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## Influence of N and non-N salts on atmospheric methane oxidation by upland boreal forest and tundra soils

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**Abstract** The short-term (24 h) and medium-term (30 day) influence of N salts ( $\text{NH}_4\text{Cl}$ ,  $\text{NaNO}_3$  and  $\text{NaNO}_2$ ) and a non-N salt ( $\text{NaCl}$ ) on first-order rate constants,  $k$  ( $\text{h}^{-1}$ ) and thresholds ( $C_{\text{Th}}$ ) for atmospheric  $\text{CH}_4$  oxidation by homogenized composites of upland boreal forest and tundra soils was assessed at salt additions ranging to  $20 \mu\text{mol g}^{-1}$  dry weight (dw) soil. Additions of  $\text{NH}_4\text{Cl}$ ,  $\text{NaNO}_3$  and  $\text{NaCl}$  to  $0.5 \mu\text{mol g}^{-1}$  dw soil did not significantly decrease  $k$  relative to watered controls in the short term. Higher concentrations significantly reduced  $k$ , with the degree of inhibition increasing with increasing dose. Similar doses of  $\text{NH}_4\text{Cl}$  and  $\text{NaCl}$  gave comparable decreases in  $k$  relative to controls and both soils showed low native concentrations of  $\text{NH}_4^+\text{-N}$  ( $\leq 1 \mu\text{mol g}^{-1}$  dw soil), suggesting that the reduction in  $k$  was due primarily to a salt influence rather than competitive inhibition of  $\text{CH}_4$  oxidation by exogenous  $\text{NH}_4^+\text{-N}$  or  $\text{NH}_4^+\text{-N}$  released through cation exchange. The decrease in  $k$  was consistently less for  $\text{NaNO}_3$  than for  $\text{NH}_4\text{Cl}$  and  $\text{NaCl}$  at similar doses, pointing to a strong inhibitory effect of the  $\text{Cl}^-$  counteranion. Thresholds for  $\text{CH}_4$  oxidation were less sensitive to salt addition than  $k$  for these three salts, as significant increases in  $C_{\text{Th}}$  relative to controls were only observed at concentrations  $\geq 1.0 \mu\text{mol g}^{-1}$  dw soil. Both soils were more sensitive to  $\text{NaNO}_2$  than to other salts in the short term, showing a significant decrease in  $k$  at an addition of  $0.25 \mu\text{mol NaNO}_2 \text{ g}^{-1}$  dw soil that was clearly attributable to  $\text{NO}_2^-$ . Soils showed no recovery from  $\text{NaCl}$ ,  $\text{NH}_4^+\text{-N}$  or  $\text{NaNO}_3$  addition with respect to atmospheric  $\text{CH}_4$  oxidation after 30 days. However, soils amended with  $\text{NaNO}_2$  to  $1.0 \mu\text{mol NaNO}_2 \text{ g}^{-1}$  dw showed values of  $k$  that were not significantly different

from controls. Recovery of  $\text{CH}_4$ -oxidizing ability was due to complete oxidation of  $\text{NO}_2^-\text{-N}$  to  $\text{NO}_3^-\text{-N}$ . Analysis of soil concentrations of N salts necessary to inhibit atmospheric  $\text{CH}_4$  oxidation and regional rates of N deposition suggest that N deposition will not decrease the future sink strength of upland high-latitude soils in the atmospheric  $\text{CH}_4$  budget.

**Key words** Methane oxidation · Nitrogen fertilization · Ammonium · Tundra soil · Upland boreal soil

### Introduction

A contemporary increase in the atmospheric concentration of the radiatively important greenhouse gas  $\text{CH}_4$  is well documented (IPCC 1996).  $\text{CH}_4$  currently contributes approximately 20% to global warming (Bouman 1990), but on a mass basis has a global warming potential 56 times that of  $\text{CO}_2$  in a 20-year time horizon (IPCC 1996). Microbial oxidation in well-drained soils is the only identified biological sink for atmospheric  $\text{CH}_4$  and accounts for 3–9% of total annual atmospheric  $\text{CH}_4$  destruction (Prather et al. 1995). This is similar in magnitude to the current atmospheric increase of  $37 \text{ Tg CH}_4 \text{ year}^{-1}$  (IPCC 1996). Consequently, alterations of the soil sink strength are a significant determinant of the rate of change in the atmospheric  $\text{CH}_4$  concentration (Prather et al. 1995) and absence of this sink will cause the atmospheric  $\text{CH}_4$  concentration to increase at 1.5 times the current rate (Duxbury 1994).

Influences on rates of atmospheric  $\text{CH}_4$  oxidation include soil temperature, moisture and N status (King 1992, 1997; Mancinelli 1995) as well as land use patterns (e.g. Ojima et al. 1993; Willison et al. 1995; Hudgens and Yavitt 1997; Boeckx et al. 1998). Results of studies assessing the influence of N fertilization on  $\text{CH}_4$  consumption in aerobic soils give conflicting results between soils, the form and mass of N applied and the duration of inhibition (e.g. Nesbit and Breitenbeck

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1992; Adamsen and King 1993; Hütsch et al. 1993, 1994; Dunfield et al. 1995). N fertilization frequently reduces soil CH<sub>4</sub> consumption (e.g. Steudler et al. 1989; Mosier et al. 1991; King and Schnell 1994a). However, some studies report no effect of N fertilization (Bronson and Mosier 1993; Tate and Striegl 1993; Flessa et al. 1995; Delgado and Mosier 1996) or even a stimulatory effect on CH<sub>4</sub> consumption (Knightly et al. 1995; Christensen et al. 1997; Börjesson and Nohrstedt 1998; De Visscher et al. 1999) and induction of CH<sub>4</sub> oxidizing activity (Bender and Conrad 1995). Further, positive correlations has been observed between NH<sub>4</sub><sup>+</sup> concentration and CH<sub>4</sub> consumption (Goldman et al. 1995) and between soil fertility and CH<sub>4</sub> consumption (Castro et al. 1995) for temperate forest soils. Clearly, the relationship between soil N and oxidation of atmospheric CH<sub>4</sub> in aerobic soils is not straightforward and broad generalizations across ecosystems are not possible.

The tundra and boreal forest are circumpolar and occupy ~17% of the earth's continental surface (Schultz 1995). These regions presently play a major role in the atmospheric CH<sub>4</sub> budget. Upland forest and tundra soils consume atmospheric CH<sub>4</sub> (Whalen et al. 1991), while poorly drained areas emit CH<sub>4</sub> to the atmosphere (Whalen and Reeburgh 1992). The fraction of the earth's terrestrial surface under agriculture is increasing (Adams et al. 1990) and agricultural soils have lower area-based rates of CH<sub>4</sub> consumption than aerated forests, shrublands and savannah (reviewed by King 1997). Inasmuch as tundra and boreal forests are generally unsuitable for this land use, aerated soils of these regions may be critical in slowing future increases in atmospheric CH<sub>4</sub>. Accurate prediction of future trends in soil CH<sub>4</sub> oxidation in these high-latitude soils requires an improved understanding of the physiology and controls of this microbial process.

Anthropogenic alteration of the global N cycle has resulted in a worldwide increase in atmospheric N deposition on terrestrial and aquatic environments (Matthews 1994; Galloway et al. 1995). Increased N input may affect rates of CH<sub>4</sub> oxidation in high-latitude soils.

Consequently, this laboratory study was aimed at assessing the N sensitivity of microbial communities oxidizing atmospheric CH<sub>4</sub> in representative tundra and boreal forest soils. In particular, the immediate influence of different forms of N and non-N salts and mass of salt applied on rates and thresholds for CH<sub>4</sub> oxidation and the recovery of CH<sub>4</sub>-oxidizing activity after 30 day were determined.

## Materials and methods

### Field sites

Soils were collected in September 1995 from representative boreal forest (BS2) and upland tundra T3 sites in Alaska. Sites BS2 and T3 are located in the Bonanza Creek Experimental Forest (64°N, 148°W) and in the Arctic Long Term Experimental Re-

search area near Toolik Lake (68°N, 149°W), respectively. Site BS2 is a north-facing black spruce (*Picea mariana*) stand with a continuous ground cover of feather mosses (*Pleurozium* spp. and *Hylocomium* spp.) invaded by lowbush cranberry (*Vaccinium vitis-idaea*) and lichens. The organic horizon extends to about 10 cm. Soils are a well-drained, stone-free micaceous loess. Site T3 is a moderately exposed, south-facing sandstone outcrop on a late Wisconsin moraine. Moss (*Sphagnum* spp.), avens (*Dryas octopetala*), willow (*Salix* spp.) and lichens (*Cetraria* spp., *Cladonia* spp.) dominate the plant community. A 2-cm organic layer overlies soils that are a sandy loam rich in rock fragments. Bedrock occurs at a depth of 10–20 cm. These sites have no soil zone of methanogenesis and are atmospheric CH<sub>4</sub> sinks (Whalen et al. 1991 and unpublished data).

### Experimental

Bulk soils were collected from the zone of maximum CH<sub>4</sub>-oxidizing activity at BS2 (10–20 cm) and T3 (5–15 cm). Soils were sieved (4 mm mesh) and homogenized and the influence of N salts (NaNO<sub>3</sub>, NaNO<sub>2</sub>, NH<sub>4</sub>Cl), a non-N salt consisting of the counterions of the selected N salts (NaCl), and concentration (0.25, 0.5, 1, 2, 5, 10 and 20 μmol N or NaCl g<sup>-1</sup> dw soil) on the threshold and rate of CH<sub>4</sub> oxidation were assessed. Reduced concentrations of NaNO<sub>2</sub> (0.025, 0.05, 0.075, 0.10, 0.15, 0.20 and 0.25 μmol N g<sup>-1</sup> dw soil) were also tested. The high (0.25–20 μmol N g<sup>-1</sup> dw soil) and low range (0.025–0.25 μmol N g<sup>-1</sup> dw soil) NaNO<sub>2</sub> additions were designated experiments A and B, respectively. Ten grams dry weight equivalent of soil was added to each of several 160-cm<sup>3</sup> serum vials. One milliliter of deionized H<sub>2</sub>O (control) or chemical solution was evenly spread over the surface of soils to give three replicates for controls and each N or NaCl concentration listed above. Perforated foil was placed on the tops of serum vials to allow air exchange between the headspace and the atmosphere and samples were acclimated overnight (~12 h) at room temperature with no evaporative water loss. On the following morning, all vial headspaces were equilibrated with the laboratory atmosphere (~1.9 μl CH<sub>4</sub> l<sup>-1</sup>) for 2 h and experiments to assess the short-term effect of N amendment commenced by sealing the vials with butyl stoppers. Immediately after sealing and at 1.5-h intervals to 4.5 h, 3 cm<sup>3</sup> N<sub>2</sub> was injected into each vial and a 3-cm<sup>3</sup> sample was immediately removed for CH<sub>4</sub> analysis. Following the 4.5-h sampling, each vial was re-equilibrated with the laboratory atmosphere and sealed. A sample was withdrawn after 24 h to measure the threshold for CH<sub>4</sub> oxidation (C<sub>Th</sub>). Previous experience (Whalen et al. 1992) indicated that after this time headspace CH<sub>4</sub> concentrations would be reduced to the point where these soils would not consume additional CH<sub>4</sub>. Vials remained sealed and headspaces were adjusted to 10 μl CH<sub>4</sub> l<sup>-1</sup>.

Samples were stored at 15 °C (a typical summer soil temperature; Whalen et al. 1991) for 30 days. Headspaces of randomly chosen controls were analyzed for CH<sub>4</sub> at 2- to 4-day intervals and when concentrations were reduced to 1 μl CH<sub>4</sub> l<sup>-1</sup> all vials were flushed and headspaces were readjusted to 10 μl CH<sub>4</sub> l<sup>-1</sup>. At 30 days, time courses for CH<sub>4</sub> consumption were again assessed as described above (experiment B for NaNO<sub>2</sub> excluded) to give a measure of the medium-term effect of these salts on CH<sub>4</sub> oxidation. Following this second time course, the triplicate samples for each level and form of salt added were composited and analyzed for NO<sub>3</sub><sup>-</sup>-N + NO<sub>2</sub><sup>-</sup>-N (hereafter referred to as NO<sub>3</sub><sup>-</sup>-N), NO<sub>2</sub><sup>-</sup>-N and NH<sub>4</sub><sup>+</sup>-N.

### Methane analysis

CH<sub>4</sub> was measured using a Shimadzu GC8A flame ionization detection gas chromatograph fitted with a 0.5-ml sample loop. Gases were separated on a 1-m molecular sieve 5A column operated at 70 °C with an N<sub>2</sub> (33 ml min<sup>-1</sup>) carrier. The gas chromatograph was calibrated with commercial CH<sub>4</sub>-air blends following verification of stated concentrations with National Institute of Standards

and Technology standards. The precision of analysis expressed as a coefficient of variation for 10 replicate injections of standards (1.01–9.23  $\mu\text{l CH}_4 \text{ l}^{-1}$ ) was 0.3%. The detection limit was 0.05  $\mu\text{l CH}_4 \text{ l}^{-1}$ .

#### Soil physicochemical properties

Extractable (2M KCl; 10:1 volume/soil wet weight)  $\text{NO}_3^-$ -N,  $\text{NO}_2^-$ -N and  $\text{NH}_4^+$ -N were determined by the copperized cadmium reduction, modified Griess-Ilosvay and indophenol blue methods, respectively (Keeney and Nelson 1982), following sample filtration through a Whatman no. 42 filter. Soil pH was measured potentiometrically on 1:2 soil-deionized water slurries equilibrated for 24 h. Gravimetric water-holding capacity (WHC) was determined as the difference in weight between water-saturated and oven-dried (105 °C) soils. Organic content was determined as loss on ignition (550 °C) of oven-dried soils. Soil texture was assessed hydrometrically and total N and total C were determined by dry combustion (Carlo Erba NA 1500 Elemental Analyzer). Cation exchange capacity was determined by the  $\text{NH}_4\text{OAc}$  method (Thomas 1982).

#### Statistical analyses and calculations

All  $\text{CH}_4$  concentrations were corrected for dilution by added  $\text{N}_2$  and the effect of previous sampling (Breitenbeck 1990) in time course experiments and first-order rate constants for  $\text{CH}_4$  oxidation,  $k$  ( $\text{h}^{-1}$ ), were calculated from linear regression of  $\ln$ -transformed data for concentration versus time. The influence of concentration on  $k$  and  $C_{\text{Th}}$  was evaluated by single-factor analysis of variance (ANOVA) for each salt. Multiple comparisons of treatment means were performed by Tukey's honestly significant difference procedure. Treatments that totally inhibited  $\text{CH}_4$  oxidation were excluded from the ANOVA and were simply assumed to differ significantly from other treatments. The remaining data satisfied assumptions of normality and homoscedasticity without transformation. Statistical analyses were performed using the SYSTAT (SYSTAT Inc. Chicago, Ill.) statistical package for personal computers. A significance level  $\alpha=0.05$  was used for all tests.

## Results

Soils at both sites were acidic and generally comparable with respect to measured chemical properties (Table 1). Most of the inorganic N was as  $\text{NH}_4^+$ -N. Site T3 soils were relatively wetter (52% WHC) than BS2 soils (29% WHC at the time of collection). Liquid amendments during experimentation increased soil moisture to 66% and 41% WHC for T3 and BS2 soils, respectively. The major difference between soils related to texture, as BS2 was predominantly silt while T3 showed a high sand content.

Soils from T3 showed more vigorous  $\text{CH}_4$  oxidation than soils from BS2. First-order rate constants for  $\text{CH}_4$  oxidation by watered controls were higher for all treatments by a factor of 3 for T3 than for BS2 soils (Tables 2, 3).

Addition of  $\text{NH}_4\text{Cl}$  or  $\text{NaCl}$  at  $\leq 0.5 \mu\text{mol g}^{-1}$  dw soil did not significantly decrease  $k$  with respect to controls in either soil in short-term response experiments (Tables 2, 3). Similar data for  $\text{NaNO}_3$  showed no effect at  $\leq 1.0 \mu\text{mol N g}^{-1}$  dw soil and  $\leq 0.5 \mu\text{mol N g}^{-1}$  dw

**Table 1** Physicochemical characteristics of boreal forest (BS2) and arctic tundra (T3) soils

Characteristic	BS2	T3
pH ( $\text{H}_2\text{O}$ )	4.5	4.2
WHC (% dry weight) <sup>a</sup>	80	66
% WHC <sup>b</sup>	29	52
Organic matter (% w/w)	7.4	8.4
Total C (% w/w)	2.2	3.0
Total N (% w/w)	0.12	0.17
$\text{NH}_4^+$ ( $\mu\text{mol N g}^{-1}$ dw)	0.8	1.0
$\text{NO}_3^-$ ( $\mu\text{mol N g}^{-1}$ dw)	0.0	0.1
Sand/silt/clay (%)	9/88/3	69/26/5
CEC <sup>c</sup>	15.8	12.3

<sup>a</sup> Water holding capacity

<sup>b</sup> Field-moist soils

<sup>c</sup> Cation exchange capacity (mEq 100  $\text{g}^{-1}$  dw at pH 7)

soil for BS2 and T3 soils, respectively. Thus, there was no clear indication of greater sensitivity of  $k$  to any of these salts at additions to at least  $0.5 \mu\text{mol g}^{-1}$  dw soil. Following the onset of inhibition of  $\text{CH}_4$  oxidation by these three salts (72–89% of control values of  $k$ ), the degree of inhibition increased with increasing dose in all cases. However, there was no clear evidence of consistently greater relative inhibition (i.e. lower % of control) by  $\text{NH}_4\text{Cl}$  than  $\text{NaCl}$  at comparable doses for either soil. In contrast, relative inhibition was consistently greater for  $\text{NH}_4\text{Cl}$  and  $\text{NaCl}$  than  $\text{NaNO}_3$  at comparable doses from 1.0 to  $20 \mu\text{mol g}^{-1}$  dw soil for both soils.

In contrast to the other salts, the lowest level of added  $\text{NaNO}_2$  in experiment A ( $0.25 \mu\text{mol g}^{-1}$  dw soil) significantly decreased  $k$  relative to controls in both soils (Tables 2, 3). An additional experiment (B) involving lower additions of  $\text{NaNO}_2$  showed that  $k$  values for concentrations up to and including  $0.25 \mu\text{mol N g}^{-1}$  dw soil were not clearly distinguishable from controls in both soils. Collectively, the results of experiments A and B indicate that the onset of inhibition of  $\text{CH}_4$  oxidation by  $\text{NaNO}_2$  occurs at or slightly above  $0.25 \mu\text{mol NaNO}_2 \text{ g}^{-1}$  dw soil.

The highest concentration of  $\text{NH}_4\text{Cl}$  and  $\text{NaCl}$  tested ( $20 \mu\text{mol g}^{-1}$  dw soil) terminated  $\text{CH}_4$  oxidation for BS2 soil (Table 2). However,  $k$  for this dose was 23–25% of controls for T3 soils, pointing to a lower sensitivity to high additions of these salts (Table 3). Likewise, T3 soil was less sensitive to high levels of  $\text{NaNO}_2$  addition in experiment A. The  $k$  value was reduced to zero at  $1.0 \mu\text{mol NaNO}_2 \text{ g}^{-1}$  dw soil in T3 soil compared with  $0.5 \mu\text{mol NaNO}_2 \text{ g}^{-1}$  dw soil in BS2 soil. Soils were essentially identical in sensitivity to  $\text{NaNO}_3$  additions in excess of  $5.0 \mu\text{mol g}^{-1}$  dw soil.

Soil from T3 showed generally lower values for  $C_{\text{Th}}$  after addition of  $\text{NH}_4\text{Cl}$ ,  $\text{NaNO}_3$  or  $\text{NaCl}$  at comparable doses than soil from BS2 in short-term response experiments (Tables 4, 5). Values for  $C_{\text{Th}}$  and  $k$  were equally sensitive to high level  $\text{NaNO}_2$  addition (experiment A), as the significant changes in both occurred at the same

**Table 2** Mean ( $n=3$ ) first-order rate constants ( $k$ ) for  $\text{CH}_4$  oxidation and (% of control) for BS2 soil as a function of form and concentration of added salt. First-order rate constants were deter-

mined about 12 h after salt addition. Values of  $k$  followed by the same letter within each column do not differ significantly

Concentration ( $\mu\text{mol g}^{-1} \text{dw}$ ) <sup>a</sup>	$k$ ( $\text{h}^{-1}$ )				
	$\text{NH}_4\text{Cl}$	$\text{NaNO}_3$	$\text{NaNO}_2$ (A)	$\text{NaNO}_2$ (B)	$\text{NaCl}$
0 (0) control	0.065 a	0.055 a	0.061 a	0.052 a	0.051 a
0.25 (0.025)	0.065 (100) a	0.054 (98) a	0.009 (15) b	0.055 (106) a	0.053 (104) a
0.5 (0.05)	0.058 (89) ab	0.055 (100) a	0.000 (0)	0.048 (92) ab	0.053 (104) a
1.0 (0.075)	0.051 (78) b	0.052 (95) a	0.000 (0)	0.049 (94) ab	0.040 (78) b
2.0 (0.10)	0.039 (60) c	0.040 (73) b	0.000 (0)	0.056 (108) a	0.038 (75) b
5.0 (0.15)	0.021 (32) d	0.028 (51) c	0.000 (0)	0.050 (96) ab	0.017 (33) c
10.0 (0.20)	0.016 (25) d	0.021 (38) cd	0.000 (0)	0.045 (87) ab	0.000 (0)
20.0 (0.25)	0.000 (0)	0.016 (29) d	0.000 (0)	0.042 (81) b	0.000 (0)

<sup>a</sup> Concentrations in parentheses are for  $\text{NaNO}_2$  (B), while those not in parentheses are for all other amendments

**Table 3** Mean ( $n=3$ ) first-order rate constants ( $k$ ) for  $\text{CH}_4$  oxidation and (% of control) for T3 soil as a function of form and concentration of added salt. First-order rate constants were deter-

mined about 12 h after salt addition. Values of  $k$  followed by the same letter within each column do not differ significantly

Concentration ( $\mu\text{mol g}^{-1} \text{dw}$ ) <sup>a</sup>	$k$ ( $\text{h}^{-1}$ )				
	$\text{NH}_4\text{Cl}$	$\text{NaNO}_3$	$\text{NaNO}_2$ (A)	$\text{NaNO}_2$ (B)	$\text{NaCl}$
0 (0) control	0.242 a	0.223 a	0.203 a	0.208 a	0.196 a
0.25 (0.025)	0.240 (99) a	0.218 (98) a	0.147 (72) b	0.203 (98) a	0.203 (104) a
0.5 (0.05)	0.231 (95) a	0.219 (98) a	0.041 (20) c	0.205 (99) a	0.201 (103) a
1.0 (0.075)	0.185 (76) b	0.198 (89) b	0.000 (0) d	0.198 (95) a	0.141 (72) b
2.0 (0.10)	0.143 (59) c	0.169 (76) c	0.000 (0) d	0.212 (102) a	0.106 (54) c
5.0 (0.15)	0.105 (43) d	0.122 (55) d	0.000 (0) d	0.201 (97) a	0.097 (50) c
10.0 (0.20)	0.083 (34) e	0.088 (39) e	0.000 (0) d	0.194 (93) a	0.088 (45) c
20.0 (0.25)	0.055 (23) f	0.065 (29) f	0.000 (0) d	0.195 (94) a	0.049 (25) d

<sup>a</sup> Concentrations in parentheses are for  $\text{NaNO}_2$  (B), while those not in parentheses are for all other amendments

**Table 4** Mean ( $n=3$ ) threshold for  $\text{CH}_4$  oxidation ( $C_{\text{Th}}$ ) by BS2 soil as a function of form and concentration of added salt. Threshold values followed by the same letter within each column do not differ significantly

Concentration ( $\mu\text{mol g}^{-1} \text{dw}$ ) <sup>a</sup>	$C_{\text{Th}}$ ( $\mu\text{l CH}_4 \text{l}^{-1}$ )				
	$\text{NH}_4\text{Cl}$	$\text{NaNO}_3$	$\text{NaNO}_2$ (A)	$\text{NaNO}_2$ (B)	$\text{NaCl}$
0 (0) control	0.21 a	0.22 a	0.24 a	0.24 a	0.30 a
0.25 (0.025)	0.20 a	0.20 a	1.56 b	0.26 a	0.34 a
0.5 (0.05)	0.17 a	0.23 a	1.81 c	0.29 a	0.29 a
1.0 (0.075)	0.21 a	0.23 a	1.78 c	0.39 ab	0.43 a
2.0 (0.10)	0.38 b	0.33 b	1.79 c	0.47 b	0.52 a
5.0 (0.15)	1.00 c	0.48 c	1.81 c	0.42 b	1.05 b
10.0 (0.20)	1.40 d	0.71 d	1.80 c	0.31 ab	1.39 c
20.0 (0.25)	1.71 e	0.95 e	1.81 c	0.43 b	1.70 d

<sup>a</sup> Concentrations in parentheses are for  $\text{NaNO}_2$  (B), while those not in parentheses are for all other amendments

concentration of  $\text{NaNO}_2$  within each soil. Otherwise,  $C_{\text{Th}}$  was frequently less sensitive than  $k$  to concentration of added salts. Values of  $C_{\text{Th}}$  for BS2 soil became significantly higher with respect to controls at 2.0  $\mu\text{mol g}^{-1} \text{dw}$  soil for  $\text{NH}_4\text{Cl}$  and  $\text{NaNO}_3$  and at 5.0  $\mu\text{mol g}^{-1} \text{dw}$  soil for  $\text{NaCl}$ . Similarly, a significant increase in  $C_{\text{Th}}$  relative to controls was not observed for T3 soil until concentrations of  $\text{NH}_4\text{Cl}$ ,  $\text{NaNO}_3$  and  $\text{NaCl}$  were  $\geq 5.0 \mu\text{mol g}^{-1} \text{dw}$  soil.

Determinations of  $k$  30 days after salt additions (medium-term effect) showed no clear evidence of recovery from salt amendment, with the exception of the high (experiment A)  $\text{NaNO}_2$  addition experiment (Tables 6, 7). Values of  $k$  for  $\text{NH}_4\text{Cl}$ ,  $\text{NaNO}_3$  and  $\text{NaCl}$  addition were not significantly different from controls for concentrations as high as 0.25–1.0  $\mu\text{mol g}^{-1} \text{dw}$  soil, depending on the soil and salt added. The medium-term effect of  $\text{NH}_4\text{Cl}$ ,  $\text{NaNO}_3$  and  $\text{NaCl}$  additions appeared

**Table 5** Mean ( $n=3$ ) threshold for CH<sub>4</sub> oxidation ( $C_{Th}$ ) by T3 soil as a function of form and concentration of added salt. Threshold values followed by the same letter within each column do not differ significantly

Concentration ( $\mu\text{mol g}^{-1} \text{ dw}$ ) <sup>a</sup>	$C_{Th}$ ( $\mu\text{l CH}_4 \text{ l}^{-1}$ )				
	NH <sub>4</sub> Cl	NaNO <sub>3</sub>	NaNO <sub>2</sub> (A)	NaNO <sub>2</sub> (B)	NaCl
0 (0) control	0.06 a	0.10 a	0.17 a	0.18 a	0.19 a
0.25 (0.025)	0.07 a	0.11 a	0.32 b	0.17 a	0.15 a
0.5 (0.05)	0.09 a	0.12 a	0.65 c	0.19 ab	0.17 a
1.0 (0.075)	0.09 a	0.13 a	1.81 d	0.17 a	0.17 a
2.0 (0.10)	0.10 a	0.12 a	1.82 d	0.17 a	0.20 a
5.0 (0.15)	0.13 ab	0.13 a	1.80 d	0.17 a	0.17 a
10.0 (0.20)	0.19 b	0.13 a	1.80 d	0.22 b	0.28 b
20.0 (0.25)	0.41 c	0.23 b	1.81 d	0.18 a	0.32 b

<sup>a</sup> Concentrations in parentheses are for NaNO<sub>2</sub> (B), while those not in parentheses are for all other amendments

**Table 6** Mean ( $n=3$ ) first-order rate constants ( $k$ ) for CH<sub>4</sub> oxidation and (% of control) for BS2 soil as a function of form and concentration of added salt 1 month after salt addition. Values of  $k$  followed by the same letter within each column do not differ significantly

Concentration ( $\mu\text{mol g}^{-1} \text{ dw}$ )	$k$ ( $\text{h}^{-1}$ )			
	NH <sub>4</sub> Cl	NaNO <sub>3</sub>	NaNO <sub>2</sub>	NaCl
0 (control)	0.059 a	0.059 a	0.062 a	0.059 a
0.25	0.038 (65) ab	0.050 (85) a	0.052 (84) ab	0.058 (98) a
0.5	0.045 (76) a	0.032 (54) b	0.051 (82) ab	0.054 (92) a
1.0	0.037 (63) ab	0.036 (61) b	0.047 (76) ab	0.039 (66) b
2.0	0.024 (41) b	0.029 (49) b	0.038 (61) b	0.041 (69) b
5.0	0.000 (0)	0.000 (0)	0.000 (0)	0.000 (0)
10.0	0.000 (0)	0.000 (0)	0.000 (0)	0.000 (0)
20.0	0.000 (0)	0.000 (0)	0.000 (0)	0.000 (0)

**Table 7** Mean ( $n=3$ ) first-order rate constants ( $k$ ) for CH<sub>4</sub> oxidation and (% of control) for T3 soil as a function of form and concentration of added nutrient 1 month after nutrient addition. Values of  $k$  followed by the same letter within each column do not differ significantly

Concentration ( $\mu\text{mol g}^{-1} \text{ dw}$ )	$k$ ( $\text{h}^{-1}$ )			
	NH <sub>4</sub> Cl	NaNO <sub>3</sub>	NaNO <sub>2</sub>	NaCl
0 (control)	0.223 a	0.212 a	0.202 a	0.190 a
0.25	0.186 (83) ab	0.177 (83) a	0.207 (102) a	0.195 (103) a
0.5	0.200 (90) ab	0.172 (81) a	0.200 (99) a	0.196 (103) a
1.0	0.168 (75) bc	0.164 (77) a	0.218 (108) a	0.075 (39) b
2.0	0.139 (62) c	0.126 (59) b	0.184 (91) a	0.078 (41) b
5.0	0.000 (0)	0.000 (0)	0.000 (0)	0.032 (17) c
10.0	0.000 (0)	0.000 (0)	0.000 (0)	0.000 (0)
20.0	0.000 (0)	0.000 (0)	0.000 (0)	0.000 (0)

to be greater than the short-term effect at high concentrations. Doses  $\geq 5.0 \mu\text{mol g}^{-1} \text{ dw}$  soil for these salts totally inhibited CH<sub>4</sub> oxidation in both soils, except for NaCl amendment to T3, where only the 10.0 and 20.0  $\mu\text{mol N g}^{-1} \text{ dw}$  soil additions gave no CH<sub>4</sub> oxidation.

In contrast to the medium-term response to the other salt additions, CH<sub>4</sub> oxidation recovered from the lowest amendments in the high-level NaNO<sub>2</sub> addition experiment. Values of  $k$  for additions of  $\leq 1.0 \mu\text{mol NaNO}_2 \text{ g}^{-1} \text{ dw}$  soil (BS2) and  $\leq 2.0 \mu\text{mol NaNO}_2 \text{ g}^{-1} \text{ dw}$  soil (T3) were not significantly different from controls. Nonetheless, concentrations  $\geq 5.0 \mu\text{mol NaNO}_2 \text{ g}^{-1} \text{ dw}$  soil still fully inhibited CH<sub>4</sub> oxidation.

The point of total inhibition for salts was sharply defined in most cases in these medium-term experiments (Tables 6, 7), unlike the gradual decrease in  $k$  with in-

creasing dose that was characteristic for most salt amendments in short-term response experiments (Tables 2, 3). With the exception of NaCl for T3 soil,  $k$  ranged from 41% to 91% of controls at the dose immediately preceding that which effected total inhibition of atmospheric CH<sub>4</sub> oxidation.

Nutrient analysis of soils following the 30-day determination of  $k$  indicated that all of the NO<sub>2</sub>-N in the high-level NaNO<sub>2</sub> addition experiment (experiment A) had been nitrified to NO<sub>3</sub>-N. N transformations were minimal in both soils for all other treatments. Observed concentrations were within 10% of expected concentrations (added N plus native N) for each form and level of N if no transformations are assumed.

## Discussion

The three-fold higher values of  $k$  consistently observed for T3 soil relative to BS2 soil (Tables 2, 3, 6, 7) likely stems from differences in soil texture (Table 1) and size of the resident methanotroph populations. Bender and Martin (1994) reported a higher oxidative capacity for fine sand than for silt and clay due to facilitated diffusion and preference of methanotrophs for the coarser-grained soil fractions.

Inhibition of atmospheric  $\text{CH}_4$  oxidation by  $\text{NH}_4^+$  salts has been demonstrated to involve competitive interactions between  $\text{CH}_4$  and  $\text{CH}_4$  monooxygenase as well as the toxic effects of  $\text{NO}_2^-$  and other intermediates (King and Schnell 1994b; Schnell and King 1994). The reported inhibition of atmospheric  $\text{CH}_4$  oxidation by other N and non-N salts (Nesbit and Breitenbeck 1992; Crill et al. 1994; Knightly et al. 1995; Nedwell 1996) may involve multiple mechanisms that include desorption of bound  $\text{NH}_4^+$ -N (King and Schnell 1998), a decrease in soil water potential (Schnell and King 1996) or other non-specific salt effects (Gulledge and Schimel 1998).

At concentrations at and above the point of inhibition of  $\text{CH}_4$  oxidation by  $\text{NH}_4\text{Cl}$  and  $\text{NaCl}$  in my short-term response experiments (generally  $\geq 1.0 \mu\text{mol g}^{-1}$  dw soil) with both soils, the degree of inhibition increased with increasing dose (Tables 2, 3) and the relative degree of inhibition was not consistently greater for  $\text{NH}_4\text{Cl}$  than  $\text{NaCl}$  at comparable doses. Observations of a similar or greater decrease in  $\text{CH}_4$  consumption by equimolar concentrations of a non- $\text{NH}_4^+$ -salt than an  $\text{NH}_4^+$  salt tested in parallel have often been interpreted (e.g. Gulledge et al. 1997; MacDonald et al. 1997) to indicate no effect of added  $\text{NH}_4^+$ -N, and have prompted MacDonald et al (1997) to question the competitive inhibition theory. Thus, results here suggest that an ionic effect rather than competitive  $\text{NH}_4^+$ -N inhibition depresses  $\text{CH}_4$  oxidation in the  $\text{NH}_4\text{Cl}$  addition experiment. Desorption of bound  $\text{NH}_4^+$ -N likely had little effect on  $\text{CH}_4$  oxidation in  $\text{NaCl}$ -treated soils. Although cation exchange capacities are low, exchangeable  $\text{NH}_4^+$ -N concentrations are only  $\leq 1 \mu\text{mol N g}^{-1}$  dw soil in unamended soil (Table 1).

The short-term response of  $\text{CH}_4$  oxidation to  $\text{NaNO}_3$  amendment was remarkably similar to that for  $\text{NH}_4\text{Cl}$  and  $\text{NaCl}$  addition (Tables 2, 3). Onset of inhibition occurred at a common dose and the degree of inhibition increased with increasing dose. However, the degree of inhibition for a  $\text{NaNO}_3$  was consistently less than for  $\text{NH}_4\text{Cl}$  and  $\text{NaCl}$  at a similar dose. King and Schnell (1998) reported that common anions including  $\text{Cl}^-$  and  $\text{NO}_2^-$  can vary in their effect on  $\text{NH}_4^+$ -N adsorption and desorption, and therefore have differing influences on atmospheric  $\text{CH}_4$  oxidation. Again, the low level of exchangeable  $\text{NH}_4^+$ -N in the study soils suggests little increased availability on salt addition. Gulledge and Schimel (1998) reported a severe inhibitory effect

of  $\text{Cl}^-$  salts in particular on  $\text{CH}_4$  oxidation in soils from varied environments, including a boreal forest birch stand low in exchangeable  $\text{NH}_4^+$ -N. Thus, the observation of lower inhibition of  $\text{CH}_4$  oxidation by  $\text{NaNO}_3$  than  $\text{NaCl}$  and  $\text{NH}_4\text{Cl}$  at similar doses may stem simply from a more severe non-specific salt effect associated with the  $\text{Cl}^-$  counter-anion.

More severe inhibition in  $\text{CH}_4$  oxidation at low  $\text{CH}_4$  concentrations by  $\text{NO}_2^-$ -N than  $\text{NH}_4^+$ -N at comparable doses in both soils (Tables 2, 3) is consistent with a similar report (Schnell and King 1994) for an acid forest soil that showed 59% and 42% inhibition following addition of  $\text{NaNO}_2$  and  $\text{NH}_4\text{Cl}$ , respectively, at  $1 \mu\text{mol N g}^{-1}$  fresh weight (fw) soil. Otherwise, Hütsch (1998) reported 84% inhibition of  $\text{CH}_4$  oxidation in a loamy arable soil within 24 h after  $\text{NaNO}_2$  addition at  $2.9 \mu\text{mol N g}^{-1}$  fw soil. The soils in this study are apparently extremely sensitive to  $\text{NO}_2^-$ -N, as concentrations similar to those tested in these other studies gave total inhibition of  $\text{CH}_4$  oxidation. Moreover, inhibition was clearly due to  $\text{NO}_2^-$ , as the effect was evident at doses where equinormal  $\text{NaCl}$  and other N salts had no influence on  $\text{CH}_4$  oxidation relative to controls.

Thresholds for atmospheric  $\text{CH}_4$  consumption by controls (Tables 4, 5) are consistent with other reports from high-latitude environments (Whalen et al. 1991, 1992) and elsewhere (Whalen et al. 1990; Yavitt et al. 1990; Koschorreck and Conrad 1993; Bender and Conrad 1993, 1995) that generally show values of  $0.1$  to  $0.3 \mu\text{l CH}_4 \text{ l}^{-1}$  for  $C_{\text{Th}}$ . The data showing a short-term increase in  $C_{\text{Th}}$  at high doses of all salts is in general agreement with the only other study assessing the influence of exogenous N on this parameter. There, Adamsen and King (1993) reported supra-atmospheric values of  $C_{\text{Th}}$  ( $>3 \mu\text{l CH}_4 \text{ l}^{-1}$ ) for a forest soil exposed to  $10 \mu\text{l CH}_4 \text{ l}^{-1}$  and amended with various  $\text{NH}_4^+$  and  $\text{NO}_3^-$ -N salts at  $1$ – $2 \mu\text{mol N g}^{-1}$  fw soil. With the exception of  $\text{NaNO}_2$ ,  $C_{\text{Th}}$  here appears to be less sensitive than  $k$  in both soils to salt amendment from the standpoint that values differ significantly from controls at higher doses within each treatment. Inasmuch as  $C_{\text{Th}}$  is probably a manifestation of the overall physiological status of methanotrophs as influenced by environmental conditions (King 1997), this observation of reduced sensitivity of  $C_{\text{Th}}$  than  $k$  to salt addition cannot be generalized. However, information concerning the sensitivity of  $C_{\text{Th}}$  to salt addition and changes in soil physicochemical variables is essential to predicting the future sink strength for the soil consumptive term in the atmospheric  $\text{CH}_4$  budget because  $C_{\text{Th}}$  can clearly be a determinant of overall  $\text{CH}_4$  consumption.

With the exception of  $\text{NaNO}_2$ , soils showed no evidence at 30 days for recovery of  $k$  from the effect of any salt addition. Studies that report the medium-term (days to months) influence of similar amendments give variable results. Nesbit and Breitenbeck (1992) and Schnell and King (1994) found persistent inhibition a month or more following addition of  $\text{NH}_4\text{Cl}$  to forest soils at  $7 \mu\text{mol N g}^{-1}$  fw soil and  $1 \mu\text{mol N g}^{-1}$  dw soil,

respectively. Similarly, Priemé and Christensen (1997) reported continued inhibition of CH<sub>4</sub> oxidation in forest soil to 27 days in response to 1.2 μmol g<sup>-1</sup> dw soil amendment with NH<sub>4</sub>Cl-N, KNO<sub>3</sub>-N and KCl while Hütsch et al. (1996) observed complete suppression of CH<sub>4</sub> oxidation to 7 days in a sandy arable soil amended with 16.8 μmol (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>-N g<sup>-1</sup> dw soil. Otherwise, data from Bronson and Mosier (1994) show either no difference or some decrease (~15%) in the degree of inhibition between 6 and 12 days for NH<sub>4</sub>Cl, KNO<sub>3</sub> or NH<sub>4</sub>NO<sub>3</sub> added to grassland soils at 1.8 μmol N g<sup>-1</sup> dw soil.

The medium-term recovery of *k* in soils amended with NaNO<sub>2</sub> at doses as high as 2 μmol g<sup>-1</sup> dw soil (Tables 6, 7) is clearly related to the complete oxidation of NO<sub>2</sub>-N to NO<sub>3</sub>-N. Hütsch (1998) also reported contemporaneous oxidation of exogenously added NO<sub>2</sub> (2.9 μmol N g<sup>-1</sup> dw soil) and a decrease in the inhibitory effect of the added substrate from 84% immediately after application to 41% after 48 h. Further, Dunfield and Knowles (1995) observed complete reversibility of NO<sub>2</sub> inhibition of CH<sub>4</sub> oxidation accompanied by the return of NO<sub>2</sub>-N to background levels 70 h after addition to humisol slurries. The low accumulation of NO<sub>3</sub>-N in NH<sub>4</sub>Cl-treated soils after 30 days and the complete recovery of NO<sub>3</sub>-N as NO<sub>3</sub>-N in NaNO<sub>2</sub>-treated soils indicates that NH<sub>4</sub><sup>+</sup>-N oxidation is the rate-limiting step in nitrification in these soils and is consistent with the observation by Gulledge et al. (1997) for other boreal forest soils that nitrifiers are slow in consuming NH<sub>4</sub><sup>+</sup>-N. Rapid oxidation of exogenous NO<sub>2</sub>-N and undetectable levels of NO<sub>2</sub>-N in NH<sub>4</sub>Cl-amended soils indicate that NO<sub>2</sub>-N will not inhibit in situ atmospheric CH<sub>4</sub> oxidation in these soils.

Partial (Hütsch 1998) or complete (Dunfield and Knowles 1995) recovery from NH<sub>4</sub><sup>+</sup>-N addition has been associated with high rates of nitrification. Except for NO<sub>2</sub>-N oxidation, rates of microbial N transformation were low in the soils in this study and concentrations of NO<sub>3</sub>-N and NH<sub>4</sub><sup>+</sup>-N were essentially static for 30 days following amendment. Thus, the observations that: (1) the normality at which *k* was significantly lower than controls was reasonably similar for NH<sub>4</sub>Cl, NaNO<sub>3</sub> and NaCl amendments in both short- and medium-term experiments; (2) the normality of salt addition that elicited the onset of inhibition essentially did not change over 30 days; (3) NO<sub>2</sub>-N oxidation in soils from experiment A resulted in recovery of CH<sub>4</sub> oxidation to levels comparable to other treatments; and (4) complete inhibition of CH<sub>4</sub> oxidation at 30 days at a level that was markedly similar among treatments all suggest that the inhibitory effects on CH<sub>4</sub> oxidation observed here for all treatments in both the short and medium term (exception: short-term NaNO<sub>2</sub> addition in experiment A) was due primarily to a salt influence.

N fertilization experiments of forest plots frequently show depressed levels of CH<sub>4</sub> consumption relative to unfertilized controls (e.g. Steudler et al. 1989; Sitaula et al. 1995) causing concern that increased atmospheric N

deposition may result in a decrease in atmospheric CH<sub>4</sub> consumption by upland soils (Castro et al. 1995). However, changes in soil CH<sub>4</sub> consumption may be an artifact in response to heavy N-fertilizer application rather than the natural reaction of the system to chronic, low-level atmospheric N deposition (Butterbach-Bahl et al. 1998). Thus, it may be equally as instructive to determine whether inhibitory levels of N can be achieved in these soils through atmospheric deposition. Data here can be unambiguously interpreted to indicate that NH<sub>4</sub><sup>+</sup> and NO<sub>3</sub><sup>-</sup> salts do not inhibit CH<sub>4</sub> oxidation regardless of the mechanism in either the short or medium-term at concentrations at least to 0.5 μmol N g<sup>-1</sup> dw (Tables 2, 3, 6 and 7). These concentrations represent 62% and 50% increases over prefertilization NH<sub>4</sub><sup>+</sup>-N levels for BS2 and T3 soils, respectively (Table 1). Assuming annual precipitation of 30 cm at both sites (Whalen et al. 1991; Kling et al. 1992), a soil bulk density of 0.91 g cm<sup>-3</sup> (Whalen et al. 1992), an average inorganic-N (NH<sub>4</sub><sup>+</sup>-N plus NO<sub>3</sub><sup>-</sup>-N) concentration of 5 μmol l<sup>-1</sup> in bulk precipitation (Galloway et al. 1984; Whalen and Cornwell 1985; Bormann et al. 1989) and distribution of the entire atmospheric N load homogeneously in a subsurface, 10-cm zone of CH<sub>4</sub> oxidation (Whalen et al. 1992), annual atmospheric N loading will be 0.016 μmol N g<sup>-1</sup> dw soil. A doubling of this annual load will represent 6% of the inorganic-N concentration necessary to inhibit atmospheric CH<sub>4</sub> oxidation. This simplistic analysis does not consider the influence of increased atmospheric deposition on N turnover, which may be more important than absolute concentration in determining sensitivity of atmospheric CH<sub>4</sub> oxidation to increased N loading (Mosier et al. 1991). Nonetheless, it suggests that substantial increases in atmospheric N deposition and N delivery to the active methanotrophic zone without plant assimilation will be necessary to inhibit atmospheric CH<sub>4</sub> oxidation in these soils. This corroborates a previous report (Whalen et al. 1991) that showed no effect of N fertilization (1:1 mix of urea-N and (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> up to 20 g N m<sup>-2</sup> year<sup>-1</sup>) on CH<sub>4</sub> oxidation in field plots at BS2.

In conclusion, NH<sub>4</sub>Cl, NaNO<sub>3</sub> and NaCl salts had no effect on atmospheric CH<sub>4</sub> oxidation by upland boreal forest and tundra soils in the short or medium term at amendments to 0.5 μmol g<sup>-1</sup> dw soil. Inhibition at higher amendments (to 20 μmol g<sup>-1</sup> dw soil) is probably due largely to a nonspecific salt effect and *k* appears to be more sensitive than C<sub>Th</sub> to salt addition in the short-term. Soils were extremely sensitive to NaNO<sub>2</sub> addition, but recovered the ability to oxidize atmospheric CH<sub>4</sub> after 30 days, when the exogenously supplied NO<sub>2</sub>-N had been oxidized to NO<sub>3</sub>-N. In contrast, suppression of CH<sub>4</sub> oxidation remained constant over 30 days in soils amended with high levels of NH<sub>4</sub>Cl, NaNO<sub>3</sub> and NaCl. Increased atmospheric N loading will likely not diminish the CH<sub>4</sub>-oxidizing capability of these soils.

## References

- Adams RM, Rosenzweig C, Peart RM, Ritchie JT, McCarl BA, Glycer JD, Curry RB, Jones JW, Boote KJ, Allen LH Jr (1990) Global climate change and U.S. agriculture. *Nature* 345:219–224
- Adamsen APS, King GM (1993) CH<sub>4</sub> consumption in temperate and subarctic forest soils: rates, vertical zonation and responses to water and nitrogen. *Appl Environ Microbiol* 59:485–490
- Bender M, Conrad R (1993) Kinetics of methane oxidation in oxic soils. *Chemosphere* 26:687–696
- Bender M, Conrad R (1994) Methane oxidation activity in various soils and freshwater sediments: occurrence, characteristics, vertical profiles, and distribution on grain size fractions. *J Geophys Res* 99:16531–16540
- Bender M, Conrad R (1995) Effect of CH<sub>4</sub> concentrations and soil conditions on the induction of CH<sub>4</sub> oxidation activity. *Soil Biol Biochem* 12:1517–1527
- Boeckx P, Van Cleemput O, Meyer T (1998) The influence of land use and pesticides on methane oxidation in some Belgian soils. *Biol Fertil Soils* 27:293–298
- Börjesson G, Nohrstedt H-Ö (1998) Short- and long-term effects of nitrogen fertilization on methane oxidation in three Swedish forest soils. *Biol Fertil Soils* 27:113–118
- Bormann BT, Tarrant RF, McClellan HM, Savage T (1989) Chemistry of rainwater and cloud water at remote sites in Alaska and Oregon. *J Environ Qual* 18:149–152
- Bouwman AF (1990) Introduction. In: Bouwman AF (ed) *Soils and the greenhouse effect*. Wiley, Chichester, pp 25–32
- Breitenbeck GA (1990) Sampling the atmosphere of small vessels. *Soil Sci Soc Am J* 54:1794–1797
- Bronson KF, Mosier AR (1993) Effect of nitrogen fertilizer and nitrification inhibitors on methane and nitrous oxide fluxes in irrigated corn. In: Oremland RS (ed) *Biogeochemistry of global change: radiatively active trace gases*. Chapman and Hall, New York, pp 278–289
- Bronson KF, Mosier AR (1994) Suppression of methane oxidation in aerobic soils by nitrogen fertilizers, nitrification inhibitors, and urease inhibitors. *Biol Fertil Soils* 17:263–268
- Butterbach-Bahl K, Gasche R, Huber CH, Kreutzer K, Papen H (1998) Impact of N-input by wet deposition on N-trace gas fluxes and CH<sub>4</sub>-oxidation in spruce forest ecosystems of the temperate zone of Europe. *Atmos Environ* 32:559–564
- Castro MS, Steudler PA, Melillo JM, Aber JD, Bowden RD (1995) Factors controlling atmospheric methane consumption by temperate forest soils. *Global Biogeochem Cycles* 9:1–10
- Christensen TR, Michelsen A, Jonasson S, Schmidt IK (1997) Carbon dioxide and methane exchange of a subarctic heath in response to climate change related environmental manipulations. *Oikos* 79:33–44
- Crill PM, Martikainen PJ, Nykänen H, Silvola J (1994) Temperature and N fertilization effects on methane oxidation in a drained peatland soil. *Soil Biol Biochem* 26:1331–1339
- Delgado JA, Mosier AR (1996) Mitigation alternatives to decrease nitrous oxide emissions and urea-nitrogen loss and their effect on methane flux. *J Environ Qual* 25:1105–1111
- De Visscher A, Thomas D, Boeckx P, Van Cleemput O (1999) Methane oxidation in a simulated landfill cover soil environment. *Environ Sci Technol* 33:1854–1859
- Dunfield P, Knowles R (1995) Kinetics of inhibition of methane oxidation by nitrate, nitrite, and ammonium in a humisol. *Appl Environ Microbiol* 61:3129–3135
- Dunfield PF, Topp E, Archambault C, Knowles R (1995) Effect of N fertilisers and moisture content on CH<sub>4</sub> and N<sub>2</sub>O fluxes in a humisol: measurements in the field and intact soil cores. *Biogeochemistry* 29:199–222
- Duxbury JM (1994) The significance of agricultural sources of greenhouse gases. *Fert Res* 38:151–163
- Flessa H, Dörsch P, Beese F (1995) Seasonal variations of N<sub>2</sub>O and CH<sub>4</sub> fluxes in differently managed arable soils in southern Germany. *J Geophys Res* 100:23115–23124
- Galloway JN (1984) Deposition of sulfur and nitrogen from the remote atmosphere. In: Galloway JN, Charlson RJ, Andreae MO, Rodhe H (eds) *The biogeochemical cycling of sulfur and nitrogen in the remote atmosphere*. Reidel, Dordrecht, pp 143–175
- Galloway JN, Schlesinger WH, Levy H II, Michaels A, Schnoor JL (1995) Nitrogen fixation: anthropogenic enhancement-environmental response. *Global Biogeochem Cycles* 9:235–252
- Goldman MB, Groffman PM, Pouyat RV, McDonnell MJ, Pickett STA (1995) CH<sub>4</sub> uptake and N availability in forest soils along an urban to rural gradient. *Soil Biol Biochem* 27:281–286
- Gulledge J, Schimel JP (1998) Moisture control over atmospheric CH<sub>4</sub> consumption and CO<sub>2</sub> production in diverse Alaskan soils. *Soil Biol Biochem* 30:1127–1132
- Gulledge J, Doyle AP, Schimel JP (1997) Different NH<sub>4</sub><sup>+</sup> inhibition patterns of soil CH<sub>4</sub> consumption: a result of distinct CH<sub>4</sub>-oxidizer populations across sites? *Soil Biol Biochem* 29:13–21
- Hudgens DE, Yavitt JB (1997) Land-use effects on soil methane and carbon dioxide fluxes in forests near Ithaca, New York. *Ecoscience* 4:214–222
- Hütsch BW (1998) Methane oxidation in arable soil as inhibited by ammonium, nitrite, and organic manure with respect to soil pH. *Biol Fertil Soils* 28:27–35
- Hütsch B, Webster WCP, Powlson DS (1993) Long-term effects of nitrogen fertilization on methane oxidation in soil of the Broadbalk wheat experiment. *Soil Biol Biochem* 25:1307–1315
- Hütsch B, Webster WCP, Powlson DS (1994) Methane oxidation in soils as affected by land use, soil pH and N fertilization. *Soil Biol Biochem* 26:1613–1622
- Hütsch BW, Russel P, Mengel K (1996) CH<sub>4</sub> oxidation in two temperate arable soils as affected by nitrate and ammonium application. *Biol Fertil Soils* 23:86–92
- IPCC (1996) Technical summary. In: Houghton, JT, Miera Filho LG, Callandar BA, Harriss N, Kattenberg A, Maskell K (eds) *Climate change 1995. The science of climate change*. Cambridge University Press, Cambridge, pp 1–49
- Keeney DR, Nelson DW (1982) Nitrogen – inorganic forms. In: Page AL (ed) *Methods of soil analysis – part 2. Chemical and microbiological methods*. American Society of Agronomy, Madison, Wis., pp 643–698
- King GM (1992) Ecological aspects of methane oxidation, a key determinant of global methane dynamics. *Adv Microb Ecol* 12:431–468
- King GM (1997) Responses of atmospheric methane consumption by soils to global climate change. *Global Change Biol* 3:351–362
- King GM, Schnell S (1994a) Effect of increasing atmospheric, methane concentration on ammonium inhibition of soil methane consumption. *Nature* 370:282–284
- King GM, Schnell S (1994b) Ammonium and nitrite inhibition of methane oxidation by *Methylobacter albus* BG8 and *Methylosinus trichosporium* OB3b at low methane concentrations. *Appl Environ Microbiol* 60:3508–3513
- King GM, Schnell S (1998) Effects of ammonium and non-ammonium salt additions on methane oxidation by *Methylosinus trichosporium* OB3b and Maine forest soils. *Appl Environ Microbiol* 64:253–257
- Kling GW, Kipphut GW, Miller MC (1992) The flux of CO<sub>2</sub> and CH<sub>4</sub> from lakes and rivers in arctic Alaska. *Hydrobiologia* 240:23–36
- Knightly D, Nedwell DB, Cooper M (1995) Capacity for methane oxidation in landfill cover soils measured in laboratory-scale soil microcosms. *Appl Environ Microbiol* 61:592–601
- Koschorreck M, Conrad R (1993) Oxidation of atmospheric methane in soil: Measurements in the field, in soil cores and in soil samples. *Global Biogeochem Cycles* 7:109–121



- MacDonald JA, Skiba U, Sheppard LJ, Ball B, Roberts JD, Smith KA, Fowler D (1997) The effect of nitrogen deposition and seasonal variability on methane oxidation and nitrous oxide emission rates in an upland spruce plantation and moorland. *Atmos Environ* 31:3693–3706
- Mancinelli RL (1995) The regulation of methane oxidation in soil. *Annu Rev Microbiol* 49:581–605
- Matthews E (1994) Nitrogenous fertilizers: global distribution of consumption and associated emissions of nitrous oxide and ammonia. *Global Biogeochem Cycles* 8:411–439
- Mosier AR, Schimel D, Valentine D, Bronson K, Parton W (1991) Methane and nitrous oxide fluxes in native, fertilized and cultivated grasslands. *Nature* 350:330–332
- Nedwell DB (1996) Methane production and oxidation in soils and sediments. In: Murrell JC, Kelly DP (eds) *Microbiology of atmospheric trace gases*. NATO ASI Series I, vol 39. NATO, Brussels, pp 33–49
- Nesbit SP, Breitenbeck GA (1992) A laboratory study of factors influencing CH<sub>4</sub> uptake by soils. *Agric Ecol Environ* 41:39–54
- Ojima DS, Valentine DW, Mosier AR, Parton WJ, Schimel DS (1993) Effect of land use change on methane oxidation in temperate forest and grassland soils. *Chemosphere* 26:675–685
- Prather M, Derwent R, Ehalt D, Fraser P, Sanhueza E, Zhou X (1995) Other trace gases and atmospheric chemistry. In: Houghton JT, Meiro Filho LG, Bruce J, Lee J, Callandar BA, Haites E, Harris N, Maskell K (eds) *Climate change 1994*. Cambridge University Press, Cambridge, pp 77–126
- Priemé A, Christensen S (1997) Seasonal and spatial variation of methane oxidation in a Danish spruce forest. *Soil Biol Biochem* 29:1165–1172
- Schnell S, King GM (1994) Mechanistic analysis of ammonium inhibition of atmospheric methane consumption in forest soils. *Appl Environ Microbiol* 60:3514–3521
- Schnell S, King GM (1996) Responses of methanotrophic activity in soils and cultures to water stress. *Appl Environ Microbiol* 62:3203–3209
- Schultz J (1995) *The ecozones of the world*. Springer, Berlin Heidelberg New York
- Sitaula BK, Bakken LR, Abrahamsen G (1995) CH<sub>4</sub> uptake by temperate forest soil: effect of N input and soil acidification. *Soil Biol Biochem* 27:871–880
- Stuedler PA, Bowden RD, Melillo JM, Aber JD (1989) Influence of nitrogen fertilization on methane uptake in temperate forest soils. *Nature* 341:314–316
- Tate CM, Striegl RG (1993) Methane consumption and carbon dioxide emission in tallgrass prairie: effects of biomass burning and conversion to agriculture. *Global Biogeochem Cycles* 7:735–748
- Thomas GW (1982) Exchangeable cations. In: Page AL (ed) *Methods of soil analysis – part 2. Chemical and microbiological methods*. American Society of Agronomy, Madison, Wis., pp 159–165
- Whalen SC, Cornwell JC (1985) Nitrogen, phosphorus and organic carbon cycling in an arctic lake. *Can J Fish Aquat Sci* 42:797–808
- Whalen SC, Reeburgh WS (1992) Interannual variations in tundra methane emission: a 4-year time series at fixed sites. *Global Biogeochem Cycles* 6:139–159
- Whalen SC, Reeburgh WS, Sandbeck KA (1990) Rapid methane oxidation in a landfill cover soil. *Appl Environ Microbiol* 56:3405–3411
- Whalen SC, Reeburgh WS, Kiser KS (1991) Methane consumption and emission by taiga. *Global Biogeochem Cycles* 5:261–273
- Whalen SC, Reeburgh WS, Barber VA (1992) Oxidation of methane in boreal forest soils: a comparison of seven measures. *Biogeochemistry* 16:181–211
- Willison TW, Webster CP, Goulding KWT, Powlson DS (1995) Methane oxidation in temperate soils: effects of land use and the chemical form nitrogen fertilizer. *Chemosphere* 30:539–546
- Yavitt JB, Downey DM, Lang GE, Sextone AJ (1990) Methane consumption in two temperate forest soils. *Biogeochemistry* 9:39–52