ORIGINAL PAPER

Blood oxygen stores of olive ridley sea turtles, *Lepidochelys olivacea* **are highly variable among individuals during arribada nesting**

B.Gabriela Arango¹[®] · Martha Harfush-Meléndez² · José Alejandro Marmolejo-Valencia³ · **Horacio Merchant‑Larios3 · Daniel E. Crocker1**

Received: 16 April 2020 / Revised: 14 September 2020 / Accepted: 29 September 2020 / Published online: 16 October 2020 © Springer-Verlag GmbH Germany, part of Springer Nature 2020

Abstract

Sea turtles dive with a full lung of air and these O_2 stores are supplemented by O_2 stored in blood and muscle. Olive ridley sea turtles exhibit polymorphic nesting behavior, mass nesting behavior called arribada, where thousands of turtles will nest at once, and solitary nesting behavior. The potential physiological diferences between the individuals using these strategies are not well understood. We measured blood volume and associated variables, including blood hemoglobin content and hematocrit, to estimate total blood O_2 stores. There were no significant differences in mean values between nesting strategies, but arribada nesting individuals were more variable than those performing solitary nesting. Mass-specifc plasma volume was relatively invariant among individuals but mass specifc blood volume and blood oxygen stores varied widely, twofold and threefold, respectively. Blood O_2 stores represented 32% of total body O_2 stores. Under typical mean diving conditions of 26 °C and high levels of activity, blood stores confer \sim 14 min to aerobic dive times and are likely critical for the long duration, deep diving exhibited by the species. Individual differences in blood $O₂$ stores strongly impact estimated aerobic dive limits and may constrain the ability of individuals to respond to changes on ocean climate.

Keywords Blood oxygen stores · Olive ridley · cADL · Arribada nesting · Solitary nesting

Abbreviations

Communicated by G. Heldmaier.

Electronic supplementary material The online version of this article [\(https://doi.org/10.1007/s00360-020-01321-1\)](https://doi.org/10.1007/s00360-020-01321-1) contains supplementary material, which is available to authorized users.

 \boxtimes B. Gabriela Arango bg.arango@berkeley.edu

- ¹ Biology Department, Sonoma State University, 1801 East Cotati Ave, Rohnert Park, CA 94928, USA
- ² Centro Mexicano de La Tortuga, Mazunte, Oaxaca, Mexico
- ³ Instituto de Investigaciones Biomédicas, Universidad Nacional Autónoma de México, Mexico, Mexico

Introduction

Air-breathing diving animals display a variety of anatomical, physiological, and behavioral adaptations that allow them to increase their time underwater used for foraging, migrating, mating and predator avoidance. These animals maximize time at depth through the use of aerobic metabolism and enhance dive durations through increased body $O₂$ stores (Kooyman [1989](#page-8-0)). Total body O_2 stores are dependent on several variables, including diving lung volume, blood volume, hemoglobin (Hb) concentration, muscle mass, and myoglobin (Mb) concentration (Ponganis [2011](#page-9-0)). While the $O₂$ stores and the physiology of breath-hold diving have been investigated extensively in endotherms, less is known about individual and inter-specific variation in diving $O₂$ stores in marine reptiles, including sea turtles (Berkson [1966;](#page-8-1) Wells and Baldwin [1994](#page-9-1); McMahon et al. [2007\)](#page-8-2).

As ectotherms, body temperature and rates of $O₂$ consumption are also infuenced by water temperature in the Cheloniidae sea turtles (Hochscheid et al. [2004](#page-8-3)). Understanding the factors that limit sea turtle breath-hold capacity is important to predicting their ability to respond to potential changes in water temperature and prey distributions that result from climate change (Yang et al. [2019\)](#page-9-2) and the potential for mortality due to incidental by-catch in fshing nets, the main anthropogenic threat to sea turtles (Polovina et al. [2003](#page-9-3), [2004](#page-9-4); Wallace et al. [2011\)](#page-9-5).

Another key diference among marine divers is whether or not they make use of their lung O_2 stores. The deepest divers, including many marine mammals, dive below the depth of lung collapse and prioritize tissue O_2 stores. Some deep divers, like phocid seals, avoid respiratory gas exchange at depth by diving after exhalation (Ponganis [2011](#page-9-0)). Other marine divers, such as sea turtles and penguins, dive following inhalation (Wood et al. 1984), and lung $O₂$ stores are an important component of diving metabolism (McDonald and Ponganis [2013\)](#page-8-4). In sea turtles, the lung serves as the major $O₂$ store during diving and limited measurements of tissue $O₂$ stores have suggested similar values to non-diving vertebrates (Lutz and Bentley [1985;](#page-8-5) Lutcavage et al. [1990,](#page-8-6) [1992](#page-8-7)). For this reason, the literature in sea turtles has de-emphasized measurements of blood O_2 stores with most published values from small samples, captive animals or juveniles.

Several recent studies suggest the importance of including blood O_2 stores in the consideration of breath-hold ability in sea turtles (Wells and Baldwin [1994](#page-9-1); Hochscheid et al. [2007](#page-8-8); McMahon et al. [2007;](#page-8-2) Chambault et al. [2016\)](#page-8-9)*.* Biologging data have revealed deep, long-duration dives in some species that may exceed the capacity of lung O_2 stores to support aerobic metabolism (McMahon et al. [2007](#page-8-2); Chambault et al. [2016](#page-8-9)). Lung collapse may occur in deep-diving turtles due to their compliant respiratory system on the deepest dives evident in biologging studies (Tenney et al. [1974](#page-9-7); Lutcavage et al. [1989\)](#page-8-10) and pulmonary shunts during diving may limit access to lung $O₂$, stores (Garcia Párraga et al. [2018](#page-8-11)). Together, these fndings suggest that contribution of tissue $O₂$ stores to aerobic limits may be critical for some dives. Further, recent studies on the sympathetic autonomic control of pulmonary shunts to limit decompression injuries in sea turtles may limit access to lung O_2 stores in some contexts and the lung stores are accessed using intermittent perfusion (Garcia Párraga et al. [2018](#page-8-11)).

The aerobic dive limit (ADL) is an estimation of physiological and energetic constraints on the dive durations of air-breathing diving animals. ADL is the maximum amount of time an animal can spend underwater before the product of anaerobic respiration, lactate, rises beyond resting levels in their blood (Costa et al. [2001\)](#page-8-12). However, establishing a direct and precise ADL value through measurements of blood lactate is difficult to achieve. Therefore, ADL is calculated as cADL, which is the sum of the total usable amount of O_2 stored in the body $(O_2$ from blood, muscles and lungs), divided by the diving metabolic rate (Ponganis et al. [2011](#page-9-8)). Values for cADL based on measurements of oxygen stores have only been reported in leatherbacks (*Dermochelys coriacea*) (Lutcavage et al. [1992;](#page-8-7) Southwood et al. [1999;](#page-9-9) Wallace et al. [2005;](#page-9-10) Bradshaw et al. [2007](#page-8-13)) and loggerheads (*Caretta caretta*) (Hochscheid et al. [2005\)](#page-8-14).

Despite being the smallest sea turtle, olive ridleys (*Lepidochelys olivacea),* exhibit long duration, deep dives when foraging pelagically. Mean dive duration for a large sample of adult females was 46 min (Chambault et al. [2016](#page-8-9)). Dives as long as 200 min (3.3 h) and as deep as 420 m have been reported (Polovina et al. [2003](#page-9-3); McMahon et al. [2007](#page-8-2); Da Silva et al. [2011](#page-8-15); Chambault et al. [2016\)](#page-8-9). To our knowledge, data for the calculation of total blood O_2 stores in the sea turtle, olive ridley have not been reported in the literature, except for one small sample $(n=2)$ of blood volume measurements (Thorson [1968\)](#page-9-11).

Olive ridley turtles exhibit two types of nesting behavior, solitary nesting and arribada nesting (Bernardo and Plotkin [2007](#page-8-16)). Solitary nesting occurs throughout the year and females are widely spaced, returning at varying inter-nesting intervals (Dornfeld et al. [2015\)](#page-8-17). During arribada nesting, thousands of females synchronize to nest together, during the third-quarter moon for a 2–7 day arribada period (Bernardo and Plotkin [2007\)](#page-8-16). Studies have suggested that arribada nesting may confer anti-predator benefts from predator–satiation (Eckrich and Owen [1995\)](#page-8-18) and may confer ftness benefts through multiple matings and paternity (Williamson et al. [2019\)](#page-9-12). However, offspring mortality may be increased at very high densities (Ocana et al. [2012](#page-9-13)) and density-dependent efects on environmental variables may have played a role in the decline of arribadas in some locations (Honarvar et al. [2008](#page-8-19)). In general, hatchling success is much lower at arribada beaches than in solitary nesters $(< 35 \text{ vs } > 75\%)$, suggesting solitary nesting may play important roles in maintaining populations (Bézy et al. [2014;](#page-8-20) Dornfeld et al. [2015](#page-8-17)). Additionally, arribada beaches might be important in the production of females, given their higher nesting temperatures (Valverde et al. [2010\)](#page-9-14), while solitary beaches might be important in the production of males for the opposite reason (Dornfeld et al. [2015\)](#page-8-17). Arribada and solitary nesters have distinct inter-nesting intervals, the time between two successful nesting events, (3 vs. 4 weeks) (Williamson et al. [2019](#page-9-12)). However, the physiological and behavioral diferences between individuals exhibiting the two strategies are not well understood.

Our primary objectives were to (1) measure plasma and blood volume and estimate blood O_2 stores in large sample of free- ranging adult female olive ridley sea turtles during nesting; (2) compare blood O_2 stores and associated variables between the arribada and solitary nesting strategies; (3) estimate cADL for the sampled olive ridley sea turtles at a range of water temperatures and activity levels and compare these estimates to published diving behavior.

Methods

Field procedures

Fig. 1 Marine Protected Area La Escobilla. Field site located in the Pacifc Ocean in Oaxaca, Mexico. Figure by E. Albavera-

Padilla

All animal handling procedures were approved by the Sonoma State University IACUC and performed under collecting permit SGPA/DGVS/12915/16. Samples were exported for laboratory analysis to Sonoma State University under exporting/importing permits CITES MX88143 and CITES 19US85728C/9. Nesting olive ridley females were sampled at the marine protected area of La Escobilla, Oaxaca, Mexico (15° 47′ N; 96° 44′ W; Fig. [1](#page-2-0)) during arribada in November 2017 ($n = 13$), and solitary nesting turtles were sampled both at Campamento Tortuguero Palmarito, Puerto Escondido, Oaxaca, México (15° 53′ 26.3″ N; 97° 07′ 52.2″ W), and at La Escobilla during solitary nesting in February 2017 (*n*=10). Handling and sampling procedures were performed on nesting turtles, after digging their nest, since this is when females enter

a 'trance-nesting period' (Dutton [1996\)](#page-8-21) and we were less likely to disrupt nesting. None of the females were disturbed from their nesting, nor returned to the sea without laying their eggs.

Straight carapace length and width were measured for all females. Females sampled during solitary nesting were weighed using a hand-held scale $(\pm 0.1 \text{ kg})$. Because of equipment failure, it was not possible to weigh the females sampled during arribada nesting. For these females, mass was estimated using a regression from published data on olive ridley morphometrics (Espinoza-Romo et al. [2018](#page-8-22); $n = 59$, mass = $-47.44 + 1.13 \times$ straight carapace length (SCL), $r^2 = 0.70$, $p < 0.001$.). This equation predicted the mass of the turtles that were weighed with a mean error of 4%.

All blood samples were collected from the cervical vein (Owens and Ruiz [1980\)](#page-9-15). An initial blood sample was taken into a 5 ml heparin Vacutainer tube. This sample was used for all hematological and plasma measurements. For plasma volume measurements, females were injected with Evan's blue dye (~ 1 mg/kg) and 3 sequential samples were taken into heparin tubes at \sim 7 min intervals after dye injection. Blood samples were kept on ice for 20 min until centrifuged

 \mathcal{D} Springer

at the feld site. The initial sample was used to measure hematocrit in triplicate using a clinical hematocrit centrifuge. All plasma samples were frozen after centrifugation at −20 °C until transported on dry ice to the laboratory at Sonoma State University. After transport, samples were stored at −80 °C until analysis. All samples were analyzed within 1 month of return.

Laboratory procedures

Blood O₂ stores

Hemoglobin (Hb) concentration was measured using the cyanmethemoglobin method (Sigma D5941, St. Louis, MO). Mean corpuscular hemoglobin concentration (MCHC) was calculated as Hb/Hct and expressed as 100 Hb Hct⁻¹. Plasma Volume (PV) was estimated using the dilution curve of the injected Evan's Blue dye (El-Sayed et al. [1995\)](#page-8-23). The syringes used to inject Evan's blue were calibrated gravimetrically to calculate the exact injection volume. Absorbance of Evans blue dye in the plasma was determined at 624 and 740 nm. Using the pre-injection samples, values at 740 nm were used to calculate the blank optical density at 624 nm and correct for hemolysis and precipitate in the postinjection samples (Foldager and Blomqvist [1991\)](#page-8-24). Values from serially collected samples were log-transformed, ft to a regression line, and the *y*-intercept was used to determine the instantaneous dilution volume by comparing to a serial dilution of the injectate (El-Sayed et al. [1995](#page-8-23)). Blood volume (BV) was calculated as PV $(1 - \text{Het}/100)$, where PV is absolute plasma volume. Total available blood O_2 stores were calculated assuming one-third of blood was arterial and two-thirds of blood was venous (Lutz and Bentley [1985](#page-8-5)), 95 and 80% saturation, respectively (Lutcavage et al. [1990,](#page-8-6) [1992](#page-8-7)), that arterial stores were depleted to 20% saturation and a Hb O₂ carrying capacity of 1.34 ml O₂ g⁻¹.

Total O₂ stores and cADL

Muscle and lung O_2 stores were estimated from mass and combined with usable blood O_2 stores to estimate total O_2 stores. No data for muscle myoglobin (Mb) exist for olive ridleys, so we used a published value from their closes relative, *Lepidochelys kempii*. Muscle O₂ stores were estimated assuming 22% of mass is muscle (Lutz and Bentley [1985\)](#page-8-5) and a Mb concentration of 3.1 mg g^{-1} muscle tissue (Stabenau and Heming 1994). Lung $O₂$ stores were estimated using an allometric equation for lung volume (Hochscheid et al. [2007](#page-8-8)),

 $lungO_2$ (*ml*) = 0.174(113.6 × *Mass*^{0.923})

and assuming the inspired air was 17.4% O₂ (Berkson [1966](#page-8-1)). O_2 consumption rate was estimated using the equation developed for captive loggerhead turtles that incorporates water temperatures and activity levels (Kinoshita et al. [2018\)](#page-8-25). Finally, cADL was calculated as the total usable body O_2 stores divided by the O_2 consumption rate. We calculated cADL for each individual turtle at 100% activity and a mean water temperature of 26 °C, the average temperature recorded for free-ranging olive ridleys (Chambault et al. [2016](#page-8-9)). We calculated and plotted cADL for total O_2 stores and for the contribution of blood $O₂$ stores across a range of activities from 0–100% and water temperatures ranging from 18.7 to 30 °C, the range of water temperatures encountered by free-ranging olive ridleys (McMahon et al. [2007;](#page-8-2) Chambault et al. [2016\)](#page-8-9).

Stress indicators

As an indicator of physiological stress levels in individuals, we measured serum corticosterone and plasma glucose and lactate concentrations. Corticosterone was measured in duplicate using a commercially available ELISA (Cayman Chemical, Ann Arbor, MI, USA; kit #500655). This assay platform was validated for use in olive ridley sea turtles. Diluted samples showed parallelism to the standard curve, linearity of measured concentrations (β = 1.04, r^2 = 0.99) and accuracy of recovered standards $(1.05 \pm 0.04\%)$. Intra-assay cv% was 4.71%. Glucose and lactate were measured in duplicate using a YSI-2300 STAT glucose/lactate autoanalyzer.

Statistical analysis

Statistical analysis was performed using JMP Pro 14 (SAS Institute, USA). Variables of interest were compared between the arribada and solitary nesting samples using two-sample *t* tests with pooled or unequal variance. Unequal variance was assessed using Levene's test. Relationships to body mass were assessed using simple linear regression. Statistical significance was considered at $p = 0.05$.

Results

Blood O₂ stores

Mean values for olive ridleys' mass, Hb, Hct, MCHC, PV and BV in wild nesting solitary females (*n*=10), wild nesting arribada females (*n*=13) are presented in Table [1](#page-4-0). Blood Hb did not vary between nesting strategies $(p=0.14)$ but was more variable during the arribada period (Levene's test, $p = 0.03$). Hct was similar during the 2 nesting strategies ($p = 0.38$). The variation in Hb content was unrelated

Table 1 Values from arribada nesting, solitary nesting and their averages

	Arribada nesting	Solitary nesting	Average	Range
Mass (kg)	31.5 ± 1.3	$29.6 + 0.9$	30.7 ± 0.7	$22.3 - 40.0$
Hb (g dL $^{-1}$)	$8.9 + 0.5$	$9.7 + 0.2$	$9.2 + 0.3$	$6.0 - 12.9$
$Hct (\%)$	$30.6 + 2.4$	$32.9 + 0.8$	$31.6 + 1.4$	$18 - 50$
MCHC $(g dL^{-1})$	0.31 ± 0.03	$0.30 + 0.01$	$0.31 + 0.01$	$0.18 - 0.43$
PV (mL)	$1768.1 + 44.8*$	$1506.5 + 43.6$	$1654.4 + 41.5$	
PV (mL $100 g^{-1}$)	5.8 ± 0.1	5.1 ± 0.1	5.4 ± 0.4	$4.4 - 6.6$
BV (mL)	$2593.3 + 121.8*$	$2247.9 + 68.7$	2443.1 ± 82.12	
BV (mL 100 g^{-1})	8.4 ± 0.1	7.6 ± 0.1	8.1 ± 0.1	$5.8 - 11.4$
Blood O_2 (mL kg ⁻¹)	7.7 ± 0.6	$7.8 + 0.3$	$7.7 + 0.4$	$3.6 - 10.8$

Values presented are least square means \pm standard error mean. Mass (kg)

Hb hemoglobin, *Hct* hematocrit, *MCHC* mean corpuscular hemoglobin concentration, *PV* plasma volume, *BV* blood volume

*Denotes significant differences between the nesting strategies $(p < 0.05)$

Fig. 2 Plasma volume increased with body mass. Open circles denote arribada nesting and closed circles denote solitary nesting $(r^2 = 0.31,$ p <0.0001). Plasma volume was calculated from dilution of Evan's Blue dye

to Hct $(p=0.33)$, due to individual differences in calculated MCHC. MCHC did not vary between nesting strategies $(p=0.58)$ but was more variable during the arribada period (Levene's test, $p=0.03$).

Mean PV increased significantly with increased body mass (PV = 28.3*mass + 795.4, r^2 = 0.31, F_{1,21} = 9.37 $p < 0.006$; Fig. [2\)](#page-4-1). PV was 5.4 ± 0.4 (SD) % of body mass and this percentage was greater during the arribada period $(t = 3.04, df = 21 p = 0.006)$. BV, calculated using PV and Hct was not related to body mass ($p = 0.35$ $p = 0.35$ $p = 0.35$; Fig. 3). BV was 8.1 ± 1.3 (SD) % of body mass and this percentage did not vary between nesting strategies $(p = 0.17)$. Estimated mass specific blood O_2 stores were 7.74 ± 1.8 (SD) ml O₂ kg⁻¹. This value did not vary between nesting strategies $(p = 0.92)$, but was more variable during the arribada period (Levene's test, $p = 0.008$; Fig. [4](#page-4-3)).

Fig. 3 Blood volume did not vary with body mass $(p=0.35)$. Open circles denote arribada nesting and closed circles denote solitary nesting. Blood volume was calculated as PV/ $[(100 - \text{Hct})^{-1}]$

Fig. 4 Mass-specific blood O_2 stores calculated from plasma volume and Hct. Stores were more variable during the arribada nesting period (Levene's test, $p=0.04$)

Fig. 5 Estimated cADL from total O_2 stores (**a**) and blood O_2 stores (**b**) at water temperatures ranging from 21.5–30.0 °C in 0.1 °C increments and 0–100% activity in 1% increments during the dive. Diving

metabolic rate was estimated using Eq. 1 from Williams et al., [2019](#page-9-17)

cADL and diving behavior cADL (min) from estimated total O₂ stores was calculated for water temperatures ranging 18.7–30.0 °C in 0.1 °C intervals (McMahon et al. [2007;](#page-8-2) Chambault et al. [2016\)](#page-8-9) and for activity levels of 0–100% of the dive in 1% intervals (Kinoshita et al. [2018](#page-8-25); Williams et al. [2019\)](#page-9-17) (Fig. [5a](#page-5-0)). At 26 °C, the mean temperature value encountered by olive ridley's during their post-nesting migration (Chambault et al. [2016\)](#page-8-9), cADL ranged from 40.5 ± 2.8 to 85.8 ± 5.9 min depending on activity levels. Blood O_2 stores represented $32.2 \pm 5.2\%$ of estimated total body O_2 stores. Thus, blood O₂ stores added 13.52 ± 2.9 to 27.9 ± 6.2 min to aerobic dive times in 26 °C water. In the coldest water (18.7 °C) and at rest (0% activity), blood O_2 stores added 42.7 ± 9.4 min to aerobic dive times (Fig. [5](#page-5-0)b).

Corticosterone and metabolites

Corticosterone concentrations were higher in arribada females (3.37 vs 2.69 ng/mL, *t*=3.14, *p*=0.004). Glucose concentrations were also higher in arribada females (4.56 vs 3.12 mmol/L , $t = 2.82$, $p = 0.01$). Lactate concentrations were highly variable among individuals but did not difer between the two breeding strategies (10.25 ± 4.54 mmol/L, $p=0.27$).

Discussion

For a large sample of free-ranging nesting olive ridley sea turtles, blood volume was 8.1 ml 100 g−1 body mass and blood O₂ stores were 7.7 ml O₂ kg⁻¹. This represents ~ 32% of body $O₂$ stores when making standard assumptions on lung volume and muscle stores. These stores may represent a critical component of aerobic capacity on the longest and deepest dives exhibited by individuals of the species. Massspecifc blood volume was similar to that reported for leatherbacks (7.7%; Table [2\)](#page-5-1) and was slightly higher than that reported previously for other chelonidae (6.6–6.7%; Thorson [1968\)](#page-9-11). However, as previously reported, blood volumes

Hb values are expressed as (g/100 mL blood). Blood volume is presented as mL 100 g⁻¹ (mass%) to facilitate comparison to published values

¹Thorson [1968,](#page-9-11) ²Davis [1991](#page-8-26), ³Marquez-M [1994,](#page-8-27) ⁴Keller et al. [2004,](#page-8-28) ⁵Muñoz-Pérez et al. [2017,](#page-8-29) ⁶Wood et al. [1984](#page-9-18), ⁷Wood and Ebanks 1984, ⁸Lutcavage et al. [1990,](#page-8-6) ⁹Lutcavage et al. [1992](#page-8-7)

were not remarkable when compared to terrestrial species (Thorson [1968\)](#page-9-11).

Mass-specific blood volume and blood O_2 stores varied widely among individuals, twofold and threefold, respectively. Plasma volume was relatively invariant and most of these diferences were due to variation in hematocrit, Hb and associated MCHC content. This wide individual variation in blood $O₂$ stores emphasizes the importance of physiological state to aerobic capacity and the value of having a large sample size for assessing capacity of a species. Hb concentrations decreased weakly with body mass, but the drivers of widely varying hematocrit and most of the variation in Hb content of blood are unknown. Large sample size studies of sea turtle hematology yield similarly wide ranges of Hct among individuals (e.g. Lewbart et al. [2014](#page-8-30); Rousselet et al. [2013](#page-9-19); Wood and Ebanks [1984](#page-9-18)) and have demonstrated associations of Hct with stress, contaminant burdens, ectoparasites and tumor scores (Work and Balazs [1999;](#page-9-20) Keller et al. [2004](#page-8-28); Stamper et al. [2005](#page-9-21); García-Párraga et al. [2014](#page-8-31)). A signifcant correlation between Hct and the straight carapace length of sea turtles has been previously reported (Wood and Ebanks [1984\)](#page-9-18) but we found no relationship to mass or size in our study.

We compiled published body mass, Hct, Hb content, BV and MCHC values for fve species of sea turtles (Table [2](#page-5-1)). When compared to the other species, olive ridley sea turtle has the lowest Hct and Hb values. MCHC values were relatively consistent among other species, with the exception of the deep-diving *D. coriacea*. Closely related *L. kempii* and *L. olivacea* (Crawford et al. [2015\)](#page-8-32) had highly similar MCHC values but showed inter-specifc variation in blood volume consistent with their diving patterns, *L. kempii* exhibit the shallowest dives of all sea turtles, with the deepest recorded dive of 5.3 m (Sasso and Witzell [2006\)](#page-9-22). *D. coriacea* is the species that had the largest values of mass-specifc blood volume, consistent with their behavior as the deepest diver from all seven species (Lutcavage et al. [1992;](#page-8-7) Fossette et al. [2010](#page-8-33)). What was most surprising was the striking similarity on BV values between *D. coriacea* (7.7 ml 100 g^{-1}) *and L. olivacea* (7.6 ml 100 g^{-1}) sea turtles. This supports previous observations of *L. olivacea* as being the second deepest diver (420 m; Da Silva et al. [2011\)](#page-8-15) of all species of sea turtles after *D. coriacea* (1280 m; López-Mendilaharsu et al. 2008). Blood O_2 stores may be especially critical if pulmonary shunts are activated as part of the dive response to avoid decompression injuries on the deepest and longest dives (Garcia Párraga et al. [2018\)](#page-8-11). Interestingly, olive ridley sea turtles newly calculated blood O_2 stores are similar to those reported for the best avian divers, emperor penguins (*Aptenodytes forsteri*) (Ponganis et al. [2011\)](#page-9-8), with blood stores representing 31% of body O_2 stores.

We found no significant difference in blood O_2 stores or associated variables between the two nesting strategies.

A previous study found that Hct is higher in migrating turtles compared to nesting values (Yang et al. [2019\)](#page-9-2). This diference was thought to be due to conditioning efects from longer, deeper dives during the migratory phase. We found no similar evidence of conditioning diferences between arribada and solitary turtles despite diferences in nesting strategies. It has been demonstrated that olive ridley populations difer widely in habitat use in that some individuals do not exhibit migrations and forage neriticly (Polovina et al. [2004;](#page-9-4) Chambault et al. [2016](#page-8-9)). We found no evidence of diferences in physiological capacity that suggests consistent at-sea behavioral diferences between the two strategies.

Although there were no signifcant diferences in mean values of hematological variables between nesting strategies, several measured variables were more variable during the arribada nesting. Hct, Hb, MCHC, mass-specifc PV and mass-specific blood O_2 stores were all more variable during arribada nesting. The underlying cause of this variability was not due to diferences in mass and likely refects physiological state. Some of these diferences may refect high levels of stress experienced by arribada nesters that might be a trade-off to the benefits of large aggregations. A recent study on transport stress in Kemp's ridley showed that the stress of transport increased variability in Hct in association with changes in corticosterone and glucose concentrations (Hunt et al. [2019](#page-8-35)). A study on diferent stress responses between solitary and nesting females found that arribada nesting turtles exhibited slower elevation of corticosterone in response to a stress test but did not compare baseline levels (Valverde et al. [1999\)](#page-9-23). Our data suggest signifcantly higher corticosterone and glucose concentrations in arribada females. This fnding was consistent with the idea that increased social stress during arribada may underlie the more highly variable Hct, but diferences in the variability of other variables (e.g. MCHC and PV%) are less likely to have arisen from acute stress.

Based on measures of diving behavior in olive ridley sea turtles, blood O_2 stores are likely a critical component of aerobic respiration to many dives. Lung stores are similar among species, suggesting that diving diferences might be most likely due to individual variation in blood O_2 stores rather than in muscle stores, given low myoglobin contents across species, \sim 4% of total body O₂ stores (Berkson [1966](#page-8-1); Lutz and Bentley [1985;](#page-8-5) Stabenau and Heming [1994](#page-9-16)). Previous investigations reporting deep and long-dive behaviors exhibited by olive ridley sea turtles (Polovina et al. [2004](#page-9-4); McMahon et al. [2007;](#page-8-2) Chambault et al. [2016\)](#page-8-9) demonstrated that 77% of dive time represents active foraging behaviors, and that some dives exceed the presumed depths of lung collapse (~120 m, Berkson [1967](#page-8-36)). Thus, selection for increased blood O_2 stores may represent an important physiological adaptation for deep diving.

Our cADL estimates are limited by the paucity of information on olive ridleys. We lacked individual data on lung volume and muscle Mb content and estimated these values using data from other species. We also estimated Hb saturation at the beginning of the dive and arterial/venous diferences using data from another species and used an assumed level of potential arterial desaturation. Despite this imprecision, our estimates clearly show the importance of blood O_2 stores to support the long-duration dives exhibited by the species. Our cADL estimates that incorporate blood stores are \sim 41 min for dives with high levels of activity and at mean water temperatures as reported for foraging dives. Blood O_2 stores provide ~ 14 min of that dive time and are likely necessary to support the diving behavior on many dives. Recorded average dive duration in olive ridleys is \sim 46 min and 40% of dives ranged between 30 and 50 min (Chambault et al. [2016](#page-8-9)). All dives are likely aerobic as their short and invariant post-dive surface times do not suggest that turtles are incurring a lactate debt. Use of pulmonary shunts to avoid decompression injury on some deep dives may further emphasize the importance of blood stores (Garcia Parraga et al. [2018\)](#page-8-11).

Bycatch, the primary mortality threat to sea turtles, results in death from drowning or signs of decompression sickness in 43% of incidental captures (García-Párraga et al. [2014](#page-8-31)). The evidence for decompression injuries in incidentally captured turtles, emphasizes the importance of modulation of lung perfusion during diving in sea turtles and the potential for loss of access to lung stores during all or parts of dives. Individual and inter-specifc variation in blood $O₂$ stores may be an important factor in avoiding drowning when captured in fishing nets.

The primary determinant of rates of O_2 use in diving sea turtles is activity levels (Williams et al. [2019\)](#page-9-17). However, given high rates of activity on most foraging dives, water temperature is also an important modulator of diving metabolism. Changes in water temperature due to global warming can directly afect ectothermic body temperature and lower breath-hold capacity. Based on our calculations, a 1-degree change in mean water temperature reduces aerobic dive time by~6% at high activity levels. Further, changes in thermal structure, currents and prey distribution in response to climate change may require substantial alterations to diving behavior (Hawkes et al. [2009\)](#page-8-37). Individual and inter-specifc variation in blood O_2 stores may be a crucial component of enabling behavioral plasticity in response to these changes. Given that this study only measured blood O_2 stores directly and used previously published values of lungs and muscles to cADL, direct measures of ADL in the future would be more accurate to determine the exact ADL on olive ridley sea turtles.

This first large sample study on blood O_2 stores in olive ridley sea turtles revealed that these stores represent a critical component of total body O_2 stores. Mass-specific blood volumes were similar to that reported for larger deeper diving leatherback sea turtles and appear to be essential to support the long-duration diving behavior reported for olive ridleys. The wide individual variation in blood $O₂$ stores has important implications for intra-specifc variation in breathhold ability. This variation was greater during the arribada nesting period due to increased variation in hematocrit, PV and MCHC. High blood O_2 stores may be a critical component of the behavioral plasticity needed to respond to changes in prey distribution, thermal structure and currents that result from climate change.

Acknowledgments We are profoundly grateful to *Centro Mexicano de la Tortuga* and *Campamento Totuguero Palmarito* personnel, especially to Ernesto Albavera-Padilla for his assistance with the logistics and useful discussions; to MVZ Elpidio Marcelino López-Reyes, Carmelo Ambrosio, Alberto Jarquín and Antonio Santiago for their assistance during feld work. In addition, we are thankful to *Cooperativa la Tortuga Felíz*, Heradio-Santillán and Sóstenes Rodríguez-Reyes from *La Escobilla Campamento Tortuguero,* and to MVZ Javier Beltran-Robledo for their assistance with feldwork and sample processing. Lastly, we thank Stacey Pelton from Sonoma State McNair's Scholarship Program for invaluable support. B. G. Arango was funded in part by Sally Casanova and McNair Scholar programs. José Alejandro Marmolejo-Valencia and Horacio Merchant-Larios were funded by UNAM grant PAPIIT-IN201218.

Authors' contributions All authors contributed to the study conception and design. Material preparation, data collection and analysis were performed by B. Gabriela Arango, Martha Harfush-Meléndez and Daniel E. Crocker. The frst draft of the manuscript was written by B. Gabriela Arango and all authors commented on previous versions of the manuscript. All authors read and approved the fnal manuscript.

Funding B. Gabriela Arango was funded in part by Sally Casanova and McNair Scholar programs. José Alejandro Marmolejo-Valencia and Horacio Merchant-Larios were funded by UNAM grant PAPIIT-IN201218.

Data accessibility Data deposited in the Dryad Digital Repository: <https://doi.org/10.5061/dryad.nk98sf7s0> (Arango, Gabriela [2020](#page-7-0)).

Ethics approval Work was approved by the Sonoma State University IACUC and performed under collecting permit SGPA/ DGVS/12915/16. Samples were exported/imported under CITES MX88143 and CITES 19US85728C/9.

Consent to participate/publication and availability of data/code Not applicable.

Conflict of interest The authors declare no confict of interest.

References

Arango, Gabriela (2020) Olive ridley blood volume by nesting strategy. Dryad, Dataset. <https://doi.org/10.5061/dryad.nk98sf7s0>

- Berkson H (1966) Physiological adjustments to prolonged diving in the Pacifc green turtle (*Chelonia mydas* agassizii). Comp Biochem Physiol 18:101–119
- Berkson H (1967) Physiological adjustments to deep diving in the Pacifc green turtle (*Chelonia mydas* agassizzii). Comp Biochem Physiol 21:507–524
- Bernardo J, Plotkin PT (2007) An evolutionary perspective on the arribada phenomenon and reproductive behavioral polymorphism of olive ridley sea turtles (Lepidochelys olivacea). In: Plotkin PT (ed) Biology and conservation of Ridley Sea Turtles. The Johns Hopkins University Press, Baltimore, pp 59–87
- Bézy VS, Valverde RA, Plante CJ (2014) Olive ridley sea turtle hatching success as a function of microbial abundance and the microenvironment of in situ nest sand at Ostional, Costa Rica. J Mar Biol 2014:1–10. [https://doi.org/10.1371/journ](https://doi.org/10.1371/journal.pone.0118579) [al.pone.0118579](https://doi.org/10.1371/journal.pone.0118579)
- Bradshaw CJA, McMahon CR, Hays GC (2007) Behavioral inference of diving metabolic rate in free-ranging leatherback turtles. PhysiolBiochemZool 80:209–219.<https://doi.org/10.1086/511142>
- Chambault P, de Thoisy B, Heerah K et al (2016) The infuence of oceanographic features on the foraging behavior of the olive ridley sea turtle *Lepidochelys olivacea* along the Guiana coast. Prog Oceanogr 142:58–71. [https://doi.org/10.1016/j.pocea](https://doi.org/10.1016/j.pocean.2016.01.006) [n.2016.01.006](https://doi.org/10.1016/j.pocean.2016.01.006)
- Costa DP, Gales NJ, Goebel ME (2001) Aerobic dive limit: how often does it occur in nature? Comp BiochemPhysiol A Mol IntegrPhysiol 129:771–783. [https://doi.org/10.1016/S1095-6433\(01\)00346-4](https://doi.org/10.1016/S1095-6433(01)00346-4)
- Crawford NG, Parham JF, Sellas AB et al (2015) A phylogenomic analysis of turtles. Mol Phylogenet Evol 83:250–257. [https://doi.](https://doi.org/10.1016/j.ympev.2014.10.021) [org/10.1016/j.ympev.2014.10.021](https://doi.org/10.1016/j.ympev.2014.10.021)
- Da Silva ACCD, Dos Santos EAP, Oliveira FLDC et al (2011) Satellite-tracking reveals multiple foraging strategies and threats for olive ridley turtles in Brazil. Mar Ecol Prog Ser 443:237–247. <https://doi.org/10.3354/meps09427>
- Davis BJ (1991) Developmental changes in the blood oxygen transport system of Kemp's ridley sea turtle, *Lepidochelys kempii*. Can J Zool 69:2660–2666
- Dornfeld TC, Robinson NJ, Tomillo PS, Paladino FV (2015) Ecology of solitary nesting olive ridley sea turtles at Playa Grande, Costa Rica. Mar Biol 162:123–139. [https://doi.org/10.1007/s0022](https://doi.org/10.1007/s00227-014-2583-7) [7-014-2583-7](https://doi.org/10.1007/s00227-014-2583-7)
- Dutton PH (1996) Methods for collection and preservation of samples for sea turtle genetic studies. In: Bowen BW, Witzell WN (eds) Proceedings of the International Symposium. Miami, FL, pp 17–24
- Eckrich CE, Owen DW (1995) Solitary versus Arribada nesting in the olive ridley sea turtles (*Lepidochelys olivacea*): a test of the predator-satiation hypothesis. Herpetologica 51:349–354
- El-Sayed H, Goodall SR, Hainsworth R (1995) Re-evaluation of Evans blue dye dilution method of plasma volume measurement. Clin Lab Haematol 17:189–194
- Espinoza-Romo BA, Sainz-Hernández JC, Ley-Quiñónez CP et al (2018) Blood biochemistry of olive ridley (*Lepidochelys olivacea*) sea turtles foraging in northern Sinaloa, Mexico. PLoS ONE 13:1–12.<https://doi.org/10.1371/journal.pone.0199825>
- Foldager N, Blomqvist C (1991) Repeated plasma volume determination with the Evans blue dye dilution technique: the method and a computer program. Comput Biol Med 21:35–41
- Fossette S, Gleiss AC, Myers AE et al (2010) Behaviour and buoyancy regulation in the deepest-diving reptile: the leatherback turtle. J Exp Biol 213:4074–4083.<https://doi.org/10.1242/jeb.048207>
- García-Párraga D, Crespo-Picazo JL, Belnaldo De Quirós Y et al (2014) Decompression sickness ('the bends') in sea turtles. Dis Aquat Organ 111:191–205. <https://doi.org/10.3354/dao02790>
- Garcia Párraga D, Moore M, Fahlman A (2018) Pulmonary ventilation-perfusion mismatch: a novel hypothesis for how diving

vertebrates may avoid the bends. Proc R Soc B BiolSci. [https://](https://doi.org/10.1098/rspb.2018.0482) doi.org/10.1098/rspb.2018.0482

- Hawkes LA, Broderick AC, Godfrey MH, Godley BJ (2009) Climate change and marine turtles. Endanger Species Res 7:137–154. [https](https://doi.org/10.3354/esr00198) [://doi.org/10.3354/esr00198](https://doi.org/10.3354/esr00198)
- Hochscheid S, Bentivegna F, Hays GC (2005) First records of dive durations for a hibernating sea turtle. Biol Lett 1:82–86. [https://](https://doi.org/10.1098/rsbl.2004.0250) doi.org/10.1098/rsbl.2004.0250
- Hochscheid S, Bentivegna F, Speakman JR (2004) Long-term cold acclimation leads to high Q 10 efects on oxygen consumption of Loggerhead Sea Turtles *Caretta caretta*. Physiol Biochem Zool 77:209–222.<https://doi.org/10.1086/381472>
- Hochscheid S, McMahon CR, Bradshaw CJA et al (2007) Allometric scaling of lung volume and its consequences for marine turtle diving performance. Comp BiochemPhysiol A Mol IntegrPhysiol 148:360–367. <https://doi.org/10.1016/j.cbpa.2007.05.010>
- Honarvar S, O'Connor MP, Spotila JR (2008) Density-dependent efects on hatching success of the olive ridley turtle, *Lepidochelys olivacea*. Oecologia 157:221–230. [https://doi.org/10.1007/s0044](https://doi.org/10.1007/s00442-008-1065-3) [2-008-1065-3](https://doi.org/10.1007/s00442-008-1065-3)
- Hunt KE, Innis C, Merigo C et al (2019) Ameliorating transport-related stress in endangered Kemp's ridley sea turtles (*Lepidochelys kempii*) with a recovery period in saltwater pools. Conserv Physiol 7:coy065.<https://doi.org/10.1093/conphys/coy065>
- Keller JM, Kucklick JR, Stamper MA et al (2004) Associations between organochlorine contaminant concentrations and clinical health parameters in loggerhead sea turtles from North Carolina, USA. Environ Health Perspect 112:1074–1079. [https://doi.](https://doi.org/10.1289/ehp.6923) [org/10.1289/ehp.6923](https://doi.org/10.1289/ehp.6923)
- Kinoshita C, Fukuoka T, Niizuma Y et al (2018) High resting metabolic rates with low thermal dependence induce active dives in overwintering Pacifc juvenile loggerhead turtles. J Exp Biol 221:jeb175836. <https://doi.org/10.1242/jeb.175836>
- Kooyman GL (1989) Diverse divers: physiology and behavior. Springer-Verlag, Berlin
- Lewbart GA, Hirschfeld M, Denkinger J et al (2014) Blood gases, biochemistry, and hematology of galapagos green turtles (*Chelonia mydas*). PLoS ONE 9:1–7. [https://doi.org/10.1371/journ](https://doi.org/10.1371/journal.pone.0096487) [al.pone.0096487](https://doi.org/10.1371/journal.pone.0096487)
- López-Mendilaharsu M, Rocha CFD, Domingo A et al (2008) Prolonged, deep dives by the leatherback turtle *Dermochelys coriacea*: pushing their aerobic dive limits. J Mar Biol Assoc 2(2):1–3. <https://doi.org/10.1017/S1755267208000390>
- Lutcavage ME, Bushnell PG, Jones DR (1992) Oxygen stores and aerobic metabolism in the leatherback sea turtle. Can J Zool 70:348–351
- Lutcavage ME, Bushnell PG, Jones DR (1990) Oxygen transport in the Leatherback Sea Turtle *Dermochelys coriacea*. Physiol Zool 63:1012–1024
- Lutcavage ME, Lutz PL, Baier H (1989) Respiration mechanics of the loggerhead sea turtle, *Caretta caretta*. Respir Physiol 76:13–24
- Lutz PL, Bentley TB (1985) Respiratory physiology of diving in the sea turtle. Copeia 3:671–679
- Marquez-MR (1994) Synopsis of Biological Data on the Kemp's Ridley Turtle, *Lepidochelys kempii* (Garman, 1880), pp 1–91
- McDonald BI, Ponganis PJ (2013) Insights from venous oxygen profles: oxygen utilization and management in diving California sea lions. J Exp Biol 216:3332–3341. [https://doi.org/10.1242/](https://doi.org/10.1242/jeb.085985) [jeb.085985](https://doi.org/10.1242/jeb.085985)
- McMahon CR, Bradshaw CJA, Hays GC (2007) Satellite tracking reveals unusual diving characteristics for a marine reptile, the olive ridley turtle *Lepidochelys olivacea*. Mar Ecol Prog Ser 329:239–252. <https://doi.org/10.3354/meps329239>
- Muñoz-Pérez JP, Lewbart GA, Hirschfeld M et al (2017) Blood gases, biochemistry and haematology of Galápagos hawksbill turtles (*Eretmochelys imbricata*). Conserv Physiol 5:1–9
- Ocana M, Harfush-Melendez M, Heppell S (2012) Mass nesting of olive ridley sea turtles Lepidochelys olivacea at La Escobilla, Mexico: linking nest density and rates of destruction. Endanger Species Res 16:45–54. <https://doi.org/10.3354/esr00388>
- Owens DW, Ruiz GJ (1980) New methods of obtaining blood and cerebrospinal fuid from marine turtles. Herpetologica 36:17–20
- Polovina JJ, Balazs GH, Howell EA et al (2004) Forage and migration habitat of loggerhead (*Caretta caretta*) and olive ridley (*Lepidochelys olivacea*) sea turtles in the central North Pacifc Ocean. Fish Oceanogr 13:36–51
- Polovina JJ, Howell E, Parker DM, Balazs GH (2003) Dive-depth distribution of loggerhead (*Carretta carretta*) and olive ridley (*Lepidochelys olivacea*) sea turtles in the central North Pacifc: might deep longline sets catch fewer turtles? Fish Bull 101:189–193
- Ponganis PJ (2011) Diving mammals. ComprPhysiol 1:447–465. [https](https://doi.org/10.1002/cphy.c091003) [://doi.org/10.1002/cphy.c091003](https://doi.org/10.1002/cphy.c091003)
- Ponganis PJ, Meir JU, Williams CL (2011) In pursuit of Irving and Scholander: a review of oxygen store management in seals and penguins. J Exp Biol 214:3325–3339. [https://doi.org/10.1242/](https://doi.org/10.1242/jeb.031252) ieb.031252
- Rousselet E, Levin M, Gebhard E et al (2013) Evaluation of immune functions in captive immature loggerhead sea turtles (*Caretta caretta*). Vet Immunol Immunopathol 156:43–53. [https://doi.](https://doi.org/10.1016/j.vetimm.2013.09.004) [org/10.1016/j.vetimm.2013.09.004](https://doi.org/10.1016/j.vetimm.2013.09.004)
- Sasso CR, Witzell WN (2006) Diving behaviour of an immature Kemp's ridley turtle (*Lepidochelys kempii*) from Gullivan Bay, ten thousand Islands, south-west Florida. J Mar Biol Assoc UK 86:919–925.<https://doi.org/10.1017/S0025315406013877>
- Southwood AL, Andrews RD, Lutcavage ME et al (1999) Heart rates and diving behavior of leatherback sea turtles in the Eastern Pacifc ocean. J Exp Biol 202:1115–1125
- Stabenau EK, Heming TA (1994) The in vitro respiratory and acid-base properties of blood and tissue from the Kemp's ridley sea turtle, *Lepidochelys kempii*. Can J Zool 72:1403–1408
- Stamper MA, Harms C, Epperly SP et al (2005) Relationship between barnacle epibiotic load and hematologic parameters in loggerhead sea turtles (*Caretta caretta*), a comparison between migratory and residential animals in Pamlico Sound, North Carolina. J Zoo Wildl Med 36:635–641.<https://doi.org/10.1638/04-074.1>
- Tenney SM, Bartlett J, Farber JP, Remmers JE (1974) Mechanics of the respiration cycle in the green turtle (*Chelonia mydas*). Respir Physiol 22:361–368
- Thorson TB (1968) Body fluid partitioning in Reptilia. Copeia 3:592–601
- Valverde RA, Owens DW, Mackenzie DS, Amoss MS (1999) Basal and stress-induced corticosterone levels in olive ridley sea turtles

(*Lepidochelys olivacea*) in relation to their mass nesting behavior. J Exp Zool 284:652–662. [https://doi.org/10.1002/\(SICI\)1097-](https://doi.org/10.1002/(SICI)1097-010X(19991101)284:6<652:AID-JEZ7>3.0.CO;2-U) [010X\(19991101\)284:6<652:AID-JEZ7>3.0.CO;2-U](https://doi.org/10.1002/(SICI)1097-010X(19991101)284:6<652:AID-JEZ7>3.0.CO;2-U)

- Valverde RA, Wingard S, Gómez F et al (2010) Field lethal incubation temperature of olive ridley sea turtle *Lepidochelys olivacea* embryos at a mass nesting rookery. Endanger Species Res 12:77– 86.<https://doi.org/10.3354/esr00296>
- Wallace BP, DiMatteo AD, Bolten AB et al (2011) Global conservation priorities for Marine turtles. PLoS ONE. [https://doi.org/10.1371/](https://doi.org/10.1371/journal.pone.0024510) [journal.pone.0024510](https://doi.org/10.1371/journal.pone.0024510)
- Wallace BP, Williams CL, Paladino FV et al (2005) Bioenergetics and diving activity of internesting leatherback turtles Dermochelys coriacea at Parque Nacional Marino Las Baulas, Costa Rica. J Exp Biol 208:3873–3884.<https://doi.org/10.1242/jeb.01860>
- Wells RM, Baldwin J (1994) Oxygen transport in Marine Green Turtle (*Chelonia mydas*) hatchlings—blood viscosity and control of hemoglobin oxygen—affinity. J Exp Biol 188:103-114
- Williams CL, Sato K, Ponganis PJ (2019) Activity not submergence explains diving heart rates of captive loggerhead turtles. J Exp Biol 222:jeb.200824.<https://doi.org/10.1242/jeb.200824>
- Williamson SA, Evans RG, Robinson NJ, Reina RD (2019) Synchronised nesting aggregations are associated with enhanced capacity for extended embryonic arrest in olive ridley sea turtles. Sci Rep 9:9783.<https://doi.org/10.1038/s41598-019-46162-3>
- Wood FE, Ebanks GK (1984) Blood cytology and hematology of the green sea turtle, *Chelonia mydas*. Herpetologica 40:331–336
- Wood SC, Gatz RN, Glass ML (1984) Oxygen transport in the green sea turtle. J Comp Physiol B 154:275–280. [https://doi.](https://doi.org/10.1007/BF02464407) [org/10.1007/BF02464407](https://doi.org/10.1007/BF02464407)
- Work TM, Balazs GH (1999) Relating tumor score to hematology in green turtles with Fibropapillomatosis in Hawaii. J Wildl Dis 35:804–807.<https://doi.org/10.7589/0090-3558-35.4.804>
- Yang T, Haas HL, Patel S et al (2019) Blood biochemistry and haematology of migrating loggerhead turtles (*Caretta caretta*) in the Northwest Atlantic: reference intervals and intra-population comparisons. Conserv Physiol 7:1–15. [https://doi.org/10.1093/conph](https://doi.org/10.1093/conphys/coy079) [ys/coy079](https://doi.org/10.1093/conphys/coy079)

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.