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Which muscarinic receptor is important in the bladder?

Abstract Antimuscarinic agents are the most widely used therapy for urge incontinence, but have side effects such as constipation, tachycardia and dry mouth, resulting from a lack of selectivity for the bladder. M_2 receptors are the predominant cholinergic receptors present in urinary bladder, but mainly the minor population of M_3 receptors mediate its contraction. M_2 receptors modulate detrusor contraction by several mechanisms, and may contribute more to contraction of the bladder in pathological states such as bladder denervation or spinal cord injury. Prejunctional inhibitory M_2 or M_4 receptors and prejunctional facilitatory muscarinic M_1 receptors in the bladder have all been reported. In clinical studies, tolterodine, a non-selective muscarinic antagonist, has been reported to be as effective as oxybutynin but inducing less dry mouth. Thus, although it is not certain which antimuscarinic drugs have the better efficacy and tolerability, the non-selective antimuscarinic drugs seem to be better than M_3 -selective antagonists in their clinical efficacies. However, controlled release, or intravesical, intravaginal, or rectal administrations of oxybutynin have been reported to cause fewer side effects. Darifenacin, a new M_3 selective antagonist, has been reported to have selectivity for the bladder over the salivary gland *in vivo*. To verify which antimuscarinic drugs selective for the muscarinic subtypes have the best efficacy and

tolerability, comparative clinical trials between M_3 selective antagonists and non-selective compounds, such as tolterodine, are required in the future.

Key words muscarinic receptor · urinary bladder · urinary incontinence · therapy

The lower urinary tract has two functions, i.e. storing and emptying urine. Failure to store urine may lead to various forms of incontinence (mainly urge and stress urinary incontinence). A 33–61% prevalence of an overactive bladder in the elderly over the age of 65 years old has been reported [5]. Bladder contraction is predominantly under the control of the parasympathetic nervous system where input is through muscarinic receptors [5, 37].

Antimuscarinic preparations are the most widely used treatment agents for urge incontinence, but these have side effects including accommodation paralysis, constipation, tachycardia, and dryness of mouth [5]. These side effects result from a lack of selectivity for the bladder resulting in actions on other organs such as the iris, intestine, and salivary gland.

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Which muscarinic receptor subtypes are present in the urinary bladder?

The urinary bladder receives cholinergic innervation via the pelvic nerves and adrenergic innervation via the hypogastric nerves. The density of muscarinic receptors is greater in the bladder body than in the base, and cholinergic stimulation produces a contraction of the bladder body of significantly greater magnitude than that of the bladder base [38].

Five different muscarinic subtypes (currently upper case nomenclature M_1 – M_5 has been recommended rather than m_1 – m_5) have been cloned, and M_1 – M_4

subtypes correlate well with the M₁–M₄ gene products pharmacologically [15]. M₁ receptors prevail in neuronal tissues (cerebral cortex, hippocampus, sympathetic ganglia) and are also present in glands. M₂ receptors are present in the heart, hindbrain and smooth muscle. M₃ receptors prevail in exocrine glands and are also found in smooth muscle and the brain. M₄ receptors are found in the basal forebrain and striatum. It has been reported that M₅ receptors are expressed in the substantia nigra [15]. A pharmacological correlate of the M₅ gene has been defined, but because of the similarity in pharmacological profiles of M₅ and M₃ subtypes and the current lack of a high affinity M₅ selective antagonist, the identification of a functional correlate has been complicated [15, 21, 74].

The urinary bladder, like the majority of other smooth muscles from many species exhibit heterogeneous populations of muscarinic receptors [29, 40]. In studies employing northern blot hybridization analysis, the presence of mRNA encoding the M₂ and M₃ subtypes, but not the M₁, M₂, M₄ receptors, has been identified in the bladder of the rat and pig [42]. In the reverse transcriptase-polymerase chain reaction experiments the presence of only M₂ and M₃ subtypes, with a ratio of 1.06:1 has been detected in human urinary bladder [76], but Braverman et al. [9] identified the presence of M₁, M₂, M₃, and M₄ transcripts in rat bladder. At protein level using receptor binding, M₁, M₂, and M₃-receptors have been detected in human detrusor muscle [35], but other studies have detected only M₂ receptors [25] or a mixed M₂ and M₃ receptor population [47, 77]. Similarly in immunoprecipitation studies, only the M₂ and M₃ subtypes have been precipitated in rat, rabbit, guinea pig, and human [73].

Thus, a predominance of the M₂ muscarinic receptor subtype, with a minor population of M₃-receptors has been reported for urinary bladder smooth muscle for several species. Immunoprecipitation data, subtype-selective antisera, and radioligand binding studies all indicate that the proportion of muscarinic M₂ and M₃ receptors is approximately 9:1 in the rat bladder [47, 70, 73], and approximately 3:1 in bladders of humans, guinea pigs, rabbits, and pigs [77, 73].

The functional role of muscarinic receptor subtypes for urinary bladder *in vitro*

M₃-muscarinic receptors

Elucidation of the muscarinic receptor subtypes responsible for mediating detrusor responses to cholinergic agonists has been hampered by the lack of subtype-selective agonists and antagonists [40]. However, pharmacological characterisation of muscarinic receptors mediating contraction of detrusor muscle in rat [40], rabbit [16, 48], guinea-pig [52], pig [58], and human [20, 78] bladder suggests the singular involvement of M₃

receptors. The best correlation between the antagonist affinities at the muscarinic receptor in rabbit [16], pig [58] and human [19, 78] bladders and the affinities at human recombinant receptors has been obtained at M₃ receptors. A significant correlation has also been found with the M₅ receptors which reflects the lack of selective muscarinic antagonists which can discriminate M₃ and M₅ subtypes [16]. However, the correlation is better at M₃ than M₅ subtypes when using darifenacin and oxybutynin [58, 78], agents which display some selectivity for M₃ receptors over the M₅ subtype [21, 74]. Furthermore, the M₅ gene has not been identified in the bladder [9, 76, 73]. The predominant role of the M₃ subtype in mediating contraction of the bladder has been confirmed by the experiments using mutant mice lacking the receptor gene for the M₃ subtype [44]. Although the hormonal state and gender may influence the sensitivity of the bladder to muscarinic stimulation, there are no differences in the affinity (pA₂) values of muscarinic antagonists, indicating that M₃ receptors mediate the contraction of the rat urinary bladder [39].

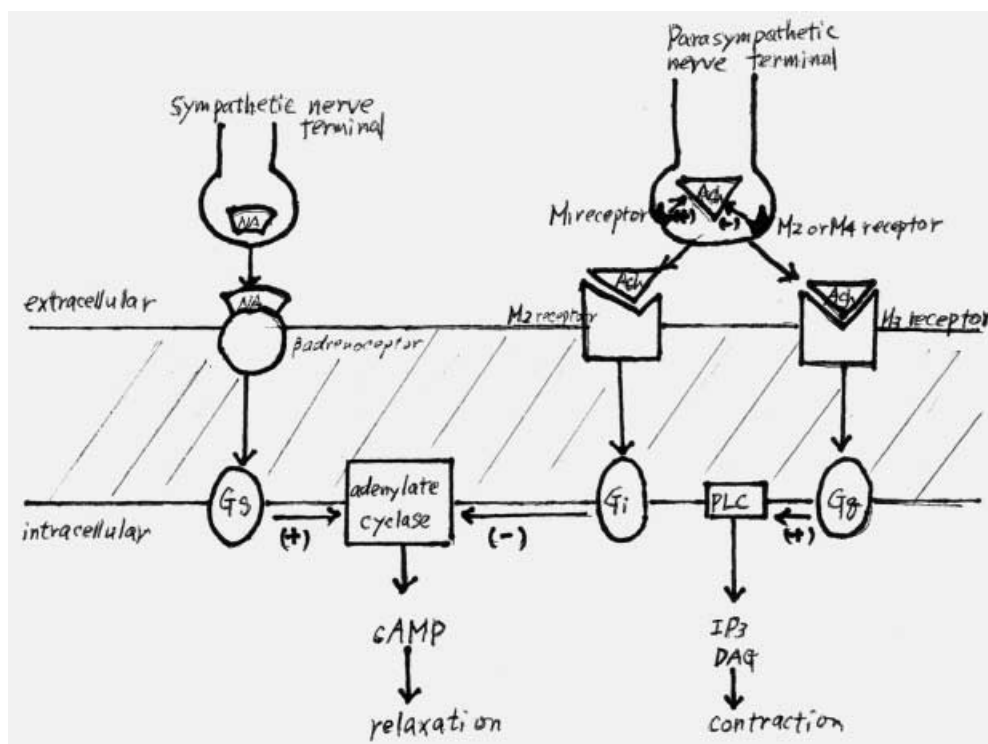
These observations suggest that it is a minor population of M₃-receptors which mediates contraction of the detrusor muscle and that M₂ receptors are not directly involved in contraction. Muscarinic M₃-receptor stimulation has been shown to stimulate phosphoinositide hydrolysis causing release of intracellular calcium in guinea pig [52] and human [4, 27] bladder, and this is most likely the signaling mechanism responsible for the direct contractile responses to muscarinic agonists in this tissue [29] (Fig. 1).

Dry mouth being the most common, the adverse effects of antimuscarinic drugs in the treatment of overactive bladder may lead to withdrawal of medication [1]. Thus, drugs which have more selectivity for the bladder is desirable. Because M₃ receptor subtypes have been reported to mediate salivary gland secretion, drugs selective for M₃ receptors have been considered to cause dry mouth. Indeed, oxybutynin, a selective M₃ receptor antagonist, has caused more dry mouth than tolterodine, a non-selective antimuscarinic agent [1, 6]. In radioligand binding studies, oxybutynin has been reported to have a higher affinity for muscarinic receptors in the parotid gland than in the bladder, and darifenacin (a M₃ selective antagonist) a two fold higher affinity in parotid gland than in the guinea pig bladder [24] (Table 1). However, Wallis and Napier [71] have reported that darifenacin exhibits functional tissue selectivity for intestinal smooth muscle over the salivary gland. They have also suggested a role for M₅ receptors in the control of salivary secretion [71], although this has been disputed [21].

M₂-muscarinic receptors

M₂ receptors couple to the pertussis toxin-sensitive guanine nucleotide regulatory protein G_i and inhibit adenylyl cyclase activation [14, 22, 26, 64] (Fig. 1). Although

Fig. 1 The role of prejunctional and postjunctional muscarinic receptor subtypes in urinary bladder. *NA* noradrenaline, *ACh* acetylcholine, *PLC* phospholipase C, *cAMP* adenosine 3',5'-cyclic monophosphate, *IP₃* inositol (1,4,5)-triphosphate, *DAG* diacylglycerol



no direct contractile response of detrusor smooth muscle to M_2 receptor activation can be demonstrated, an indirect influence on contraction via inhibition of cAMP-mediated smooth muscle relaxation by β -adrenoceptors, forskolin, 5-hydroxytryptamine, or vasoactive intestinal peptide has been reported [14, 23, 26, 53].

M_2 -receptors have also been shown to activate smooth muscle by decreasing the probability of opening K^+ -channels [8, 49]. Other postulated mechanisms of M_2 -mediated contraction include opening of non-specific cation channels resulting in depolarisation and calcium influx, and stimulation of Rho proteins causing Ca^{2+} sensitization [15, 28, 33, 66].

Recently, an M_2 -mediated contraction from muscarinic stimulation has been demonstrated following selective M_3 -receptor inactivation and elevation of cAMP levels in guinea-pig ileum [53, 64], and in the rat [29] and

pig [77] bladder, an effect which manifests as a re-contraction.

Thus, M_2 -receptors may mediate the dominant parasympathetic control over smooth muscle tone under conditions of high sympathetic activity or where M_3 -receptors are dysfunctional [22]. β -Adrenoceptors predominate over α -adrenoceptors in the urinary bladder body, where their tonic stimulation is thought to facilitate the storage phase of micturition by relaxing the detrusor smooth muscle [38]. Cholinergic activity is inhibited during this filling phase. In contrast, during the voiding phase, sympathetic nerve activity is inhibited and acetylcholine is released. Activation of M_2 receptors during the voiding phase may oppose inhibitory sympathetic activation via β -adrenoceptors, resulting in more efficient bladder emptying or initiation of voiding [40, 51, 77]. Furthermore, M_2 -receptors have been reported to directly contribute to contraction of the rat denervated urinary bladder following bilateral major pelvic ganglion removal or spinal cord compression at T9 level [10, 11]. However, Krichevsky et al. [36] have reported that the affinity (pA_2) values for the M_3 -selective antagonist 4-DAMP are not significantly different in normal and obstructed rat urinary bladder, suggesting that M_3 -receptors still mediate contraction of the bladder with bladder outlet obstruction.

Recently, the role of M_2 and M_3 receptors in mediating contraction of the urinary bladder has been confirmed in experiments using muscarinic receptor subtype knockout mice. Stengel et al. [62] have reported that carbachol-induced bladder contractions in the M_4 receptor knockout mice are similar to wild-type littermates, suggesting that M_4 receptors do not participate in

Table 1 Affinity estimates (pK_{is}) of antagonists in radioligand binding assays at recombinant human muscarinic receptor subtypes expressed in CHO cells. Values are mean K_{is} estimated in a Tris-Krebs

Antagonist	M_1	M_2	M_3	M_4	M_5
Atropine ^a	9.1	8.9	9.5	9.2	9.1
4-DAMP ^a	9.2	8.1	9.3	8.4	8.9
Darifenacin ^a	7.8	7.0	8.9	7.7	8.1
Methoctramine ^a	6.6	7.6	6.1	6.9	6.4
Oxybutynin ^b	8.5	7.8	8.7	8.2	7.6
Pirenzepine ^a	8.0	6.3	6.8	7.0	6.9
Tolterodine ^b	8.5	8.4	8.5	8.1	8.6

^a Data from Hegde et al [28]

^b Data from Eglén and Nahorski [21]

bladder contraction. In contrast, in urinary bladders from M_2 receptor knockout mice, the potency of carbachol was significantly reduced by a factor of ~2, and the affinity of AF-DX116 (an M_2 selective antagonist) in inhibiting carbachol-induced bladder was significantly reduced compared with wild-type littermates. The authors concluded that M_3 receptors are the predominant muscarinic subtype that mediate contractions of smooth muscle including the urinary bladder, and that M_2 but not M_4 receptors also play a role in carbachol-induced contractions of the bladder.

Matsui et al. [44] have reported that mutant mice lacking the receptor gene for the M_3 subtype show impaired salivary secretion to cholinergic stimuli, urinary retention in male mutants, and greatly reduced detrusor contractions from carbachol stimulation (5% of those obtained in the wild-type muscles), suggesting that M_3 receptors play key roles in salivary secretion and bladder contraction. They also reported that the potency of a number of antagonists that block the residual carbachol-induced contractions in these mutant mice is consistent with M_2 receptors in both sexes, suggesting that the M_2 receptors directly mediate the residual contraction of the detrusor of the M_3 -knockout mice.

The functional role of muscarinic receptor subtypes in vivo

Although many investigators report that M_3 receptors are the predominant receptor subtype mediating contraction of the bladder in vitro [19, 58, 78], the role of the other muscarinic receptor subtypes in vivo remains to be established. Hegde et al. [29] have reported that the in vivo potency of muscarinic antagonists in rat bladder correlates best with the affinity of such agents for M_2 receptors, and that pretreatment with propranolol (a β -receptor antagonist) decreases the potency of methoctramine (an M_2 receptor antagonist) but not that of darifenacin (an M_3 receptor antagonist) implying an obligatory role for β -adrenoceptors in M_2 -receptor mediated effects.

Nilvebrant et al. [51] and Gillberg et al. [24] have shown that tolterodine (a non-selective muscarinic antagonist), oxybutynin (an M_1 and M_3 antagonist) and darifenacin (an M_3 selective antagonist) are equally effective at inhibiting bladder contraction in vivo, but that tolterodine is more potent at inhibiting contraction of the bladder than salivation in the cat. In contrast, oxybutynin and darifenacin have more effect on salivation than on bladder contraction, suggesting that a selectivity for M_3 over M_2 receptors results in a more potent effect on salivation than on bladder contraction in vivo. Interestingly, similar profiles have been demonstrated for tolterodine and AQ-RA 741 (an M_2 selective antagonist); these drugs at low doses are more potent at inhibiting bladder contraction than salivation in the anesthetized cat. These results suggest that M_2

receptors have a role in bladder contraction in vivo [24]. From these in vivo data it has been suggested that it is possible to separate the effects of muscarinic antagonists on the bladder and salivary gland [24, 51]. Newgreen et al. [50] have reported that darifenacin and tolterodine have an 9 and 5 fold selectivity, respectively, for the bladder over the salivary glands in the dog in vivo. Wallis and Napier [74] have also reported that darifenacin is more effective in inhibiting salivary secretion than in reducing micturition pressure in the conscious rat.

Characterisation of presynaptic muscarinic receptor subtypes in the urinary bladder

Although the presence of M_1 , M_2 and M_4 receptors has been questioned, especially in human bladder [71], the presence of M_1 , M_2 , M_3 , and M_4 receptors has been identified by reverse transcriptase-polymerase chain reaction in the rat bladder [9]. The activation of inhibitory presynaptic M_2 or M_4 receptors and facilitatory presynaptic M_1 receptors on cholinergic terminals in urinary bladder have been identified in studies investigating [3 H] acetylcholine (ACh) release and contractile response to electrical field stimulation. D'Agostino et al. [17] have reported the presence of inhibitory presynaptic muscarinic receptors in rat bladder, while Somogi and deGroat [59] have characterised this inhibitory presynaptic receptor as the M_2 -receptor subtype. Somogi et al. [61, 60] have also shown that continuous stimulation of postganglionic cholinergic nerves leads to the activation of presynaptic facilitatory M_1 receptors which enhance [3 H]ACh release in the rat, cat, and human bladder. This facilitatory mechanism is thought to be upregulated after spinal cord injury and mediated by M_3 subtype [60]. The prejunctional facilitatory M_1 receptors and inhibitory M_2 receptors regulating acetylcholine release and contractions from electric field stimulation have also been reported in rat and rabbit bladder [9, 30]. However, Alberts [2] has reported that the potency of a number of muscarinic antagonists at enhancing the secretion of [3 H]ACh in the guinea pig and rat bladder correlate best with values for the M_4 muscarinic receptor subtype. This suggests the prevalence of a presynaptic M_4 receptor, a muscarinic receptor subtype that is not coupled to adenylate cyclase. D'Agostino et al. [18, 19] have confirmed the role of presynaptic M_4 receptors in rat and human bladder, using new muscarinic antagonists that can distinguish between M_2 and M_4 receptors (Fig. 1).

Clinical effectiveness and muscarinic subtype selectivity of anticholinergic drugs for the treatment of overactive bladder

Propantheline bromide, emepronium bromide and trospium chloride, are quaternary ammonium compounds,

and are non-selective anti-muscarinic drugs. *Propantheline bromide* and *tropium chloride* also have ganglionic blocking action. Although the clinical effects of these non-selective anticholinergics have been reported [7, 13, 43], several reports have shown insufficient efficacy for treatment of urinary incontinence [55, 65, 72]. However, *tropium chloride* has been found to be effective in the treatment of detrusor hyperreflexia in a placebo-controlled double-blind study; it was as effective as oxybutynin but with fewer side effects [41, 63].

Recently, the tertiary amines such as oxybutynin, propiverine, and tolterodine have been commonly used for bladder overactivity.

Oxybutynin chloride has seven- to tenfold higher affinity for M_1 - and M_3 -receptors than for M_2 -receptors [24, 51]. It has been used for the treatment of urge incontinence for more than 20 years [3] and its efficacy has been established in double-blind studies [45, 65]. It also has a direct muscle-relaxant (spasmolytic) effect, calcium antagonistic actions and local anaesthetic actions [32]. However, it has been reported that the concentrations required for its direct smooth muscle relaxant effects are greater (500 fold) than those at which its antimuscarinic effects are exerted [32]. Also the effects of oxybutynin on the bladder *in vivo* correlate significantly only with its antimuscarinic activity, and do not appear to be related to its direct smooth muscle relaxant effects or local anaesthetic effects [52]. Thus when given systemically, oxybutynin acts mainly as an antimuscarinic drug [5]. Oxybutynin has higher affinity for muscarinic receptors in the parotid gland than the bladder in radioligand binding studies, and has more selectivity for salivation than for bladder contraction *in vivo* [51].

Oral oxybutynin has an extensive first-pass metabolism. Its metabolite, N-desethyl-oxybutynin is equally potent as oxybutynin in terms of antimuscarinic activity and selectivity profile *in vitro* [69]. The serum concentrations of N-desethyl-oxybutynin are two- to fivefold higher and persist longer than the parent compound. The affinity of N-desethyl-oxybutynin for the muscarinic receptors of the parotid gland is significantly higher than for those of the bladder [69], and it has been suggested that the adverse effects of oxybutynin may be mediated to a large extent by N-desethyl-oxybutynin [12, 57, 69]. The optimal application of oxybutynin would allow continuous absorption without variations in serum levels [57]. Thus, intravesical oxybutynin [12], oxybutynin rectal suppositories [75], vaginal inserts [57] and oxybutynin patches are considered to cause fewer side effects due to reduced plasma concentration of this metabolite. Similarly, controlled release oxybutynin chloride, a slow release formation of oxybutynin, has also been reported to have an equivalent effects on urge incontinence yet cause less dry mouth [3].

Propiverine has a dual mode of action which includes both non-selective antimuscarinic and calcium antagonistic properties. These two effects occur at similar concentrations of propiverine in rat and guinea pig bladder *in vitro* [67].

Tolterodine is a new drug for the treatment of overactive bladder. Neither tolterodine nor its major active 5-hydroxymethyl metabolite (5-HM) has any selectivity for muscarinic receptor subtypes [51]. Radioligand binding data has shown that tolterodine has eight times less potency than oxybutynin at the muscarinic receptor in the guinea-pig parotid gland [51]. In functional studies *in vivo*, tolterodine is found to be equipotent to oxybutynin, but has selectivity for the bladder over salivary glands in the cat [51] and dog [50].

The clinical efficacy of tolterodine has been verified in double-blind, placebo-controlled studies [1, 6, 20, 31, 37, 46, 54]. In urodynamic studies, an increase in maximum cystometric capacity and volume at first contraction have been reported for patients with detrusor overactivity [31, 37, 54] and detrusor hyperreflexia [68]. In a placebo-controlled study comparing tolterodine and oxybutynin, these drugs were found to be equally effective in treatment of frequency and incontinence [1]. Tolterodine at a dose of 2 mg twice daily and oxybutynin at 5 mg three times daily have been reported to be equivalent in their effectiveness [6, 20]. However, tolterodine appeared to be better tolerated than oxybutynin, particularly with regard to dry mouth, where 69–80%, 30–50% and 15–21% of patients treated with oxybutynin, tolterodine, and placebo, respectively, reported this side effect [1, 6, 20, 68]. However, cardiovascular side effects would also be anticipated for tolterodine due to its actions at M_2 receptors. Appell [6] has reported, from meta-analysis, no significant difference in the incidence of cardiovascular adverse events among individuals receiving 2 mg tolterodine (4%), 5 mg oxybutynin (6%), and placebo (8%), although 1 mg tolterodine produced a higher incidence (12%) of these events. A slight dose-dependent increase in heart rate has been reported for tolterodine [37, 54]. However, no dose-related changes in ECGs (especially QTc interval) have been demonstrated in tolterodine treated groups [6, 37, 46, 54]. Thus, this drug is considered well-tolerated and cost-effective in the treatment of overactive bladder [34].

Darifenacin has a 7-fold higher affinity for M_3 -receptors than for M_2 -receptors [24], but has a marginal selectivity for the bladder over the salivary gland [50, 71]. The effectiveness of this drug for detrusor instability has been reported as measured by ambulatory urodynamics [56]. Clinical trials to determine its efficacy for overactive bladder are currently being considered.

Discussion

From the experimental results we can consider that M_2 receptors are the predominant receptors present in the urinary bladder, but that M_3 receptors mainly mediate contraction. Since agents with a selective action on the bladder are considered preferable for the treatment of urinary incontinence, the selective M_3 antagonists have been thought to be useful for the treatment of this

condition. However, the side effects such as dry mouth caused by the M₃ selective agent oxybutynin are considerable because M₃ receptors also mediate contraction of the iris and intestine, and secretion of the salivary gland.

Other muscarinic receptor subtypes may also have roles for the contraction of urinary bladder. M₂ receptors modulate detrusor contraction by inhibiting the relaxation mediated by β -adrenoceptors, and they may contribute more to the contraction of the bladder in vivo or with pathological states e.g. bladder denervation or spinal cord injury.

In addition, the prejunctional inhibitory M₂ or M₄ receptors and prejunctional muscarinic M₁ receptors may be important targets for anticholinergic drugs. The role of muscarinic receptor subtypes in the central nervous system is poorly understood, and the actions of muscarinic receptor antagonists in this area should also be investigated. From this standpoint, non-selective muscarinic antagonists such as tolterodine may be more effective, with fewer of those side effects that are mediated by M₃ receptors, although non-selective antagonists may have other side effects: M₂-receptor antagonism may have effects on heart rate and M₁-receptor antagonism on gastric acid secretion and cognitive function [71]. Thus, although it is not known which antimuscarinic drugs selective for the muscarinic subtypes have better efficacy and tolerability, the non-selective antimuscarinic drugs seem to be better than M₃-selective antagonists in clinical efficacy.

However, with regard to M₃ selective antagonists, controlled release, or intravesical, intravaginal, or rectal (suppository) administration have been reported to cause fewer side effects, at least for oxybutynin. Darifenacin, a new M₃ selective antagonist has been reported to have selectivity for the bladder over the salivary gland in vivo [50, 71]. A clinical study to determine its efficacy and safety has been under consideration.

To verify which antimuscarinic drugs selective for the muscarinic subtypes have best efficacy and tolerability, comparative clinical trials between M₃ selective antagonists, such as darifenacin or controlled release oxybutynin, and non-selective compounds, such as tolterodine, are required in the future [71].

References

- Abrams P, Freeman R, Anderström C, Matiasson A (1998) Tolterodine, a new antimuscarinic agent: as effective but better tolerated than oxybutynin in patients with an overactive bladder. *BJU Int* 81: 801–810
- Alberts P (1995) Classification of the presynaptic muscarinic receptor subtype that regulates ³H-Acetylcholine secretion in the guinea pig urinary bladder in vitro. *J Pharmacol Exp Ther* 274: 458–468
- Anderson RU, Mobley D, Blank B, Saltzsten D, Susset J, Brown JS (1999) Once daily controlled versus immediate release oxybutynin chloride for urge urinary incontinence. *J Urol* 161: 1809–1812
- Andersson K-E, Holmquist F, Fovaeus M, Hedlund H, Sundler R (1991) Muscarinic receptor stimulation of phosphoinositide hydrolysis in the human isolated urinary bladder. *J Urol* 146: 1156–1159
- Andersson K-E, Appell R, Cardozo LD, Chapple C, Drutz HP, Finkbeiner AE, Haab F, Navarrete V (1999) The pharmacological treatment of urinary incontinence. *BJU Int* 84: 923–947
- Appell RA (1997) Clinical efficacy and safety of tolterodine in the treatment of overactive bladder: a pooled analysis. *Urology* 50 Suppl: 90–96
- Blaivas JG, Labib KB, Michalik SJ, Zayed AAH (1980) Cystometric response to propantheline in detrusor hyperreflexia: therapeutic implications. *J Urol* 124: 259–262
- Bonev AD, Nelson MT (1993) Muscarinic inhibition of ATP-sensitive K⁺ channels by protein kinase C in urinary bladder smooth muscle. *Am J Physiol* 265: C1723–C1728
- Braverman AS, Kohn IJ, Luthin GR, Ruggieri MR (1998) Prejunctional M₁ facilitatory and M₂ inhibitory muscarinic receptors mediate rat urinary bladder contractility. *Am J Physiol* 274: R517–R523
- Braverman AS, Luthin GR, Ruggieri MR (1998) M₂ muscarinic receptor contributes to contraction of the denervated rat urinary bladder. *Am. J. Physiol.*, 275: R1654–R1660
- Braverman AS, Legos J, Young W, Luthin G, Ruggieri M (1999) M₂ receptors in genito-urinary smooth muscle pathology. *Life Sci* 64: 429–436
- Buyse G, Waldeck K, Verpoorten C, Bjork H, Casaer P, Andersson K-E (1998) Intravesical oxybutynin for neurogenic bladder dysfunction: less systemic side effects due to reduced first pass metabolism. *J. Urol* 160: 892–896
- Cardozo LD, Stanton SL (1979) An objective comparison of the effects of parenterally administered drug in patients suffering from detrusor instability. *J Urol* 122: 58–59
- Caulfield MP (1993) Muscarinic receptors – characterization, coupling and function. *Pharmacol Ther* 58: 319–379
- Caulfield MP, Birdsall NJM (1998) International union of pharmacology: XVII. Classification of muscarinic acetylcholine receptors. *Pharmacol Rev* 50: 279–290
- Choppin A, Eglén RM, Hegde SS (1998) Pharmacological characterization of muscarinic receptors in rabbit isolated iris sphincter muscle and urinary bladder smooth muscle. *Br J Pharmacol* 124: 883–888
- D'agostino G, Kilbinger H, Chiari MC, Grana E (1986) Presynaptic inhibitory muscarinic receptors modulating [³H] acetylcholine release in the rat urinary bladder. *J Pharmacol Exp Ther* 239: 522–528
- D'agostino G, Barbieri A, Chiossa E, Tonini M (1997) M₄ muscarinic autoreceptor-mediated inhibition of [³H] acetylcholine release in the rat isolated urinary bladder. *J Pharmacol Exp Ther*. 283:750–756
- D'agostino G, Bolognesi ML, Lucchelli A, Vicini D, Balestra B, Spelta V, Melchiorre C, Tonini M (2000) Prejunctional muscarinic inhibitory control of acetylcholine release in the human isolated detrusor: involvement of the M₄receptor subtype. *Br J Pharmacol* 129: 493–500
- Drutz H, Appell RA, Gleason D, Klimberg I, Radomski S (1999) Clinical efficacy and safety of tolterodine compared to oxybutynin and placebo in patients with overactive bladder. *Int Urogynecol J Pelvic Floor Dysfunct* 10: 283–289
- Eglén, RM, Nahorski SR (2000) The muscarinic receptor: a silent or emerging subtype? *Br J Pharmacol* 130: 13–20
- Eglén RM, Reddy H, Watson N, Challis RAJ (1994) Muscarinic acetylcholine receptor subtypes in smooth muscle. *Trends. Pharmacol Sci* 15: 114–119
- Eglén, RM, Hegde SS, Watson, N (1996). Muscarinic receptor subtypes and smooth muscle function. *Pharmacol Rev* 15: 114–119
- Gillberg PG, Sundquist S, Nilvebrant L (1998) Comparison of the in vitro and in vivo profiles of tolterodine with those of subtype-selective muscarinic receptor antagonists. *Eur J Pharmacol* 349: 285–292
- Goepel M, Wittmann A, Rübber H, Michel MC (1997) Comparison of adrenoceptor subtype expression in porcine and human bladder and prostate. *Urol Res* 25: 199–206

26. Griffin MT, Ehlert FJ (1992) Specific inhibition of isoproterenol-stimulated cyclic AMP accumulation by M₂ muscarinic receptors in rat intestinal smooth muscle. *J Pharmacol Exp Therap* 263: 221–225
27. Harriss DR, Marsh KA, Birmingham AT, Hill SJ (1995) Expression of muscarinic M₃-receptors coupled to inositol phospholipid hydrolysis in human detrusor cultured smooth muscle cells. *J Urol* 154: 1241–1245
28. Hegde SS, Eglén RM (1999) Muscarinic receptor subtypes modulating smooth muscle contractility in the urinary bladder. *Life Sci* 64: 419–428
29. Hegde SS, Choppin A, Bonhaus D, Briaud S, Loeb M, Moy TM, Loury D, Eglén RM (1997). Functional role of M₂ and M₃ muscarinic receptors in the urinary bladder of rats in vitro and in vivo. *Br J Pharmacol* 120: 1409–1418
30. Inadome A, Yoshida M, Takahashi W, Yono M, Seshita H, Miyamoto Y, Kawano T, Ueda S (1998) Prejunctional muscarinic receptors modulating acetylcholine release in rabbit detrusor smooth muscles. *Urol Int* 61: 135–141
31. Jonas U, Höfner K, Madersbacher H, Holmdahl TH (1997) Efficacy and safety of two doses of tolterodine versus placebo in patients with detrusor overactivity and symptoms of frequency, urge incontinence and urgency: urodynamic evaluation. *World J Urol* 15: 144–151
32. Kachur JF, Peterson JS, Carter JP, Rzeszutarsky WJ, Hanson RC, Noronha-Blob L (1988) R and S enantiomers of oxybutynin: pharmacological effects in guinea pig bladder and intestine. *J Pharmacol Exp Ther* 247: 867–872
33. Kim SJ, Koh EM, Kang TM, Kim YC, So I, Isenberg G, Kim KW (1998) Ca²⁺ influx through carbachol-activated non-selective cation channels in guinea-pig gastric myocytes. *J Physiol* 513: 749–760
34. Kobelt G, Jönsson L, Mattiasson A (1998) Cost-effectiveness of new treatments for overactive bladder: the example of tolterodine, a new muscarinic agent: a Markov model. *Neurourol Urodyn* 17: 599–611
35. Kondo S, Morita T, Tashima Y (1995) Muscarinic cholinergic receptor subtypes in human detrusor muscle studied by labeled and nonlabeled pirenzepine, AFDX-116 and 4DAMP. *Urol Int* 54: 150–153
36. Krichevsky VP, Pagala MK, Vaydovsky I, Damer V, Wise GJ (1999) Function of M₃ muscarinic receptors in the rat urinary bladder following partial outlet obstruction. *J Urol* 161: 1644–1650
37. Larsson G, Hallén B, Nilvebrant L (1999) Tolterodine in the treatment of overactive bladder: analysis of the pooled phase II efficacy and safety data. *Urology* 53: 990–998
38. Levin RM, Ruggieri, MR, Wein AJ (1988). Identification of receptor subtypes in the rabbit and human urinary bladder by selective radio-ligand binding. *J Urol* 139: 844–848
39. Longhurst PA, Levendusky M (2000) Influence of gender and the oestrous cycle on *in vitro* contractile responses of the rat urinary bladder to cholinergic stimulation. *Br J Pharmacol* 131: 177–184
40. Longhurst PA, Leggett RE, Briscoe AK (1995) Characterization of the functional muscarinic receptors in the rat urinary bladder. *Br J Pharmacol* 116: 2279–2285
41. Madersbacher H, Stöhrer M, Richter R, Burgdörfer H, Hachen HJ, Mürtz G (1995) Trospium chloride versus oxybutynin: a randomized, double-blind, multicentre trial in the treatment of detrusor hyper-reflexia. *Br J Urol* 75: 452–456
42. Maeda A, Tubo T, Mishima M, Numa S (1988) Tissue distribution of mRNAs encoding muscarinic acetylcholine receptor subtypes. *FEBS Lett* 239: 339–342
43. Massey JA, Abrams P (1986) Dose titration in clinical trials. An example using emepromonium carrageenate in detrusor instability. *Br J Urol* 58: 125–128
44. Matsui M, Motomura D, Karasawa H, Fujikawa T, Jiang J, Komiya Y, Takahashi S, Takeo MM (2000) Multiple functional defects in peripheral autonomic organs in mice lacking muscarinic acetylcholine receptor gene for the M₃ subtype. *P Natl Acad Sci USA* 97: 9577–9584
45. Milani R, Scalabrino S, Milia R (1993) Double-blind crossover comparison of flavoxate and oxybutynin in women affected by urinary urge syndrome. *Int Urogynecol J* 4: 3–8
46. Millard R, Tuttle J, Moore K, Susset J, Clarke B, Dwyer P, Davis BE (1999) Clinical efficacy and safety of tolterodine compared to placebo in detrusor overactivity. *J Urol* 161: 1551–1555
47. Monferini E, Giraldo E, Ladinsky H (1988) Characterization of the muscarinic receptor subtypes in the rat urinary bladder. *Eur J Pharmacol* 147: 453–458
48. Mutoh S, Latifpour J, Saito M, Weiss RM (1997) Evidence for the presence of regional differences in the subtype specificity of muscarinic receptors in rabbit lower urinary tract. *J Urol* 157: 717–721
49. Nakamura T, Yamaguchi O, Kimura J (1998) Muscarinic M₂ receptors regulate Ca²⁺-activated K channels in bladder smooth muscle cells. *Neurourol Urodyn* 17: 400–401
50. Newgreen DT, Anderson CWP, Carter AJ, Naylor AM (1995) Darifenacin – a novel bladder-selective agent for the treatment of urge incontinence. *Neurourol Urodyn* 14: 555–557
51. Nilvebrant L, Andersson K-E, Gillberg P-G, Stahl M, Sparf B (1997) Tolterodine – a new bladder selective antimuscarinic agent. *Eur J Pharmacol* 327: 195–207
52. Noronha-Blob L, Lowe VC, Peterson JS, Hanson RC (1989) Anticholinergic activity of agents indicated for urinary incontinence is an important property for effective control of bladder dysfunction. *J Pharmacol Exp Ther* 251: 586–593
53. Reddy H, Watson N, Ford APDW, Eglén RM (1995) Characterization of the interaction between muscarinic M₂ receptors and β -adrenoceptor subtypes in guinea-pig isolated ileum. *Br J Pharmacol* 114: 49–56
54. Rentzhog L, Stanton SL, Cardozo L, Nelson E, Fall M, Abrams P (1998) Efficacy and safety of tolterodine in patients with detrusor instability: a dose ranging study. *Br J Urol* 81: 42–48
55. Ritch AES, Castleden CM, George CF, Hall MRP (1977) A second look at emepromonium bromide in urinary incontinence. *Lancet* 1: 504–506
56. Rosario DJ, Leaker BR, Smith DJ, Chapple CR (1999) A pilot study of the effects of multiple doses of the M₃ muscarinic receptor activity in patients with detrusor instability. *Neurourol Urodyn* 14: 464–465
57. Schröder A, Levin RM, Kogan BA, Das AK, Kay F, Mahashabde A (2000) Absorption of oxybutynin from vaginal inserts: drug blood levels and the response of the rabbit bladder. *J Urol* 56: 1063–1067
58. Sellers DJ, Yamanishi T, Chapple CR, Couldwell C, Yasuda K, Korstanje C, Chess-Williams R (2001) M₃ muscarinic receptors but not M₂ mediate contraction of the porcine detrusor muscle in vitro. *J Auton Pharmacol* in press
59. Somogyi GT, de Groat WC (1992) Evidence for nicotinic and facilitatory muscarinic receptors in cholinergic nerve terminals of the rat urinary bladder. *Auton Nerv Syst* 37: 89–98
60. Somogyi GT, de Groat WC (1999) Function, signal transduction mechanisms and plasticity of presynaptic muscarinic receptors in the urinary bladder. *Life Sci* 64: 411–418
61. Somogyi GT, Tanowitz M, de Groat WC (1994) M₁ muscarinic receptor-mediated facilitation of acetylcholine release in the rat urinary bladder. *J Physiol* 140: 81–89
62. Stengel PW, Gomeza J, Wess J, Cohen ML (2000) M₂ and M₄ receptor knockout mice: muscarinic receptor function in cardiac and smooth muscle in vitro. *J Pharmacol Exp Ther* 292: 877–885
63. Stöhrer M, Bauer P, Giannetti BM, Richter R, Burgdörfer H, Mürtz G (1991) Effect of trospium chloride on urodynamic parameters in patients with detrusor hyperreflexia due to spinal cord injuries: a multicentre placebo controlled double-blind trial. *Urol Int* 47: 138–143
64. Thomas EA, Baker SA, Ehlert FJ (1993) Functional role for the M₂ muscarinic receptor in smooth muscle of guinea pig ileum. *Mol Pharmacol* 44: 102–110
65. Thüroff JW, Bunke B, Ebner A, Faber P, de Geeter P, Hannappel J, Heidler H, Madersbacher H, Melchior H,

- Schäfer W, Schwenzer T, Stöckle M (1991) Randomized, double-blind, multicenter trial on treatment of frequency, urgency and incontinence related to detrusor hyperactivity: oxybutynin versus propantheline versus placebo. *J Urol* 145: 813–817
66. Togashi H, Emala CW, Hall IP, Hirshman CA (1998) Carbachol-induced actin reorganization involves Gi activation of Rho in human airway smooth muscle cells. *Am J Physiol* 274: L 803-L809
67. Tokuno H, Chowdhury JU, Tomita T (1993) Inhibitory effects of propiverine on rat and guinea-pig urinary bladder muscle. *Naunyn-Schmiedeberg's Arch Pharmacol* 348: 659–662
68. van Kerrebroek PEVA, Amarenco G, Thüroff JW, Madersbacher HG, Lock MTWT, Messelink EJ, Soler JM (1998) Dose finding study of tolterodine in patients with detrusor hyperreflexia. *Neurourol Urodyn* 17: 499–512
69. Waldeck K, Larsson B, Andersson K-E (1997) Comparison of oxybutynin and its active metabolite, N-desethyl-oxybutynin, in the human detrusor and parotid gland. *J Urol* 157: 1093–1097
70. Wall SJ, Yasuda RP, Li M, Wolfe BB (1991) Development of an antiserum against m3 muscarinic receptors: distribution of m3 receptors in rat tissues and clonal cell. *Mol Pharmacol* 40: 783–789
71. Wallis RM, Napier CM (1999) Muscarinic antagonists in development for disorders of smooth muscle function *Life Sci* 64: 395–401
72. Walter S, Hansen J, Hansen L, Maegaard E, Meyhoff HH, Nordling J (1982) Urinary incontinence in old age. A controlled trial of emepronium bromide. *Br J Urol* 54: 249–251
73. Wang P, Luthin GR, Ruggieri MR (1995) Muscarinic acetylcholine receptor subtypes mediating urinary bladder contractility and coupling to GTP binding proteins. *J Pharmacol Exp Therap* 273: 959–966
74. Watson N, Daniels DV, Ford APDW, Eglen RM, Hegde SS (1999) Comparative pharmacology of recombinant human M₃ and M₅ muscarinic receptors expressed on CHO-K1 cells. *Br J Pharmacol* 127: 590–596
75. Winkler HA, Sand PK (1998) Treatment of detrusor instability with oxybutynin rectal suppositories. *Int Urogynecol J* 9: 100–102
76. Yamaguchi O, Shishido K, Tamura K, Ogawa T, Fujimura T, Ohshita M (1996) Evaluation of mRNAs encoding muscarinic receptor subtypes in human detrusor muscle. *J Urol* 156: 1208–1213
77. Yamanishi T, Yasuda K, Chapple CR, Chess-Williams R (2000) The role of M2-muscarinic receptors in mediating contraction of the pig urinary bladder in vitro *Br J Pharmacol* 131: 1482–1488
78. Yamanishi T, Sellers DJ, Chess-Williams R, Yasuda K, Chapple CR (2000) Does the minor population of M₃ receptors in human detrusor muscle mediate contractions? *BJU Int* 86 (Suppl 3): 244