



Lignan exposure: a worldwide perspective

Lucia Rizzolo-Brime¹ · Elida M. Caro-Garcia¹ · Cynthia A. Alegre-Miranda¹ · Mireia Felez-Nobrega² · Raul Zamora-Ros¹

Received: 6 May 2021 / Accepted: 3 November 2021 / Published online: 20 November 2021
© The Author(s), under exclusive licence to Springer-Verlag GmbH Germany 2021

Abstract

Dietary lignans are phytoestrogens that are mostly found in plant-based foods, especially whole grains, seeds, nuts, legumes and vegetables. An accurate assessment of lignan exposure is crucial to evaluate their potential health benefits and to establish future recommendations and dietary guidelines. This narrative review aimed to (i) summarize the pros and the cons of the current main assessment methods for lignan exposure—i.e., dietary questionnaires, food composition tables and biomarkers, (ii) describe the individual lignans more consumed from a worldwide perspective, as well as their main food sources, (iii) determine the lignans concentrations in both urine and blood, and explore their heterogeneity among countries, and finally (iv) discuss the main determinants of lignan exposure.

Keywords Lignan · Intake · Biomarker · Urine · Plasma · Serum · Enterolignan

Abbreviations

24-HDR	24-H dietary recall
END	Enterodiol
ENL	Enterolactone
EPIC	European Prospective Investigation into Cancer and Nutrition
FFQ	Food frequency questionnaire
LARI	Lariciresinol
MATA	Matairesinol
MEDI	Medioresinol
PINO	Pinoresinol
SECO	Secoisolariciresinol
SYRI	Syringaresinol

Introduction

Chemistry and bioavailability

Lignans are secondary plant metabolites widely distributed in many plant-derived foods, such as whole grains, seeds, nuts, legumes, vegetables, and drinks (e.g., tea, coffee, or wine) [1]. Lignans are bioactive compounds well-known by their ability to mimic or modulate the action of endogenous estrogens [2]. Thus, they have been suggested to play a role in the prevention of several chronic and hormone-related diseases such as cardiovascular disease [1, 3], breast cancer [4, 5], osteoporosis [6], and menopausal symptoms [7, 8]. Lignans are chemically polyphenolic compounds derived from two β - β' -linked phenylpropane (C6–C3) units. Based on the way in which oxygen is incorporated into the skeleton and cyclization patterns, they can be classified into eight subgroups: furans, furofurans, dibenzylbutanes, dibenzylbutyrolactones, dibenzocyclooctadienes, dibenzylbutyrolactols, aryltetralins, and arylnaphthalenes. The most common lignans consumed and for which the evidence has shown the most compelling benefits for health are secoisolariciresinol (SECO), lariciresinol (LARI), pinoresinol (PINO), matairesinol (MATA); although other lignans are also frequently consumed [e.g., sesamol, sesamin, syringaresinol (SYRI) and medioresinol (MEDI)] [9].

In nature, lignans are generally linked to other molecules, mainly as glycosylated derivatives [10]. Lignan glycosides

✉ Raul Zamora-Ros
rzamora@idibell.cat

¹ Unit of Nutrition and Cancer, Cancer Epidemiology Research Program, Catalan Institute of Oncology, Bellvitge Biomedical Research Institute (IDIBELL), Av Gran Via 199-203, 08908 L'Hospitalet de Llobregat, Barcelona, Spain

² Research and Development Unit, Parc Sanitari Sant Joan de Déu, Barcelona, Spain

are absorbed in the gastrointestinal tract after being metabolized by gut mucosa and/or colonic microbiota into lignan aglycones and further converted into enterolignans [i.e., enterolactone (ENL) and enterodiol (END)] [1, 11]. The efficacy of this conversion depends on several factors, especially on the microbiota composition and function, and differs considerably among individuals. In an *in vitro* fecal microbiota metabolism system, 100% of LARI, 72% of SECO and 55% of PINO were converted to END; while approximately half of END and 62% of MATA were transformed to ENL [12]. Enterolignans, also called mammalian lignans, are efficiently absorbed and conjugated to glucuronide and/or sulfates by enterocytes. Finally, enterolignans are detected in blood (8–10 h half-life) and excreted 30% through urine (residence time approximately 24 h) and 50% via enterohepatic circulation and feces [11]. Only small amounts of LARI, MATA, PINO, SECO, and SYRI have been found in blood and urine [13] (Fig. 1).

In plant-derived foods, the richest sources of lignans are sesame seed oil (1294 mg/100 g), flaxseed meal (867 mg/100 g), and sesame seed meal (776 mg/100 g), followed to a lesser extent by whole grains and virgin olive oil (<5 mg/100 g). The lignan content of other or plant-derived foods is generally minimal with concentrations lower than 1 mg/100 g [14]. Similarly, only negligible amounts of enterolignans have been detected in specific animal foods (i.e., milk, eggs, and derived products), which are produced by the intestinal bacterial metabolism in the animals' guts after eating a diet rich in lignans [15]. A list of the top 25

richest foods of the main six individual lignans is shown in the Supplementary Table 1.

Exposure assessment

In nutritional studies, lignan exposure has been assessed using either dietary questionnaires or nutritional biomarkers. Both methodologies have advantages and disadvantages. On one hand, dietary questionnaires [e.g., food frequency questionnaires (FFQ), 24-h dietary recalls (24-HDR), and food diaries] are inexpensive, easy to administer and can estimate a lot of dietary data simultaneously, including dietary patterns, foods, nutrients and non-nutrients [16]. On the other hand, dietary questionnaires are susceptible to random and systematic reporting errors since they are based on subjects' memory and their ability to estimate food portion sizes. Moreover, a food composition database is needed to convert food consumption into lignan intake. Phenol-Explorer [17] is the most comprehensive database on polyphenols that include all individual lignans ($n \sim 30$) present in habitual foods. Other studies have used other food composition databases from Canada [18], the Netherlands [19], UK [20–22] and Finland [23]; although these only usually include the four main individual lignans. The main limitations of using these databases are a large amount of unknown values, the limited quantity of food items included, and the absence of composition data on cooked foods. Thus, the estimation of lignan intake may be inaccurate and tends to be

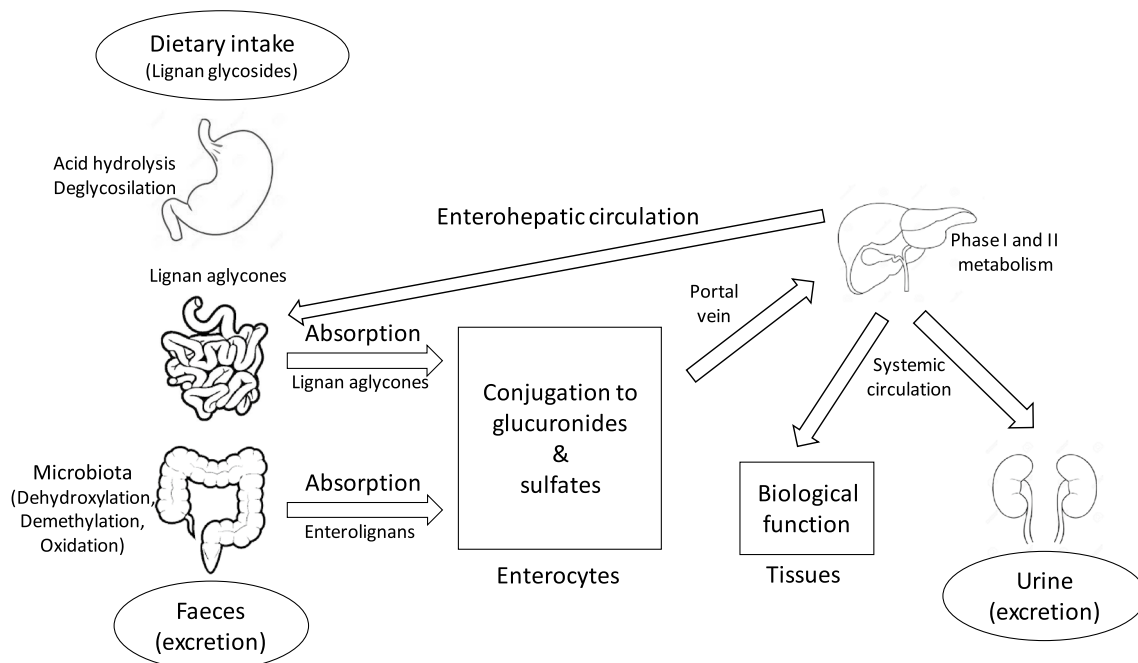


Fig. 1 Scheme of the human bioavailability of dietary lignans

underestimated. To improve the accuracy of self-reported dietary estimates, researchers are using new technologies, which are practical, have lower costs and burden for both researchers and participants (e.g., mobile phone applications) [24]. Moreover, they are using databases that are regularly updated, allowing to increase the number of available foods and individual lignans.

Nutritional biomarkers have become an alternative or complementary method for estimating dietary intake. An ideal dietary biomarker would accurately reflect its dietary intake and be specific, sensitive, and applicable to many populations. Their main advantage is that they are objective, take into account bioavailability, and offer more accurate assessment since they do not rely on subject's memory. In contrast, their disadvantages include the requirement of biological samples, the complexity of the analytical methodology, and the elevated cost [25]. During the last two decades, lignans and especially enterolignans have been measured in blood and urine samples as potential biomarkers of dietary lignans. Currently, the analytical method generally used is liquid chromatography coupled to a tandem mass spectrometer (LC–MS/MS); although gas chromatography GC–MS and time-resolved fluorescence immunoassay have also been successfully used. These analytical methodologies allow us to have limits of detections below 0.1 mg/L [26].

Concentrations of enterolignans in plasma and urine have been extensively investigated as potential biomarkers of dietary lignan intakes. In a pooled analysis, urinary ENL levels have been highly correlated with MATA and SECO intake ($r=0.78$), but not urinary END ($r=-0.14$) [27]. However, in individual studies, correlations between lignan intake (sum of MATA and SECO) and urinary enterolignans (sum of ENL and END) were moderate ($r=0.40-0.46$) in 26 Canadian women [28] and low ($r=0.16-0.25$) in 195 adults from the California Teachers Study [29]. Weak associations between lignan intake and plasma END ($r=0.09$) and ENL ($r=0.18$) were observed in a Dutch study [30]. Similarly, correlations between lignan intake and sum of plasma/serum enterolignans were low ($r=0.1-0.22$) [31]. These low correlations could be due to the constraints to accurately assess dietary lignan intake (such as the aforementioned limitations of dietary questionnaires and food composition databases) or to difficulties to analyze the lignan content in foods, particularly in the extraction since they are usually bounded to dietary fiber [32]. It is also probable that a low correlation may exist due to the high inter- and intra-individuality in the absorption, metabolism and excretion of lignans or in the average lifetime of enterolignans in biospecimens (plasma and urine) [11]. Despite these results, concentrations of enterolignans, especially in urine, are considered suitable and reliable alternative measurements of lignan exposure.

Worldwide dietary lignan intake

Geographical differences in the intake of lignans and their food sources

Due to differences in dietary patterns worldwide, lignan intakes vary considerably by geographical region, with mean intakes mostly ranging from 0.2 to 6.4 mg/d in adults (Table 1, Fig. 2) [9, 33]. It is important to highlight that comparing results and estimates across studies presents several challenges due to differences in the amount of individual lignans included, and both the composition database and the dietary assessment method used. However, some studies used similar methodologies that allow us to compare results more easily.

Europe

Europe is the continent with more studies estimating the intake of lignans (Table 1). In adults, the mean intake ranged from 0.2 mg/d to 5.2 in France [34] and Latvia [35], respectively. Unsurprisingly, the highest intake of lignans (9.1 mg/d) was reached in a vegetarian/vegan UK population, since lignan is almost exclusively found in plant-based foods [9]. Despite the differences between studies, the existence of large multi-center studies such as the European Prospective Investigation into Cancer and Nutrition (EPIC) and the Healthy Lifestyle in Europe by Nutrition in Adolescents (HELENA) allows to compare lignan intakes across Europe using the same methodology [9, 36, 37]. Data from the EPIC study, that used Phenol-Explorer database, indicates that Mediterranean countries have a higher intake than the non-Mediterranean ones [9, 36]. However, the HELENA study, which used the Dutch database, showed a small decreasing north-to-south gradient [37].

Data from studies using different methodology and databases indicates that the highest lignan intake in Europe usually occurs in northern countries, including Scandinavian and Baltic countries (Table 1). Considering the assessment of at least 6 individual lignans (LARI, MATA, PINO, SECO, SYRI, and MEDI), the average of overall lignan intake ranged between 2.3 and 5.2 mg/d. Intake estimates were lower (0.9–1.8 mg/d) if only LARI, MATA, PINO, and SECO were considered. LARI, PINO and SECO were usually the individual lignans more consumed, although SYRI was also common. The main food sources of lignans in this region were whole grain cereals (especially rye, oat, and wheat), bread, flaxseeds, and berries.

The mean intake of lignans in Central European countries, such as UK, Poland, Germany, and the Netherlands,

Table 1 Characteristics of the studies included in the review of dietary lignan intake

Author (Reference)	Year	Country	Population		Dietary survey	FCDB	TOTAL LIGNANS			
			N	Sex			Individual lignans	Intake (mg/d)	Food sources	
Wisnuwarda-ni [36]	2006–2007	MED countries	915	F (53%)	12–18	2 × 24-HDR	Phenol Explorer	All ^d	1.2 (0.0) ^a	Breads (71%), fruit (8%), vegetables (7%)
			1513							0.9 (0.0) ^a
Zamora-Ros [9]	1995–2000	MED countries	11,285	F (64%)	35–74	24-HDR	Phenol Explorer	All ^d : SECO (18%), LARI (14%), Sesamolol (12%), Sesamin (12%)	3.6 (0.1) ^a	Vegetable oils (26%), cakes and biscuits (20%), breads (12%)
			24,443							2.3 (0.1) ^a
		UK healthy	309						9.1 (0.9) ^a	Seeds (48%), vegetable oils (10%), vegetables (9%)
Tetens [37]	2000–2002	Denmark	2463	F (53%)	25–64	7-DR	Dutch DB	LARI (43%), PINO (32%), SECO (22%), MATA (3%)	1.5 ^b	Cereals (27–30%), fruit and berries (18–25%), coffee and tea (21%), vegetables (19–20%)
			2007	F (55%)	25–64	48-HDR		LARI (43%), PINO (37%), SECO (17%), MATA (2%)	1.1 ^b	Cereals (27–36%), fruit and berries (22–31%), vegetables (16–20%), coffee and tea (17%)
	1994–1996	Italy	1268	F (54%)	25–64	7-DR		LARI (45%), PINO (42%), SECO (13%), MATA (1%)	1.1 ^b	Fruit and berries (42–46%), vegetables (26–28%), cereals (17%)
	1987–1990	Sweden	83,760	F (45%)	45–79	FFQ		PINO (44%), LARI (41%), SECO (13%), MATA (2%)	1.8 ^b	Cereals (26–42%), vegetables (18–30%), fruit and berries (15–23%), coffee and tea (18–19%)
	2000–2001	UK	1724	F (56%)	19–64	7-DR		LARI (43%), PINO (39%), SECO (16%), MATA (2%)	1.2 ^b	Coffee and tea (30–32%), vegetables (23–25%), fruit and berries (15–20%), cereals (15–17%)
Kilkinen [57]	1997	Finland	1359	M	25–63	24-HDR	Finish DB	MATA (73%), SECO (27%)	0.2 ^b	Cereals (49%), fruits (25%), vegetables (12%)
			1493	F	25–64			MATA (80%), SECO (20%)	0.2 ^b	Fruits (39%), cereals (35%), vegetables (13%)
Nurmi [90]	1995	Finland	100	M	58(6) ^a	4-DR	Dutch DB	LARI (40%), PINO (38%), SECO (14%), MATA (7%)	1.2 (0.5) ^c	Rye products, berries, coffee, tea, vegetable roots

Table 1 (continued)

Author (Reference)	Year	Country	Population		Dietary survey	FCDB	TOTAL LIGNANS		Food sources
			N	Sex			Individual lignans	Intake (mg/d)	
Hedelin [63]	1991–1992	Sweden	48,268	F	FFQ	Finish DB II	LARI, MATA, PINO, SECO, SYRI, MEDI	2.3 (1.8–2.8) ^c	Rye bread (57%), wheat bread (27%), cereals (8%)
Hedelin [62]	1991–1992	Sweden	46,977	F	FFQ	Finish DB II	LARI, MATA, PINO, SECO, SYRI, MEDI	2.3 (1.0–4.0) ^c	Rye bread, wheat bread, cereals, berries
Suzuki [55]	1987–1990	Sweden	51,823	F	FFQ	Own DB	LARI, MATA, PINO, SECO	0.9 (0.7–1.0) ^c	–
Hedelin [91]	2001–2002	Sweden	1130	M	FFQ	Finish DB II	SECO (38%), SYRI (30%), PINO (15%), LARI (13%), MEDI (12%), MATA (1%)	4.9 ^b	Flaxseed (36%), Rye bread (39%), wheat bread (15%)
Mejja [35]	2009–2011	Latvia	172	M	FFQ	Canadian DB	SECO (58%), SYRI (22%), PINO (11%), LARI (6%), MATA (1%), MEDI (1%)	5.2 (6.4) ^a	Seed and rye bread (86%), flaxseed (7%); Seed and rye bread (57%), flaxseed (35%)
Bhakta [92]	1995–1999	UK	108	F	≥ 9×24HDR	Finish DB II	SECO (93%), MATA (7%)	0.1 (0.1) ^a	Breads (75%), vegetables (9%), fruit and fruit juices (7%)
Bhakta [93]	1995–1999	UK (Asian)	221	F	≥ 4×24HDR	Own DB	SECO (93%), MATA (7%)	0.1 (0.1) ^a	Breads (70%), vegetables (12%)
Mulligan [94]	1993–1997	UK	9680	M	7d DR	Own DB	SECO (93%), MATA (7%), MATA, SECO, Shonanin	0.2 (0.1) ^a 0.3 (0.2) ^a 0.3 (0.1) ^a	Breads (60%), fruit and fruit juices (21%) Tea and coffee (33%), beer (12%), vegetables (9%) Tea and coffee (37%), vegetables (12%), fruits (9%)
Grosso [38]	1993–1997	Poland	10,477	F (50%)	FFQ	Phenol Explorer	All ^d	0.6 (1.2) ^a	Seeds (51%) tea (27%), dark bread (8%)
Witkowska [95]	2003–2014	Poland	1683	F	24-HDR	Dutch DB	SECO (45%), LARI (26%), PINO (26%), MATA (3%)	1.1 (4.4) ^a	Vegetables (38%), flaxseed (22%), tea (12%)
Witkowska [39]	2003–2005	Poland	6661	F (53%)	24-HDR	Phenol Explorer	All ^d	12.1 ^b	Cucumber (41%), red cabbage (22%)

Table 1 (continued)

Author (Reference)	Year	Country	Population		Dietary survey	FCDB	TOTAL LIGNANS		Food sources	
			N	Sex			Individual lignans	Intake (mg/d)		
Linseisen [96]	1992–1995	Germany	666	F	43(6) ^a	FFQ	Own DB	SECO (94%), MATA (6%)	0.6 (0.3–1.3) ^c	Nuts and seeds (75%), vegetables (7%), coffee (6%)
Boker [97]	1993–1997	Netherlands	17,140	F	50–69	FFQ	Dutch DB	SECO (93%), MATA (7%)	1.0 ^b	Breads (41%), coffee and tea (23%), fruits (14%)
Milder [61]	1997–1998	Netherlands	4661	F (55%)	≥ 19	2-DR	Dutch DB	LARI (43%), PINO (32%), SECO(24%), MATA(0.6%)	1.2 (2.1) ^a	Tea and coffee (37%), nuts and seeds (14%),
Milder [30]	1997–2002	Netherlands	306	F (56%)	19–75	FFQ	Dutch DB	LARI (47%), PINO (35%), SECO (18%), MATA (1%)	1.1 (0.4) ^a	Vegetables and black tea (> 20%), whole-grain bread, fruits, wine
Milder [98]	1985–1995	Netherlands	570	M	64–84	DH	Dutch DB	LARI (48%), PINO (36%), SECO (15%), MATA (1%)	1.0 (0.8–1.0) ^c	Tea (28%), vegetables (27%), bread (14%)
Pérez-Jiménez [99]	1994–2001	France	4942	F (47%)	35–60	6×24-HDR	Phenol Explorer	All ^d	0.4 (0.2) ^a	Coffee (21%), refined wheat products (18%), whole-grain wheat products (16%)
Lefèvre-Arbogast [100]	1999–2000	France	1329	F (62%)	≥ 65	24-HDR	Phenol Explorer	All ^d	0.4 (0.3) ^a	Wine (65%), olive oil (12%), tea and infusion (9%), soy products (8%)
Adriouch [34]	1994–1996	France	3903	F (47%)	35–60	≥ 6×24-HDR	Phenol Explorer	All ^d	0.2 (0.1) ^a	Bread (30%), red wine (29%), olive oil (15%), tea (9%)
Pellegrini [101]	2002–2003	Italy	242	F (38%)	60(8) ^a	3D-WR	Dutch DB	SECO (52%), LARI (27%), PINO (17%), MATA (3%)	0.7 (0.3) ^a	Red wine, fruits and vegetables (80%)
Pounis [58]	2005–2010	Italy	14,029	F (50%)	35 ^b	FFQ	Eurofir-eBASIS	–	80 (60–106) ^c	Seasonal fruits (41%), grain and pod vegetables (11%)
Godos [102]	2014–2015	Italy	1947	F (33%)	> 18	FFQ	Phenol Explorer	All ^d	2.8 (2.6) ^a	Citrus fruits (44%), red orange (32%), garlic (11%)
Godos [103]	2014–2015	Italy	1936	F (28%)	> 18	FFQ	Phenol Explorer	All ^d	1.4 (1.1–2.0) ^a	Citrus fruits, garlic, olive oil, bread
Russo [40]	2015–2016	Italy	340	M	> 18	FFQ	Phenol Explorer	LARI (54%), PINO (34%), SECO (4%), MATA (1%)	3.1 (2.7) ^a	Cereals, fruits, vegetables, grains, nuts

Table 1 (continued)

Author (Reference)	Year	Country	Population		Dietary survey		FCDB	TOTAL LIGNANS	
			N	Sex	Age (y)	Intake (mg/d)		Food sources	
González [104]	–	Spain	127	M	73(7) ^a	FFQ	Phenol Explorer	All ^d	0.5 (0.3) ^a Olive oil, white bread, and red wine (93%)
Peñalvo [56]	1998–2000	Spain	177	F	77(6) ^a	24-HDR	Alignia DB	PINO (42%), SECO (17%), LARI (13%), MATA (1%)	0.4 (0.2) ^a 0.8 (0.5–1.3) ^c Olive oil (27%), refined wheat bread (17%), whole-grain wheat bread (8%)
Zamora-Ros [105]	1996–1998	Spain	401	M	65 (12) ^a	FFQ	UK DB	SECO, MATA, LARI, PINO	0.7 (0.5–1.0) ^c Fruit (32%), vegetables (31%), cereals products (10%)
Tresserra-Rimbau [106]	2003–2009	Spain	7200	MandF	55–80	FFQ	Phenol Explorer	All ^d	0.9 (0.4) ^a Olive oil (47%), virgin olive oil (25%), whole-grain wheat-flour bread (6%)
Mendonça [107]	1999–	Spain	17,065	F (61%)	20–89	FFQ	Phenol Explorer	All ^d	0.6 (0.4) ^a Olive oil, dried fruits, gazpacho, bread
Petrick [108]	1997–2000	US	183	MandF	20–80	FFQ	Canadian DB	SECO, MATA	0.6 (0.1) ^a Coffee (31), wine (12), and citrus juice (9%)
Petrick [109]	1993–1995	US	662	MandF	30–79	FFQ	Canadian DB	SECO, MATA	0.07 (0.03) ^a Coffee (35%), citrus juice (13%), wine (10%)
Williams [110]	2003–2008	US	216	F	55 (13) ^a	FFQ	Canadian DB	SECO, MATA, LARI, PINO	0.1(0.1–0.2) ^c –
Carmichael [42]	1997–2005	US	3118	F	~18–40	FFQ	UK DB	SECO (87%), MATA (13%)	0.2 (0.05–0.3) ^c Coffee and tea, alfalfa sprouts, flaxseed
Waetjen [111]	2008	US	1459	F	42–52	FFQ	Own DB	SECO, MATA, LARI, PINO	0.2 ^b –
Bandera [112]	2004–2008	US	391	F	~21	FFQ	Own DB	SECO (89%), LARI (6%), PINO (4%), MATA (1%)	1.0 ^a –
Chang [113]	1995–2007	US	110,215	F	20–84	FFQ	Canadian DB	SECO, MAT, LARI, PINO	0.8 (0.3–1.3) ^c Vegetables, fruits, whole grains
McCann [114]	1996–2001	US	1122	F	35–79	FFQ	Canadian DB	SECO (50%), PINO (21%), MATA (3%), LARI (3%)	0.2 (0.1) ^a Whole-grain bread, peaches, orange juice, coffee, onions, string beans, tea
Fink [43]	1996–1997	US	1500	F	<65	FFQ	Own DB	SECO, MATA	6.4 (4.7) ^a Tea (99%), strawberries (0.5%), whole grain products(0.3%)
Horn-Ross [59]	1996–1999	US	470	F	35–79	FFQ	Own DB	SECO (77.9%), MATA (16.9%)	0.2 (0.1–0.2) ^c –

Table 1 (continued)

Author (Reference)	Year	Country	Population		Dietary survey		FCDB	TOTAL LIGNANS		Food sources
			N	Sex	Age (y)	24-HDR		Phenol Explorer	Individual lignans	
Mervish [115]	2004–2014	US	1044	F	6–8	24-HDR	Phenol Explorer	All ^d	0.4 ^b	Orange juice (35%), strawberries (17%), broccoli (8%)
van der Schouw [116]	1994	US	468	M	47–83	FFQ	US DB	SECO (97%), MATA (3%)	0.7 (0.5–0.9) ^c	Tea and coffee (28%), alcoholic beverages (9%), cereals and grains (7%)
Horn-Ross [117]	1992–1998	US	558	F	20–74	FFQ	Own DB	SECO (71%), MATA (29%)	0.1 (0.1–0.2) ^c	–
McCann [118]	1986–1991	US	696	F	40–85	FFQ	US DB	SECO, MATA	0.5 (0.3) ^a	Coffee, carrots, cucumbers, strawberries
Schabath [60]	1995–2003	US	1735	F (49%)	–	FFQ	US DB	SECO, MATA	5.3 (3.4–9.7) ^c	Coffee 52%, tea 30%, flaxseed (6%)
de Kleijn [119]	1991–1994	US	964	F	–	FFQ	US DB	SECO (97%), MATA (3%)	0.6 (0.4–0.8) ^c	Other fruits (13%), cereals and grains (11%), berries (8%)
Cotterchio [44]	2001	Canada	1890	F (47%)	20–74	FFQ	Own DB	SECO, MATA	0.2 (0.1–0.3) ^c	Legumes, seeds, cereals/grains, berries, dried fruit, vegetables
Morisset [45]		Canada	115	F	≤70	FFQ	Canadian DB	SECO, MAT, LARI, PINO	0.4 (3.8) ^a	–
Chávez-Suárez [120]	2012–2017	Mexico	100	F	25–80	FFQ,	Own DB	SECO (73%), END (18%), ENL (7%), MATA (2%)	1.1 (1.6) ^a	–
Hernández-Ramírez [46]	2004–2005	Mexico	478	F (46%)	>20	FFQ	Own DB	SECO (75%), END (16%), ENL (5%), MATA (4%)	0.4 (1.8) ^a	–
Zamora-Ros [33]	2006–2011	Mexico	106,466	F	≥20	FFQ	Phenol Explorer	LARI (54%), PINO (26%), SECO (20%), MATA (0.1%)	0.3 (0.2–0.5) ^c	Vegetables, fruits, legumes
Nascimento-Souza [47]	2016	Brasil	620	F (70%)	60–98	24-HDR	Phenol Explorer	All4: LARI (46%), PINO (21%), SECO (18%)	0.1 (0.03–0.2) ^c	Broccoli and cauliflower (11%), strawberries (9%), fruit-flavoured water (6%)
Miranda [48]	2008–2009	Brasil	550	F (65%)	≥12	24-HDR	Phenol Explorer	All4: LARI(50%)	13.6 (25.5) ^a	Orange (16%), broccoli (15%), flaxseed (15%)
								All ^d	0.1 (0.1–0.2) ^c	Sesame seed oil (71%), nuts (20%), sesame seeds (4%)

Table 1 (continued)

Author (Reference)	Year	Country	Population		Dietary survey		FCDB	TOTAL LIGNANS		
			N	Sex	Age (y)			Individual lignans	Intake (mg/d)	Food sources
Miranda [49]	2008	Brasil	1103	F (54%)	20	24-HDR	Phenol Explorer	All ^d	2.3(0.7) ^a	Cereals oil (71%), nuts (26%), olive oil (2%)
Lahmann [50]	2002–2007	Australia	2078	F	18–79	FFQ	Canadian and UK DB	SECO (68%), LARI (12%), MATA (10%), PINO (8%)	0.7 (0.3) ^a	–
Hanna [51]		Australia	511	F	40–80	FFQ	AusNut DB		2.7 (3.0) ^a	Soy, linseed
Sohrab [52]	2006–2008	Iran	2618	F (56%)	19–84	FFQ	Phenol Explorer	All ^d	0.2 (0.1–0.3) ^c	Nuts, whole grains
Sohrab [53]	1999	Iran	1265	F (56%)	19–74	FFQ	Phenol Explorer	All ^d	3.8 (2.4–5.7) ^c	–
Jang [54]	2004	Korea	48	F PreM	40–51	24-HDR	US DB	END (34%), ENL (33%), SECO (28%), MATA (5%)	1.5 (0.3) ^a	–
			53	F PostM	41–57			END (36%), ENL (32%), SECO (25%), MATA (7%)	1.8 (0.5) ^a	–

24-HDR 24-h dietary recall; DB database; DH dietary history; DR dietary record; F female; FCDB Food Composition DataBase; FFQ food frequency questionnaire; LARI lariciresinol; M male; MATA matairesinol; MED Mediterranean; MEDI medioresinol; PINO pinioresinol; PostM post-menopausal; PreM pre-menopausal; SECO secoisolariciresinol; SYRI syringaresinol

^{a,b,c}Type of estimation: ^aMean (SD), ^bMean, ^cMedian (p25–p75)

^dAll lignans, including: 1-AcetoxyPINO, 7-HydroxyMATA, 7-HydroxySECO, 7-OxoMATA, Anhydro-SECO, Aretigenin, Conidendrin, CycloLARI, DimethylMATA, Episesamin, Episesaminol, IsohydroxyMATA, IsoLARI, LARI, LARI-sesquillignan, MATA, MEDI, Nortrachelogenin, PINO, SECO, SECO di-O-glucoside, SECO-sesquillignan, Sesamin, Sesaminol, Sesamol, Sesamolol, Sesamololol, SYRI, Todolactol A, Trachelogenin

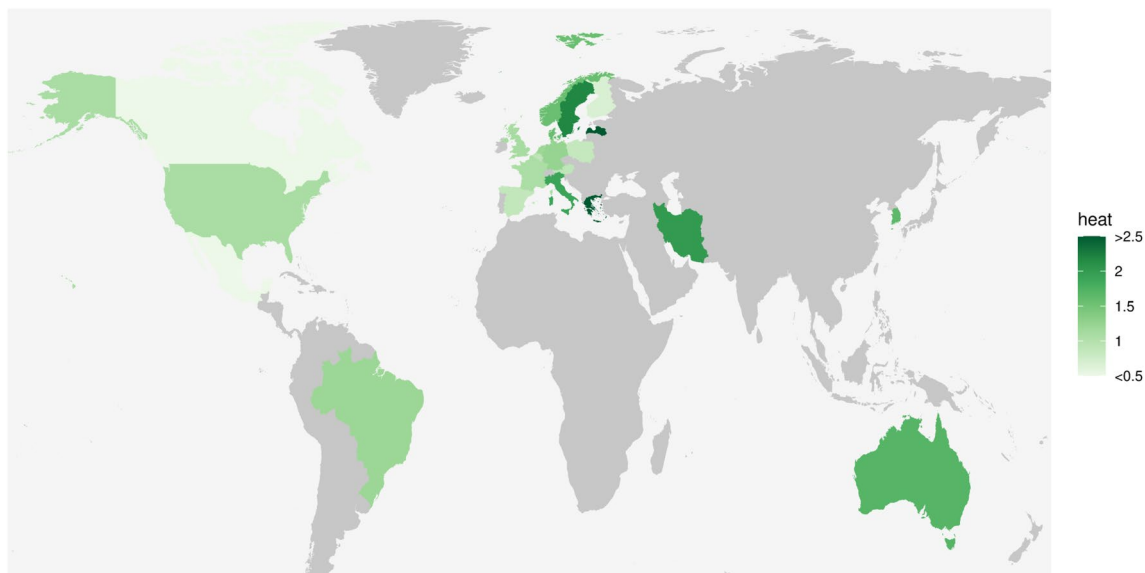


Fig. 2 Mean of means/medians of total dietary lignan intake (mg/d) by country

ranged between 0.6 [38] and 2.3 mg/d [9]. Most of the studies in this region only assessed LARI, MATA, PINO, and SECO, and therefore, the intakes may be slightly underestimated. In a Polish study [39] the mean intake of lignans was extremely high (12.1 mg/day) due to a Phenol-Explorer error in the lignans content of some specific vegetables [17] that were the main food sources in this Polish study (such as cucumber). In Central European countries, LARI, PINO and SECO were the main individual lignans consumed. Bread, seeds, and vegetables were the most common food sources of lignans in this region.

Finally, southern European countries, also referred as Mediterranean countries, had a highly variable intake, ranging from 0.2 mg/day in France [36] to 4.3 mg/day in Greece [9]. France and Spain had relatively low intakes (0.2–2.1 mg/d), while Italy and Greece generally had a high consumption (0.7–4.3 mg/d) [9, 36]. In an Italian study [40] the mean intake was extremely high (80 mg/d). Although the authors did not provide any rationale for such results, it is possible that this could be due to a processing error in the Eurofir-eBASIS food composition database [41]. LARI, PINO and SECO were also the most consumed individual lignans in this region; although depending on the study, the proportions largely vary. These countries typically follow a Mediterranean dietary pattern, where the main food sources of lignans are derived from olive oil, vegetables, fruits (mostly citrus fruit), wine (predominantly red wine) and in a minor percentage bread and cereal products.

Americas

In the US, there is also a great quantity of studies describing the lignan intake (Table 1). Most of these studies used the Canadian database [18] which only contains data on the four traditional individual lignans: LARI, MATA, PINO, and SECO. The mean intake of total lignans ranged between 0.1 and 6.4 mg/d [42, 43] although in the majority of these studies, their intake was < 1 mg/d. In this region, the main food sources were tea and coffee, probably due to a lower consumption of fruits, vegetables and whole grains compared to Europe. In the US, SECO was clearly the most consumed individual lignan, followed by far by LARI and PINO. In two Canadian studies, the intake of total lignans was slightly lower than in the US, ranging from 0.2 to 0.4 mg/d [44, 45] and the main food sources were legumes, seeds, cereals and grains, and berries. To date, only SECO and MATA were assessed in Canada, which clearly underestimate total lignan intake.

To our knowledge, the existing data in Latin-American countries is limited to Mexico [33, 46] and Brazil [47–49]. The mean intake of total lignans was similar in both countries, varying from 0.1 to 2.3 mg/d. A Brazilian study [47] was not included in the current review, since its mean intake was exceptionally high 13.6 mg/d, possibly due to an error in data calculation. As in Europe, SECO, LARI and PINO were the main contributors to total lignans in this region. Main food sources were generally vegetables, fruits, nuts, seeds and vegetable oils. However, there is

a potential underestimation of lignan intakes in Latin American countries due to the limited food composition data on some tropical foods [33], such as mamey, zapote, papaya, sweet potato, nopal, guava, jicama, and prickly pears. Those are frequently consumed in this region, but their lignan content is not available in any food composition database yet.

Other continents

In Australia, two studies estimated the intake of total lignans in women only [50, 51]. Their mean intake ranged from 0.7 to 2.7 mg/d. SECO was the major individual lignan consumed and the main food sources were soy and linseed [51].

In Asian countries, lignan intake was estimated only in two Iranian-based [52, 53] and one Korean-based [54] studies. In Iran, the mean intake of total lignans, including all individual lignans, varied between 0.2 and 2.4 mg/d; whereas in Korea, including only MAT and SECO, the mean intake was 1.5–1.8 mg/d. Data on main food sources were not available in this region.

Determinants of lignan intake

Lignans were positively correlated to total energy intake [55]; therefore, participants consuming more energy were more likely to be those with a higher intake of total lignans. Although a Latvian study [35] showed a greater consumption of total lignans in men compared to women; data from EPIC showed that women had a higher intake of lignans after adjusting for total energy consumption (3.6 mg/d in women vs. 2.5 mg/d in men) [9]. Interestingly, one Korean study [54] observed slight differences between menopausal statuses in women (1.8 mg/d in postmenopausal women vs. 1.5 mg/d in premenopausal women). In the EPIC study [9], results indicated that lignan intake also increased with age. For instance, young adults (35–44 years) had a lower intake of total lignans (2.8 mg/d) than older adults (65–74 years; 3.5 mg/d) [9]. In children and adolescents, the two available European studies [36, 56] found that the mean intake was higher in adolescents (15–18 years) than in children (2–15 years), 0.98–1.10 vs. 0.61–1.00 mg/d, respectively.

The results by lifestyle factors and other sociodemographic variables are controversial. For example, some studies showed that subjects with obesity had a higher intake of lignans [9, 36, 45, 57–60] than individuals with normal weight; whereas in other studies occurred the opposite [9, 35, 61–63]. Discrepancies were also observed comparing lignan intake by educational level, smoking status, physical activity, and alcohol consumption.

Worldwide enterolignans concentrations

Geographical differences in total enterolignans concentrations

Concentrations of lignan metabolites (END and ENL) in biospecimens, as potential biomarkers of lignan intake, are useful indicators of lignan exposures across populations. To straightforwardly compare concentrations of enterolignans, all estimates have been converted into the same units (nmol/L) in Tables 2 and 3. These summarize the most representative studies assessing urinary and blood (i.e., serum or plasma) enterolignan concentrations, respectively. Levels of urinary enterolignans were usually 100-fold higher than those found in blood (serum or plasma). The mean urinary END concentrations worldwide ranged from 38 [64] to 763 nmol/L [65] and for ENL from 148 [66] to 3651 nmol/L [67] (Table 2, Fig. 3). In the case of plasma and serum, END concentrations varied between 0.2 [68] and 7.0 nmol/L [69] while ENL levels ranged from 4.9 [68] to 39.2 nmol/L [69]. Levels of enterolignans in plasma and serum were similar (Table 3, Fig. 3). Mean concentrations of END were between 2 to 13 times lower than ENL in both urine and blood.

Europe

Few studies ($n = 8$) have measured urinary enterolignans in Europe (Table 2). Northern European countries tend to have the highest levels of enterolignans (ENL = 768–3267 nmol/L) [65, 70] followed by Central European countries (END = 204–288 and ENL = 2414–3333 nmol/L) [71–75]. Data for Mediterranean countries were limited. There is only one study from Italy, that reported a high urinary concentration (END = 763 and ENL = 1577 nmol/L) [76].

Most of the studies measuring enterolignan concentrations in blood specimens, of which 20 were conducted in plasma and 10 in serum, were performed in Central and Northern European countries (Table 3). The lowest concentrations of END and ENL were 0.2 and 4.9 nmol/L, respectively, in a UK-based study [68]; while the highest levels were derived from a Dutch population: 7.0 nmol/L for END and 39.2 nmol/L for ENL [69]. Comparing studies that used the same analytical methodology, in general, concentrations in Central European countries (e.g., the Netherlands, Germany, UK) were slightly lower than in Scandinavian countries [68, 77]. However, when all studies were considered independently of lignan assessments, levels of enterolignans in central European countries were very heterogeneous [68, 69]. The lowest mean

Table 2 Characteristics of the studies included in the review of urinary lignan excretions

Author (Reference)	Data collection	Country	N	Sex	Age	Type of study	Urine	Analytical method	END (nmol/L)	ENL (nmol/L)
Zamora-Ros [71]	1995–1999	Europe ^d	475	F (58%)	33–77	Cohort	24 h	LC-MS/MS	247 ^b	2080 ^b
Uehar [65]	–	Finland	126	F	24–65	C-S	24 h	TR-FIA	–	3267 ^a
Krogholm [70]	2002–2004	Denmark	84	M	38–63	Cohort	24 h	LC-MS	–	768 ^a
			107	F			Overnight		–	696 ^a
							24 h		–	1050 ^a
							Overnight		–	970 ^a
Ward [72]	1993–1997	UK	828	M	45–75	NCC	–	GC-MS	204 ^b	2953 ^b
			889	F (43%)					210 ^b	3333 ^b
Low [73]	1993–1997	UK	125	F	45–76	Cohort	Spot	LC-MS	288 ^a	2561 ^a
Grace [74]	1993–1997	UK	219	F	45–75	NCC	Spot	GC-MS	274 ^a	2792 ^a
Low [75]	1993–1997	UK	267	M	45–75	Cohort	Spot	GC-MS	207 ^a	2414 ^a
Durazzo [76]	–	Italy	13	F	48–58	CT	24 h	LC-CEAD	763 ^a	1577 ^a
									348 ^a	1092 ^a
Park [121]	2001–2006	US	404	M	45–75	NCC	Spot	LC-MS/MS	–	1313 ^b
Hu [122]	1997–2010	US	1111	F	25–55	Cohort	Spot	LC-MS	159 ^a	2938 ^a
Reger [123]	1999–2010	US	6009	F (52%)	>40	C-S	Spot	LC-MS/MS	248 ^a	2041 ^a
Martínez Steele [124]	2009–2010	US	2692	M/F	>6	C-S	Spot	LC-MS/MS	133 ^b	728 ^b
Adlercreutz [125]	–	US	10	F	58 ^a	C-S	24 h	GC-MS	267 ^c	2120 ^c
			10						213 ^c	1533 ^c
			7						140 ^c	693 ^c
Miles [126]	2006	US	80	F (50%)	18–45	CT	24 h	GC-MS	533 ^a	3000 ^a
									267 ^a	1933 ^a
Rybak [64]	2003–2006	US	2873	M	≥20	C-S	Spot	LC-MS/MS	41 ^a	302 ^a
				F					38 ^a	285 ^a
Reger [127]	1999–2004	US	5179	F (52%)	>18	C-S	Spot	LC-MS	133 ^b	1178 ^b
Eichholzer [128]	1999–2004	US	2028	F (49%)	>18	C-S	Spot	LC-MS	147 ^b	1507 ^b
	2005–2008		2628	F (48%)					164 ^b	1683 ^b
Xu [67]	2001–2010	US	694	M	12–19	C-S	Spot	LC-MS	278 ^a	2246 ^a
			600	F					463 ^a	2618 ^a
			1273	M	20–60				552 ^a	2950 ^a
			1226	F					609 ^a	3319 ^a
			578	M	>60				533 ^a	3651 ^a
			584	F					386 ^a	2907 ^a
Valentín-Blasini [129]	1999–2000	US	334	F (52%)	6–11	C-S	Spot	LC-MS	89 ^c	802 ^c
			757		12–19				84 ^c	852 ^c
			1496		≥20				93 ^c	758 ^c

Table 2 (continued)

Author (Reference)	Data collection	Country	N	Sex	Age	Type of study	Urine	Analytical method	END (nmol/L)	ENL (nmol/L)
Valentín-Blasini [80]	1988–1994	US	199	F (61%)	20–58	C-S	Spot	LC-MS/MS	209 ^a	1718 ^a
Sun [130]	1995–2001	US	452	F	53–79	NCC	Spot	LC-MS	123 ^a	2506 ^a
Kunisue [83]	2005–2009	US	655		32–52				77 ^a	2172 ^a
			10	M	24–63	C-S	24 h	LC-MS/MS	43 ^a	738 ^a
			6	F	23–48				129 ^a	872 ^a
Levine [131]	2005–2009	US	471	F	18–40	Cohort	Spot	LC-MS/MS	94 ^b	754 ^b
Simon [78]	–	Jamaica	171	F	20–75	CC	Spot	TR-FIA	–	2671 ^b
Liu [66]	2000–2001	Japan	500	F	20–70	C-S	Spot	GC-MS	95 ^c	148 ^c
Uehar [65]	–	Japan	111	F	24–65	C-S	24 h	TR-FIA	–	ND
Kunisue [83]	2005	Japan	15	M	22–54				126 ^a	1376 ^a
			11	F	21–35				80 ^a	1074 ^a
		Vietnam	31	M	20–78				133 ^a	772 ^a
			32	F	21–73				245 ^a	1678 ^a
		Vietnam	14	M	21–74				80 ^a	503 ^a
			14	F	33–74				182 ^a	705 ^a
		Cambodia	13	M	21–48				60 ^a	571 ^a
			24	F	21–46				86 ^a	671 ^a
		India	16	M	27–62				179 ^a	1141 ^a
			23	F	20–70				119 ^a	940 ^a
		India	18	M	26–55				255 ^a	1443 ^a
			24	F	20–48				205 ^a	1342 ^a
Talaei [82]	1999–2004	Singapore	564	F (58%)	45–74	NCC	Spot	LC-MS/MS	228 ^a	1140 ^a

CC case-control; C-S cross-sectional; CT clinical trial; END Enterodiol; ENL enterolactone; F female; HCC hospital-based case-control; GC-MS gas chromatography-mass spectrometry; LC-CEAD liquid chromatography-coulometric electrode array detector; LC-MS liquid chromatography-mass spectrometry; M male; NCC nested case-control; ND non detected; PCC population-based case-control; TR-FIA Sensitive time-resolved fluoroimmunoassay

^{a,b,c}Type of estimation: ^aMean, ^bMedian, ^cGeometric mean

^dFrance, Italy, Greece, and Germany

ENL and END concentrations have been converted into nmol/L from the original studies

Table 3 Characteristics of the studies included in the review of blood lignan concentrations

Reference	Data collection	Country	N	Sex	Age	Type of study	Specimen	Methods	END (nmol/L)	ENL (nmol/L)
Travis [77]	1992–2000	Europe ^d	1,042	M/F	60.1 ^a	NCC	Plasma	LC-MS/MS	1.0 ^b	12.4 ^b
Peeters [26]	1992–2012	Europe ^d	1,344	F (51%)	54–55	Cohort	Plasma	LC-MS	1.0 ^c	8.7 ^c
Pérez-Cornago [68]	1992–2000	UK healthy	70	F (49%)	59.6 ^a	NCC	Plasma	LC-MS/MS	3.6 ^c	17.8 ^c
	1993–1997	Europe ^d	1,042	M	64.7 ^a				1.0 ^c	11.2 ^c
	1981–1991	UK	130	M	46.5 ^a			TR-FIA	0.2 ^c	4.9 ^c
		Finland, Norway, Sweden	2,209	M					–	5.8 ^c
		Sweden	1,664	M	60.0 ^a				–	9.6 ^c
	1985–2017	Sweden	514	M	58.0 ^a				–	14.6 ^c
Uehara [65]	–	Finland	87	F	24–65	C-S	Plasma	TR-FIA	–	25.0 ^a
Stumpf [132]	1983	Finland	85	M/F	35–49	CT	Plasma	TR-FIA	–	19.5 ^b
Pietinen [133]	1990–1995	Finland	75	F PreM	25–75	PCC	Serum	TR-FIA	–	20.7 ^a
			133	F PostM					–	28.9 ^a
Vanharanta [134]	2005	Finland	167	M	42–60	NCC	Serum	TR-FIA	–	23.5 ^a
Kilkinen [135]	1986–1999	Finland	420	M	50–69	Case-Cohort	Serum	GC-MS	–	18.1 ^a
Vanharanta [136]	1995	Finland	100	M	58.6 ^a	CT	Serum	TR-FIA	–	16.6 ^a
Kilkinen [137]	1997	Finland	1,168	M	25–64	C-S	Serum	TR-FIA	–	13.8 ^b
			1,212	F					–	16.6 ^b
Vanharanta [138]	1998–2000	Finland	1,889	M	42–60	Cohort	Serum	TR-FIA	–	17.1 ^a
Hedelin [91]	2002	Sweden	1,130	M	67.8 ^a	PCC	Plasma	TR-FIA	–	24.0 ^a
Sonestedt [139]	1991–1996	Sweden	728	F	56.3 ^a	NCC	Plasma	TR-FIA	–	16.3 ^b
Stattin [140]	2001	Sweden	525	M	59.9 ^a	NCC	Plasma	TR-FIA	–	15.0 ^a
Lin [141]	2003–2004	Sweden	135	F	55–75	Cohort	Serum	TR-FIA	–	23.2 ^a
Hultén [142]	1986–1994	Sweden	308	F	51.2 ^a	NCC	Plasma	TR-FIA	–	22.9 ^a
	1995–2000		185		58.1 ^a				–	20.4 ^a
Aarestrup [143]	1993–1997	Denmark	149	F	50–64	Case-Cohort	Plasma	TR-FIA	–	31.0 ^a
Eriksen [144]	1993–1997	Denmark	850	F (40%)	50–64	Case-Cohort	Plasma	LC-MS/MS	–	10.9 ^b
Johnsen [88]	1993–1997	Denmark	857	F	50–64	NCC	Plasma	TR-FIA	–	38.0 ^a
Kuijsten [69]	–	Netherlands	3	F (25%)	28–53	C-S	Plasma	LC-MS	7.0 ^a	39.2 ^a
Milder [30]	1997–2002	Netherlands	637	F (55%)	19–75	PCC	Plasma	LC-MS/MS	1.4 ^a	11.3 ^a
Verheus [145]	1993–1997	Netherlands	87	F PreM	51.6 ^a	NCC	Plasma	LC-MS	0.6 ^a	8.9 ^a
			296	F PostM	58.6 ^a				0.6 ^a	8.9 ^a
Heald [146]	1998 2001	Scotland	483	M	50–74	PCC	Serum	GC-MS	–	16.2 ^b
Bhakta [92]	1995–1999	UK	58	F	25–75	PCC	Plasma	TR-FIA	–	13.7 ^a
Ward [72]	1993–1997	UK	815	M	45–75	NCC	Serum	LC-MS	0.7 ^b	18.1 ^b
			877	F (43%)					0.3 ^b	17.4 ^b

Table 3 (continued)

Reference	Data collection	Country	N	Sex	Age	Type of study	Specimen	Methods	END (nmol/L)	ENL (nmol/L)
Morton [85]	–	UK	36	M	41–74	C-S	Plasma	GC-MS	–	13.1 ^a
Low [73]	–	Portugal	50		35–71				1.2 ^a	13.1 ^a
Grace [74]	1993–1997	UK	109	F	45–76	NCC	Serum	GC-MS	1.3 ^a	12.4 ^a
Low [75]	1993–1997	UK	187	F	45–75	NCC	Plasma	LC-MS	1.3 ^a	12.8 ^a
Xie [81]	1993–1997	UK	267	M	45–75	Cohort	Plasma	LC-MS/MS	1.0 ^a	12.8 ^a
Bhakta [93]	1996–1999	US	802	F	25–42	NCC	Plasma	LC-MS	–	11.5 ^b
	–	UK	40	F	25–75	PCC	Plasma	TR-FIA	–	28.5 ^a
	–	UK (Asian)	100						–	13.9 ^a
Piller [147]	1992–1995	Germany	237	F	≤ 50	PCC	Plasma	TR-FIA	–	9.7 ^a
Zeleniuch-Jacquotte [79]	1985–1991	US	60	F	34–65	Cohort	Serum	CG-MS	1.5 ^b	21.2 ^b
Valentin-Biasimi [80]	1988–1994	US	199	F (61%)	20–58	C-S	Serum	LC-MS	6.0 ^a	11.9 ^a
Horner [148]	–	US	78	M	20–40	C-S	Plasma	TR-FIA	–	11.0 ^c
	–		115	F					–	13.3 ^c
Uehar [65]	–	Japan	111	F	40–60	C-S	Plasma	TR-FIA	–	13.3 ^a
Morton [87]	–	Japan	102	M	40–85	C-S	Plasma	GC-MS	–	32.7 ^a
	–		125	F	40–89				–	22.8
Morton [85]	–	China	53	M	31–85	C-S	Plasma	GC-MS	5.6 ^a	20.8 ^a
Liu [84]	2010–2012	China	264	F (71%)	35–60	NCC	Plasma	LC-MS	16.4 ^b	2.0 ^b
Ko [149]	1993–2004	Korea	206	F	60.4 ^a	HCC	Plasma	LC-MS	–	249.3 ^a
	–		185	M					–	177.8 ^a
	2003–2007	Vietnam	114	F	54.5 ^a				–	10.2 ^a
	–		92	M					–	10.4 ^a

ENL and END concentrations have been converted into nmol/L from the original studies

C-S cross-sectional; CT clinical trial; END enteridiol; ENL enterolactone; HCC hospital-based case-control; GC-MS gas chromatography-mass spectrometry; LC-MS liquid chromatography-mass spectrometry; NCC nested case-control; PCC population-based case-control; PostM post-menopausal; PreM pre-menopausal; TR-FIA sensitive time-resolved fluoroimmunoassay

^{a,b,c}Type of estimation: ^aMean, ^bMedian, ^cGeometric mean

^dEurope: Denmark, France, Germany, Greece, Italy, Netherlands, Spain, Sweden, UK

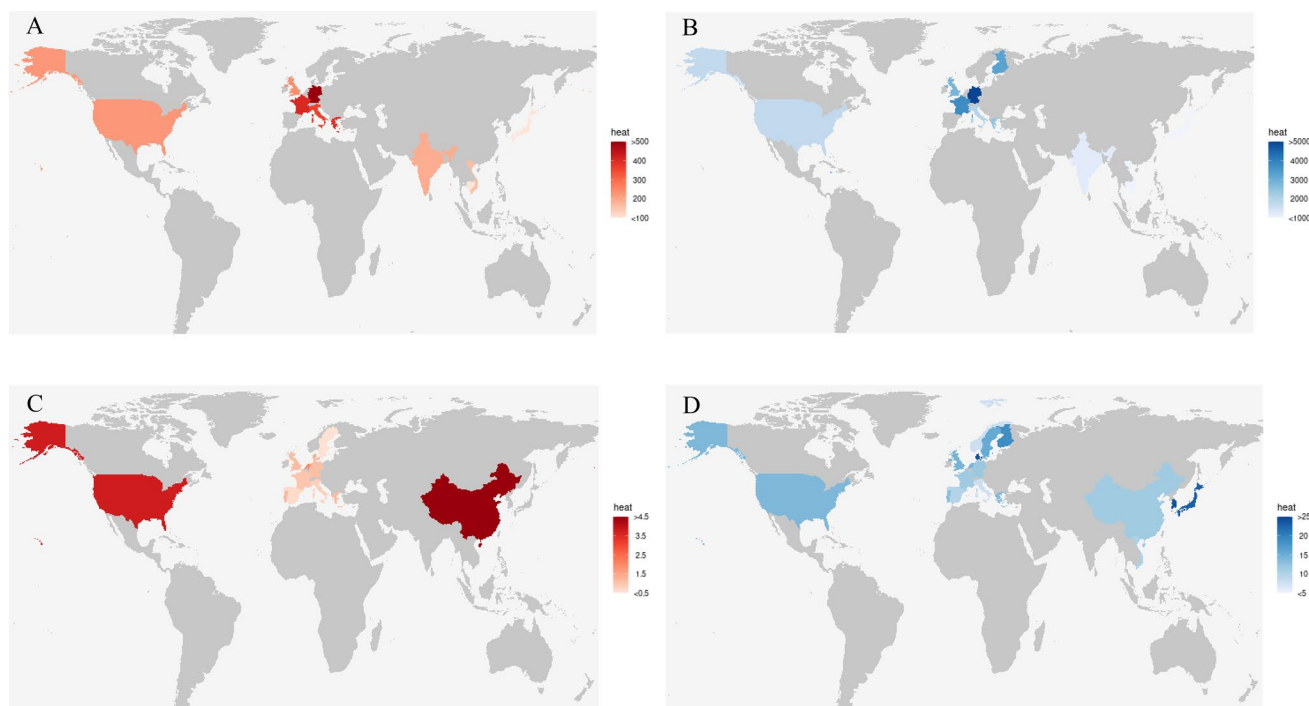


Fig. 3 Mean of means/medians of urinary and blood enterolignan concentrations (nmol/L) by country; **A** urinary enterolactone, **B** urinary enterodiols, **C** blood enterolactone, **D** blood enterodiols

enterolignan concentrations were found in Mediterranean countries: 0.3 nmol/L for END and 6.7–7.8 nmol/L for ENL [77]. Italy was the Mediterranean country with the highest END (1.3 nmol/L) and ENL (9.1 nmol/L) concentrations in plasma [77], which is similar to intake estimations.

Americas

To our knowledge, only US data were available from both North and South American continents, with the exception of a Jamaican study. In the US, several studies assessed enterolignan concentrations in urine ($n = 15$) (Table 2), plasma ($n = 2$), and serum ($n = 2$) (Table 3). Both urinary END and ENL excretions varied considerably among US studies from 38 [64] to 609 nmol/L [67] for END, and from 285 [64] to 3651 nmol/L [67] for ENL. Indeed, US populations included the worldwide minimum mean of END levels (285 nmol/L) and the worldwide maximum mean of ENL excretions (609 nmol/L). In the Jamaican study, the mean intake of END was in the upper side of the interval of the US studies (2671 nmol/L) [78].

Similarly, a high variability in blood END levels was observed among US studies, ranging between 1.5 [79] and 6.0 nmol/L [80] while the range of mean levels for ENL was narrower from 11.5 [81] to 22.5 nmol/L [79].

Asia

To date, urinary concentrations of enterolignans in Asia were measured in Singapore [82], Japan [66, 83], Vietnam [83], Cambodia [83] and India [83]. The mean of urinary END concentrations varied from 60 nmol/L in Cambodia [83] to 245 nmol/L [83] in Vietnam. For ENL, the highest mean value was found in Vietnam (1678 nmol/L) [83] while the lowest excretion was identified in a Japanese study (148 nmol/L) [66].

Several studies in East Asia (such as Japan, China, Korea and Vietnam) assessed enterolignans in plasma and showed a relatively low variation in their mean concentrations (~threefold variation). Thus, END concentration means ranged from 2.0 [84] to 5.6 nmol/L [85] in the two Chinese studies. Mean ENL concentrations in blood samples were between 10.2 [86] and 32.7 nmol/L [87] in Vietnam and Japan, respectively. In the study of Liu et al. [84] median plasma concentrations of ENL (2.0 nmol/L) and END (16.4 nmol/L) seem to be exchanged. Mean ENL concentrations in Korea were extremely high (177.8 nmol/L in women and 249.3 nmol/L in men), around tenfold higher than values found in any other study from other continents.

Determinants of the total enterolignans concentrations

Data from studies that analysed separately men and women showed that urinary concentrations of enterolignans were slightly higher in women than in men [67, 70, 83], with one exception [64]. Urinary ENL and END excretions were the highest in adults (20–60 years), followed by the elderly (> 60 years) and, finally, by adolescents (12–19 years) [67]. This pattern according to age and sex is consistent with findings from dietary lignans adjusted for energy intake. A Danish study suggested that smoking and higher BMI were associated with lower concentrations of ENL [88]. No other information was found for concentrations of enterolignans (in both urine and blood) and other determinants, such as educational level and physical activity.

Strengths and limitations

Dietary data

The main limitation of this review was that each study used a different methodology to estimate lignan intake. First, differences in both the type of dietary questionnaire (FFQ, 24 h dietary recall, history of diet) and the amount of food items included in the questionnaire could complicate comparisons in the habitual estimation of individual foods, particularly lignan-rich products. Although, the vast majority of studies used validated FFQs; very few of these questionnaires were specifically validated for lignans. Secondly, available food composition tables/databases were not complete. They have missing data on several foods and, especially, on some individual lignans. Only Phenol-Explorer [17] contains data on all commonly consumed lignans; while others only have data on two (MATA and SECO) or four individual lignans (MATA, SECO, LARI, and PINO). These four lignans are the most abundant ones accounting for at least 50% of total lignan intake in Europe [9]. Thirdly, most of the presented studies were not representative of the entire population, so the results may not be totally generalizable. However, the inclusion of several medium-to-large size studies from the same geographical area enhances generalizability. Fourth, studies evaluating the reliability of enterolignans as biomarkers of lignan intake are limited; especially those investigating all individual lignans, and correlations were moderate for urinary concentrations [27–29] and low for plasma/serum concentrations [31]. Therefore, inconsistent results have been observed comparing results using dietary conventional dietary questionnaires and biomarkers. For example, a recent meta-analysis showed no associations between dietary lignan intake and cancer outcomes; while a higher

concentration of serum/plasma ENL was inversely associated with overall cancer survival [89].

Biomarker data

Variability in results due to differences in procedures and methods in the analysis of concentrations of enterolignans in blood and urine were relatively minor, since all analytical methodologies were validated. The main limitation was that the studies only analyzed one sample per subject. It is well-known that enterolignans are relatively short-term nutritional biomarkers [11] and therefore multiple measurements would be recommended to estimate habitual exposure at an individual level. However, the mean of a single punctual measure in a large quantity of subjects was a suitable way to reflect the habitual mean of lignan concentrations at population level. Another limitation was the relatively small size of all studies and therefore the limited generalizability of the results.

Conclusions

Overall, common mean intakes of total lignans worldwide ranged from 1 to 5 mg/d, with a higher intake in vegetarian populations (9.1 mg/d). There was a large heterogeneity in the estimations of lignan intake across studies partially due to real differences among geographical areas and populations and to differences between dietary assessment methods used. Food sources also varied across regions, although the most typical ones were whole-grain cereal products, seeds, vegetables, and fruits.

As expected, similar trends and differences between regions were observed using dietary and biomarker data. END concentrations were usually tenfold lower than ENL levels in both urine and blood. Results of enterolignans in plasma and serum were equivalent. END and ENL concentrations in urine were approximately 100 times higher than in blood.

More food composition data are warranted to update current databases on lignans and improve dietary intake estimations. Data from some regions, particularly in low- and middle-income countries (Africa, Latin America, and some areas in Asia), was scarce or null; therefore, further studies combining both dietary and biomarker data in these regions are requested to improve data coverage globally.

Finally, an accurate estimation of lignan exposure is essential to better understand associations between lignan intake and the risk of chronic diseases. In our opinion, although, current estimations of dietary lignan intake are getting more precise, they are often underestimated. Thus, concentrations of enterolignans in blood and urine are still preferable to estimate lignan exposure in epidemiological

studies. This data will be crucial for setting and improving current dietary recommendations for populations.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s00394-021-02736-4>.

Funding This research was funded by the Instituto de Salud Carlos III through the grants CP15/00100 and PI18/00191 (Co-funded by the European Regional Development Fund. ERDF, a way to build Europe); and by La Marató de TV3 (project 201943-30). We thank CERCA Program/Generalitat de Catalunya for institutional support. LR-B, MF-N and RZ-R would like to thank the program were supported by the “PFIS” (FI20/00006), “Sara Borrell” (CD20/00036) and the “Miguel Servet” (CPII20/00009) programs from the Institute of Health Carlos III (Co-funded by the European Social Fund (ESF)—ESF investing in your future), respectively.

Declarations

Conflict of interest The authors are not aware of any conflicts of interest.

References

- Peterson J, Dwyer J, Adlercreutz H, Scalbert A, Jacques P, McCullough ML (2010) Dietary lignans: physiology and potential for cardiovascular disease risk reduction. *Nutr Rev* 68:571–603. <https://doi.org/10.1017/S0007114515005012>
- Kuiper GG, Lemmen JG, Carlsson B, Corton JC, Safe SH, van der Saag PT, van der Burg B, Gustafsson JA (1998) Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor beta. *Endocrinology* 139:4252–4263
- Grosso G, Micek A, Godos J, Pajak A, Sciacca S, Galvano F, Giovannucci EL (2017) Dietary flavonoid and lignan intake and mortality in prospective cohort studies: systematic review and dose-response meta-analysis. *Am J Epidemiol* 185(12):1304–1316. <https://doi.org/10.1093/aje/kww207>
- Touillaud MS, Thiébaud AC, Fournier A, Niravong M, Boutron-Ruault MC, Clavel-Chapelon F (2007) Dietary lignan intake and postmenopausal breast cancer risk by estrogen and progesterone receptor status. *J Natl Cancer Inst* 99(6):475–486. <https://doi.org/10.1093/jnci/djk096>
- Buja A, Pierbon M, Lago L, Grotto G, Baldo V (2020) Breast cancer primary prevention and diet: an umbrella review. *Int J Environ Res Public Health* 17(13):4731. <https://doi.org/10.3390/ijerph17134731>
- Ma ZP, Zhang ZF, Yang YF, Yang Y (2019) Sesamin promotes osteoblastic differentiation and protects rats from osteoporosis. *Med Sci Monit* 25:5312–5320. <https://doi.org/10.12659/MSM.915529>
- Pruthi S, Qin R, Terstreip SA et al (2012) A phase III, randomized, placebo-controlled, double-blind trial of flaxseed for the treatment of hot flashes: North Central Cancer Treatment Group N08C7. *Menopause* 19(1):48–53. <https://doi.org/10.1097/gme.0b013e318223b021>
- Adlercreutz H (2007) Lignans and human health. *Crit Rev Clin Lab Sci* 44:483–525
- Zamora-Ros R, Knaze V, Rothwell JA et al (2016) Dietary polyphenol intake in Europe: the European Prospective Investigation into Cancer and Nutrition (EPIC) study. *Eur J Nutr* 55:1359–1375. <https://doi.org/10.1007/s00394-015-0950-x>
- Smeds AI, Eklund PC, Sjöholm RE, Willför SM, Nishibe S, Deyama T, Holmbom BR (2007) Quantification of a broad spectrum of lignans in cereals, oilseeds, and nuts. *J Agric Food Chem* 55:1337–1346
- Clavel T, Doré J, Blaut M (2006) Bioavailability of lignans in human subjects. *Nutr Res Rev* 19:187–196. <https://doi.org/10.1017/S0954422407249704>
- Heinonen S, Nurmi T, Liukkonen K, Poutanen K, Wähälä K, Deyama T, Nishibe S, Adlercreutz H (2001) In vitro metabolism of plant lignans: new precursors of mammalian lignans enterolactone and enterodiol. *J Agric Food Chem* 49:3178–3186
- Nurmi T, Voutilainen S, Nyyssönen K, Adlercreutz H, Salonen JT (2003) Liquid chromatography method for plant and mammalian lignans in human urine. *J Chromatogr B Analyt Technol Biomed Life Sci* 798:101–110
- Pérez-Jiménez J, Neveu V, Vos F, Scalbert A (2010) Systematic analysis of the content of 502 polyphenols in 452 foods and beverages: an application of the phenol-explorer database. *J Agric Food Chem* 58:4959–4969. <https://doi.org/10.1021/jf100128b>
- Kuhnle GGC, Dell’Aquila C, Aspinall SM, Runswick SA, Mulligan AA, Bingham SA (2008) Phytoestrogen content of foods of animal origin: dairy products, eggs, meat, fish, and seafood. *J Agric Food Chem* 56:10099–10104. <https://doi.org/10.1021/jf801344x>
- Zamora-Ros R, Rabassa M, Llorach R, González CA, Andres-Lacueva C (2012) Application of dietary phenolic biomarkers in epidemiology: past, present, and future. *J Agric Food Chem* 60:6648–6657. <https://doi.org/10.1021/jf204742e>
- Neveu V, Perez-Jiménez J, Vos F, Crespy V, du Chaffaut L, Mennen L, Knox C, Eisner R, Cruz J, Wishart D, Scalbert A (2010) Phenol-Explorer: an online comprehensive database on polyphenol contents in foods. *Database (Oxford)* 2010:bap024. <https://doi.org/10.1093/database/bap024>
- Thompson LU, Boucher BA, Liu Z, Cotterchio M, Kreiger N (2006) Phytoestrogen content of foods consumed in Canada, including isoflavones, lignans, and coumestrol. *Nutr Cancer* 54:184–201
- Milder IEJ, Arts ICW, van de Putte B, Venema DP, Hollman PCH (2005) Lignan contents of Dutch plant foods: a database including lariciresinol, pinoresinol, secoisolariciresinol and matairesinol. *Br J Nutr* 93:393–402
- Kuhnle GGC, Dell’Aquila C, Aspinall SM, Runswick SA, Joosen AMCP, Mulligan AA, Bingham SA (2009) Phytoestrogen content of fruits and vegetables commonly consumed in the UK based on LC–MS and ¹³C-labelled standards. *Food Chem* 116:542–554
- Kuhnle GGC, Dell’Aquila C, Aspinall SM, Runswick SA, Mulligan AA, Bingham SA (2008) Phytoestrogen content of beverages, nuts, seeds, and oils. *J Agric Food Chem* 56:7311–7315. <https://doi.org/10.1021/jf801534g>
- Kuhnle GGC, Dell’Aquila C, Aspinall SM, Runswick SA, Mulligan AA, Bingham SA (2009) Phytoestrogen content of cereals and cereal-based foods consumed in the UK. *Nutr Cancer* 61:302–309. <https://doi.org/10.1080/01635580802567141>
- Valsta LM, Kilkkinen A, Mazur W, Nurmi T, Lampi A-M, Ovaskainen M-L, Korhonen T, Adlercreutz H, Pietinen P (2003) Phyto-oestrogen database of foods and average intake in Finland. *Br J Nutr* 89(Suppl 1):S31–S38
- Illner A-K, Freisling H, Boeing H, Huybrechts I, Crispim SP, Slimani N (2012) Review and evaluation of innovative technologies for measuring diet in nutritional epidemiology. *Int J Epidemiol* 41:1187–1203. <https://doi.org/10.1093/ije/dys105>
- Potischman N (2003) Biologic and methodologic issues for nutritional biomarkers. *J Nutr* 133(Suppl 3):875S–880S. <https://doi.org/10.1093/jn/133.3.875S>

26. Peeters PHM, Slimani N, van der Schouw YT et al (2007) Variations in plasma phytoestrogen concentrations in European adults. *J Nutr* 137:1294–1300
27. Pérez-Jiménez J, Hubert J, Hooper L, Cassidy A, Manach C, Williamson G, Scalbert A (2010) Urinary metabolites as biomarkers of polyphenol intake in humans: a systematic review. *Am J Clin Nutr* 92:801–809. <https://doi.org/10.3945/ajcn.2010.29924>
28. French MR, Thompson LU, Hawker GA (2007) Validation of a phytoestrogen food frequency questionnaire with urinary concentrations of isoflavones and lignan in premenopausal women. *J Am Coll Nutr* 26(1):76–82. <https://doi.org/10.1080/07315724.2007.10719588>
29. Horn-Ross PL, Barnes S, Lee VS et al (2006) Reliability and validity of an assessment of usual phytoestrogen consumption (United States). *Cancer Causes Control* 17(1):85–93. <https://doi.org/10.1007/s10552-005-0391-6>
30. Milder IEJ, Kuijsten A, Arts ICW, Feskens EJM, Kampman E, Hollman PC, Van 't Veer P (2007) Relation between plasma enterodiol and enterolactone and dietary intake of lignans in a Dutch endoscopy-based population. *J Nutr* 137:1266–1271
31. Lin Y, Wolk A, Håkansson N, Peñalvo JL, Lagergren J, Adlercreutz H, Lu Y (2013) Validation of FFQ-based assessment of dietary lignans compared with serum enterolactone in Swedish women. *Br J Nutr* 109(10):1873–1880. <https://doi.org/10.1017/S000711451200387X>
32. Liggins J, Grimwood R, Bingham SA (2000) Extraction and quantification of lignan phytoestrogens in food and human samples. *Anal Biochem* 287(1):102–109. <https://doi.org/10.1006/abio.2000.4811>
33. Zamora-Ros R, Biessy C, Rothwell JA, Monge A, Lajous M, Scalbert A, López-Ridaura R, Romieu I (2018) Dietary polyphenol intake and their major food sources in the Mexican Teachers' Cohort. *Br J Nutr* 120:353–360. <https://doi.org/10.1017/S0007114518001381>
34. Adriouch S, Kesse-Guyot E, Feuillet T, Touvier M, Olié V, Andreeva V, Hercberg S, Galan P, Fezeu LK (2018) Total and specific dietary polyphenol intakes and 6-year anthropometric changes in a middle-aged general population cohort. *Int J Obes (Lond)* 42:310–317. <https://doi.org/10.1038/ijo.2017.227>
35. Meija L, Söderholm P, Samaletdin A, Ignace G, Siksa I, Joffe R, Lejnieks A, Lietuviētis V, Krams I, Adlercreutz H (2013) Dietary intake and major sources of plant lignans in Latvian men and women. *Int J Food Sci Nutr* 64:535–543. <https://doi.org/10.3109/09637486.2013.765835>
36. Wisnuwardani R, Henauw S, Androustos O et al (2018) Estimated dietary intake of polyphenols in European adolescents: the HELENA study. *Eur J Nutr* 58:2345–2363. <https://doi.org/10.1007/s00394-018-1787-x>
37. Tetens I, Turrini A, Tapanainen H, Christensen T, Lampe JW, Fagt S, Håkansson N, Lundquist A, Hallund J, Valsta LM (2013) Dietary intake and main sources of plant lignans in five European countries. *Food Nutr Res*. <https://doi.org/10.3402/fnr.v57i0.19805>
38. Grosso G, Stepaniak U, Topor-Mądry R, Szafraniec K, Pająk A (2014) Estimated dietary intake and major food sources of polyphenols in the Polish arm of the HAPIEE study. *Nutrition* 30:1398–1403. <https://doi.org/10.1016/j.nut.2014.04.012>
39. Witkowska AM, Zujko ME, Waśkiewicz A, Terlikowska KM, Piotrowski W (2015) Comparison of various databases for estimation of dietary polyphenol intake in the population of Polish adults. *Nutrients* 7:9299–9308. <https://doi.org/10.3390/nu7115464>
40. Russo GI, Di Mauro M, Regis F, Reale G, Campisi D, Marranzano M, Lo Giudice A, Solinas T, Madonia M, Cimino S, Morgia G (2018) Association between dietary phytoestrogens intakes and prostate cancer risk in Sicily. *Aging Male* 21:48–54. <https://doi.org/10.1080/13685538.2017.1365834>
41. Plumb J, Pigat S, Bompola F, Cushen M, Pinchen H, Nørby E, Astley S, Lyons J, Kiely M, Finglas P (2017) eBASIS (bioactive substances in food information systems) and bioactive intakes: major updates of the bioactive compound composition and beneficial bioeffects database and the development of a probabilistic model to assess intakes in Europe. *Nutrients* 9:E320. <https://doi.org/10.3390/nu9040320>
42. Carmichael SL, Cogswell ME, Ma C, Gonzalez-Feliciano A, Olney RS, Correa A, Shaw GM (2013) Hypospadias and maternal intake of phytoestrogens. *Am J Epidemiol* 178:434–440. <https://doi.org/10.1093/aje/kws591>
43. Fink BN, Steck SE, Wolff MS, Kabat GC, Gammon MD (2006) Construction of a flavonoid database for assessing intake in a population-based sample of women on Long Island, New York. *Nutr Cancer* 56:57–66
44. Cotterchio M, Boucher BA, Manno M, Gallinger S, Okey A, Harper P (2006) Dietary phytoestrogen intake is associated with reduced colorectal cancer risk. *J Nutr* 136:3046–3053
45. Morisset A-S, Lemieux S, Veilleux A, Bergeron J, John Weisnagel S, Tchernof A (2009) Impact of a lignan-rich diet on adiposity and insulin sensitivity in post-menopausal women. *Br J Nutr* 102:195–200. <https://doi.org/10.1017/S0007114508162092>
46. Hernández-Ramírez RU, Galván-Portillo MV, Ward MH, Agudo A, González CA, Oñate-Ocaña LF, Herrera-Goepfert R, Palma-Coca O, López-Carrillo L (2009) Dietary intake of polyphenols, nitrate and nitrite and gastric cancer risk in Mexico City. *Int J Cancer* 125:1424–1430. <https://doi.org/10.1002/ijc.24454>
47. Nascimento-Souza MA, de Paiva PG, Pérez-Jiménez J, do Carmo Castro Franceschini S, Ribeiro AQ, (2018) Estimated dietary intake and major food sources of polyphenols in elderly of Viçosa, Brazil: a population-based study. *Eur J Nutr* 57:617–627. <https://doi.org/10.1007/s00394-016-1348-0>
48. Miranda AM, Steluti J, Fisberg RM, Marchioni DM (2016) Association between polyphenol intake and hypertension in adults and older adults: a population-based study in Brazil. *PLoS ONE* 11:e0165791. <https://doi.org/10.1371/journal.pone.0165791>
49. Miranda AM, Steluti J, Fisberg RM, Marchioni DM (2016) Dietary intake and food contributors of polyphenols in adults and elderly adults of Sao Paulo: a population-based study. *Br J Nutr* 115:1061–1070. <https://doi.org/10.1017/S0007114515005061>
50. Lahmann PH, Hughes MC, Ibiebele TI, Mulligan AA, Kuhnle GGC, Webb PM (2012) Estimated intake of dietary phyto-oestrogens in Australian women and evaluation of correlates of phyto-oestrogen intake. *J Nutr Sci* 1:e11. <https://doi.org/10.1017/jns.2012.11>
51. Hanna KL, O'Neill S, Lyons-Wall PM (2010) Intake of isoflavone and lignan phytoestrogens and associated demographic and lifestyle factors in older Australian women. *Asia Pac J Clin Nutr* 19:540–549
52. Sohrab G, Hosseinpour-Niazi S, Hejazi J, Yuzbashian E, Mirmiran P, Azizi F (2013) Dietary polyphenols and metabolic syndrome among Iranian adults. *Int J Food Sci Nutr* 64:661–667. <https://doi.org/10.3109/09637486.2013.787397>
53. Sohrab G, Ebrahimof S, Hosseinpour-Niazi S, Yuzbashian E, Mirmiran P, Azizi F (2018) Association of dietary intakes of total polyphenol and its subclasses with the risk of metabolic syndrome: tehran lipid and glucose study. *Metab Syndr Relat Disord* 16:274–281. <https://doi.org/10.1089/met.2017.0140>
54. Jang J-H, Yoon J-Y, Cho S-H (2007) Intake of dietary phytoestrogen and indices of antioxidant and bone metabolism of pre- and post-menopausal Korean women. *Nutr Res Pract* 1:305–312. <https://doi.org/10.4162/nrp.2007.1.4.30>
55. Suzuki R, Rylander-Rudqvist T, Saji S, Bergkvist L, Adlercreutz H, Wolk A (2008) Dietary lignans and postmenopausal breast

- cancer risk by oestrogen receptor status: a prospective cohort study of Swedish women. *Br J Cancer* 98:636–640. <https://doi.org/10.1038/sj.bjc.6604175>
56. Peñalvo JL, Moreno-Franco B, Ribas-Barba L, Serra-Majem L (2012) Determinants of dietary lignan intake in a representative sample of young Spaniards: association with lower obesity prevalence among boys but not girls. *Eur J Clin Nutr* 66:795–798. <https://doi.org/10.1038/ejcn.2012.45>
 57. Kilkkinen A, Valsta LM, Virtamo J, Stumpf K, Adlercreutz H, Pietinen P (2003) Intake of lignans is associated with serum enterolactone concentration in Finnish men and women. *J Nutr* 133:1830–1833
 58. Pounis G, Di Castelnuovo A, Bonaccio M, Costanzo S, Persichillo M, Krogh V, Donati MB, de Gaetano G, Iacoviello L (2016) Flavonoid and lignan intake in a Mediterranean population: proposal for a holistic approach in polyphenol dietary analysis, the Moli-sani Study. *Eur J Clin Nutr* 70:338–345. <https://doi.org/10.1038/ejcn.2015.178>
 59. Horn-Ross PL, John EM, Canchola AJ, Stewart SL, Lee MM (2003) Phytoestrogen intake and endometrial cancer risk. *J Natl Cancer Inst* 95:1158–1164
 60. Schabath MB, Hernandez LM, Wu X, Pillow PC, Spitz MR (2005) Dietary phytoestrogens and lung cancer risk. *JAMA* 294:1493–1504
 61. Milder IE, Feskens EJ, Arts IC, Bueno de Mesquita HB, Hollman PC, Kromhout D (2005) Intake of the plant lignans secoisolariciresinol, matairesinol, lariciresinol, and pinoresinol in Dutch men and women. *J Nutr* 135:1202–1207
 62. Hedelin M, Löf M, Andersson TM-L, Adlercreutz H, Weiderpass E (2011) Dietary phytoestrogens and the risk of ovarian cancer in the women's lifestyle and health cohort study. *Cancer Epidemiol Biomark Prev* 20:308–317. <https://doi.org/10.1158/1055-9965.EPI-10-0752>
 63. Hedelin M, Löf M, Sandin S, Adami H-O, Weiderpass E (2016) Prospective study of dietary phytoestrogen intake and the risk of colorectal cancer. *Nutr Cancer* 68:388–395. <https://doi.org/10.1080/01635581.2016.1152380>
 64. Rybak ME, Sternberg MR, Pfeiffer CM (2013) Sociodemographic and lifestyle variables are compound- and class-specific correlates of urine phytoestrogen concentrations in the U.S. population. *J Nutr* 143:986S–994S. <https://doi.org/10.3945/jn.112.172981>
 65. Uehar M, Arai Y, Watanabe S, Adlercreutz H (2000) Comparison of plasma and urinary phytoestrogens in Japanese and Finnish women by time-resolved fluoroimmunoassay. *BioFactors* 12:217–225
 66. Liu W, Tanabe M, Harada KH, Koizumi A (2013) Levels of urinary isoflavones and lignan polyphenols in Japanese women. *Environ Health Prev Med* 18:394–400. <https://doi.org/10.1007/s12199-013-0338-6>
 67. Xu C, Liu Q, Zhang Q, Gu A, Jiang Z-Y (2015) Urinary enterolactone is associated with obesity and metabolic alteration in men in the US National Health and Nutrition Examination Survey 2001–10. *Br J Nutr* 113:683–690. <https://doi.org/10.1017/S0007114514004115>
 68. Perez-Cornago A, Appleby PN, Boeing H et al (2018) Circulating isoflavone and lignan concentrations and prostate cancer risk: a meta-analysis of individual participant data from seven prospective studies including 2,828 cases and 5,593 controls. *Int J Cancer* 143:2677–2686. <https://doi.org/10.1002/ijc.31640>
 69. Kuijsten A, Buijsman MN, Arts IC, Mulder PP, Hollman PC (2005) A validated method for the quantification of enterodiol and enterolactone in plasma using isotope dilution liquid chromatography with tandem mass spectrometry. *J Chromatogr B Analyt Technol Biomed Life Sci* 822:178–184
 70. Krogholm KS, Bysted A, Brantsæter AL, Jakobsen J, Rasmussen SE, Kristoffersen L, Toft U (2012) Evaluation of flavonoids and enterolactone in overnight urine as intake biomarkers of fruits, vegetables and beverages in the Inter99 cohort study using the method of triads. *Br J Nutr* 108:1904–1912. <https://doi.org/10.1017/S0007114512000104>
 71. Zamora-Ros R, Achaintre D, Rothwell JA et al (2016) Urinary excretions of 34 dietary polyphenols and their associations with lifestyle factors in the EPIC cohort study. *Sci Rep* 6:26905. <https://doi.org/10.1038/srep26905>
 72. Ward H, Chapelais G, Kuhnle GG, Luben R, Khaw K-T, Bingham S (2008) Lack of prospective associations between plasma and urinary phytoestrogens and risk of prostate or colorectal cancer in the European Prospective into Cancer-Norfolk study. *Cancer Epidemiol Biomark Prev* 17:2891–2894. <https://doi.org/10.1158/1055-9965.EPI-08-0335>
 73. Low Y-L, Taylor JI, Grace PB, Dowsett M, Scollen S, Dunning AM, Mulligan AA, Welch AA, Luben RN, Khaw KT, Day NE, Wareham NJ, Bingham SA (2005) Phytoestrogen exposure correlation with plasma estradiol in postmenopausal women in European Prospective Investigation of Cancer and Nutrition-Norfolk may involve diet-gene interactions. *Cancer Epidemiol Biomark Prev* 14:213–220
 74. Grace PB, Taylor JI, Low Y-L, Luben RN, Mulligan AA, Botting NP, Dowsett M, Welch AA, Khaw KT, Wareham NJ, Day NE, Bingham SA (2004) Phytoestrogen concentrations in serum and spot urine as biomarkers for dietary phytoestrogen intake and their relation to breast cancer risk in European prospective investigation of cancer and nutrition-norfolk. *Cancer Epidemiol Biomark Prev* 13:698–708
 75. Low Y-L, Taylor JI, Grace PB, Dowsett M, Folkard E, Doody D, Dunning AM, Scollen S, Mulligan AA, Welch AA, Luben RN, Khaw KT, Day NE, Wareham NJ, Bingham SA (2005) Polymorphisms in the CYP19 gene may affect the positive correlations between serum and urine phytoestrogen metabolites and plasma androgen concentrations in men. *J Nutr* 135:2680–2686
 76. Durazzo A, Carcea M, Adlercreutz H et al (2014) Effects of consumption of whole grain foods rich in lignans in healthy postmenopausal women with moderate serum cholesterol: a pilot study. *Int J Food Sci Nutr* 65:637–645. <https://doi.org/10.3109/09637486.2014.893283>
 77. Travis RC, Spencer EA, Allen NE et al (2009) Plasma phytoestrogens and prostate cancer in the European prospective investigation into cancer and nutrition. *Br J Cancer* 100:1817–1823. <https://doi.org/10.1038/sj.bjc.6605073>
 78. Simon GA, Fletcher HM, Golden K, McFarlane-Anderson ND (2015) Urinary isoflavone and lignan phytoestrogen levels and risk of uterine fibroid in Jamaican women. *Maturitas* 82:170–175. <https://doi.org/10.1016/j.maturitas.2015.06.041>
 79. Zeleniuch-Jacquotte A, Adlercreutz H, Akhmedkhanov A, Toniolo P (1998) Reliability of serum measurements of lignans and isoflavonoid phytoestrogens over a two-year period. *Cancer Epidemiol Biomark Prev* 7:885–889
 80. Valentín-Blasini L, Blount BC, Caudill SP, Needham LL (2003) Urinary and serum concentrations of seven phytoestrogens in a human reference population subset. *J Expo Anal Environ Epidemiol* 13:276–282
 81. Xie J, Tworoger SS, Franke AA, Terry KL, Rice MS, Rosner BA, Willett WC, Hankinson SE, Eliassen AH (2013) Plasma enterolactone and breast cancer risk in the nurses' health study II. *Breast Cancer Res Treat* 139:801–809. <https://doi.org/10.1007/s10549-013-2586-y>
 82. Talaei M, Lee BL, Ong CN, van Dam RM, Yuan JM, Koh WP, Pan A (2016) Urine phyto-oestrogen metabolites are not significantly associated with risk of type 2 diabetes: the Singapore

- Chinese health study. *Br J Nutr* 115:1607–1615. <https://doi.org/10.1017/S0007114516000581>
83. Kunisue T, Tanabe S, Isobe T, Aldous KM, Kannan K (2010) Profiles of phytoestrogens in human urine from several Asian countries. *J Agric Food Chem* 58:9838–9846. <https://doi.org/10.1021/jf102253j>
 84. Liu J, Mi S, Du L, Li X, Li P, Jia K, Zhao J, Zhang H, Zhao W, Gao Y (2018) The associations between plasma phytoestrogens concentration and metabolic syndrome risks in Chinese population. *PLoS ONE* 13:e0194639. <https://doi.org/10.1371/journal.pone.0194639>
 85. Morton MS, Chan PS, Cheng C, Blacklock N, Matos-Ferreira A, Abranches-Monteiro L, Correia R, Lloyd S, Griffiths K (1997) Lignans and isoflavonoids in plasma and prostatic fluid in men: samples from Portugal, Hong Kong, and the United Kingdom. *Prostate* 32:122–128
 86. Ko K-P, Yeo Y, Yoon J-H, Kim C-S, Tokudome S, Ngoan LT, Koriyama C, Lim YK, Chang SH, Shin HR, Kang D, Park SK, Kang CH, Yoo KY (2018) Plasma phytoestrogens concentration and risk of colorectal cancer in two different Asian populations. *Clin Nutr* 37:1675–1682. <https://doi.org/10.1016/j.clnu.2017.07.014>
 87. Morton MS, Arisaka O, Miyake N, Morgan LD, Evans BA (2002) Phytoestrogen concentrations in serum from Japanese men and women over forty years of age. *J Nutr* 132:3168–3171
 88. Johnsen NF, Hausner H, Olsen A, Tetens I, Christensen J, Knudsen KE, Overvad K, Tjønnelund A (2004) Intake of whole grains and vegetables determines the plasma enterolactone concentration of Danish women. *J Nutr* 134:2691–2697. <https://doi.org/10.1093/jn/134.10.2691>
 89. Micek A, Godos J, Brzostek T et al (2021) Dietary phytoestrogens and biomarkers of their intake in relation to cancer survival and recurrence: a comprehensive systematic review with meta-analysis. *Nutr Rev* 79(1):42–65. <https://doi.org/10.1093/nutrit/nuaa043>
 90. Nurmi T, Mursu J, Peñalvo JL, Poulsen HE, Voutilainen S (2010) Dietary intake and urinary excretion of lignans in Finnish men. *Br J Nutr* 103:677–685. <https://doi.org/10.1017/S0007114509992261>
 91. Hedelin M, Klint A, Chang ET, Bellocchio R, Johansson J-E, Andersson SO, Heinonen SM, Adlercreutz H, Adami HO, Grönberg H, Bälter KA (2006) Dietary phytoestrogen, serum enterolactone and risk of prostate cancer: the cancer prostate Sweden study (Sweden). *Cancer Causes Control* 17:169–180
 92. Bhakta D, dos Santos SI, Higgins C, Sevak L, Kassam-Khamis T, Mangtani P, Adlercreutz H, McMichael A (2005) A semi-quantitative food frequency questionnaire is a valid indicator of the usual intake of phytoestrogens by south Asian women in the UK relative to multiple 24-h dietary recalls and multiple plasma samples. *J Nutr* 135:116–123
 93. Bhakta D, Higgins CD, Sevak L, Mangtani P, Adlercreutz H, McMichael AJ, dos Santos SI (2006) Phyto-oestrogen intake and plasma concentrations in South Asian and native British women resident in England. *Br J Nutr* 95:1150–1158
 94. Mulligan AA, Kuhnle GG, Lentjes MA, van Scheltinga V, Powell NA, McTaggart A, Bhaniani A, Khaw KT (2013) Intakes and sources of isoflavones, lignans, enterolignans, coumestrol and soya-containing foods in the Norfolk arm of the European Prospective Investigation into Cancer and Nutrition (EPIC-Norfolk), from 7 d food diaries, using a newly updated database. *Public Health Nutr* 16:1454–1462. <https://doi.org/10.1017/S1368980012003904>
 95. Witkowska AM, Waśkiewicz A, Zujko ME, Szcześniewska D, Stepaniak U, Pająk A, Drygas W (2018) Are total and individual dietary lignans related to cardiovascular disease and its risk factors in postmenopausal Women? A nationwide study. *Nutrients* 10:865. <https://doi.org/10.3390/nu10070865>
 96. Linseisen J, Piller R, Hermann S, Chang-Claude J (2004) Dietary phytoestrogen intake and premenopausal breast cancer risk in a German case-control study. *Int J Cancer* 110:284–290
 97. Boker LK, Van der Schouw YT, De Kleijn MJ, Jacques PF, Grobbee DE, Peeters PH (2002) Intake of dietary phytoestrogens by Dutch women. *J Nutr* 132:1319–1328
 98. Milder IE, Feskens EJ, Arts IC, Bueno-de-Mesquita HB, Holman PC, Kromhout D (2006) Intakes of 4 dietary lignans and cause-specific and all-cause mortality in the Zutphen elderly study. *Am J Clin Nutr* 84:400–405
 99. Pérez-Jiménez J, Fezeu L, Touvier M, Arnault N, Manach C, Hercberg S, Galan P, Scalbert A (2011) Dietary intake of 337 polyphenols in French adults. *Am J Clin Nutr* 93:1220–1228. <https://doi.org/10.3945/ajcn.110.007096>
 100. Lefèvre-Arbogast S, Gaudout D, Bensalem J, Letenneur L, Dartigues JF, Hejblum BP, Féart C, Delcourt C, Samieri C (2019) Pattern of polyphenol intake and the long-term risk of dementia in older persons. *Neurology* 90:e1979–e1988. <https://doi.org/10.1212/WNL.0000000000005607>
 101. Pellegrini N, Valtueña S, Ardigò D, Brighenti F, Franzini L, Del Rio D, Scazzina F, Piatti PM, Zavaroni I (2010) Intake of the plant lignans matairesinol, secoisolariciresinol, pinoresinol, and lariciresinol in relation to vascular inflammation and endothelial dysfunction in middle age-elderly men and post-menopausal women living in Northern Italy. *Nutr Metab Cardiovasc Dis* 20:64–71. <https://doi.org/10.1016/j.numecd.2009.02.003>
 102. Godos J, Marventano S, Mistretta A, Galvano F, Grosso G (2017) Dietary sources of polyphenols in the mediterranean healthy eating, aging and lifestyle (MEAL) study cohort. *Int J Food Sci Nutr* 68:750–756. <https://doi.org/10.1080/09637486.2017.1285870>
 103. Godos J, Bergante S, Satriano A, Pluchinotta FR, Marranzano M (2018) Dietary phytoestrogen intake is inversely associated with hypertension in a cohort of adults living in the mediterranean area. *Molecules* 23:E368. <https://doi.org/10.3390/molecules23020368>
 104. González S, Fernández M, Cuervo A, Lasheras C (2014) Dietary intake of polyphenols and major food sources in an institutionalised elderly population. *J Hum Nutr Diet* 27:176–183. <https://doi.org/10.1111/jhn.12058>
 105. Zamora-Ros R, Not C, Guinó E, Luján-Barroso L, García RM, Biondo S, Salazar R, Moreno V (2013) Association between habitual dietary flavonoid and lignan intake and colorectal cancer in a Spanish case-control study (the Bellvitge Colorectal Cancer Study). *Cancer Causes Control* 24:549–557. <https://doi.org/10.1007/s10552-012-9992-z>
 106. Tresserra-Rimbau A, Medina-Remón A, Pérez-Jiménez J (2013) Dietary intake and major food sources of polyphenols in a Spanish population at high cardiovascular risk: the PREDIMED study. *Nutr Metab Cardiovasc Dis* 23:953–959. <https://doi.org/10.1016/j.numecd.2012.10.008>
 107. Mendonça RD, Carvalho NC, Martin-Moreno JM, Pimenta AM, Lopes ACS, Gea A, Martinez-Gonzalez MA, Bes-Rastrollo M (2019) Total polyphenol intake, polyphenol subtypes and incidence of cardiovascular disease: the SUN cohort study. *Nutr Metab Cardiovasc Dis* 29:69–78. <https://doi.org/10.1016/j.numecd.2018.09.012>
 108. Petrick JL, Steck SE, Bradshaw PT, Chow W-H, Engel LS, He K, Risch HA, Vaughan TL, Gammon MD (2015) Dietary flavonoid intake and Barrett's esophagus in western Washington State. *Ann Epidemiol* 25:730–735. <https://doi.org/10.1016/j.annepidem.2015.05.010>
 109. Petrick JL, Steck SE, Bradshaw PT, Trivers KF, Abrahamson PE, Engel LS, He K, Chow WH, Mayne ST, Risch HA, Vaughan TL, Gammon MD (2015) Dietary intake of flavonoids

- and oesophageal and gastric cancer: incidence and survival in the United States of America (USA). *Br J Cancer* 112:1291–1300. <https://doi.org/10.1038/bjc.2015.25>
110. Williams AM, Bonner M, Ochs-Balcom HM, Hwang H, Morrison C, McCann SE (2015) Dietary lignan intake and androgen receptor expression in breast tumors. *Cancer Causes Control* 26:311–317. <https://doi.org/10.1007/s10552-014-0504-1>
 111. Waetjen LE, Leung K, Crawford SL, Huang M-H, Gold EB, Greendale GA (2013) The relationship between dietary phytoestrogens and development of urinary incontinence in midlife women. *Menopause* 20:428–436. <https://doi.org/10.1097/gme.0b013e3182703c9c>
 112. Bandera EV, King M, Chandran U, Paddock LE, Rodriguez-Rodriguez L, Olson SH (2011) Phytoestrogen consumption from foods and supplements and epithelial ovarian cancer risk: a population-based case control study. *BMC Womens Health* 11:40. <https://doi.org/10.1186/1472-6874-11-40>
 113. Chang ET, Canchola AJ, Clarke CA, Lu Y, West DW, Bernstein L, Wang SS, Horn-Ross PL (2011) Dietary phytochemicals and risk of lymphoid malignancies in the California Teachers Study cohort. *Cancer Causes Control* 22:237–249. <https://doi.org/10.1007/s10552-010-9692-5>
 114. McCann SE, Thompson LU, Nie J, Dorn J, Trevisan M, Shields PG, Ambrosone CB, Edge SB, Li HF, Kasprzak C, Freudenheim JL (2010) Dietary lignan intakes in relation to survival among women with breast cancer: the Western New York Exposures and Breast Cancer (WEB) Study. *Breast Cancer Res Treat* 122:229–235. <https://doi.org/10.1007/s10549-009-0681-x>
 115. Mervish NA, Teitelbaum SL, Pajak A, Windham GC, Pinney SM, Kushi LH, Biro FM, Wolff MS (2017) Peripubertal dietary flavonol and lignan intake and age at menarche in a longitudinal cohort of girls. *Pediatr Res* 82:201–208. <https://doi.org/10.1038/pr.2017.34>
 116. van der Schouw YT, Sampson L, Willett WC, Rimm EB (2005) The usual intake of lignans but not that of isoflavones may be related to cardiovascular risk factors in U.S. men. *J Nutr* 135:260–266
 117. Horn-Ross PL, Hoggatt KJ, Lee MM (2002) Phytoestrogens and thyroid cancer risk: the San Francisco Bay Area thyroid cancer study. *Cancer Epidemiol Prev Biomark* 11:43–49
 118. McCann SE, Freudenheim JL, Marshall JR, Graham S (2003) Risk of human ovarian cancer is related to dietary intake of selected nutrients, phytochemicals and food groups. *J Nutr* 133:1937–1942
 119. de Kleijn MJ, van der Schouw YT, Wilson PW, Adlercreutz H, Mazur W, Grobbee DE, Jacques PF (2001) Intake of dietary phytoestrogens is low in postmenopausal women in the United States: the Framingham study. *J Nutr* 131:1826–1832
 120. Chávez-Suárez KM, Ortega-Vélez MI, Valenzuela-Quintanar AI et al (2017) Phytoestrogen concentrations in human urine as biomarkers for dietary phytoestrogen intake in Mexican women. *Nutrients* 9:E1078. <https://doi.org/10.3390/nu9101078>
 121. Park S-Y, Wilkens LR, Franke AA, Le Marchand L, Kakazu KK, Goodman MT, Murphy SP, Henderson BE, Kolonel LN (2009) Urinary phytoestrogen excretion and prostate cancer risk: a nested case-control study in the Multiethnic Cohort. *Br J Cancer* 101:185–191. <https://doi.org/10.1038/sj.bjc.6605137>
 122. Hu Y, Song Y, Franke AA, Hu FB, van Dam RM, Sun Q (2015) A prospective investigation of the association between urinary excretion of dietary lignan metabolites and weight change in US women. *Am J Epidemiol* 182:503–511. <https://doi.org/10.1093/aje/kwv091>
 123. Reger MK, Zollinger TW, Liu Z, Jones J, Zhang J (2017) Association between urinary phytoestrogens and C-reactive protein in the continuous national health and nutrition examination survey. *J Am Coll Nutr* 36:434–441. <https://doi.org/10.1080/07315724.2017.1318722>
 124. Martínez Steele E, Monteiro CA (2017) Association between dietary share of ultra-processed foods and urinary concentrations of phytoestrogens in the US. *Nutrients* 9:E209. <https://doi.org/10.3390/nu9030209>
 125. Adlercreutz H, Fotsis T, Heikkinen R, Dwyer JT, Woods M, Goldin BR, Gorbach SL (1982) Excretion of the lignans enterolactone and enterodiol and of equol in omnivorous and vegetarian postmenopausal women and in women with breast cancer. *Lancet* 2:1295–1299
 126. Miles FL, Navarro SL, Schwarz Y, Gu H, Djukovic D, Randolph TW, Shojaie A, Kratz M, Hullar MAJ, Lampe PD, Neuhauser ML, Raftery D, Lampe JW (2017) Plasma metabolite abundances are associated with urinary enterolactone excretion in healthy participants on controlled diets. *Food Funct* 8:3209–3218. <https://doi.org/10.1039/c7fo00684e>
 127. Reger MK, Zollinger TW, Liu Z, Jones J, Zhang J (2016) Urinary phytoestrogens and cancer, cardiovascular, and all-cause mortality in the continuous National Health and Nutrition Examination Survey. *Eur J Nutr* 55:1029–1040. <https://doi.org/10.1007/s00394-015-0917-y>
 128. Eichholzer M, Richard A, Nicastro HL, Platz EA, Linseisen J, Rohrmann S (2014) Urinary lignans and inflammatory markers in the US National Health and Nutrition Examination Survey (NHANES) 1999–2004 and 2005–2008. *Cancer Causes Control* 25:395–403. <https://doi.org/10.1007/s10552-014-0340-3>
 129. Valentín-Blasini L, Sadowski MA, Walden D, Caltabiano L, Needham LL, Barr DB (2005) Urinary phytoestrogen concentrations in the U.S. population (1999–2000). *J Expo Anal Environ Epidemiol* 15:509–523
 130. Sun Q, Wedick NM, Pan A, Townsend MK, Cassidy A, Franke AA, Rimm EB, Hu FB, van Dam RM (2014) Gut microbiota metabolites of dietary lignans and risk of type 2 diabetes: a prospective investigation in two cohorts of U.S. women. *Diabetes Care* 37:1287–1295. <https://doi.org/10.2337/dc13-2513>
 131. Levine LD, Kim K, Purdue-Smithe A, Sundaram R, Schisterman EF, Connell M, Devilbiss EA, Alkhalaf Z, Radoc JG, Buck Louis GM, Mumford SL (2019) Urinary phytoestrogens and relationship to menstrual cycle length and variability among healthy, eumenorrheic women. *J Endocr Soc* 4:bvz003. <https://doi.org/10.1210/jendso/bvz003>
 132. Stumpf K, Pietinen P, Puska P, Adlercreutz H (2000) Changes in serum enterolactone, genistein, and daidzein in a dietary intervention study in Finland. *Cancer Epidemiol Biomark Prev* 9:1369–1372
 133. Pietinen P, Stumpf K, Männistö S, Kataja V, Uusitupa M, Adlercreutz H (2001) Serum enterolactone and risk of breast cancer: a case-control study in eastern Finland. *Cancer Epidemiol Biomark Prev* 10:339–344
 134. Vanharanta M, Voutilainen S, Lakka TA, van der Lee M, Adlercreutz H, Salonen JT (1999) Risk of acute coronary events

- according to serum concentrations of enterolactone: a prospective population-based case-control study. *Lancet* 354:2112–2115
135. Kilkkinen A, Erlund I, Virtanen MJ, Alftan G, Ariniemi K, Virtamo J (2006) Serum enterolactone concentration and the risk of coronary heart disease in a case-cohort study of Finnish male smokers. *Am J Epidemiol* 163:687–693
 136. Vanharanta M, Voutilainen S, Nurmi T, Kaikkonen J, Roberts LJ, Morrow JD, Adlercreutz H, Salonen JT (2002) Association between low serum enterolactone and increased plasma F2-isoprostanes, a measure of lipid peroxidation. *Atherosclerosis* 160:465–469
 137. Kilkkinen A, Stumpf K, Pietinen P, Valsta LM, Tapanainen H, Adlercreutz H (2001) Determinants of serum enterolactone concentration. *Am J Clin Nutr* 73:1094–1100
 138. Vanharanta M, Voutilainen S, Rissanen TH, Adlercreutz H, Salonen JT (2003) Risk of cardiovascular disease-related and all-cause death according to serum concentrations of enterolactone: Kuopio Ischaemic Heart Disease Risk Factor Study. *Arch Intern Med* 163:1099–1104
 139. Sonestedt E, Ivarsson MIL, Harlid S, Ericson U, Gullberg B, Carlson J, Olsson H, Adlercreutz H, Wirfält E (2009) The protective association of high plasma enterolactone with breast cancer is reasonably robust in women with polymorphisms in the estrogen receptor alpha and beta genes. *J Nutr* 139:993–1001. <https://doi.org/10.3945/jn.108.101691>
 140. Stattin P, Bylund A, Biessy C, Kaaks R, Hallmans G, Adlercreutz H (2004) Prospective study of plasma enterolactone and prostate cancer risk (Sweden). *Cancer Causes Control* 15:1095–1102
 141. Lin Y, Wolk A, Håkansson N, Peñalvo JL, Lagergren J, Adlercreutz H, Lu Y (2013) Validation of FFQ-based assessment of dietary lignans compared with serum enterolactone in Swedish women. *Br J Nutr* 109:1873–1880. <https://doi.org/10.1017/S000711451200387X>
 142. Hultén K, Winkvist A, Lenner P, Johansson R, Adlercreutz H, Hallmans G (2002) An incident case-referent study on plasma enterolactone and breast cancer risk. *Eur J Nutr* 41:168–176
 143. Aarestrup J, Kyrø C, Knudsen KEB, Weiderpass E, Christensen J, Kristensen M, Würtz AM, Johnsen NF, Overvad K, Tjønneland A, Olsen A (2013) Plasma enterolactone and incidence of endometrial cancer in a case-cohort study of Danish women. *Br J Nutr* 109:2269–2275. <https://doi.org/10.1017/S0007114512004424>
 144. Eriksen AK, Kyrø C, Nørskov NP, Frederiksen K, Bach Knudsen K-E, Overvad K, Landberg R, Tjønneland A, Olsen A (2019) Pre-diagnostic plasma enterolactone concentrations are associated with lower mortality among individuals with type 2 diabetes: a case-cohort study in the Danish Diet, Cancer and Health cohort. *Diabetologia* 62:959–969. <https://doi.org/10.1007/s00125-019-4854-9>
 145. Verheus M, van Gils CH, Keinan-Boker L, Grace PB, Bingham SA, Peeters PH (2007) Plasma phytoestrogens and subsequent breast cancer risk. *J Clin Oncol* 25:648–655
 146. Heald CL, Ritchie MR, Bolton-Smith C, Morton MS, Alexander FE (2007) Phyto-oestrogens and risk of prostate cancer in Scottish men. *Br J Nutr* 98:388–396
 147. Piller R, Chang-Claude J, Linseisen J (2006) Plasma enterolactone and genistein and the risk of premenopausal breast cancer. *Eur J Cancer Prev* 15:225–232
 148. Horner NK, Kristal AR, Prunty J, Skor HE, Potter JD, Lampe JW (2002) Dietary determinants of plasma enterolactone. *Cancer Epidemiol Biomark Prev* 11:121–126
 149. Ko KP, Yeo Y, Yoon JH et al (2018) Plasma phytoestrogens concentration and risk of colorectal cancer in two different Asian populations. *Clin Nutr* 37:1675–1682. <https://doi.org/10.1016/j.clnu.2017.07.014>