5-Aminolevulinic Acid-Induced Heavy Metal Stress Tolerance and Underlying Mechanisms in Plants

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Abstract

Plants face different types of biotic and abiotic stresses during their life span. Heavy metal (HM) stress is considered as one of the most challenging and emerging threats to sustainable agricultural development and overall economic yield of various plant species. Increasing levels of HMs in arable soils is a main environmental issue due to their deleterious effects on plant growth and productivity. The exogenous application of different plant growth regulators is a well-known strategy to alleviate the adverse effects of HMs stress on plants. In the present review, the role of 5-aminolevulinic acid (ALA) in the alleviation of HM stress in different plants is elaborated. 5-Aminolevulinic acid is identified as a highly efficient ameliorating agent to sustainably neutralize the harmful effects of abiotic stresses in plants. In particular, the role of ALA has been increasingly recognized in improving plant HM stress-tolerance via ALA-mediated control of principal plant-metabolic processes. However, various underlying mechanisms that unravel ALA-induced plant HM stress-tolerance remain unexplored. The application of ALA on HM-stressed plants improves plant height, root length, chlorophyll pigments, antioxidant enzyme activities, nutrient uptake and soluble protein contents and minimizes ultra-structural damage, oxidative stress and HM uptake. Furthermore, it triggers modification of glutathione reductase, ascorbic acid and GSH contents in HM-stressed plants. The lower concentration of ALA proved to be more beneficial in stress amelioration. The cost-effectiveness and efficiency of ALA in improving growth and production of plants under varying growth conditions is still not clear. Nevertheless, over-accumulation of ALA through genetic manipulation can enhance stress-tolerance in plants which is the key area to be investigated. This review article elaborates the potential role of ALA in HM tolerance and highlights the future research dimensions in the related ambits.

Keywords 5-Aminolevulinic acid · Heavy metal stress · Antioxidant enzymes · Nutrients uptake · Stress-tolerance mechanisms

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Introduction

The group of elements in the periodic table having densities greater than 5 $g/cm³$ are regarded as heavy metals (HMs) (Alloway [1995](#page-10-0); Singh et al. [2016](#page-12-0)). Different anthropogenic activities are the major source of HM contamination in soil and water bodies thus causing serious problems for living organisms (Rouphael et al. [2008](#page-12-1); Rizwan et al. [2016](#page-12-2); Qayyum et al. [2017\)](#page-12-3). The most widespread anthropogenic sources of HMs include industries, indiscriminate utilization of agricultural inputs, metalliferous mining and smelting, military warfare and trainings, sewage sludge disposal, automobile emissions, and open waste dumping (Alloway [1995](#page-10-0); Shi et al. [2005;](#page-12-4) Rehman et al. [2015](#page-12-5); Rizwan et al. [2017](#page-12-6)). Plants take up and accumulate HM ions from the surrounding environment. The accumulation of HMs induces toxic

effects on plant growth attributes as well as on consumer health (Stobrawa and Lorenc-Plucinska [2008;](#page-12-7) Jaishankar et al. [2014](#page-11-0); Afshan et al. [2015](#page-10-1); Abbas et al. [2017\)](#page-10-2). According to the U.S. Environmental Action Group, the toxicity caused by various HMs is one of the major reasons affecting health of more than 10 million people and posing serious threats to sustainable agricultural development in eight countries such as China, Russia, Ukraine, India, Dominican Republic, Kyrgyzstan, Peru, and Zambia (Khan et al. [2015](#page-11-1)). The exposure to low doses of HMs for a long period of time disturbs plant functions and human health (Fu et al. [2008](#page-11-2); Adrees et al. [2015a;](#page-10-3) Jabeen et al. [2016](#page-11-3); Rehman et al. [2017](#page-12-8)). A number of reviews have been surveyed to observe the effects of HMs on plant growth and development (Babula et al. [2009;](#page-11-4) Adrees et al. [2015b](#page-10-4); Singh et al. [2016](#page-12-0); Farid et al. [2018](#page-11-5); Rizwan et al. [2018](#page-12-9)). HMs may exist in dissolved or suspended form. The suspended form is more harmful to plants as compared to the dissolved form (Alloway [1995](#page-10-0); Keller et al. [2015](#page-11-6)). Metals such as cadmium (Cd), copper (Cu), molybdenum (Mo), nickel (Ni), lead (Pb), zinc (Zn), and beryllium (Be) are more detrimental to crops as compared to manganese (Mn), iron (Fe), arsenic (As), and mercury (Hg) (Ali et al. [2018\)](#page-10-5). Some HMs (Cu, Mo, Mn, and Zn) at lower concentrations serve as vital nutrients to plants and animals but are toxic at higher levels (Jarup [2003](#page-11-7); Azevedo and Lea [2005;](#page-11-8) Rehman et al. [2016](#page-12-10)).

Plant growth regulators (PGRs) play important regulatory roles in signaling networks, developmental processes and engineering tolerance in plants either directly or indirectly to an array of biotic and abiotic environmental stresses (Ahmad et al. [2018;](#page-10-6) Bali et al. [2018;](#page-11-9) Jan et al. [2018](#page-11-10); Wani et al. [2016;](#page-13-0) Handa et al. [2018\)](#page-11-11). 5-Aminolevulinicacid (ALA) is a white to off white, odorless and water soluble crystalline solid PGR, having a molecular weight of 167.59 g/mol. The chemical name for ALA is 5-amino-4 oxypentanoic acid hydrochloride with structural formula $C_5H_9NO_3HCl$ (Fig. [1\)](#page-1-0). ALA is the main predecessor in the biosynthesis of porphyrins such as, chlorophyll and hemeA glutamyltRNA intermediates, in plants. ATP and NADPH as cofactor react to synthesize ALA from glutamate which, at lower concentration helps in chlorophyll biosynthesis (Castelfranco et al. [1974](#page-11-12); Beale [1990](#page-11-13); Hotta et al. [1997b\)](#page-11-14). ALA has both growth enhancing and stress-tolerance abilities. It has

been observed that ALA at lower concentrations improves plant growth and yield (Hotta et al. [1997a,](#page-11-15) [b;](#page-11-14) Roy and Vivekanandan [1998](#page-12-11); Watanabe et al. [2000\)](#page-13-1). An increase in chlorophyll content and photosynthetic ability was also noted at lower concentrations of ALA. ALA strengthens plants against abiotic stress by increased enzymatic activities or by improving the antioxidant defense system that protects the plant membrane system against reactive oxygen species (Korkmaz [2012\)](#page-11-16). It also promotes vegetative expansion (Tanaka and Tanaka [2011;](#page-12-12) Naeem et al. [2010;](#page-12-13) Zavgorodnyaya et al. [1997\)](#page-13-2) and helps the plants against harmful effects of different abiotic stresses including HMs. It is observed that enzymes involved in the production and breakdown of ALA play an important role in chlorophyll synthesis (Tanaka et al. [1992;](#page-13-3) Bogorad [1967](#page-11-17)). ALA not only stimulates plant development but also enhances yield through improvement in carbon dioxide fixation and nitrogen assimilation (Tanaka et al. [1992](#page-13-3)). Other notable changes include vegetation greenness (Maruyama-Nakashita et al. [2010](#page-12-14)), improved rate of photosynthesis (Akram and Ashraf [2011b;](#page-10-7) Hotta et al. [1997b](#page-11-14)), enhanced chlorophyll activity (Tanaka and Tanaka [2007,](#page-12-15) [2011](#page-12-12); Naeem et al. [2010](#page-12-13)), and better functioning of nitrite reductase (Tanaka and Tanaka [2007;](#page-12-15) Mishra and Srivastava [1983\)](#page-12-16). Moreover, ALA was effective in terms of promoting plant growth and development at lower concentrations (Chakraborty and Tripathy [1990;](#page-11-18) Balestrasse et al. [2010](#page-11-19)). On the contrary, ALA at higher concentrations promotes increased production of reactive oxygen species (ROS), which cause oxidative stress in plants (Pattanayak and Tripathy [2011](#page-12-17); Balestrasse et al. [2010\)](#page-11-19). However, the underlying mechanisms of stress-tolerance and the extent of crop yield improvement by ALA is not fully exposed yet. Considering the importance of ALA, the present review was accomplished to understand the cost-effectiveness and efficacy of ALA-mediated improvement in plant growth and development under different HM stress conditions (Fig. [2](#page-2-0)).

Plant Response to Environmental Stresses

Different biotic and abiotic stresses (HM, drought, salinity, temperature, pathogen and pest) lead to physiological, morphological, biochemical, and molecular changes in plants. HMs, temperature, salinity, and drought stimulate cellular damage by orchestrating the rate of reactive oxygen species (ROS) production. ROS are produced in the apoplast of plant cells via NADPH oxidases and accumulate in the mitochondria, chloroplasts and even in the nucleus (Van Breusegem et al. [2008](#page-13-4); Czarnocka and Karpiński [2018\)](#page-11-20) as demonstrated in Fig. [3](#page-3-0). HM stress, like drought and salinity, leads to distraction of ion circulation, the photosynthetic mechanism, and growth of the cells (Rucinska-Sobkowiak [2016\)](#page-12-18). Oxidative stresses which normally harmonize with

Fig. 2 Biosynthesis of ALA from various intermediates and vice versa

drought, salinity, and temperature can cause protein denaturation resulting in altered configurations and functions. Consequently, these adverse environmental stresses stimulate identical cell signaling corridors and cellular reactions. The biochemistry of the cells is changed during biochemical alteration in plants. These adaptations give rise to metabolic pathways in the form of lower molecular weight metabolites, generation of particular proteins, a mechanism of toxicity removal, and variations in phytohormone levels. The aptitude of living organisms to cope with varying environmental conditions enhances their endurance and reproduction capacity (Fujita et al. [2006;](#page-11-21) Smirnoff [1995\)](#page-12-19).

HMs are taken up by the plants from the external environment via specific transporters (Fig. [3\)](#page-3-0) like zinc–iron permease (ZIP), heavy metal transport ATPase (CPxandP1B-ATPase), natural resistant associated macrophage protein (NRAMP), cation diffusion facilitator (CDF), and ATP-binding cassette (ABC) transporters which are either present at the plasma membrane or vacuolar membranes (Park et al. [2012\)](#page-12-20). In addition to transporters, there are cysteine-rich metal binding peptides like phytochelatins (PCs) or metallothionines

(MTs) and glutathione (GSH) which are also important players in the metal transport system in plants (Jalmi et al. [2018\)](#page-11-22) (Fig. [3](#page-3-0)). Plants require both essential metals (micronutrients) and metalloids (B, Cu, Fe, Mn, Mo, Ni, and Zn) for appropriate metabolic and physiological system functioning in optimal and suboptimal environmental conditions. However, HMs that are essential for growth and metabolism of plants cause toxic effects on metabolic pathways at higher concentrations. The HMs exert toxicity by overcrowding of functional groups like enzymes, nutrients and the ion transport system or substitution of important ions on cellular sites, enzyme denaturing, and distraction of cell reliability. HMs also impose toxicity by free radical formation, that is, ROS overproduction (Smirnoff [1995\)](#page-12-19).

Fig. 3 Uptake of various HMs ions through various transporters, their sequestration

Importance of ALA in Plant Growth Constraints

In recent years, the use of plant growth regulators to mitigate HM stress has been regarded as a good strategy for sustainable agriculture production (Sakpirom et al. [2018](#page-12-21)). 5-Aminolevulinic acid has great importance in many physiological and biochemical processes in plants (Yossouf and Awad [2008;](#page-11-23) Tanaka et al. [1992;](#page-13-3) Beale and Castelfranco [1974](#page-11-24); Sasaki et al. [2002](#page-12-22); Korkmaz et al. [2010;](#page-11-25) Balestrasse et al. [2010](#page-11-19); Li et al. [2011](#page-11-26)). A number of studies have shown that ALA enhanced plant growth by increasing chlorophyll levels and photosynthesis rates in different plants like rice (Hotta et al. [1997a](#page-11-15)), barley, garlic, potato (Tanaka et al. [1992](#page-13-3)), and date palm (*Phoenix dactylefera*) (Al-Khateeb et al. [2006\)](#page-10-9). ALA has also been observed to improve the carbon to nitrogen ratio (Maruyama-Nakash-ita et al. [2010\)](#page-12-14). ALA at a higher concentration (\geq 5 mM) also showed herbicidal effect on plants (Kumar et al. [1999](#page-11-27); Tanaka and Tanaka [2007](#page-12-15)). Normally, antioxidants (enzymatic and non-enzymatic) have extensive roles in promoting plant growth by controlling the production of reactive oxygen species (ROS) during stress and non-stress conditions (Shahbaz and Ashraf [2008](#page-12-23); Mittler [2002;](#page-12-24) Ashraf and Akram [2009](#page-10-10); Perveen et al. [2010](#page-12-25), [2011,](#page-12-26) [2012;](#page-12-27) Akram et al. [2012](#page-10-11)). ALA also plays an important role in the plant antioxidant defense system (Xu et al. [2009;](#page-13-5) Akram et al. [2012](#page-10-11); Memon et al. [2009\)](#page-12-28). It was observed that ALA helps to enhance antioxidant enzyme activities like peroxidase (POD), catalase (CAT), and superoxide dismutase (SOD) in pakchoi (*Brassica campestris*) leaves. Moreover, an increase in chlorophyll content and photosynthetic rate was also recorded in *Brassica campestris* (Memon et al. [2009](#page-12-28)). ALA at lower concentrations stimulated the production of antioxidant enzymes such as POD, CAT, ascorbate peroxidase (APX) and SOD in *Ginkgo biloba* (Xu et al. [2009\)](#page-13-5). The elevated levels of non-enzymatic antioxidants like glutathione (GSH) have also been observed under ALA treatment. However, a decline in dehydroascorbate (DHA) and no alteration in GSSG level were also evident in *Ginkgo biloba* as a result of ALA treatment. According to these observations, ALA also behaves as a light capturing pigment (Senge [1993;](#page-12-29) Burnham and Lascelles [1963](#page-11-28); Sano and Granick [1961\)](#page-12-30). Reports indicate that ALA has an important role in mineral nutrient uptake and transformation in kudzu (*Pueraria phaseoloides*). Substantial improvements in kudzu dry biomass, chlorophyll content, photosynthetic rate, stomatal conductance, starch,

nutrients, and sugar levels were observed as a result of ALA application (Xu et al. [2010\)](#page-13-6). Awad ([2008\)](#page-11-23) reported that ALA appreciably enhanced photosynthetic pigments and the chlorophyll a/b ratio in date palm.

Significance of ALA against Abiotic Stress

ALA application is also used to mitigate the effects of abiotic stresses on plants. Induction of temperature stress-tolerance has also been reported in rice, potato and soybean with the application of ALA (Balestrasse et al. [2010](#page-11-19); Hotta et al. [1998](#page-11-29)). ALA treatment under reduced light conditions enhanced chilling resistance by controlling constraints in chlorophyll biosynthesis, photosynthesis and respiration (Wang et al. [2004](#page-13-7)). ALA applications improved grain yield of wheat (Al-Thabet [2006\)](#page-10-12) and barley under drought stress (Al-Khateeb et al. [2006](#page-10-9)). Its applications also reduced salt stress in crops like date palm (Youssef and Awad [2008\)](#page-13-8), oilseed rape (Naeem et al. [2010\)](#page-12-13), potato (Zhang et al. [2006](#page-13-9)), pakchoi (Wang et al. [2005\)](#page-13-10), and spinach (Nishihara et al. [2003](#page-12-31)). The ALA treatment controls many physiological processes related to plant growth under salinity stress like seed germination, photosynthesis, and absorption of nutrients (Hotta et al. [1997a,](#page-11-15) [b;](#page-11-14) Youssef and Awad [2008](#page-13-8)). ALA application also enhanced photosynthetic activity in cucumber under salt stress (Wu et al. [2018\)](#page-13-11). The application of ALA not only enhanced nutrient uptake in plants under stressed and non-stressed conditions (Akram et al. [2018\)](#page-10-13) but also improved the antioxidant defense system under chilling stress (Liu et al. [2018\)](#page-12-32).

ALA as Stress‑Tolerance Agent

ALA has the ability to mitigate harmful effects of stress on plant growth and production (Akram et al. [2012](#page-10-11)). However, the physiological mechanism of stress alleviation by ALA still remains unclear. Different abiotic stresses such as salinity, temperature, drought and heavy metals, and so on, modify the ALA status in plants (Hodgins and van Huystee [1986](#page-11-30); Cornah et al. [2003](#page-11-31); Genisel et al. [2017\)](#page-11-32). However, it is not totally obvious which metabolic process is disturbed in plants with modified ALA status.

ALA Against Heavy Metal Stress

Plants under HM stress experience different harmful effects. ALA acts as a plant growth regulator and resists the toxic effects of heavy metals. The ameliorative role of ALA under HM stress in plants is discussed below (Table [1](#page-5-0)).

Plant Growth, Biomass, and Chlorophyll Contents

Usually, HMs at higher concentrations disturb plant growth attributes, biomass and chlorophyll contents. A considerable decline has been reported in plant height, root length, number of leaves, dry and fresh weight of roots, stem and leaves, and root morphological parameters like root diameter, root tips, root surface area, and root volume of *Brassica napus* at higher Cd concentrations. However, ALA application at the 25 mg/L concentration significantly mitigated the adverse effects of Cd and appreciably improved all the above-mentioned plant growth attributes (Ali et al. [2013a,](#page-10-14) [b](#page-10-15); Xu et al. [2015](#page-13-12)). ALA application under Cd stress extensively enhanced chlorophyll a, b and total chlorophyll and considerably reduced metal concentrations in shoots and roots of *B. napus* (Ali et al. [2013a\)](#page-10-14). In a recently reported study, sunflower plants under Cr stress had significantly reduced plant growth attributes and photosynthetic pigments. The application of ALA markedly enhanced plant growth under Cr stress (Farid et al. [2018\)](#page-11-5). The application of ALA in cauliflower enhanced plant growth, photosynthetic pigments, and antioxidant enzymes activities under Cr stress (Ahmad et al. [2017\)](#page-10-16). Higher concentrations of Pb also extensively reduced the fresh and dry weight of *Brassica napus* leaf, stem and roots. However, ALA application alleviated the toxic effects of Pb and enhanced plant biomass under Pb stress. ALA application without Pb stress showed no alteration in plant biomass (Ali et al. [2014\)](#page-10-17). Tian et al. ([2014](#page-13-13)) reported that combined application of ALA and Pb promoted root surface area, root diameter, root volume, number of root tips and total chlorophyll content as compared to Pb treatment only. Lead considerably reduced chlorophyll contents in *B. napus*. However, ALA application under Pb stress showed no changes in chlorophyll contents (Ali et al. [2014](#page-10-17)). By increasing the concentration of ALA Xiaomenget al. ([2018](#page-13-14)) reported an increase in chlorophyll a/b in *Matricaria recutita* plants. The possible reason for enhancement in plant growth might be due to the potential of ALA to resist HM-induced stress and reduce lipid peroxidation (Youssef and Awad [2008](#page-13-8)) by triggering the heme-based antioxidant system to scavenge overproduced ROS (Nishi-hara et al. [2003\)](#page-12-31). The application of ALA confers tolerance against metal stress by reducing the uptake of toxic metals by the plant, improving the nutrient level, photosynthetic pigments and upholding of the photosynthetic mechanism. Generally, ALA has a promotive role in regulating a number of metabolic processes and improves plant growth and yield under HM toxicity.

Variations in Photosynthetic Traits and Carotenoid Contents

It is well documented that photosynthetic traits like stomatal conductance, net photosynthesis and transpiration rate

Table 1 Promotive effect of ALA on plant physio-chemical traits under heavy metal stress

HM stress	ALA dosage	Plant species Effect		References
Cd at 100 μ M concentration 25 mg/L		B. napus L	Increase in fresh biomass 47.07%, dry biomass 65.58%, root diameter 7.66%, root surface area 20.31%, number of root tips 23.32%, root volume 10.99%, decrease in MDA content by producing 9.97 nmol/mg protein	Ali et al. (2013b)
Cd at 500 μ M concentration 25 mg/L		B. napus L	Enhanced fresh biomass by 115.14%, dry biomass 165.51%, root diameter 42.35%, root surface area 51.75%, number of root tips 19.20%, root volume 99.01%, decrease MDA content by producing 15.31 nmol/mg protein	Ali et al. (2013b)
Cd at 100μ M concentration 25 mg/L		B. napus L	Decrease H ₂ O ₂ by 34.24%, O ₂ ⁻ 10.47%, ⁻ OH 63.04%	Ali et al. (2013b)
Cd at 500 µM concentration 25 mg/L		B. napus L	Decrease H_2O_2 by 36.26%, O_2 ⁻ 63.47%, ⁻ OH 50.01%	Ali et al. (2013b)
Cd at 100 μ M concentration 25 mg/L		B. napus L	Increase in fresh biomass 47.07%, dry biomass 65.58%, root diameter 7.66%, root surface area 20.31%, number of root tips 23.32%, root volume 10.99%, decrease in MDA content by producing 9.97 nmol/mg protein	Ali et al. (2013b)
Cd at 500 μ M concentration 25 mg/L		B. napus L	Enhanced fresh biomass by 115.14%, dry biomass 165.51%, root diameter 42.35%, root surface area 51.75%, number of root tips 19.20%, root volume 99.01%, decrease MDA content by producing 15.31 nmol/mg protein	Ali et al. (2013b)
Cd at 100μ M concentration 25 mg/L		B. napus L	Decrease H_2O_2 by 34.24%, O_2 ⁻ 10.47%, ⁻ OH 63.04%	Ali et al. (2013b)
Cd at 500 μ M concentration 25 mg/L		B. napus L	Decrease H ₂ O ₂ by 36.26%, O ₂ ⁻ 63.47%, ⁻ OH 50.01%	Ali et al. (2013b)
Pb at 100 and 400 μ M conc 25 mg/L		B. napus L	No improvement in leaves chlorophyll content, improve plant biomass significantly and reduce Pb stress, enhance macronu- trient $(N.P,K)$ in shoot and root	Ali et al. (2014)
Pb at 100 and 400 μ M conc 25 mg/L		B. napus L	Decreased ROS, MDA in leaves and roots and also alleviated Pb stress, stimulated APX activity and no significant effect on CAT activity in shoot and root, no considerable effect on glutathione reductase (GR) and total soluble protein (TSP) in leaves, increased TSP in roots, enhanced ascorbic acid (AsA) in roots, improvement in GSH contents in leaves and root	Ali et al. (2014)
Pb at 400 µM conc	25 mg/L	B. napus L	Enhanced total glutathione by 20.43% and 34.69% in the leaves and roots	Ali et al. (2014)
Cd at 100 µM conc	2 mg/L	B. napus L	Longest shoot of 6.76 cm and longest root 2.77 cm, increase GR activity by 26%	Ali et al. (2013b)
Cd at 500 µM conc	2 mg/L	B. napus L	Longest shoot of 6.12 cm and root 2.36 cm, reduce MDA and H_2O_2 , increase GR activity by 18%	Ali et al. (2013b)
Cd at 100 and 500 μ M conc 0.4 and 2 mg/L B. napus L			Enhance Chl a, Chl b, total chlorophyll and carotenoids	Ali et al. (2013b)
	2 mg/L		increased SOD and POD activity, ultra-structural changes in nucleus and chloroplast	

decline under Cd stress. However, application of ALA, with or without Cd stress, shows improvement in photosynthetic traits. The ameliorative effect of ALA under Cd stress has been observed at the 25 mg/L concentration (Ali et al. [2013a](#page-10-14)). The combination of ALA and Pb also enhanced stomatal conductance, net photosynthetic rate and transpiration rate in oilseed rape (Tian et al. [2014\)](#page-13-13). The above-mentioned results indicate the positive impact of ALA on photosynthetic traits under HM stress.

Effects on Malondialdehyde and Reactive Oxygen Species

Malondialdehyde (MDA) is a signal molecule of free radicals in plant tissues. Its concentration may vary under varying environmental stresses. The MDA contents in roots were higher under Cd stress whereas ALA application with Cd stress extensively reduced MDA production. Minimum production of MDA was observed at 25 mg/L of ALA (Ali et al. [2013b](#page-10-15), [2015\)](#page-10-18). Cadmium stress enhanced the production of ROS in oilseed rape (*Brassica napus* L.). ALA at 25 mg/L reduced H_2O_2 , O_2 and OH contents under Cd stress (Ali et al. [2013b\)](#page-10-15). However, ALA application without Cd stress showed no decrease in MDA and ROS production in the leaves and roots of *B. napus*. Like Cd, Pb stress also enhanced MDA contents in oilseed rape. However, ALA application with Pb stress considerably declined MDA contents in the leaves and roots of *B. napus*. Lead at higher concentrations alone increased ROS production. ALA application with 100 and 400 µM concentrations of Pb reduced ROS contents in the leaves and roots of *B. napus* (Ali et al. [2014](#page-10-17)).

The above reports showed that MDA and ROS generation was increased under HM stress. However, ALA application reduced MDA and ROS generation and conferred resistance against HM stress by improving antioxidant enzyme activity.

Changes in Antioxidant Enzymes and Total Soluble Protein Content

The HMs like Cd without ALA application reduced SOD and POD activities in *B. napus*. However, combined application of ALA and Cd significantly enhanced SOD and POD activities. Application of ALA (25 mg/L) alone showed momentous alteration in APX activity. But combined application of ALA and Cd treatments enhanced APX and CAT activities in *B. napus* (Ali et al. [2013b\)](#page-10-15). Contrary to that, increasing Pb concentrations without ALA application enhanced POD and declined SOD activity. ALA applications mitigated the negative effect of Pb. The ALA application under Pb stress promoted SOD activity in the leaves and roots of *B. napus*. ALA at higher Pb concentrations showed no change in POD activity. It was observed that Pb at 100 and 400 µM concentrations declined APX activity in the leaves and roots of plants. However, ALA application under Pb stress enhanced APX activity in the leaves and roots of *B. napus*. Increasing Pb concentrations enhanced CAT activity in the roots and declined in the leaves of *B. napus*. Reports showed that ALA application had no momentous effect on CAT activity under Pb stress in the leaves and roots of *B. napus*. ALA application without metal stress showed no appreciable effects on antioxidant enzyme activities (Ali et al. [2014\)](#page-10-17). Higher Pb concentrations reduced total soluble protein extensively in the leaves and roots of *B. napus*. The ALA application under Pb stress enhanced total soluble proteins in the roots, whereas no change was observed in the leaves of *B. napus* (Ali et al. [2014](#page-10-17)). Ali et al. [\(2013b\)](#page-10-15) investigated that ALA and Cd applications, alone or in combination, showed no impact on total soluble protein in *B. napus* roots. From the above reports, it was found that ALA application in combination with HMs enhanced the activity of antioxidant enzymes. It is assumed that plants under ALA application might be more efficient at neutralizing excess ROS.

Alteration in Nutrients Uptake

Lead stress enhances nitrogen concentrations in plant leaves as compared with roots. ALA application under Pb stress reduces Na⁺ uptake in plant roots. ALA application without metal stress considerably increased Ca^{2+} uptake in *B. napus* leaves. Combined application of ALA and Pb extensively enhanced potassium and calcium ions in leaves and roots, and magnesium in leaves of *B. napus* (Ali et al. [2014](#page-10-17)). Application of Pb considerably decreased micro and macronutrients in the leaves and roots of *B. napus* but at lower concentrations showed no noticeable change in Cu and Zn in leaves or Cu and S in roots. The ALA application without metal stress showed no alteration in nutrient concentrations. However, ALA application alone also reduced Fe in leaves but Zn and P in roots of *B. napus*. The ALA along with Pb applications notably enhanced the macronutrients N, P, and S concentrations in the leaves and roots of *B. napus*. Similarly, Zn concentration was enhanced in the roots of *B. napus* under ALA and Pb combined application. The ALA application with lower concentrations of Pb improved Fe content in *B. napus* roots (Ali et al. [2014\)](#page-10-17). The above results elucidate that HM stress disrupts nutrient uptake dynamics in plants. However, ALA application mitigates the toxic effects of HMs and enhances the plant's ability to accumulate micro and macronutrients to maintain optimum nutrient assimilation.

Modification in Glutathione Reductase (GR), Ascorbic Acid (AsA) and GSH Contents

Reports showed that Pb at lower and higher concentrations without ALA application declined GR activity in plant roots. However ALA application under Pb stress showed no considerable effect on GR activity in the leaves of *B. napus*. Higher Pb concentration alone notably decreased AsA contents in the leaves and roots of *B. napus*. The ALA application significantly mitigated Pb toxicity and promoted AsA contents in the roots of *B. napus* (Ali et al. [2014\)](#page-10-17). AsA demonstrated no considerable change under ALA application alone. While at 25 mg/L of ALA dosage, appreciable enhancement in AsA content was noted (Ali et al. [2013b](#page-10-15)). Higher concentrations of Pb declined GSH and GSSG contents in the leaves. The ALA application with Pb stress enhanced GSH content in the leaves and roots of *B. napus*. No change in GSH/GSSG ratio of leaves and roots was observed under Pb stress. Nonetheless, ALA application under Pb stress showed a promotive effect on the total glutathione contents (Ali et al. [2014](#page-10-17)). Cadmium stress showed a steady decline in glutathione contents. ALA at the 25 mg/L concentration slowly enhanced GSH contents under higher Cd concentrations (Ali et al. [2013b](#page-10-15)). From the above discussion it is clear that ALA application under metal stress enhanced GSH, AsA, and total glutathione content. The ameliorative effect of ALA might be due to enhanced resistive ability of plants against ROS that triggers the production of GSH and total glutathione.

Effect on Metal Uptake

ALA application reduced Pb uptake in shoots and roots of *B. napus*. ALA application at 25 mg/L decreased Pb uptake in shoots and roots at higher Pb concentration (Tian et al.

[2014](#page-13-13)). Report showed that ALA mitigates the toxic effects of metals by improving plants resistance against HM stress.

Ultra-structural Changes

Higher Pb concentration totally injured mesophyll cells and reduced intercellular spaces. Lead at a higher concentration also showed harmful effects on chloroplast thylakoid membranes. ALA application alleviated the damaging effect of Pb on leaf ultrastructure in rapeseed plant. ALA at 25 mg/L showed appreciable changes in root cell tips under Pb stress (Tian et al. [2014\)](#page-13-13). ALA application under HM stress improves antioxidant enzyme activity and reduces ROS production, avoiding ultra-structural damage in plant roots as documented by Castelfranco and Jones ([1975\)](#page-11-33).

ALA Application Methods

Different approaches of ALA application at various concentrations play a considerable part in the plant physio-chemical attributes under stress conditions (Youssef and Awad [2008](#page-13-8); Hotta et al. [1997a](#page-11-15), [b;](#page-11-14) Akram and Ashraf [2011a](#page-10-19), [b;](#page-10-7) Naeem et al. [2010](#page-12-13); Tanaka and Tanaka [2011\)](#page-12-12). The following are the different modes of ALA applications as plant growth promoter.

Foliar Method

One of the methods used to enable plants to remain healthy against stressful conditions is the foliar application of ALA, which was found helpful in enhancing growth and related attributes of various plants.

Non-stress Condition

Foliar application of ALA (50–250 mg/L) in pakchoi plants increased photosynthetic rates. Antioxidant enzyme activities and total chlorophyll contents were improved by ALA under normal conditions (Memon et al. [2009](#page-12-28)). An improvement in plant biomass, photosynthetic rate, chlorophyll contents, nutrient level and stomatal conductance were also observed by foliar application of ALA under a non-stressed environment in kudzu plants (Xu et al. [2010](#page-13-6)). Foliar ALA application increased antioxidant enzyme activities, GSH and H₂O₂ under normal conditions in the *Ginkgo biloba* plant (Xu et al. [2009](#page-13-5)). ALA foliar application also enhanced chlorophyll content in date palm (Al-Khateeb et al. [2006](#page-10-9)). The above reports showed that foliar applications of ALA at lower concentrations considerably promoted plant growth attributes in terms of photosynthetic traits, nutrients, antioxidant enzyme activity, and chlorophyll content under no stress conditions.

Stress Condition

Foliar application of ALA under drought stress conditions improved plant growth traits and yield in wheat (Al-Thabet [2006](#page-10-12)). Under cold stress, foliar application of ALA enhanced plant biomass, proline, and chlorophyll contents in pepper. Low temperature stress also enhanced SOD activity in pepper (Korkmaz et al. [2010](#page-11-25)). ALA also induced tolerance against chilling stress by improving photosynthetic attributes, chlorophyll b content and stomatal opening in melon (*Cucumis melo*) plants (Wang et al. [2004\)](#page-13-7). Foliar application of ALA under heat stress enhanced plant growth traits by reducing MDA content, superoxide radicals $(O^{2−})$, and H_2O_2 in cucumber plants (Zhen et al. [2012](#page-13-15)). Salt stress in spinach enhanced antioxidant enzyme activities and photosynthetic traits under ALA foliar application (Nishihara et al. [2001](#page-12-33), [2003](#page-12-31)). Foliar application of ALA at 20–80 mg/L enhanced plant growth attributes in sunflower (Akram and Ashraf [2011a\)](#page-10-19).

The above reports showed the prospective advantages of foliar applied ALA in mitigating the harmful effects of unfavorable environments in different plants. Reports also suggested that foliar application of ALA might have a great effect at the early growing stages like germination and seedling establishment.

Seed Treatment Before Sowing

Seed treatment with ALA before sowing enhances plant development in addition to physiological attributes under stressed and non-stressed conditions (Korkmaz and Korkmaz [2009](#page-11-34); Wang et al. [2005;](#page-13-10) Korkmaz et al. [2010\)](#page-11-25).

Non-stressed Condition

An improvement in plant growth traits, chlorophyll content, photosynthetic attributes and yield was noticed without stress conditions in seed treatment of *Vigna* species (Roy and Vivekanandan [1998\)](#page-12-11). Pre-sowing treatment of rice and horseradish seeds with ALA, enhanced plant growth traits, total chlorophyll content, and yield under normal conditions (Hotta et al. [1997a](#page-11-15)).

Stress Condition

Reports showed that pre-sowing seed treatment with ALA application under cold stress improved the germination rate in pepper (Korkmaz and Korkmaz [2009\)](#page-11-34). Pre-sowing seed treatment with ALA enhanced seed germination under salinity stress in pakchoi (Wang et al. [2005](#page-13-10)). The ALA seed treatment promoted photosynthetic ability, antioxidant enzyme activity, and chlorophyll contents under low light conditions in watermelon (Sun et al. [2009\)](#page-12-34). The above reports suggested that seed priming with ALA appreciably stimulated seed germination, plant growth parameters, the antioxidant defense system, and improved yield in different plant species under stress and normal conditions. Practically, seed priming with ALA could be used as a valuable tool to promote plant growth and production under different stresses.

Root Medium

Soil application of ALA has been reported to promote plant growth attributes and production under stressed and normal conditions.

Non-stressed Condition

An enhancement in chlorophyll a and total chlorophyll content in date palm was noted under normal growth conditions and ALA as a Pentakeep fertilizer (Awad [2008\)](#page-11-23). Yoshida et al. ([2005\)](#page-13-16) noted improvement in tulip bulbs under normal conditions at 2000–5000 times dilution of Pentakeep.

Stress Condition

Under cold stress, ALA applied in the root medium of soybean enhanced chlorophyll and antioxidant enzyme activity (Balestrasse et al. [2010](#page-11-19)). The ALA treatment enhanced photosynthetic traits, stomatal conductance and chlorophyll content in *Cucumis melo* (Wang et al. [2004\)](#page-13-7). Under salinity stress, ALA treatment promotes tuber formation in potato (Zhang et al. [2006](#page-13-9)). ALA application under cold stress enhances rice dry weight and seedling growth (Balestrasse et al. [2010](#page-11-19)). Similarly, ALA application enhanced photosynthetic traits, chlorophyll contents, stomatal conductance, and respiration rate under cold stress in melon (Wang et al. [2004](#page-13-7)). ALA treatments through the roots neutralize salinity stress and enhance antioxidant enzymes activities in cucumber (Zhen et al. [2012](#page-13-15)). The above reports suggest that ALA application in the plant growth medium under different stresses could mitigate the toxic effects by improving plant physio-chemical traits. ALA addition in the soil medium is not economical as a large amount of soil is required for this purpose. Hence, further study is needed to modify the effectiveness of ALA in the soil medium to suppress undesirable effects of environmental stresses.

Assessment of ALA Application Methods

All the above-mentioned methods were reported to be beneficial in improving plant growth attributes and physiological processes (Fig. [4](#page-9-0)). ALA is an expensive compound and present in plants in minute quantities. So application methods that require large quantities of ALA are not cost-effective.

Foliar application usually depends on the leaf and plant growth stage (Akram et al. [2012;](#page-10-11) Ashraf and Foolad [2007](#page-11-35); Ashraf et al. [2010a](#page-11-36)). Chlorophyll production or its use is more dynamic at the early developmental stages in plants (Xu et al. [2010;](#page-13-6) Memon et al. [2009](#page-12-28); Naeem et al. [2010;](#page-12-13) Tanaka and Tanaka [2011\)](#page-12-12). Therefore, the application of ALA as a foliar spray appreciably affects early growth stages like germination and seedling establishment. ALA at a low dosage (1–100 mg L^{-1}) promotes plant growth and yield while at higher concentration (\geq 150 mg L⁻¹) causes inhibitory effects on plants (Tanaka and Tanaka [2007](#page-12-15); Sasaki et al. [2002](#page-12-22)).

Genetic Treatment

Crop resistance against environmental stresses by genetic manipulation is one of the concerned issues being discussed nowadays. Numerous plants have been developed for improvement in stress resistance by enhancing hormone production, organic osmolytes, antioxidants and nutrients (Mittler [2002](#page-12-24); Yamaguchi and Blumwald [2005;](#page-13-17) Ashraf and Akram [2009;](#page-10-10) Ashraf et al. [2010a](#page-11-36); Munns and Tester [2008](#page-12-35)). ALA application also enhanced the plant's ability to resist environmental stress. The application of ALA synthase in fungi enhanced the ALA level in this organism as documented by Elrod et al. ([2000](#page-11-37)). Fewer reports are available regarding stress resistance in plants and their ALA accumulation level. Jung et al. [\(2004\)](#page-11-38) reported the addition of the *Bradyrhizobium japonicum* aminolevulinic acid synthase (ALA-S) gene in rice. Synthesis of ALA was enhanced significantly causing improvement in chlorophyll content. The authors conclude that ALA is the main precursor in chlorophyll biosynthesis. Increase in ALA-mediated chlorophyll biosynthesis can lead to enhanced net photosynthesis. Endogenous ALA biosynthesis can also optimize different growth and developmental processes in plants. The above reports showed enhancement in ALA production by genetic manipulation of plants which could be exploited as a tool to impart ALA-mediated HM stress-tolerance in plants (Fig. [5](#page-9-1)).

Conclusions and Future Prospects

In this review, we intended to investigate ALA effects on plant growth traits and the mechanisms involved in plant promoting processes under HM stress. A detailed review from the available literature showed that HM stress restricts plant growth and productivity, posing a threat to food security for the ever-increasing population. ALA has the potential to reduce HM stress and enhance plant growth attributes and yield. It also reduces HM toxicity by increasing hemebased antioxidant enzyme activities. Application of ALA

Fig. 4 ALA application methods in controlling various morpho-physiological activities in plants

enhances plant growth traits under HM stress. Foliar application was found to be the most effective mode of application in promoting plant growth attributes. However, it was not determined what factors could enhance ALA uptake when applied exogenously. The pattern of ALA uptake needs to be investigated. Although ALA application showed an ameliorative effect on plant growth traits, there is still a gap in knowledge regarding the ALA physiological response and its function in signal transduction pathways in plants growing under HM stress. The accurate role of ALA in HM stress alleviation is needed in soil based environmental conditions. A broad range of study is needed at the molecular level to determine how ALA interacts with plant physio-chemical processes and promotes nutrient uptake under HM stress. Genetic exploitation is another area to be investigated with the objective of enhancing ALA production against HM stress. ALA is an expensive compound and naturally occurs in a very low quantity in plants. ALA is extracted from different microorganisms like bacteria and algae, etc. This is the need of the time to develop plants that have the potential to serve as a cheap source of ALA. Furthermore, a deep insight of the ALA-mediated defense networks/plant stresstolerance mechanism as well as its relationship with other defense signaling pathways in HM-stressed plants can be further investigated by exploring and expanding the knowledge of various approaches, that is, biotechnology, biochemistry, plant stress molecular biology, genomics, proteomics, ionomics, bioinformatics techniques, metabolomics, and computational biology.

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Compliance with Ethical Standards

Conflict of interest All the authors declare no conflicts of interest.

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