REVIEW

How low can you go? An adaptive energetic framework for interpreting basal metabolic rate variation in endotherms

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Abstract Adaptive explanations for both high and low body mass-independent basal metabolic rate (BMR) in endotherms are pervasive in evolutionary physiology, but arguments implying a direct adaptive benefit of high BMR are troublesome from an energetic standpoint. Here, we argue that conclusions about the adaptive benefit of BMR need to be interpreted, first and foremost, in terms of energetics, with particular attention to physiological traits on which natural selection is directly acting. We further argue from an energetic perspective that selection should always act to reduce BMR (i.e., maintenance costs) to the lowest level possible under prevailing environmental or ecological demands, so that high BMR per se is not directly adaptive. We emphasize the argument that high BMR arises as a correlated response to direct selection on other physiological traits associated with high ecological or environmental

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costs, such as daily energy expenditure (DEE) or capacities for activity or thermogenesis. High BMR thus represents elevated maintenance costs required to support energetically demanding lifestyles, including living in harsh environments. BMR is generally low under conditions of relaxed selection on energy demands for high metabolic capacities (e.g., thermoregulation, activity) or conditions promoting energy conservation. Under these conditions, we argue that selection can act directly to reduce BMR. We contend that, as a general rule, BMR should always be as low as environmental or ecological conditions permit, allowing energy to be allocated for other functions. Studies addressing relative reaction norms and response times to fluctuating environmental or ecological demands for BMR, DEE, and metabolic capacities and the fitness consequences of variation in BMR and other metabolic traits are needed to better delineate organismal metabolic responses to environmental or ecological selective forces.

Keywords Basal metabolic rate · Endotherms · Selection · Evolutionary physiology · Daily energy expenditure · Energetics

Introduction

Comparisons of energetic traits among endotherms across various environments and life history strategies require standardized measurements of metabolic rates, with basal metabolic rate (BMR) being by far the most commonly used metric for comparative analyses. Measurements of BMR have indeed been conducted for decades (e.g., Riddle et al. [1932;](#page-15-0) Scholander et al. [1950a](#page-16-0); McKechnie [2008](#page-14-0); McKechnie and Swanson [2010](#page-14-1); Fristoe et al. [2015](#page-13-0)). BMR is defined as the minimum metabolic rate of a normothermic,

resting endotherm, within the thermoneutral zone, and in the absence of digestive, circadian, or other increments in metabolic heat production (Benedict [1938](#page-12-0); Kleiber [1961](#page-14-2); Bligh and Johnson [1973](#page-12-1); McNab [1997](#page-14-3); McKechnie [2008](#page-14-0)). Because BMR is the lowest measurable rate of energy expenditure in a resting animal, it is widely considered as a measure of physiological maintenance costs. The strict definition of BMR also includes only non-growing, nonreproductive individuals (Benedict [1938;](#page-12-0) Kleiber [1961](#page-14-2); Aschoff and Pohl [1970;](#page-12-2) McNab [1997\)](#page-14-3), as growth and reproduction have energetic costs and thereby elevate metabolic rate above minimal levels (e.g., Vézina and Salvante [2010](#page-16-1); Careau et al. [2013a](#page-12-3)). The conditions necessary to standardize metabolic measurements to conform with the definition of BMR are generally achievable only in laboratory studies, and likely do not reflect conditions that animals experience in their natural environment on a consistent basis. These standardized conditions do, however, allow for studying flexibility of energetic traits within individuals, with BMR varying with season (cold), migration, and other aspects of the annual cycle (reviewed by McKechnie [2008](#page-14-0); McKechnie and Swanson [2010](#page-14-1); Swanson [2010](#page-16-2)). We also suggest that growing and reproductive phases of life cycles similarly have minimum energetic costs and we suggest that these can be considered as BMR for these phases, thereby facilitating intraspecific comparisons of minimum metabolic costs among different phases of the annual cycle. Furthermore, dichotomous categorizations of reproductive and growth states are often difficult in practice, particularly for tropical and subtropical species in which the phenologies of breeding, growth, and processes, such as molt, are not strongly seasonal. Nevertheless, it remains critical that interspecific comparisons are based on data sets, where the reproductive and growth status of individuals are standardized as far as possible, which will usually require the exclusion of data collected during breeding seasons or periods of ontogenetic growth to avoid confounding variables. For this paper, we interpret BMR as the minimum maintenance energetic costs for any given stage of the life cycle of an organism, with the caveat that life stages (e.g., breeding vs. non-breeding, molting vs. non-molting, growing vs. nongrowing) may sometimes be difficult to precisely delineate.

Correlations of BMR with other metabolic traits

Adaptive explanations for a high BMR are pervasive despite evidence that natural selection acts on field-active metabolic rates or maximal metabolic capacities (Hayes and O'Connor [1999](#page-13-1); Rezende et al. [2004;](#page-15-1) Sears et al. [2006](#page-16-3); Boratyński and Koteja [2009](#page-12-4); Clavijo-Baquet and Bozinovic [2012](#page-13-2); Zub et al. [2014](#page-17-0); Pauli et al. [2016](#page-15-2); Petit et al. [2017](#page-15-3)) rather than on BMR directly. Raichlen et al. ([2010\)](#page-15-4) articulated this idea, suggesting that "there may be adaptive reasons for relatively high or low BMR at a given body mass" (p. 302). Comparative studies of endotherm energetics often measure BMR as a proxy for overall energy turnover (more accurately defined by daily energy expenditure, DEE, or some similar metric), and then infer potential adaptive differences in energetics from patterns of variation in BMR. To complicate this issue further, evidence for the inferred link between BMR and DEE is inconsistent among studies (e.g., Ricklefs et al. [1996](#page-15-5); White and Seymour [2004](#page-16-4); Speakman et al. [2004;](#page-16-5) Speakman and Król [2011](#page-16-6); Careau et al. [2013b](#page-12-5); Welcker et al. [2015;](#page-16-7) Portugal et al. [2016\)](#page-15-6), and, moreover, is mechanistically difficult to reconcile with the observations that different tissues are largely responsible for variation in BMR and DEE (Konarzewski and Diamond [1995](#page-14-4); Chappell et al. [1999;](#page-13-3) Selman et al. [2001](#page-16-8); Książek et al. [2009;](#page-14-5) Bacigalupe et al. [2010](#page-12-6); but see; Green et al. [2013\)](#page-13-4). Furthermore, the tissues influencing variation in BMR and DEE may vary from one life-history stage to another (Vézina and Williams [2003;](#page-16-9) Vézina et al. [2009](#page-16-10)). Similarly, evidence for phenotypic correlations between basal and maximal aerobic metabolic rates (either thermogenic or exercise maximal rates) is also often inconsistent. Such studies typically find positive correlations at the interspecific level (e.g., Hayes and Garland [1995](#page-13-5); Dutenhoffer and Swanson [1996](#page-13-6); Rezende et al. [2002](#page-15-7), [2004](#page-15-1); White and Seymour [2004](#page-16-4); but see; Clavijo-Baquet and Bozinovic [2012](#page-13-2)), but variable relationships within species, with either positive correlations, positive correlations for raw, but not body mass-corrected values, or non-significant correlations (Boily [2002;](#page-12-7) Sadowska et al. [2005;](#page-16-11) Vézina et al. [2006](#page-16-12); Gębczyński and Konarzewski [2009;](#page-13-7) Swanson et al. [2012\)](#page-16-13). Speakman et al. [\(2004](#page-16-5)) also reviewed the functional significance of individual variation in BMR, examining the occurrence of high BMR as a means to support increased energy intake rates and sustained metabolic rates (SusMR; central limits hypothesis) or increased sustained activity levels (peripheral limits hypothesis), concluding that between-state comparisons (e.g., non-reproductive vs. lactating mice) of BMR were generally consistent with links between BMR and sustained maximal metabolic rates, but that these relationships disappeared at the individual level, at least after accounting for variation in body mass. The reasons for the differences between interspecific and intraspecific studies of these phenotypic correlations have not yet been resolved, but may involve different selective forces acting on BMR and DEE or MMR (maximal metabolic rate; see below), or may represent a statistical artifact associated with much lower levels of variation in body masses and metabolic rates for intraspecific than for interspecies studies. Supporting the idea of a phenotypic link between maximal and basal metabolic rates for interspecific comparisons, factorial aerobic scope (i.e., MMR/ BMR) tends to remain fairly constant across vertebrate groups (Killen et al. [2016](#page-14-6); Nespolo et al. [2017](#page-15-8)). For these reasons, we urge caution in developing adaptive explanations for BMR, either alone or as a proxy for overall endotherm energetics.

If BMR evolves as a correlated response to selection acting on other metabolic traits, such as DEE or maximal metabolic capacities for exercise or thermogenesis (summit metabolic rate, M_{sum}), then these traits should be heritable and the phenotypic correlation between them should also reflect underlying genetic correlations. A number of studies have used quantitative genetic analyses to examine such correlations in captive animals originating from both long-term laboratory-bred and wild populations, as well as narrow sense heritability $(h^2$ = the ratio of the additive genetic variance to the total phenotypic variance) of metabolic traits. In a review of heritabilities of metabolic traits in mammals, Konarze-wski et al. ([2005](#page-14-7)) found that support for significant heritability of metabolic traits was equivocal, with some studies showing very low and non-significant heritability, but others showing high and significant heritabilities. Subsequent mammalian studies have also supported this conclusion, as some show significant heritabilities of metabolic traits (Sadowska et al. [2005;](#page-16-11) Nespolo et al. [2005;](#page-15-9) Wone et al. [2009;](#page-17-1) Boratyński et al. [2013](#page-12-8)), but others do not (Nespolo et al. [2005;](#page-15-9) Boratyński et al. [2013](#page-12-8)). There are relatively few studies of heritability of avian metabolic traits, but Rønning et al. ([2007\)](#page-15-10) and Tieleman et al. [\(2009](#page-16-14)) found significant heritability for BMR in zebra finches (*Taeniopygia guttata*) and stonechats (*Saxicola* spp.), respectively. Moreover, heritabilities may differ between BMR and other metabolic traits (exercise MMR, M_{sum} , SusMR) (Dohm et al. [2001](#page-13-8); Bacigalupe et al. [2004](#page-12-9); Nespolo et al. [2005](#page-15-9)).

Positive genetic correlations of BMR with other metabolic traits, such as exercise MMR, M_{sum} , or SusMR, or with other traits (e.g., locomotor movements, growth rates, and litter size) potentially related to metabolic capacities, occur in some endotherms (Dohm et al. [2001](#page-13-8); Sadowska et al. [2005,](#page-16-11) [2009;](#page-16-15) Wone et al. [2009](#page-17-1), [2015](#page-17-2); Careau et al. [2011;](#page-12-10) Boratyński et al. [2013\)](#page-12-8). In contrast, non-significant correlations of BMR with other metabolic traits also occur in endotherms (Książek et al. [2004](#page-14-8); Nespolo et al. [2005](#page-15-9); Sadowska et al. [2005](#page-16-11); Gębczyński and Konarzewski [2009\)](#page-13-7). Thus, although these studies collectively provide some support for the idea that metabolic traits are heritable and that BMR and other metabolic traits are genetically correlated, support for this hypothesis is far from universal. Further studies are needed to illuminate the genetic architecture underlying metabolic traits.

BMR and thermal conditions

Many endotherms, especially small species inhabiting temperate or polar climates, have lower critical temperatures (T_{lc} = the temperature below which regulatory thermogenesis must occur to balance heat loss, and, therefore, the lower limit of the thermoneutral zone) that are substantially higher than the environmental temperatures they typically encounter (Rubner [1883](#page-16-16); Scholander et al. [1950b](#page-16-17); Schmidt-Nielsen [1984](#page-16-18); Porter and Kearney [2009](#page-15-11)). As a consequence, these animals should almost always be outside of their thermoneutral zones under natural conditions (Fig. [1\)](#page-2-0), so rarely does metabolic rate in the wild reach levels approximating BMR. However, we emphasize that few data directly address the proportion of time endotherms spent inside or outside of their thermoneutral zones in natural thermal environments, which encompass all avenues of heat exchange between the animal and the environment (but see Liwanag et al. [2009\)](#page-14-9). Behavioral mechanisms, such as basking or huddling, may mollify thermal demands to some extent (Maldonado et al. [2009\)](#page-14-10). Nevertheless, small endotherms likely still operate outside of their thermoneutral zones under most natural environmental conditions, although even very small species can substantially modify roost temperatures by occupying thermally buffered microsites and/or using communal roosting (see Walsberg [1990](#page-16-19) for an example). Larger endotherms, because of their lower surface area to volume ratios, thicker pelages

Fig. 1 Relationship between metabolic rate (*solid line* measured as oxygen consumption) and environmental operative temperature for a hypothetical small endotherm from a temperate-zone environment. The lower critical temperature (T_{lc}) , below which regulatory thermogenesis is required, occurs at 23 °C. The *dashed line* represents the hypothetical distribution of operative temperatures encountered by this animal over the entire annual cycle. Note that only a small percentage of these temperatures (filled area of the *curve*) occur within the thermoneutral zone of this animal

or plumages, and consequent lower thermal conductances, more often encounter thermal conditions within their thermoneutral zones, so metabolic rates may more frequently approach BMR compared to the situation in smaller endotherms (Rubner [1883;](#page-16-16) Schmidt-Nielsen [1984;](#page-16-18) Porter and Kearney [2009;](#page-15-11) Liwanag et al. [2009\)](#page-14-9). If endotherms only rarely experience thermoneutral conditions, it is difficult to envision natural selection acting directly on BMR, which by definition is associated with these temperatures. Activity also increases metabolic rates above BMR, with DEE usually averaging two to fourfold higher than BMR (Ricklefs et al. [1996;](#page-15-5) Piersma and van Gils [2011;](#page-15-12) Careau et al. [2013b](#page-12-5)). Therefore, because animals are most often not simply "idling" during their activity periods, it again becomes difficult to envision selection acting directly on BMR under conditions necessitating metabolic rates elevated above basal levels.

These issues raise a fundamental question pertinent to comparative energetic studies of endotherms: Does BMR, especially a high BMR, possess direct adaptive value? Dawson and Marsh ([1989\)](#page-13-9) and Dawson and O'Connor [\(1996](#page-13-10)) raised this issue directly in considering BMR flexibility in small birds wintering in cold climates, stating that the "… adaptive value [of a high BMR] … is not immediately obvious" and "… at first glance … [a high BMR is] a needless encumbrance," respectively. Rezende and Bacigalupe [\(2015](#page-15-13), p. 719) echoed this sentiment, noting that for endotherms, in general, "the adaptive significance of higher BMR per se remains unclear."

Selective forces acting on metabolic rates

Burton et al. ([2011\)](#page-12-11) developed three hypotheses explaining the persistence of variable BMR in natural populations, which collectively provide adaptive explanations for low, high, and variable BMR. The compensation hypothesis posits that low BMR increases fitness, because individuals can devote more energy to processes other than maintenance (e.g., growth and reproduction). The increased intake hypothesis suggests that high BMR increases fitness, because individuals will have larger internal organs capable of supporting higher activity or thermoregulatory capacities. Finally, the context-dependent hypothesis states that interactions between resource availability, environmental conditions, and BMR are such that high BMR is favored in some conditions and low BMR is favored in others. This latter hypothesis is akin to a stabilizing selection hypothesis (Nespolo et al. [2017](#page-15-8)), where fluctuating environmental conditions favor high metabolic capacities (and thereby high maintenance costs) in some years and low metabolic capacities others (Nilsson and Nilsson [2016](#page-15-14)). Such fluctuating selection might produce an optimum BMR which functions

well over a wide range of environmental and ecological conditions. Although these hypotheses do help explain why BMR variation might persist among populations, we argue here that natural selection acts directly on BMR only to reduce it. Such a selective pattern is consistent with predictions of the compensation hypothesis and the low end of the context-dependent hypothesis, particularly for conditions, where selection on activity or thermoregulatory capacities is relaxed. For the increased energy intake and the high end of the context-dependent hypotheses, we argue that natural selection is operating directly on other traits (e.g., activity or thermoregulatory capacities) and that BMR merely tags along as a correlated response. Thus, our hypothesis is that, from an energetic standpoint, natural selection should always act to minimize BMR while still meeting maintenance requirements for a given set of environmental or ecological demands.

Is a high BMR adaptive?

A few scenarios exist where a high BMR has been hypothesized to have direct adaptive value. These include a reduction in the lower critical temperature, protection of the tissues against cold temperatures, or a correlated response resulting in an increased general metabolic level (Dawson and Marsh [1989;](#page-13-9) Dawson and O'Connor [1996;](#page-13-10) Fristoe et al. [2015](#page-13-0)). In the first scenario, the argument is that an elevation in BMR would reduce T_{lc} so that regulatory thermogenesis is postponed until lower temperatures (e.g., Fristoe et al. [2015\)](#page-13-0). From an energetic standpoint, this argument fails, because energy is merely redistributed to BMR (i.e., maintenance costs) rather than to regulatory thermogenesis. Whether metabolic rate is elevated due to higher BMR, regulatory thermogenesis, or other factors that may substitute for thermoregulatory heat production, such as activity or the heat increment of feeding (e.g., Hindle et al. [2003](#page-13-11); Enstipp et al. [2008;](#page-13-12) Humphries and Careau [2011\)](#page-13-13), the overall energetic cost of maintaining body temperature would remain the same. A higher BMR and subsequent reductions in T_{lc} would delay the onset of shivering, but active shivering, even at relatively high intensities, does not seem to be injurious to muscle (Swanson and Thomas [2007](#page-16-20)), so we see little selective advantage to this strategy. In theory, if the upper limit to thermogenesis (summit metabolism, M_{sum}) was a fixed, invariant multiple of BMR, and conductance remained constant, then increases in BMR could directly lead to improved cold tolerance by reducing the minimum environmental temperature at which a body temperature setpoint can be regulated. However, empirical studies of seasonal changes in BMR, M_{sum} and metabolic expansibility (i.e., M_{sum}/BMR) do not support the idea of metabolic expansibility remaining sufficiently constant for BMR to directly influence cold tolerance in this way (Swanson [2010](#page-16-2); Petit et al. [2013](#page-15-15)). Finally, the adaptive advantage of such a scenario still directly results from an increase in *M*_{sum}, rather than BMR.

The protective response to cold idea is also not a particularly appealing explanation, as such protection is precisely why regulatory thermogenesis exists—to increase metabolic heat production in the face of declining temperatures to maintain a high, stable body temperature. This leaves us with the correlated response suggestion, where selection acts to increase BMR with a correlated increase in general metabolic level. We argue, however, that the reverse, selection acting on maximal or sustained maximal capacities for activity or thermoregulation, with BMR increasing as a correlated response, is far more likely than selection to increase BMR, with a correlated response in metabolic capacities. Indeed, the former scenario was the basis for the aerobic capacity model for the evolution of endothermy (Bennett and Ruben [1979](#page-12-12)). In such a scenario, BMR is not a direct target of selection. Thus, none of these ideas for why natural selection should act to increase BMR directly seems obviously adaptive.

Potential adaptive explanations for a high BMR

To further explore this issue, we will examine some environmental or ecological conditions, where BMR is typically high and consider the selective factors that could be driving these high levels of BMR. Environmental temperature is a strong predictor of BMR, with endotherms living in cold environments typically having higher BMR than those living in warm climates (White and Seymour [2004](#page-16-4); Wiersma et al. [2007;](#page-17-3) Jetz et al. [2008](#page-13-14); Naya et al. [2013](#page-15-16); Fristoe et al. [2015](#page-13-0); Luna et al. [2017](#page-14-11); but see also; Londoño et al. [2015](#page-14-12)). In addition, BMR is often elevated in winter relative to summer for birds wintering in cold climates or for cold acclimated relative to warm-acclimated birds (McKechnie [2008](#page-14-0); McKechnie and Swanson [2010](#page-14-1)). This pattern of winter or cold-induced increments in BMR is also characteristic of several winter-active small mammals (e.g., Heldmaier and Steinlechner [1981](#page-13-15); Li and Wang [2005](#page-14-13)) and of intermediate-sized (100–1000 g) mammals in general (Lovegrove [2005\)](#page-14-14). In contrast, for endotherms from tropical or subtropical climates, lesser seasonal temperature variation and warmer winter temperatures than in coldtemperate and polar climates result in low energy demands in winter. Under such conditions, energetic demands during summer associated with reproductive workloads are greater than in winter, so BMR may be greater in summer than in winter if BMR is positively correlated with energy demands (Wells and Schaeffer [2012;](#page-16-21) McKechnie et al. [2015](#page-14-15)). Moreover, subtropical sites may show differential seasonal patterns of temperature variation, with some sites showing colder winters than summers and some sites with less seasonal variation (van de Ven et al. [2013](#page-16-22); McKechnie et al. [2015;](#page-14-15) Noakes et al. [2017\)](#page-15-17). Consequently, seasonal patterns of thermoregulatory costs may vary among subtropical sites, which may lead to variation in seasonal metabolic patterns (McKechnie et al. [2015](#page-14-15); Noakes et al. [2017](#page-15-17)). In addition, lower latitudes may present a greater diversity of metabolic niches (Londoño et al. [2015](#page-14-12)), whereas higher latitudes with cold winters produce much greater thermoregulatory demands in winter than in summer, thereby producing common seasonal patterns of metabolic variation, with higher metabolic rates in winter than in summer (Swanson [2010](#page-16-2)). Londoño et al. [\(2015](#page-14-12)) also found no correlation between temperature and BMR in a large comparative study of birds from the New World tropics, suggesting that tropical birds are more variable in their responses to temperature than birds from higher latitudes.

Identifying latitudinal patterns in the seasonal BMR adjustments of mammals is significantly complicated by the more widespread occurrence of torpor and hibernation in mammals compared to birds. The most recent analysis of which we are aware (Lovegrove [2005\)](#page-14-14) revealed that the magnitude and direction of mammalian seasonal metabolic adjustments are strongly size-dependent. Among small mammals (<100 g), metabolic rates generally decrease in winter, a process driven both by body mass reductions as well as reductions in mass-specific BMR. In the Lovegrove [\(2005](#page-14-14)) analysis, neither winter body mass nor BMR reductions were correlated with latitude and predictions concerning the possible existence of latitudinal gradients in the variability of seasonal BMR adjustments were not tested. However, none of the figures in this paper give any cause to suspect a similar trend as occurs in birds, whereby patterns of seasonal adjustments are more variable in subtropical or tropical regions compared to high latitudes (McKechnie et al. [2015\)](#page-14-15).

For species wintering in cold climates, the winteror cold-induced increases in BMR are accompanied by increases in thermogenic capacity via shivering and nonshivering thermogenesis. In fact, for both mammals and birds, such cold-induced increases in thermogenic capacity are more commonly detected than increases in BMR (see Rezende et al. [2004;](#page-15-1) McKechnie and Swanson [2010;](#page-14-1) Swanson [2010](#page-16-2) for reviews). Thus, the strong effect of cold temperatures on endotherm BMR appears mediated by thermoregulatory demands, with selection acting to increase thermogenic capacity, and BMR increasing as a correlated response to support life in the cold. One exception to this general rule, however, involves mammals wintering in cold climates with scarce food resources, many of which enter torpor and/or hibernation. Such mammals may differ from other endotherms in seasonal patterns of cold acclimation or winter acclimatization, as they often show a reduction in BMR, which is usually interpreted as a mechanism of energy conservation at a time when food resources are low (e.g., Geiser and Baudinette [1987](#page-13-16); Boratyński et al. [2016](#page-12-13)). Birds also use torpor, but the mean latitude for birds using daily torpor is 23°, with a 95% confidence interval extending from 19° to 27° (Ruf and Geiser [2015\)](#page-16-23). Thus, heterothermic birds generally do not inhabit very cold winter climates, at least compared to heterothermic mammals, so selection for winter energy conservation is likely lower than for mammals wintering in cold climates with marked reductions in resource availability. Interestingly, minimum environmental temperatures are negatively correlated with M_{sum} , but not with BMR, in heterothermic rodents, suggesting a decoupling of any phenotypic correlation between M_{sum} and BMR in these rodents where selection forces may act to conserve energy during torpor but maintain high *M*_{sum} for arousal during inter-torpor bouts (Careau [2013](#page-12-14)). Such decoupling may be mediated through large contributions to arousal metabolic rates of non-shivering thermogenesis by brown adipose tissue by torpid rodents, rather than primary reliance on muscular shivering (e.g., Jefimow et al. [2004;](#page-13-17) Oelkrug et al. [2011\)](#page-15-18).

If selection acts directly to increase BMR under conditions of high energy demand, such as thermoregulation in cold environments, then BMR should be correlated with fitness components to a greater degree than other metabolic traits. The relationship of BMR to fitness has been examined for several birds and mammals, but correlations between BMR and reproductive output or survival are variable, with positive, negative, or no correlations detected (see Rønning et al. [2016](#page-15-19) for review). Such variability could result from non-directional (e.g., stabilizing, fluctuating, and disruptive) selection (Nilsson and Nilsson [2016](#page-15-14); Rønning et al. [2016](#page-15-19)). Alternatively, variability in the direction and strength of correlations between BMR and fitness could result from selection acting directly on other physiological traits, such as DEE or activity or thermogenic capacities, rather than BMR, as we argue here. In this case, correlations between BMR and fitness traits may differ from the corresponding correlations for physiological traits under direct selection. In support of the idea that selection acts directly to promote phenotypic flexibility of thermogenic performance in cold environments, rather than on BMR, Petit et al. [\(2017](#page-15-3)) found that black-capped chickadees from eastern Canada with a winter M_{sum} above a threshold level showed improved winter survival, whereas BMR variation did not similarly impact survival (Fig. [2](#page-5-0); but see Nilsson and Nilsson [2016](#page-15-14)). In this specific system, massindependent M_{sum} increased by 34% between summer and winter, but mass-independent BMR only changed by 6%, and variations in M_{sum} and BMR were not temporally matched (Petit et al. [2013](#page-15-15)). These results suggest that these traits could reflect different body components

Fig. 2 Winter survival probability of *black*-capped chickadees (*Poecile atricapillus*) as a function of BMR (*top*) and M_{sum} (*bottom*). BMR was not significantly related to overwinter survival, but M_{sum} was significantly and asymptotically associated with survival, with the asymptote occurring above 1.4 W. BMR values are corrected for the effect of body mass, while M_{sum} values are corrected for the effects of date of capture and measurement and body mass (see Petit et al. [2017](#page-15-3) for details)

responding to different demands associated with the winter season. Barceló et al. ([2017](#page-12-15)) subsequently tested the uncoupling of BMR and M_{sum} in captive white-throated sparrows (*Zonotrichia albicollis*) using a combination of experimental diets and temperature treatments. Both BMR (15%) and M_{sum} (19%) increased in the cold (−8 °C) relative to the warm (28 °C) treatment. However, although both traits varied with lean body mass and were correlated, the components of lean mass that exerted the greatest influence on BMR variation were the digestive and excretory organs through the influence on food intake. In this specific experiment, temperature did not influence muscle size per se, but cold acclimated individuals with larger hearts and skeletal muscles also had higher thermogenic capacity. These observations, therefore, support the thesis that BMR reflects support functions (energy acquisition) in a thermal acclimation context and that fitness consequences stem primarily from direct selection on other metabolic traits.

Another factor often correlated with BMR is personality or social status, which in turn may be related to "pace of life" along a fast-slow continuum. A fast pace of life is characterized by high reproductive output, high activity, and relatively short life spans. These features tend to be positively correlated with BMR, such that a high BMR is an associated characteristic of the fast lifestyle (e.g., Wiersma et al. [2007](#page-17-3); Williams et al. [2010;](#page-17-4) Versteegh et al. [2012;](#page-16-24) but see; Speakman [2005](#page-16-25)). Regarding personality traits, high levels of exploratory behaviors and social dominance are typical of fast lifestyles and are often (Car-eau et al. [2008](#page-12-16); Møller [2009](#page-15-20); Careau and Garland [2012](#page-12-17); Maldonado et al. [2012](#page-14-16); Mathot et al. [2015](#page-14-17)), but not always (Lewden et al. [2012\)](#page-14-18), positively correlated with BMR. Behaviors and activities associated with fast lifestyles may involve higher support costs, such as larger reproductive or other organs (Wiersma et al. [2012\)](#page-17-5), which, in turn, may result in higher BMR relative to individuals on the slow end of the pace of life continuum. Therefore, a high BMR would allow a fast pace of life, which may be adaptive in variable environments by allowing rapid exploitation of resources when they become available. As we see it, however, these arguments are ambiguous with respect to causation—does high BMR drive a fast pace of life or does a fast pace of life result in a high BMR (e.g., Glazier [2015](#page-13-18))? We suspect that the latter scenario is more likely, as natural selection should act directly on personality traits that may relate to fitness, such as aggression or exploratory behavior, with BMR reflecting higher maintenance costs associated with the exploitative lifestyle (Réale et al. [2010\)](#page-15-21).

Potential adaptive explanations for a low BMR

We argue here that, as a general rule, natural selection does not act directly to increase BMR. Because BMR typically comprises about 30–50% of total energy expenditure for endotherms (Raichlen et al. [2010;](#page-15-4) Burton et al. [2011\)](#page-12-11), it represents a significant component of the overall energy budget. Energetically, it would, therefore, make sense to reduce BMR to the lowest level possible, while still effectively supporting overall metabolic demands, to minimize energy expended on maintenance functions. As such, cases where BMR does seem directly adaptive are scenarios where selection acts to reduce BMR. These include scenarios where animals live in warm climates or scenarios involving conditions that promote energy (or water) conservation, such as hot deserts, sedentary lifestyles, low food resources, torpor use, or low-risk behavioral strategies (Møller [2009](#page-15-20); McNab [2012;](#page-14-19) Pauli et al. [2016\)](#page-15-2). All of these cases should favor maintaining maintenance energy expenditures, and, therefore, BMR, at the lowest feasible levels. To examine this idea further, we will briefly review examples of where selection on high energy demands is relaxed, which could allow direct selection for reduction of BMR.

Food habits are related to BMR variation in endotherms (McNab [1988](#page-14-20), [2008;](#page-14-21) Cruz-Neto and Bozinovic [2004](#page-13-19); Bozinovic and Sabat [2010;](#page-12-18) Sabat et al. [2010](#page-16-26); Naya et al. [2013\)](#page-15-16). The general mechanistic explanation for this relationship is that animals on low-energy diets adopt an energy conserving lifestyle, including low BMR, to deal with low dietary energy intake or low availability of food resources (see McNab [2008](#page-14-21); Bozinovic and Sabat [2010](#page-12-18) for reviews). Endotherms with more abundant, better-quality, and/or more predictable food can adopt a more energetically expensive lifestyle, since they are less constrained by the requirement for energy conservation. Again, however, we emphasize that direct selection on BMR occurs only to drive BMR downward in the low resource situation. When food is readily available, we contend that selection likely acts on field metabolic rates, activity, or thermogenic capacities, with BMR increasing as a correlated response.

Related to both abiotic factors (e.g., aridity) and food resources, environmental productivity has also been postulated to influence BMR. This correlation is thought to be driven by reduced food availability in environments where productivity is low, thereby promoting an energy-conservation strategy for lifestyle and energetics (Bozinovic et al. [2009](#page-12-19)). The well-known trend of lower BMR in desert endotherms than in those from more mesic environments (e.g., Lovegrove [2003](#page-14-22); Tieleman et al. [2003;](#page-16-27) McNab [2012\)](#page-14-19) supports this idea. Reduced maintenance energetic costs also have the added benefit of reducing respiratory evaporative water loss and, thereby, water requirements in arid environments, which is especially advantageous in hot deserts where evaporative cooling for thermoregulation is particularly crucial (McNab [2012](#page-14-19)). Indeed, rodents from hot deserts tend to have lower BMR than allometrically predicted, but rodents from cold deserts tend to have higher BMR than allometrically predicted (Rezende et al. [2004](#page-15-1)). Camelids also have a low BMR for mammals and are associated with arid environments with low food resources (Dittmann et al. [2014\)](#page-13-20). Positive correlations between net primary productivity and BMR occur for several intraspecific studies of endotherms (Mueller and Diamond [2001](#page-15-22); Bozinovic et al. [2009\)](#page-12-19). Nevertheless, net primary productivity is often not a prominent factor associated with high BMR in broad comparative multivariate analyses (White et al. [2007](#page-16-28); Jetz et al. [2008](#page-13-14); Naya et al. [2013;](#page-15-16) Luna et al. [2017](#page-14-11); but see; Tieleman et al. [2003](#page-16-27); Rezende et al. [2004](#page-15-1)). In these studies, variation in abiotic factors, especially temperature, tends to swamp effects of habitat productivity or precipitation on BMR, suggesting that temperature acting

on thermoregulatory capacities and their associated support costs is generally a more important driver of metabolic variation. The trends of low BMR in desert environments, where energy conservation is a priority, and the primary importance of temperature as a driver of metabolic variation, with BMR elevated in the cold in support of higher energy requirements, collectively fit the idea that natural selection serves to drive BMR to the lowest levels that environmental or ecological circumstances permit.

Another factor potentially selecting for a low BMR is the heat produced during daily activities, for example, the metabolic heat production associated with breeding activities in warm climates, which must be dissipated. This factor forms the basis for the heat dissipation limit hypothesis for upper limits to energy expenditure in endotherms (Speakman and Król [2010](#page-16-29)). Under many (perhaps most) environmental conditions, with the possible exception of small endotherms breeding in cold climates, a downregulation of maintenance functions during activity would be selectively advantageous, as it would reduce the overall heat load needing to be dissipated. For small endotherms in cold climates, a high BMR might be required to support energetically expensive breeding activities or thermoregulation, but if energy availability is not limiting, selection to reduce BMR to lower the amount of heat dissipation required may be relaxed. Even under these conditions, however, a high BMR would necessitate increased energy intake and, thereby, increased foraging efforts, which might expose animals to increased ecological risks from predation or competition. Thus, even under conditions when energy is not limiting, we argue that selection may be expected to reduce BMR to the lowest level possible.

Some of the most convincing evidence for selection for low BMR and consequent energy conservation comes from endothermic taxa in which the capacity of individuals to effectively thermoregulate in the absence of thermally buffered microsites and/or communal roosting with conspecifics has apparently been lost. The classic mammalian example is the naked mole rat (*Heterocephalus glaber*, Buffenstein and Yahav [1991](#page-12-20)). This species lives in subterranean burrows with warm stable temperatures and practices behaviors, such as huddling, that limit thermoregulatory demands (Yahav and Buffenstein [1991\)](#page-17-6). Consequently, thermogenic ability is downregulated along with maintenance costs (Hislop and Buffenstein [1994](#page-13-21); Woodley and Buffenstein [2002](#page-17-7)). A broadly analogous avian example involves mousebirds (Coliiformes). Mousebirds are highly sociable birds that exhibit well-developed communal roosting behavior in which groups form tight clusters while clinging to branches. In addition to leading to substantial energy savings (Prinzinger [1988;](#page-15-23) McKechnie and Lovegrove [2001](#page-14-23)), the clustering behavior of mousebirds provides the basis for the defense of a normothermic body temperature setpoint; individuals prevented from clustering with conspecifics exhibit labile body temperature that decreases during the night (McKechnie and Lovegrove [2001;](#page-14-23) McKechnie et al. [2006\)](#page-14-24). Moreover, the fact that mousebirds have long been viewed as a highly heterothermic taxon that routinely uses torpor (Bartholomew and Trost [1970;](#page-12-21) Prinzinger et al. [1981](#page-15-24); Finke et al. [1995\)](#page-13-22) appears to largely represent an artifact of studies investigating thermoregulation in individual birds without the presence of conspecifics. Data collected under laboratory and free-ranging conditions suggest that torpor is very rare in clustering mousebirds (McKechnie and Lovegrove [2001](#page-14-23); McKechnie et al. [2004](#page-14-25), [2006\)](#page-14-24). Although McKechnie and Lovegrove [\(2001](#page-14-23)) argued that the thermoregulatory characteristics of mousebirds reflect a plesiomorphic trait linked to their ancient phylogenetic position, this argument was based on the placement of the Coliiformes in the phylogeny proposed by Sibley and Ahlquist ([1990\)](#page-16-30). More recent work suggests that the mousebirds belong to a more-recently derived clade (Hackett et al. [2008;](#page-13-23) Prum et al. [2015](#page-15-25)), supporting the idea that the thermal physiology of this group arose from selection for energy conservation acting on maintenance energy costs.

Functional correlates of BMR

We are proposing here that selection acts generally to reduce BMR, which begs the question of the mechanisms by which such a reduction might occur. In the following, we review several mechanisms potentially available to endotherms for mediating variation in BMR.

Body size and BMR

Body size is the greatest determinant of metabolic rate variation in animals (Schmidt-Nielsen [1984;](#page-16-18) McNab [2012](#page-14-19)). Consequently, adjustment of body size represents one potential mechanism for adjustment of BMR to varying environmental or ecological demands, although variation in body size will have opposite effects on mass-specific and whole-organism metabolic rates. A decrease in body size, for example, will result in a reduction in overall metabolic costs, but an increase in mass-specific metabolic rates, so selection could act on either whole-organism or mass-specific rates, with different selective forces potentially focusing on one or the other of these metabolic traits. One example of how body size might be adjusted to environmental demands is Bergmann's rule. Bergmann's rule codifies a general relationship between body size and climate for endotherms, with body size expected to increase in colder climates as a function of lower surface areas for heat loss (Bergmann [1847\)](#page-12-22). Whether Bergmann's rule is valid for endotherms is a contentious issue, with some studies of endotherms supporting the rule (e.g., Ashton et al. [2000;](#page-12-23) Ashton [2002](#page-12-24); Meiri and Dayan [2003](#page-14-26); Blackburn and Hawkins [2004](#page-12-25); Olson et al. [2009](#page-15-26)), but others not (e.g., Geist [1987](#page-13-24); Fristoe et al. [2015\)](#page-13-0), and some studies finding support in cold climates, but not in warmer climates (Rodríguez et al. [2006,](#page-15-27) [2008\)](#page-15-28). Nevertheless, the general reasoning behind Bergmann's rule serves as the basis for recent suggestions that reductions in body size may serve as a general response of endotherms and other vertebrate animals to climate warming (Gardner et al. [2011;](#page-13-25) Sheridan and Bickford [2011;](#page-16-31) Goodman et al. [2012](#page-13-26)). Thus, adjustment of body size in response to changing environmental or ecological demands may be one mechanism for adaptive adjustment of BMR, although such adjustment is likely to be accompanied by variation in body composition (e.g., Piersma and van Gils [2011\)](#page-15-12) and the direction of adjustment may vary depending on the selective forces involved.

Body temperature and BMR

Another potential mechanism for adjusting BMR is to increase or decrease body temperature (T_b) . Because resting metabolic rates in endotherms have Q_{10} values (i.e., the change in metabolic rate for a 10^oC change in T_b) ranging from about 2–3 (Clarke et al. [2010](#page-13-27)), a relatively small change in the T_b setpoint could impact metabolic rates. As such, a reduction in normothermic T_b might serve as an effective mechanism to reduce BMR generally. Substantial variation in T_b occurs among different endotherm taxa; among extant mammals, T_b varies by approximately 10^oC (Lovegrove [2012](#page-14-27)). Clarke and Rothery ([2008\)](#page-13-28) argued that T_b may be influenced by ecology but constrained by physiology, suggesting that selection may adjust BMR through T_b variation or, alternatively, that T_b differences are a consequence of selection on BMR or other metabolic traits. Because metabolic rates during thermoregulation or activity are equivalent to maintenance costs plus an increment due to thermoregulatory or activity costs, higher maintenance costs would elevate the metabolic rate for thermoregulation or activity generally if costs for these actions were purely additive (e.g., no substitution of heat produced by BMR for thermoregulation). Under such conditions, if thermal conductance and environmental temperatures were kept constant, the higher BMR should result in a higher T_b , so an elevated BMR could help to maintain elevated T_b (as suggested by the thermoregulatory model for the evolution of endothermy; Crompton et al. [1978;](#page-13-29) McNab [1978](#page-14-28)). For example, Clarke et al. (2010) (2010) modeled T_b in mammals as functions of body mass and average ambient temperature (T_a) within the animals' range and found that T_a was inversely related to T_b , such that T_b increased by 1 °C for every 13 °C decrease in temperature. Thus, a polar animal

at an average annual T_a of -13° C would be predicted to have a T_b 2.7 °C higher than that of a tropical mammal living at an average annual T_a of 25 °C (Clarke et al. [2010](#page-13-27)). Such an increase in T_b should result in higher BMR for endotherms from cold climates than from warm climates, which is indeed the case (e.g., Weathers [1979;](#page-16-32) Rezende et al. [2004;](#page-15-1) Wiersma et al. [2007](#page-17-3)), although the precise contribution of T_b variation to the relationship of BMR to climate in endotherms is not known. From an energetic standpoint, however, the level of BMR should not affect the overall cost for thermoregulation, because absolute heat production is the same whether it is BMR or regulatory thermogenesis that results in the higher metabolic heat production. Thus, from an energetic standpoint, increasing T_b to elevate BMR for thermoregulation seems unnecessary.

Organ masses and BMR

BMR is primarily thought to be functionally derived from the metabolism of central organs and brain, with high maintenance activities at rest reflecting costs for protein turnover, gluconeogenesis, ion pumping, enzyme activities, nitrogenous waste synthesis, and mitochondrial membrane proton transport (Rolfe and Brown [1997;](#page-15-29) Burton et al. [2011](#page-12-11); Nespolo et al. [2011\)](#page-15-30). Elia ([1992\)](#page-13-30) estimated contributions of various organs and tissues to BMR for humans, with central organs (liver, kidney, heart, and "other," including the gut) accounting for 56%, brain accounting for 20% and skeletal muscles accounting for 16–22% of organismal BMR. Rolfe and Brown ([1997\)](#page-15-29) also estimated contribution of organs and tissues to BMR for humans, with central organs (liver, gut, kidney, heart, and lung) accounting for 48%, brain for 20%, and skeletal muscles for 20% of organismal BMR. Similar values for rats (*Rattus norvegicus*) were 36% for central organs (liver, gut, kidney, heart, and lung), 3% for brain, and 30% for skeletal muscles (Rolfe and Brown [1997](#page-15-29)). Many studies also reveal positive correlations between the masses of central organs and variation in BMR (see Swanson [2010;](#page-16-2) Williams et al. [2010](#page-17-4); Burton et al. [2011](#page-12-11); Wiersma et al. [2012;](#page-17-5) Careau et al. [2014](#page-12-26) for reviews), although such positive correlations are not necessarily universal among all selective forces. For example, Williams et al. [\(2001](#page-17-8)) found that marine carnivores had substantially higher BMRs than carnivorous terrestrial mammals, and these elevated BMRs were associated with large digestive tracts, which were necessary for supporting the energetic demands and varied diet of the marine lifestyle. In addition, increasing energy demands associated with winter, acclimation to cold or short photoperiod, and lactation or endurance training in mammals result in BMR variation that is often positively associated with changes in some or all central organ masses (i.e., digestive tract, liver, kidney, heart) (Hammond and Kristan [2000;](#page-13-31) Nespolo et al.

[2002](#page-15-31); Zhu et al. [2012\)](#page-17-9). In contrast, an inverse relationship between BMR and intestine mass occurred for South American field mouse (*Abrothrix andinus*, Bozinovic et al. [1990](#page-12-27)), masses of small intestine, liver, kidney, and heart were not significantly correlated with BMR in leafeared mice (*Phyllotis darwini*, Bacigalupe et al. [2010](#page-12-6)), and masses of heart, lungs, gut, liver, and kidney were not significantly correlated with altitude-induced BMR variation in deer mice (*Peromyscus maniculatus*, Hammond et al. [1999](#page-13-32)).

Another approach to studying the relationship between BMR and organ masses and other functional correlates in mammals is artificial selection experiments, which have focused primarily on laboratory mice. Genetic lines selected for high BMR or high food intake tend to produce increases in BMR (Konarzewski and Diamond [1995](#page-14-4); Selman et al. [2001](#page-16-8); Książek et al. [2004;](#page-14-8) Brzęk et al. [2007](#page-12-28)). Genetic lines selected for high wheel running activity or high running endurance increase energy expenditure relative to control lines and often increase maximum metabolic rates, but do not necessarily increase BMR. Nevertheless, such artificial selection treatments can alter central organ masses which might affect BMR. For example, masses for some or all central organs (intestine, liver, heart, and kidney) increase in mice lines selected for high BMR (Książek et al. [2004](#page-14-8); Brzęk et al. [2007\)](#page-12-28). Inbred mice strains with high BMR showed higher liver, small intestine, kidney, and heart masses relative to strains with low BMR (Konarzewski and Diamond [1995](#page-14-4)). Mice lines selected for high food intake resulted in increased BMR and increased liver, but not digestive tract, mass (Selman et al. [2001](#page-16-8)). In contrast, mice lines selected for high wheel-running activity increased maximum metabolic rates, but not BMR, and masses of gastrocnemius muscle, heart, and liver also did not vary with selection for wheel-running behavior (Swal-low et al. [2005](#page-16-33); Kane et al. [2008](#page-14-29)). Finally, mice lines selected for high aerobic capacity (increased endurance) exhibited higher maximum metabolic rates, but not BMR, relative to control lines, and these changes were accompanied by increases in masses of gastrocnemius muscle and heart, but not gut, liver, or kidney (Gębczyński and Konarzewski [2009](#page-13-7)). Collectively, these artificial selection studies provide general support for the idea that central organ masses are positively correlated with BMR, but such a correlation is not inviolate.

Birds also show generally positive relationships between central organ masses and BMR, although as for mammals, this relationship is not inviolate (see Swanson [2010](#page-16-2); Wiersma et al. [2012](#page-17-5) for reviews) and may vary depending on the predominant selective forces. For example, Tiele-man et al. ([2003\)](#page-16-27) found that BMR was reduced in lark species from arid environments compared to those from mesic environments, but central organ masses did not vary with aridity. Larks from both arid and mesic environments, however, increased BMR and central organ masses in response to cold acclimation (Williams and Tieleman [2000;](#page-17-10) Tieleman et al. [2003](#page-16-27)). Skeletal muscle is generally thought to be less important than central organs as an effector of BMR, since it demonstrates low metabolic rates at rest. However, because skeletal muscle makes up a large percentage of body mass in most endotherms, perhaps especially birds, it may nevertheless be an important determinant of BMR (Elia [1992](#page-13-30); Weber and Piersma [1996;](#page-16-34) Chappell et al. [1999](#page-13-3); Vézina and Williams [2003;](#page-16-9) Raichlen et al. [2010](#page-15-4); Petit et al. [2014](#page-15-32)). Several recent studies, however, highlight the variability of the relationships between central organ masses, skeletal muscle masses, and BMR in birds. Petit et al. [\(2014](#page-15-32)) studied seasonally acclimatized black-capped chickadees (*Poecile atricapillus*) from Quebec, Canada, finding that BMR increased during winter and that BMR was positively correlated with skeletal muscle, liver, and kidney masses, but not digestive tract mass, with skeletal muscle mass being the strongest predictor of BMR. Liang et al. [\(2015](#page-14-30)) exposed Chinese bulbuls (*Pycnonotus sinensis*) to short-term (7-day) food restriction, resulting in a reduction in BMR which was accompanied by reductions in liver and heart masses, but not kidney and digestive tract masses. Bai et al. ([2016\)](#page-12-29) examined three species of Chinese passerines in spring, including both migrants and residents, and found that BMR was positively correlated only with total digestive tract mass, whereas pectoralis muscle, heart, liver, kidney, gizzard, and small intestine masses were not significantly correlated with BMR. Finally, Barceló et al. ([2017\)](#page-12-15) exposed white-throated sparrows (*Zonotrichia albicollis*) to experimental cold and high fiber diet treatments. BMR in these sparrows increased during cold exposure, as did heart, digestive tract, liver, and kidney, but not pectoralis muscle, masses (Barceló et al. [2017\)](#page-12-15). Thus, various central organ masses may often be positively correlated with BMR variation in birds, but this is not always the case and pectoralis muscle mass may or may not be positively associated with BMR in birds.

Cellular metabolic intensity and BMR

In addition to the overall size of organs, cellular metabolic rates, especially in central organs with high maintenance activities, could also influence BMR. Maximal activities of key aerobic regulatory enzymes (citrate synthase [CS] or cytochrome C oxidase [COX]) in various tissues are often used as indices of cellular metabolic intensities, but these are more effective measures of maximal cellular metabolic capacity, rather than rates occurring at BMR. Mitochondrial volume density, basal cellular metabolic rates in isolated cells, or State 4 respiration in isolated mitochondria (an index of resting mitochondrial respiration rates) might be more appropriate measures for studying effectors of BMR variation, since these may be better measures of resting cellular aerobic flux. Several mammalian studies document positive associations between either BMR or conditions that increase energy demand (e.g., winter acclimatization, cold acclimation, or endurance training), which might be expected to increase BMR, and/or CS or COX activities, state 4 respiration or mitochondrial density in liver, brown adipose tissue, or skeletal muscle (Wickler [1981](#page-17-11); Hoppeler et al. [1995;](#page-13-33) Li et al. [2001;](#page-14-31) Liu et al. [2009](#page-14-32)). In contrast, some mammalian studies fail to detect positive correlations between BMR and these variables in liver, skeletal muscle, or brown adipose tissue under conditions of increased energy demand (Li et al. [2001](#page-14-31); Villarin et al. [2003](#page-16-35)). To complicate the issue further, indices of cellular metabolic intensity sometimes show variable relationships with BMR. Mineo et al. ([2012\)](#page-15-33) found that CS activities in skeletal muscle and brown adipose tissue increased with cold acclimation, but State 4 respiration did not. Artificial selection experiments in mice and rats have focused mainly on cellular metabolic intensity of hindlimb muscle and heart in lines selected for high running activity and generally show increased CS and/or COX activities in these tissues (Houle-Leroy et al. [2000](#page-13-34); Koch and Britton [2005](#page-14-33); Kelly et al. [2017\)](#page-14-34). These cellular changes, however, may not be positively correlated with BMR changes (Kane et al. [2008](#page-14-29)). Finally, BMR in humans was not correlated with skeletal muscle mitochondrial density or CS activity, but was positively correlated with mitochondrial oxygen affinity, which, in turn, was mediated in part by COX activity (Larsen et al. [2011\)](#page-14-35). Thus, variation in cellular metabolic intensity in various tissues may contribute to BMR variation in mammals, but such variation is far from universal.

A similar situation exists for birds, where pectoralis muscle activities of key aerobic enzymes are often, although not always, positively correlated with maximal metabolic capacities (Swanson [2010;](#page-16-2) Liknes and Swanson [2011](#page-14-36); Price et al. [2011\)](#page-15-34), but whether correlations also exist for BMR is less studied. In addition, cellular metabolic intensities of central organs are likely to be important to BMR variation. Avian aerobic enzyme activities are substantially higher in skeletal muscle and heart than for central organs, such as liver and kidney (Weber and Piersma [1996](#page-16-34); Vézina and Williams [2005\)](#page-16-36), but, as mentioned before, these activities provide measures of maximal flux through these pathways, so a better measure of resting cellular metabolic intensity is State 4 mitochondrial respiration. Correlations of BMR with both aerobic enzyme activities and State 4 mitochondrial respiration have been examined for a number of bird species under conditions differing in energy demands and results generally fall into two patterns: positive correlations with BMR or no relationship with BMR. Positive correlations of BMR with CS, COX, and/or State

4 mitochondrial respiration in pectoralis and/or liver occur for a number of bird species under conditions of winter acclimatization, cold acclimation, geographic temperature variation, or photoperiod variation (Liu et al. [2008;](#page-14-37) Zheng et al. [2008a,](#page-17-12) [b](#page-17-13), [2010](#page-17-14), [2013](#page-17-15), [2014a](#page-17-16), [b](#page-17-17); Peña-Villalobos et al. [2014](#page-15-35); Bai et al. [2016](#page-12-29); Zhou et al. [2016](#page-17-18); Hu et al. [2017](#page-13-35)), although not all indices of cellular metabolic intensity in all organs produced significant correlations with BMR in these studies. In contrast, BMR was not correlated with CS activity in pectoralis, kidney, heart, or liver in breeding European starlings (*Sturnus vulgaris*, Vézina and Williams [2005](#page-16-36)). In addition, cellular metabolic rates of temperate birds are higher than for tropical birds, consistent with broad BMR differences between these groups (Jimenez et al. [2014\)](#page-14-38). Thus, for birds, variation in cellular metabolic intensities in different tissues often covary with BMR, but such relationships are not universal for birds in all tissues or under all selective regimes.

An additional potential mechanism for increasing cellular metabolic intensity could involve elevated plasma levels or turnover of triiodothyronine (T_3) , which are generally positively correlated with BMR in endotherms (e.g., Kim [2008\)](#page-14-39), although the cause–effect relationship between *T*3 and BMR is not known (see Hulbert and Else [2004](#page-13-36) for review). Plasma T_3 levels are positively correlated with BMR under conditions of varying energy demand in both birds (Liu et al. [2006](#page-14-40); Zheng et al. [2008b](#page-17-13); [2010](#page-17-14), [2013;](#page-17-15) but see; Welcker et al. [2013](#page-16-37)) and mammals (e.g., Seidel et al. [1988](#page-16-38); Li et al. [2001](#page-14-31); Liu et al. [2009](#page-14-32); Brinkmann et al. [2016](#page-12-30); Rimbach et al. [2017\)](#page-15-36). Modifications of plasma membrane fatty acids can also influence cellular metabolic intensity and BMR, generally with increasing levels of unsaturation leading to increasing permeability of membranes to ions and increased costs of maintaining ionic gradients (Rolfe and Brown [1997;](#page-15-29) Hulbert and Else [2000](#page-13-37)). Indeed, this relationship underlies the membrane pacemaker hypothesis for the connection between metabolic rates and life history in endotherms (Hulbert and Else [1999,](#page-13-38) [2000\)](#page-13-37). Membrane fatty acid unsaturation is also mechanistically associated with the metabolic rate–body size relationship in endotherms, with smaller animals having a generally greater degree of unsaturation and higher mass-specific metabolic rates (Hulbert and Else [2004](#page-13-36)). A number of endotherm studies offer variable support for the membrane pacemaker hypothesis, with some endotherms showing unsaturation of membrane fatty acids in various tissues positively correlated with BMR or MMR, but others not, and correlations of different fatty acids in different tissues with either BMR or MMR in different organisms (Guglielmo et al. [2002](#page-13-39); Pierce et al. [2005](#page-15-37); Brzęk et al. [2007](#page-12-28); Haggerty et al. [2008;](#page-13-40) Wone et al. [2013](#page-17-19)). Thus, membrane fatty acid composition can influence BMR or MMR, although not in a particularly consistent manner across all organisms or all selective forces.

Because BMR and MMR are largely products of different tissues (see "[Organ masses and BMR](#page-8-0)") and selection on BMR (reduction) and MMR (elevation in many cases) may be operating in different directions, a dissociation of the phenotypic correlation between these traits might be expected under current selective regimes (Fig. [3](#page-11-0)). According to the aerobic capacity model for the evolution of endothermy (Bennett and Ruben [1979\)](#page-12-12), both traits increased during the evolution of endothermy, as sustained activity capacities and concomitant support costs both increased. Such selective trends might not continue to operate under present-day conditions, as selection on MMR may now represent stabilizing selection instead of selection for further increases (Swanson and Garland [2009;](#page-16-39) Raichlen et al. [2010;](#page-15-4) Swanson and Bozinovic [2011;](#page-16-40) Nilsson and Nilsson [2016](#page-15-14); Nespolo et al. [2017](#page-15-8)), while selection should act to minimize BMR, as long as maintenance costs are covered. In other words, the selective advantage of low BMR, as long as necessary support costs are met, stays consistent through evolutionary time, but selection on high sustained activity does not. This could dissociate selective trends on the two traits under current conditions (Fig. [3](#page-11-0)). In summary, adjustments of body size, body temperature, and organ masses and/or cellular metabolic intensities in endotherms, especially adjustments associated with central organs, are likely to influence BMR. However, we emphasize that such adjustments promoting increases in BMR arise from selection for activity or maximal aerobic metabolic rates (MMR) rather than for high BMR directly.

Summary

In conclusion, we contend that BMR can be subject to directional selection for reduction under conditions favoring energy conservation and a low BMR can, therefore, be directly adaptive. High BMR, however, seems unlikely to be directly adaptive and instead appears to be largely a correlated response to selection on physiological traits associated with increased capacities for activity or thermoregulation. We postulate that the selection for maintaining BMR as low as possible is a general selective feature for BMR in endotherms. We further contend that variation in endotherm energetics arises from selection on other traits and that these other factors promote higher maintenance costs (i.e., higher BMR) under energetically demanding conditions. Nevertheless, even under energetically demanding conditions, BMR should always be as low as possible under existing environmental or ecological constraints. Otherwise, the animal is "wasting" energy (i.e., the "needless encumbrance" of Dawson and O'Connor [1996\)](#page-13-10). Studies examining concurrent variation in BMR and DEE (or activity or thermoregulatory capacities) and their mechanistic underpinnings (e.g., organ masses, cellular metabolic capacities, variation in transcriptomes, proteomes, or metabolomes) in response to natural or experimentally manipulated energetic demands should help to resolve how these metabolic traits are adjusted (perhaps differentially) to changing energy demands. In addition, studies of the relative timing of such variation will be important in delineating selective actions on these traits. In addition,

Fig. 3 Graphical representation for how erosion of the phenotypic correlation between basal and maximal metabolic rates might occur. The aerobic capacity model for the evolution of endothermy posits that both basal and maximal metabolic rates increased in parallel, leading to a positive relationship between these variables (**a**). Selec-

tion patterns for maximal metabolic rates, however, may now be stabilizing instead of directional selection for further increases, while selection should act to minimize BMR, as long as maintenance costs are met, potentially leading to dissociation of the phenotypic correlation between BMR and metabolic capacities (**b**)

studies examining relative fitness consequences of BMR and DEE or activity or thermogenic capacities (e.g., Petit et al. [2017\)](#page-15-3) are needed to determine whether fitness is more closely associated with BMR or with other physiological traits. Finally, experimental manipulation of BMR, perhaps through thyroid hormone treatment or other mechanisms, without changing DEE or metabolic capacities (Bishop et al. [1995;](#page-12-31) Burger and Denver [2002;](#page-12-32) Liu et al. [2006](#page-14-40); Welcker et al. 2013 ; King and Swanson 2013), followed by measurement of fitness consequences under conditions of variable energy demand could reveal whether, and under what conditions, selection acts directly on BMR or on other metabolic traits.

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References

- Aschoff J, Pohl H (1970) Rhythmic variations in energy metabolism. Fed Proc 291:1541–1552
- Ashton KG (2002) Patterns of within-species body size variation of birds: strong evidence for Bergmann's rule. Global Ecol Biogeogr 11:505–523
- Ashton KG, Tracy MC, de Queiroz A (2000) Is Bergmann's rule valid for mammals? Am Nat 156:391–415
- Bacigalupe LD, Nespolo RF, Bustamante DM, Bozinovic F (2004) The quantitative genetics of sustained energy budget in a wild mouse. Evolut Int J Org Evolut 58:421–429
- Bacigalupe LD, Bustamante DM, Bozinovic F, Nespolo RF (2010) Phenotypic integration of morphology and energetic performance under routine capacities: a study in the leaf-eared mouse *Phyllotis darwini*. J Comp Physiol B 180:293–299
- Bai M, Wu X, Cai K, Zheng W, Liu J-S (2016) Relationships between interspecific differences in the mass of internal organs, biochemical markers of metabolic activity and the thermogenic properties of three small passerines. Avian Res 7:11.
- Barceló G, Love OP, Vézina F (2017) Uncoupling basal and summit metabolic rates in white-throated sparrows: digestive demand drives maintenance costs but changes in muscle mass are not needed to improve thermogenic capacity. Physiol Biochem Zool 90:153–165
- Bartholomew GA, Trost CH (1970) Temperature regulation in the speckled mousebird, *Colius striatus*. Condor 72:141–146
- Benedict FG (1938) Vital energetics: a study in comparative basal metabolism. Carnegie Inst, Washington (**Publication 503**)
- Bennett AF, Ruben JA (1979) Endothermy and activity in vertebrates. Science 206:649–654
- Bergmann C (1847) Ueber die Verhältnisse der Wärmeökonomie der Thiere zu ihrer Grösse. Gottinger Studien 3:595–708
- Bishop CM, Butler PJ, Atkinson NM (1995) The effect of elevated levels of thyroxine on the aerobic capacity of locomotor muscles of the tufted duck. Aythya fuligula J Comp Physiol B 164:618–621
- Blackburn TM, Hawkins BA (2004) Bergmann's rule and the mammal fauna of northern North America. Ecography 27:715–724
- Bligh J, Johnson KG (1973) Glossary of terms for thermal physiology. J Appl Physiol 35:941–961
- Boily P (2002) Individual variation in metabolic traits of wild ninebanded armadillos (*Dasypus novemcinctus*), and the aerobic capacity model for the evolution of endothermy. J Exp Biol 205:3207–3214
- Boratyński Z, Koteja P (2009) The association between body mass, metabolic rates and survival of bank voles. Funct Ecol 23:330–339
- Boratyński Z, Koskela E, Mappes T, Schroderus E (2013) Quantitative genetics and fitness effects of basal metabolism. Evol Ecol 27:301–314
- Boratyński JS, Jefimow M, Wojciechowski MS (2016) Phenotypic flexibility of energetics in acclimated Siberian hamsters has a narrower scope in winter than in summer. J Comp Physiol B 186:387–402
- Bozinovic F, Sabat P (2010) On the intraspecific variability in basal metabolism and the food habits hypothesis in birds. Curr Zool 56:759–766
- Bozinovic F, Novoa FF, Veloso C (1990) Seasonal changes in energy expenditure and digestive tract of *Abrothrix andinus* in the Andes Range. Physiol Zool 63:1216–1231
- Bozinovic F, Rojas JM, Broitman BR, Vásquez RA (2009) Basal metabolism is correlated with habitat productivity among populations of degus (*Octodon degus*). Comp Biochem Physiol A 152:560–564
- Brinkmann L, Gerken M, Hambly C, Speakman JR, Ried A (2016) Thyroid hormones correlate with field metabolic rate in ponies, *Equus ferus caballus*. J Exp Biol 219:2559–2566
- Brzęk P, Bielawska K, Książek A, Konarzewski M (2007) Anatomic and molecular correlates of divergent selection for basal metabolic rate in laboratory mice. Physiol Biochem Zool 80:491–499
- Buffenstein R, Yahav S (1991) Is the naked mole-rat *Heterocephalus glaber* an endothermic yet poikilothermic mammal? J Therm Biol 16:227–232
- Burger MF, Denver RJ (2002) Plasma thyroid hormone concentrations in a wintering passerine bird: their relationship to geographic variation, environmental factors, metabolic rate and body fat. Physiol Biochem Zool 75:187–199
- Burton T, Killen SS, Armstrong JD, Metcalfe NB (2011) What causes intraspecific variation in resting metabolic rate and what are its ecological consequences. Proc Roy Soc B Biol Sci 278:3465–3473
- Careau V (2013) Basal metabolic rate, maximum thermogenic capacity and aerobic scope in rodents: interaction between environmental temperature and torpor use. Biol Lett 9:20121104
- Careau V, Garland T Jr (2012) Performance, personality and energetics: correlation, causation, and mechanism. Physiol Biochem Zool 85:543–571
- Careau V, Thomas D, Humphries MM, Réale D (2008) Energy metabolism and animal personality. Oikos 117:641–653
- Careau V, Thomas D, Pelletier F, Turki L, Landry F, Garant D, Réale D (2011) Genetic correlation between resting metabolic rate and exploratory behaviour in deer mice (*Peromyscus maniculatus*). J Evol Biol 24:2153–2163
- Careau V, Bergeron P, Garant D, Réale D, Speakman JR, Humphries MM (2013a) The energetic and survival costs of growth in freeranging eastern chipmunks. Oecologia 171:11–23
- Careau V, Réale D, Garant D, Pelletier F, Speakman JR, Humphries MM (2013b) Context-dependent correlation between resting metabolic rate and daily energy expenditure in wild chipmunks. J Exp Biol 216:418–426
- Careau V, Killen SS, Metcalfe NB (2014) Adding fuel to the "fire of life": energy budgets across levels of variation in ectotherms and endotherms. In: Martin LB, Ghalambor CK, Woods HA (eds) Integrative organismal biology. Wiley, Hoboken, pp 219–233
- Chappell M, Bech C, Buttemer W (1999) The relationship of central and peripheral organ masses to aerobic performance variation in house sparrows. J Exp Biol 202:2269–2279
- Clarke A, Rothery P (2008) Scaling of body temperature in mammals and birds. Funct Ecol 22:58–67
- Clarke A, Rothery P, Isaac NJB (2010) Scaling of basal metabolic rate with body mass and temperature in mammals. J Anim Ecol 79:610–619
- Clavijo-Baquet S, Bozinovic F (2012) Testing the fitness consequences of the thermoregulatory and parental care models for the origin of endothermy. PLoS One 7:e37069
- Crompton AW, Taylor CR, Jagger JA (1978) Evolution of homeothermy in mammals. Nature 272:333–336
- Cruz-Neto AP, Bozinovic F (2004) The relationship between diet quality and basal metabolic rate in endotherms: insights from intraspecific analyses. Physiol Biochem Zool 77:877–889
- Dawson WR, Marsh RL (1989) Metabolic acclimatization to cold and season in birds. In: Bech C, Reinertsen RE (eds) Physiology of cold adaptation in birds. Plenum Life Sciences, New York, pp. 83–94
- Dawson WR, O'Connor TP (1996) Energetic features of avian thermoregulatory responses. In: Carey C (ed) Avian energetics and nutritional ecology. Chapman & Hall, New York, pp 85–124
- Dittmann MR, Hummel J, Runge U, Galeffi C, Kreuzer M, Clauss M (2014) Characterising an artiodactyl family inhabiting arid habitats by its metabolism: low metabolism and maintenance requirements in camelids. J Arid Env 107:41–48
- Dohm MR, Hayes JP, Garland T Jr (2001) The quantitative genetics of maximal and basal rates of oxygen consumption in mice. Genetics 159:267–277
- Dutenhoffer MS, Swanson DL (1996) Relationship of basal to summit metabolic rate in passerine birds and the aerobic capacity model for the origin of endothermy. Physiol Zool 69:1232–1254
- Elia M (1992) Organ and tissue contribution to metabolic rate. In: Kinney JM, Tucker HN (eds) Energy metabolism: tissue determinants and cellular corollaries. Raven Press, New York, pp 61–77
- Enstipp MR, Grémillet D, Jones DR (2008) Heat increment of feeding in double-crested cormorants (*Phalacrocorax auritis*) and its potential for thermal substitution. J Exp Biol 211:49–57
- Finke C, Misovic A, Prinzinger R (1995) Growth, the development of endothermy, and torpidity in blue-naped mousebirds *Urocolius macrourus*. Ostrich 66:1–9
- Fristoe TS, Burger JR, Balk MA, Khaliq I, Hof C, Brown JH (2015) Metabolic heat production and thermal conductance are massindependent adaptations to thermal environment in birds and mammals. Proc Natl Acad Sci USA 52:15934–15939
- Gardner JL, Peters A, Kearney MR, Joseph L, Heinsohn R (2011) Declining body size: a third universal response to warming? Trends Ecol Evol 26:285–291
- Gębczyński AK, Konarzewski M (2009) Locomotor activity of mice divergently selected for basal metabolic rate: a test of hypotheses on the evolution of endothermy. J Evol Biol 22:1212–1220
- Geiser F, Baudinette RV (1987) Seasonality of torpor and thermoregulation in three dasyurid marsupials. J Comp Physiol B 157:335–344
- Geist V (1987) Bergmann's rule is invalid. Can J Zool 65:1035–1038
- Glazier DS (2015) Is metabolic rate a universal 'pacemaker' for biological processes? Biol Rev 90:377–407
- Goodman RE, Lebuhn G, Seavy NE, Gardali T, Bluso-Demers JD (2012) Avian body size changes and climate change: warming or increasing variability? Global Change Biol 18:63–73
- Green JA, Aitken-Simpson EJ, White CR, Bunce A, Butler PJ, Frappell PB (2013) An increase in minimum metabolic rate and not activity explains field metabolic rate changes in a breeding seabird. J Exp Biol 216:1726–1735
- Guglielmo CG, Williams TD, Zwingelstein G, Brichon G, Weber J-M (2002) Plasma and muscle phospholipids are involved in the metabolic response to long-distance migration in a shorebird. J Comp Physiol B 172:409–417
- Hackett SJ, Kimball RT, Reddy S, Bowie RCK, Braun EL, Braun MJ, Chojnowski JL, Cox WA, Han K-L, Harshman J, Huddleston CJ, Marks BD, Miglia KJ, Moore WS, Sheldon FH, Steadman DW, Witt CC, Yuri T (2008) A phylogenomic study of birds reveals their evolutionary history. Science 320:1763–1768
- Haggerty C, Hoggard N, Brown DS, Clapham JC, Speakman JR (2008) Intra-specific variation in resting metabolic rate in MF1 mice is not associated with membrane desaturation in the liver. Mech Ageing Dev 129:129–137
- Hammond KA, Kristan DM (2000) Responses to lactation and cold exposure by deer mice (*Peromyscus maniculatus*). Physiol Biochem Zool 73:547–556
- Hammond KA, Roth J, Janes DN, Dohm MR (1999) Morphological and physiological responses to altitude in deer mice *Peromyscus maniculatus*. Physiol Biochem Zool 72:613–622
- Hayes JP, Garland T Jr (1995) The evolution of endothermy: testing the aerobic capacity model. Evolut Int J org Evolut 49:836–847
- Hayes JP, O'Connor CS (1999) Natural selection on thermogenic capacity of high-altitude deer mice. Evolut Int J org Evolut 53:1280–1287
- Heldmaier G, Steinlechner S (1981) Seasonal control of energy requirements for thermoregulation in the Djungarian hamster (*Phodopus sungorus*), living in natural photoperiod. J Comp Physiol 142:429–437
- Hindle AG, McIntyre IW, Campbell KL, MacArthur RA (2003) The heat increment of feeding and its thermoregulatory implications in the short-tailed shrew (Blarina brevicauda). Can J Zool 81:1445–1453
- Hislop MS, Buffenstein R (1994) Noradrenaline induces nonshivering thermogenesis in both the naked mole-rat (*Heterocephalus glaber*) and the Damara mole-rat (*Cryptomys damarensis*) despite very different modes of thermoregulation. J Therm Biol $19.25 - 32$
- Hoppeler H, Altpeter E, Wagner M, Turner DL, Hokanson J, König M, Stalder-Navarro VP, Weibel ER (1995) Cold acclimation and endurance training in guinea pigs: changes in lung, muscle and brown fat tissue. Resp Physiol 101:189–198
- Houle-Leroy P, Garland T Jr, Swallow JG, Guderley H (2000) Effects of voluntary actiity and genetic selection on muscle metabolic capacities in house mice *Mus domesticus*. J Appl Physiol 89:1608–1616
- Hu S-N, Zhu Y-Y, Lin L, Zheng W-H, Liu J-S (2017) Temperature and photoperiod as environmental cues affect body mass and thermoregulation in Chinese bulbuls *Pycnonotus sinensis*. J Exp Biol 220:844–855
- Hulbert AJ, Else PL (1999) Membranes as possible pacemakers of metabolism. J Theor Biol 199:257–274
- Hulbert AJ, Else PL (2000) Mechanisms underlying the cost of living in animals. Annu Rev Physiol 62:207–235
- Hulbert AJ, Else PL (2004) Basal metabolic rate: history, composition, regulation and usefulness. Physiol Biochem Zool 77:869–876
- Humphries MM, Careau V. (2011) Heat for nothing or activity for free? Evidence and implications of activity-thermoregulatory heat substitution. Integr Comp Biol 51:419–431
- Jefimow M, Wojciechowski M, Masuda A, Oishi T (2004) Correlation between torpor frequency and capacity for non-shivering thermogenesis in the Siberian hamster (*Phodopus songorus*). J Therm Biol 29:641–647
- Jetz W, Freckleton RP, McKechnie AE (2008) Environment, migratory tendency, phylogeny and basal metabolic rate in birds. PLoS One 3:e3261
- Jimenez AG, Van Brocklyn J, Wortman M, Williams JB (2014) Cellular metabolic rate is influenced by life-history traits in tropical and temperate birds. PLoS One 9:e87349
- Kane SL, Garland T Jr, Carter PA (2008) Basal metabolic rate of aged mice is affected by random genetic drift but not be selective breeding for high early-age locomotor activity or chronic wheel access. Physiol Biochem Zool 81:288–300
- Kelly SA, Gomes FR, Kolb EM, Malisch JL, Garland T Jr (2017) Effects of activity, genetic selection, and their interaction on muscle metabolic capacities and organ masses in mice. J Exp Biol 220:1038–1047
- Killen SS, Glazier DS, Rezende EL, Clark TD, Atkinson D, Willener AST, Halsey LG (2016) Ecological influences and morphological correlates of resting and maximal metabolic rates across teleost fish species. Am Nat 187:592–606
- Kim B (2008) Thyroid hormone as a determinant of energy expenditure and the basal metabolic rate. Thyroid 18:141–144
- King MO, Swanson DL (2013) Activation of the immune system incurs energetic costs but has no effect on the thermogenic performance of house sparrows during acute cold challenge. J Exp Biol 216:2097–2102

Kleiber M (1961) The fire of life. Wiley, New York

- Koch LG, Britton SL (2005) Divergent selection for aerobic capacity in rats as a model for complex disease. Integr Comp Biol 45:405–415
- Konarzewski M, Diamond J (1995) Evolution of basal metabolic rate and organ masses in laboratory mice. Evolut Int J org Evolut 49:1239–1248
- Konarzewski M, Książek A, Lapo IB (2005) Artificial selection of metabolic rates and related traits in rodents. Integr Comp Biol 45:416–425
- Książek A, Konarzewski M, Lapo IB (2004) Anatomic and energetic correlates of divergent selection for basal metabolic rate in laboratory mice. Physiol Biochem Zool 77:890–899
- Książek A, Czerniecki J, Konarzewski M (2009) Phenotypic flexibility of traits related to energy acquisition in mice divergently selected for basal metabolic rate (BMR). J Exp Biol 212:808–814
- Larsen FJ, Schiffer TA, Sahlin K, Ekblom B, Weitzberg E, Lundberg JO (2011) Mitochondrial oxygen affinity predicts basal metabolic rate in humans. FASEB J 25:2843–2852
- Lewden A, Petit M, Vézina F (2012) Dominant black-capped chickadees pay no maintenance energy costs for their wintering status and are not better at enduring cold than subordinate individuals. J Comp Physiol B 182:381–392
- Li X-S, Wang D-H (2005) Seasonal adjustments in body mass and thermogenesis in Mongolian gerbils (*Meriones unguiculatus*): the roles of short photoperiod and cold. J Comp Physiol B 175:593–600
- Li Q, Sun R-Y, Huang C, Wang Z, Liu X, Hou J, Liu J-S, Cai L, Li N, Zhang S, Wang Y (2001) Cold adaptive thermogenesis in small mammals from different geographical zones of China. Comp Biochem Physiol A 129:949–961
- Liang Q-J, Zhao L, Wang J-Q, Chen Q, Zheng W-H, Liu J-S (2015) Effect of food restriction on the energy metabolism of the Chinese bulbul (*Pycnonotus sinensis*). Zool Res 36:79–87
- Liknes ET, Swanson DL (2011) Phenotypic flexibility in passerine birds: seasonal variation of aerobic enzyme activities in skeletal muscle. J Therm Biol 36:430–436
- Liu J-S, Chen Y-Q, Li M (2006) Thyroid hormones increase liver and muscle thermogenic capacity in little buntings (*Emberiza pusilla*). J Therm Biol 31:386–393
- Liu J-S, Li M, Shao S-L (2008) Seasonal changes in thermogenic properties of liver and muscle in tree sparrows *Passer montanus*. Acta Zool Sinica 54:777–784
- Liu J-S, Yang M, Sun R-Y, Wang D-H (2009) Adaptive thermogenesis in Brandt's vole (*Lasiopodomys brandti*) during cold and warm acclimation. J Therm Biol 34:60–69
- Liwanag HEM, Williams TM, Costa DP, Kanatous SB, Davis RW, Boyd IL (2009) The effects of water temperature on the energetic costs of juvenile and adult California sea lions (*Zalophus californianus*): the importance of skeletal muscle thermogenesis for thermal balance. J Exp Biol 212:3977–3984
- Londoño GA, Chappell MA, Castañeda MR, Jankowski JE, Robinson SK (2015) Basal metabolism in tropical birds: latitute, altitude, and the "pace of life". Funct Ecol 29:338–346
- Lovegrove BG (2003) The influence of climate on the basal metabolic rate of small mammals: a slow-fast metabolic continuum. J Comp Physiol B 173:87–112
- Lovegrove BG (2005) Seasonal thermoregulatory responses in mammals. J Comp Physiol B 175:231–247
- Lovegrove BG (2012) The evolution of endothermy in Cenozoic mammals: a plesiomorphic-apomorphic continuum. Biol Rev 87:128–162
- Luna F, Naya H, Naya DE (2017) Understanding evolutionary variation in basal metabolic rate: an analysis in subterranean rodents. Comp Biochem Physiol A 206:87–94
- Maldonado K, Cavieres G, Veloso C, Canals M, Sabat P (2009) Physiological responses in rufous-collared sparrows to thermal acclimation and seasonal acclimatization. J Comp Physiol B 179:335–343
- Maldonado K, van Dongen WFD, Vásquez RA, Sabat P (2012) Geographic variation in the association between exploratory behavior and physiology in rufous-collared sparrow. Physiol Biochem Zool 85:618–624
- Mathot KJ, Nicolaus M, Araya-Ajoy YG, Dingemanse NJ, Kempenaers B (2015) Does metabolic rate predict risk-taking behaviour? A field experiment in a wild passerine bird. Funct Ecol 29:239–249
- McKechnie AE (2008) Phenotypic flexibility in basal metabolic rate and the changing view of avian physiological diversity: a review. J Comp Physiol B 178:235–247
- McKechnie AE, Lovegrove BG (2001) Thermoregulation and the energetic significance of clustering behavior in the whitebacked mousebird (*Colius colius*). Physiol Biochem Zool 74:238–249
- McKechnie AE, Swanson DL (2010) Sources and significance of variation in basal, summit and maximal metabolic rates in birds. Curr Zool 56:741–758
- McKechnie AE, Körtner G, Lovegrove BG (2004) Rest-phase thermoregulation in free-ranging white-backed mousebirds. Condor 106:144–150
- McKechnie AE, Körtner G, Lovegrove BG (2006) Thermoregulation under semi-natural conditions in specked mousebirds: the role of communal roosting. Afr Zool 41:155–163
- McKechnie AE, Noakes MJ, Smit B (2015) Global patterns of seasonal acclimatization in avian resting metabolic rates. J Ornithol 156(Suppl 1):S367–S376
- McNab BK (1978) The evolution of endothermy in the phylogeny of mammals. Am Nat 112:1–21
- McNab BK (1988) Food habits and the basal rate of metabolism in birds. Oecologia 77:343–349
- McNab BK (1997) On the utility of uniformity in the definition of the basal rate of metabolism. Physiol Zool 70:718–720
- McNab BK (2008) An analysis of the factors that influence the level and scaling of mammalian BMR. Comp Biochem Physiol A 151:5–28
- McNab BK (2012) Extreme measures: the ecological energetics of birds and mammals. University of Chicago Press, Chicago
- Meiri S, Dayan T (2003) On the validity of Bergmann's rule. Global Ecol Biogeogr 30:331–351
- Mineo PM, Cassell EA, Roberts ME, Schaeffer PJ (2012) Chronic cold acclimation increases thermogenic capacity, non-shivering thermogenesis and muscle citrate synthase activity in both wildtype and brown adipose tissue deficient mice. Comp Biochem Physiol A 161:395–400
- Møller AP (2009) Basal metabolic rate and risk taking behaviour in birds. J Evol Biol 22:2420–2429
- Mueller P, Diamond J (2001) Metabolic rate and environmental productivity: well-provisioned animals evolved to run and idle fast. Proc Natl Acad Sci USA 98:12550–12554
- Naya DE, Spangenberg L, Naya H, Bozinovic F (2013) How does evolutionary variation in basal metabolic rates arise? A statistical assessment and a mechanistic model. Evolut Int J org Evolut 67:1463–1476
- Nespolo RF, Bacigalupe LD, Sabat P, Bozinovic F (2002) Interplay among energy metabolism, organ mass and digestive enzyme activity in the mouse-opossum *Thylamys elegans*: the role of thermal acclimation. J Exp Biol 205:2697–2703
- Nespolo RF, Bustamante DM, Bacigalupe LD, Bozinovic F (2005) Quantitative genetics of bioenergetics and growth-related traits in the wild mammal, *Phyllotis darwini*. Evolut Int J org Evolut 59:1829–1837
- Nespolo RF, Baciagalupe LD, Figueroa CC, Koteja P, Opazo JC (2011) Using new tools to solve an old problem: the evolution of endothermy in vertebrates. Trends Ecol Evol 26:414–423
- Nespolo RF, Solano-Iguaran JJ, Bozinovic F (2017) Phylogenetic analysis supports the aerobic-capacity model for the evolution of endothermy. Am Nat 189:13–27
- Nilsson JF, Nilsson J-A (2016) Fluctuating selection on basal metabolic rate. Ecol Evolut 6:1197–1202
- Noakes MJ, Wolf BO, McKechnie AE (2017) Seasonal metabolic acclimatization varies in direction and magnitude among populations of an Afrotropical passerine bird. Physiol Biochem Zool 90:178–189
- Oelkrug R, Heldmaier G, Meyer CW (2011) Torpor patterns, arousal rates, and temporal organization of torpor entry in wildtype and UCP1-ablated mice. J Comp Physiol B 181:137–145
- Olson VA, Davies RG, Orme CDL, Thomas GH, Meiri S, Blackburn TM, Gaston KJ, Owens IPF, Bennett PM (2009) Global biogeography and the ecology of body size in birds. Ecol Lett 12:249–259
- Pauli JN, Peery MZ, Fountain ED, Karasov WH (2016) Arboreal folivores limit their energetic output, all the way to slothfulness. Am Nat 188:196–204
- Peña-Villalobos I, Nuñez-Villegas M, Bozinovic F, Sabat P (2014) Metabolic enzymes in seasonally acclimatized and cold acclimated rufous-collared sparrows inhabiting a Chilean Mediterranean environment. Curr Zool 60:338–350
- Petit M, Lewden A, Vézina F (2013) Intra-seasonal flexibility in avian metabolic performance highlights the uncoupling of basal metabolic rate and thermogenic capacity. PLoS One 8:e68292
- Petit M, Lewden A, Vézina F (2014) How does flexibility in body composition relate to seasonal changes in metabolic performance in a small passerine wintering at northern latitude? Physiol Biochem Zool 87:539–549
- Petit M, Clavijo-Baquet S, Vézina F (2017) Increasing winter maximal metabolic rate improves intra-winter survival in small birds. Physiol Biochem Zool 90:166–177
- Pierce BJ, McWilliams SR, O'Connor TP, Place AR, Guglielmo CG (2005) Effect of dietary fatty acid composition on depot fat and exercise performance in a migrating songbird, the red-eyed vireo. J Exp Biol 208:1277–1285
- Piersma T, van Gils J (2011) The flexible phenotype: A body-centred integration of ecology, physiology, and behavior. Oxford University Press, Oxford
- Porter WP, Kearney M (2009) Size, shape and the thermal niche of endotherms. Proc Natl Acad Sci USA 106(suppl 2):19666–19672
- Portugal SJ, Green JA, Halsey LG, Arnold W, Careau V, Dann P, Frappell PB, Grémillet D, Handrich Y, Martin GR, Ruf T, Guillemette MM, Butler PJ (2016) Associations between resting, activity, and daily metabolic rate in free-living endotherms: no universal rule in birds and mammals. Physiol Biochem Zool 89:251–261
- Price ER, Staples JF, Milligan CL, Guglielmo CG (2011) Carnitine palmitoyl transferase activity and whole muscle oxidation rates vary with fatty acid substrate in avian flight muscle. J Comp Physiol B 181:565–573
- Prinzinger R (1988) Energy metabolism, body-temperature and breathing parameters in non-torpid blue-naped mousebirds *Urocolius macrourus*. J Comp Physiol B 157:801–806
- Prinzinger R, Göppel R, Lorenz A, Kulzer E (1981) Body temperature and metabolism in the red-backed mousebird (*Colius castanotus*) during fasting and torpor. Comp Biochem Physiol A 69:689–692
- Prum RO, Berv JS, Dornburg A, Field DJ, Townsend JP, Lemmon EM Lemmon AR (2015) A comprehensive phylogeny of birds (Aves) using targeted next-generation DNA sequencing. Nature 526:569–573
- Raichlen DA, Gordon AD, Muchlinski MN, Snodgrass JJ (2010) Causes and significance of variation in mammalian basal metabolism. J Comp Physiol B 180:301–311
- Réale D, Garant D, Humphries MM, Bergeron P, Careau V, Montiglio P-O (2010) Personality and the emergence of the pace-of-life syndrome concept at the population level. Phil Trans Roy Soc B 36:4051–4063
- Rezende EL, Bacigalupe LD (2015) Thermoregulation in endotherms: physiological principles and ecological consequences. J Comp Physiol B 185:709–727
- Rezende EL, Swanson DL, Novoa FF, Bozinovic F (2002) Passerines versus nonpasserines: so far, no statistical differences in the scaling of avian energetics. J Exp Biol 20:101–107
- Rezende EL, Bozinovic F, Garland, T Jr (2004) Climatic adaptation and the evolution of basal and maximal rates of metabolism in rodents. Evolut Int J org Evolut 58:1361–1374
- Ricklefs RE, Konarzewski M, Daan S (1996) The relationship between basal metabolic rate and daily energy expenditure in birds and mammals. Am Nat 147:1047–1071
- Riddle O, Smith GC, Benedict FG (1932) The basal metabolism of the mourning dove and some of its hybrids. Am J Physiol 101:206–267
- Rimbach R, Pillay N, Schradin C (2017) Both thyroid hormone levels and resting metabolic rate decrease in African striped mice when food availability decreases. J Exp Biol 220:837–843
- Rodríguez MA, López-Sañudo IL, Hawkins BA (2006) The geographic distribution of mammal body size in Europe. Global Ecol Biogeogr 15:173–181
- Rodríguez MA, Olalla-Tárraga, Hawkins BA (2008) Bergmann's rule and the geography of mammal body size in the Western Hemisphere. Global Ecol Biogeogr 17:274–283
- Rolfe DFS, Brown GC (1997) Cellular energy utilization and molecular origin of standard metabolic rate in mammals. Physiol Rev 77:731–758
- Rønning B, Jensen H, Moe B, Bech C (2007) Basal metabolic rate: heritability and genetic correlations with morphological traits in the zebra finch. J Evol Biol 20:1815–1822
- Rønning B, Broggi J, Bech C, Moe B, Ringsby TH, Pärn H, Hagen IJ, Saether B-E, Jensen H (2016) Is basal metabolic rate associated with recruit production and survival in free-living house sparrows? Funct Ecol 30:1140–1148
- Rubner M (1883) Ueber den Eifluss der Körpergrösse auf Sttoffund Kraftwechsel. Z Biol 19:535–562
- Ruf T, Geiser F (2015) Daily torpor and hibernation in birds and mammals. Biol Rev 90:891–926
- Sabat P, Ramirez-Otarola N, Barceló G, Salinas J, Bozinovic F (2010) Comparative basal metabolic rate among passerines and the food habit hypothesis. Comp Biochem Physiol A 157:35–40
- Sadowska ET, Labocha MK, Baliga K, Stanisz A, Wróblewska AK, Jagusiak W, Koteja P (2005) Genetic correlations between basal and maximum metabolic rates in a wild rodent: consequences for evolution of endothermy. Evolut Int J org Evolut 59:672–681
- Sadowska ET, Baliga-Klimczyk K, Labocha MK, Koteja P (2009) Genetic correlations in a wild rodent: grass-eaters and fastgrowers evolve high basal metabolic rates. Evolut Int J org Evolut 63:1530–1539
- Schmidt-Nielsen K (1984) Scaling. Why is animal size so important? Cambridge Univ Press, Cambridge
- Scholander PF, Hock R, Walters V, Johnson F, Irving L (1950a) Adaptation to cold in arctic and tropical mammals and birds in relation to body temperature, insulation, and basal metabolic rate. Biol Bull 99:259–271
- Scholander PF, Hock R, Walters V, Johnson F, Irving L (1950b) Heat regulation in some arctic and tropical mammals and birds. Biol Bull 99:237–258
- Sears MW, Hayes JP, O'Connor CS, Geluso K, Sedinger JS (2006) Individual variation in thermogenic capacity affects aboveground activity of high-altitude deer mice. Funct Ecol 20:97–104
- Seidel A, Heldmaier G, Schulz F (1988) Effect of triiodothyronine, thyrotropin-releasing hormone and propylthiouracil on the thermogenic capacities of Djangarian hamsters living in natural photoperiod. J Therm Biol 13:49–51
- Selman C, Lumsden S, Bünger L, Hill WG, Speakman JR (2001) Resting metabolic rate and morphology in mice (*Mus musculus*) selected for high and low food intake. J Exp Biol 204:777–784
- Sheridan JA, Bickford D (2011) Shrinking body size as an ecological response to climate change. Nat Climate Change 1:401–406
- Sibley CG, Ahlquist JE (1990) Phylogeny and classification of birds. Yale University Press, New Haven
- Speakman JR (2005) Body size, energy metabolism and lifespan. J Exp Biol 208:1717–1730
- Speakman JR, Król E (2010) Maximal heat dissipation capacity and hyperthermia risk: neglected key factors in the ecology of endotherms. J Anim Ecol 79:726–746
- Speakman JR, Król E (2011) Limits to sustained energy intake. XIII. Recent progress and future perspectives. J Exp Biol 214:230–241
- Speakman JR, Król E, Johnson MS (2004) The functional significance of individual variation in basal metabolic rate. Physiol Biochem Zool 77:900–915
- Swallow JG, Rhodes JS, Garland T Jr (2005) Phenotypic and evolutionary plasticity of organ masses in response to voluntary exercise in house mice. Integr Comp Biol 45:426–437
- Swanson DL (2010) Seasonal metabolic variation in birds: functional and mechanistic correlates. Curr Ornithol 17:75–129
- Swanson DL, Bozinovic F (2011) Metabolic capacity and the evolution of biogeographic patterns in oscine and suboscine passerine birds. Physiol Biochem Zool 84:185–194
- Swanson DL, Garland T Jr (2009) The evolution of high summit metabolism and cold tolerance in birds and its impact on present-day distributions. Evolut Int J org Evolut 63:184–194
- Swanson DL, Thomas NE (2007) The relationship of plasma indicators of lipid metabolism and muscle damage to overnight temperature in winter-acclimatized small birds. Comp Biochem Physiol A 146:87–94
- Swanson DL, Thomas NE, Liknes ET, Cooper SJ (2012) Intraspecific correlations of basal and maximal metabolic rates in birds and the aerobic capacity model for the evolution of endothermy. PLoS One 7:e34271
- Tieleman BI, Williams JB, Buschur ME, Brown CR (2003) Phenotypic variation of larks along an aridity gradient: are desert birds more flexible. Ecology 84:1800–1815
- Tieleman BI, Versteegh MA, Helm B, Dingemanse NJ (2009) Quantitative genetics parameters show partial independent evolutionary potential for body mass and metabolism in stonechats from different populations. J Zool Lond 279:129–136
- van de Ven TMFN, Mzilikazi N, McKechnie AE (2013) Seasonal metabolic variation in two populations of an Afrotropical Euplectid bird. Physiol Biochem Zool 86:19–26
- Versteegh MA, Schwabl I, Jaquier S, Tieleman BI (2012) Do immunological, endocrine and metabolic traits fall on a single Paceof-Life axis? Covariation and constraints among physiological systems. J Evol Biol 25:1864–1876
- Vézina F, Salvante KG (2010) Behavioral and physiological flexibility are used by birds to manage energy and support investment in the early stages of reproduction. Curr Zool 56:767–792
- Vézina F, Williams TD (2003) Plasticity in body composition in breeding birds: what drives the metabolic costs of egg production? Physiol Biochem Zool 76:716–730
- Vézina F, Williams TD (2005) Interactions between organ mass and citrate synthase activity as an indicator of tissue maximal oxidative capacity in breeding European starlings: implications for metabolic rate and organ mass relationships. Funct Ecol 19:119–128
- Vézina F, Jalvingh KM, Dekinga A, Piersma T (2006) Acclimation to different thermal conditions in a northerly wintering shorebird is driven by body mass-related changes in organ size. J Exp Biol 209:3141–3154
- Vézina F, Love OP, Lessard M, Williams TD (2009) Shifts in metabolic demands in growing altricial nestlings illustrate contextspecific relationships between basal metabolic rate and body composition. Physiol Biochem Zool 82:248–257
- Villarin JJ, Schaeffer PJ, Markle RA, Lindstedt SL (2003) Chronic cold exposure increases liver oxidative capacity in the marsupial *Monodelphis domestica*. Comp Biochem Physiol A 136:621–630
- Walsberg GE (1990) Communal roosting in a very small bird: consequences for the thermal and respiratory environments. Condor 92:795–798
- Weathers WW (1979) Climatic adaptation in avian standard metabolic rate. Oecologia 42:81–89
- Weber TP, Piersma T (1996) Basal metabolic rate and the mass of tissues differing in metabolic scope: migration-related covariation between individual knots, *Calidris canutus*. J Avian Biol 27:215–224
- Welcker J, Chastel O, Gabrielsen GW, Guillaumin J, Kitaysky AS, Speakman JR, Tremblay Y, Bech C (2013) Thyroid hormones correlate with basal metabolic rate but not field metabolic rate in a wild bird species. PLoS One 8:e56229
- Welcker J, Speakman JR, Elliott KH, Hatch SA, Kitaysky AS (2015) Resting and daily energy expenditure during reproduction are adjusted in opposite directions in free-living birds. Funct Ecol 29:250–258
- Wells ME, Schaeffer PJ (2012) Seasonality of peak metabolic rate in non-migrant tropical birds. J Avian Biol 43:481–485
- White CR, Seymour RS (2004) Does basal metabolic rate contain a useful signal? Mammalian BMR allometry and correlations with a selection of physiological, ecological and life-history variables. Physiol Biochem Zool 77:929–941
- White CR, Blackburn TM, Martin GR, Butler PJ (2007) Basal metabolic rate of birds is associated with habitat temperature and

precipitation, not primary productivity. Proc Roy Soc B Biol Sci 274:287–293

- Wickler SJ (1981) Seasonal changes in enzymes of aerobic heat production in the white-footed mouse. Am J Physiol Reg Integr Comp Physiol 240:R289–R294
- Wiersma P, Muñoz-Garcia A, Walker A, Williams JB (2007) Tropical birds have a slow pace of life. Proc Nal Acad Sci USA 104:9340–9345
- Wiersma P, Nowak B, Williams JB (2012) Small organ size contributes to the slow pace of life in tropical birds. J Exp Biol 215:1662–1669
- Williams JB, Tieleman BI (2000) Flexibility in basal metabolic rate and evaporative water loss among hoopoe larks exposed to different environmental temperatures. J Exp Biol 203:3153–3159
- Williams TM, Haun J, Davis RW, Fuiman LA, Kohn S (2001) A killer apetite: metabolic consequences of carnivory in marine mammals. Comp Biochem Physiol A 129:785–796
- Williams JB, Miller RA, Harper JM, Wiersma P (2010) Functional linkages for the pace of life, life history and environment in birds. Integr Comp Biol 50:855–868
- Wone B, Sears MW, Lobacha MK, Donovan ER, Hayes JP (2009) Genetic variances and covariances of aerobic metabolic rates in laboratory mice. Proc R Soc B 276:3695–3704
- Wone B, Donovan ER, Cushman JC, Hayes JP (2013) Metabolic rates associated with membrane fatty acids in mice selected for increased maximal metabolic rate. Comp Biochem Physiol A 165:70–78
- Wone B, Madsen P, Donovan ER, Lobacha MK, Sears MW, Downs CJ, Sorenson DA, Hayes JP (2015) A strong response to selection on mass-independent maximal metabolic rate without a correlated response in basal metabolic rate. Heredity 114:419–427
- Woodley R, Buffenstein R (2002) Thermogenic changes with chronic cold exposure in the naked mole-rat (*Heterocephalus glaber*). Comp Biochem Physiol A 133:827–834
- Yahav S, Buffenstein R (1991) Huddling behavior facilitates homeothermy in the naked mole rat *Heterocephalus glaber*. Physiol Zool 64:871–884
- Zheng W-H, Li M, Liu J-S, Shao S-L (2008a) Seasonal acclimatization of metabolism in Eurasian tree sparrows (*Passer montanus*). Comp Biochem Physiol A 151:519–525
- Zheng W-H, Liu J-S, Jang XH, Fang YY, Zhang G-K (2008b) Seasonal variation in metabolism and thermoregulation in the Chinese bulbul. J Therm Biol 33:315–319
- Zheng W-H, Fang YY, Jiang X-H, Zhang G-K, Liu J-S (2010) Comparison of thermogenic character of liver and muscle in Chinese bulbul *Pycnonotus sinensis* between summer and winter. Zool Res 31:319–327
- Zheng W-H, Fang YY, Jiang X-H, Li M (2013) Geographic variation in basal thermogenesis in little buntings: relationship to cellular thermogenesis and thyroid hormone concentrations. Comp Biochem Physiol A 164:483–490
- Zheng W-H, Liu J-S, Swanson DL (2014a) Seasonal phenotypic flexibility of body mass, organ masses, and tissue oxidative capacity and their relationship to resting metabolic rate in Chinese bulbuls. Physiol Biochem Zool 87:432–444
- Zheng W-H, Li M, Liu J-S, Xu X-J, Shao S-L, Xu X-J (2014b) Seasonal variation of metabolic thermogenesis in Eurasian tree sparrows (*Passer montanus*) over a latitudinal gradient. Physiol Biochem Zool 87:704–718
- Zhou L-M, Xia S-S, Chen Q, Wang R-M, Zheng W-H, Liu J-S (2016) Phenotypic flexibility of thermogenesis in hwamei (*Garrulax canorus*): responses to cold acclimation. Am J Physiol Regul Integr Comp Physiol 310:R330–R336
- Zhu W-L, Zhang H, Wang Z-K (2012) Seasonal changes in body mass and thermogenesis in tree shrews (*Tupaia belangeri*): the roles of photoperiod and cold. J Therm Biol 37:479–484
- Zub K, Borowski Z, Szafransk PA, Wieczorek M, Konarzewski M (2014) Lower body mass and higher metabolic rate enhance winter survival in root voles, *Microtus oeconomus*. Biol J Linnaean Soc 113:297–309