes: provocative insights from epigenomics. Nat Rev Genet 9(6):465–476
- <span id="page-14-27"></span>168. Suzuki N, Maroof AM, Merkle FT, Koszka K, Intoh A, Armstrong I et al (2013) The mouse C9ORF72 ortholog is enriched in neurons known to degenerate in ALS and FTD. Nat Neurosci 16(12):1725–1727
- <span id="page-14-37"></span>169. Szczygielski J, Mautes A, Steudel WI, Falkai P, Bayer TA, Wirths O (2005) Traumatic brain injury: cause or risk of Alzheimer's disease? A review of experimental studies. J Neural Transm 112(11):1547–1564
- <span id="page-14-15"></span>170. Szulwach KE, Li X, Li Y, Song CX, Wu H, Dai Q et al (2011) 5-hmC-mediated epigenetic dynamics during postnatal neurodevelopment and aging. Nat Neurosci 14(12):1607–1616
- <span id="page-14-16"></span>171. Tahiliani M, Koh KP, Shen Y, Pastor WA, Bandukwala H, Brudno Y et al (2009) Conversion of 5-methylcytosine to 5-hydroxymethylcytosine in mammalian DNA by MLL partner TET1. Science 324(5929):930–935
- <span id="page-14-26"></span>172. Therrien M, Rouleau GA, Dion PA, Parker JA (2013) Deletion of C9ORF72 results in motor neuron degeneration and stress sensitivity in *C. elegans*. PLoS ONE 8(12):e83450
- <span id="page-14-24"></span>173. Toivonen JM, Manzano R, Olivan S, Zaragoza P, Garcia-Redondo A, Osta R (2014) MicroRNA-206: a potential circulating biomarker candidate for amyotrophic lateral sclerosis. PLoS ONE 9(2):e89065
- <span id="page-14-23"></span>174. Tremolizzo L, Messina P, Conti E, Sala G, Cecchi M, Airoldi L et al (2014) Whole-blood global DNA methylation is increased in amyotrophic lateral sclerosis independently of age of onset. Amyotroph Lateral Scler Frontotemporal Degener 15(1–2):98–105
- <span id="page-14-33"></span>175. Tsankova NM, Berton O, Renthal W, Kumar A, Neve RL, Nestler EJ (2006) Sustained hippocampal chromatin regulation in a mouse model of depression and antidepressant action. Nat Neurosci 9(4):519–525
- <span id="page-14-38"></span>176. Valdmanis PN, Kabashi E, Dyck A, Hince P, Lee J, Dion P et al (2008) Association of paraoxonase gene cluster polymorphisms with ALS in France, Quebec, and Sweden. Neurology 71(7):514–520
- <span id="page-14-5"></span>177. van Blitterswijk M, DeJesus-Hernandez M, Rademakers R (2012) How do C9ORF72 repeat expansions cause amyotrophic lateral sclerosis and frontotemporal dementia: can we learn from other noncoding repeat expansion disorders? Curr Opin Neurol 25(6):689–700
- <span id="page-14-25"></span>178. van Blitterswijk M, Gendron TF, Baker MC, DeJesus-Hernandez M, Finch NA, Brown PH et al (2015) Novel clinical associations with specific C9ORF72 transcripts in patients with repeat expansions in C9ORF72. Acta Neuropathol 130(6):863–876
- <span id="page-14-7"></span>179. van Es MA, Van Vught PW, Blauw HM, Franke L, Saris CG, Andersen PM et al (2007) ITPR2 as a susceptibility gene in sporadic amyotrophic lateral sclerosis: a genome-wide association study. Lancet Neurol 6(10):869–877
- <span id="page-14-9"></span>180. Varriale A (2014) DNA methylation, epigenetics, and evolution in vertebrates: facts and challenges. Int J Evol Biol 2014:475981
- 181. Varriale A, Bernardi G (2006) DNA methylation and body temperature in fishes. Gene 385:111–121
- <span id="page-14-10"></span>182. Varriale A, Bernardi G (2006) DNA methylation in reptiles. Gene 385:122–127
- <span id="page-15-11"></span>183. Vasanthakumar A, Godley LA (2015) 5-hydroxymethylcytosine in cancer: significance in diagnosis and therapy. Cancer Genet 208(5):167–177
- <span id="page-15-9"></span>184. Veerappan CS, Sleiman S, Coppola G (2013) Epigenetics of Alzheimer's disease and frontotemporal dementia. Neurotherapeutics 10(4):709–721
- <span id="page-15-3"></span>185. Voinnet O, Baulcombe DC (1997) Systemic signalling in gene silencing. Nature 389(6651):553
- <span id="page-15-8"></span>186. Wassenegger M, Heimes S, Riedel L, Sanger HL (1994) RNAdirected de novo methylation of genomic sequences in plants. Cell 76(3):567–576
- <span id="page-15-0"></span>187. Xi Z, Rainero I, Rubino E, Pinessi L, Bruni AC, Maletta RG et al (2014) Hypermethylation of the CpG-island near the C9orf72 G(4)C(2)-repeat expansion in FTLD patients. Hum Mol Genet 23(21):5630–5637
- <span id="page-15-7"></span>188. Xi Z, van Blitterswijk M, Zhang M, McGoldrick P, McLean JR, Yunusova Y et al (2015) Jump from pre-mutation to pathologic expansion in C9orf72. Am J Hum Genet 96(6):962–970
- <span id="page-15-6"></span>189. Xi Z, Yunusova Y, van Blitterswijk M, Dib S, Ghani M, Moreno D et al (2014) Identical twins with the C9orf72 repeat expansion are discordant for ALS. Neurology 83(16):1476–1478
- <span id="page-15-1"></span>190. Xi Z, Zhang M, Bruni AC, Maletta RG, Colao R, Fratta P et al (2015) The C9orf72 repeat expansion itself is methylated in ALS and FTLD patients. Acta Neuropathol 129(5):715–727
- <span id="page-15-2"></span>191. Xi Z, Zinman L, Moreno D, Schymick J, Liang Y, Sato C et al (2013) Hypermethylation of the CpG island near the G4C2 repeat in ALS with a C9orf72 expansion. Am J Hum Genet 92(6):981–989
- <span id="page-15-4"></span>192. Yang Y, Gozen O, Vidensky S, Robinson MB, Rothstein JD (2010) Epigenetic regulation of neuron-dependent induction of astroglial synaptic protein GLT1. Glia 58(3):277–286
- <span id="page-15-10"></span>193. Zeier Z, Esanov R, Belle KC, Volmar CH, Johnstone AL, Halley P et al (2015) Bromodomain inhibitors regulate the C9ORF72 locus in ALS. Exp Neurol 271:241–250
- <span id="page-15-5"></span>194. Zhang Z, Almeida S, Lu Y, Nishimura AL, Peng L, Sun D et al (2013) Downregulation of microRNA-9 in iPSC-derived neurons of FTD/ALS patients with TDP-43 mutations. PLoS ONE 8(10):e76055