

Patients with *IDH1* wild type anaplastic astrocytomas exhibit worse prognosis than *IDH1*-mutated glioblastomas, and *IDH1* mutation status accounts for the unfavorable prognostic effect of higher age: implications for classification of gliomas

Christian Hartmann · Bettina Hentschel · Wolfgang Wick · David Capper · Jörg Felsberg · Matthias Simon · Manfred Westphal · Gabriele Schackert · Richard Meyermann · Torsten Pietsch · Guido Reifenberger · Michael Weller · Markus Loeffler · Andreas von Deimling

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Abstract WHO grading of human brain tumors extends beyond a strictly histological grading system by providing a basis predictive for the clinical behavior of the respective neoplasm. For example, patients with glioblastoma WHO grade IV usually show a less favorable clinical course and receive more aggressive first-line treatment than patients with anaplastic astrocytoma WHO grade III. Here we provide evidence that the *IDH1* status is more prognostic for overall survival than standard histological criteria that differentiate high-grade astrocytomas. We sequenced the isocitrate dehydrogenase 1 gene (*IDH1*) at codon 132 in 382 patients with anaplastic astrocytoma and glioblastoma from

the NOA-04 trial and from a prospective translational cohort study of the German Glioma Network. Patients with anaplastic astrocytomas carried *IDH1* mutations in 60%, and patients with glioblastomas in 7.2%. *IDH1* was the most prominent single prognostic factor (RR 2.7; 95% CI 1.6–4.5) followed by age, diagnosis and *MGMT*. The sequence from more favorable to poorer outcome was (1) anaplastic astrocytoma with *IDH1* mutation, (2) glioblastoma with *IDH1* mutation, (3) anaplastic astrocytoma without *IDH1* mutation and (4) glioblastoma without *IDH1* mutation ($p < 0.0001$). In this combined set of anaplastic astrocytomas and glioblastomas both, *IDH1* mutation and

C. Hartmann · D. Capper · A. von Deimling (✉)
Department of Neuropathology, Institute of Pathology,
Ruprecht-Karls-University Heidelberg, Im Neuenheimer Feld
220/221, 69120 Heidelberg, Germany
e-mail: andreas.vondeimling@med.uni-heidelberg.de

C. Hartmann · W. Wick · A. von Deimling
Clinical Cooperation Unit Neuropathology, German Cancer
Research Center, 69120 Heidelberg, Germany

B. Hentschel · M. Loeffler
Institute for Medical Informatics, Statistics and Epidemiology,
Universität Leipzig, Härtelstr. 16-18, 04107 Leipzig, Germany

W. Wick
Department of Neurooncology, Neurology,
Ruprecht-Karls-University Heidelberg,
Im Neuenheimer Feld 400, 69120 Heidelberg, Germany

J. Felsberg · G. Reifenberger
Department of Neuropathology, Heinrich Heine University,
Moorenstr. 5, 40225 Düsseldorf, Germany

M. Simon
Department of Neurosurgery, University of Bonn,
Sigmund Freud Str. 25, 53105 Bonn, Germany

M. Westphal
Department of Neurosurgery, University Hamburg-Eppendorf,
Martinistr. 52, 20251 Hamburg, Germany

G. Schackert
Department of Neurosurgery, University Dresden,
Fetscherstr. 74, 01307 Dresden, Germany

R. Meyermann
Brain Research Institute, Eberhard Karls-University Tübingen,
Calwerstr. 3, 72076 Tübingen, Germany

T. Pietsch
Department of Neuropathology, University of Bonn,
Sigmund-Freud-Str. 25, 53105 Bonn, Germany

M. Weller
Department of Neurology, University Hospital Zurich,
Frauenklinikstr. 26, 8091 Zurich, Switzerland

IDH1 expression status were of greater prognostic relevance than histological diagnosis according to the current WHO classification system. Our data indicate that much of the prognostic significance of patient age is due to the predominant occurrence of *IDH1* mutations in younger patients. Immunohistochemistry using a mutation-specific antibody recognizing the R132H mutation yielded similar results. We propose to complement the current WHO classification and grading of high-grade astrocytic gliomas by the *IDH1* mutation status and to use this combined histological and molecular classification in future clinical trials.

Keywords Grading · Classification · Anaplastic astrocytoma · Glioblastoma · *IDH1* mutation · *MGMT* · Age · Immunohistochemistry · Prognosis

Introduction

WHO classification and grading of human brain tumors is a dynamic classification system undergoing regular updating. Classification of brain tumors relies on the presumed recognition of cell lineages giving rise to specific tumor entities. WHO classification of brain tumors distinguishes diffuse astrocytomas and glioblastomas among astrocytic tumors on grounds of their predominant astrocytic differentiation. Grading of astrocytomas relies on an evaluation of anaplasia, dedifferentiation and malignancy. The WHO criteria for anaplasia in diffuse astrocytomas include increased cellularity, distinct nuclear atypia and mitotic activity. Glioblastomas are defined as anaplastic, cellular gliomas composed of poorly differentiated, often pleomorphic astrocytic tumor cells with marked nuclear atypia and brisk mitotic activity. Prominent microvascular proliferation and/or necrosis are essential diagnostic features. Malignancy, however, extends beyond the term of anaplasia because it evokes clinical parameters such as tumor recurrence and clinical outcome. The WHO classification of brain tumors has emphasized a malignancy scale underlying its brain tumor grading rather than a purely histology based algorithm [11]. Thus, the identification of glioma subgroups exhibiting significant differences in clinical outcome within an established WHO tumor entity would necessitate an adjustment of the WHO grading system. At present, the distinction between anaplastic astrocytoma and glioblastoma has clinical implications regarding the choice of treatments with more aggressive treatments for glioblastoma. Hence, novel prognostic factors may influence clinical decision making regarding treatment.

Cytosolic isocitrate dehydrogenase 1 (*IDH1*) mutations were initially detected in a fraction of glioblastomas [14] followed by the observation that they are present in the majority of diffuse astrocytomas, oligodendrogliomas and

mixed oligoastrocytomas of WHO grades II and III [1, 7, 9, 22, 26]. Consistently, the *IDH1* mutation rate is also high in secondary glioblastoma developing from previously diagnosed astrocytoma. While approximately 70% of the diffuse astrocytomas and secondary glioblastomas carry *IDH1* mutations, this alteration is observed in less than 10% of primary glioblastomas. These observations so far provide the strongest molecular evidence for different origins of primary glioblastoma and secondary glioblastoma [13]. On the other hand, the occurrence of *IDH1* mutations in the majority of both diffuse astrocytomas and oligodendroglial tumors requires reevaluation of the relation between these tumor entities. This contrasts findings suggesting a separation of astrocytic and oligodendroglial tumors on grounds of characteristic genomic alterations, mainly *TP53* mutations in diffuse astrocytoma and combined 1p/19q losses in oligodendroglial tumors [10]. These observations indicate that *IDH1* mutational status may be a marker distinguishing tumor lineages and point to a common origin of diffuse astrocytoma including secondary glioblastomas and oligodendroglial tumors. Determination of *IDH1* mutation frequency in these tumors was followed by clinical analyses revealing that this mutation constitutes a favorable prognostic factor for both patients with anaplastic astrocytoma [25] and glioblastoma [23]. In fact, the prognostic power of presence or absence of this mutation exceeded that of other markers such as *MGMT* promoter methylation status.

The role of *IDH1* mutations in tumorigenesis is under intense investigation. These mutations, which nearly always affect codon 132 in gliomas and which nearly always occur in a heterozygous manner leaving one parental allele unaffected, strongly compromise the ability of the enzyme to decarboxylate isocitrate to α -ketoglutarate and to generate NADPH [9, 26]. In contrast, mutated *IDH1* protein gains a novel function enabling the conversion of α -ketoglutarate to 2-hydroxyglutarate in a NADPH-consuming manner [5, 21]. While 2-hydroxyglutarate appears to increase the levels of reactive oxygen species, its role for tumor development is not clear. The mutation causes reduced catalytic generation of α -ketoglutarate, which combined with its additional consumption due to the gained function may inhibit prolyl hydroxylases thereby resulting in activation of the transcription factor, hypoxia-inducible factor (HIF) [27].

The objective of our present study was to investigate whether the *IDH1* mutation status provides an essential contribution to delineate the individual prognosis in patients with anaplastic astrocytoma or primary glioblastoma, respectively, and to assess whether the WHO grading system should be amended in this regard. For this purpose, we assembled a large cohort of anaplastic astrocytoma and glioblastoma patients who were treated and prospectively

followed up within the NOA-04 trial or the German Glioma Network (GGN). The prognostic impact of *IDH1* mutation and *MGMT* promoter methylation in these patients was assessed in relation to the histological classification. Based on our results, we propose a refinement of the current WHO classification of high-grade astrocytic gliomas that considers the *IDH1* mutation status in addition to the classic histological parameters.

Materials and methods

Patients, clinical and molecular data and immunohistochemistry

A group of 382 patients with anaplastic astrocytoma ($n = 145$) or primary glioblastoma ($n = 237$) forms the basis of this study. Clinical and molecular data are derived from 94 patients with anaplastic astrocytoma treated in the NOA-04 trial [25] and from 51 patients with anaplastic astrocytoma and 237 patients with glioblastoma included in the prospective translational cohort study of the GGN [23]. None of the glioblastoma patients in this study had a history of previous manifestation as diffuse astrocytoma WHO grade II or anaplastic astrocytoma and all patients from both studies were centrally reviewed by the same neuropathologist (T.P.) at the German Brain Tumor Reference Center of the German Society for Neuropathology and Neuroanatomy (DGNN) according to the revised WHO 2000 and 2007 classifications [11], assuring identical histological classification and grading. Data from both studies were compiled in a joint database and centrally analyzed. Data from the NOA-04 trial [25] and data from the ongoing translational study of the GGN [23] have been published previously.

The methods and conditions for determining the mutational status of *IDH1* by direct sequencing of PCR products has been described previously [23, 25]. Methods and conditions for detecting the *IDH1R132H* mutation by IHC with mouse monoclonal antibody H09 (Dianova, catalog number DIA H09, Hamburg, Germany) on an automated immunostainer (BenchMark, Ventana Medical Systems, Tuscon, AZ, USA) have been described in detail elsewhere [3, 4]. Analysis for *MGMT* promoter methylation by methylation-specific PCR (MSP) was performed as previously described [23, 25].

Patients in this study were recruited between January 2004 and August 2008. The GGN data were used with a cut-off value for last information of 31 December 2009. The extent of resection was assessed by early (<72 h) magnetic resonance imaging (MRI) or computed tomography (CT) but was not centrally reviewed. Complete resection was scored whenever no residual tumor was

detected; subtotal resection corresponded to removal of 50–99% of the tumor and partial resection to removal of <50% of the tumor. Overall survival (OS) was the primary endpoint of most analyses which is most appropriate for this retrospective analysis of pooled patients from a clinical trial (NOA-04) and a large cohort study (GGN). Data on postsurgical first-line treatment and treatment at first recurrence of the patients were collected. The median observation time for the entire cohort was 41 months.

Statistics

The association of clinical data and molecular markers were tested by χ^2 test and Fisher's exact test. Predictive values, sensitivity and specificity with 95% CI were determined to assess the validity of IHC to *IDH1* mutations in comparison to DNA sequencing as gold standard. OS, the primary endpoint, was calculated from the day of first surgery until death or end of follow up. Logrank test was used to compare outcome data. Cox regression models were fitted to assess the independent impact of the *IDH1* mutation (mutated vs. wild type), adjusting for age (>60 vs. ≤ 60), diagnosis (anaplastic astrocytoma vs. glioblastoma), *MGMT* status (methylated yes vs. not methylated), and extent of resection (total vs. not total). Interaction between *IDH1* mutation and histological diagnosis as well *MGMT* promoter methylation and histological diagnosis were evaluated in multivariate models. Data were analyzed by PASW Statistics 18 (Version 18.0.0) and StatXact-8 (Cytel Studio Version 8.0.0).

Results

Patient characteristics

Patient characteristics are given in Table 1. As expected, patients with anaplastic astrocytoma were 17 years younger (median) than patients with glioblastoma, had better performance status, experienced somewhat less radical surgery and less frequently received combined modality treatment as first-line treatment. Hence any analysis of prognostic factors needed adjustment for these imbalances in order to separate the contribution of each single factor.

IDH1 sequencing and immunohistochemical data

IDH1 mutations were detected in 104 of 382 high-grade malignant astrocytic gliomas. In 145 anaplastic astrocytomas, we found 87 mutations (60.0%) with 78 of the R132H, 6 of the R132C, 2 of the R132S and 1 of the R132G types. In 237 glioblastomas, we found 17 *IDH1* mutations (7.2%), which were all of the R132H type. All

Table 1 Patient characteristics

	A III <i>n</i> = 145 (100%)	GBM <i>n</i> = 237 (100%)
Age in years (median, range)	43.7 (18.3–77.8)	60.9 (19.2–86.5)
Gender (male/female)	89/56	144/93
Extent of resection <i>n</i> (%)		
Complete	48 (33.1%)	112 (47.3%)
Subtotal (50–99%)	53 (36.6%)	81 (34.2%)
Partial (<50%)	43 (29.7%)	23 (9.7%)
Biopsy	1 (0.7%)	21 (8.9%)
KPS (≤ 70 / > 70)	10/127	51/181
First-line treatment		
RT alone	63 (43.4%)	60 (25.3%)
RT and CT (alkylating agents)	29 (20.0%)	164 (69.2%)
CT alone (alkylating agents)	49 (33.8%)	10 (4.2%)
None	4 (2.8%)	3 (1.3%)
Treatment at first recurrence		
RT alone	42 (29.0%)	2 (0.8%)
RT and CT (alkylating agents)	1 (0.7%)	15 (6.3%)
CT alone (alkylating agents)	45 (31.0%)	54 (22.8%)
Other CT +/- RT	0	12 (5.1%)
None	57 (39.3%)	154 (65.0%)
<i>IDH1</i> mutated	87 (60.0%)	17 (7.2%)
≤ 60 years	78 (53.8%)	15 (6.2%)
> 60 years	9 (6.2%)	2 (1.0%)
<i>MGMT</i> promoter methylated	58/105 (55.2%)	110/233 (47.2%)
≤ 60 years	50 (47.6%)	53 (22.7%)
> 60 years	8 (7.6%)	57 (24.5%)
<i>IDH1</i> and <i>MGMT</i> (<i>n</i> = 338)	<i>n</i> = 105	<i>n</i> = 233
<i>IDH1</i> wt and <i>MGMT</i> meth –	28 (26.7%)	118 (50.6%)
<i>IDH1</i> wt and <i>MGMT</i> meth +	17 (16.2%)	98 (42.1%)
<i>IDH1</i> mut and <i>MGMT</i> meth –	19 (18.1%)	5 (2.1%)
<i>IDH1</i> mut and <i>MGMT</i> meth +	41 (39.0%)	12 (5.2%)

RT radiotherapy, CT chemotherapy, wt wild type, mut mutated, meth – unmethylated, meth + methylated, A III anaplastic astrocytoma WHO grade III, GBM glioblastoma WHO grade IV

mutations in anaplastic astrocytomas and glioblastomas were heterozygous. *IDH2* mutations were analyzed in 365 patients of the 382 patients. Only a single glioblastoma patient carried an *IDH2* R172K mutation.

A total of 197 patients consisting of 80 anaplastic astrocytomas and 117 glioblastomas were analyzed by IHC with monoclonal antibody H09 specific for mutant *IDH1* protein encoded by the *IDH1R132H* mutation. We detected mutant *IDH1* protein in 40 anaplastic astrocytomas and in 19 glioblastomas. Seven mutations that were recognized by sequencing were not detected by the mutation-specific antibody. These included five patients with four R132C and one R132S mutations who were not recognized by the mutation-specific antibody and two patients who were

sequenced with the *IDH1R132H* mutation but did not exhibit antibody binding. On the other hand, three patients were detected with the antibody but had escaped initial sequencing due to a signal below threshold. However, re-sequencing confirmed the mutation. Thus, all IHC-positive patients were confirmed by sequencing.

Hence, the positive predictive value (PPV) of IHC for *IDH1* R132H mutations is 100% (95% CI 93.4–100), while the negative predictive value of a negative IHC for excluding *IDH1* mutations is 94.9% (95% CI 89.8–97.4). Sensitivity is 89.4% (95% CI 79.4–95.6) and specificity 100% (95% CI 97.2–100). This implies that IHC is highly predictive of the sequencing result and can therefore be used as a very powerful surrogate marker of the *IDH1* sequence status.

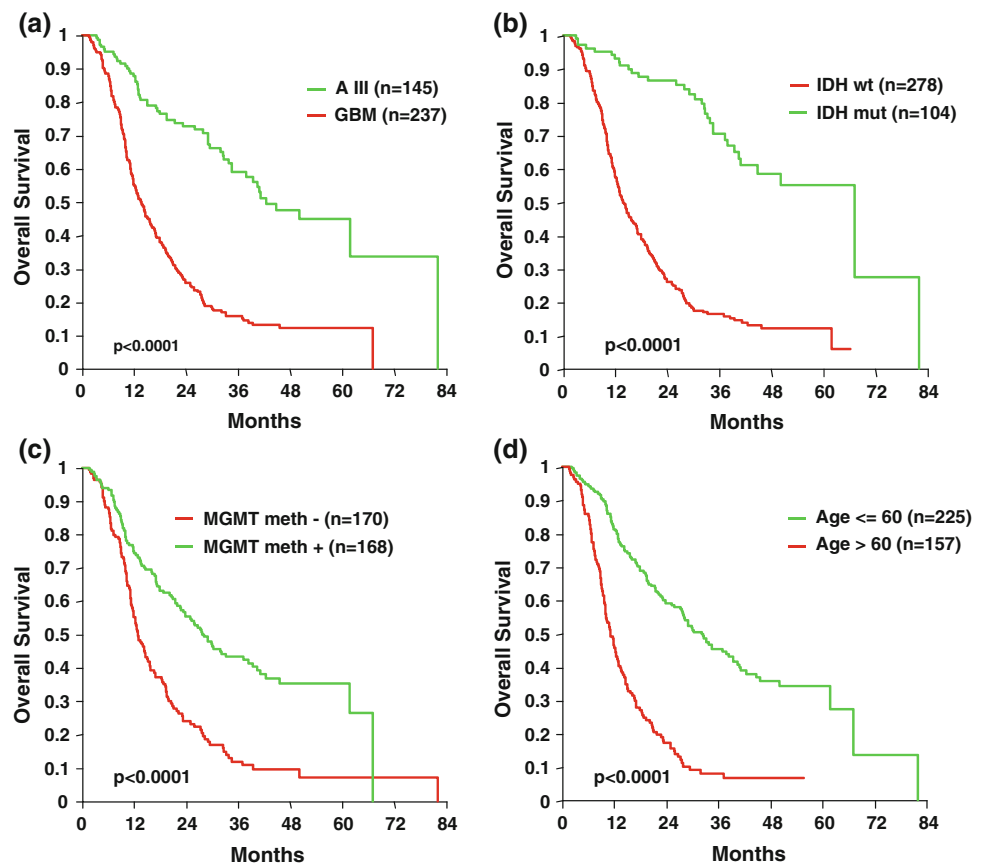
MGMT promoter methylation

The *MGMT* promoter methylation status was determined in 338 patients including 105 anaplastic astrocytoma and 233 glioblastoma patients. Among anaplastic astrocytoma patients, 58 exhibited *MGMT* promoter methylation while 47 did not. Among glioblastomas, 110 exhibited a methylated *MGMT* promoter while 123 did not. *MGMT* and *IDH1* status in the 105 anaplastic astrocytomas and 233 glioblastomas are given in Table 1. This table also provides the relationships between molecular findings for both genes and patient age groups. It is noteworthy that *MGMT* promoter methylation was more frequent in *IDH1* mutated tumors (69%, 53/77) than in wild type-*IDH1*-tumors (44%, 115/261), irrespective of the histological diagnosis ($p = 0.0001$).

Survival (univariate analysis)

Figure 1 shows OS estimates in a univariate breakdown regarding the main factors diagnosis, age, *IDH1*- and *MGMT* status. Patients with anaplastic astrocytoma lived significantly longer than patients with glioblastoma ($p < 0.0001$) (Fig. 1a). The median OS was 42.5 months (95% CI 31.9–53.2) for anaplastic astrocytoma and 13.6 months (95% CI 11.6–15.6) for glioblastoma. Patients with *IDH1* mutations lived longer than patients without ($p < 0.0001$) (Fig. 1b), Patients with *MGMT* promoter methylation lived longer than patients without ($p < 0.0001$) (Fig. 1c) and furthermore, we confirmed younger age at diagnosis as a significant prognostic factor associated with longer OS ($p < 0.0001$) (Fig. 1d). When we calculated the relative risks (RR) for these pair-wise comparisons, we found the largest risk for *IDH1* wild type versus mutant (RR = 5.1; 95% CI 3.5–7.5) followed by glioblastoma versus anaplastic astrocytoma (RR = 3.4; 95% CI 2.5–4.6), old versus young age (RR = 3.3; 95% CI

Fig. 1 Kaplan–Meier plots of the series of 382 anaplastic astrocytomas and glioblastomas showing the association with overall survival of histological WHO diagnosis (a), *IDH1* mutation status (b), *MGMT* methylation status (c) and age (d)



2.6–4.3) and *MGMT* non-methylated versus methylated (RR = 2.3; 95% CI 1.7–3). These findings are already indicative of a strong prognostic contribution of the *IDH1* mutation status. However, these univariate contrasts are confounded by the unbalanced compositions of the groups. Hence, multivariate modeling is required to adjust for these effects.

Survival (multivariate analysis)

Figures 2, 3, 4 provide insights into the multivariate structure of the survival data. For this purpose, we separated the OS estimates in subgroups using two (or three) main factors. Co-evaluation of diagnosis and *MGMT* status showed that both factors contribute to prognosis. *MGMT* splits the cohort into the most favorable clinical course for 47 anaplastic astrocytoma patients with *MGMT* promoter methylation, followed by 58 anaplastic astrocytoma patients without *MGMT* promoter methylation, followed by 110 glioblastoma patients with and trailed by 123 glioblastoma patients without *MGMT* promoter methylation (Fig. 2a). There is also an indication that the prognostic split associated with *MGMT* is slightly more pronounced in patients with glioblastoma compared to patients with anaplastic astrocytoma (see below).

Co-evaluation of diagnosis and age indicates that both factors contribute to prognosis. Anaplastic astrocytoma patients aged 60 years or younger had the most favorable clinical course. The poorest survival occurred in glioblastoma patients older than 60 years. The age-effect seems to be more pronounced among anaplastic astrocytoma patients (Fig. 2b).

Co-evaluation of histological diagnosis and *IDH1* status showed that both factors contribute to prognosis. The most favorable OS was found in 87 anaplastic astrocytoma patients with *IDH1* mutation, followed by 17 glioblastoma patients with *IDH1* mutation ($p = 0.014$). The difference between the 17 glioblastoma patients with *IDH1* mutation and 58 anaplastic astrocytoma patients without *IDH1* mutation was not significant ($p = 0.222$). However, the Kaplan–Meier plots indicate a trend towards an inferior OS for patients with anaplastic astrocytoma patients without *IDH1* mutation. These patients showed a longer OS than 220 glioblastoma patients without *IDH1* mutation ($p = 0.003$). The differences in OS across the whole cohort were highly significant ($p < 0.0001$) (Fig. 3a). The separation of these four groups became even more prominent when PFS was analyzed instead of OS (Fig. 3b), but, as indicated above, PFS may not be an appropriate endpoint for a pooled analysis of a clinical trial where single

Fig. 2 Kaplan–Meier plot of the series of 338 anaplastic astrocytomas and glioblastomas showing the association with overall survival of the combination of WHO diagnosis and *MGMT* methylation status (a). Association of overall survival with the effect of the combination of WHO diagnosis and age are shown on the series of 382 patients (b)

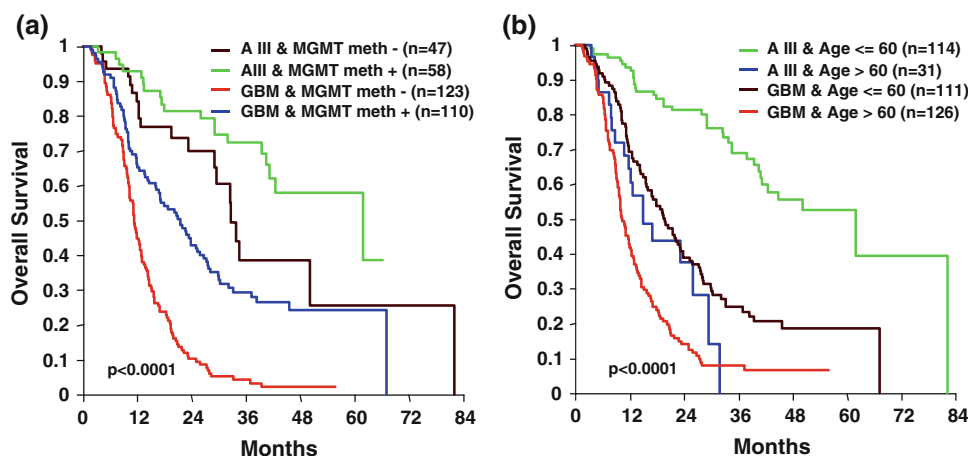
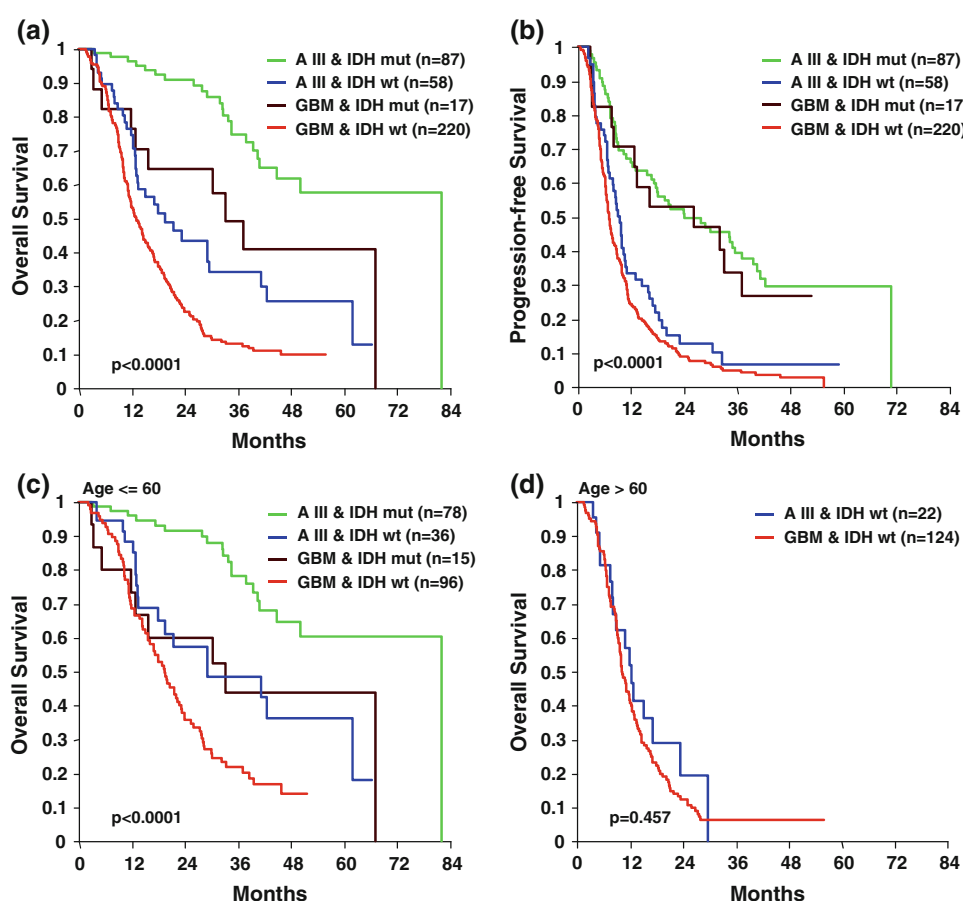


Fig. 3 Kaplan–Meier plot of the series of 382 anaplastic astrocytomas and glioblastomas patients showing the association of *IDH1* status with overall survival (a) and progression-free survival (b). The same analysis regarding overall survival is shown for patients aged 60 years or younger (c). For patients older than 60 years, overall survival only for *IDH1* wild type is shown (d)



modality treatment was the rule (NOA-04) and a cohort where single modality treatment was more likely given to patients with a more malignant tumor with less favorable prognostic factors (GGN).

An analysis restricted to patients aged 60 years or younger again showed best OS in 78 anaplastic astrocytomas with *IDH1* mutation. Worst outcome was seen in 96 glioblastoma patients without *IDH1* mutation ($p < 0.0001$) (Fig. 3c). If we restricted the analysis to

patients older than 60 years, we find *IDH1* mutations only rarely ($n = 2$ for glioblastoma, $n = 9$ for anaplastic astrocytoma) and these curves are not shown. However, we found that there was no apparent prognostic difference for anaplastic astrocytoma and glioblastoma patients without *IDH1* mutation ($p = 0.457$) (Fig. 3d). Hence, elderly anaplastic astrocytoma patients without *IDH1* mutation seem to have the same poor prognosis as glioblastoma patients.

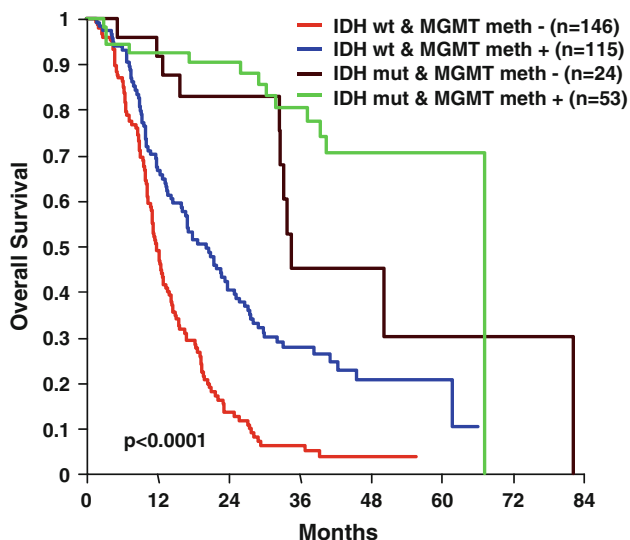


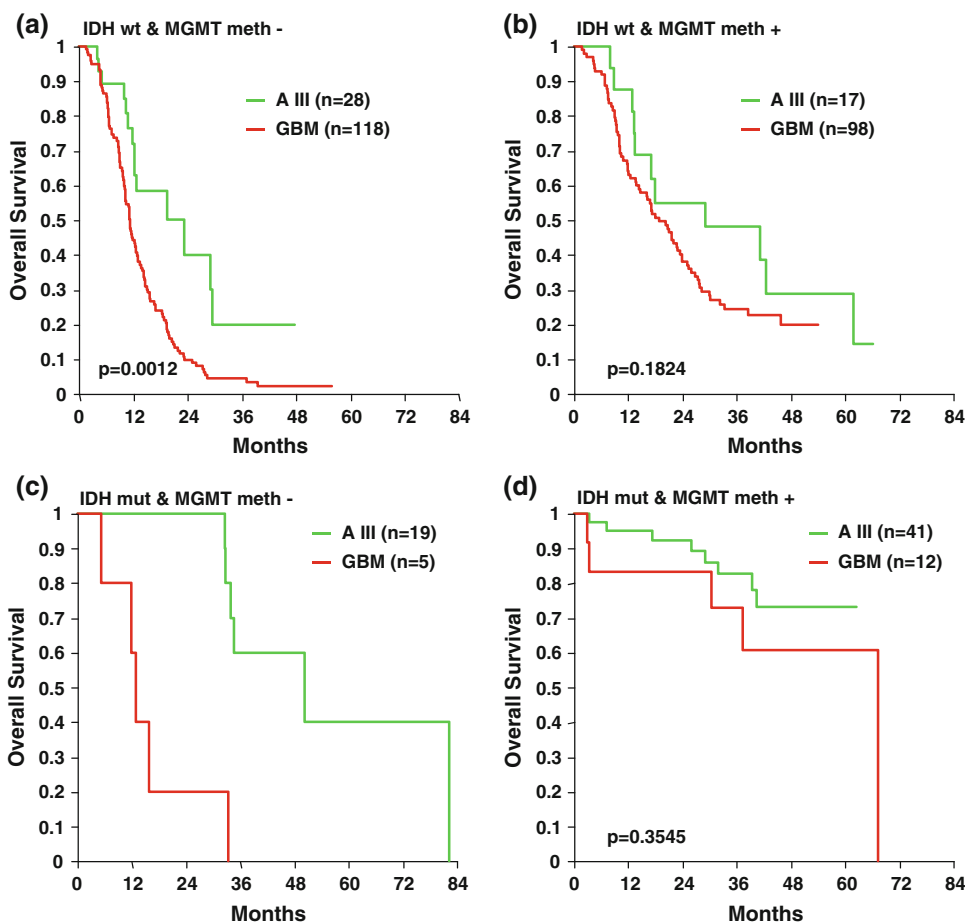
Fig. 4 Kaplan–Meier plot of the series of 338 anaplastic astrocytomas and glioblastomas showing the combined associations of *IDH1* and *MGMT* status with overall survival

Co-evaluation of *IDH1* and *MGMT* status revealed that *MGMT* permits to separate the prognostic difference by *IDH1* mutations further. However, patients with *IDH1*

mutation performed better than patients without *IDH1* mutation irrespective of *MGMT* status. Patients without *IDH1* mutation but with *MGMT* promoter methylation fared worse and poorest survival was seen in patients showing neither *IDH1* mutation nor *MGMT* promoter methylation (Fig. 4).

Figure 5 shows the co-evaluation of the three main factors diagnosis, *IDH1* and *MGMT* status. Clearly the dominant prognostic factors are related to the molecular status. There is a very poor prognostic group without *IDH1* mutation and with unmethylated *MGMT* at one end of the spectrum (with few long-term survivors) and a small but very favorable group of *IDH1* mutant and *MGMT*-methylated patients (among whom long-term survival seems frequent). In most of the four groups, WHO diagnosis seemed to retain a small influence on OS. Among patients without *IDH1* mutation and without *MGMT* promoter methylation, the 28 patients with anaplastic astrocytoma fared better ($p = 0.0012$) than the 118 with glioblastoma (Fig. 5a). There was no significant difference ($p = 0.1824$) in survival between 17 anaplastic astrocytoma and 98 glioblastoma patients without *IDH1* mutation and with *MGMT* promoter methylation (Fig. 5b). Similarly, 41 anaplastic astrocytoma patients with *IDH1* mutation and

Fig. 5 Kaplan–Meier plots of OS of anaplastic astrocytoma and glioblastoma patients stratified according to histological diagnosis within the individual groups defined by the *IDH1* mutation status in combination with the *MGMT* promoter methylation status



with *MGMT* promoter methylation did not live significantly longer ($p = 0.3534$) than 12 glioblastoma patients with the same molecular findings (Fig. 5d). The group defined by patients with *IDHI* mutation and without *MGMT* promoter methylation included only 19 anaplastic astrocytoma and 5 glioblastoma patients and, therefore, was too small to yield conclusive results (Fig. 5c).

Multivariate modeling

Multivariate modeling was undertaken to estimate the RR for age, extent of resection, diagnosis, *IDHI* mutation status and *MGMT* status. The results for the main effects are given in Table 2. *IDHI* mutation was the dominant prognostic factor with a RR of 2.7 (95% CI 1.6–4.5). Histology and *MGMT* status were also prominent with RR of 2.2, respectively. Since RR can be multiplied, it becomes evident that this implies a very powerful system spanning a RR range of about 13 in total ($2.7 \times 2.2 \times 2.2$).

Finally, in a sensitivity analysis, we also introduced two interaction terms into the main effects model. The model fit (i.e. log-likelihood) improved significantly. This model provides an indication that the *IDHI* effect is quantitatively more pronounced in anaplastic astrocytoma (anaplastic astrocytoma: RR = 4.1; glioblastoma: RR = 1.8) while the

MGMT effect is more pronounced in glioblastoma (anaplastic astrocytoma: RR = 1.3; glioblastoma: RR = 2.5).

Discussion

Classification and grading of high-grade astrocytic gliomas according to WHO is accompanied by a considerable interobserver variation [19]. Reasons are guidelines allowing for subjective interpretation. On the one hand, this may be due to a subjective interpretation of borderline histological features by different neuropathologists. On the other hand, malignant gliomas are regionally heterogeneous tumors and incomplete tissue sampling thus may lead to the underestimation of a tumor's true malignancy grade when important histological features of anaplasia are restricted to focal areas and not represented in the evaluable tissue specimens. Molecular analyses are expected to reduce diagnostic interobserver variation due to providing clear yes/no answers in many instances and detecting molecular changes present in all tumor cells, thereby reducing the problems of subjectivity in histological assessment and incomplete tissue sampling. In the present study, we focus on the role of the *IDHI* status in assisting classification and grading of anaplastic astrocytomas and

Table 2 Multivariate models

	Model with main effects only			Model: main effects and interactions with diagnosis		
	Relative risk	95% CI	<i>p</i> value	Relative risk	95% CI	<i>p</i> value
Age		1.6–2.8	<0.001	Age	1.7–2.9	<0.001
≤60	1			≤60	1	
>60	2.2			>60	2.2	
Resection		0.5–0.9	0.009	Resection	0.5–0.9	0.011
No total	1			No total	1	
Total	0.7			Total	0.7	
Diagnosis		1.5–3.2	<0.001	Diagnosis	1.1–6.5	0.024
A III	1			A III	1	
GBM	2.2			GBM	2.7	
<i>IDHI</i> general		1.6–4.5	<0.001	<i>IDHI</i> in A III	Test for interaction: $p = 0.099$	
mut	1			mut	1	
wt	2.7			wt	4.1	
				<i>IDHI</i> in GBM		
				mut	1	
				wt	1.8	
<i>MGMT</i> general		1.7–2.9	<0.001	<i>MGMT</i> in A III	Test for interaction: $p = 0.067$	
meth +	1			meth +	1	
meth –	2.2			meth –	1.3	
				<i>MGMT</i> in GBM		
				meth +	1	
				meth –	2.5	

wt wild type, mut mutant, meth – unmethylated, meth + methylated, A III anaplastic astrocytoma WHO grade III, GBM glioblastoma WHO grade IV

glioblastomas. The data set analyzed here was obtained by joining the primary data sets of the anaplastic astrocytoma patients treated within the NOA-04 randomized phase III trial [25], the glioblastoma cohort of the GGN [23], and a new subset of anaplastic astrocytomas from the GGN. In our series, 87/145 (60%) anaplastic astrocytomas and 17/137 (7.2%) glioblastomas carried a mutation in *IDH1*. The frequency of *IDH1* mutations in anaplastic astrocytomas in our series is within the spectrum described in previous series ranging from 52 [9] to 78% [22]. The frequency of *IDH1* mutation detected in glioblastomas has generally been reported to range below 10% in primary glioblastoma [1, 9, 22, 26].

The impact of *IDH1* mutations on clinical outcome has been demonstrated in prospective clinical studies including anaplastic astrocytomas and glioblastomas [20, 23, 25]. It has also been detected in various retrospective series including diffuse gliomas of WHO grades II, III and IV recently reviewed [15]. There is consensus on patients with *IDH1* mutations performing better than those without [6, 9, 14, 17, 26]. Our data provide compelling evidence that the *IDH1* status separates anaplastic astrocytomas and glioblastomas in two sub-entities each with significantly different clinical outcomes. Further, our data indicate a close clinical and biological relation of anaplastic astrocytomas and glioblastomas with *IDH1* mutation versus anaplastic astrocytomas and glioblastomas without *IDH1* mutations. The sequence from more favorable to poorer outcome was (1) *IDH1* mutant anaplastic astrocytoma, (2) *IDH1* mutant glioblastoma, (3) *IDH1* wild type anaplastic astrocytoma and (4) *IDH1* wild type glioblastoma, respectively. The missing significance between groups 2 and 3 ($p = 0.222$) might be due to the low number of patients with *IDH1* mutant glioblastoma ($n = 17$). The prognostic relevance of *IDH1* status in comparison with histopathological evaluation of high-grade astrocytomas is demonstrated in Figs. 1a, b, 5 and particularly in the multivariate modeling result.

Using the *IDH1* mutation status as a separator to this predefined set of tumors, we were able to separate two patient groups with even more pronounced differences in median OS than those stratified according to conventional histological features. Thus, *IDH1* analysis in this set of tumors is a more powerful prognostic marker than current WHO classification and grading for high-grade astrocytomas. Notably, *IDH1* wild type anaplastic astrocytoma patients demonstrated not only shorter OS when compared to *IDH1* mutant anaplastic astrocytoma patients but even survived shorter than patients with *IDH1* mutant glioblastoma.

Apart from the WHO diagnosis, patient age has been established as a powerful parameter for the prognosis of patients with anaplastic astrocytomas and glioblastomas [2]. Therefore, we also examined the relationship between

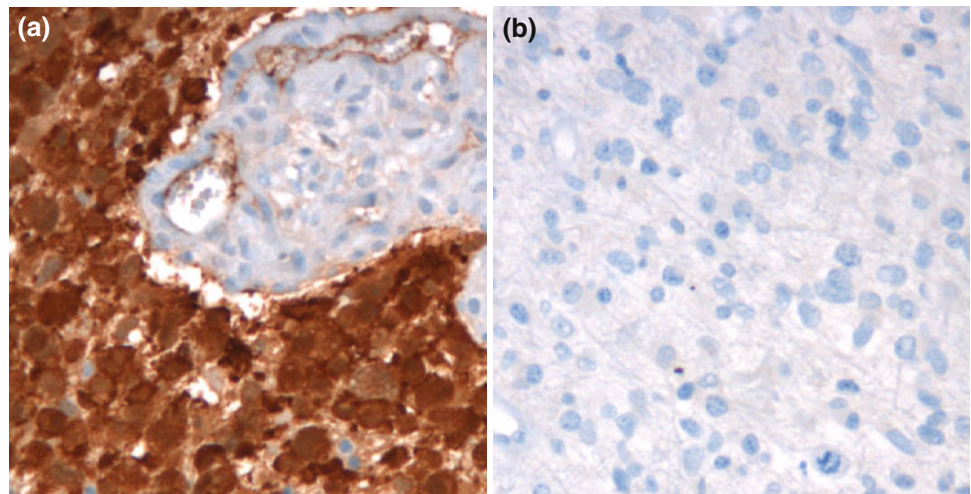
IDH1 status and patient age. Analysis in patients aged 60 years or younger or over 60 revealed that in the younger age group, anaplastic astrocytoma patients without *IDH1* mutations exhibited OS very similar to that of glioblastoma patients with *IDH1* mutations (Fig. 3c). Importantly, in patients over 60 years the absence of *IDH1* mutation in anaplastic astrocytoma was indicative of poor OS that was similar to the OS of older patients with *IDH1* wild type glioblastoma (Fig. 3c). This may be of relevance for treatment decisions.

A yet unsolved issue relates to the impact of *IDH2* mutation on OS. So far, none of the previous series provided reliable data on the clinical relevance of *IDH2* mutations alone [20]. *IDH2* mutations occur infrequently in astrocytic gliomas. In a large series including astrocytomas, oligodendrogliomas and oligoastrocytomas, the frequency of *IDH2* mutations was 3.1%. Of note, *IDH2* mutations clustered in patients with oligodendroglial tumors but were found in <1% of astrocytomas [7]. We examined 132 anaplastic astrocytoma patients and 233 glioblastoma patients from the present series for *IDH2* mutation in codon 172 and detected only a single R172K mutation in a glioblastoma patient. Because *IDH2* mutations are so rare in anaplastic astrocytomas and glioblastomas, this molecular alteration was not further considered in the clinical correlations performed in the present study.

MGMT promoter methylation status has been demonstrated as a powerful prognostic and predictive marker for patients with glioblastomas [8, 24]. Its relevance was confirmed in our series to be independent of the *IDH1* status (Figs. 2a, 5). For all four groups, stratified for diagnosis and presence or absence of *IDH1* mutation, OS was longer in patients with *MGMT* promoter methylation. These differences were significant for glioblastomas with or without *IDH1* mutation, and for anaplastic astrocytomas with *IDH1* mutation. In contrast, the *MGMT* status was not associated with differential survival in anaplastic astrocytomas without *IDH1* mutation. Interestingly, the rate of patients with *MGMT* methylation was higher in both anaplastic astrocytomas and glioblastomas with *IDH1* mutation than in anaplastic astrocytomas and glioblastomas without *IDH1* mutation. Upon sorting our series of 338 patients into four groups defined by combined *IDH1* and *MGMT* status, it became evident that the histological distinction between anaplastic astrocytomas and glioblastomas had only a moderate effect on OS (Fig. 5a–d). Noteworthy is the very similar gradient of the curves for both histological diagnoses within each group. This analysis underscores the importance of combined *IDH1* and *MGMT* analysis to predict survival in these patients.

We recently developed a monoclonal mouse antibody (m*IDH1*R132H/clone H09) to mutant *IDH1* protein of the R132H type [3, 4]. The R132H mutation constitutes >90% of the *IDH1* mutations seen in gliomas [7]. Therefore, we

Fig. 6 Representative images of a primary glioblastoma with *IDH1*R132H mutation detected by immunohistochemistry with monoclonal antibody H09 (**a left**), and of an anaplastic astrocytoma without *IDH1* mutation (**b right**). Original magnification $\times 200$



performed immunohistochemistry with this antibody on 197 tumors of our series, which revealed high positive and negative predictive values of H09 IHC for detecting *IDH1* mutation, thus corroborating this method as a surrogate approach to DNA sequencing. Representative data for IHC with mouse monoclonal antibody H09 is shown in Fig. 6. Interestingly, we detected in three samples mutant *IDH1* protein of the R132H type that showed no *IDH1* mutation by initial sequencing. However, these *IDH1* mutations were confirmed by re-sequencing. On the other hand, two *IDH1* R132H mutations were not detected by the m*IDH1*R132H/clone H09 antibody. These findings show that both methods do not have a 100% detection rate but that detection rate is comparable.

Based on the presented data we propose to consider a refinement of the current WHO classification and grading system for anaplastic astrocytomas and glioblastomas by subdividing each tumor entity into two molecularly and clinically distinct subgroups according to the *IDH1* mutation status. This implies that this molecular parameter will become part of the tumor classification and thus needs to be determined on a regular basis. In detail, *IDH1* mutant anaplastic astrocytomas should be separated from *IDH1* wild type anaplastic astrocytomas, and likewise, glioblastomas with and without *IDH1* mutation should be distinguished. It remains to be seen whether further molecular subdivision of glioblastomas according to the *MGMT* promoter methylation status will be applicable in the routine diagnostic setting. In contrast to the technically simple and reliable detection of *IDH1* mutations by DNA sequencing or immunohistochemistry, current methods for the molecular analysis of *MGMT* promoter methylation are technically more sophisticated, difficult to standardize, and thus suffer from considerable interlaboratory variability [15]. We expect an extended sub-classification of anaplastic astrocytomas and glioblastomas to be of immediate clinical impact. For example, our data clearly indicate that

patients with *IDH1* wild type anaplastic astrocytomas have poor clinical outcome. Thus, one could consider treating these patients more aggressively with first-line combined radiochemotherapy corresponding to the current standard of care for glioblastoma treatment. On the other hand, the rare *IDH1* mutant primary glioblastomas may require less aggressive first-line treatment similar to that usually administered to anaplastic astrocytoma patients, that is, either radiotherapy or alkylating chemotherapy, but no combined modality treatment [25]. As *IDH1* mutant glioblastomas usually affect younger patients and are associated with a higher likelihood of long-term survival, a less aggressive up-front treatment may be beneficial also in terms of reducing treatment-associated neurotoxicity in these patients. It is not yet clear whether anaplastic astrocytomas without *IDH1* mutation generally should be considered as underdiagnosed glioblastomas. While such an approach would yield a tighter correlation of diagnosis and OS than the current procedure, it does not account for the still better OS of anaplastic astrocytoma patients without *IDH1* mutation than glioblastoma patients without *IDH1* mutations. We also assume that our data will have major impact on reporting results from clinical trials and on the study designs for novel trials focusing on anaplastic astrocytomas and glioblastomas. The circumstance that the presence of *IDH1* mutations is the strongest indicator for more favorable clinical outcome will make it inevitable to determine this parameter in order to test the influence of the experimental treatment applied. Furthermore, this molecular parameter likely will have an impact on the inclusion criteria of future trials.

How could the WHO classification be developed in the future with regard to the optimal prognostic and clinically most helpful stratification of patients with high-grade astrocytic gliomas? There are several possibilities for this separation, such as the simple addition of the *IDH1* status to the histological diagnosis to indicate that the respective

tumor is associated with a more or less favorable clinical course. A more radical approach would be to disregard necrosis and microvascular proliferation as the basis for distinction between anaplastic astrocytomas and glioblastomas, and substitute these histological parameters by the *IDH1* mutation status, that is high-grade astrocytic tumors with *IDH1* mutation would be termed anaplastic astrocytomas while all high-grade astrocytic tumors without *IDH1* mutation would be termed glioblastomas. Such a procedure would result in a significantly tighter association of grading with clinical course and, therefore, would better suit the intention of glioma grading to provide a prognostically meaningful stratification that may guide the postoperative treatment. It would emphasize on biological properties inherent of distinct tumor cell lineages, i.e. astrocytoma deriving from a tumor precursor cell characterized by *IDH1* mutation. However, it would also break with a long and well received tradition of purely histological grading by devaluating the prognostic importance of necrosis or microvascular proliferation. In fact, this approach would miss some prognostically relevant information, as indicated by the survival differences between anaplastic astrocytomas and glioblastomas within the group of *IDH1* wild type tumors. *IDH1* mutation is frequent in secondary glioblastomas derived by progression from astrocytomas. It has been suggested that *IDH1* mutant primary glioblastoma in fact represent secondary glioblastoma that rapidly evolved from lower grade precursor tumors and thus have not been diagnosed before [13]. Thus, secondary glioblastomas and *IDH1* mutant primary glioblastomas could be the endpoints of a disease following the same pathogenetic pathway. This hypothesis is supported by similar chromosomal and genetic aberration profiles in these two types of glioblastomas [13, 18] and might be taken into account by terming all *IDH1* mutant glioblastomas secondary glioblastomas, thereby distinguishing them from the more common primary glioblastomas that are *IDH1* wild type. However, there undoubtedly is a fraction of true secondary glioblastomas that progressed from a lower grade astrocytoma but lack *IDH1* mutation, consistent with the lack of *IDH1* mutations in approximately 20% of lower grade astrocytomas [1, 9, 22, 26]. These tumors argue against redefining the clinically established term of secondary glioblastoma. Another alternative might be to use the traditional designation “anaplastic astrocytoma WHO grade III” only for those anaplastic astrocytomas that carry an *IDH1* mutation, while the term “anaplastic astrocytoma WHO grade IV” may be used for *IDH1* wild type anaplastic astrocytomas. However, this designation is problematic as glioblastoma is traditionally regarded as the only type of astrocytic glioma corresponding to WHO grade IV. Furthermore, the WHO grade IV group would then cover a spectrum of malignant gliomas variably defined on histological and/or molecular

features, and associated with different prognoses, i.e. *IDH1* mutant and wild type glioblastomas, glioblastomas with oligodendroglial component or “anaplastic oligoastrocytoma WHO grade IV” [16], which show an intermediate prognosis between anaplastic oligoastrocytoma and glioblastoma [12], and *IDH1* wild type anaplastic astrocytoma. Thus, for the time being and for reasons of simplicity, we propose to continue using the conventional histological terms of anaplastic astrocytoma and glioblastoma, respectively, supplemented by the *IDH1* mutation status, and alert the physicians treating the patients concerning the prognostic implications of this molecular marker.

In conclusion, we demonstrate that the *IDH1* mutation status distinguishes anaplastic astrocytomas and glioblastomas into clinically meaningful prognostically distinct subgroups that possibly require different first-line treatment. Therefore, determination of *IDH1* status is essential for a comprehensive neuropathological assessment of high-grade astrocytic gliomas and should be considered in the design and evaluation of future clinical trials. The differential distribution of *IDH1* mutations across age groups largely explains the prognostic impact of age in high-grade astrocytoma patients. We propose to revise the current WHO classification of these tumors by inclusion of the *IDH1* mutation status, which can be easily and reliably assessed by DNA sequencing or IHC.

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References

- Balss J, Meyer J, Mueller W et al (2008) Analysis of the *IDH1* codon 132 mutation in brain tumors. *Acta Neuropathol* 116:597–602
- Burger PC, Vogel FS, Green SB et al (1985) Glioblastoma multiforme and anaplastic astrocytoma, pathologic criteria and prognostic implications. *Cancer* 56:1106–1111
- Capper D, Weißert S, Balss J et al (2010) Characterization of R132H mutation specific *IDH1* antibody binding in brain tumors. *Brain Pathol* 20:245–254

4. Capper D, Zentgraf H, Balsl J et al (2009) Monoclonal antibody specific for IDH1 R132H mutation. *Acta Neuropathol* 118:599–601
5. Dang L, White DW, Gross S et al (2009) Cancer-associated IDH1 mutations produce 2-hydroxyglutarate. *Nature* 462:739–744
6. Gravendeel LA, Kloosterhof NK, Bralten LB et al (2010) Segregation of non-p.R132H mutations in IDH1 in distinct molecular subtypes of glioma. *Hum Mutat* 31:E1186–E1199
7. Hartmann C, Meyer J, Balsl J et al (2009) Type and frequency of IDH1 and IDH2 mutations are related to astrocytic and oligodendroglial differentiation and age: a study of 1010 diffuse gliomas. *Acta Neuropathol* 118:469–474
8. Hegi ME, Diserens AC, Gorlia T et al (2005) MGMT gene silencing and benefit from temozolomide in glioblastoma. *N Engl J Med* 352:997–1003
9. Ichimura K, Pearson DM, Kocalkowski S et al (2009) IDH1 mutations are present in the majority of common adult gliomas but are rare in primary glioblastomas. *Neuro Oncol* 11:341–347
10. Kraus JA, Koopmann J, Kaskel P et al (1995) Shared allelic losses on chromosomes 1p and 19q suggest a common origin of oligodendroglioma and oligoastrocytoma. *J Neuropathol Exp Neurol* 54:91–95
11. Louis D, Ohgaki H, Wiestler O et al (eds) (2007) World health organization classification of tumours of the central nervous system, 4 edn. In: Bosman F, Jaffe E, Lakhani S et al (eds) World health organization classification of tumours. IARC, Lyon
12. Miller CR, Dunham CP, Scheithauer BW et al (2006) Significance of necrosis in grading of oligodendroglial neoplasms: a clinicopathologic and genetic study of newly diagnosed high-grade gliomas. *J Clin Oncol* 24:5419–5426
13. Nobusawa S, Watanabe T, Kleihues P et al (2009) IDH1 mutations as molecular signature and predictive factor of secondary glioblastomas. *Clin Cancer Res* 15:6002–6007
14. Parsons DW, Jones S, Zhang X et al (2008) An integrated genomic analysis of human glioblastoma multiforme. *Science* 321:1807–1812
15. Riemenschneider MJ, Jeuken JW, Wesseling P et al (2010) Molecular diagnostics of gliomas: state of the art. *Acta Neuropathol* 120:567–584
16. Scheithauer BW, Fuller GN, VandenBerg SR (2008) The 2007 WHO classification of tumors of the nervous system: controversies in surgical neuropathology. *Brain Pathol* 18:307–316
17. Sonoda Y, Kumabe T, Nakamura T et al (2009) Analysis of IDH1 and IDH2 mutations in Japanese glioma patients. *Cancer Sci* 100:1996–1998
18. Toedt G, Barbus S, Wolter M et al (2010) Molecular signatures classify astrocytic gliomas by IDH1 mutation status. *Int J Cancer* (epub ahead of print)
19. van den Bent MJ (2010) Interobserver variation of the histopathological diagnosis in clinical trials on glioma: a clinician's perspective. *Acta Neuropathol* 120:297–304
20. van den Bent MJ, Dubbink HJ, Marie Y et al (2010) IDH1 and IDH2 mutations are prognostic but not predictive for outcome in anaplastic oligodendroglial tumors: a report of the European Organization for Research and Treatment of Cancer Brain Tumor Group. *Clin Cancer Res* 16:1597–1604
21. Ward PS, Patel J, Wise DR et al (2010) The common feature of leukemia-associated IDH1 and IDH2 mutations is a neomorphic enzyme activity converting alpha-ketoglutarate to 2-hydroxyglutarate. *Cancer Cell* 17(3):225–234
22. Watanabe T, Nobusawa S, Kleihues P et al (2009) IDH1 mutations are early events in the development of astrocytomas and oligodendrogliomas. *Am J Pathol* 174:653–656
23. Weller M, Felsberg J, Hartmann C et al (2009) Molecular predictors of progression-free and overall survival in patients with newly diagnosed glioblastoma: a prospective translational study of the German Glioma Network. *J Clin Oncol* 27:5743–5750
24. Weller M, Stupp R, Reifenberger G et al (2010) MGMT promoter methylation in malignant gliomas: ready for personalized medicine? *Nat Rev Neurol* 6:39–51
25. Wick W, Hartmann C, Engel C et al (2009) NOA-04 randomized phase III trial of sequential radiochemotherapy of anaplastic glioma with procarbazine, lomustine, and vincristine or temozolomide. *J Clin Oncol* 27:5874–5880
26. Yan H, Parsons DW, Jin G et al (2009) IDH1 and IDH2 mutations in gliomas. *N Engl J Med* 360:765–773
27. Zhao S, Lin Y, Xu W et al (2009) Glioma-derived mutations in IDH1 dominantly inhibit IDH1 catalytic activity and induce HIF-1alpha. *Science* 324:261–265