REVIEW

Global strategies for the prevention of neural tube defects through the improvement of folate status in women of reproductive age

Homero Martinez1,2 [·](http://orcid.org/0000-0002-1435-0306) Adriana Benavides‑Lara³ · Anastasia Arynchyna‑Smith4 · Kemel A. Ghotme5,6 [·](http://orcid.org/0000-0001-6802-4373) Mandana Arabi¹ · Alexander Arynchyn[7](http://orcid.org/0000-0001-9808-1774)

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Abstract

Introduction Neural tube defects represent a global public health problem, mainly in countries where effective prevention strategies are not yet in place. The global prevalence of neural tube defects is estimated at 18.6/10,000 (uncertainty interval: 15.3–23.0) live births, where ~ 75% of cases result in under-five mortality. Most of the mortality burden is in low- and middle-income countries. The main risk factor for this condition is insufficient folate levels in women of reproductive age. **Methods** This paper reviews the extent of the problem, including the most recent global information on folate status in women of reproductive age and the most recent estimates of the prevalence of neural tube defects. Additionally, we provide an overview of the available interventions worldwide to reduce the risk of neural tube defects by improving folate status in the population, including dietary diversification, supplementation, education, and fortification.

Results Large-scale food fortification with folic acid is the most successful and effective intervention to reduce the prevalence of neural tube defects and associated infant mortality. This strategy requires the coordination of several sectors, including governments, the food industry, health services providers, the education sector, and entities that monitor the quality of the service processes. It also requires technical knowledge and political will. An international collaboration between governmental and non-governmental organizations is essential to succeed in saving thousands of children from a disabling but preventable condition.

Discussion We propose a logical model for building a national-level strategic plan for mandatory LSFF with folic acid and explain the actions needed for promoting sustainable system-level change.

Keywords Neural tube defects · Prevention · Folic acid · Food fortification · Low- and middle-income countries

Abbreviations

 \boxtimes Homero Martinez hmartinez@nutritionintl.org

- ¹ Global Technical Services, Nutrition International, 180 Elgin St. suite 1000, ON, Ottawa, Canada
- ² Hospital Infantil de México Federico Gómez, Mexico City, Mexico
- ³ Costa Rican Birth Defects Register Center (CREC), Costa Rican Institute of Research and Education in Nutrition and Health (INCIENSA), Cartago, Costa Rica

- Division of Pediatric Neurosurgery, Department of Neurosurgery, University of Alabama at Birmingham, Birmingham, AL, USA
- ⁵ Translational Neuroscience Research Lab, Faculty of Medicine, Universidad de La Sabana, Chia, Colombia
- ⁶ Department of Neurosurgery, Fundación Santa Fe de Bogota, Bogota, Colombia
- ⁷ Division of Preventive Medicine, Department of Medicine, University of Alabama at Birmingham, Birmingham, AL, USA

Introduction

Neural tube defects (NTD) represent a failure of the neural tube to close properly in the early phase of gestation (around day 28 post-conception) [[1](#page-14-0)]. The most common forms of NTD include spina bifida, encephalocele, and anencephaly. The latter is the most severe form, as this defect is incompatible with extrauterine life. Children presenting with other forms survive but often require neurosurgery to close the defect, either prenatally or postnatally. About 75% of them die before reaching the age of 5 years, mainly in low- and middle-income countries (LMIC) [[2\]](#page-14-1). A common complication of spina bifida is hydrocephalus. Managing these patients requires early and recurrent neurosurgical, urological, and orthopedic follow-up, with multiple surgeries, hospitalizations, and clinic follow-ups [\[1\]](#page-14-0). These children live with a life-long disability that impacts them and their families. Given the scarcity of resources, this public health problem is especially burdensome in LMIC.

NTD have several causes, as summarized in Table [1.](#page-1-0) However, there is ample scientific evidence that the major contributing factor is insufficient folate levels, as reflected by red blood cell (RBC) folate. There is a dose–response relationship showing an inverse response between RBC folate and NTD risk [[3,](#page-14-2) [4\]](#page-14-3), with an inflection point of folate in RBC~1000 nmol/L (determined by a microbiological assay or MBA, Table [2](#page-2-0)), above which risk of folate-responsive NTD-affected pregnancies is the lowest [\[3](#page-14-2)]. The World Health Organization (WHO) has identified a cut-off of 906 nmol/L of RBC folate at a population level to provide maximum protection against NTD [[5\]](#page-14-4). The scientific evidence expresses population protection level as an NTD prevalence of 5–6 cases per 10,000 live births, as is found in every country that has implemented a successful large-scale food fortification (LSFF) program with folic acid [[2,](#page-14-1) [6](#page-14-5)].

The objectives of this paper are (1) to review the extent of the problem, including the most recent global information on folate status in women of reproductive age and NTD prevalence estimates, (2) to provide an overview of available interventions to reduce NTD risk by improving folate status in the population, and (3) to propose activities that a country should consider in their strategic plan to establish a mandatory large-scale food fortification (LSFF) program with folic acid.

Identifying the extent of the problem

Assessment of folate status

RBC and serum/plasma folate levels are the most widely used biomarkers for assessing folate status. RBC folate is a biomarker of long-term folic acid intake, whereas serum/ plasma folate reflects short-term folic acid intake [\[4,](#page-14-3) [15,](#page-14-6) [16](#page-15-0)]. RBC folate level is preferred to assess population-level folate status. The WHO recommends using a harmonized MBA to measure RBC folate to obtain reliable, comparable results across countries and time [\[5](#page-14-4)].

Folate *deficiency* increases the risk of developing megaloblastic anemia, with cut-off values of<7 nmol/L for serum folate and $\langle 227 \text{ nmol/L}$ for RBC folate (Table [2\)](#page-2-0) [[5\]](#page-14-4). The folate concentration needed to support rapid cell division at the neural tube closure is much higher; therefore, it requires a higher cut-off to provide maximum protection against folate-dependent NTD. Folate levels below this cut-off are referred to as folate *insufficiency*. According to WHO guidelines, the most significant protective level against NTD is when RBC folate in the population reaches a threshold of 906 nmol/L or the corresponding value (748 nmol/L) when using a different calibrator for the MBA (Table [2](#page-2-0)) [[16](#page-15-0)].

Environmental exposures $[7-10]$	Infectious diseases $[11]$	Drugs $[12]$		Metabolic disorders $[13]$ Genetic susceptibility $[14]$	Nutritional factors [3]
Organic solvents Arsenic Pesticides Paints X-radiation	Malaria	Thalidomide Methotrexate Antiepileptics (carbamazepine, valproic acid) Antimalarials (sulfadiazine / pyrimethamine)	Diabetes mellitus Phenyl-ketonuria	Polymorphisms of the methylenetetra- hydrofolate reductase (MTHFR) enzyme	Deficiency of B vitamins: riboflavin (B_2) , pyridoxine (B_6) , and B_{12} Insufficiency of folate (B_0)

Table 1 Potential factors contributing to the occurrence of neural tube defects

Table 2 Deficiency and insufficiency of serum and RBC folate – defnition and threshold values

RBC red blood cell

^aAs recommended by World Health Organization [\[5](#page-14-4)]/Centers for Disease Control and Prevention [[21](#page-15-4)]

b Determined by a homogenized microbiological assay using wild-type strain and folic acid standard as calibrator c Determined by a homogenized microbiological assay using a chloramphenicol-resistant strain of *Lactobacillus rhamnosus* and folic acid standard as calibrator

d Determined by a homogenized microbiological assay using a chloramphenicol-resistant strain of *Lactobacillus rhamnosus* and 5-methyl-tetrahydrofolate standard as calibrator

Folate concentration shows a dose–response relationship concerning NTD risk [[17](#page-15-1)].

Current folate status in women of reproductive age

There is a dearth of information about folate status globally. A recent systematic review article by Rogers et al. (2018) identified all nationally representative surveys that reported folate deficiency or insufficiency worldwide between 2000 and 2014 $[22]$ $[22]$. Most (>70%) of the available information represented high- or upper-middle-income countries. Overall, folate insufficiency prevalence was > 40% in most countries. This review highlighted several shortfalls present in understanding folate status globally. They included the paucity of data (only 39 countries had conducted this evaluation during the study period, providing 45 surveys overall), scarcity of data in LMIC, scarcity of information on folate insufficiency, and heterogeneity of laboratory methods used (only ten surveys used the recommended harmonized MBA to measure RBC folate). The main caveat was related to the laboratory methods used to measure folate status, which required the authors to devise an "assay factor" to assess whether the survey was likely to overestimate, underestimate, or was likely to report correct results, taking the MBA (calibrated with 5-methyl-THF) recommended by the Centres for Disease Control and Prevention (CDC) as reference (Table [3\)](#page-3-0).

Prevalence of NTD

Estimating NTD prevalence in LMIC has proved difficult and, in most cases, imprecise, often resulting in an underestimation of true values. This challenge is due to multiple factors; most LMICs do not have high-quality birth defects surveillance programs, including information on stillbirths and abortions. Additionally, elective termination of pregnancy for fetal anomalies (ETOPFA) is illegal in many LMICs, so the prevalence is measured by the rate of NTD cases out of 10,000 live births. Most recent estimates of NTD prevalence come from a systematic analysis by Blencowe et al. (2018) that estimated global prevalence based on available country birth registries and published literature [\[2](#page-14-1)]. This analysis showed that in 2015, there were an estimated 260,100 (uncertainty interval (UI): 213,800–322,000) new NTD globally (prevalence 18.6/10,000 (UI:15.3–23.0) live births). Out of this, 75% of cases (117,900) resulted in under-five mortality, with a majority in LMIC. The authors also report NTD prevalence rates per 10,000 live births without folic acid fortification by region: Southern Asia (31.96), East Asia (19.44), Northern Africa and Western Asia (17.45), and Sub-Saharan Africa (15.27) [\[2](#page-14-1)].

NTD prevention strategies

The following sections summarize the strategies available to improve folate status, highlighting their strengths and caveats. Figure [1](#page-3-1) illustrates the interconnectivity and complementarity of the preventive strategies.

Dietary diversification

Naturally occurring folate is found in different foods, including green leafy vegetables (folic in Latin means leaf), beans, peas, lentils, asparagus, broccoli, beets, avocado, nuts and seeds, wheat germ, in some fruits (papaya, bananas, citrus fruits), and animal products like eggs and beef liver. While nutrition education at the individual or the community levels should include information on sources of folate and reasons to include these foods in the diet, it may be challenging to obtain the required daily intake of folate from only food. Current folic acid daily dose recommendations build on folate intake provided by a regular diet [[23\]](#page-15-3).

Table 3 Global folate deficiency and insufficiency in women of reproductive age (2000–2014)^a

Data source: Rogers et al. [[22](#page-15-2)]

a Reporting "likely correct prevalence" as defned by the authors based on pre-specifed values of the ratio of survey assay to the microbiological assay proposed by the US Centers for Disease Control and Prevention

b Prevalence based on red blood cell folate < 305 nmol/L using folic acid calibrator or serum folate < 7 nmol/L using folic acid calibrator, considering only those surveys reporting on a likely correct assessment of folate defciency prevalence

c Prevalence based on red blood cell folate < 906 nmol/L using folic acid calibrator, considering only those surveys reporting on a likely correct assessment of folate insufficiency prevalence

^dPrevalence of folate insufficiency in women of reproductive age in upper-middle-income countries was based on only two surveys; one likely overestimated prevalence and one likely underestimated prevalence

Periconceptional oral folic acid supplementation for primary prevention

In the early 1980s, Smithells et al. provided the earliest evidence of the efficacy of primary prevention of NTD with multivitamin supplementation, including folic acid [\[24](#page-15-8)[–26](#page-15-9)]. Later studies identified folic acid as the primary contributor to the preventive effect $[27-30]$ $[27-30]$ $[27-30]$ $[27-30]$. A systematic review of randomized or quasi-randomized clinical trials evaluating the effect of periconceptional folate supplementation in women, independent of age and parity, reporting on 7391 women (2033 with a history of an NTD-affected pregnancy and 5358 without NTD history) demonstrated that oral supplementation prevents the occurrence and recurrence of NTD by up to 70% [[31\]](#page-15-12).

WHO and the US Centers for Disease Control and Prevention (CDC) recommend that all women of reproductive age get 400 µg of folic acid daily and folate in the diet. For women with diabetes or receiving an anticonvulsant treatment, the daily recommended dose is 5000 µg, in addition to dietary advice [\[23](#page-15-3)].

A recent systematic review on the impact of folic acid fortification and supplementation policies worldwide determined that government supplementation campaigns or recommendations effectively reduced NTD prevalence [[32](#page-15-13)]. They also highlighted the role of education and adherence to recommendations. However, the unintended pregnancy rate in LMIC is high and is negatively associated with income compared to HIC. In 2015–2019, the unintended pregnancy rate for HIC was 34 per 1000 women of 15–49 years, for middle-income countries (MIC) was 66 per 1000 and for LIC was 96 per 1000 [\[33](#page-15-14)]. There is a potential disparity in those who access prenatal care. Women seeking such care may be more affluent, more educated, or have a history of complicated pregnancies [\[34](#page-15-15), [35\]](#page-15-16). Supplementation is not an effective strategy for unintended pregnancies [\[33](#page-15-14)].

Periconceptional oral folic acid supplementation for recurrence prevention

NTD recurrence is an NTD-affected pregnancy in a mother with a previously NTD-affected pregnancy. The American

Fig. 1 Interconnected and complementary strategies for the prevention of neural tube defects. Source: authors' work using Microsoft's PowerPoint SmartArt

- Dietary diversification and
- practice of healthy eating habits Health literacy including
- awareness of neural tube defects risk related to poor folate intake and how to improve folate status
- Awareness of the importance of peri-conceptional care

- in the population Mandatory large scale food fortification with folic acid that may include one or more foods/food vehicles
- Promote stakeholders involvement
- \triangleright Neural tube defects surveillance Awareness and education

campaigns

MULTILEVEL STRATEGIES FOR PREVENTION OF NEURAL TUBE DEFECTS

Periconceptional folic acid supple-

CLINICAL PRACTICE LEVEL

 \triangleright Identification and control of other risk factors for neural tube defects

mentation

> Periconceptional care

College of Medical Genetics, CDC, and the American College of Obstetricians and Gynecologists recommend intake of 4000 µg per day of folic acid, starting 1 month before the intended pregnancy and continuing for the first trimester of pregnancy to prevent NTD recurrence in the subsequent pregnancy [\[23](#page-15-3), [36](#page-15-17)[–38](#page-15-18)].

Grosse and Collins' (2007) systematic review reports a 4% recurrence risk after one previously NTD-affected pregnancy and 11.1% after two NTD-affected pregnancies from pooled data in Great Britain, North America, and Europe [[39\]](#page-15-19). Four randomized trials focused on NTD recurrence prevention via folic acid supplements showed a pooled NTD rate of 0.6% compared to those who did not take folic acid supplements of 4.1%, a risk reduction of 87% [\[39\]](#page-15-19). Five observational studies provide a range of 85–100% recurrence risk reduction [[39\]](#page-15-19). [https://www.msn.com/en-ca/feed.](https://www.msn.com/en-ca/feed)

Folic acid weekly supplementation

No randomized controlled trials have tested the efficacy of weekly periconceptional folate supplementation, specifically on NTD prevention rates. However, three studies have documented improvements in folate status (as a surrogate for NTD prevention) in response to weekly folic acid supplements, with one documenting reductions in NTD following a pre-post analysis.

The first study enrolled 74 women from rural and urban areas in the state of Nuevo Leon, Mexico, including 39 who had a previously NTD-affected pregnancy and 35 who had a normal pregnancy. All women received 5000 µg (5 mg) of folic acid weekly for 3 months. Results showed that 90% of women significantly increased RBC and plasma folate concentrations [[40\]](#page-15-20). Following these encouraging results, the state scaled up the intervention to include awareness education to the public, focusing on women affected by an NTD pregnancy (recurrence prevention). The program provided free supplementation to 250,000 low-income women of childbearing age. Follow-up at 28 months drew data from an active hospital-based NTD surveillance program, complemented by fetal and child mortality from death records. Of 209 women enrolled in the recurrence prevention intervention, 30 became pregnant, with 0 recurrences. The state-wide evaluation showed a significant decrease in NTD prevalence from 10.04/10,000 live births at baseline to 5.8/10,000 live births at 28 months [[41\]](#page-15-21).

A three-arm randomized control trial in New Zealand enrolled 114 women of reproductive age (WRA) for 12 weeks in a (2800 µg weekly, 400 µg daily, placebo daily) and assessed the effect of a once-a-week supplementation dose on RBC folate concentrations. Results showed a protective increase in folate levels in 49% of women, yet this was less effective compared to 74% of women in the daily 400 µg group [[42](#page-15-22)].

More recently, a three-arm randomized controlled trial in Malaysia documented the response of 331 young women (median age 18 years), randomly assigned to receive weekly iron and folic acid supplementation or WIFAS in a dose of 60 mg iron and either 2800 μ g 400 μ g, or 0 μ g, for 16 weeks. This trial found that two groups with folic acid showed a higher mean RBC than those receiving none, and those receiving the higher dose of folic acid (2800 µg) had a higher mean RBC folate than those receiving the lower (400 µg) dose (mean difference of 271 nmol/L). More importantly, women who received the 2800 µg were seven times (*RR*=7.3, 95% CI:3.9–13.7) more likely to show RBC folate levels above the cut-off sufficiency point (748 nmol/L) [\[43](#page-15-23)].

Education campaigns on folic acid supplementation

Lack of knowledge and awareness of the protective effect of folic acid against NTD is the leading cause of inadequate folic acid intake among WRA [\[44](#page-15-24)]. Prenatal consultation is a valuable strategy for those who plan to conceive, but often it is not timely. Most women find out they are pregnant after 5 weeks when they miss their menstrual period. As neural tube closure occurs within the first 4 weeks post-conception, initiating the consumption of folic acid at this stage is too late. A total of 44% of pregnancies worldwide were unintended from 2010 to 2014 [[45\]](#page-15-25), and in LMIC, planned pregnancies are even lower among adolescents and women with limited access to information.

Furthermore, supplement use is deficient even among women planning their pregnancy. Werler et al. found that among 60% of planned pregnancies in the USA and Canada, only 8% of mothers with previously NTD-affected pregnancies and 13% in healthy pregnancies used folic acid supplements before getting pregnant [[30](#page-15-11)]. Similarly, HIC demonstrate the same gap between supplement use in planned pregnancy: 3%/77% (UK) [[46](#page-15-26)],14%/76% (Denmark) [\[47](#page-15-27)], and 31%/80% (Norway) [\[48](#page-15-28)].

A systematic review by Stockley et al. (2008) identified several challenges related to this strategy, including young age, ethnicity, and low socioeconomic status. Isolated educational campaigns that use printed resources or mass media were not effective long term, while healthcare-based initiatives could be effective as long as supplements were easily accessible. Most effective campaigns focus on women in a state of vulnerability, yet they reach only 50% of women [[49\]](#page-15-29).

Large‑scale food fortification (LSFF)

LSFF refers to the process of adding essential micronutrients (vitamins and minerals) at the production stage (i.e., milling) to foods or other food vehicles (i.e., salt) widely consumed by the general population [[50](#page-16-0)]. LSFF's primary goal is to

improve the population's health status through increased nutrient intake without significant changes to habitual food consumption. LSFF is a population-level intervention and has several advantages: (1) no need to change diet at the individual level; (2) feasibility of fortifying foods with multiple micronutrients at once; (3) high return-on-investment on population-level health; and (4) safe [[50\]](#page-16-0).

LSFF can be especially effective in improving the population's nutritional status if well planned, executed, and maintained by respective governmental entities. LSSF does not replace other methods of food fortification such as targeted food fortification, point-of-use or home fortification, or biofortification but can work in combination with those. Effective implementation of LSFF programs implies efficacy and effectiveness research that provides evidence about the actual impact of LSSF on health outcomes [[51](#page-16-1)].

LSFF with folic acid is successful and highly effective in increasing folate levels in WRA and reducing NTD prevalence in LMIC [[52\]](#page-16-2). Table [4](#page-6-0) summarizes the best available information from peer-reviewed publications on the effectiveness of LSFF with folic acid on NTD prevalence, NTDassociated mortality, and folate levels in LMIC and selected HIC countries.

As shown in Table [4,](#page-6-0) LSFF with folic acid is consistently successful and highly effective in increasing folate levels in WRA, leading to a reduced NTD prevalence, including NTDassociated infant mortality. Several MIC such as Costa Rica [\[53](#page-16-3)[–55\]](#page-16-4), Brazil [[56,](#page-16-5) [57](#page-16-6)], Mexico [[57\]](#page-16-6), Argentina [\[58](#page-16-7)], Iran [\[59\]](#page-16-8), South Africa $[60]$ $[60]$, and Cameroon $[61]$ $[61]$ have demonstrated such benefits after the implementation of mandatory fortification. Furthermore, NTD prevalence reduction ranged from 30 to 31% (Brazil and South Africa) [\[56,](#page-16-5) [58,](#page-16-7) [60](#page-16-9)] and 58–59% (Costa Rica, Mexico) [[54,](#page-16-11) [57\]](#page-16-6), mainly due to the fortification of wheat flour with folic acid at levels greater than 1.8 mg/kg or corn flour at 1.3–2 mg/kg. Costa Rica demonstrated one of the highest rates of reduction in NTD prevalence by fortifying different foods as a strategy to reach different populations, including corn and wheat flour (1.8 mg/ kg), rice (1.8 mg/kg), and milk (400 µg/200 ml) [\[53](#page-16-3), [54\]](#page-16-11).

In China, NTD prevalence has remained high despite mandatory supplementation programs. However, a study in Shanxi province showed a 68% reduction in NTD prevalence in villages with mandatory wheat flour fortification compared to those without fortification [[62\]](#page-16-12).

Australia provides a singular case study, as there are published evaluations showing changes in NTD prevalence over time, corresponding to a pre-fortification period with no intervention (1980–1992), followed by the introduction of widely available folic acid supplementation (1993–1995), voluntary fortification (1996–2008), and lastly by mandatory fortification (September 2009). Comparing the nointervention period with the promotion of the supplementation period, the prevalence of anencephaly, spine bifida,

and encephalocele per 10,000 births remained significantly unchanged (PR of 1.1, 0.9, and 1.2, respectively). After voluntary fortification, anencephaly and spina bifida prevalence significantly reduced by 32% and 23%, respectively [[71\]](#page-16-13). However, the reduction in NTD was only significant for the population of non-Aboriginal origin (Table [4\)](#page-6-0). They achieved a significant decrease in NTD prevalence across all population groups (i.e., Aboriginal and Torres Strait Islanders) only after mandatory fortification. Hilder et al. [\[71](#page-16-13)] documented a drop from 10.2 per 10,000 births in the voluntary fortification period to 9.4 per 10,000 births in the mandatory fortification period (*RR* = 0.86, 95%CI 0.74–0.99). There was a higher reduction in the population of Aboriginal origin, from 19.6 per 10,000 to 5.1 per 10,000 births (*RR*=0.26, 95%CI 0.12–0.55) [[72\]](#page-16-14). These results agree with those reported by Brown et al. who demonstrated an 85% reduction in folate insufficiency (RBC less than 310 nmol/L) in a survey of public hospitals in Australia, which decreased from 3.4% after voluntary fortification to 0.5% after mandatory fortification [\[72](#page-16-14)].

Though data are scarce, the reduction in NTD-related mortality after mandatory LSFF ranged between 66% (South Africa) [\[60\]](#page-16-9), 68% (Argentina) [\[56\]](#page-16-5), and 71% (Costa Rica) [\[54\]](#page-16-11).

Building a national‑level strategic plan for mandatory LSFF with folic acid

Enacting, promulgating, and enforcing a national regulation for LSFF with folic acid is an attainable goal, but challenges may exist. Establishing a strategic plan requires the coordination of different sectors, including governments, the food industry, health services providers, the education sector, and entities that monitor the quality of the service processes. It also requires technical knowledge, political will, and partnership with national entities and supranational organizations. We describe the main elements of a strategic plan for a national-level mandatory LSFF, summarized in a logical model presented in Fig. [2.](#page-11-0)

Building the evidence to support LSFF

Establishing a mandatory food fortification program in a specific country or population must be based on three factors: (1) food/micronutrient intake, (2) micronutrient status (using the relevant biomarkers), and (3) identification of a suitable food vehicle for fortification [\[50,](#page-16-0) [73\]](#page-16-15).

Dietary surveys tailored for each country or culture assess food consumption habits and folate intake. A fortification rapid assessment tool (FRAT) provides the necessary information about food consumption patterns at the household level. This tool combines a simplified 24-h recall and food consumption questionnaires [\[74](#page-16-16), [75\]](#page-16-17) to support an informed

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Table 4 (continued)

NTD neural tube defects, SB spina bifida, WRA women of reproductive age, EAR estimated average requirement $*_{p} > 0.05$; $**_{p} > 0.05$; $***$ $p < 0.001$ *NTD* neural tube defects, *SB* spina bifda, *WRA* women of reproductive age, *EAR* estimated average requirement **p >* 0.05; ***p <* 0.05; *** *p <* 0.001

Table 4 (continued)

Fig. 2 A logic model for large-scale food fortifcation as an evidence-based policy to prevent neural tube defects (NTD). Source: authors' work using Microsoft's PowerPoint SmartArt

decision about the most appropriate food vehicle(s) to fortify [[76](#page-16-26)]. Cultural and food habits are crucial factors in explaining the prevalence of folate insufficiency in different regions. Importantly, if the population has outliers due to ethnic origin or eating habits, this should also be considered in the sampling strategy [\[77\]](#page-16-27).

Biological indicators such as serum and RBC folate help assess folate status (Table [2](#page-2-0)). The case study to measure folate status at the population level should have a representative number of WRA from different regions to evaluate the prevalence of folate insufficiency in the whole country. It is good practice to select a representative sample of rural and urban areas and, within them, select women from regions or localities with high and low NTD prevalence to serve as sentinel groups when testing the effectiveness of the intervention.

The first requirement for an appropriate vehicle for fortification is that it should be consumed by most of the population. Secondly, this food should be processed by a centralized industry (as opposed to being processed in multiple small entities, such as small mills in rural areas). Additionally, the added micronutrient(s) must not alter the accepted organoleptic characteristics of the food, including flavor, smell, taste, and, in most instances, color. Once added to food, the micronutrient should not be significantly affected by regular packaging and storage or traditional cooking practices. It is also critical that the price not be significantly impacted due to the fortification to assure vast consumption of the fortified food [[50\]](#page-16-0).

The most common staple foods used for LSFF with folic acid include cereals (wheat, corn (maize), and rice), but other fortified foods help improve folic acid delivery to the population, including milk and oil. There are, however, other foods and condiments that may constitute relevant alternatives for LSFF. A recent article has identified many foods that have been fortified with folic acid and have some evaluation, including retention of the micronutrient after production or storage, acceptability, and food and sensory characteristics after fortification. These foods include dairy products (lowfat milk, whole milk, powder milk, yogurt), condiments (salt, sugar, bouillon cubes), fruit juices, meat and eggs, fruit, other cereals (finger millet, teff), and candy [\[78](#page-16-28)]. Table salt and bouillon have attracted attention due to their potential to reach large segments of the population [[79,](#page-16-29) [80](#page-16-30)].

A few countries have successful experience fortifying several foods simultaneously, like Costa Rica, which fortifies wheat flour, corn flour, milk, and rice, intending to reach different populations. This strategy has proved successful, as it is complementary and universal and has significantly reduced NTD prevalence, as different foods were added to the program in a stepwise manner [[53](#page-16-3)].

Population reach and quality control are critical points in implementing any LSFF program. LSFF may be a voluntary practice (where food manufacturers add one or more micronutrients to their products, usually to increase profit but not regulated by the government) or a mandatory one (where the government mandates and regulates the fortification standards, sets up regulatory and monitoring practices and enforces them).

In settings of high coverage, poor functional results may be due to poor fortification quality and the absence of quality assurance/control programs that verify that food fortification meets the required standards. For example, one study from 12 countries with 20 national food fortification programs combined revealed that more than 50% of tested samples had micronutrients below the recommended levels [\[81](#page-16-31)]. An appropriate standardized legal framework for LSFF and well-established quality control mechanisms are needed for LSFF to effectively address the prevention goals at the population level [[82\]](#page-16-32).

Strategy definition, political will, approval, and legislation

Developing political will to fortify food with folic acid in LMIC is often challenging, as it requires efforts and funds to set up the fortification program. Given other priorities, allocating money to a food fortification program may be difficult [\[83](#page-16-33)]. However, mandatory LSFF is a government policy, so it is crucial to develop and ensure political will to move ahead with this policy. There is ample evidence to show that identifying one or more champions is a determining factor in increasing awareness and ensuring political will [[83\]](#page-16-33).

The next step is to build partnerships with stakeholders, including the food industry, universities, researchers, the public health sector (ministry of health), and the political sector, to help define the country-specific strategy. In LMIC, where resources are limited, collaborative work is crucial. The proposed fortification strategy should lead to the specific country promulgating a law appropriate to their context, including implementing, monitoring, and verifying the quality, safety, and efficacy criteria.

Legislation is the cornerstone of food fortification program implementation. It regulates all aspects of food fortification, including the list of foods to be fortified, micronutrients to be added, their level, and the program to control the efficiency. It regulates food manufacturing, packaging and labeling, quality assurance, and control of food production, storage, and transportation while promoting people's awareness and education. Legislation could be via state law or other governmental decisions. The local health ministry should prepare legislation with the other ministries and departments such as education, food production, transportation, and foreign affairs, among others. After adopting the law, food fortification becomes a mandatory process. However, it is necessary to allow a certain period so the industry may adapt to the regulation. After this, the process must remain under permanent supervision and strict control by the government. Monitoring and evaluation are essential to ensure industry compliance. Quality assurance procedures are needed to ensure fortification standards. Periodic surveys of folate levels will provide information about the effectiveness of the fortification program and may provide information to make any necessary corrections at the early stages of the process. Nationwide food-intake surveys will help amend the regulations most effectively.

A government dependency with a regulatory function such as a Food and Drug Administration will state that the codified policy or legislative regulations include situations and conditions in which the fortification of food with the nutrients listed is considered appropriate [[84](#page-17-0)]:

- \checkmark to correct a dietary insufficiency recognized by the scientific community to exist and known to result in nutrient deficiency disease or condition, such as NTD in the case of folate insufficiency
- \checkmark to restore such nutrient(s) to a level(s) representative of the food prior to storage, handling, and processing
- \checkmark in proportion to the total caloric content of the food, to balance the vitamin, mineral, and protein content
- \checkmark to replace traditional food in the diet to avoid nutritional inferiority.

Public safety and awareness

Public safety is the highest priority when considering mandatory legislation. Safety in food fortification implies a reasonable certainty that the food additive is not harmful, although there is no absolute proof of the harmlessness of any substance in the present state of scientific knowledge. Safety can be determined by consideration of the following factors [\[85](#page-17-1)]:

- the estimated consumption of the additive with food based on its use
- the estimated cumulative effect of the additive on the diet
- the existence of safety factors generally recognized as appropriate

Nutrient mixtures added to food should be stable, physiologically available, present at a level that will not lead to excessive intake, suitable for fortification purposes (at the industrial level), and acceptable for food safety regulations. Food labeling cannot be false or misleading [[86](#page-17-2), [87](#page-17-3)].

The addition of different minerals and vitamins must be under strict control. Not every food is suitable for fortification with additives, depending on its chemical composition and potential interaction with food additives. Iodine, for example, is not stable and could quickly evaporate during food processing, such as cooking. Direct sunlight can also decrease the amount of iodine in salt. That is why the fortification of salt with iodine is going through the addition of potassium iodate, which is more stable and efficient in preventing iodine insufficiency. Folic acid is a more stable chemical agent applicable to frequently consumed foods such as flour and salt [\[87](#page-17-3)]. Also, the permissible level of micronutrients must be determined and constantly verified by monitoring programs.

Monitoring and evaluation

After enacting and implementing mandatory fortification, it is necessary to gather adequate evidence on the program's effectiveness, safety, and sustainability. This evidence is attainable through nutritional surveys, measurements of folate levels in WRA, and NTD prevalence through country-wide surveillance databases. Based on the experience of several LMICs and HIC that have successfully implemented food fortification with folic acid, the impact on NTD prevalence and folate levels became evident 2 to 3 years after their implementation [\[53,](#page-16-3) [59](#page-16-8)[–64,](#page-16-19) [67](#page-16-22)[–72\]](#page-16-14). After the first evaluation, the bottlenecks and weaknesses of the processes become evident and can be corrected.

In addition, it will be possible to identify the populations least and most benefited from the intervention and propose additional foods to fortify or complementary strategies which enhance the perinatal intake of folates in these populations. For example, in Costa Rica, universities and researchers, government entities in charge of public health, and the food industry have collaborated to generate evidence and strategies for continuous intervention improvement and maximizing resources.

The long-term sustainability of LSFF programs requires stakeholders' intervention at the system level, as illustrated in Fig. [3.](#page-13-0)

Building international collaboration

Convincing local government and medical officials to fortify a staple food or condiment with folic acid is a complex goal that requires coordinating multiple efforts. International

professional and non-governmental organizations such as WHO, Nutrition International, Food Fortification Initiative, Global Alliance for Improved Nutrition, International Society for Pediatric Neurosurgery, and the Global Alliance for Prevention of Spina Bifida-F (GAPSBi-F) often play crucial advocacy roles [\[52,](#page-16-2) [88](#page-17-4)]. These organizations utilize available epidemiological and clinical data about NTD to convince local government officials on the urgency of LSFF with folic acid, using knowledge translation, transfer, and science diplomacy methods.

An international collaboration of local governments with WHO, UNICEF, medical institutions, and other organizations is essential in establishing high-quality standards for fortification programs. The local government can set up a liaison office with competent personnel to communicate with international organizations to provide logistic and scientific support. The involvement of local medical institutions will help get reliable NTD incidence and prevalence data, including an ETOPFA rate due to NTD, and evaluate the food fortification's success with folic acid through the follow-up programs. The leading local medical institution could serve as a collaborating center with the WHO that will encourage medical staff to improve public health through participation in joint research projects. Educational and research exchanges between physicians and medical researchers supported financially by WHO or other international organizations will serve this purpose.

An important aspect to consider, especially in LMIC, is the need for a reliable laboratory to conduct the recommended MBA to measure RBC folate. Given that the procedure requires a well-honed technique by adequately trained laboratory technicians, it is necessary to provide such training when conducting a national or subnational assessment. Furthermore, having an external reference laboratory checking for quality control samples is ideal. When folate surveys are not frequent in a given country, the trained laboratory may lose its expertise. To address these challenges, the Division of Laboratory Sciences of the US CDC has proposed a

Fig. 3 Actions needed for promoting sustainable systemlevel change on large-scale food fortifcation policies to prevent neural tube defects. Source: authors' work using Microsoft's PowerPoint

strategy to develop a global network of appropriately trained regional laboratories, which are certified, and periodically re-certified to conduct the folate MBA. These resource laboratories could work fee-for-service in different countries, ensuring quality results. By having different countries requiring these services, the reference laboratories will have a steady stream of work, thus maintaining their proficiency, ensured through the afore-mentioned re-certification program. Continued demand for their services would eventually lead to lower costs per sample. Furthermore, these laboratories could also help train other regional laboratories, expanding and strengthening the global network [[89](#page-17-5)].

Likewise, international collaboration is vital in the implementation, standardization, and improvement of NTD surveillance systems in LMIC to have more reliable and genuine figures to characterize the NTD burden and assess the impact of the intervention.

Conclusion

Every year, thousands of infants and young children affected by NTD die or suffer permanent disability worldwide, and this condition disproportionately affects LMIC. However, this public health problem can be effectively prevented by implementing mandatory LSFF with folic acid. This intervention has improved folate levels in WRA in every lowmiddle- or high-income country that has enacted such a policy. This intervention is the most cost-effective way to improve micronutrient status in the population, providing a sufficient level of folate that can reduce NTD prevalence to 5–6 per 10,000 live births in every setting where the program is mandatory. Our review should encourage those countries which have not yet established an LSFF program to consider this intervention to reduce the burden of several micronutrient deficiencies, including nutritional anemia, iodine deficiency disorders, and NTD. LSFF can effectively reduce newborn, infant, and young child morbidity and mortality rates, thus improving the population's health, particularly in the most vulnerable segments.

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Data availability All data provided in this article can be accessed through the corresponding references in the bibliography.

Declarations

Ethical approval This review paper dealt only with published material and did not include the participation of human subjects or animals. Therefore, the research was not submitted for review to a Human Subjects Protection Committee or Institutional Review Board. The authors state that the review of such published material was carried out with full respect and veracity of statements and facts and thus represents an authoritative assessment of the evidence reviewed.

Conflict of interest The authors declare no competing interests.

References

- 1. Botto LD, Moore CA, Khoury MJ, Erickson JD (1999) Neuraltube defects. N Engl J Med 341(20):1509–1519
- 2. Blencowe H, Kancherla V, Moorthie S, Darlison MW, Modell B (2018) Estimates of global and regional prevalence of neural tube defects for 2015: a systematic analysis. Ann N Y Acad Sci 1414(1):31–46
- 3. Daly LE, Kirke PN, Molloy A, Weir DG, Scott JM (1995) Folate levels and neural tube defects. Implications for prevention. Jama 274(21):1698–1702
- 4. Crider KS, Devine O, Hao L, Dowling NF, Li S, Molloy AM, Li Z, Zhu J, Berry RJ (2014) Population red blood cell folate concentrations for prevention of neural tube defects: Bayesian model. BMJ 349:g4554
- 5. World Health Organization (2015) Guideline: optimal serum and red blood cell folate concentrations in women of reproductive age for prevention of neural tube defects. WHO, Geneva
- 6. Castillo-Lancellotti C, Tur JA, Uauy R (2013) Impact of folic acid fortification of flour on neural tube defects: a systematic review. Public Health Nutr 16(5):901–911
- 7. Lacasaña M, Vázquez-Grameix H, Borja-Aburto VH, Blanco-Muñoz J, Romieu I, Aguilar-Garduño C, García AM (2006) Maternal and paternal occupational exposure to agricultural work and the risk of anencephaly. Occup Environ Med 63(10):649–656
- 8. Nieuwenhuijsen MJ, Martinez D, Grellier J, Bennett J, Best N, Iszatt N, Vrijheid M, Toledano MB (2009) Chlorination disinfection byproducts in drinking water and congenital anomalies: review and meta-analyses. Environ Health Perspect 117(10):1486–1493
- 9. Sever LE (1995) Looking for causes of neural tube defects: where does the environment fit in? Environ Health Perspect 103 Suppl 6(Suppl 6):165–71
- 10. Mazumdar MIHM, Hamid R et al (2015) Arsenic is associated with reduced effect of folic acid in myelomeningocele prevention: a case control study in Bangladesh. Environ Health 14:34
- 11. Metz J (2007) Folic acid metabolism and malaria. Food Nutr Bull 28(4_suppl4):S540-S9
- 12. Nzila A, Okombo J, Molloy AM (2014) Impact of folate supplementation on the efficacy of sulfadoxine/pyrimethamine in preventing malaria in pregnancy: the potential of 5-methyl-tetrahydrofolate. J Antimicrob Chemother 69(2):323–330
- 13. Ray JG, Vermeulen MJ, Meier C, Cole DE, Wyatt PR (2004) Maternal ethnicity and risk of neural tube defects: a population-based study. CMAJ 171(4):343–345
- 14. Crider KS, Zhu JH, Hao L, Yang QH, Yang TP, Gindler J, Maneval DR, Quinlivan EP, Li Z, Bailey LB, Berry RJ (2011) MTHFR 677C->T genotype is associated with folate and homocysteine concentrations in a large, population-based, double-blind trial of folic acid supplementation. Am J Clin Nutr 93(6):1365–1372
- 15. Cordero AM, Crider KS, Rogers LM, Cannon MJ, Berry RJ (2015) Optimal serum and red blood cell folate concentrations in women of reproductive age for prevention of neural tube defects:

World Health Organization guidelines. MMWR Morb Mortal Wkly Rep 64(15):421–423

- 16. Bailey LB, Hausman DB (2018) Folate status in women of reproductive age as basis of neural tube defect risk assessment. Ann N Y Acad Sci 1414(1):82–95
- 17. Duffy ME, Hoey L, Hughes CF, Strain JJ, Rankin A, Souverein OW, Dullemeijer C, Collings R, Hooper L, McNulty H (2014) Biomarker responses to folic acid intervention in healthy adults: a meta-analysis of randomized controlled trials. Am J Clin Nutr 99(1):96–106
- 18. Institute of Medicine (1998) DRI dietary reference values for thiamin, riboflavin, niacin,vitamin B6, folate, vitamin B12, pantothenic acid, biotin, and choline. Washington, D C
- 19. Chen MY, Rose CE, Qi YP, Williams JL, Yeung LF, Berry RJ, Hao L, Cannon MJ, Crider KS (2019) Defining the plasma folate concentration associated with the red blood cell folate concentration threshold for optimal neural tube defects prevention: a population-based, randomized trial of folic acid supplementation. Am J Clin Nutr 109(5):1452–1461
- 20. Pfeiffer CM, Sternberg MR, Hamner HC, Crider KS, Lacher DA, Rogers LM, Bailey RL, Yetley EA (2016) Applying inappropriate cutoffs leads to misinterpretation of folate status in the US population. Am J Clin Nutr 104(6):1607–1615
- 21. Rabinowitz D, Zhang M, Paladugula N, LaVoie D, Pfeiffer C, editors. A fresh look at the folate microbiological assay, including dried blood spots and preanalytical conditions for whole blood samples. Clinical Chemistry; 2009: Amer Assoc Clinical Chemistry 2101 L Street NW, suite 202, Washington, DC
- 22. Rogers LM, Cordero AM, Pfeiffer CM, Hausman DB, Tsang BL, De-Regil LM, Rosenthal J, Razzaghi H, Wong EC, Weakland AP, Bailey LB (2018) Global folate status in women of reproductive age: a systematic review with emphasis on methodological issues. Ann N Y Acad Sci 1431(1):35–57
- 23. Centers for Disease Control (CDC) (1991) Use of folic acid for prevention of spina bifida and other neural tube defects--1983–1991. MMWR Morb Mortal Wkly Rep 40(30):513–516. PMID: 2072886
- 24. Hibbard ED, Smithells R (1965) Folic acid metabolism and human embryopathy. Lancet 1
- 25. Smithells RW, Sheppard S, Schorah CJ, Seller MJ, Nevin NC, Harris R, Read AP, Fielding DW (1980) Possible prevention of neural-tube defects by periconceptional vitamin supplementation. Lancet 1(8164):339–340
- 26. Smithells RW, Sheppard S, Schorah CJ (1976) Vitamin deficiencies and neural tube defects. Arch Dis Child 51(12):944–950
- 27. Berry RJ, Li Z, Erickson JD, Li S, Moore CA, Wang H, Mulinare J, Zhao P, Wong LY, Gindler J, Hong SX, Correa A (1999) Prevention of neural-tube defects with folic acid in China. China-U.S. Collaborative Project for Neural Tube Defect Prevention. N Engl J Med 341(20):1485–90
- 28. Bower C, Stanley FJ (1989) Dietary folate as a risk factor for neural-tube defects: evidence from a case-control study in Western Australia. Med J Aust 150(11):613–619
- 29. Mulinare J, Cordero JF, Erickson JD, Berry RJ (1988) Periconceptional use of multivitamins and the occurrence of neural tube defects. JAMA 260(21):3141–3145
- 30. Werler MM, Shapiro S, Mitchell AA (1993) Periconceptional folic acid exposure and risk of occurrent neural tube defects. JAMA 269(10):1257–1261
- 31. De-Regil LM, Peña-Rosas JP, Fernández-Gaxiola AC, Rayco-Solon P (2015) Effects and safety of periconceptional oral folate supplementation for preventing birth defects. Cochrane Database Syst Rev 2015(12):Cd007950
- 32. Shlobin NA, LoPresti MA, Du RY, Lam S (2020) Folate fortification and supplementation in prevention of folate-sensitive neural tube defects: a systematic review of policy. J Neurosurg Pediatr 1–17
- 33. Bearak J, Popinchalk A, Ganatra B, Moller AB, Tunçalp Ö, Beavin C, Kwok L, Alkema L (2020) Unintended pregnancy and abortion

by income, region, and the legal status of abortion: estimates from a comprehensive model for 1990–2019. Lancet Glob Health 8(9):e1152–e1161

- 34. Elsinga J, van der Pal-de BK, le Cessie S, de Jong-Potjer L, Verloove-Vanhorick S, Assendelft W (2006) Preconception counselling initiated by general practitioners in the Netherlands: reaching couples contemplating pregnancy [ISRCTN53942912]. BMC Fam Pract 7:41
- 35. Czeizel AE (2004) The primary prevention of birth defects: multivitamins or folic acid? Int J Med Sci 1(1):50–61
- 36. Toriello HV (2011) Policy statement on folic acid and neural tube defects. Genet Med 13(6):593–596
- 37. ACOG Practice Bulletin (2005) Clinical management guidelines for obstetrician-gynecologists. Number 60, March 2005. Pregestational diabetes mellitus. Obstet Gynecol 105(3):675–85
- 38. Petersen JM, Parker SE, Benedum CM, Mitchell AA, Tinker SC, Werler MM (2019) Periconceptional folic acid and risk for neural tube defects among higher risk pregnancies. Birth Defects Res 111(19):1501–1512
- 39. Grosse SD, Collins JS (2007) Folic acid supplementation and neural tube defect recurrence prevention. Birth Defects Res A Clin Mol Teratol 79(11):737–742
- 40. Martínez-de Villarreal LE, Limón-Benavides C, Valdez-Leal R, Sánchez-Peña MA, Villarreal-Pérez JZ (2001) Efecto de la administración semanal de ácido fólico sobre los valores sanguíneos. salud pública de méxico 43:103–7
- 41. Martínez de Villarreal L, Pérez JZ, Vázquez PA, Herrera RH, Campos Mdel R, López RA, Ramírez JL, Sánchez JM, Villarreal JJ, Garza MT, Limón A, López AG, Bárcenas M, García JR, Domínguez AS, Nuñez RH, Ayala JL, Martínez JG, González MT, Alvarez CG, Castro RN (2002) Decline of neural tube defects cases after a folic acid campaign in Nuevo León, México. Teratology 66(5):249–56
- 42. Norsworthy B, Skeaff CM, Adank C, Green TJ (2004) Effects of once-a-week or daily folic acid supplementation on red blood cell folate concentrations in women. Eur J Clin Nutr 58(3):548–554
- 43. Samson KLI, Loh SP, Lee SS, Sulistyoningrum DC, Khor GL, Shariff ZBM, Ismai IZ, Yelland LN, Leemaqz S, Makrides M, Hutcheon JA, Roche ML, Karakochuk CD, Green TJ (2020) Weekly iron-folic acid supplements containing 2.8 mg folic acid are associated with a lower risk of neural tube defects than the current practice of 0.4 mg: a randomised controlled trial in Malaysia. BMJ Glob Health 5(12)
- 44. Lolowa AM, Selim N, Alkuwari M, Salem IM (2019) Knowledge and intake of folic acid among teachers of childbearing age in the State of Qatar: a cross-sectional study. BMJ Open 9(4):e025005
- 45. Bearak J, Popinchalk A, Alkema L, Sedgh G (2018) Global, regional, and subregional trends in unintended pregnancy and its outcomes from 1990 to 2014: estimates from a Bayesian hierarchical model. Lancet Glob Health 6(4):e380–e389
- 46. Inskip HM, Crozier SR, Godfrey KM, Borland SE, Cooper C, Robinson SM (2009) Women's compliance with nutrition and lifestyle recommendations before pregnancy: general population cohort study. BMJ 338:b481
- 47. Knudsen VK, Orozova-Bekkevold I, Rasmussen LB, Mikkelsen TB, Michaelsen KF, Olsen SF (2004) Low compliance with recommendations on folic acid use in relation to pregnancy: is there a need for fortification? Public Health Nutr 7(7):843–850
- 48. Nilsen RM, Mastroiacovo P, Gunnes N, Alsaker ER, Bjørke-Monsen AL, Eussen SJ, Haugen M, Johannessen A, Meltzer HM, Stoltenberg C, Ueland PM, Vollset SE (2014) Folic acid supplementation and interpregnancy interval. Paediatr Perinat Epidemiol 28(3):270–274
- 49. Stockley L, Lund V (2008) Use of folic acid supplements, particularly by low-income and young women: a series of systematic reviews to inform public health policy in the UK. Public Health Nutr 11(8):807–821
- 50. Allen L, De Benoist B, Dary O, Hurrell R (2006) Guidelines on food fortification with micronutrients. World Health Organization. ISBN: 9241594012
- 51. Keats EC, Neufeld LM, Garrett GS, Mbuya MNN, Bhutta ZA (2019) Improved micronutrient status and health outcomes in lowand middle-income countries following large-scale fortification: evidence from a systematic review and meta-analysis. Am J Clin Nutr 109(6):1696–1708
- 52. Martinez H, Pachón H, Kancherla V, Oakley GP (2021) Food fortification with folic acid for prevention of spina bifida and anencephaly: the need for a paradigm shift in evidence evaluation for policy-making. Am J Epidemiol 190(10):1972–1976
- 53. Barboza-Argüello Mde L, Umaña-Solís LM, Azofeifa A, Valencia D, Flores AL, Rodríguez-Aguilar S, Alfaro-Calvo T, Mulinare J (2015) Neural tube defects in Costa Rica, 1987–2012: origins and development of birth defect surveillance and folic acid fortification. Matern Child Health J 19(3):583–590
- 54. Barboza Argüello Mde L, Umaña Solís LM (2011) [Impact of the fortification of food with folic acid on neural tube defects in Costa Rica]. Rev Panam Salud Publica 30(1):1–6
- 55. Ministerio de Salud Costa Rica (2012) Encuesta nacional de nutrición. Fascículo 2. Micronutrientes. Available from: [https://www.binasss.sa.cr/opac-ms/media/digitales/Encuesta%](https://www.binasss.sa.cr/opac-ms/media/digitales/Encuesta%20nacional%20de%20nutrici%C3%B3n.%20Fasc%C3%ADculo%202.%20Micronutrientes.pdf) 20nac [ional%20de%20nutrici%C3%B3n.%20Fasc%C3%](https://www.binasss.sa.cr/opac-ms/media/digitales/Encuesta%20nacional%20de%20nutrici%C3%B3n.%20Fasc%C3%ADculo%202.%20Micronutrientes.pdf) [ADculo%202.%20Micronutrientes.pdf.](https://www.binasss.sa.cr/opac-ms/media/digitales/Encuesta%20nacional%20de%20nutrici%C3%B3n.%20Fasc%C3%ADculo%202.%20Micronutrientes.pdf) Accessed 4 May 2022
- 56. Santos LM, Lecca RC, Cortez-Escalante JJ, Sanchez MN, Rodrigues HG (2016) Prevention of neural tube defects by the fortification of flour with folic acid: a population-based retrospective study in Brazil. Bull World Health Organ 94(1):22–29
- 57. Rosenthal J, Casas J, Taren D, Alverson CJ, Flores A, Frias J (2014) Neural tube defects in Latin America and the impact of fortification: a literature review. Public Health Nutr 17(3):537–550
- 58. Calvo EB, Biglieri A (2008) Impact of folic acid fortification on women's nutritional status and on the prevalence of neural tube defects. Arch Argent Pediatr 106(6):492–498
- 59. Abdollahi Z, Elmadfa I, Djazayery A, Golalipour MJ, Sadighi J, Salehi F, Sadeghian SS (2011) Efficacy of flour fortification with folic acid in women of childbearing age in Iran. Ann Nutr Metab 58(3):188–196
- 60. Sayed AR, Bourne D, Pattinson R, Nixon J, Henderson B (2008) Decline in the prevalence of neural tube defects following folic acid fortification and its cost-benefit in South Africa. Birth Defects Res A Clin Mol Teratol 82(4):211–216
- 61. Engle-Stone R, Nankap M, Ndjebayi AO, Allen LH, Shahab-Ferdows S, Hampel D, Killilea DW, Gimou MM, Houghton LA, Friedman A, Tarini A, Stamm RA, Brown KH (2017) Iron, zinc, folate, and vitamin B-12 status increased among women and children in Yaoundé and Douala, Cameroon, 1 year after introducing fortified wheat flour. J Nutr 147(7):1426–1436
- 62. Wang H, De Steur H, Chen G, Zhang X, Pei L, Gellynck X, Zheng X (2016) Effectiveness of folic acid fortified flour for prevention of neural tube defects in a high risk region. Nutrients 8(3):152
- 63. Honein MA, Paulozzi LJ, Mathews TJ, Erickson JD, Wong LY (2001) Impact of folic acid fortification of the US food supply on the occurrence of neural tube defects. JAMA 285(23):2981–2986
- 64. Williams J, Mai CT, Mulinare J, Isenburg J, Flood TJ, Ethen M, Frohnert B, Kirby RS (2015) Updated estimates of neural tube defects prevented by mandatory folic acid fortification - United States, 1995–2011. MMWR Morb Mortal Wkly Rep 64(1):1–5
- 65. Dietrich M, Brown CJ, Block G (2005) The effect of folate fortification of cereal-grain products on blood folate status, dietary folate intake, and dietary folate sources among adult non-supplement users in the United States. J Am Coll Nutr 24(4):266–274
- 66. Hertrampf E, Cortés F (2008) National food-fortification program with folic acid in Chile. Food Nutr Bull 29(2 Suppl):S231–S237
- 67. De Wals P, Tairou F, Van Allen MI, Uh SH, Lowry RB, Sibbald B, Evans JA, Van den Hof MC, Zimmer P, Crowley M, Fernandez B,

Lee NS, Niyonsenga T (2007) Reduction in neural-tube defects after

- folic acid fortification in Canada. N Engl J Med 357(2):135–142 68. Pardo R, Vilca M, Villarroel L, Davalji T, Obrycki JF, Mazumdar M, Avila C, Mellado C (2022) Neural tube defects prevalence does not increase after modification of the folic acid fortification program in Chile. Birth Defects Res 114(7):259–266
- 69. Liu S, West R, Randell E, Longerich L, O'Connor KS, Scott H, Crowley M, Lam A, Prabhakaran V, McCourt C (2004) A comprehensive evaluation of food fortification with folic acid for the primary prevention of neural tube defects. BMC Pregnancy Childbirth 4(1):20
- 70. Bower C, D'Antoine H, Stanley FJ (2009) Neural tube defects in Australia: trends in encephaloceles and other neural tube defects before and after promotion of folic acid supplementation and voluntary food fortification. Birth Defects Res A Clin Mol Teratol 85(4):269–273
- 71. Hilder L (2016) Neural tube defects in Australia 2007–2011: national perinatal epidemiology and statistics unit, University of New South Wales. Available from: [https://npesu.unsw.edu.au/sites/](https://npesu.unsw.edu.au/sites/default/files/npesu/surveillances/NTD%20Australia%200711_1.pdf) [default/files/npesu/surveillances/NTD%20Australia%200711_1.](https://npesu.unsw.edu.au/sites/default/files/npesu/surveillances/NTD%20Australia%200711_1.pdf) [pdf](https://npesu.unsw.edu.au/sites/default/files/npesu/surveillances/NTD%20Australia%200711_1.pdf). Accessed 4 May 2022
- 72. Brown RD, Langshaw MR, Uhr EJ, Gibson JN, Joshua DE (2011) The impact of mandatory fortification of flour with folic acid on the blood folate levels of an Australian population. Med J Aust 194(2):65–67
- 73. Arinchin A, Gembicki M, Moschik K, Skalyzhenko A, Khmara I, Korytko N, Petrenko S, Gomolko N, Balakleevskaya V, Laptenok S, Bertollini R (2000) Goiter prevalance and urinary iodine excretion in Belarus children born after the Chernobyl accident. IDD Newsletter 16:7–9
- 74. Głąbska D, Książek A, Guzek D (2017) Development and validation of the brief folate-specific food frequency questionnaire for young women's diet assessment. Int J Environ Res Public Health 14(12)
- 75. Willett WC, Sampson L, Stampfer MJ, Rosner B, Bain C, Witschi J, Hennekens CH, Speizer FE (1985) Reproducibility and validity of a semiquantitative food frequency questionnaire. Am J Epidemiol 122(1):51–65
- 76. Institute of Medicine (US) Standing Committee on the Scientific Evaluation of Dietary Reference Intakes and its Panel on Folate, Other B Vitamins, and Choline (1998) Dietary reference intakes for thiamin, riboflavin, niacin, vitamin B6, folate, vitamin B12, pantothenic acid, biotin, and choline. National Academies Press (US), Washington (DC). PMID: 23193625
- 77. Imhoff-Kunsch B, Flores R, Dary O, Martorell R (2007) Wheat flour fortification is unlikely to benefit the neediest in Guatemala. J Nutr 137(4):1017–1022
- 78. Tsang B, Stadnik C, Duong M, Pachon H, Martinez H (2020) Novel foods with potential for folic acid fortification in LMIC. Micronutrient Forum CONNECTED! 5th Global Conference; Nov 9–13, 2020; Bangkok, Thailand
- 79. Matthias D, McDonald CM, Archer N, Engle-Stone R (2022) The role of multiply-fortified table salt and Bouillon in food systems transformation. Nutrients 14(5):989
- 80. Kancherla V, Tsang B, Wagh K, Dixon M, Oakley GP Jr (2020) Modeling shows high potential of folic acid-fortified salt to accelerate global prevention of major neural tube defects. Birth Defects Res 112(18):1461–1474
- 81. Luthringer CL, Rowe LA, Vossenaar M, Garrett GS (2015) Regulatory monitoring of fortified foods: identifying barriers and good practices. Glob Health Sci Pract 3(3):446–461
- 82. Osendarp SJM, Martinez H, Garrett GS, Neufeld LM, De-Regil LM, Vossenaar M, Darnton-Hill I (2018) Large-scale food fortification and biofortification in low- and middle-income countries: a review of programs, trends, challenges, and evidence gaps. Food Nutr Bull 39(2):315–331
- 83. Martinez H, Weakland AP, Bailey LB, Botto LD, De-Regil LM, Brown KH (2018) Improving maternal folate status to prevent infant neural tube defects: working group conclusions and a framework for action. Ann N Y Acad Sci 1414(1):5–19
- 84. 45FR6323. as amended at 58 FR 2228, Jan. 6, 1993. Jan. 25, 1980
- 85. 42FR14483. Title 21 Chapter I Subchapter B Part 170 CFR, 2022, Title 21 Chapter I Subchapter B Part 170. Mar. 15, 1977
- 86. 21CFR104.20; Subpart B. 45 FR 6323, Jan. 25, 1980, as amended at 58 FR 2228, Jan. 6, 1993
- 87. Institute of Medicine, Food and Nutrition Board (1998) The national academies collection: reports funded by national institutes of health. Dietary reference intakes: a risk assessment model for establishing upper intake levels for nutrients. Washington (DC): National Academies Press (US), National Academy of Sciences
- 88. Kancherla V, Botto LD, Rowe LA, Shlobin NA, Caceres A, Arynchyna-Smith A, Zimmerman K, Bount J, Kibruyisfaw A, Ghotme KA, Karmarkar S, Fieggen G, Roozen S, Oakley GPJr, Rosseau G, Berry RJ (2022) Preventing birth defects, saving lives,

and promoting health equity: an urgent call to action for universal mandatory food fortification with folic acid. Lancet Glob Health

89. Pfeiffer CM, Zhang M, Jabbar S (2018) Framework for laboratory harmonization of folate measurements in low- and middle-income countries and regions. Ann N Y Acad Sci 1414(1):96–108

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