



Body mass and cuticular hydrocarbon profiles, but not queen number, underlie worker desiccation resistance in a facultatively polygynous harvester ant (*Pogonomyrmex californicus*)

Madeleine M. Ostwald¹ · Sandra Tretter² · Jan Buellesbach² · Juliana M. Calixto¹ · Jennifer H. Fewell¹ · Jürgen Gadau^{1,2} · Kaitlin M. Baudier³

Received: 19 September 2022 / Revised: 17 March 2023 / Accepted: 14 April 2023 / Published online: 29 April 2023
© The Author(s), under exclusive licence to Springer-Verlag GmbH Germany, part of Springer Nature 2023

Abstract

As small-bodied terrestrial organisms, insects face severe desiccation risks in arid environments, and these risks are increasing under climate change. Here, we investigate the physiological, chemical, and behavioral mechanisms by which harvester ants, one of the most abundant arid-adapted insect groups, cope with desiccating environmental conditions. We aimed to understand how body size, cuticular hydrocarbon profiles, and queen number impact worker desiccation resistance in the facultatively polygynous harvester ant, *Pogonomyrmex californicus*. We measured survival at 0% humidity of field-collected worker ants sourced from three closely situated populations within a semi-arid region of southern California. These populations vary in queen number, with one population dominated by multi-queen colonies (primary polygyny), one population dominated by single-queen colonies, and one containing an even mix of single- and multi-queen colonies. We found no effect of population on worker survival in desiccation assays, suggesting that queen number does not influence colony desiccation resistance. Across populations, however, body mass and cuticular hydrocarbon profiles significantly predicted desiccation resistance. Larger-bodied workers survived longer in desiccation assays, emphasizing the importance of reduced surface area-to-volume ratios in maintaining water balance. Additionally, we observed a positive relationship between desiccation resistance and the abundance of *n*-alkanes, supporting previous work that has linked these high-melting point compounds to improved body water conservation. Together, these results contribute to an emerging model explaining the physiological mechanisms of desiccation resistance in insects.

Keywords Pleometrosis · Water balance · *n*-alkanes · Gas chromatography

Introduction

Maintaining water balance is a major homeostatic challenge shared by all organisms. As small-bodied terrestrial organisms, insects face particular water balance challenges due to their larger body surface area to volume ratios (Hadley

1994; Kühnel et al. 2017). Despite the critical ecological value of insects in terrestrial ecosystems, the mechanisms by which they resist desiccation remain underexplored. As a complex functional trait, desiccation resistance is shaped by interactions between phylogeny, environment, and life history traits (Chung and Carroll 2015; Hadley 1994; Sprenger et al. 2018; Wang et al. 2021). Here, we experimentally link major physiological and colony characteristics of harvester ants, a common arid-adapted insect, to their survival outcomes under low humidity conditions, with the aim of disentangling the physiochemical and behavioral mechanisms governing insect desiccation resistance.

The capacity to withstand desiccating conditions can vary widely among and within species (Bujan et al. 2016; Hadley 1994; Hoffmann and Harshman 1999). Animals living in arid environments, in particular, may reduce water loss through behavioral and/or morphological adaptations, for example,

Communicated by P. Withers.

✉ Madeleine M. Ostwald
ostwald.madeleine@gmail.com

- ¹ School of Life Sciences, Arizona State University, Tempe, AZ, USA
- ² Institute for Evolution and Biodiversity, University of Münster, Münster, Germany
- ³ School of Biological, Environmental, and Earth Sciences, University of Southern Mississippi, Hattiesburg, MS, USA

by restricting activity to the coolest times of day (Schultheiss and Nooten 2013), or by exploiting body morphologies that reduce their surface area-to-volume ratio (Hood and Tschinkel 1990; Prange and Pinshow 1994). Larger-bodied organisms not only have a reduced surface area, they also may have proportionally larger water reserves, additionally buffering against desiccation (Lighton et al. 1994; Prange and Pinshow 1994). Positive relationships between body size and desiccation resistance have been demonstrated across insect taxa (Chown and Klok 2003; Fouet et al. 2012; Kærsgaard et al. 2004; Le Lagadec et al. 1998) suggesting potential selective pressures for increased body size in insects experiencing water conservation challenges.

Other physiological adaptations to resist desiccation are determined by the chemical composition of the insects' epicuticle, which consists predominantly of structurally diverse lipids known as cuticular hydrocarbons (CHCs). This waxy barrier serves as the primary interface through which insects exchange water with the environment (Blomquist and Bagnères 2010). The chemical composition of CHC profiles can vary considerably among species and populations, as do their waterproofing capabilities (Blomquist and Bagnères 2010; Sprenger and Menzel 2020). CHCs generally protect against water loss below a certain critical temperature, which is determined by the chemical structure of individual CHC compounds (Gibbs 1998, 2002). Generally, molecules with longer chain lengths, fewer methyl groups, and fewer double bonds have higher melting points (Gibbs and Pomonis 1995). Several studies have associated particular CHC signatures with desiccation resistance (especially linear alkanes; Chung 2014; Chung and Carroll 2015; Ferveur et al. 2018; Whyte et al. unpub.) or with adaptation to arid environments (Buellesbach et al. 2018; Menzel et al. 2018; Sprenger et al. 2018), though these patterns appear to vary considerably across taxa.

Seed harvester ants represent the numerically dominant animals in arid ecosystems globally (Brown et al. 1979; La Richelière et al. 2022), and thus hold particular promise for illuminating the adaptive and physiological mechanisms underlying desiccation resistance. One such seed harvester, *Pogonomyrmex californicus* (Formicidae: Myrmicinae), is abundant in arid to semi-arid environments including deserts, grasslands, and scrublands of southwestern North America. This species has served as a valuable model for studying the evolution of cooperation due to its intraspecific variation in queen social strategy (i.e., cooperative/primary polygynous vs. non-cooperative/monogynous); (Cahan and Fewell 2004; Clark and Fewell 2014; Haney and Fewell 2018; Overson et al. 2014, 2016; Shaffer et al. 2016). Polygyny has the potential to interact with worker and colony-level functional traits (Greenberg et al. 1985; Haney and Fewell 2018; Hölldobler and Wilson 1990; Pisarski 1981), potentially by increasing worker genetic diversity

or by mediating offspring quality, though this relationship has rarely been examined for physiological traits. A recent study showed that multi-queen colonies of *P. californicus* produced workers with higher and more uniform heat tolerance (Baudier et al. 2022), raising questions about implications for desiccation resistance, which interacts with thermal tolerance in complex and variable ways not yet fully elucidated (Bujan et al. 2016; Klok et al. 2006; Renault et al. 2005). Importantly, the relationship between nest founding strategy (polygynous vs. monogynous) and worker desiccation resistance has not been previously studied. Despite the prominence of this model system, the chemical composition of *P. californicus*' CHC profile has also not yet been formally described. Likewise, despite suggestions in other harvester ants that water may be limiting during colony founding (Johnson 2000, 2021), little is known of the chemical mechanisms workers employ to resist desiccation.

In this study, we use *P. californicus* as a model for investigating the physiological, chemical, and behavioral drivers of desiccation resistance. We assay survival under desiccation stress and characterize chemical profiles of worker ants sampled simultaneously across distinct populations that vary in queen number (monogynous vs. polygynous) within a small geographic area with homogenous environmental conditions (Overson et al. 2016). In doing so, we examine effects of queen number on colony desiccation resistance while controlling for major seasonal and habitat effects. Across this variation, we highlight physiological and chemical correlates of desiccation resistance with the aim of clarifying the integrated set of mechanisms these animals use to cope with water balance challenges.

Methods

Study populations and field collections

To assess desiccation resistance in *P. californicus* workers, we sampled worker ants from three sites in San Diego County, California: at Lake Henshaw (33.23176°N, – 116.7553°W, elevation: 825 m), Pine Valley, CA (32.82342°N, – 116.5278°W, elevation: 1130 m), and Cuyamaca Rancho State Park, hereafter “Cuyamaca” (32.93798°N, – 116.5625°W, elevation: 1295 m). The three sites span a ~46 km range of *P. californicus* populations that vary in queen number, from predominantly single-queen colonies at Lake Henshaw to predominantly multi-queen colonies at Pine Valley (Overson et al. 2016). The Cuyamaca site is situated geographically between these two regions and contains a relatively even mix of monogynous and polygynous colonies (Haney et al. 2017). We did not quantify queen number in colonies of this site, which can range from one to eight in mature colonies, and which could impact worker genetic diversity (beyond the

effects of polyandry, as queens are multiply mated, Overson et al. 2016). However, queen number in polygynous populations is both relatively uniform (mean = 4.1; std. error = 0.66) and temporally stable (Haney and Fewell 2018).

We collected workers for desiccation assays and chemical analyses from a total of 30 colonies across the three sites. For desiccation assays, we collected foraging workers from 10 colonies at each site on the mornings of June 22 and 23, 2021. Workers were immediately placed in a high humidity chamber (a 15 mL tube containing 5 mL water separated by a cotton ball, from which ants could drink ad libitum) with only their nestmates. For chemical analysis, we additionally collected foraging workers from each of the same 30 colonies across the three sites between June 20 and 28, 2021. Workers were collected into glass vials and freeze-killed within 2 h of collection then stored at $-20\text{ }^{\circ}\text{C}$ for later analysis.

Desiccation resistance assays

To standardize the initial hydration level of individual ants, we kept all field-collected workers in tubes with ad libitum water for approximately 5 h following collection (described above). We then marked all workers for individual identification with a single, small mark on the gaster using oil-based paint pens (Sharpie®, Newell Brands, Atlanta, GA, USA). Because the paint mark covered only a small proportion of total body surface area, we do not expect it to meaningfully impact desiccation resistance. We assayed ten workers from each colony, for all ten colonies from each of the three sites, for a total treatment sample size of 300 workers.

Immediately after marking, ants were placed in desiccation treatment chambers consisting of 15 mL tubes containing 5 mL calcium sulfate (Drierite, W.A. Hammond Drierite Co Ltd, Xenia, OH, USA). Ants were placed in a 5 mL space in the uppermost portion of the tube, separated from the Drierite by two cotton balls, between which was an additional layer of air (as in Whyte et al. unpub.). Each desiccation chamber held five nestmate workers, such that we had two treatment chambers per colony to hold 10 workers. We chose to maintain workers in groups to mitigate the confounding negative effects of social isolation on ant survival and energy consumption (Boulay et al. 1999; Koto et al. 2015). To control for other possible causes of death over the experimental period, we additionally established a high-humidity control in the same design as our desiccation chambers, but replacing Drierite with water. We used Hygrochron humidity-logging iButtons® (DS1923, Maxim Integrated, San Jose, CA, USA) to confirm that desiccation chambers consistently provided 0% relative humidity conditions.

We visually inspected all treatment and control workers for survival every three hours from the start of the experiment until the death of the final surviving treatment ant. We

defined the time of death as the point at which an individual ant did not move in response to agitation of the chamber. Ants from all treatment were then dried in a drying oven at $55\text{ }^{\circ}\text{C}$ for 3 days and weighed with a microbalance (AandD GR-200; repeatability 0.0001 g) to obtain dry mass as a proxy for body size and therefore surface area to volume ratio (Kühnel et al. 2017).

Chemical analysis

We extracted chemical surface profiles from 150 freeze-killed, field-collected ants from 30 colonies (5 workers from 10 colonies at each of 3 sites). We added 100 μL HPLC-grade hexane to 2 mL HPLC screwcap vials (Agilent Technologies, Waldbronn, Germany) containing single ants, and extracted their surface profiles for 10 min on an orbital shaker at 240 revolutions/min (IKA KS 130 Basic, Staufen, Germany). We then transferred extracts to a conical 250 μL GC insert (Agilent Technologies, Waldbronn, Germany) and evaporated the solvent completely under a flow of CO_2 . We then resuspended the dried extracts in 10 μL of a hexane solution containing 7.5 ng/ μL dodecane (C12) as an internal standard.

To identify and quantify chemical compounds from the surface profiles, we used a gas chromatograph (GC: 7890B) coupled to a flame ionization detector (FID: G3440B) and a tandem mass spectrometer (MS/MS: 7010B, all from Agilent Technologies, Waldbronn, Germany). The system was equipped with a fused silica column (DB-5MS ultra inert; 30 m \times 250 μm \times 0.25 μm ; Agilent JandW columns, Santa Clara, CA, USA) at a temperature of $300\text{ }^{\circ}\text{C}$ with helium used as a carrier gas under a constant flow of 1.8 mL/min. The FID had a temperature of $300\text{ }^{\circ}\text{C}$ and used nitrogen with a 20 mL/min flow rate as make-up gas and hydrogen with a 30 mL/min flow rate as fuel gas. The column was split at an auxiliary electronic pressure control (Aux EPC) module into an additional deactivated fused silica column piece (0.9 m \times 250 μm \times 0.25 μm) with a flow rate of 0.8 mL/min leading into the FID detector, and another deactivated fused silica column piece (1.33 m \times 250 μm \times 0.25 μm) at a flow rate of 1.33 mL/min into the mass spectrometer. An initial column temperature of $60\text{ }^{\circ}\text{C}$ was maintained for 5 min and then gradually increased ($20\text{ }^{\circ}\text{C}/\text{min}$ to $240\text{ }^{\circ}\text{C}$, $1\text{ }^{\circ}\text{C}/\text{min}$ to $294\text{ }^{\circ}\text{C}$, $4\text{ }^{\circ}\text{C}/\text{min}$ to $325\text{ }^{\circ}\text{C}$) to a final temperature of $325\text{ }^{\circ}\text{C}$ held for 2 min.

We used the resulting chromatograms to identify cuticular hydrocarbon (CHC) compounds from their mass spectra, diagnostic ions, retention indices using the program Agilent MassHunter Workstation Software (Version B.09.00/Build 9.0.647.0, Agilent Technologies, Santa Clara, CA, USA). We included only compounds identified as CHCs in our analysis. Absolute CHC quantities (in ng) were obtained by calibrating each compound according to a dilution series at

four concentrations (5 ng/ μ l, 10 ng/ μ l, 20 ng/ μ l, 30 ng/ μ l) based on the closest eluting *n*-alkane from a C21-40 standard series (Merck, KGaA, Darmstadt, Germany). We excluded five of 150 chromatograms from analysis due to apparent extraction failure, leaving a final sample size of 145 workers (Lake Henshaw $n = 49$; Cuyamaca $n = 49$; Pine Valley $n = 47$).

Environmental conditions at sampling sites

The three sampling sites are situated in chaparral-dominated, lower montane shrub and woodland ecoregions in southern California. We characterized the environmental conditions at these three sampling sites using publicly available bioclimatic data (WorldClim2, (Fick and Hijmans 2017)). We analyzed average monthly climatic patterns over a 30-year time period (1970–2000) and found that the three sites do not differ significantly in annual precipitation (Kruskal–Wallis test: $P = 0.388$), water vapor pressure ($P = 0.320$), or temperature ($P = 0.615$); (Supplementary Fig. 1; Supplementary Table 1). This confirmed our assumption that the three sites did not substantially differ in climatic conditions related to desiccation.

Statistical analysis

To understand predictors of survival in the desiccation resistance assays, we performed a nonparametric mixed effect Cox regression (“coxme” and “survival” packages; (Therneau 2020, 2022)) with survival time in desiccation assays as our response variable, population and log dry mass as our predictor variables, and colony as a random effect. We log-transformed the dry mass data to improve normality. We calculated lethal time (LT₅₀) as the median time to death. We used a Kruskal–Wallis test to compare colony-level standard deviation in survival time across populations. Colony-level standard deviation was the standard deviation of the survival time values across all workers in each colony. We then used post-hoc pairwise Wilcoxon tests with a Bonferroni correction for multiple comparisons.

To visualize CHC data, we used a principal component analysis (PCA) of absolute CHC abundances with centered log-ratio transformation using the R package “vegan” (Oksanen et al. 2020). To assess the relationship between desiccation resistance and CHC profiles, we used a linear model with mean colony LT₅₀ as our response variable and with the log abundance of linear *n*-alkanes and the log abundance of methyl-branched alkanes (the two major structural classes of compounds we found in the CHC profiles) as predictor variables. Because the desiccation assays are intrinsically stressful (and ultimately lethal), and likely to impact workers’ CHC composition (Menzel et al. 2018), we did not extract CHCs from the same individual workers that had undergone

desiccation assays. Instead, we extracted CHCs from workers that were frozen immediately after field sampling. As a consequence, our CHC and desiccation resistance were not taken from the same individual ants. Instead, we compared individual ant CHC profiles (predictor variable) to the mean colony-level LT₅₀ value for a given worker’s colony of origin (response variable). We log-transformed CHC abundance data to improve normality, as assessed by Q–Q plots of model residuals, and confirmed that data met assumptions homoscedasticity by inspecting plots of fitted values vs. residuals.

Finally, to analyze WorldClim data to assess environmental conditions at sampling sites, we used the R packages “raster” (Hijmans 2021), “rgdal” (Bivand et al. 2021), and created maps using the package “maps” (Becker et al. 2018). All statistical analyses were conducted in R 4.1.9 (Team, 2021).

Results

Effects of population and body mass on desiccation resistance

All ants in the 0% humidity treatment had died by 81 h post-introduction to the desiccation chambers (LT₅₀ = 36). Survival was high (86.7%) for workers in our high-humidity controls over the 81-h experimental duration, suggesting that the majority of death in our treatment groups can be attributed to desiccating conditions, as opposed to other stressors such as starvation. Resistance to desiccation increased significantly with log dry mass (Mixed-effect Cox regression: $X^2 = 44.36$; $df = 4$; $P = 0.004$; mean dry mass = 1.595 ± 0.019); (Fig. 1). The effect of population on desiccation was non-significant ($P = 0.096$), with workers from Lake Henshaw (majority monogynous pop.) and

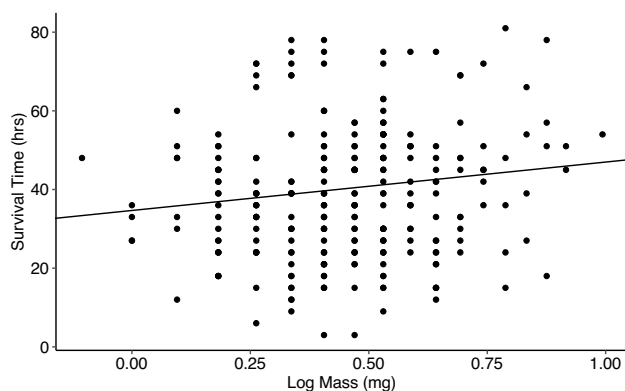


Fig. 1 The log dry mass of workers (mg) was positively related to survival time (hrs) in desiccation assays (slope = 12.305; Mixed-effect Cox regression: $X^2 = 44.36$; $df = 4$; $P = 0.004$)

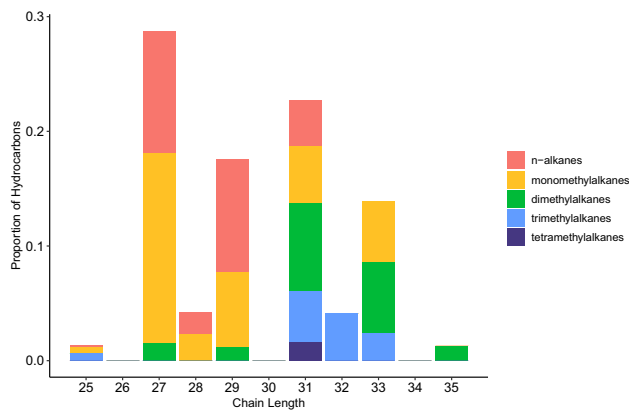


Fig. 2 Cuticular hydrocarbon profiles of *P. californicus* workers. Bars indicate proportions of the total hydrocarbon profile represented by each compound class for a given chain length (i.e., number of carbon atoms in the hydrocarbon chain). Proportions were calculated by pooling data from all 145 individuals

Cuyamaca (mixed mono/polygynous pop.) surviving slightly longer than workers from Pine Valley (majority polygynous pop.); ($LT_{50,LH} = 45$; $LT_{50,CM} = 45$; $LT_{50,PV} = 39$); (Supplementary Fig. 2). Within-colony variance in desiccation resistance did not differ across populations (Wilcoxon tests: Cuyamaca vs. Lake Henshaw, $P = 0.157$; Cuyamaca vs. Pine Valley, $P = 1.000$), though there is a trend toward differentiation in the comparison between Pine Valley and Lake Henshaw colonies (Wilcoxon test: $P = 0.056$).

Cuticular hydrocarbon profiles of *P. californicus* workers

We identified 48 unique CHC compounds in the cuticular surface profiles of *P. californicus* workers (Supplementary Table 2). Compounds consisted of linear and methyl-branched alkanes, ranging in carbon chain length from 25 to 35 (Fig. 2). CHC profiles were dominated by methyl-branched alkanes (72.0% of CHCs), especially monomethylalkanes, which account on average for 38.1% of the total CHC profile of analyzed workers. Principal component analysis revealed similarities in CHC profiles across sampling populations, shown by overlap in 95% confidence intervals (Fig. 3). In the PCA, the first two principal components explained the majority of variation in CHC abundance (PC1: 44.4%; PC2: 12.3%).

Effects of CHC profiles on colony desiccation resistance

Cuticular hydrocarbon profiles significantly predicted desiccation resistance in our model of absolute CHC abundances by compound class (linear and methyl-branched alkanes; Adj. $R^2 = 0.064$; $P = 0.004$). Colony LT_{50} was significantly

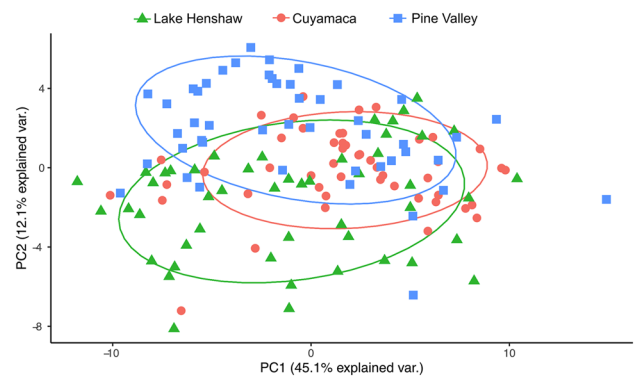


Fig. 3 Principal component analysis of the cuticular hydrocarbon profiles in workers from three sampling populations, plotted for the first two components, which explain 44.4% and 12.3% of the total variance, respectively. CHC abundance data are center log-ratio transformed. Ellipses show the 95% confidence intervals around the centroids for each population

and positively correlated to the log absolute abundance of linear *n*-alkanes (ANOVA: Adj. $R^2 = 0.062$; $P = 0.002$); (Fig. 4), but not to the log abundance of methyl-branched alkanes ($P = 0.474$).

Discussion

Water conservation presents a fundamental physiological challenge for terrestrial organisms, particularly small-bodied ectotherms like insects. Desiccation is one of the principal risks of insect foraging, especially in arid environments and for insects that forage on exposed ground surfaces in high-temperature boundary layers (Bernstein 1974; Oke 1978). As such, activity and distributions of insects are often shaped by moisture availability (Kaspari and Weiser 2000; Menke and Holway 2006), though the mechanisms supporting these

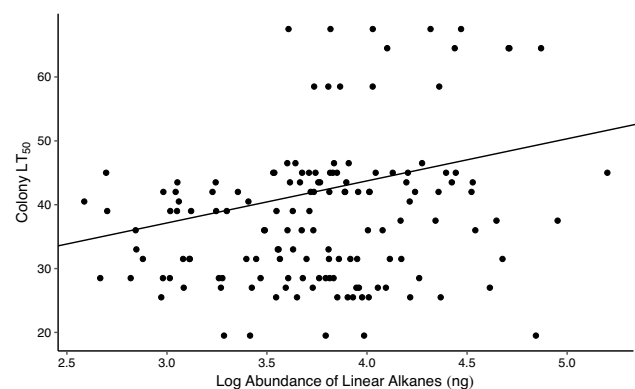


Fig. 4 The log abundance of linear alkanes (ng) in worker cuticular hydrocarbon profiles significantly predicted colony LT_{50} in desiccation trials (ANOVA: Adj. $R^2 = 0.062$; $P = 0.002$)

patterns have received little attention relative to temperature effects (Chown et al. 2011). Here, we present evidence for the roles of body mass and cuticular hydrocarbon (CHC) composition in determining the desiccation resistance of *P. californicus* workers from a semi-arid habitat. These results provide insights into the physiological mechanisms supporting insect survival in low-humidity conditions.

One of the clearest patterns in insect desiccation resistance is a positive relationship between body size and water conservation (Chown and Klok 2003; Fouet et al. 2012; Hadley 1994; Kærsgaard et al. 2004; Le Lagadec et al. 1998). We experimentally assayed desiccation resistance in *P. californicus* workers and found that larger ants survived longer at 0% humidity. Differential survival according to body size was likely driven by reduced surface area-to-volume ratios in larger-bodied ants (Hood and Tschinkel 1990; Kühnel et al. 2017), and perhaps proportionally larger water reserves (Lighton et al. 1994; Prange and Pinshow 1994). Unlike ant species that forage heavily on water-based foods like nectar (Nicolson 2007), *P. californicus* colonies subsist primarily on seeds, and to a lesser extent, other arthropods (Macmahon et al. 2000). This seed-based diet suggests that water intake during foraging is minimal. Future work measuring body water content in *P. californicus* in tandem with desiccation resistance could disentangle these effects.

The chemical composition of epicuticular lipids, especially CHCs, influences water loss in insects by determining the permeability of the cuticle to water (Gibbs 2002; Rourke and Gibbs 1999). Lipid melting increases transpiration of water through the cuticle, rendering insects vulnerable to desiccation (Gibbs 1998). Melting occurs over a range of temperatures determined by the chemical structure of CHC compounds; methyl-branching and double bonds both disrupt hydrocarbon packing, leading to lower melting points (Gibbs and Pomonis 1995). In line with this proposed mechanism for cuticular permeability, we found that desiccation resistance was positively correlated with absolute abundances of *n*-alkanes, but not methyl-branched alkanes. This association between desiccation resistance and the abundance of saturated, straight-chain molecules emphasizes the link between the physical properties of CHCs and their waterproofing performance under desiccating conditions.

These results also highlight the taxon-specific nature of chemical mechanisms for desiccation resistance. In line with our findings, foragers of the congener *Pogonomyrmex barbatus* have been found to possess higher proportions of *n*-alkanes than nest workers, likely as an adaptation to hot/dry conditions outside of the nest (Wagner et al. 1998). Menzel et al. (2018) exposed *Temnothorax* ants to low humidity stress and observed resulting increases in the abundance of *n*-alkanes and decreases in methyl-branched alkanes. Conversely, in other species, methyl-branched alkanes may play an important role in desiccation resistance. For example,

long-chain methyl-branched alkanes were positively associated with desiccation resistance in Argentine ants (*Linepithema humile*; Whyte et al. unpub.) and *Drosophila serrata* (Chung 2014). Despite containing methyl groups, which are associated with lower melting points (Gibbs and Pomonis 1995), the length of these compounds may lower the melting point sufficiently to protect against desiccation at typical environmental temperatures (Whyte et al. unpub.). It may be that the quantities of *n*-alkanes observed in our study are sufficiently protective against desiccation regardless of the quantities of methyl-branched alkanes in the rest of the CHC profile.

This interspecific variation in chemical mechanisms for desiccation resistance may be a consequence of constraints arising from the dual role of CHCs in waterproofing and communication (Gibbs 2002; Menzel et al. 2019). In social insects, CHCs additionally play a major role in nestmate recognition (Blomquist and Bagnères 2010; Leonhardt et al. 2016). In Argentine ants, methyl-branched alkanes are important both for colony recognition and desiccation resistance (Whyte et al. unpub., (Brandt et al. 2009; van Wilgenburg et al. 2010). The use of a particular compound class for one function (e.g., nestmate recognition) may constrain its use in another role (e.g., waterproofing). Our study represents the first characterization of the CHC profile of *P. californicus*. As such, it is unclear at this stage how these compounds relate to other behaviors (colony recognition, individual recognition, division of labor, etc.) in this species. Further, we present CHC information only for workers, not queens or males. Compared to workers, queens may exhibit different or additional chemical mechanisms for desiccation resistance, likely due to the additional role of CHCs in fertility signaling (Menzel et al. 2018). Characterizing the CHC profiles of *Pogonomyrmex* queens would be particularly interesting given their vulnerability to desiccation via cuticular abrasion during nest excavation (Johnson 2000; Johnson and Gibbs 2004). This desiccation stress during colony founding may represent an important selective pressure leading to the formation of foundress associations in this species (Cahan and Fewell 2004). As such, future studies more broadly clarifying the chemical/behavioral ecology of *P. californicus* have potential to shed additional light on the physiology of desiccation resistance in this species.

Interestingly, we observed relative homogeneity in desiccation resistance and CHC profiles across populations differing in queen social organization (monogyny vs. polygyny). The evolution of polygyny has often been linked to harsh environmental conditions (Bourke and Franks 1995; Heinze et al. 2017), but the physiological mechanisms underlying the adaptive benefits of polygyny remain unclear. In our study, worker desiccation resistance did not differ significantly across populations despite strong population-level variation in queen number (Overson et al.

2016). Importantly, we were not able to characterize individual colonies in our study as monogynous or polygynous. Instead, we took advantage of known variation in colony founding life history across a small and climatically homogeneous geographic region, with populations varying from predominantly monogynous (Lake Henshaw) to predominantly polygynous (Pine Valley); (Haney and Fewell 2018; Overson et al. 2016). A minority of colonies in these populations adopt the opposite founding strategy, and our study does not identify these cases. However, if queen number were an important driver of desiccation resistance in this species, we would still expect to see strong differentiation in survival outcomes across populations, a pattern we did not observe.

Polygyny can shape colony phenotypes not only by directionally shifting worker trait distributions, but also by impacting behavioral and genetic variation (Herbers 1986; Hölldobler and Wilson 1990). Little is known about how polygyny might affect variance in worker physiological tolerance, though it is feasible that a workforce derived from multiple matrilineal lines could show greater variation in physiological traits. Contrary to this expectation, colonies from the Pine Valley (polygynous) population were no more variable in desiccation resistance than colonies from Lake Henshaw (monogynous). Indeed, a marginally non-significant trend suggested the opposite: that colonies from Lake Henshaw may be more variable in their desiccation responses than Pine Valley colonies. A similar pattern was observed in a comparison of experimental associations of Pine Valley queens, where single-queen colonies were more variable in worker heat tolerance relative to multi-queen colonies (Baudier et al. 2022). It may be that within-colony genetic diversity is already high due to high levels of polyandry in this species (Overson et al. 2016), such that multiple matrilineal lines do not meaningfully increase worker phenotypic diversity. Together, these results call into question the intuition that multiple matrilineal lines should increase phenotypic variation in workers and highlight the need for further investigation of the relationship between polygyny and variation in environmental tolerance.

In conclusion, we found that body size and the abundance of *n*-alkanes predicted survival responses in low humidity conditions, across three populations of *P. californicus*. Despite associations between polygyny and survival in harsh conditions (Cahan 2001; Heinze 1993; Heinze and Hölldobler 1994; Rueppell et al. 2015), we found that workers from a majority-polygynous population were no more resistant to desiccation than those in a majority-monogynous population. Together, these results suggest that individual physiology and CHC composition are more important drivers of desiccation resistance than queen social phenotype. These results align with previous studies highlighting the role of body size and unsaturated, straight-chain hydrocarbons in adaptive responses to water conservation challenges.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s00360-023-01488-3>.

Acknowledgements We thank Dr. Rebecca Clark, Carol Iskander, Kathryn Myers, and Natalie Matis for assistance with field collections. We especially thank Weizhao Sun for help with calibrations. We gratefully acknowledge Dr. Neil Tsutsui and Brian Whyte for their expertise and helpful discussions about ant desiccation resistance. We also thank Lake Henshaw Resort for permission to collect on their premises. This research was supported by a German Research Foundation (DFG) award (part of the SFB TRR 212 (NC³)—TP C04, project numbers 316099922 and 396780988; to JG), startup funding provided by the University of Southern Mississippi (to KMB), an NSF Graduate Research Grant (to MMO), a Momental Foundation Mistletoe Research Fellowship (to MMO), and an ASU Graduate and Professional Student Association Grant (to MMO).

Data availability Data is made available in the supplementary information.

References

- Baudier KM, Ostwald MM, Haney BR, Calixto JM, Cossio FJ, Fewell JH (2022) Social factors in heat survival: multiqueen desert ant colonies have higher and more uniform heat tolerance. *Physiol Biochem Zool* 95(5):379–389. <https://doi.org/10.1086/721251>
- Becker R, Wilks A, Brownrigg R, Minka T, Deckman A (2018) maps: draw geographical maps. R package version 3.3.0.
- Bernstein R (1974) Seasonal food abundance and foraging activity in some desert ants. *Am Nat* 108(962):490–498
- Bivand R, Keitt T, Rowlingson B (2021) gdal: bindings for the “Geospatial” data abstraction library. R package version 1.5-23
- Blomquist G, Bagnères A (eds) (2010). Cambridge University Press, Cambridge
- Bonebrake TC, Mastrandrea MD (2010) Tolerance adaptation and precipitation changes complicate latitudinal patterns of climate change impacts. *PNAS* 107(28):12581–12586. <https://doi.org/10.1073/pnas.0911841107>
- Boulay R, Quagebeur M, Godzińska EJ, Lenoir A (1999) Social isolation in ants: evidence of its impact on survivorship and behavior in *Camponotus fellah* (Hymenoptera, Formicidae). *Sociobiology* 33(2):111–124
- Bourke A, Franks NR (1995) Social evolution in ants. Princeton University Press
- Brandt M, van Wilgenburg E, Sulc R, Shea KJ, Tsutsui ND (2009) The scent of supercolonies: the discovery, synthesis, and behavioural verification of ant colony recognition cues. *BMC Biol* 7:71. <https://doi.org/10.1186/1741-7007-7-71>
- Brown J, Reichman O, Davidson D (1979) Granivory in desert ecosystems. *Ann Rev Ecol Evol Syst* 10:201–227
- Buellesbach J, Whyte BA, Cash E, Gibson JD, Scheckel KJ, Sandidge R, Tsutsui N (2018) desiccation resistance and micro-climate adaptation: cuticular hydrocarbon signatures of different Argentine ant supercolonies across California. *J Chem Ecol* 44:1101–1114. <https://doi.org/10.1007/s10886-018-1029-y>
- Bujan J, Yanoviak SP, Kaspari M (2016) Desiccation resistance in tropical insects: causes and mechanisms underlying variability in a Panama ant community. *Ecol Evol* 6(17):6282–6291. <https://doi.org/10.1002/ece3.2355>
- Cahan S (2001) Cooperation and conflict in ant foundress associations: insights from geographical variation. *Anim Behav* 61(4):819–825. <https://doi.org/10.1006/anbe.2000.1671>

- Cahan S, Fewell J (2004) Division of labor and the evolution of task sharing in queen associations of the harvester ant *Pogonomyrmex californicus*. *Behav Ecol Sociobiol* 56(1):9–17. <https://doi.org/10.1007/s00265-003-0746-5>
- Chown SL, Klok C (2003) Water-balance characteristics respond to changes in body size in subantarctic weevils. *Physiol Biochem Zool* 76(5):634–643. <https://doi.org/10.1086/376919>
- Chown SL, Sørensen JG, Terblanche JS (2011) Water loss in insects: an environmental change perspective. *J Insect Physiol* 57(8):1070–1084. <https://doi.org/10.1016/j.jinsphys.2011.05.004>
- Chung H (2014) A single gene affects both ecological divergence and mate choice in *Drosophila*. *Science* 343:1148. <https://doi.org/10.1126/science.1249998>
- Chung H, Carroll S (2015) Wax, sex and the origin of species: dual roles of insect cuticular hydrocarbons in adaptation and mating. *Bioessays*. <https://doi.org/10.1002/bies.201500014>
- Clark R, Fewell J (2014) Social dynamics drive selection in cooperative associations of ant queens. *Behav Ecol* 25(1):117–123. <https://doi.org/10.1093/beheco/art093>
- Cook B, Mankin J, Anchukaitis K (2018) Climate change and drought: from past to future. *Curr Clim Change Rep* 4:164–179
- Ferveur JF, Cortot J, Rihani K, Cobb M, Everaerts C (2018) Desiccation resistance: effect of cuticular hydrocarbons and water content in *Drosophila melanogaster* adults. *PeerJ* 2018(2):1–23. <https://doi.org/10.7717/peerj.4318>
- Fick S, Hijmans R (2017) WorldClim 2: New 1 km spatial resolution climate surfaces for global land areas. *Int J Climatol* 37(12):4302–4315
- Fouet C, Gray E, Besansky NJ, Costantini C (2012) Adaptation to aridity in the malaria mosquito *Anopheles gambiae*: Chromosomal inversion polymorphism and body size influence resistance to desiccation. *PLoS ONE*. <https://doi.org/10.1371/journal.pone.0034841>
- Fung F, Lopez A, New M (2011) Water availability in + 2 °C and + 4 °C worlds. *Phil Trans R Soc A* 369:99–116. <https://doi.org/10.1098/rsta.2010.0293>
- Gibbs A (1998) Water-proofing properties of cuticular. *Lipids* 482:471–482
- Gibbs A (2002) Lipid melting and cuticular permeability: New insights into an old problem. *J Insect Physiol* 48:391–400
- Gibbs A, Pomonis J (1995) Physical properties of insect cuticular hydrocarbons: the effects of chain length, methyl-branching and unsaturation. *Comp Biochem Physiol* 112B(2):243–249
- Greenberg L, Fletcher DJC, Vinson S (1985) Differences in worker size and mound distribution in monogynous and polygynous colonies of the fire ant *Solenopsis invicta* buren. *J Kansas Entomol Soc* 58(1):9–18
- Hadley N (1994) Water relations of terrestrial arthropods. Academic Press
- Haney B, Fewell J (2018) Ecological drivers and reproductive consequences of non-kin cooperation by ant queens. *Oecologia* 187(3):643–655. <https://doi.org/10.1007/s00442-018-4148-9>
- Haney B, Fewell J, Cole B, Gadau J, Hölldobler B, Rutowski R (2017) Ecological drivers and reproductive consequences of queen cooperation in the California harvester ant *Pogonomyrmex californicus*. Arizona State University, Arizona
- Harvell CD, Mitchell CE, Ward JR, Altizer S, Dobson P, Ostfeld RS, Samuel MD (2002) Climate warming and disease risks for terrestrial and marine biota. *Science* 296(5576):2158–2162
- Heinze J (1993) Life histories of subarctic ants. *Arctic* 46(4):354–358
- Heinze J, Hölldobler B (1994) Ants in the cold. *Memorabilia Zoologica* 48:99–108
- Heinze J, Kellner K, Seal J (2017) Sociality in ants. In: Rubenstein DR, Abbot P (eds) *Comparative social evolution*. Cambridge University Press, pp 21–49
- Herbers J (1986) Ecological genetics of queen number in *Leptothorax longospinosus* (Hymenoptera: Formicidae). *Entomologia Generalis* 11:119–123
- Hijmans R (2021) raster: geographic data analysis and modeling. R package version 3.4-13
- Hoffmann A, Harshman L (1999) Desiccation and starvation resistance in *Drosophila*: patterns of variation at the species, population and intrapopulation levels. *Heredity* 83(6):637–643. <https://doi.org/10.1046/j.1365-2540.1999.00649.x>
- Hölldobler B, Wilson E (1990) The ants. Harvard University Press, In the ants
- Hood W, Tschinkel W (1990) Desiccation resistance in arboreal and terrestrial ants. *Physiol Entomol* 15(1):23–35. <https://doi.org/10.1111/j.1365-3032.1990.tb00489.x>
- Johnson R (2000) Water loss in desert ants: Caste variation and the effect of cuticle abrasion. *Physiol Entomol* 25(1):48–53. <https://doi.org/10.1046/j.1365-3032.2000.00170.x>
- Johnson R (2021) Desiccation limits recruitment in the pleometrotic desert seed-harvester ant *Veromessor pergandei*. *Ecol Evol* 11:294–308. <https://doi.org/10.1002/ece3.7039>
- Johnson R, Gibbs AG (2004) Effect of mating stage on water balance, cuticular hydrocarbons and metabolism in the desert harvester ant, *Pogonomyrmex barbatus*. *J Insect Physiol* 50(10):943–953. <https://doi.org/10.1016/j.jinsphys.2004.07.006>
- Kærsgaard CW, Holmstrup M, Malte H, Bayley M (2004) The importance of cuticular permeability, osmolyte production and body size for the desiccation resistance of nine species of *Collembola*. *J Insect Physiol* 50(1):5–15. <https://doi.org/10.1016/j.jinsphys.2003.09.003>
- Kaspari M, Weiser M (2000) Ant activity along moisture gradients in a Neotropical forest. *Biotropica* 32(4):703–711. <https://doi.org/10.1111/j.1744-7429.2000.tb00518.x>
- Kellermann V, Hoffmann AA, Overgaard J, Loeschcke V, Sgro CM (2018) Plasticity for desiccation tolerance across *Drosophila* species is affected by phylogeny and climate in complex ways. *Proc R Soc B* 285:20180048
- Klok CJ, Terblanche JS, Chown SL, Krafur ES (2006) Phenotypic plasticity and geographic variation in thermal tolerance and water loss of the tsetse *Glossina pallidipes* (Diptera: Glossinidae): Implications for distribution modeling. *Am J Trop Med Hyg* 74(5):786–794. <https://doi.org/10.4269/ajtmh.2006.74.786>
- Koto A, Mersch D, Hollis B, Keller L (2015) Social isolation causes mortality by disrupting energy homeostasis in ants. *Behav Ecol Sociobiol* 69(4):583–591. <https://doi.org/10.1007/s00265-014-1869-6>
- Kühnel S, Brückner A, Schmelzle S, Heethoff M, Blüthgen N (2017) Surface area—volume ratios in insects. *Insect Sci* 24:829–841. <https://doi.org/10.1111/1744-7917.12362>
- La Richelière F, Muñoz G, Guénard B, Dunn RR, Economo EP, Powell S, Sanders NJ, Weiser MD, Abouheif E, Lessard JP (2022) Warm and arid regions of the world are hotspots of superorganism complexity. *Proc R Soc B: Biol Sci*. <https://doi.org/10.1098/rspb.2021.1899>
- Le Lagadec MD, Chown SL, Scholtz CH (1998) Desiccation resistance and water balance in southern African keratin beetles (Coleoptera, Trogidae): the influence of body size and habitat. *J Comp Physiol B Biochem Syst Environ Physiol* 168(2):112–122. <https://doi.org/10.1007/s003600050127>
- Leonhardt S, Menzel F, Nehring V, Schmitt T (2016) Ecology and evolution of communication in social insects. *Cell* 164(6):1277–1287. <https://doi.org/10.1016/j.cell.2016.01.035>
- Lighton J, Quinlan M, Feener D (1994) Is bigger better? Water balance in the polymorphic desert harvester ant *Messor pergandei*. *Physiol Entomol* 19(4):325–334. <https://doi.org/10.1111/j.1365-3032.1994.tb01059.x>

- Macmahon JA, Mull J, Crist T (2000) Harvester ants (*Pogonomyrmex* spp.): their community and ecosystem influences. *Annu Rev Ecol Syst* 31:265–291. <https://doi.org/10.1146/annurev.ecolsys.31.1.265>
- Menke SB, Holway DA (2006) Abiotic factors control invasion by Argentine ants at the community scale. *J Anim Ecol* 75:368–376. <https://doi.org/10.1111/j.1365-2656.2006.01056.x>
- Menzel F, Zumbusch M, Feldmeyer B (2018) How ants acclimate: Impact of climatic conditions on the cuticular hydrocarbon profile. *Funct Ecol* 32:657–666. <https://doi.org/10.1111/1365-2435.13008>
- Menzel F, Morsbach S, Martens JH, Ra P, Hadjaje S, Poizat M (2019) Communication versus waterproofing the physics of insect cuticular hydrocarbons. *J Exp Biol*. <https://doi.org/10.1242/jeb.210807>
- Nicolson S (2007) Nectar consumers. In: Nicolson S, Nepi M, Pacini E (eds) *Nectaries and Nectar*. Springer, Dordrecht, pp 289–342
- Oke T (1978) *Boundary layer climates*, 2nd edn. Methuen and Co., Ltd
- Oksanen J, Guillaume Blanchet F, Friendly M, Kindt R, Legendre P, McGlinn D, Minchin P, O'Hara R, Simpson G, Solymos P, Stevens M, Szoecs E, Wagner H (2020) *vegan: community ecology package*. R Package version 2.5-7
- Otte T, Hilker M, Geiselhardt S (2018) Phenotypic plasticity of cuticular hydrocarbon profiles in insects. *J Chem Ecol* 44:235–247
- Overson R, Gadau J, Clark R, Pratt S, Fewell J (2014) Behavioral transitions with the evolution of cooperative nest founding by harvester ant queens. *Behav Ecol Sociobiol* 68(1):21–30. <https://doi.org/10.1007/s00265-013-1618-2>
- Overson R, Fewell J, Gadau J (2016) Distribution and origin of intraspecific social variation in the California harvester ant *Pogonomyrmex californicus*. *Insectes Soc* 63(4):531–541. <https://doi.org/10.1007/s00040-016-0497-8>
- Parmesan C (2006) Ecological and evolutionary responses to recent climate change. *Annu Rev Evol Syst* 37:637–669. <https://doi.org/10.1146/annurev.ecolsys.37.091305.110100>
- Pisarski B (1981) Intraspecific variations in ants of the genus *Formica* L. In: *Biosystematics of social insects*, vol 19. Academic Press, Amsterdam, pp 17–25
- Pörtner HO, Farrell AP (2008) Physiology and climate change. *Science* 322(5902):690–692
- Prange H, Pinshow B (1994) Thermoregulation of an unusual grasshopper in a desert environment: the importance of food source and body size. *J Therm Biol* 19(1):75–78
- Renault D, Vernon P, Vannier G (2005) Critical thermal maximum and body water loss in first instar larvae of three Cetoniidae species (Coleoptera). *J Therm Biol* 30(8):611–617. <https://doi.org/10.1016/j.jtherbio.2005.09.003>
- Ring R, Block W, Somme L, Worland M (1990) Body water content and desiccation resistance in some arthropods from subarctic South Georgia. *Polar Biol* 10:581–588
- Rourke B, Gibbs A (1999) Effects of lipid phase transitions on cuticular permeability: model membrane and in situ studies. *J Exp Biol* 202:3255–3262
- Rueppell O, Königseder F, Heinze J, Schrepf A (2015) Intrinsic survival advantage of social insect queens depends on reproductive activation. *J Evol Biol* 28(12):2349–2354. <https://doi.org/10.1111/jeb.12749>
- Sanderson M, Hemming D, Betts R (2011) Regional temperature and precipitation changes under high-end ($\geq 4^\circ\text{C}$) global warming. *Phil Trans R Soc A* 369:85–98. <https://doi.org/10.1098/rsta.2010.0283>
- Schultheiss P, Nooten SS (2013) Foraging patterns and strategies in an Australian desert ant. *Austral Ecol* 38(8):942–951. <https://doi.org/10.1111/aec.12037>
- Shaffer Z, Sasaki T, Haney B, Janssen M, Pratt S, Fewell J (2016) The foundress's dilemma: group selection for cooperation among queens of the harvester ant, *Pogonomyrmex californicus*. *Sci Rep* 6:29828. <https://doi.org/10.1038/srep29828>
- Sprenger PP, Burkert LH, Bérengère A, Federle W, Menzel F (2018) Coping with the climate: cuticular hydrocarbon acclimation of ants under constant and fluctuating conditions. *J Exp Biol*. <https://doi.org/10.1242/jeb.171488>
- Sprenger PP, Menzel F (2020) Cuticular hydrocarbons in ants (Hymenoptera: Formicidae) and other insects: how and why they differ among individuals, colonies, and species. *Myrmecol News* 30:1–26. <https://doi.org/10.25849/myrmecol.news>
- Stinziano JR, Sové RJ, Rundle HD, Sinclair BJ (2015) Rapid desiccation hardening changes the cuticular hydrocarbon profile of *Drosophila melanogaster*. *Comp Biochem Physiol Part A: Mol Integr Physiol* 180:38–42. <https://doi.org/10.1016/j.cbpa.2014.11.004>
- Tauber MJ, Tauber CA, Nyrop JP, Villani MG (1998) Moisture, a vital but neglected factor in the seasonal ecology of insects: hypotheses and tests of mechanisms. *Environ Entomol* 27(3):523–530. <https://doi.org/10.1093/ee/27.3.523>
- Team RC (2021) R: a language and environment for statistical computing. In: R Foundation for Statistical Computing
- Therneau T (2020) *Coxme: mixed effects cox models*. R package version 2.2-16
- Therneau T (2022) *A package for survival analysis in R*. R package version 3.3-0
- van Wilgenburg E, Sulc R, Shea KJ, Tsutsui ND (2010) Deciphering the chemical basis of nestmate recognition. *J Chem Ecol* 36:751–758. <https://doi.org/10.1007/s10886-010-9812-4>
- Wagner D, Brown MJF, Broun P, Cuevas W, Moses LE, Chao DL, Gordon DM (1998) Task-related differences in the cuticular hydrocarbon composition of harvester ants, *Pogonomyrmex barbatus*. *J Chem Ecol* 24(12):2021–2037. <https://doi.org/10.1023/A:1020781508889>
- Wang Y, Ferveur JF, Moussian B (2021) Eco-genetics of desiccation resistance in *Drosophila*. *Biol Rev* 96(4):1421–1440. <https://doi.org/10.1111/brv.12709>
- Woodrow RJ, Grace JK, Nelson LJ, Haverty MI (2000) Modification of cuticular hydrocarbons of *Cryptotermes brevis* (Isoptera: Kalotermitidae) in response to temperature and relative humidity. *Environ Entomol* 29(6):1100–1107. <https://doi.org/10.1603/0046-225X-29.6.1100>

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.