Claudia Penna Daniele Mancardi Francesca Tullio Pasquale Pagliaro

# Postconditioning and intermittent bradykinin induced cardioprotection require cyclooxygenase activation and prostacyclin release during reperfusion

Received: 27 July 2007

Returned for 1. Revision: 6 August 2007 1. Revision received: 16 November 2007 Returned for 2. Revision: 22 November 2007 2. Revision received: 23 November 2007 Accepted: 27 November 2007 Published online: 9 January 2008

C. Penna · D. Mancardi · F. Tullio Dr. P. Pagliaro (⊠) Dipartimento di Scienze Cliniche e Biologiche Università di Torino Regione Gonzole 10 10043 Orbassano (TO), Italy Tel.: +39-11/6705430 ext -5450 Fax: +39-11/9038639 E-Mail: pasquale.pagliaro@unito.it **Abstract** Postconditioning (PostC), obtained with brief intermittent cycles of ischemia alternating with reperfusion applied after the ischemic event, has been shown to reduce infarct size. Recently, we have shown that PostC triggering includes  $B_2$  receptor activation and its downstream pathway. Moreover, we showed that BK intermittent infusion induces a cardioprotection similar to PostC. The aim of this study was to investigate the involvement of cyclooxygenase-(COX)-derivated prostaglandins, such as prostacyclin (PGI<sub>2</sub>) pathway in the cardioprotective action mediated by intermittent BK infusion.

Isolated rat hearts underwent 30 min ischemia and 120 min reperfusion. Myocardial damage was evaluated using nitro-blue-tetrazolium staining. The production of metabolite of PGI2, 6-keto-PGF1a, was evaluated with EIA assay on the samples collected during reperfusion. The perfusion pressure and the left ventricular pressure were monitored. In Control hearts, the infarct size was  $64\% \pm 4\%$  of risk area. PostC reduced significantly the infarct size (28%  $\pm$  4% *P* < 0.001 Vs. Control). BK intermittent protocol to mimic PostC, attenuated infarct size  $(40\% \pm 2\% P < 0.01$  Vs. Control). The BK-intermittent and PostC protections were abolished with COX-inhibition. Intermittent BK and PostC enhanced the release of prostacyclin metabolite, 6-keto-PGF1a, in the late phase of reperfusion (i.e., 6-keto-PGF1 $\alpha$  peaked 30 min after protective maneuvers). Also the stable PGI<sub>2</sub> analogue, Iloprost, given in the early reperfusion reduced infarct size and improved post-ischemic heart function. In conclusion, protection by PostC and intermittent BK requires COX activation and PGI<sub>2</sub> release during late reperfusion. These data suggest that COX must not be inhibited to have PostC protection. This finding should be kept present by future clinical studies on PostC.

**Key words** bradykinin B<sub>2</sub> receptors – ischemia/reperfusion – postconditioning – prostacyclin – rat

# Introduction

BRC

It has been reported that Postconditioning (PostC), i.e., *repetitive cycles* of reperfusion and coronary

occlusion following an ischemic insult, cause massive salvage of the myocardium similar to that of ischemic preconditioning (IP) [35, 36, 38]. Moreover, PostC involves signal transduction pathways which are similar to those seen in IP [12, 16, 23-26, 31-40].

Virtually in all of the species in which different PostC protocols have been tested they have been protective, including humans [e.g., 4, 31, 32, 38], with the exception of recent works conducted on a rodent model [5]. In this respect, it is intriguing that PostC protection is present in wildtype mice [2, 13] and it is not lost in connexin 43-deficient mice [13]; yet an age-related loss of PostC protection in mice has been described, which seems to be related to a reduced level of signal transducer and activator of transcription 3 (STAT3) with increasing age [2]. Remote ischemic PostC has been also induced by cycles of few minutes of ischemia/ reperfusion applied to a distal artery territory (femoral or renal artery) [15, 40]. Importantly, the protection by postconditioning represent a long-term protective effect and not a mere attenuation of event involved in early reperfusion injury [21]. Therefore, postconditioning has impressive clinical potential and to precisely clarify the involved pathway signaling is of paramount importance.

Recently, we have demonstrated that the intermittent infusion of bradykinin (BK) at the beginning of reperfusion can trigger PostC-like cardioprotection against infarct size via  $B_2$  BK receptors activation [24].

Cyclooxygenase enzymes (COX) are constitutively expressed in most tissues, including myocardium [10, 11]. It is reported that  $B_2$  BK receptors activation results in de novo synthesis of prostacyclin (PGI<sub>2</sub>) [34], which has been demonstrated to attenuate ischemia-reperfusion injury [4]. In particular, Berti et al. [1] and Rossoni et al. [28] have reported that both indomethacin and aspirin aggravated ischemiainduced ventricular dysfunction in perfused rabbit hearts and this was associated with inhibition of PGI<sub>2</sub> production in the cardiac tissue. Yet, either administration of the stable PGI<sub>2</sub> analogue Iloprost [7] or prostaglandin  $E_1$  [8] during ischemia have been seen to attenuate myocardial stunning in open chest dogs.

It seems that  $PGI_2$  is produced by jeopardized myocardium [33] and that the rate of its formation increases particularly during the first 5–10 min of reperfusion declining thereafter [1, 6]. Endogenous prostaglandins are also involved in ischemic preconditioning [10].

Therefore, we hypothesized that intermittent BK may enhance  $PGI_2$  release during reperfusion and that COX activation is involved in PostC.

To verify the role of COX pathway in the cardioprotection by PostC and intermittent BK, we studied the effect of COX inhibition during PostC maneuvers and during the intermittent infusion of BK. Moreover, we studied the effects of the administration of the stable  $PGI_2$  analogue Iloprost during early reperfusion. Finally, we measured the release of 6-ketoPGF1 $\alpha$ , a  $PGI_2$  metabolite, which is used to evaluate the activation of  $B_2$  BK receptor, during reperfusion.

# Materials and methods

Male Wistar rats (n = 105; body weight 450–550 g) received humane care in compliance with Italian law (DL-116, 27 Jan. 1992) and in accordance with the Guide for the Care and Use of Laboratory Animals published by the US National Institutes of Health (NIH Publication No. 85-23, revised 1996).

#### Isolated heart perfusion

The methods were similar to those previously described [22–25]. In brief, each animal was anesthetized. The chest was opened 10 min after heparin treatment and the heart was rapidly excised. Isolated rat hearts were weighed, attached to the perfusion apparatus and retrogradely perfused with oxygenated Krebs–Henseleit buffer (127 mM NaCl, 17.7 mM NaHCO<sub>3</sub>, 5.1 mM KCl, 1.5 mM CaCl<sub>2</sub>, 1.26 mM MgCl<sub>2</sub>, 11 mM D-glucose and gassed with 95% O<sub>2</sub> and 5% CO<sub>2</sub>). A constant flow was adjusted with a proper pump to obtain a typical coronary perfusion pressure (PP) of 80–85 mmHg during the initial part of stabilization. Thereafter the same flow level (9  $\pm$  1 ml/min/g) was maintained throughout the experiment.

A small hole in the left ventricular wall allowed drainage of the thebesian flow, and a polyvinyl-chloride balloon was placed into the left ventricle and connected to an electromanometer for the recording of left ventricular pressure (LVP). The balloon was filled with saline to achieve an end-diastolic LVP of 5 mmHg. Coronary perfusion pressure, coronary flow and LVP were monitored to asses the preparation conditions. LVP was used to calculate the positive  $(dP/dt_{max})$  and negative  $(dP/dt_{min})$  first derivative of pressure during systole and diastole, respectively.

The hearts were electrically paced at 280 bpm and kept in a temperature-controlled chamber (37°C).

All chemicals were purchased from Sigma (USA).

#### Experimental protocols (Fig. 1)

Each heart was allowed to stabilize for 20 min. After the stabilization period, hearts were subjected to a specific protocol, which included in all groups a 30 min of global no-flow ischemia. A period of 120min reperfusion followed the 30 min ischemia in all groups (see below). Pacing was discontinued on initiation of ischemia and restarted after the third minute of reperfusion in all groups [16, 22–25].

Group 1: Control								
Stabilization	Ischemia	Reperfusion						
Group 2: PostC								
Stabilization	Ischemia	Reperfusion						
Groups 3 and 7: Intermittent BK or Intermittent ILOPROST								
Stabilization	Ischemia	Reperfusion						
Groups 4 and 5 : PostC + INDO or Intermittent BK + INDO								
Stabilization	Ischemia	Reperfusion						
Group 6: INDO								
Stabilization	Ischemia	Reperfusion						
Group 8: ILOPROST Continous 3 min								
Stabilization	Ischemia	Reperfusion						

**Fig. 1** Experimental design. The isolated, Langendorff-perfused hearts were stabilized for 20 min, and then subjected to 30 min of normothermic global ischemia followed by 120 min of reperfusion. *PostC* postconditioning; *BK* Bradykinin; *INDO* Indomethacin; *ILOPROST* analogous of prostacyclin. For further explanation see text

Experimental protocols are described in Fig. 1.

After stabilization the hearts of the *Control* group (Group 1, n = 20) were exposed to 30 min ischemia and then to 120 min reperfusion only.

In Group 2 (*PostC* group; n = 20) after the 30 min ischemia, the hearts immediately underwent a protocol of PostC. This consisted of five cycles of 10 s reperfusion and 10 s global ischemia [23–25].

In Group 3 the intermittent infusion of BK (*Intermittent BK* + *intermittent buffer*; n = 15) was performed after 30 min ischemia, this intermittent protocol consisted of five cycles of 10 s of oxygenated/ non-oxygenated buffer with BK (100 nM) [24, 27].

To evaluate the role of COX pathway hearts were co-infused with an inhibitor of COX, *Indomethacin* (INDO, 10  $\mu$ M) [27] during the initial 3 min of reperfusion, either during the PostC maneuvers (Group 4, n = 10) or during intermittent BK protocol (Group 5, n = 10). To determine the effects of the antagonist, in Group 6 (*INDO group*, n = 10) indomethacin was only infused during the initial 3 min of reperfusion. Moreover, in Group 7 Iloprost (12 nM), an analogous of PGI<sub>2</sub> [8, 10], was infused intermittently in lieu of BK (*Intermittent ILOPROST* + *intermittent buffer*; n = 10) after the 30 min ischemia, as in Group 3; in Group 8 Iloprost (12 nM) was infused in continuous (*Continuous ILOPROST*; n = 10) for 3 min immediately after the 30 min ischemia.

Assessment of myocardial injury

Infarct areas were assessed at the end of the experiment as previously described [22–25]. In brief, immediately after reperfusion each heart was rapidly removed from the perfusion apparatus and the left ventricle (LV) was dissected into 2–3 mm circumferential slices. Following 20 min of incubation at 37°C in 0.1% solution of nitro-blue tetrazolium in phosphate buffer, unstained necrotic tissue was carefully separated from stained viable tissue by an independent observer. The necrotic mass was expressed as a percentage of total left ventricular mass. In fact, though in this model the whole heart underwent normothermic ischemia, only the LV had a fixed volume and pre-load; therefore only the LV mass was considered as a risk area [22–25].

#### 6-keto-PGF1α assay

In the hearts of Group 1, Group2 and Group 3, the PGI<sub>2</sub>-releasing capacity of the isolated heart was measured by evaluating the concentration of its stable metabolite 6-keto-PGF1 $\alpha$  in perfusate samples, collected before ischemia and after 3, 5, 10, 30, 60, 90 and 120 min of reperfusion. The amount of 6-keto-PGF1 $\alpha$  released was determined with an enzyme-immuno-assay kit (Amersham International plc, UK).

# Statistical analysis

All data are expressed as means  $\pm$  SEM One-way ANOVA and Newman–Keuls multiple comparison test (for post-ANOVA comparisons) have been used to compare infarct size. One-way ANOVA has been also used to compare area under the curve of 6-keto-PGF1 $\alpha$  release. Two-way ANOVA (variables treatment groups and time point) and one-way ANOVA for multiple measure (post test Newman–Keuls multiple comparison test) have been used for the analysis of PP and LVP data. A *P* value <0.05 was considered statistically significant.

#### Results

## Infarct size (Fig. 2)

Total infarct size, expressed as a percentage of left ventricular mass, was  $64\% \pm 4\%$  in Control Group 1. PostC (Group 2) significantly (P < 0.001) reduced infarct size to  $28\% \pm 4\%$  of risk area. This high infarct size in Control group and the strong protective effect of PostC are features of the constant flow model of isolated rat heart, as suggested in our previous work [23].

In intermittent BK, Group 3, the infarct size was 40%  $\pm$  2%; a value similar to that obtained with PostC and significantly (P < 0.01) lower than that of Control Group 1. The cardioprotective effect of PostC and that of intermittent-BK was abolished during the co-infusion with the COX-inhibitor, INDO. In fact, the infarct areas of Group 4 (PostC + INDO, 65%  $\pm$  2%) and Group 5 (intermittent BK + INDO, 57%  $\pm$  6%) were similar to that of the Control Group 1.

The treatment with INDO alone did not significantly change infarct size, which was  $57 \pm 6$ , i.e., similar to that of Control Group 1, and significantly higher than that of intermittent BK (P < 0.05) and PostC (P < 0.001) groups (Fig. 2).

Iloprost reduced infarct size both when infused intermittently (Group 7, Infarct size  $39 \pm 6$ , P < 0.01



**Fig. 2** Infarct size. The amount of necrotic tissue is expressed as percent of the left ventricle (*LV*), which is considered the risk area. *PostC* postconditioning; *BK* Bradykinin; *INDO* Indomethacin; *ILOPROST* analogous of prostacyclin. *NS* not significant; \*\*\*P < 0.001, \*\*P < 0.01 and \*P < 0.05 Vs. Control; #P < 0.001 Vs. PostC and NS Vs. Control. For further explanation see text

Table 1	Baseline	hemodynamic	parameters
---------	----------	-------------	------------

Vs. Control) and in continuous for three min (Group 8, infarct size  $42 \pm 7$ , P < 0.05 Vs. Control).

# Perfusion pressure and left ventricular pressure (Table 1; Figs. 3, 4, 5)

Perfusion pressure is represented as percent variation with respect to baseline level (Fig. 3). Perfusion pressure increased in all groups during reperfusion. Two-way ANOVA revealed that there is not interaction between the considered variables (treatment groups and time point), whereas both treatment groups and time point affect the results (P < 0.01 for both). The post ANOVA comparison revealed that the only group that showed a significantly (P < 0.01) blunted increase in pressure (i.e., a reduced postischemic vasoconstriction) was the Intermittent Iloprost group (Group 7).

Systolic function is represented by develop LVP and  $dP/dt_{max}$ , which are reported as percent variation with respect to baseline level in Fig. 4. For both parameters the two-way ANOVA revealed that there is not interaction between the considered variables (treatment groups and time point), whereas both treatment groups and time point affect the results (P < 0.01 for both). In reperfusion, the left ventricular function was not statistically different among groups. The post ANOVA comparison revealed that the only group that performed significantly better than Control group (P < 0.01) in terms of  $dP/dt_{max}$  (Fig. 4B) was the Intermittent Iloprost group.

*Diastolic function* is represented by diastolic LVP and  $dP/dt_{min}$ , which are reported in Fig. 5 as mmHg and as percent variation with respect to baseline level, respectively. For both parameters the two-way ANOVA revealed that there is no interaction between the considered variables (treatment groups and time point), whereas both treatment groups and time point affect the results (P < 0.01 for both). The post ANO-VA comparison revealed that in reperfusion there were groups that reached higher diastolic LVP (higher

	PP (mmHg)	LVEDP (mmHg)	Dev LVP (mmHg)	dP/dt <sub>max</sub> (mmHg/s)	dP/dt <sub>min</sub> (mmHg/s)
Control (Group 1) PostC (Group 2) Intermittent BK (Group 3) PostC + INDO (Group 4) Intermittent BK + INDO (Group 5) INDO (Group 6) Intermittent ILOPROST (Group 7) ILOPROST 3 min (Group 8)	$87 \pm 482 \pm 286 \pm 389 \pm 282 \pm 286 \pm 380 \pm 282 \pm 282 \pm 282 \pm 282 \pm 2$	$\begin{array}{l} 4.8 \pm 0.9 \\ 4.2 \pm 0.7 \\ 4.0 \pm 1.0 \\ 4.7 \pm 0.7 \\ 4.2 \pm 0.7 \\ 4.3 \pm 0.4 \\ 5.0 \pm 0.7 \\ 4.5 \pm 0.9 \end{array}$	$80 \pm 3  78 \pm 5  75 \pm 4  70 \pm 6  78 \pm 5  75 \pm 4  81 \pm 3  82 \pm 3$	$\begin{array}{l} 1950 \pm 60 \\ 2021 \pm 111 \\ 2187 \pm 121 \\ 2078 \pm 174 \\ 2019 \pm 120 \\ 2188 \pm 114 \\ 1965 \pm 89 \\ 1987 \pm 93 \end{array}$	$\begin{array}{c} -1,578 \pm 130 \\ -1,788 \pm 122 \\ -1,679 \pm 120 \\ -1,578 \pm 164 \\ -1,754 \pm 144 \\ -1,689 \pm 120 \\ -1,779 \pm 94 \\ -1,680 \pm 85 \end{array}$

PP perfusion pressure; LVEDP left ventricular end diastolic pressure; Dev LVP developed left ventricular pressure; dP/dt<sub>max</sub> maximum rate of increase of LVP during systole; dP/dt<sub>min</sub> maximum rate of decrease of LVP during diastole. Other acronyms as text

**Fig. 3** Perfusion pressure. Percent variation of perfusion pressure with respect to baseline level for each group, during the 30 min ischemia and 120 min reperfusion. Time 0 correspond to the beginning of reperfusion. *PostC* postconditioning; *BK* Bradykinin; *INDO* Indomethacin; *ILOPROST* analogous of prostacyclin. \*P < 0.01 between Control and lloprost intermittent groups. For further explanation see text



contracture) and others that reached lower levels of diastolic LVP (less contracture) than Control group. Again the group that performed significantly better than Control group (P < 0.01) in terms of both diastolic LVP (Fig. 4A) and  $dP/dt_{min}$  (Fig. 4B) was the Intermittent Iloprost group, which showed less contracture and an increased lusitropism. Yet, the group that performed worst was the INDO group, both in terms of contracture (diastolic LVP) and relaxation  $(dP/dt_{min})$ .

## **6**-keto PGF1 $\alpha$ release during reperfusion (Fig. 6)

To easily compare the three groups in which 6-keto PGF1 $\alpha$  release was measured, in Fig. 6 results are presented as normalized data. In the Control group, the release of 6-keto PGF1 $\alpha$  slightly increased during the initial part of reperfusion, then started to decline after 30 min of reperfusion, reaching a value not different from baseline level at the end of reperfusion. On the contrary, in the Intermittent BK group the release of 6-keto PGF1 $\alpha$  started to increase after BK-treatment, showing a sharp increase at 30 min reperfusion. Thereafter it slightly declined, and at the end of reperfusion was still higher than baseline. A similar trend was observed in PostC group. The analysis of the

area under the curve revealed that the Control curve was significantly lower than that of Intermittent BK curve (P < 0.05), but not different from that of PostC curve. The areas under Intermittent BK curve and PostC curve were not statistically different.

# Discussion

We have previously shown that cardioprotection triggered by postconditioning involves endogenous activation of B<sub>2</sub> receptors. Using intermittent infusion of BK in the early phase of reperfusion, we triggered cardioprotection akin to postconditioning. We demonstrated that B<sub>2</sub> receptors, nitric oxide, protein kinase G, mitochondrial K<sub>ATP</sub> channels and reactive oxygen species are also integral to the protection triggered by intermittent BK [24].

In the present study our goals were (1) to show that cardioprotection triggered by postconditioning or intermittent BK involves activation of COX; (2) to show that intermittent BK trigger the release of  $PGI_2$  and (3) to show that  $PGI_2$  analogous can mimic postconditioning.

Here, we show that the cardioprotection induced by PostC or by intermittent BK infusion is dependent **Fig. 4** Systolic function. **A** Percent variation of Developed left ventricular (*LV*) pressure with respect to baseline level for each group, during the 30 min ischemia and 120 min reperfusion. **B** Percent variation of first derivative of LV pressure during systole ( $dP/dt_{max}$ ) with respect to baseline level for each group, during the 30 min ischemia and 120 min reperfusion. Time 0 correspond to the beginning of reperfusion. *PostC* postconditioning; *BK* Bradykinin; *INDO* Indomethacin; *ILOPROST* analogous of prostacyclin. \**P* < 0.01 between Control and lloprost intermittent groups. For further explanation see text



on COX activation and augmentation of  $PGI_2$  release in the late phase of reperfusion. Moreover, Iloprost, a stable  $PGI_2$  analogous, given in early reperfusion induced a cardioprotective effect either when infused intermittently or when infused in continuous.

Regardless of the fact that protective stimuli (Intermittent BK or PostC) are applied only for few seconds at the beginning of reperfusion the endogenous release of PGI<sub>2</sub> metabolite is enhanced during the late period of reperfusion. It has been, previously, reported that during reperfusion in non-protected hearts the rate of PGI<sub>2</sub> formation increases particularly during the first 5–10 min of reperfusion declining rapidly thereafter [1, 6]. Here we confirm this finding. We suggest that the potentially protective effect of PGI<sub>2</sub> release cannot operate in naive ischemia/reperfusion because the enhanced production during the first phase of reperfusion rapidly decline in the second phase of reperfusion, when reperfusion injury occurs. On the contrary the PostC maneuvers and intermittent BK triggers the above reported protective cascade [24], and may allow COX activity and PGI<sub>2</sub> release in the late phase of reperfusion, thus preventing reperfusion injury. In other words, we suggest that PostC or Intermittent BK triggers protection, which allows COX activity and subsequent PGI<sub>2</sub> formation.

It is worth of note that the protective effect of Intermittent BK on infarct size seems to be less than that produced by PostC; yet the Intermittent BK seems to increase prostacyclin to a greater level than PostC. Although these differences are not statistically **Fig. 5** Diastolic function. **A** LV diastolic (mmHg) during the 30 min ischemia and 120 min reperfusion. **B** Percent variation of first derivative of LV pressure during diastole  $(dP/dt_{min})$  with respect to baseline level for each group, during the 30 min ischemia and 120 min reperfusion. Time 0 correspond to the beginning of reperfusion. *PostC* postconditioning; *BK* Bradykinin; *INDO* Indomethacir; *ILOPROST* analogous of prostacyclin. \*P < 0.01 between Control and Iloprost intermittent groups; #P < 0.01 between Control and INDO groups. For further explanation see text



significant, these data are in line with the fact that other factors (e.g., adenosine and opioids), other than BK and COX, are involved in the PostC-induced protection [e.g., 12, 26, 35, 39].

Moreover, Intermittent BK and Iloprost (intermittently and in continuous) achieved similar degree of protection in terms of infarct size reduction. Once again suggesting that other factors, other than BK and prostaglandin, are involved in the PostC-induced protection. The fact that Iloprost may induce protection in reperfusion is in agreement with previous studies which have shown a protective role of prostaglandins in reperfusion [7, 8, 10]. With respect to this, it is intriguing that the involvement of COX has been very recently demonstrated in the additive effect of late preconditioning and PostC protection [29]. However, the fact that Iloprost is able to induce protection when infused intermittently and in continuous in the early reperfusion, whereas the endogenous prostaglandin **Fig. 6** 6-keto PGF1 $\alpha$  release. The PGI2-releasing capacity of the isolated heart is estimated by the percent variation of the concentration of its stable metabolite 6-keto-PGF1 $\alpha$  in perfusate samples during baseline and reperfusion. Each value is represented as percent with respect to the mean baseline data of each group. Time 0 correspond to the beginning of reperfusion. *PostC* postconditioning; *BK* Bradykinin. For further explanation see text



release is enhanced in the late phase of reperfusion by PostC, suggests different mechanisms of protection by endogenous prostaglandin(s) release triggered by PostC and infusion of exogenous prostaglandin analogue. Differences might be due to molecule stability differences and/or pharmacodynamic features; further studies may clarify the reasons of these differences.

The more robust end-point in analyzing I/R injury is infarct size [9, 20]. Therefore, we focused our attention on myocardial damages evaluated by infarct size. As for preconditioning [9, 20], systolic function (developed LV pressure and  $dP/dt_{max}$ ) may not be an appropriate end-point to study PostC protection. The improvement of function during reperfusion in preconditioned rat hearts has been attributed to the reduction of adenosine release during this phase [9]. On the contrary, in post-conditioned hearts an accumulation of adenosine has been reported [16]. These differences may explain our findings of an unclear effect on systolic function by PostC treatment. Yet, it is hard to distinguish the impairment of function due to necrosis and/or to stunning of viable tissue. In order to understand the role of PostC on myocardial stunning appropriate studies are required. Moreover, it has been suggested that in rodent models contracture development, rather than systolic function, may be a more appropriate end-point to understand the protective effects of cardioprotective maneuvers [9]. In the present study the only conditions that clearly improves post-ischemic heart performance is the treatment with Intermittent Iloprost, which also improved contracture. On the other hand, the only treatment that clearly worsens diastolic heart performance is Indomethacin treatment. On the base of these effects on contracture, but keeping in mind the model limits above reported, we can argue that COX is involved not only in reducing infarct size, but also in improving heart function recovery of viable tissue. This is also in line with the reduced post-ischemic vasoconstriction (less increase in PP) observed in the present study. Moreover, the beneficial effect by Iloprost is in agreement with the involvement of endogenous prostaglandins in ischemic preconditioning [10], with the anti-stunning effect by Iloprost observed in a canine model of ischemia/reperfusion [8] and with the involvement of COX in late preconditioning, which clearly improves post-ischemic stunning [29].

Differently from previous studies [8, 10, 17, 19, 26, 30, 37], which used BK or other agents in continuous for a long period during reperfusion, we were able to reproduce PostC by means of few seconds (50 s in total) treatment with BK only when given in an intermittent manner in the very early period of reperfusion [24] and we confirm that late PGI<sub>2</sub> release is mandatory to limit reperfusion injury by PostC.

Another novelty of the present study is that the inhibitor, indomethacin, has been given for 3 min only to prevent the triggering of protection by PostC or intermittent BK, confirming that the initial phase of reperfusion is of paramount importance for triggering cardioprotective pathways. Also the stable  $PGI_2$  analogue, iloprost, is able to induce protection when applied for few minutes at the beginning of reperfusion.

Kinins are produced during ischemia reperfusion by the action of a kininogenase activated by the acidosis. Recently, PostC has been seen to induce cardioprotection by maintaining acidosis during the first minutes of reperfusion as reoxygenated myocardium produces reactive oxygen species that activate protective signaling [3]. Bradykinin was believed to be the only kinin acting on the B<sub>2</sub> receptors in rats; however, it has been demonstrated that isolated rat hearts also releases Arg-kallidin, a kallidin-like peptide, which acts on  $B_2$  receptors too [14, 18, 19]. Importantly, Arg-kallidin release increases during ischemia and mediates IP and preconditioning-like protection, via B<sub>2</sub> receptor activation, in isolated rat hearts [18, 19]. Whether the endogenous mediator of PostC protection is BK or Arg-kallidin was beyond the aims of the present study.

In conclusion, the major new findings in this study are: (1) postconditioning triggered cardioprotection involves endogenous activation of COX and release of PGI<sub>2</sub>; (2) intermittent BK infusion also triggers the COX activation and induces PGI<sub>2</sub> formation in the late phase of reperfusion; (3) exogenous infusion of the stable PGI<sub>2</sub> analogue, Iloprost, is able to induce cardioprotection when infused in the early reperfusion either continuously or intermittently.

We suggest that protection exerted by postconditioning and intermittent BK is not solely dependent on the activation of the so called RISK (reperfusion injury salvage kinase [12, 24, 25, 38], but it also depends on the late release of  $PGI_2$ . That is, during PostC maneuvers the heart may release kinins that accumulate in an intermittent manner and triggers a protective pathway that lead to a protected state, which include COX activation and  $PGI_2$  release.

# Clinical implication

One goal in applying reperfusion therapeutic strategies is to pharmacologically mimic postconditioning. It is likely that one drug or maneuver cannot target all of the mechanisms of reperfusion injury, but a combined therapy integrating pharmacological agents (e.g., BK and Iloprost) and maneuvers, such as intermittence of infusion, in the very early phase of reperfusion may provide a broader spectrum approach to attenuate reperfusion injury. From a clinical point of view, it is important that ischemia is not necessary, whereas intermittence of drug infusion is able to trigger PostC. Equally important, COX must not be inhibited to obtain cardioprotection by ischemic postconditioning during reperfusion.

**Acknowledgments** We thank Regione Piemonte; Compagnia di San Paolo"; the Consorzio Interuniversitario per le Ricerche Cardiovascolari (CIRC) and the Italian Ministry of University and Research (MIUR, PRIN). We Thank Dr. Giovanni Berta for insightful suggestions.

# References

- Berti F, Rossoni G, Magni F, Caruso D, Omini C, Puglisi L, Galli G (1988) Nonsteroidal antiinflammatory drugs aggravate acute myocardial ischemia in the perfused rabbit heart: a role for prostacyclin. J Cardiovas Pharmacol 12:438-444
- 2. Boengler K, Buechert A, Heinen Y, Roeskes C, Hilfiker-Kleiner D, Heusch G, Schulz R (2008) Cardioprotection by ischemic postconditioning is lost in aged and STAT3-deficient mice. Circ Res 102:131-135
- Cohen MV, Yang XM, Downey JM (2007) The pH hypothesis of postconditioning: staccato reperfusion reintroduces oxygen and perpetuates myocardial acidosis. Circulation 115:1895-1903
- Darling CE, Solari PB, Smith CS, Furman MI, Przyklenk K (2007) 'Postconditioning' the human heart: multiple balloon inflations during primary angioplasty may confer cardioprotection. Basic Res Cardiol 102:274–278
- Dow J, Kloner RA (2007) Postconditioning does not reduce myocardial infarct size in an in vivo regional ischemia rodent model. J Cardiovasc Pharmacol Ther 12:153–163
- Engels W, Van Bilsen M, De Groot MJ, Lemmens PJ, Willemsen PH, Reneman RS, Van der Vusse GJ (1990) Ischemia and reperfusion induced formation of eicosanoids in isolated rat hearts. Am J Physiol 258:H1865–H1871
- Farber NE, Gross GJ (1989) Prostaglandin E1 attenuates postischemic contractile dysfunction after brief coronary occlusion and reperfusion. Am Heart J 118:17–24

- 8. Farber NE, Pieper GM, Thomas JP, Gross GJ (1988) Beneficial effects of iloprost in the stunned canine myocardium. Circ Res 62:204-215
- 9. Gelpi RJ, Morales C, Cohen MV, Downey JM (2002) Xanthine oxidase contributes to preconditioning's preservation of left ventricular developed pressure in isolated rat heart: developed pressure may not be an appropriate end-point for studies of preconditioning. Basic Res Cardiol 97:40-46
- Gres P, Schulz R, Jansen J, Umschlag C, Heusch G (2002) Involvement of endogenous prostaglandins in ischemic preconditioning in pigs. Cardiovasc Res 55:626-632
- 11. Grosser T, Fries S, FitzGerald GA (2006) Biological basis for the cardiovascular consequences of COX-2 inhibition: therapeutic challenges and opportunities. J Clin Invest 116:4–15

- 12. Hausenloy DJ, Tsang A, Yellon DM (2005) The reperfusion injury salvage kinase pathway: a common target for both ischemic preconditioning and postconditioning. Trends Cardiovasc Med 15:69–75
- Heusch G, Buchert A, Feldhaus S, Schulz R (2006) No loss of cardioprotection by postconditioning in connexin 43-deficient mice. Basic Res Cardiol 101:354–356
- 14. Hilgenfeldt U, Stannek C, Lukasova M, Schnolzer M, Lewicka S (2005) Rat tissue kallikrein releases a kallidin-like peptide from rat low-molecular-weight kininogen. Br J Pharmacol 146:958–963
- Kerendi F, Kin H, Halkos ME, Jiang R, Zatta AJ, Zhao ZQ, Guyton RA, Vinten-Johansen J (2005) Remote postconditioning. Brief renal ischemia and reperfusion applied before coronary artery reperfusion reduces myocardial infarct size via endogenous activation of adenosine receptors. Basic Res Cardiol 100:404-412
- 16. Kin H, Zatta AJ, Lofye MT, Amerson BS, Halkos ME, Kerendi F, Zhao ZQ, Guyton RA, Headrick JP, Vinten-Johansen J (2005) Postconditioning reduces infarct size via adenosine receptor activation by endogenous adenosine. Cardiovasc Res 67:124-133
- Leesar MA, Stoddard MF, Manchikalapudi S, Bolli R (1999) Bradykinin-induced preconditioning in patients undergoing coronary angioplasty. J Am Coll Cardiol 34:639–660
- 18. Liu X, Lukasova M, Zubakova R, Lewicka S, Hilgenfeldt U (2005) A kallidinlike peptide is a protective cardiac kinin, released by ischaemic preconditioning of rat heart. Br J Pharmacol 146:952–957
- Liu X, Lukasova M, Zubakova R, Lewicka S, Hilgenfeldt U (2006) Kallidinlike peptide mediates the cardioprotective effect of the ACE inhibitor captopril against ischaemic reperfusion injury of rat heart. Br J Pharmacol 148:825-832
- 20. Lochner A, Genade S, Moolman JA (2003) Ischemic preconditioning: infarct size is a more reliable endpoint than functional recovery. Basic Res Cardiol 98:337-346
- Mykytenko J, Kerendi F, Reeves JG, Kin H, Zatta AJ, Jiang R, Guyton RA,Vinten-Johansen J, Zhao ZQ (2007) Longterm inhibition of myocardial infarction by postconditioning during reperfusion. Basic Res Cardiol 102:90– 100

- 22. Pagliaro P, Mancardi D, Rastaldo R, Penna C, Gattullo D, Miranda KM, Feelisch M, Wink DA, Kass DA, Paolocci N (2003) Nitroxyl affords thiolsensitive myocardial protective effects akin to early preconditioning. Free Radic Biol Med 34:33–43
- 23. Penna C, Cappello S, Mancardi D, Raimondo S, Rastaldo R, Gattullo D, Losano G, Pagliaro P (2006) Post-conditioning reduces infarct size in the isolated rat heart: role of coronary flow and pressure and the nitric oxide/ cGMP pathway. Basic Res Cardiol 101:168–179
- 24. Penna C, Mancardi D, Rastaldo R, Losano G, Pagliaro P (2007) Intermittent activation of bradykinin B2 receptors and mitochondrial KATP channels trigger cardiac postconditioning through redox signaling. Cardiovasc Res 75:168–177
- 25. Penna C, Rastaldo R, Mancardi D, Raimondo S, Cappello S, Gattullo D, Losano G, Pagliaro P (2006) Post-conditioning induced cardioprotection requires signaling through a redoxsensitive mechanism, mitochondrial ATP-sensitive K+ channel and protein kinase C activation (see comment). Basic Res Cardiol 101:180–189
- 26. Philipp S, Yang XM, Cui L, Davis AM, Downey JM, Cohen MV (2006) Postconditioning protects rabbit hearts through a protein kinase C-adenosine A2b receptor cascade. Cardiovasc Res 70:308–314
- 27. Rastaldo R, Paolocci N, Chiribiri A, Penna C, Gattullo D, Pagliaro P (2001) Cytochrome P-450 metabolite of arachidonic acid mediates bradykinin-induced negative inotropic effect. Am J Physiol Heart Circ Physiol 280:H2823– H2832
- 28. Rossoni G, Berti M, Colonna VD, Bernareggi M, Del Soldato P, Berti F (2000) Myocardial protection by the nitroderivative of aspirin, NCX 4016: in vitro and in vivo experiments in the rabbit. Ital Heart J 1:146–155
- 29. Sato H, Bolli R, Rokosh GD, Bi Q, Dai S, Shirk G, Tang XL (2007) The cardioprotection of the late phase of ischemic preconditioning is enhanced by postconditioning via a COX-2-mediated mechanism in conscious rats. Am J Physiol Heart Circ Physiol 293:H2557-2564
- 30. Schulz R, Post H, Vahlhaus C, Heusch G (1998) Ischemic preconditioning in pigs: a graded phenomenon: its relation to adenosine and bradykinin. Circulation 98:1022–1029

- 31. Sivaraman V, Mudalgiri NR, Di Salvo C, Kolvekar S, Hayward M, Yap J, Keogh B, Hausenloy DJ, Yellon DM (2007) Postconditioning protects human atrial muscle through the activation of the RISK pathway. Basic Res Cardiol 102:453-459
- 32. Staat P, Rioufol G, Piot C, Piot C, Cottin Y, Cung TT, L'Huillier I, Aupetit JF, Bonnefoy E, Finet G, Andre-Fouet X, Ovize M (2005) Postconditioning the human heart. Circulation 112:2143– 2148
- 33. van Bilsen M, Engels W, van der Vusse GJ, Reneman RS (1989) Significance of myocardial eicosanoid production. Mol Cell Biochem 88:113–121
- 34. Veeravalli KK, Akula A, Kota MK (2003) Nitric oxide- and prostaglandinmediated cardioprotection by bradykinin in myocardial ischemia and reperfusion injury. Pol J Pharmacol 55:1021– 1029
- 35. Vinten-Johansen J, Zhao ZQ, Zatta AJ, Kin H, Halkos ME, Kerendi F (2005) Postconditioning-a new link in nature's armor against myocardial ischemia-reperfusion injury. Basic Res Cardiol 100:295-310
- 36. Yang XM, Downey JM, Cohen MV (2005) Postconditioning's protection is not dependent on circulating blood factors or cells but requires PI3-kinase and guanylyl cyclase activation. Basic Res Cardiol 100:57–63
- 37. Yang XP, Liu YH, Scicli GM, Webb CR, Carretero OA (1997) Role of kinins in the cardioprotective effect of preconditioning: study of myocardial ischemia/reperfusion injury in B2 kinin receptor knockout mice and kininogen-deficient rats. Hypertension 30:735-740
- Yellon DM, Opie LH (2006) Postconditioning for protection of the infarcting heart. Lancet 367:456–458
- 39. Zhao ZQ, Corvera JS, Halkos ME, Kerendi F, Wang NP, Guyton RA, Vinten-Johansen J (2003) Inhibition of myocardial injury by ischemic postconditioning during reperfusion: comparison with ischemic preconditioning. (erratum appears in Am J Physiol Heart Circ Physiol. 2004 Jan;286(1):H477). Am J Physiol Heart Circ Physiol 285:H579-H588
- Zhao ZQ, Vinten-Johansen J (2006) Postconditioning: reduction of reperfusion-induced injury. Cardiovasc Res 70:200-211