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Sven Marhan . Stefan Scheu

# Mixing of different mineral soil layers by endogeic earthworms affects carbon and nitrogen mineralization

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Abstract The effect of the endogeic earthworm species Octolasion tyrtaeum (Savigny) on decomposition of uniformly 14C-labelled lignin (lignocellulose) was studied in microcosms with upper mineral soil (Ah-horizon) from two forests on limestone, representing different stages of succession, a beech- and an ash-tree-dominated forest. Microcosms with and without lower mineral soil (Bw-horizon) were set-up; one O. tyrtaeum was added to half of them. It was hypothesised that endogeic earthworms stabilise lignin and the organic matter of the upper mineral soil by mixing with lower mineral soil of low C content. Cumulative C mineralization was increased by earthworms and by the addition of lower mineral soil. Effects of the lower mineral soil were more pronounced in the beech than in the ash forest. Cumulative mineralization of lignin was strongly increased by earthworms, but only in the beech soil (+24.6%). Earthworms predominantly colonized the upper mineral soil; mixing of the upper and lower mineral soils was low. The presence of lower mineral soil did not reduce the rates of decomposition of organic matter and lignin; however, the earthworm-mediated increase in mineralization was less pronounced in treatments with (+8.6%) than in those without  $(+14.1\%)$  lower mineral soil. These results indicate that the mixing of organic matter with C-unsaturated lower mineral soil by endogeic earthworms reduced microbial decomposition of organic matter in earthworm casts.

S. Marhan  $(\boxtimes)$ Institut für Bodenkunde und Standortslehre, Universität Hohenheim, Emil-Wolff-Strasse 27, 70593 Stuttgart, Germany e-mail: marhan@uni-hohenheim.de Tel.: +49-711-4593118 Fax: +49-711-4593117

S. Marhan . S. Scheu Institut für Zoologie, Technische Universität Darmstadt, Schnittspahnstr. 3, 64287 Darmstadt, Germany

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## Introduction

Endogeic species of earthworms are of great importance for bioturbation of soils. Ultimately, the activity of endogeic earthworms leads to the formation of mull-type soils (Müller [1950](#page-6-0); Bal [1982](#page-5-0); Scheu [1987b](#page-6-0); Bernier [1998](#page-5-0)). Mull-type soils are common in European deciduous forests stocking on non-acidic parent rock. Mull soils are characterised by a complete mixing of fragmented litter into the mineral soil by the soil macrofauna, resulting in an accumulation of soil organic matter in the mineral soil horizons (Kubiena [1948](#page-6-0)). Humus accumulation during secondary succession has been studied widely (Kögel-Knabner [1993](#page-6-0); Swift [2001](#page-6-0); Six et al. [2002](#page-6-0)). The accumulation of soil organic matter starts in the upper soil layers followed by the enrichment of the lower mineral soil in later successional stages. Lower mineral soil layers become enriched in organic matter until the inorganic matrix of the mineral soil is saturated (Hassink et al. [1997\)](#page-6-0). Although earthworms are prominent soil-forming agents, their role in stabilisation processes of soil organic matter by mixing material rich in organic matter with a C-unsaturated inorganic matrix is poorly understood.

The present study investigates the effects of earthworms on the mineralization of organic matter in the absence or presence of a lower mineral soil with low organic matter content, i.e. with a C-unsaturated inorganic matrix. By using upper mineral soils from two forest ecosystems with different stages of humus accumulation, the effects of the degree of saturation with organic matter on mineralization processes were investigated. For studying the effects of earthworms on these processes, the endogeic species *Octo*lasion tyrtaeum (Savigny) was chosen because O. tyrtaeum is one of the dominant endogeic earthworm species in the studied forests. The use of  $14C$ -labelled lignin allowed detailed analysis of C mineralization of one of the major structural carbon components of litter. Lignin has been

assumed to be an important precursor of humus materials in soils (Haider [1988;](#page-6-0) Kögel-Knabner [1993](#page-6-0)).

# Material and methods

#### Sampling

Soil samples were taken from a beech and ash forest representing two different stages of secondary succession in April 2002. The forests are located on a limestone plateau east of Göttingen at 420 m above sea level. The beech forest (Fagus sylvatica L.) is ca. 130 years old (c.f. Schaefer [1991b\)](#page-6-0) and forms the climax stage on limestone in the submontane belt of Central Europe. On the same limestone plateau an earlier stage of forest succession, represented by an ash-dominated (Fraxinus excelsior L.) forest is present. The ash forest developed from a fallow field left uncultivated for about 60 years (Scheu [1990\)](#page-6-0). The soils of the two forests are of the *Rendzina* type, shallow and containing some loess. The mineral soil of the beech forest (Ah-horizon) is rich in organic matter to the depth where it reaches the limestone bedrock, typically at 15– 30 cm. In contrast, in the ash forest, only the upper mineral soil of 5–10 cm (Ah-horizon) is rich in humus, overlaying a mineral soil layer of low organic matter content (Bwhorizon). The ash forest soil has been assumed to be in a state of humus accumulation (Thöle and Meyer [1979](#page-6-0); Scheu [1990\)](#page-6-0). Soil of the Ah-horizons of both forests was taken from the top 0- to 10-cm layer after removal of the litter material. Soil from the Bw-horizon was taken only from the ash forest from about 20 cm below the soil surface. The soils were passed through a 4-mm sieve, homogenised, and plant roots and smaller stones were removed by hand. To kill any earthworms, the soil was frozen at −28°C for 2 weeks. Data on soil C and N contents and C/N ratios of the soil materials used in the experiment are given in Table 1. Earthworms were collected by hand in the beech forest at the time when the soil samples were taken. The earthworms were stored in plastic buckets filled with soil from the beech forest at 5°C until the experiment was set-up.

Table 1 Content of C and N and C/N ratio of the soil materials used; means and SD of three replicates

	C(%)		N(%		$C/N$ ratio	
	Mean	-SD	Mean SD		Mean	SD
Beech forest						
Upper mineral soil $9.01$ $0.35$ $0.72$ $0.01$					12.54	0.34
Ash forest						
Upper mineral soil $6.60$		0.09	0.58	0.01	11.39	0.13
Lower mineral soil 1.97		0.03	0.20	0.01	9.73	0.02

#### The main experiment

Before the experiment was set-up soil samples were kept at 5°C for 1 week, and for acclimation, a further week at 20°C. Incubation was in microcosms consisting of Perspex tubes (height 150 mm, diameter 60 mm) fixed airtight on ceramic plates. Leached water was sampled in vessels placed underneath the microcosms in a box. The microcosms were closed at the top by a lid which had a small vessel attached to the underside, which can be filled with alkali to absorb  $CO<sub>2</sub>$  evolved from the soil.

The microcosms were filled with upper and/or lower mineral soil equivalent to 50 g dry wt. per layer. The following treatments were established: (1) ash forest upper mineral soil only (ash), (2) ash forest upper mineral soil on top of lower mineral soil from the ash forest (ash  $+$  min), (3) beech forest upper mineral soil only (beech) and (4) beech forest upper mineral soil on top of lower mineral soil from the ash forest (beech  $+$  min). Ten replicates were established per treatment.

After soil placement the microcosms were watered weekly for 4 weeks. Leachates from this pre-incubation period were discarded. To each microcosm 100 mg <sup>14</sup>Clabelled lignin was added and mixed homogeneously into the upper mineral soil. <sup>14</sup>C-labelled lignin (lignocellulose) material was obtained by incubating beech twigs with homogeneously <sup>14</sup>C-labelled vanillic acid as described in more detail in Scheu ([1993b\)](#page-6-0). The specific activity of the labelled material was 155.6 kBq  $g^{-1}$ . One individual O. tyrtaeum was placed into each microcosm from half of the microcosms of each treatment 1 day after the  $^{14}$ C material had been added. Before the earthworms were added they were placed on moist filter paper to void their guts for 3 days. The initial earthworm body mass was 189  $(\pm 10)$  mg fresh wt.

The microcosms were placed in a climate chamber at 20°C and incubated in darkness for 100 days. Each microcosm was irrigated with 10 ml distilled  $H_2O$  at 6-day intervals. Leached water was collected, and pooled water samples over 4 weeks were analysed for the amounts of ammonium (NH $_4^+$ –N) and nitrate (NO<sub>3</sub>–N) in the leachate. The total N leached  $(N_{\text{min}})$  during the incubation was calculated by summing the amounts of  $NH_4^+$ -N and NO<sub>3</sub>–N. Concentrations of NH<sub>4</sub><sup>+</sup> and NO<sub>3</sub> in the leachate were measured by ion-selective electrodes (ISEs) (Winlab, Windaus, Germany).

For measurement of carbon dioxide  $(CO<sub>2</sub>)$  production the microcosms were closed with rubber stoppers and incubated for 6 days. Carbon dioxide evolved during this period was trapped in 3 ml 1 N KOH and measured titrimetrically after the precipitation of carbonate with saturated  $BaCl<sub>2</sub>$  solution using 0.1 N HCl (Macfadyen [1970](#page-6-0)). An aliquot of the alkali (0.5 ml) was taken from each microcosm and mixed with 4.5 ml scintillation fluid (Luma-Safe Plus, Lumac-LSC, The Netherlands) to measure the amount of  $^{14}$ C evolved from the labelled lignocellulose. Radioactivity was measured immediately after sampling using a liquid scintillation analyser (Model 1600 TR; Packard, Meriden, USA).

At the end of the experiment earthworms were removed and individual live body masses were measured. Total soil C ( $C_{org}$ ) and N ( $N_{org}$ ) were determined by an elemental analyser (Model 1400; Carlo Erba, Milan, Italy).

## Statistical analyses

Data on cumulative  $CO_2$  production,  ${}^{14}CO_2$ –C production and mineral N  $(N_{min})$  leaching after 100 days of incubation were analysed by a three-factor analysis of variance (ANOVA). Factors were earthworms (without and with O. tyrtaeum), type of upper mineral soil (beech and ash) and lower mineral soil (with and without lower mineral soil of the ash forest). Data on the water content were analysed separately for treatments with and without lower mineral soil by a two-factor ANOVA. Changes in earthworm live body masses were analysed by two-factor analysis of covariance (ANCOVA) using the initial earthworm live body masses as covariate. Data were inspected for homogeneity of variance (Levene test) and log-transformed if required. Tukey's honestly significant difference (HSD) test was used for comparison of means. Statistical analysis was done using the STATISTICA 6.0 software package (Statsoft, Hamburg, Germany).

# **Results**

#### Earthworm burrowing

By the end of the experiment the upper soil layers had been almost entirely transformed into casts by O. tyrtaeum. In contrast, only 20–40% of the lower soil layer was processed, and only a few earthworm burrows crossed this layer. The water content of the upper soil was generally higher in the beech treatments without *O. tyrtaeum* (43.6%) dry wt.) than in the respective ash treatments (34.4%;  $F_{1,16}=2,764.17, P<0.001$ ). Earthworms increased the soil water content in both upper soils to 45.2 and 36.6% for beech and ash treatments, respectively  $(F_{1,16}=127.87)$ ,  $P<0.001$ ). In treatments with lower mineral soil the water content also differed between beech and ash soil  $(F_{1,14}$ = 1,692.60,  $P<0.001$ ) and was increased by earthworms  $(F_{1,14}=158.16, P<0.001)$ . Soil water content in the beech + min (35.7%) was higher than in the ash  $+$  min treatment  $(29.9\%)$ .

## Earthworm body mass

All individuals of *O. tyrtaeum* survived until the end of the experiment and increased in body mass. However, the increases in the ash  $(89.2 \pm 20.9\%)$  and ash + min treatments (81.0±33.4%) were significantly higher than in beech (22.3 $\pm$ 16.3%) and beech + min treatments (9.8 $\pm$ 10.7%;  $F_{1,15}$ =56.59, P<0.001 for the effect of type of upper mineral soil).

Nitrogen leaching

Leaching of mineral  $N(N_{min})$  decreased during the course of the experiment. In particular the amounts of N leached as  $NO<sub>3</sub>$  strongly decreased during the first 65 days, whereas the amounts of  $NH_4^+$ -N in the leachate remained more constant (data not shown). O. tyrtaeum increased significantly the amounts of NH<sub>4</sub><sup>--</sup>N ( $F_{1,30}$ =32.4, P<0.001) and NO<sub>3</sub>N ( $F_{1,30}$ =13.1,  $P<0.01$ ) leached during the experiment on average by 9.8 and 14.8%, respectively, but the effects of earthworms accounted for only 0.7 and 5.3% of the total variation.

The amount of  $N_{\text{min}}$  leached relative to the total amount of soil N in the microcosms (mg  $N_{\text{min}} g^{-1}$  N) was not affected by "lower mineral soil" in treatments without earthworms. This indicated that N bound in the lower mineral soil of the ash forest was mobilised to a similar extent than that bound in upper mineral soils.

## $CO<sub>2</sub>$  production

Rates of  $CO<sub>2</sub>$  production in treatments without earthworms decreased throughout the experiment (Fig. [1](#page-3-0)a) except in ash forest soil treatments where they increased initially. Carbon dioxide production in soils from earthworm treatments generally exceeded those of the control soils (Fig. [1](#page-3-0)b). The effects of *O. tyrtaeum* on  $CO<sub>2</sub>$  production increased early in the experiment, reaching a maximum by day 20. Then, rates of  $CO<sub>2</sub>$  production decreased, but the decrease was faster in ash than in beech treatments.

Cumulative  $CO<sub>2</sub>$  production during the experiment ranged between 160 and 196 mg  $CO<sub>2</sub>–C$  microcosm<sup>-1</sup> (Fig. [2a](#page-3-0)). Each of the factors studied affected total  $CO<sub>2</sub>$ production significantly. Overall, earthworms increased total CO<sub>2</sub> production by 9.1%  $(F_{1,32} = 121.4, P < 0.001)$ , which accounted for 48.8% of the variation. The earthworm-mediated increases differed between treatments with and without lower mineral soil (earthworm  $\times$  lower mineral soil interaction;  $F_{1,32}$ =4.3,  $P<0.05$ ), i.e. the earthwormmediated increase in  $CO<sub>2</sub>$  production was less pronounced in ash (10.0%) and ash + min (5.0%) than in beech (12.5%) and beech  $+$  min (9.1%). The presence of lower mineral soil also increased  $CO<sub>2</sub>$  production ( $F<sub>1,32</sub>=57.8$ ,  $P<0.001$ ) but only in the beech treatments  $(+10.8\%)$ ; the increase in the ash treatments was negligible  $(+1.8\%)$ . The interactions between type of upper mineral soil and lower mineral soil were highly significant  $(F_{1,32}=28.4, P<0.001)$ and accounted for an additional 11.4% of the variation.

Mineralization of  $^{14}$ C-labelled lignin

The amount of  ${}^{14}C$  in leaching water was below detection limit, suggesting that leaching of lignin-derived C as dissolved organic carbon (DOC) was negligible. Changes in mineralization rates of lignin differed between beech and ash treatments but were little affected by the presence of lower mineral soil. Mineralization rates increased strongly

<span id="page-3-0"></span>Fig. 1 Effects of type of upper mineral soil (ash and beech forest) and lower mineral soil (without,  $-Min$  and with,  $+Min$ ) on a rates of C mineralisation (mg  $CO<sub>2</sub>$ –C per day and microcosm) in treatments without earthworms, and b the effect of earthworms on C mineralization plotted relative to respective treatments without earthworms. Means of five replicates with 1 SD



during the first 15–20 days of incubation, reaching 0.25 and 0.17% of initial  ${}^{14}$ C day<sup>-1</sup> in the ash and beech treatments without earthworms, respectively (Fig. [3](#page-4-0)a). Thereafter, mineralization rates decreased slowly but continuously in beech treatments to a level of about  $0.13\%$  of initial  $14$ C  $day^{-1}$ . In contrast, in ash treatments, <sup>14</sup>CO<sub>2</sub>–C production



Fig. 2 Effects of type of upper mineral soil (ash and beech forest), lower mineral soil (without, –*Min* and with, +*Min*) and earthworms (without,  $-EW$  and with,  $+EW$ ) on cumulative C mineralization (mg  $CO<sub>2</sub>$ –C per microcosm) during 100 days of incubation. Bars sharing the same letter are not significantly different (Tukey's HSD test,  $P<0.05$ )

decreased sharply for 12 days and then increased again, reaching a second maximum of 0.24% of initial <sup>14</sup>C dav<sup>-1</sup> on day 49. After this mineralization rates declined slowly to a level similar to that in beech treatments.

Octolasion tyrtaeum strongly increased the rates of mineralization of lignin for 42 and 82 days in the ash and beech treatments, respectively (Fig. [3](#page-4-0)b). Later, mineralization rates in the earthworm treatments were lower than in the respective treatments without earthworms at the end of the experiment by ca. 20% in both the ash and beech treatments.

In the ash treatments a total of 8.4% of the initial lignin C was mineralized during the experiment. In the beech treatments O. tyrtaeum increased cumulative lignin mineralization (from 6.6 to 8.3% of initial). Earthworms significantly  $(F_{1,32} = 98.1, P < 0.001)$  increased lignin mineralization on average by 11.3%, accounting for 27.5% of the total variation. The type of upper soil accounted for 39.8% of the total variation  $(F_{1,32}=144.3, P<0.001)$ . The effects of the earthworms were much more pronounced in beech treatments (by 24.6%). The earthworm  $\times$  type of upper mineral soil interaction was highly significant  $(F_{1,32}$ = 80.9, P<0.001) and accounted for an addition of 22.3% of the variation. The presence of lower mineral soil affected the cumulative mineralization of lignin, but the effect depended on the presence of earthworms ( $F_{1,32}$ =5.5, P<0.05;

<span id="page-4-0"></span>Fig. 3 Effects of type of upper mineral soil (ash and beech forest) and lower mineral soil (without,  $-Min$  and with,  $+Min$ ) on a mineralization rates of  ${}^{14}C$ labelled lignin (% of initial per day) in treatments without earthworms and b mineralization of  ${}^{14}$ C-labelled lignin in treatments with earthworms plotted relative to respective treatments without earthworms. Means of five replicates with 1 SD



accounting for an addition of 1.5% of the total variation). The increases in mineralization caused by O. tyrtaeum were less pronounced in the treatments with lower mineral soil  $(+8.6\%)$  than in those without  $(+14.1\%)$ .

Mineralization of lignin relative to total  $CO<sub>2</sub>$  production was on average 11.9% less in the upper mineral soil from the beech than from the ash forest  $(F_{1,32}=116.8, P<0.001)$ . The presence of lower mineral soil decreased the relative  $14$ C mineralization by 3.7% in the ash and 10.3% in the beech treatments (significant type of upper mineral soil  $\times$ lower mineral soil interaction;  $F_{1,32}$ =7.1, P<0.05). The effects of earthworms on the relative mineralization of lignin differed between ash and beech treatments (significant earthworm  $\times$  type of upper mineral soil interaction;  $F_{1,32}$ =57.5, P<0.001). In the ash treatments, earthworms reduced relative mineralization rates (−6.1%), whereas in the beech treatments, rates were increased (+12.5%).

# **Discussion**

Forest soils often contain large amounts of soil organic matter which is formed predominantly by the accumulation

of stable humus substances such as lignin (Schaefer [1991a](#page-6-0); Joergensen [1991](#page-6-0)). The potential of the soil matrix to bind organic material such as lignin is important for the accumulation and stabilisation of soil organic matter (Six et al. [2002\)](#page-6-0). In the present experiment, two forest soils were compared, representing different stages of humus accumulation. The total mineralization of lignin-derived C was 1.3 times higher in treatments containing ash forest soil, which is supposed to be in a state of humus accumulation, than in those containing humus-rich soil from a matured beech forest. Therefore, cumulative  $CO<sub>2</sub>$  production from the ash soil slightly exceeded that from the beech soil. Microorganisms are predominantly responsible for the degradation and mineralization of soil organic matter (Swift et al. [1979](#page-6-0)). It is known that few soil micro-organisms are able to attack the lignin molecule (Crawford [1981\)](#page-5-0), and lignin degradation was shown to be uncorrelated with microbial biomass (Entry et al. [1987](#page-6-0)). Therefore, the extent of lignin mineralization depends strongly on the composition of the soil microflora. This suggests that the microbial community of the upper mineral soil from the ash forest contained more micro-organisms that were able to mineralize lignin than that from the beech forest. However, lignin-miner<span id="page-5-0"></span>alizing micro-organisms are also affected by resource availability, and it has been shown that lignin mineralization is increased if easily degradable C resources such as carbohydrates are available (Reid [1979;](#page-6-0) Kirk and Farrell [1987](#page-6-0)). Presumably, the higher contents of easily available C resources in the upper mineral soil in the ash forest soil contributed to the higher rates of lignin decomposition than in the beech forest soil. The content of easily available and degradable C resources was not measured in the present experiment. However, the more pronounced increase in body mass of earthworms in the ash than in beech soils suggests that the content of easily available C resources in the ash soil exceeded those in the beech soil. The diet of endogeic earthworms depends substantially on easily available C resources such as glucose (Tiunov and Scheu [2004\)](#page-6-0).

Octolasion tyrtaeum strongly increased lignin mineralization during the first 6 weeks of incubation in the ash forest soil, whereas in the beech forest soil, the increase lasted almost twice as long. The enzyme system of earthworms is unable to degrade the lignin molecule (Neuhauser et al. [1978](#page-6-0)). Therefore, the different mineralization patterns were likely caused by changes in the microbial community structure (Scheu [1993a\)](#page-6-0). Microbial growth is often limited by the availability of nutrients, mainly by N and/or P (Anderson and Domsch 1978; Joergensen and Scheu [1999](#page-6-0)). Microbial growth in the ash forest soil has been shown to be limited by C and P, whereas in the beech soil, growth is primarily limited by P (Scheu [1987a](#page-6-0), [1990\)](#page-6-0). Lignin mineralization also is known to be limited by the availability of P (Reid [1979\)](#page-6-0). Endogeic earthworms mobilise P as demonstrated by increased amounts of phosphate in earthworm casts (Graff [1971](#page-6-0); Aldag and Graff 1975; Haynes et al. [1999\)](#page-6-0). Therefore, the earthworm-mediated increase in mineralization of soil organic matter and lignin in the beech forest soil may have been due to an increased availability of P. Similarly, the initial stimulation of lignin mineralization in the ash forest soil with earthworms was likely caused by an increased availability of nutrients. However, increased C availability may also have contributed to the initially increased mineralization of lignin in the presence of earthworms. Substantial amounts of C are deposited by the earthworms as mucus, which is mainly concentrated in the burrow linings and in casts, providing easily available C resources for micro-organisms (Scheu [1991](#page-6-0); Schmidt et al. [1999\)](#page-6-0).

Reduced lignin mineralization rates in the later stages of incubation in the earthworm treatments may have been caused by reduced P and C availability. The steepest decline occurred in ash forest soil. Presumably, the nutrients necessary for lignin decomposition became limited due to leaching and microbial immobilisation; leaching of mineral N decreased later in the experiment. In addition, O. tyrtaeum may have efficiently exploited easily available carbon resources in soil and competed with micro-organisms for carbon, thereby reducing lignin mineralization.

The incorporation of soil organic matter into cast aggregates is assumed to reduce rates of C mineralization (Lee [1985](#page-6-0); Edwards and Bohlen 1996). The incorporated organic particles were shown to become encrusted and

serve as nuclei for the formation of stable aggregates during the earthworm gut passage (Shipitalo and Protz [1989](#page-6-0)). Lignin particles enclosed in cast aggregates are protected from microbial attack due to physical separation (Guggenberger et al. [1995](#page-6-0)). Access of micro-organisms to nutrients inside cast aggregates is reduced, and this is assumed to limit the decomposition of soil organic matter (Wolters [2000\)](#page-6-0). This may explain the reduced lignin mineralization rates in treatments with upper mineral soil later in the experiment.

The lower mineral soil also affected rates of lignin mineralization; the increase in cumulative lignin mineralization in the presence of O. tyrtaeum was less pronounced when lower mineral soil was present. The close association of organic particles with a C-unsaturated inorganic matrix was shown to increase the stabilisation of organic matter (Six et al. [2002](#page-6-0)). Carbon accumulation in the mineral-associated fraction of organic matter is suggested to be caused by a rapid association of organic matter with particles or colloids, resulting in the formation of a physically protected C pool (Jastrow [1996\)](#page-6-0). Lignin molecules in the present study may have been stabilised and protected against microbial degradation by close association with the C-unsaturated lower mineral soil matrix due to intimate mixing during passage through the earthworm gut. In the long term, the pool of stabilised C presumably increases until the C-unsaturated matrix becomes saturated. In natural systems such as forests, the formation of mull soils rich in organic matter is fostered by the earthwormmediated incorporation of lower mineral soil into the upper mineral soil horizon rich in humus (Ah). However, O. tyrtaeum burrowed very little into the lower mineral soil layer, suggesting that lower mineral soil was an unfavourable habitat for endogeic earthworms, which is probably related to low food quality. Therefore, although earthworms may turn over large quantities of humus-rich upper mineral soil, the formation of mull soils by the mixing of plant residues, upper and lower mineral soil by endogeic earthworms probably takes decades.

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#### References

- Aldag R, Graff O (1975) N-Fraktionen in Regenwurmlosung und deren Ursprungsboden. Pedobiologia 15:151–153
- Anderson JPE, Domsch KH (1978) A physiological method for the quantitative measurement of microbial biomass in soils. Soil Biol Biochem 10:215–221
- Bal L (1982) Zoological ripening of soils. Centre for Agricultural Publishing and Documents, Wageningen
- Bernier N (1998) Earthworm feeding activity and development of the humus profile. Biol Fertil Soils 26:215–223
- Crawford RL (1981) Lignin biodegradation and transformation. Wiley, New York
- Edwards CA, Bohlen P (1996) Biology and ecology of earthworms. Chapman and Hall, New York
- <span id="page-6-0"></span>Entry JA, Stark NM, Lowenstein H (1987) Timber harvesting: effects on degradation of cellulose and lignin. For Ecol Manag 22:79–88
- Graff O (1971) Stickstoff, Phosphor und Kalium in der Regenwurmlosung auf der Wiesenversuchsfläche des Sollingprojektes. Ann Zool 4:503–512
- Guggenberger G, Zech W, Thomas RJ (1995) Lignin and carbohydrate alteration in particle-size separates of an oxisol under tropical pastures following native savanna. Soil Biol Biochem 27:1629–1638
- Haider K (1988) Der mikrobielle Abbau des Lignins. Forum Mikrobiol 11:477–482
- Hassink J, Whitmore AP, Kubat J (1997) Size and density fractionation of soil organic matter and the physical capacity of soils to protect organic matter. Eur J Agron 7:189–199
- Haynes RJ, Fraser PM, Tregurtha RJ, Piercy JE (1999) Size and activity of the microbial biomass and N, S and P availability in earthworm casts derived from arable and pastoral soil and arable soil amended with plant residues. Pedobiologia 43:568– 573
- Jastrow JD (1996) Soil aggregate formation and the accrual of particulate and mineral-associated organic matter. Soil Biol Biochem 28:665–676
- Joergensen RG (1991) Organic matter and nutrient dynamics of the litter layer on a forest Rendzina under beech. Biol Fertil Soils 11:163–169
- Joergensen RG, Scheu S (1999) Response of soil microorganisms to the addition of carbon, nitrogen and phosphorus in a forest Rendzina. Soil Biol Biochem 31:859–866
- Kirk TK, Farrell RL (1987) Enzymatic "combustion": the microbial degradation of lignin. Annu Rev Microbiol 41:465–505
- Kögel-Knabner I (1993) Biodegradation and humification processes in forest soil. In: Bollag J-M, Stotzky G (eds) Soil Biochemistry, vol 8. Dekker, New York, pp 105–135
- Kubiena WL (1948) Entwicklungslehre des Bodens. Springer, Wien Lee KE (1985) Earthworms—their ecology and relationships with
- soils and land use. Academic, Sydney Macfadyen A (1970) Simple methods for measuring and maintaining the proportion of carbon dioxide in air, for use in ecological studies of soil respiration. Soil Biol Biochem 2:9–18
- Müller PE (1950) Forest soil studies, a contribution to silvicultural theory. III. On compacted ground deficient in mull, especially in beech forest. Dan Skovforen Tidsskr 1:10–619
- Neuhauser EF, Hartenstein R, Connors J (1978) The role of soil macroinvertebrates in the degradation of vanillin, cinnamic acid, and lignins. Soil Biol Biochem 10:431–435
- Reid ID (1979) The influence of nutrient balance on lignin degradation by the white-rot fungus Phanerochaete chrysosporium. Can J Bot 57:2050–2058
- Schaefer M (1991a) Ecosystem processes: secondary production and decomposition. In: Röhrig E, Ulrich B (eds) Temperate deciduous forests. Ecosystems of the world. Elsevier, Amsterdam, pp 175–218
- Schaefer M (1991b) Animals in European temperate deciduous forest. In: Röhrig E, Ulrich B (eds) Temperate deciduous forests. Ecosystems of the world. Elsevier, Amsterdam, pp 503–525
- Scheu S (1987a) Microbial activity and nutrient dynamics in earthworm casts (Lumbricidae). Biol Fertil Soils 5:230–234
- Scheu S (1987b) The role of substrate feeding earthworms (Lumbricidae) for bioturbation in a beechwood soil. Oecologia 72: 192–196
- Scheu S (1990) Changes in the microbial nutrient status during secondary succession and its modification by earthworms. Oecologia 84:351–358
- Scheu S (1991) Mucus excretion and carbon turnover of endogeic earthworms. Biol Fertil Soils 12:217–220
- Scheu S (1993a) Litter microflora–soil macrofauna interactions in lignin decomposition: a laboratory experiment with 14-Clabelled lignin. Soil Biol Biochem 25:1703–1711
- Scheu S (1993b) Cellulose and lignin decomposition in soils from different ecosystems on limestone as affected by earthworm processing. Pedobiologia 37:167–177
- Schmidt O, Scrimgeour CM, Curry JP (1999) Carbon and nitrogen stable isotope ratios in body tissue and mucus of feeding and fasting earthworms (Lumbricus festivus). Oecologia 118:9–15
- Shipitalo MJ, Protz R (1989) Chemistry and micromorphology of aggregation in earthworm casts. Geoderma 45:357–374
- Six J, Conant, RT, Paul EA, Paustian K (2002) Stabilization mechanisms of soil organic matter: implications for C-saturation of soils. Plant Soil 241:155–176
- Swift RS (2001) Sequestration of carbon by soil. Soil Sci 166:858– 871
- Swift MJ, Heal OW, Anderson JM (1979) Decomposition in terrestrial ecosystems. Blackwell, Oxford
- Thöle R, Meyer B (1979) Bodengenetische und-ökologische Analyse eines repräsentativen Areals der Göttinger Muschelkalk-Schale als landschaftsökologische Planungsgrundlage. Gött Bodenkdl Ber 59:230
- Tiunov AV, Scheu S (2004) Carbon availability controls the growth of detritivores (Lumbricidae) and their effect on nitrogen mineralization. Oecologia 138:83–90
- Wolters V (2000) Invertebrate control of soil organic matter stability. Biol Fertil Soils 31:1–19